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# Molecular marker dissection of rice (*Oryza sativa* L.) plant architecture under temperate and tropical climates

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Abstract Rice (Oryza sativa L.) plants develop vertically with shoot elongation and horizontally with tillering. The purpose of this study was to identify and characterize genomic regions influencing the rice plant architecture by quantitative trait locus (QTL) analysis for the component traits: culm length (CL), panicle length (PnL), panicle number (PnN) and tiller number (TN). For this QTL analysis, 191 recombinant inbred lines (F7) derived from a cross of Milyang 23 (M23) and Akihikari (AK) were grown in 1995, 1996 and 1997 (May-Oct) in Joetsu, Japan (temperate climate), and in the 2000 dry season (Jan-Apr), the 2000 wet season (Jun-Oct) and the 2001 dry season in Los Baños, The Philippines (tropical climate). Results showed that rice plant architecture was influenced by 19 genomic regions categorized into five groups. In Group I, two regions (on chrs. 6 and 11) affected shoot elongation (CL and PnL) and tillering (PnN and TN) in opposite directions more significantly in Los Baños than in Joetsu. In Group II, two regions (chrs. 3 and 12) affected shoot elongation, whereas in Group III, five regions [chrs. 1 (two), 2, 3 and 9] affected only culm length (CL). Expressions of four regions of Group III were influenced by either tropical or temperate environments. In Group IV, seven regions (chrs. 1, 2, 4, 5, 6, 8

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Graduate School of Life Science, Tohoku University, Katahira, Aoba-ku, 980-8577 Sendai, Japan and 9) controlled panicle development (PnN or PnL), and in Group V, three regions (chrs. 1, 2 and 3) regulated tillering (PnN or TN). Characterizing these 19 genomic regions provided a detailed analysis of rice plant architecture with emphasis on the multiple effect and environmental responsive regions.

**Key words** Genomic region · Plant architecture · Quantitative trait locus (QTL) · Recombinant inbred line (RIL) · Rice (*Oryza sativa* L.)

### Introduction

Rice (Oryza sativa L.) plants develop vertically with shoot elongation and horizontally with tillering. Shoot elongation and tillering are usually represented by plant height (PH) and panicle number (PnN), respectively. PH consists of the length of the culm and panicle. Hence, culm length (CL), panicle length (PnL) and PnN are major traits composing rice plant architecture, and ultimately the grain yield (Sasahara 1997; Tanisaka 1997). Past studies have reported that these component traits are inherited quantitatively, related genetically to one another, and influenced by growing environments (Gravois and McNew 1993; Sasahara 1997; Tanisaka 1997). Because of these trait characteristics, it has been difficult to identify the genetic factors of each trait and to comprehend the genetic mechanisms of plant architecture in rice.

The recent development of molecular markers enabled researchers to identify gene loci affecting quantitative traits, such as CL, PnL and PnN, on a linkage map in rice (Yano and Sasaki 1997). Using the IR64/Azucena double-haploid population, Yan et al. (1999) identified nine and six quantitative trait loci (QTLs) for PH and PnN, respectively, and found that QTLs for both traits were detected in the same region of chr. 1. The QTLs of this region increased PH and decreased PnN with the Azucena allele. Yamamoto et al. (2001) likewise found two regions (chrs. 3 and 6) that increased CL and PnL through QTL

analysis using the Koshihikari/Kasalath  $BC_1F_3$  population. These genomic regions with multiple effects possibly show how the component traits are related to one another (Li et al. 1995; Lin et al. 1996; Yan et al. 1999). Their localization and characterization would help to understand the genetic mechanisms controlling rice plant architecture. However, QTL analyses that examined the three component traits simultaneously and focused on the multiple effect regions are not abundant.

The component traits change dramatically with growing environments; for example rapid culm elongation in floating varieties (Hamamura 1997). The changes are partly attributable to the genes (QTLs) expressed in a particular environment, in other words the interaction between QTLs and the environment (Lu et al. 1996; Yan et al. 1999; Yamamoto et al. 2000). Past QTL analyses measured CL, PnL and PnN at a fixed location (environment), thereby providing little acknowledgment of the  $QTL \times$  environment interaction. Exceptionally, Lu et al. (1996), Zhuang et al. (1997) and Yan et al. (1999) examined small mapping populations at a few locations in China (temperate and subtropical climates) and statistically quantified the effect of the QTL  $\times$  environment interaction on the component traits. Further analyses must be necessary to fully locate the QTLs interacting with the environments, especially the tropical dry and wet seasons, and to qualify the interactions.

