#### ORIGINAL RESEARCH





# Synthesis and evaluation of cyclic diamino benzamide based  $D_3$ receptor ligands

Benjamin E. Bla[s](http://orcid.org/0000-0003-2449-4503)s n<sup>[1](http://orcid.org/0000-0003-2449-4503)</sup> · Peng-Jen Chen<sup>1</sup> · Michelle Taylor<sup>2</sup> · Suzy A. Griffin<sup>2</sup> · John C. Gordon<sup>1</sup> · Robert R. Luedtke<sup>1</sup>

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#### Abstract

Dopamine (1) is a key neurotransmitter whose impact on pharmacological processes is mediated by a family of dopamine receptors designated  $D_1$ ,  $D_2$ ,  $D_3$ ,  $D_4$ , and  $D_5$ . Various diseases and conditions such as schizophrenia, drug abuse, depression, restless leg syndrome, Parkinson's disease (PD), and inflammatory diseases have been linked to aberrant  $D_3$  activity. Herein, we report a series of novel  $D_3$  ligands with improved solubility over our previous lead compound, MC25-41 (2).



Keywords Dopamine  $\cdot$  D<sub>3</sub> dopamine receptor  $\cdot$  D<sub>2</sub> dopamine receptor

## Introduction

The key neurotransmitter known as dopamine (1) was prepared synthetically for the first time in 1910 by George Barger and James Ewens [\[1](#page-14-0)] long before its pharmacological role was recognized. Over 45 years later, Katharine Montagu determined that dopamine was present in the human brain [\[2](#page-14-0)], and in 1958 Arvid Carlsson and Nils-Åke Hillarp demonstrated that this chemical acts as a neurotransmitter [[3\]](#page-14-0). Over the next several decades, the pharmacological function and the means through which dopamine exerts its impact on biological systems has been elucidated. It is known, for example, that dopamine (1) is synthesized in the brain and the periphery, and it has been conclusively linked to a wide range of physiological functions. These functions include vasodilation, modulation of renal sodium excretion, altering urine output, learning, movement, and behavioral motivations [[4\]](#page-14-0).

At the cellular level, dopamine signaling is mediated by a family of G-protein coupled receptors (GPCRs) that are designated as  $D_1$ ,  $D_2$ ,  $D_3$ ,  $D_4$ , and  $D_5$ . The  $D_1$ -like  $(D_1$  and  $D_5$ ) and  $D_2$ -like  $(D_2, D_3$  and  $D_4$ ) sub-families are based on genetic organization, amino acid homology, and pharmacological properties of the individual family members [[5\]](#page-14-0). The  $D_3$  receptor has been the subject of intense interest as a potential therapeutic target, as it has been linked to various

 $\boxtimes$  Benjamin E. Blass [Benjamin.Blass@Temple.edu](mailto:Benjamin.Blass@Temple.edu)

<sup>1</sup> Department of Pharmaceutical Sciences, Temple University School of Pharmacy, 3307 North Broad Street, Philadelphia, PA 194140, USA

Department of Pharmacology and Neuroscience, Center, University of North Texas Health Science, 3500 Camp Bowie Blvd, Fort Worth, TX 76107, USA

disease states and conditions such as schizophrenia, drug abuse, depression, restless leg syndrome [\[6](#page-14-0)], Parkinson's disease (PD) [\[7](#page-14-0), [8](#page-14-0)], and various inflammatory diseases [\[9](#page-14-0)]. We recently described our effort to identify novel, selective  $D_3$  ligands with potential utility for the treatment of cocaine use disorder. These studies led to the identification of MC25-41 (2, Fig. 1) as a potent  $D_3$  ligand that (1) possesses a high level of selectivity for  $D_3$  over other dopamine receptors and a range of other key CNS targets, (2) has a pharmacokinetic profile suitable for in vivo studies [\[10](#page-14-0)], and (3) attenuates motivation for cocaine in Sprague-Dawley rats [[11\]](#page-14-0). While MC25-41 (2) has proven to be an effective tool molecule, its limited solubility  $(2 \mu M)$  may be lead to future problems in formulation and dosing [\[12](#page-14-0)]. As part of an effort to address this issue, we have developed a new series of novel, selective  $D_3$  ligands whose solubility is significantly improved over our original lead compound (2). Specifically, we have examined the impact of inserting an ether moiety in the linker chain. This provides additional opportunities to form hydrogen bonds with water and increases aqueous solubility. In addition, we have explored replacing the piperazine ring with two bioisosteres, homopiperazine and 2,6-diazaspiro[3.3]heptane. These alternative rings systems oriented their embedded nitrogen



Scheme. 1 Synthesis of  $(3a)–(3K)$ 

atoms in manners significantly different from the orientation of the corresponding piperazine nitrogen atoms. which leads to increased basicity and increased capability to form hydrogen bonds with water. Both of these factors led to increased aqueous solubility for the homopiperazine and 2,6-diazaspiro[3.3]heptane analogs.

## Results and discussion

Our effort to improve the solubility of our lead series began with the incorporation of an oxygen atom in the central linker region (3a–3k). This addition adds an additional hydrogen bond acceptor, which could improve the solubility of our compounds, but also increases the length of the linker, which could alter  $D_3$  binding affinity and selectivity. Target compounds were prepared as outlined in Scheme 1. DMT-MM mediated coupling of biaryl acid (4) with aminoalcohol (5) provided the corresponding amide, which was converted to the corresponding bromide (6) with carbon tetrabromide and triphenylphosphine. The bromine was displaced with an aryl piperazine (7) to provide the target compounds (3a–3k).

Table [1](#page-2-0) includes the in vitro binding  $(K_i$  at  $D_3$  and  $D_2$ ) as well as the physicochemical properties (MW, TPSA, LogP) of target compounds (3a–3k). Table [2](#page-2-0) provides a comparison of the solubility of the target compounds  $(3a-3k)$  with the corresponding MC25-41 analogs (8a–8k) The compounds prepared and tested have MW, TPSA, and cLogP values that are consistent with drug like properties, but all examples have either TPSA (3a, 3c) or cLogP (3b, 3d–3k) value outside of the range suggested of BBB penetration  $(TPSA < 90, cLogP = 2-4)$  [\[13](#page-14-0)]. It is noteworthy, however, that we have previously demonstrated that MC25-41 (2) is efficacious in rat models of cocaine addiction (10 mg/kg IP) [\[11](#page-14-0)] and a marmoset model of Parkinson's disease (10 mg/ kg PO) [\[14](#page-15-0)]. This indicates that MC25-41 (2) is able to penetrate the BBB despite the fact that its clogP (4.9) value Fig. 1 Structures of dopamine (1) and MC25-41 (2) are outside of the range suggestive of BBB penetration and



\*DMT-MM =  $(4-(4.6-dimethoxy 3e = 2-CF<sub>3</sub>-Ph,$  $3d = 2-CI-Ph$ . 1,3,5-triazin-2-yl)-4-methyl- $3g = 2,4-Di-Cl-Ph$ ,  $3h = 3,5-Di-Cl-Ph$ ,  $3i = 2-OMe-Ph$ , morpholinium chloride)  $3j = 4$ -OMe-Ph,  $3k = 1-Napthyl$ 

<span id="page-2-0"></span>its and TPSA (87.6) is on the boarderline. It further suggests that related compounds may also be able to penetrate the BBB despite out-of-range values for cLogP and TPSA.

The structure-activity relationship analysis began with the 3-cyano analog (3a), which provided a direct comparison with our previous lead compound MC25-41 (2). A decrease in both  $D_3$  binding affinity  $(K_i = 128 \text{ nM})$  and selectivity over  $D_2$  (13-fold) were observed. Similar  $D_3$ potency was observed when the 3-CN (3a) was replaced with a 3-CF<sub>3</sub> (3b, D<sub>3</sub> K<sub>i</sub> = 175 nM), but selectivity decreased (3.2-fold). Relocation of the cyano substituent to the 2-position  $(3c)$  once again produced a compound with similar  $D_3$  potency (K<sub>i</sub> = 140 nM) and selectivity (3.5-fold). Replacing the 2-CN  $(3c)$  with either a 2-Cl  $(3d)$  or 2-CF<sub>3</sub> (3e) led to an increase in both  $D_3$  potency (K<sub>i</sub> = 27 nM and 37 nM respectively) and selectivity (11-fold and 17-fold respectively versus  $D_2$ ). Addition of a second chlorine atom

Table 1 In vitro screening and physicochemical properties data for  $(3a)–(3k)$ 



Table 2 Comparison of kinetic solubility of  $(3a)$ – $(3k)$  and  $(8a)$ (8k)



in either the 3-position  $(3f)$  or the 4-position  $(3g)$  caused a drop in  $D_3$  potency (K<sub>i</sub> = 127 nM and 1653 nM respectively) and selectivity (3.6-fold and 1.3-fold respectively versus  $D_2$ ).  $D_3$  binding affinity was restored when the 2 chlorine atoms were relocated to the 3- and 5-positions (3h,  $D_3 K_i = 22$  nM), and selectivity over  $D_2$  improved ( $D_2 K_i =$ 332 nM, 14.9-fold). Incorporation of a methoxy group produced mixed results depending on the positioning of the substituent. The 2-methoxy analog  $(3i)$  is a potent  $D_3$  ligand  $(K_i = 24 \text{ nM})$ , but selectivity was marginal (3.1-fold). The 4-methoxy analog (3j), on the other hand, showed little binding affinity for both  $D_3$  (K<sub>i</sub> = 15265 nM) and  $D_2$  (K<sub>i</sub> = 25435 nM). Finally, the 1-napthyl analog  $(3k)$  had moderate  $D_3$  binding affinity (K<sub>i</sub> = 79 nM) and low  $D_2$  selectivity (4fold).

A comparison of the kinetic solubility of the ether-linked compounds (3a–3k) with the corresponding MC25-41 analogs (8a–8k) indicated that this change produced mixed results. A 10-fold increase in solubility, for example, was observed with the 2-OMe analog (3i versus 8i), but when the methoxy substituent was in the 4-position (3j versus 8j), solubility did not increase. Significant increases in solubility over the MC25-41 analogs were also observed for the 3-CN (3a, 29-fold), 2-Cl (3d, 19-fold) and 2,4-di-Cl (3g, 10-fold) analogs, but the impact on solubility was smaller in the  $2-CF_3$  (3e, 6-fold), 2,3-di-Cl (3f, 5-fold), 2-CN (3c, 3-fold), and 3-CF<sub>3</sub> (3b, 2-fold) analogs. Interestingly, no improvement in solubility was observed for the 3,5-di-Cl (3h) and 1-napthyl (3k) analogs, both of which were poorly soluble (sol  $\langle 10 \mu M \rangle$ ).

We next turned our attention to replacing the piperazine ring of our lead compound MC25-41 (2) with either a homopiperazine (9a–9i) or a 2,6-diazaspiro[3.3]heptane (9j, 9k). Both of these moieties have been effectively used as piperazine bioisosteres [\[15](#page-15-0)], but there are differences that could impact  $D_3$  binding and selectivity (Fig. [2](#page-3-0)). While the distance between the nitrogen atoms in a piperazine ring (10) and homopiperazine ring  $(11)$  is similar  $(2.86 \text{ Å}$  versus 2.89 Å), this is not the case for 2,6-diazaspiro[3.3]heptane (12). The distance between nitrogen atoms is substantially



<span id="page-3-0"></span>larger (4.17 Å). In addition, the 3-dimensional shapes of the three rings systems are not the same. The shape of the piperazine ring (10) and homopiperazine (11) are similar in that the first exists in a standard chair confirmation, while the second exists in pseudo-chair conformation. The chair confirmation of the piperazine ring (10) allows for significant cross-ring interaction of the loan pairs of electrons of the two nitrogen atoms, but this interaction is substantially decreased in the homopiperazine ring (11) due to its pseudo-chair conformation. This difference makes the homopiperazine nitrogen atoms more basic and more available to participate in hydrogen bonding interactions that could increase solubility. In a similar but more pronounced manner, the nitrogen atoms of 2,6-diazaspiro[3.3]heptane ring system (12) are oriented very differently from the piperazine ring (10). In this instance, the nitrogen atoms are oriented so that their lone pairs of electrons are perpendicular to each other and therefore incapable of undergoing the interaction seen in the chair configuration of the piperazine ring (10). As a result the 2,6 diazaspiro[3.3]heptane (12) nitrogen atoms are more basic and more available to participate in hydrogen bonding interactions that could increase solubility.

The homopiperazine  $(9a-9i)$  and the 2,6-diazaspiro[3.3] heptane (9j, 9k) analogs were prepared by the methods



Fig. 2 Structures of (10), (11), and (12)

described in Schemes 2 and 3. Buchwald coupling of homopiperazine (11) with an aryl bromide (13) provided the requisite aryl homopiperzines (14a–14i), which were reacted with alkyl bromide (15) under basic conditions to provide target compounds (9a–9g). The synthesis of pyridine analogs (9h) and (9i), on the other hand, began with a displacement reaction of homopiperazine (11) with the corresponding chloropyridine (16a or 16b) to provide aryl homopiperzines (14h) and (14i), This was followed by reaction with alkyl bromide (15) under basic conditions to provide target compounds  $(9i)$  and  $(9k)$ . In a similar manner, the synthesis of 2,6-diazaspiro[3.3]heptane (9j) and (9k) began with the reaction of Boc-2,6-diazaspiro[3.3] heptane (17) with a chloropyridine (16a or 16b) followed by TFA mediated deprotection to provide (18a) and (18b). Reaction with alkyl bromide (15) under basic conditions to provide target compounds (9j) and (9k).