The objective of this study was to clarify genomic regions affecting the component traits of plant architecture, and their interactions with environments. For this objective, 191 recombinant inbred lines (RILs) derived from a cross of Milyang 23 (M23) and Akihikari (AK) (Fukuta et al. 1999) were repeatedly grown under two major rice-growing climates, temperate (Japan) and tropical (Philippines), to locate QTLs for CL, PnL, PnN and tiller number (TN). M23 is an indica variety with heavy panicles, whereas AK is a *japonica* variety with a large number of panicles. Both varieties are high yielding and have been used in actual breeding programs (Yokoo et al. 1994). QTL analyses using the M23/AK cross are possible to disclose a typical genetic nature (background) underlying the different types of high yielding varieties (Yano and Sasaki 1997).

#### Materials and methods

Plant materials and phenotypic data collection

One hundred and ninety one RILs ( $F_7$  generation) derived from a cross between M23 and AK (Fukuta et al. 1999) were grown in irrigated field conditions during May–Oct in 1995 (95 J), 1996 (96 J) and 1997 (97 J) at the former Hokuriku National Agricultural Experiment Station (HNAES, currently the Hokuriku Research Center, 138° E; 38° N), Joetsu, Japan, and during the 2000 dry season (Jan–Apr, 00DS), the 2000 wet season (Jun–Oct, 00WS), and the 2001 dry season (01DS) at the International Rice Research Institute (IRRI, 121°E; 14°N), Los Baños, The Philippines. Joetsu and Los Baños belong to the temperate and tropical zones, respectively, both having annual rainfall of >2,000 mm per year (see Table 1 for detailed climatic data). Thirty day-old seedlings

 Table 1
 Climatic data of theJoetsu season (May–Oct), and the Los
 Baños dry (Jan–Apr) and wet (Jun–Oct) seasons<sup>a</sup>

Season		Air ten	nperatur	re (°C)	Radiation	Day length		
		Mean	Max	Min	(MJ m <sup>-2</sup> )	(h)		
Joetsu Los Baños	Dry Wet	20.1 26.7 27.5	25.6 28.7 28.4	15.7 26.8 27.0	2,757 2,134 2,465	13.4 11.8 12.3		

<sup>a</sup> Maximum and minimum indicate the highest and lowest monthly averaged air temperatures. Radiation and day length are the accumulated quantity and average, respectively

were transplanted with a single plant per hill at locally accepted densities: 15.4 plants  $m^{-2}$  in Joetsu (22.2 plants  $m^{-2}$  for 1996) and 16.7 plants  $m^{-2}$  in Los Baños. Nitrogen, phosphorus and potassium fertilizers were applied 70, 70 and 70 kg ha<sup>-1</sup>, respectively, at HNAES, and at 134, 66 and 66 kg ha<sup>-1</sup> at IRRI, following the experimental-farm recommendations based on soil fertility.

More than five plants were harvested 35 days after heading (when the plants were mature) to measure CL, PnL and PnN. TN was also examined in Los Baños since tillering continued in many RILs after heading under high air temperature and abundant rainfall. Based on planting density, PnN and TN were converted to a per-square-meter basis for data comparison between the two locations.

### QTL analysis

Using 182 restriction fragment length polymorphism (RFLP) markers, Fukuta et al. (1999) determined the genotype of marker loci and constructed a complete linkage map (see Fig. 1). With the genotypic data, interval mapping was performed using the computer software QGENE ver. 3.05 (Nelson 1997) and QTLs were identified by the following two steps. For the first step, all visible LOD peaks >2.0 were located, including the one near the highest peak. This threshold is suitable for detecting relatively minor-effect genes, according to Yano and Sasaki (1997). For the second step, the LOD peaks (>2.0), of which the nearest RFLP marker showed P < 0.01, were accepted as QTLs. This threshold has also been used commonly in recent single-point QTL analyses (Yano and Sasaki 1997). To further confirm our analysis, we focused on only the genomic regions that repeatedly showed QTLs for a trait across crop seasons. These regions were designated tentatively with the letter R based on each chromosome and region number.

### **Results and discussion**

### Trait segregation and correlation

The four traits examined showed a continuous frequency distribution with transgressive segregants for all seasons (Table 2). For CL, the mean value of the RILs was higher in 95 J, 96 J, 97 J and 00WS than in 00DS and 01DS. Parental differences were only observed in the latter two seasons where the mean value of M23 was larger than that of AK. The former four seasons were from May to Oct, whereas the latter two were the dry seasons from Jan to Apr. For PnL, the mean value of the RILs was larger in Joetsu (95 J, 96 J and 97 J) than in Los Baños (00DS, 00WS and 01DS), whereas for PnN, the mean value was larger in Los Baños than in Joetsu. These data showed

**Table 2** Culm length (CL), panicle length (PnL), panicle number (PnN) and tiller number (TN) in the Milyang 23/Akihikari RILs grown during 1995 (95 J), 1996 (96 J) and 1997 (97 J) in Joetsu, Japan, and during the 2000 dry season (00DS), the 2000 wet season (00WS), and the 2001 dry season (01DS) in Los Baños, The Philippines<sup>a</sup>

Trait	Season	RILs				M23		AK	
		Mean	SD	Max	Min	Mean	SD	Mean	SD
CL	95J	70	10.5	106	43	70	2.1	71	2.1
	96J	70	9.7	105	43	64	1.4	68	1.6
	97J	72	9.6	107	50	65	2.1	72	3.7
	00DS	55	7.7	78	36	59	4.4	49	1.5
	00WS	74	9.3	98	49	75	5.2	68	2.1
	01DS	57	8.3	92	39	66	4.1	53	3.1
	Combined	68	10.9	-	-	-	_	-	_
PnL	95J	23	2.6	33	17	26	1.0	20	1.0
	96J	22	2.5	29	14	26	1.1	17	6.1
	97J	22	2.6	31	15	25	0.8	19	1.3
	00DS	20	2.6	27	15	24	0.9	19	5.9
	00WS	20	2.6	27	14	21	1.4	18	1.5
	01DS	19	2.8	29	14	22	1.8	44	0.7
	Combined	21	2.8	-	-	-	-	-	-
PnN	95J	193	47.2	325	80	208	30.2	206	28.3
	96J	275	68.0	568	62	251	57.4	284	85.1
	97J	208	58.4	397	77	176	28.0	179	13.8
	00DS	374	107.6	698	160	367	77.4	271	38.1
	00WS	302	71.7	649	156	228	40.4	259	36.2
	01DS	350	104.4	729	103	228	27.3	239	22.8
	Combined	282	103.4	-	-	-	_	-	-
TN	00DS	611	225.8	1,353	227	755	84.2	-301	39.2
	00WS	370	109.4	793	170	362	69.0	-273	36.1
	001DS	426	125.1	893	145	379	89.4	-267	33.4
	Combined	466	189.4	-	-	-	_	-	-

<sup>a</sup> SD, max and min mean standard deviation, maximum value and minimum valae, respectively. *Combined* was calculated using all the RILs measured for six seasons. Units are cm in CL and PnL, and the number per square meter in PnN and TN

that CL differed with cropping time, whereas PnL and PnN differed with cropping location.

CL was positively correlated between seasons (r=0.49-0.79). The correlation coefficients were especially high between different years in Joetsu (0.76–0.79) and between different seasons in Los Baños (0.68–0.72), and between 00WS and 95 J, 96 J and 97 J (0.67–0.75). Such a trend was also observed for the three other traits. PnL had higher coefficients (r=0.78-0.61) than CL, whereas PnN and TN had similar or lower coefficients (r=0.78-0.49 and r=0.70-0.48).

CL was positively correlated with PnL (r=0.20-0.61). The correlation coefficients were higher between 00DS and 01DS (0.43–0.61). Positive correlations were also observed between PnN and TN (r=0.36-0.93), whereas negative ones were observed between CL and PnN (r=-0.38 to -0.08) and between PnL and PnN (r=-0.54 to -0.19). These results showed that the two vertical growth components, CL and PnL, were positively correlated especially in the tropical dry season, and that these two were negatively correlated with the two horizontal growth components, PnN and TN.