Table [3](#page-4-0) includes the in vitro binding  $(K_i$  at  $D_3$  and  $D_2$ ) as well as the physicochemical properties (MW, TPSA, LogP) of target compounds (9a–9k). Table [4](#page-4-0) provides a comparison of the kinetic solubility of the target compounds (9a– 9k) with the corresponding analogs of MC25-41 (8a–8c, 8e, 8g, 8h, 8k–8o) The majority compounds prepared and tested have MW and TPSA values that are consistent with drug-like properties and suggestive of BBB penetration. The cyano-pyridine analogs (9h) and (9j) are notable exceptions, as their TPSA are 101. cLogP values of the majority of compounds are above the range suggested for orally delivered compounds. Notable exceptions include  $(9h)$ ,  $(9j)$ , and  $(9k)$ , and only one compound,  $(9j)$ , has a cLogP value within the targeted range that suggests BBB



Scheme. 3 Synthesis of (9j) and (9k)

Table 3 In vitro screening and physicochemical properties data for  $(9a)–(9k)$  $\Omega$ 

$$
Ar^{-X}
$$



Homopip homopiperazine

Table 4 Comparison of kinetic solubility of  $(9a)$ – $(9k)$  with the corresponding MC25-41 analogs (8a–8c, 8e, 8g, 8h, 8k–8o)





Homopip homopiperazine

penetration. Overall, replacing the piperazine ring with a homopiperazine ring produced better results than those observed with 2,6-diazaspiro[3.3] heptane with respect to  $D_3$ binding and selectivity over  $D_2$ . Incorporation of a cyano substituents in either the 2-position (9a) or 3-positions (9c) produced compounds that were highly potent  $(K_i = 5.7 \text{ nM})$ and 6.3 nM) and highly selective (139-fold and 114-fold versus  $D_2$ ). Replacing the cyano substituents with a  $CF_3$ group in either the 2-postion (9b) or 3-position (9d) led to a small decrease in  $D_3$  binding potency ( $K_i = 30.2$  nM and 18.6 nM), but selectivity over  $D_2$  was substantially diminished (43-fold and 47-fold). The 2,4-di-chloro (9e) and 3,5 di-chloro (9f) analogs were also less potent  $D_3$  ligands

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A comparison of the solubility of the homopiperazine (9a–9i) and the 2,6-diazaspiro[3.3]heptane based compounds with the corresponding MC25-41 analogs (8a–8c, 8e, 8g, 8h, 8k–8o) (Table 4) demonstrated improvements in

all but one of the homopiperazine analogs (**9f**, sol = 2  $\mu$ M). Solubility improved by a 2- to 13-fold in all other examples. The 2,6-diazaspiro[3.3]heptane analogs (9j and 9k) had the highest aqueous solubility (sol = 148  $\mu$ M and 109  $\mu$ M), but as noted above, these compounds also demonstrated low affinity for  $D_3$  (K<sub>i</sub> = 1093 nM ad 464 nM respectively). The 2-CN homopiperazine analog (9c) is notable as it is highly soluble (sol = 103  $\mu$ M), has high affinity for D<sub>3</sub> (K<sub>i</sub> = 6.3 nM) and is highly selective for  $D_3$  over  $D_2$  (114-fold). Follow-up studies have demonstrated that this compound is highly selective for  $D_3$  over  $D_4$  ( $D_4$  K<sub>i</sub> = 1077 nM). Additional studies on this compound (9c) to assess its affinity for  $D_1$  and  $D_5$ , as well as full in vitro ADME profiling (e.g., Cyp450 inhibition, mouse and human liver microsome stability, permeability) are on-going.

## Conclusion

In summary, we have identified analogs of our initial lead compound MC25-41 (2) that have high affinity for  $D_3$ , are selective for this receptor over  $D_2$ , and whose aqueous solubility is improved over the corresponding MC25-41 analogs  $(8a-8o)$ . In addition, we have identified  $(9c)$  as a potential next-generation lead compound, given its high  $D_3$ affinity, selectivity over  $D_2$  and  $D_4$ , and improved solubility. Future efforts will include follow-up studies on (9c) and close analogs thereof, as well as assessment of our current collection of compounds at the remaining members of the dopamine receptor family  $(D_1, D_4, and D_5)$  and in battery of in vitro ADME screens (e.g., Cyp450 inhibition, mouse and human liver microsome stability, permeability).

## Experimental methods and materials

Reagents were purchased from Fisher Scientific, VWR International, Sigma Aldrich, and Combi-Blocks, Inc. Chromatographic purification of compounds (normal phase and reverse phase) was carried out on a Teledyne Isco Combiflash RF system. H-NMR spectra were obtained on a Bruker 400-MHz NMR. Chemical shift values (δ values) were reported in ppm relative to TMS. For multiplicity,  $s =$ singlet,  $d =$  doublet,  $t =$  triplet,  $m =$  multiplet. Purity  $(\%)$ and mass spectral data were determined with a Waters Agilent 1200 HPLC/MS (Zorbax SB-C18,  $2.1 \times 30$  mm, 3.5 μm, 100% water/0.1% formic acid to 100% acetonitrile/ 0.1% formic acid over 4.0 min, 1.0 mL/min.) with a diode array detector from 210–400 nm and Agilent 6130 quadrupole MS. All compounds were purified to 95% purity or greater as determined by HPLC/MS and 1H-NMR. Melting points were recorded on a capillary melting point apparatus.



Preparation of N-(2-(2-hydroxyethoxy)ethyl)-4-(thiophen-3-yl)benzamide: A solution of 4-(thiophen-3-yl)benzoic acid  $(6 g, 29.27 mmol)$  and  $4-(4, 6$ -dimethoxy-1,3,5triazin-2-yl)-4-methylmorpholinium chloride (8.88 g, 32.09 mmol) in ethanol (200 ml) was stirred at room temperature for 1 h. After 1 h, 2-(2-aminoethoxy)-ethanol (3.66 ml, 32.2 mmol) was added into the reaction. The reaction was further stirred for 2 days. After 2 days, the reaction was filtered to give a filtrate. The filtrate was purified by reverse phase chromatography (50% acetonitrile/0.1% formic acid in water) to provide N-(2-(2-hydroxyethoxy)ethyl)-4-(thiophen-3-yl)benzamide (6.68, 78% yield): <sup>1</sup>H NMR (400 MHz, MeOD-d4)  $\delta$  7.90–7.88 (d, J = 8.52 Hz, 2H),  $7.79-7.77$  (m, 3H),  $7.53$  (d,  $J = 2.16$  Hz, 2H),  $3.72-3.67$  (quint,  $J = 4.84$ ,  $4.68$ ,  $4.28$ ,  $1.92$ ,  $1.68$ ,  $1.4$ , 0.72 Hz, 4H), 3.63–3.59 (quart, 4H); MS (LC/MS, M +  $H^+$ : 292.75.