### QTL mapping

Throughout six crop seasons (three for TN), a total of 31, 59, 38 and 14 QTLs were detected for CL, PnL, PnN and TN, respectively. The total phenotypic variance explained

by these QTLs ranged from 17.0–44.4%, 37.4–52.2%, 24.1–47.2% and 19.7–34.0% in CL, PnL, PnN and TN, respectively. Most QTLs were mapped onto 19 genomic regions (Fig. 1, Table 3). Based on the effects of these QTLs, the 19 regions were characterized and categorized into five groups.

# Group I: QTLs affecting both shoot elongation (CL and PnL) and tillering (PnN and TN)

The M23 allele for R6-2 (G2140C-XNpb12) resulted in increased CL and PnL (shoot length) and decreased PnN and TN (tillering) (Fig. 1). The LOD score peak indicated that the QTL for these four traits was present near XNpb12. This QTL showed high  $r^2$ , suggesting a large genetic effect (Table 3). Comparison of the  $r^2$  between Joetsu and Los Baños showed that the effects of R6-2 on CL and PnN were elevated by the tropical climate. A similar region was also found near the centromere on chromosome 11 (R11, C477-XNpb257). QTLs for CL and PnL were identified near N099K in the Los Baños dry seasons (00DS and 01DS), whereas two different QTLs for PnN were located near C477 and XNpb257 in all crop seasons. These results showed that R11 possibly consists of three loci: one functions for shoot elongation in the tropical dry season (Jan-Apr) and two for tillering regardless of cropping season. The above results demonstrated that R6-2 and R11 affected the balance between



**Fig. 1** Linkage map sowing QTLs for culm length (CL), panicle length (PnL), panicle number (PnN) and tiller number (TN) in the Milyang 23/Akihikari RILs grown during 1995 (95 J), 1996 (96 J) and 1997 (97 J) in Joetsu, Japan, and during the 2000 dry season (00DS), the 2000 wet season (00WS) and the 2001 dry season (01DS) in Los Baños, The Philippines. *Vertical bars* show the genomic region recording LOD >2.0. *Open and filled arrows* present the highest LOD

peak and the neighboring one, respectively. *Upward and downward arrows* indicate that the Milyang 23 and Akihikari alleles increased the value, respectively. The genomic regions that repeatedly detected QTLs for a trait across seasons were shown by *bold vertical bars* and designated tentatively with the letter R, and the chromosome and region number

**Table 3** Location and effect of QTLs for culm length (CL), panicle length (PnL), panicle number (PnN) and tiller number (TN) in the Milyang 23/Akihikari RILs grown during 1995 (95 J), 1996 (96 J)

and 1997 (97 J) in Joetsu, Japan, and during the 2000 dry season (00DS), the 2000 wet season (00WS) and the 2001 dry season (01DS) in Los Baños, The Philippines