Preparation of N-(2-(2-bromoethoxy)ethyl)-4-(thiophen-3-yl)benzamide (6): N-(2-(2-hydroxyethoxy)ethyl)-4-(thiophen-3-yl)benzamide (1.67 g, 5.71 mmol) and carbon tetrabromide (2.84 g, 8.58 mmol) were dissolved in dichloromethane. Triphenylphosphine (2.27 g, 8.58 mmol, 1.5 eq) was slowly added into the reaction in an ice bath. The reaction was warmed up to room temperature and was further stirred for 48 h. After 48 h, the reaction mixture was concentrated to provide a solid-oil residue. The resulting material was purified by normal phase chromatography (Ethyl acetate/hexane 0 to 100% gradient) to provide N-(2- (2-bromoethoxy)ethyl)-4-(thiophen-3-yl)benzamide (1.28 g, 63% yield. <sup>1</sup> H NMR (400 MHz, MeOD-d4) δ 7.92–7.90 (d,  $J = 8.43$  Hz, 2H), 7.75–7.71 (m, 3H), 7.56 (d,  $J = 2.26$  Hz, 2H), 3.75 (m, 4H), 3.57 (m, 2H), 3.48 (m, 2H), MS (LC/ MS,  $M + H^{+}$ ): 354.70.



Preparation of N-(2-(2-(4-(2-chlorophenyl)piperazin-1-yl) ethoxy)ethyl)-4-(thiophen-3-yl)benzamide (3d): A solution of N-(2-(2-bromoethoxy)-ethyl)-4-(thiophen-3-yl)benzamide  $(0.14 \text{ g}, \quad 0.3 \text{ mmol})$ , 1-(2-chlorophenyl)piperazine  $(0.06 \text{ g}, \quad 0.3 \text{ mmol})$ 0.3 mmol) and N,N-diisopropylethylamine (0.12 ml, 1.2 mmol, 4.0 eq) in anhydrous acetonitrile (6 ml) at room temperature for 48 h. After 48 h, the reaction was purified by normal phase chromatography (100% dichloromethane to 10% methanol in dichloromethane) to give a partially pure desired product. The impure product was further purified by reverse phase chromatography (10% acetonitrile/0.1% formic acid in water to 40% acetonitrile/0.1% formic acid in water) to provide N-(2- (2-(4-(2-chlorophenyl)piperazin-1-yl)ethoxy)ethyl)-4-(thiophen-3-yl) benzamide  $(38.2 \text{ mg}, 48\% \text{ yield})$ : <sup>1</sup>H NMR  $(400 \text{ MHz}, \text{CDCl}_3)$  δ 8.05 (d, J = 8.36 Hz, 2H), 7.63 (d, J = 8.8 Hz, 2H), 7.47–7.46 (m, 1H), 7.40–7.35 (m, 2H), 7.28–7.25  $(m, 1H)$ , 6.93–6.87  $(m, 2H)$ , 6.74  $(m, 1H)$ , 3.85  $(t, J =$ 4.84 Hz, 2H), 3.73–3.68 (m, 4H), 3.17 (broad s, 4H), 3.13–3.09 (broad s, 6H); MS (LC/MS,  $M + H^{+}$ ): 470.60.



Preparation of N-(2-(2-(4-(3-cyanophenyl)piperazin-1-yl) ethoxy)ethyl)-4-(thiophen-3-yl)benzamide (3a): The title compound was prepared according to the procedure for N-(2- (2-(4-(2-chlorophenyl)piperazin-1-yl)ethoxy)ethyl)-4-(thiophen-3-yl)benzamide, except 3-(piperazin-1-yl)benzonitrile was substituted for 1-(2-chlorophenyl)piperazine. The reaction was purified by normal phase chromatography (100% dichloromethane to 10% methanol in dichloromethane) to give a partially pure desired product. The impure product was further purified by reverse phase chromatography (10% acetonitrile/0.1% formic acid in water to 40% acetonitrile/ 0.1% formic acid in water) to provide N-(2-(2-(4-(3-cyanophenyl)piperazin-1-yl)ethoxy)ethyl)-4-(thiophen-3-yl)benzamide (32% yield): <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.95 (d, J  $= 8$  Hz, 2H), 7.63 (d, J  $= 8$  Hz, 2H), 7.58 (s, 1H), 7.49 (s, 1H), 7.48 (s, 1H), 7.42 (s, 1H), 7.13–6.89 (m, 3H), 6.87 (s, 1H), 3.79 (broad s, 2H), 3.70 (broad s, 4H), 3.22 (broad s, 4H), 2.98 (broad s, 6H); MS (LC/MS,  $M + H^{+}$ ): 461.65.



Preparation of 4-(thiophen-3-yl)-N-(2-(2-(4-(3-(trifluoromethyl)phenyl)piperazin-1-yl)ethoxy) ethyl)benzamide (3b): The title compound was prepared according to the procedure for  $N-(2-(2-(4-(2-chlorophenyl))$ piperazin-1-yl)ethoxy)ethyl)-4-(thiophen-3-yl)benzamide, except 1- (3-(trifluoromethyl)phenyl)-piperazine was substituted for 1-(2-chlorophenyl)piperazine. The reaction was purified by normal phase chromatography (100% dichloromethane to 10% methanol in dichloromethane) to give a partially pure desired product. The impure product was further purified by reverse phase chromatography (10% acetonitrile/0.1% formic acid in water to 40% acetonitrile/ 0.1% formic acid in water) to provide 4-(thiophen-3-yl)- N-(2-(2-(4-(3-(trifluoromethyl)phenyl)piperazin-1-yl) ethoxy)ethyl)benzamide  $(61\% \text{ yield}):$ <sup>1</sup>H NMR  $(400 \text{ MHz}, \text{CDC1}_3)$  δ 7.97 (d, J = 8.28 Hz, 2H), 7.65 (d, J  $= 8.32$  Hz, 2H), 7.49–7.48 (m, 2H), 7.42–7.7.40 (m, 1H), 7.38–7.28 (m, 1H), 7.17 (t,  $J = 8$ , 7.84 Hz, 1H), 7.09–7.05 (m, 2H), 6.85 (d,  $J = 8.08$  Hz, 1H), 3.81 (t,  $J =$ 4.84 Hz, 2H), 3.72 (s, 4H), 3.27 (t,  $J = 4.76$ , 4.36 Hz, 4H), 3.00 (t,  $J = 4.6$  Hz, 4H), 2.95 (t,  $J = 4.84$  Hz, 2H); MS (LC/MS,  $M + H^{+}$ ): 504.60.



Preparation of N-(2-(2-(4-(2-cyanophenyl)piperazin-1-yl)ethoxy)ethyl)-4-(thiophen-3-yl)benzamide (3c): The title compound was prepared according to the procedure for N-(2-(2-(4-(2-chlorophenyl)piperazin-1-yl) ethoxy)ethyl)-4-(thiophen-3-yl)benzamide, except 2- (piperazin-1-yl)benzonitrile was substituted for 1-(2 chlorophenyl)piperazine. The reaction was purified by normal phase chromatography (100% dichloromethane to 10% methanol in dichloromethane) to give a partially pure desired product. The impure product was further purified by reverse phase chromatography (10% acetonitrile/0.1% formic acid in water to 40% acetonitrile/  $0.1\%$  formic acid in water) to provide N- $(2-(2-(4-(2-\epsilon)))$ cyanophenyl)-piperazin-1-yl)ethoxy)ethyl)-4-(thiophen-3-yl)benzamide  $(27\% \text{ yield})$ : <sup>1</sup>H NMR  $(400 \text{ MHz},$ CDCl<sub>3</sub>)  $\delta$  8.08 (s, 1H), 8.07 (d, J = 8.32 Hz, 2H), 7.61 (d,  $J = 8.32$  Hz, 2H), 7.44–7.43 (t,  $J = 2.28$ , 1.64 Hz, 1H), 7.41–7.39 (dd,  $J = 6.16$ , 1.44, 1.36 Hz, 1H), 7.35–7.32  $(m, 2H), 7.00-6.96$  (t,  $J = 6.32$  Hz, 1H), 6.93-6.89 (t, J  $= 7.32$  Hz, 1H), 6.59 (d, J = 8.08 Hz, 1H), 3.91 (t, J = 4.76 Hz, 2H), 3.74 (t,  $J = 4.72$ , 4.56 Hz, 2H), 3.69–3.65 (m, 2H), 3.30 (broad s, 7H), 3.19 (m, 3H); MS (LC/MS,  $M + H^{+}$ ): 461.70.



Preparation of 4-(thiophen-3-yl)-N-(2-(2-(4-(2-(trifluoromethyl)phenyl)piperazin-1-yl)ethoxy) ethyl)benzamide (3e): The title compound was prepared according to the procedure for N-(2-(2-(4-(2-chlorophenyl)piperazin-1 yl)ethoxy)ethyl)-4-(thiophen-3-yl)benzamide, except 1-(2- (trifluoromethyl)phenyl)piperazine was substituted for 1-(2 chlorophenyl)piperazine. The reaction was purified by normal phase chromatography (100% dichloromethane to 10% methanol in dichloromethane) to give a partially pure desired product. The impure product was further purified by reverse phase chromatography (10% acetonitrile/0.1% formic acid in water to 40% acetonitrile/0.1% formic acid in water) to provide 4-(thiophen-3-yl)-N-(2-(2-(4-(2-(trifluoromethyl)phenyl)piperazin-1-yl)ethoxy)ethyl)benza-

mide (57% yield): <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.98 (d, J  $= 8.24$  Hz, 2H), 7.65 (d, J  $= 8.24$  Hz, 2H), 7.61 (broad s, 1H), 7.49 (broad s, 1H), 7.49–7.41 (m, 1H), 7.38–7.37 (m, 1H), 7.15 (t,  $J = 7.96$ , 7.84 Hz, 1H), 7.08 (d,  $J = 7.72$  Hz, 1H), 7.04 (broad s, 1H), 6.83 (d,  $J = 7.88$  Hz, 1H), 3.81 (t, J  $= 4.68$ , 4.6 Hz, 2H), 3.72 (broad s, 4H), 3.25 (t, J = 4.68 Hz, 4H), 3.05 (t, J = 4.44 Hz, 4H), 2.99 (t, J = 4.64, 4.56 Hz, 2H); MS (LC/MS,  $M + H^+$ ): 504.60.



Preparation of N-(2-(2-(4-(2,3-dichlorophenyl) piperazin–1-yl)ethoxy)ethyl)-4-(thiophen-3-yl)benzamide (3f): The title compound was prepared according to the procedure for N-(2-(2-(4-(2-chlorophenyl)piperazin-1-yl) ethoxy)ethyl)-4-(thiophen-3-yl)benzamide, except 1-(2,3 dichlorophenyl)piperazine was substituted for 1-(2-chlorophenyl)piperazine. The reaction was purified by normal phase chromatography (100% dichloromethane to 10% methanol in dichloromethane) to give a partially pure desired product. The impure product was further purified by reverse phase chromatography (10% acetonitrile/0.1% formic acid in water to 40% acetonitrile/0.1% formic acid in water) to provide N-(2-(2-(4-(2,3-dichlorophenyl)-piperazin-1-yl)ethoxy)ethyl)-4-(thiophen-3-yl)benzamide (37% yield): <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.88 (d, J = 8.36 Hz,

2H), 7.66 (d,  $J = 8.44$  Hz, 2H), 7.53–7.52 (m, 1H), 7.44–7.40 (m, 2H), 7.15–7.13 (dd,  $J = 1.4$  Hz, 1H), 7.01 (t,  $J = 8.04$  Hz, 2H), 6.82 (dd,  $J = 6.8$ , 1.32, 1.25 Hz, 1H), 3.71 (broad s, 6H), 3.03 (broad s, 4H), 2.75 (m, 6H); MS (LC/ MS,  $M + H^{+}$ ): 506.10.



Preparation of N-(2-(2-(4-(2,4-dichlorophenyl)piperazin-1-yl)ethoxy)ethyl)-4-(thiophen-3-yl)benz-amide (3g): The title compound was prepared according to the procedure for N-(2-(2-(4-(2-chlorophenyl)piperazin-1-yl)ethoxy)ethyl)-4- (thiophen-3-yl)benzamide, except 1-(2,4-dichlorophenyl) piperazine was substituted for 1-(2-chlorophenyl)piperazine. The reaction was purified by normal phase chromatography (100% dichloromethane to 10% methanol in dichloromethane) to give a partially pure desired product. The impure product was further purified by reverse phase chromatography (10% acetonitrile/0.1% formic acid in water to 40% acetonitrile/0.1% formic acid in water) to provide N-(2-(2-(4-(2,4-dichlorophenyl)piperazin-1-yl) ethoxy)ethyl)-4-(thiophen-3-yl)benzamide  $(45\%$  yield): <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.71 (d, J = 8.2 Hz, 2H), 7.48  $(d, J = 8.28 \text{ Hz}, 2H), 7.35 \text{ (s, 1H)}, 7.28-7.26 \text{ (m, 1H)}, 7.23$  $(m, 1H), 7.19$  (s, 1H), 6.92–6.90 (dd, J = 6.32, 2.32, 2.24 Hz, 1H),  $6.68$  (d, J = 8.64 Hz, 1H), 3.53 (m, 6H), 2.83 (broad s, 4H), 2.54 (broad s, 6H); MS (LC/MS,  $M + H^{+}$ ): 505.50.