Chr.	Re- gion <sup>a</sup>	Trait	Season	Posi- tion (cM) <sup>b</sup>	LOD	a <sup>c</sup>	$\begin{array}{c} R^2 \\ (\%)^d \end{array}$	Chr.	Re- gion <sup>a</sup>	Trait	Season	Posi- tion (cM) <sup>b</sup>	LOD	a <sup>c</sup>	$R^2$ (%) <sup>d</sup>
1 R1 R1	R1-1	CL	00DS 00WS	4	3.0	2.4 2.6	7.6 5.8	4	R4	PnL PnL	00DS 96J	0 145	2.3	0.6 0.7	5.8 5.3
	R1-2	PnN	95J	25 35	4.9 3.8	16.0 15.6	11.2 8.8			1	97J 00DS	143 140	2.0 2.6	0.7 0.8	4.8 6.6
			97J	20 36	5.2 4.9	22.2 21.1	11.8 11.1			PnN	00WS 00WS	141 143	3.4 2.2	0.8 19.4	7.9 5.3
			00DS 01DS	37 24	2.4 5.5	27.9 38.7	6.0 12.4			TN	01DS 00WS	144 140	2.5 3.1	29.7 33.7	5.9 7.3
		TN	00DS	34 34	5.1 2.1	40.2 57.8	11.7 5.2	50		PnL	00WS	157	2.8	0.7	6.5
			00WS	24	3.3	31.9	7.6	55 L	R5	CL PnL	95J	28	5.1 2.4	2.7 0.7	7.6 5.7
			01DS	24	5.3	46.1	12.1	2			97J 00DS	32 32	2.3 2.1	0.6 0.6	5.4 5.2
	R1-3	CL	01DS	55 63	4.4 3.3	44.9 2.4	7.6			DnN	00WS	11	2.1	0.6	5.0
		PnL	95J 97J	47 54	3.1 3.2	0.9 0.8	7.2 7.4	6	R6-1	PnL	90J 95J	24 37	4.0	0.9	9.3 9.8
			00DS 01DS	58 57	2.9 2.2	0.7 0.7	7.2 5.2				96J 97J	38 37	2.6 2.2	0.7 0.7	6.3 5.3
	R1-4	PnN CL	00DS 951	50 123	2.6 4.5	34.1 3.8	6.6 10.4				00DS 00WS	38 38	2.0	0.7	5.1
	R1 1	CL	96J 071	128	6.1	3.7	13.9		D6 0	CI	01DS	38 70	14.3	1.0	9.9
			00DS	120	3.0	4.0 2.4	7.6		K0-2		01DS	69	4.4 5.0	3.4 3.3	10.0
			00WS 01DS	122 126	5.3 7.5	3.6 3.7	12.1 16.5			PnL	95J	55 76	4.4 8.5	1.0 1.3	10.2 18.7
		PnL PnN	01DS 01DS	121 135	2.5 2.1	0.7 27.7	5.8 4.9				96J	55 76	5.9 8.2	1.1 1.3	13.5 18.2
2	R2-1	CL	95J	16	2.5	3.1	5.9				97J	58 77	4.5 8.4	0.9 13	10.3 18.4
	R2-2	PnN	97J 95J	21 30	2.5	2.4 14.2	5.9 6.6				00DS	57 77	4.2	1.0	10.2
			97J 96J	31 33	2.8 3.4	17.2 20.6	6.6 7.9				00WS	56	6.0 8.2	1.1	13.4
		CL	00DS 95J	48 71	3.5 2.4	35.4 2.9	8.8 5.6					08 77	8.2 9.0	1.3 1.4	18.0 19.6
2	<b>D2</b> 2	CL	01DS	106	2.2	2.4	5.1				01DS	56 68	8.0 10.5	1.4 1.6	17.6 22.5
2	K2-3	PnL	96J 01DS	120	2.1 2.3	0.6	5.0 5.4			PnN	96J 97J	78 76	2.1 2.6	$18.0 \\ 17.1$	4.9 6.1
		PnN	95J 96J	120 121	2.4 2.1	11.4 16.0	5.5 5.1				00DS 00WS	76 77	2.8 7.1	33.7 34 1	7.1 15 9
			97J 00WS	120 120	2.2 2.1	13.6 16.4	5.1 4.9			TN	01DS	75 76	5.6	43.4	12.6
		TN	01DS 01DS	120 120	4.2 3.1	33.2 34 7	9.6 7.2				01DS	73	4.2	43.6	19.7
3	R3-1	PnN	95J	1	2.1	11.2	5.0	7		PnN PnN	00DS 97J	86 32	2.1 2.8	26.6 17.1	5.4 6.6
		TN	00DS 00WS	18 2	3.6 2.6	70.8 29.7	8.9 6.1	0		PnL	97J	37	2.5	0.7	5.8
	R3-2	PnL CL	97J 95J	16 23	2.1 2.6	0.6 2.7	4.9 16.2	8	R8	PnL PnL	96J 95J	59 100	2.7 3.4	0.7	6.4 7.9
			01DS	27 42	2.5	2.3	5.9 7.1				96J 97J	90 101	4.0 4.3	1.1 0.9	9.4 9.8
		PnL	97J	21	2.4	0.6	5.6				00DS 00WS	194 100	2.9 5.5	0.9 1.0	7.2 12.5
			00DS	52 53	2.8	0.7	7.1		<b>D</b> 0 4		01DS	97	2.7	0.8	16.3
			00WS	67 67	2.8 2.8	0.8 0.7	7.0 6.5	9	R9-1	PnL	95J 00WS	53 52	3.0 2.5	0.7 0.6	7.0 5.9
			01DS	33 51	2.1 4.2	0.7 1.2	4.9 9.7		R9-2	TN CL	00WS 96J	59 77	2.5 2.7	31.2 2.5	5.9 6.3
	R3-3	CL	95J 96J	107 105	3.3 3.9	3.3 3.5	7.8 19.2	10		CI	97J	77	2.1	2.2	5.1
			97J 00WS	105 109	6.1 4.1	4.2 2.9	13.7 9.5	10		CL PnL	00WS 01DS	24 50	2.0 3.6	2.1 0.9	4.7 8.4

Table 3 (continued)

Chr.	Re- gion <sup>a</sup>	Trait	Season	Posi- tion (cM) <sup>b</sup>	LOD	a <sup>c</sup>	${R^2 \over (\%)^d}$
11	R11	CL	00DS	36	2.1	1.8	5.2
			OIDS	30	3.1	2.3	1.2
		PnL	96J	24	2.1	0.6	5.1
			00DS	34	3.5	0.8	8.8
			01DS	38	3.6	0.8	8.3
		PnN	95J	24	2.4	11.2	5.5
			96J	24	2.1	5.5	5.1
				41	3.7	1.1	8.8
			97J	41	3.7	17.9	38.6
			00DS	40	3.7	34.7	9.1
			00WS	43	2.7	19.5	6.3
			01DS	25	3.4	30.2	7.9
				43	4.5	36.4	10.4
		TN	00DS	37	2.2	54.1	5.5
			01DS	40	4.2	41.6	9.7
12		PnN	00DS	0	3.2	34.7	8.0
	R12	CL	95J	62	2.2	3.9	5.2
			96J	37	2.5	3.1	6.0
			01DS	69	2.1	2.8	4.9
		PnL	97J	32	2.4	0.9	5.7

<sup>a</sup> Chromosomal regions shown in Fig. 1

<sup>b</sup> Distance from the end of the short arm

<sup>c</sup> Additive effect of the Milyang 23 allele compared with the Akihikari one. Units are centimeter in CL and PnL, and number per square meter in PnN and TN

<sup>d</sup>  $\hat{R}^2$  mean the phenotypic variance explained by QTLs

shoot elongation and tillering, especially in the tropics. These two regions well explained the observed positive genetic relations between CL and PnL, or between PnN and TN, and the negative genetic relations of PnN with CL and PnL.

Past QTL analyses examined CL, PnL and PnN individually in different crop seasons or mapping populations, thereby providing limited information on genomic regions affecting both shoot elongation and tillering. Exceptionally, Yan et al. (1999) reported a region that increased PH and decreased PnN near *RZ801* on the long arm of chr. 1. However, the effects of R6-2 and R11 on these traits were identified only in this study.

### Group II: QTLs affecting shoot elongation (CL and PnL)

The M23 allele for R3-2 (*C198-R250*) resulted in increased CL and PnL. In R3-2, QTLs appeared at two different positions, *C198* and *R250*. A similar type of region was found in R12 (*XNpb304–C1069*). Two different LOD score peaks were observed near *XNpb304* and between *XNpb307–C1016*, although the peaks of PnL were <2.0 in LOD (ranging from 1.5 to 2.0 for most seasons). These results indicate that R3-2 and R12 could each have two loci affecting shoot elongation (CL and PnL). These two regions explained the observed positive genetic relations between CL and PnL.