Preparation of N-(2-(2-(4-(3,5-dichlorophenyl)piperazin-1-yl)ethoxy)ethyl)-4-(thiophen-3-yl)benzamide (3h): The title compound was prepared according to the procedure for N-(2-(2-(4-(2-chlorophenyl)piperazin-1-yl)ethoxy)ethyl)-4- (thiophen-3-yl)benzamide, except 1-(3,5-dichlorophenyl) piperazine was substituted for 1-(2-chlorophenyl)piperazine. The reaction was purified by normal phase chromatography (100% dichloromethane to 10% methanol in dichloromethane) to give a partially pure desired product. The impure product was further purified by reverse phase chromatography (10% acetonitrile/0.1% formic acid in water to 40% acetonitrile/0.1% formic acid in water) to provide N-(2-(2-(4-(3,5-dichlorophenyl)-piperazin-1-yl) ethoxy)ethyl)-4-(thiophen-3-yl)benzamide (49% yield): <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.95 (d, J = 8.28 Hz, 2H), 7.64  $(d, J = 8.24 \text{ Hz}, 2H), 7.50 \text{ (s, 2H)}, 7.41-7.37 \text{ (m, 2H)}, 6.80 \text{)}$  $(s, 1H)$ , 6.64  $(s, 2H)$ , 3.77  $(t, J = 4.8, 4.72 \text{ Hz}, 2H)$ , 3.70 (broad s, 4H), 3.21 (t, J = 4.8, 4.32 Hz, 4H), 2.97 (t, J = 4.2, 4.68 Hz, 4H), 2.93 (t,  $J = 4.88$ , 4.72 Hz, 2H); MS (LC/MS,  $M + H^{+}$ ): 504.50.



Preparation of N-(2-(2-(4-(2-methoxyphenyl)piperazin-1-yl)ethoxy)ethyl)-4-(thiophen-3-yl)benzamide (3i): The title compound was prepared according to the procedure for N-(2-(2-(4-(2-chlorophenyl)piperazin-1-yl)ethoxy)ethyl)-4- (thiophen-3-yl)benzamide, except 1-(2-methoxyphenyl) piperazine was substituted for 1-(2-chlorophenyl)piperazine. The reaction was purified by normal phase chromatography (100% dichloromethane to 10% methanol in dichloromethane) to give a partially pure desired product. The impure product was further purified by reverse phase chromatography (10% acetonitrile /0.1% formic acid in water to 40% acetonitrile/0.1% formic acid in water) to provide N-(2-(2-(4-(2-methoxyphenyl)-piperazin-1-yl) ethoxy)ethyl)-4-(thiophen-3-yl)benzamide  $(55\% \text{ yield})$ : <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 8.10 (broad s, 1H), 8.00–7.97 (d,  $J = 9.72$  Hz, 2H), 7.59–7.57 (d,  $J = 8.28$  Hz, 2H), 7.40–7.39 (m, 1H), 7.34–7.30 (m, 2H), 6.91–6.86 (m, 1H), 6.72–6.70 (d,  $J = 8.04$  Hz, 1H), 6.58–6.56 (m, 2H), 3.82–3.78 (m, 2H), 3.70 (s, 3H), 3.65–3.61 (m, 4H), 3.15 (broad s, 4H), 3.04 (broad s, 6H); MS (LC/MS,  $M + H^{+}$ ): 466.70.



Preparation of N-(2-(2-(4-(4-methoxyphenyl)piperazin-1-yl)ethoxy)ethyl)-4-(thiophen-3-yl)benzamide (3j): The title compound was prepared according to the procedure for N-(2-(2-(4-(2-chlorophenyl)piperazin-1-yl)ethoxy)ethyl)-4- (thiophen-3-yl)benzamide, except 1-(4-methoxyphenyl) piperazine was substituted for 1-(2-chlorophenyl)piperazine. The reaction was purified by normal phase chromatography (100% dichloromethane to 10% methanol in

dichloromethane) to give a partially pure desired product. The impure product was further purified by reverse phase chromatography (10% acetonitrile/0.1% formic acid in water to 40% acetonitrile/0.1% formic acid in water) to provide N-(2-(2-(4-(4-methoxyphenyl)piperazin-1-yl) ethoxy)ethyl)-4-(thiophen-3-yl)benzamide  $(68\% \text{ yield})$ : <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.87 (d, J = 8.4 Hz, 2H), 7.64  $(d, J = 8.4 \text{ Hz}, 2H)$ , 7.49–7.48 (m, 1H), 7.43–7.41 (m, 1H), 7.39–7.28 (m, 1H), 6.97 (broad s, 1H), 6.83 (d,  $J = 9.2$  Hz, 2H), 6.78 (d, J = 9.16 Hz, 2H), 3.77 (s, 3H), 3.71–3.68 (m, 6H), 3.06 (t, J = 5.12, 4.68 Hz, 4H), 2.71–2.68 (m, 6H); MS (LC/MS,  $M + H^{+}$ ): 466.60.



Preparation of N-(2-(2-(4-(naphthalen-1-yl)piperazin-1 yl)ethoxy)ethyl)-4-(thiophen-3-yl)benzamide (3k): The title compound was prepared according to the procedure for N- (2-(2-(4-(2-chlorophenyl)piperazin-1-yl)ethoxy)ethyl)-4- (thiophen-3-yl)benzamide, except 1-(naphthalen-1-yl) piperazine was substituted for 1-(2-chlorophenyl)piperazine. The reaction was purified by normal phase chromatography (100% dichloromethane to 10% methanol in dichloromethane) to give a partially pure desired product. The impure product was further purified by reverse phase chromatography (10% acetonitrile/0.1% formic acid in water to 40% acetonitrile/0.1% formic acid in water) to provide N-(2-(2-(4-(naphthalen-1-yl)piperazin-1-yl)ethoxy) ethyl)-4-(thiophen-3-yl)benzamide  $(34\%$  yield): <sup>1</sup>H NMR  $(400 \text{ MHz}, \text{ CDCl}_3)$  δ 8.21-8.17 (m, 1H), 7.88 (d, J = 1.6 Hz, 2H), 7.87–7.82 (m, 1H), 7.63 (d,  $J = 8.4$  Hz, 2H), 7.55 (d, J = 8.2 Hz, 1H), 7.48–7.45 (m, 3H), 7.42–7.40 (m, 1H),  $7.37-7.32$  (m, 1H),  $7.30-7.28$  (d,  $J = 8.28$  Hz, 1H), 7.01 (d,  $J = 6.88$  Hz, 1H), 6.96 (broad s, 1H), 3.74 (m, 6H), 3.13 (broad s, 4H), 2.84 (broad s, 3H), 2.78 (t,  $J = 5.52$ , 5.48 Hz, 2H), 2.07 (broad s, 2H); MS (LC/MS,  $M + H^{+}$ ): 487.15.



Preparation of 4-(thiophen-3-yl)-N-(4-(4-(3-(trifluoromethyl)phenyl)-1,4-diazepan-1-yl)butyl) benzamide (9b): A solution of 1-(3-(trifluoromethyl)-phenyl)- 1,4-diazepane (0.11 g, 0.466 mmol), N-(4-bromobutyl)- 4-(thiophen-3-yl)-benzamide (0.16 g, 0.466 mmol) and N,N-diisopropylethylamine (0.25 ml, 1.4 mmol, 3.0 eq) in dry tetrahydrofuran (6 ml) was refluxed at 66 °C for 48 h. After 48 h, the mixture was cooled and filtered to obtain a filtrate. The filtrate was purified by normal phase chromatography (10% methanol in dichloromethane) to provide 4-(thiophen-3-yl)-N-(4-(4-(3-(trifluoromethyl) phenyl)-1,4-diazepan-1-yl)butyl) benzamide (132.8 mg, 56% yield): <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.72 (d, J = 8.36 Hz, 2H), 7.57 (d,  $J = 8.36$  Hz, 2H), 7.45 (t,  $J =$ 2.16 Hz, 1H),  $1.59-1.52$  (m, 4H),  $1.90$  (pent,  $J = 5.16$  Hz, 2H), 2.48 (t,  $J = 7$  Hz, 2H), 2.57 (t,  $J = 5.52$  Hz, 2H), 7.33 (d,  $J = 2.16$  Hz, 2H), 7.19 (t,  $J = 8.16$  Hz, 2H), 6.80 (d,  $J = 7.64$  Hz, 1H), 6.77 (broad s, 1H), 6.72 (dd,  $J =$ 8.4, 2.48 Hz, 2H), 6.47 (broad s, 1H), 3.48 (t,  $J =$ 3.88 Hz, 2H), 3.37–3.43 (m, 4H), 2.72 (t,  $J = 4.16$  Hz, 2H); MS (LC/MS,  $M + H^+$ ): 502.60.



Preparation of N-(4-(4-(3-cyanophenyl)-1,4-diazepan-1 yl)butyl)-4-(thiophen-3-yl)benzamide (9a): The title compound was prepared according to the procedure for 4- (thiophen-3-yl)-N-(4-(4-(3-(trifluoromethyl)phenyl)-1,4-

diazepan-1-yl)butyl)benzamide, except 3-(1,4-diazepan-1 yl)benzonitrile was substituted for 1-(3-(trifluoromethyl) phenyl)-1,4-diazepane. The reaction mixture was first purified by normal phase chromatography (10% methanol in dichloromethane) to give a partially pure desired product. The impure product was further purified by reverse phase chromatography (50% acetonitrile/0.1% formic acid in water) to provide N-(4-(4-(3-cyanophenyl)-1,4-diazepan-1 yl)butyl)-4-(thiophen-3-yl)benzamide  $(48\%$  yield): <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.72–7.7.70 (d,  $J = 8.38$  Hz, 2H),  $7.56-7.54$  (d,  $J = 8.36$  Hz, 2H),  $7.44-7.43$  (t,  $J = 2.24$ , 2 Hz, 1H), 7.32 (d,  $J = 1.88$  Hz, 1H), 7.16–7.12 (m, 1H), 6.81–6.79 (d,  $J = 7.56$  Hz, 1H), 6.76–675 (m, 2H), 6.57–6.54 (t,  $J = 5.32$ , 5.28 Hz, 1H), 3.42–3.38 (m, 2H),  $3.37-3.34$  (m, 4H),  $2.67-2.65$  (t,  $J = 4.92$ ,  $4.88$  Hz, 2H), 2.52–2.49 (t,  $J = 5.56$ , 5.4 Hz, 2H), 2.44–2.41 (t,  $J = 6.96$ ,

6.84 Hz, 2H), 1.96–1.82 (m, 2H). 1.59–1.45 (m, 4H); MS  $(LC/MS, M + H^{+})$ : 459.



Preparation of N-(4-(4-(2-cyanophenyl)-1,4-diazepan-1-yl)butyl)-4-(thiophen-3-yl)benzamide (9c): The title compound was prepared according to the procedure for  $4-(\text{thiophen-3-yl})-N-(4-(4-(3-(\text{trifluoromethyl})phe$ nyl)-1,4-diazepan-1-yl)butyl)benzamide, except 2-(1,4 diazepan-1-yl)benzonitrile was substituted for 1-(3-(trifluoromethyl)-phenyl)-1,4-diazepane to provide N-(4- (4-(2-cyanophenyl)-1,4-diazepan-1-yl)butyl)-4-(thiophen-3-yl)benzamide  $(33\% \text{ yield})$ : <sup>1</sup>H NMR  $(400 \text{ MHz},$ CDCl3) δ 8.02–8.00 (m, 2H), 7.64–7.63 (m, 3H), 7.55–7.45 (m, 3H), 7.38 (broad s, 2H), 6.99–6.96 (m, 2H), 3.85 (broad s, 2H), 3.52–3.42 (m, 8H), 3.20 (broad s, 2H), 2.54 (broad s, 2H), 2.04 (broad s, 2H), 1.76 (broad s, 2H); MS (LC/MS,  $M + H^{+}$ ): 459.



Preparation of 4-(thiophen-3-yl)-N-(4-(4-(2-(trifluoromethyl)phenyl)-1,4-diazepan-1-yl)butyl)-benzamide (9d): The title compound was prepared according to the procedure for 4-(thiophen-3-yl)-N-(4-(4-(3-(trifluoromethyl)phenyl)-1,4-diazepan-1-yl)butyl)benzamide, except 1-(2-(trifluoromethyl)phenyl)-1,4 diazepane was substituted for 1-(3-(trifluoromethyl) phenyl)-1,4-diazepane to provide 4-(thiophen-3-yl)-N- (4-(4-(2-(trifluoromethyl)phenyl)-1,4-diazepan-1-yl) 1 butyl)benzamide (59% yield): H NMR (400 MHz, CDCl<sup>3</sup>) δ 7.91–7.90 (d,  $J = 7.52$  Hz, 3H), 7.60–7.59 (d,  $J = 4.92$  Hz, 3H), 7.52–7.48 (t,  $J = 7.64$ , 6.88 Hz, 2H), 7.35–7.23 (m, 4H), 3.47 (broad s, 2H), 3.40 (broad s, 2H), 3.31 (broad s, 2H), 3.23 (broad s, 2H) 3.06 (broad s, 4H), 3.17-2.13 (m, 2H), 1.85 (broad s, 2H), 1.67 (broad s, 2H); MS (LC/MS, M + H ): 502.



Preparation of N-(4-(4-(2,4-dichlorophenyl)-1,4-diazepan-1-yl)butyl)-4-(thiophen-3-yl) benzamide (9e): The title compound was prepared according to the procedure for 4-  $(thiophen-3-yl)-N-(4-(3-(trifluoromethyl)phenyl)-1,4$ diazepan-1-yl)butyl)benzamide, except 1-(2,4-dichlorophenyl)-1,4-diazepane was substituted for 1-(3-(trifluoromethyl)-phenyl)-1,4-diazepane to provide N-(4-(4- (2,4-dichlorophenyl)-1,4-diazepan-1-yl)butyl)-4-(thiophen-3-yl)benzamide (24% yield):  ${}^{1}H$  NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$ 7.97–7.95 (d,  $J = 7.92$  Hz, 2H), 7.67–7.65 (d,  $J = 7.88$  Hz, 2H),  $7.54-7.53$  (t,  $J = 2$ , 1.96 Hz, 2H),  $7.42$  (t  $J = 1.84$  Hz, 2H),  $7.38-7.37$  (d,  $J = 2.4$  Hz, 1H),  $7.20-7.17$  (dd,  $J = 6.24$ , 2.4, 2.36 Hz, 2H),  $7.02-7.00$  (d,  $J = 8.64$  Hz, 2H), 3.54 (broad s, 2H), 3.43 (broad s, 2H), 3.36 (broad s, 4H), 3.24 (broad s, 2H), 3.09 (broad s, 2H), 2.29 (broad s, 2H), 1.93 (broad s, 2H), 1.75 (broad s, 2H); MS (LC/MS,  $M +$  $H^+$ ): 503.