### Group III: QTLs affecting culm elongation (CL) only

A powerful OTL for CL was identified near C86 (R1-4). Using near-isogenic lines, Cho et al. (1994) located the semidwarf gene *sd1*, which reduces PH markedly, near the RFLP marker RG220 on chromosome 1. Since RG220 is close to C86, the powerful CL QTL near C86 could be identical to the *sd1* locus (Fukuta 1995; Tabuchi et al. 2000). Because of the large *sd1* effect, other genes with a minor effect on CL have not been investigated in detail by past genetic analyses, although they play an important role in fine-tuning CL toward the desired PH (Liang et al. 1994; Li et al. 1995). Our 6-season investigation found QTLs with a minor or intermediate effect on CL in four regions, R1-1 (C970), R2-1 (XNpb227), R3-3 (XNpb48) and R9-2 (G1445). QTLs were detected in R1-1 for the Los Baños crop seasons, in R2-1 and R9-2 for the Joetsu crop seasons, and in R3-3 for 95 J, 96 J, 97 J and 00WS. These detection patterns indicated that the QTLs of these four regions interacted with the seasonal environments. Since R6-2 and R11 affected CL more significantly in Los Baños, a total of six CL-affecting regions (QTLs) were expressed differently in temperate and tropical environments.

### Group IV: QTLs affecting panicle development (PnL and PnN)

QTLs for PnL were detected in five regions, R1-3 (R210A), R5 (C734B), R6-1 (C235), R8 (XNpb56) and R9-1 (XNpb13). No seasonal detection pattern was observed for these five regions, suggesting relatively stable expression of PnL QTLs. Generally, the heritability of PnL is higher than that of CL (Sasahara 1997; Tanisaka 1997). The observed stable QTL expression may explain the high PnL heritability. QTLs for PnL and PnN were also detected in R2-3 (XNpb317B) and R4 (XNpb331). The M23 allele of R2-3 resulted in increased PnL and decreased PnN, whereas that of R4 resulted in the opposite effect. In R4, QTLs for PnN were detected only in the tropical location of Los Baños. Previously published literature reported QTLs for panicle traits, such as spikelet fertility and spikelet number per panicle, in the vicinities of R5, R6-1 and R8 (Lin et al. 1996; Lu et al. 1996; Xiao et al. 1996; Ishimaru et al. 2001). However, QTLs for PnL or PnN were not reported previously in R1-3, R9-1, R2-3 and R4, and the effect of temperate climate on R4 was not shown.

### Group V: QTLs affecting tillering (PnN and TN)

It has been difficult to locate QTLs controlling PnN or TN because of the large environmental error (Sasahara 1997). The 6-season investigation repeatedly detected QTLs for PnN or TN in three regions, R1-2 (*R210B-XNpb90*), R2-2 (*XNpb353-XNpb132*) and R3-1 (*C74-XNpb129*). In R1-2, two high LOD peaks were observed near *R210B* and

*XNpb90*, suggesting two loci of large effect were involved. The AK allele of R1-2 resulted in increased PnN and TN. Yagi et al. (2001) reported that the M23 allele of this region resulted in an increased number of spikelets per panicle. R1-2 corresponded to the QTL of Yagi et al. (2001). These allelic effects agree well with the varietal descriptions of AK as having a large number of panicles, and M23 as having a large number of spikelets per panicle (heavy panicle). These past and current results from our research group, for the first time, shed light on R1-2 as a genetic factor that differentiates rice varieties, such as AK and M23, into panicle-number and heavy panicle types. It was also noteworthy that R3-1 showed only the TN QTLs in Los Baños. In other words, R3-1 has a locus (loci) that stimulates tillering only in the tropics.

# Conclusions

The plant architectures of M23 and AK were differentiated by at least 19 genomic regions. The 19 regions were characterized and categorized into five groups: Group I (two regions) promoted shoot elongation and suppressed tillering, Group II (two regions) affected shoot elongation, Group III (five regions) culm elongation, Group IV (seven regions) panicle development and Group V (three regions) tillering. The multiple effects of two regions in Group I were enhanced in a tropical climate. Such a genetic response to the tropical climate also occurred with two regions in Groups IV and V. Moreover, the effects of four regions in Group III were different in a tropical climate, a temperate climate, or the Joetsu and Los Baños wet seasons. Therefore, a total of eight regions interacted with the environments. These findings demonstrate that: (1) more than a dozen genes are involved in the complex genetic control of plant architecture, (2) a few genes control the balance between shoot elongation and tillering, and (3) several genes interact with environments and result in differentiated genetic mechanisms among locations. The actual number of genes in each region, and their accurate locations and effects, are being investigated by our subsequent experiments using the substitution lines in the M23 or AK background.

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