Preparation of N-(4-(4-(3,5-dichlorophenyl)-1,4-diazepan-1-yl)butyl)-4-(thiophen-3-yl) benzamide (9f): The title compound was prepared according to the procedure for 4-(thiophen-3-yl)-N-(4-(4-(3-(trifluoromethyl) phenyl)-1,4-diazepan-1-yl)butyl)benzamide, except 1- (3,5-dichlorophenyl)-1,4-diazepane was substituted for 1- (3-(trifluoromethyl)-phenyl)-1,4-diazepane to provide N-(4-(4-(3,5-dichlorophenyl)-1,4-diazepan-1-yl)butyl)-4- (thiophen-3-yl)benzamide  $(35\%$  yield): <sup>1</sup>H NMR  $(400 \text{ MHz}, \text{ CDCl}_3)$  δ 7.90–7.89 (d, J = 7.72 Hz, 2H), 7.62–7.60 (d,  $J = 7.84$  Hz, 2H), 7.50 (broad s, 1H), 7.38 (broad s, 2H), 6.70 (broad s, 2H), 6.48 (s, 2H), 3.69 (broad s, 2H), 3.43 (broad s, 4H), 3.12 (broad s, 2H), 3.04 (broad s, 2H), 2.93 (broad s, 2H), 2.28 (broad s, 2H), 1.81 (broad s, 2H), 1.65 (broad s, 2H); MS (LC/MS,  $M +$  $H^+$ ): 503.



Preparation of N-(4-(4-(naphthalen-1-yl)-1,4-diazepan-1 yl)butyl)-4-(thiophen-3-yl)benzamide (9g): The title compound was prepared according to the procedure for 4-(thiophen-3-yl)-N-(4-(4-(3-(trifluoromethyl)phenyl)-1,4 diazepan-1-yl)butyl)benzamide, except 1-(naphthalen-1-yl)- 1,4-diazepane was substituted for 1-(3-(trifluoromethyl) phenyl)-1,4-diazepane. The reaction mixture was first purified by normal phase chromatography (10% methanol in dichloromethane) to give a partially pure desired product. The impure product was further purified by normal amine phase (RediSep Rf Gold® Amine #: 69-2203-507) chromatography (100% isopropanol) to provide N-(4-(4-(naphthalen-1-yl)-1,4-diazepan-1-yl)butyl)-4-(thiophen-3-yl)benzamide (57% yield): <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.77–7.72  $(m, 3H), 7.57$  (d,  $J = 8 Hz, 2H), 7.45-7.43$  (m, 1H), 7.39–7.36 (m, 2H), 7.33 (d,  $J = 1.88$  Hz, 2H), 7.28 (t,  $J =$ 7.64 Hz, 1H), 7.03 (dd,  $J = 6.68$ , 0.72, 0.64 Hz, 1H), 6.82 (broad s, 1H), 3.46 (quart,  $J = 6.16$ , 6.12, 5.68 Hz, 2H), 3.30 (m, 2H), 3.26 (t,  $J = 5.92$  Hz, 2H), 2.91 (m, 4H), 2.63 (m, 2H), 1.98 (pent, 2H), 1.66 (m, 4H); MS (LC/MS, M +  $H^+$ ): 484.65.



Preparation of N-(4-(4-(4-cyanopyridin-2-yl)-1,4-diazepan-1-yl)butyl)-4-(thiophen-3-yl) benzamide (9h): The title compound was prepared according to the procedure for 4-(thiophen-3-yl)-N-(4-(4-(4-(trifluoromethyl)pyridin-2-yl)- 1,4-diazepan-1-yl)butyl)benzamide, except 2-(1,4-diazepan-1-yl)isonicotinonitrile was substituted for 1-(4-(trifluoromethyl)pyridin-2-yl)-1,4-diazepane. The reaction mixture was first purified by normal phase chromatography (10% methanol in dichloromethane) to give a partial pure desired product. The impure product was further purified by normal amine phase (RediSep Rf Gold® Amine #: 69-2203-

507) chromatography (100% isopropanol) to provide N-(4- (4-(4-cyanopyridin-2-yl)-1,4-diazepan-1-yl)butyl)-4-(thiophen-3-yl)benzamide  $(34\% \text{ yield})$ : <sup>1</sup>H NMR  $(400 \text{ MHz},$ CDCl<sub>3</sub>)  $\delta$  8.15 (dd, J = 4.44, 0.56, 0.48 Hz, 1H), 7.78 (d, J  $= 8.36$  Hz, 2H) 7.57 (d, J  $= 8.44$  Hz, 2H), 7.45 (t, J  $=$ 2.16 Hz, 1H), 7.33 (d,  $J = 2.12$  Hz, 2H), 6.77 (broad s, 1H), 6.62 (dd, J = 4, 1.04, 1 Hz, 1H), 6.57 (s, 1H), 3.82 (broad s, 2H), 3.51 (t,  $J = 6.2$  Hz, 2H), 3.41 (quart,  $J = 6.24$ , 5.92 Hz, 2H), 2.84 (t,  $J = 4.28$  Hz, 2H), 2.73 (broad s, 2H), 2.62 (t, J  $= 6.6$  Hz, 2H), 2.06 (broad s, 2H), 1.62 (broad s, 4H), 1.42 (d, J = 6.6 Hz, 2H); MS (LC/MS,  $M + H^{+}$ ): 460.70.



Preparation of 4-(thiophen-3-yl)-N-(4-(4-(4-(trifluoromethyl)pyridin-2-yl)-1,4-diazepan-1-yl) butyl)benzamide (9i): A solution of n-butyllithium, 1.6 M in hexane (0.43 ml, 0.69 mmol) was added dropwise into a solution of 1-  $(4-(\text{trifluorometry})\text{pyridin-2-yl}-1,4-\text{diazepane}$   $(0.17 \text{ g},$ 0.69 mmol) in anhydrous tetrahydrofuran (6 ml) at −78 °C for 1 h. After 1 h, N-(4-bromobutyl)-4-(thiophen-3-yl)benzamide (0.23 g, 0.69 mmol) was added into the reaction and was stirred at 0 °C for another 1 h. The reaction mixture was then refluxed at 66 °C for 48 h. After 48 h, the reaction mixture was cooled and purified by normal phase chromatography (100% ethyl acetate to 10% methanol in dichloromethane) to give a partial pure desired product. The impure product was further purified by reverse phase chromatography (100% acetonitrile with 0.1% formic acid) to provide 4-(thiophen-3-yl)-N-(4-(4- (4-(trifluoromethyl)pyridine-2-yl)-1,4-diazepan-1-yl)butyl)

benzamide (279 mg, 41%): <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 8.19 (d, J = 3.36 Hz, 1H), 7.82 (d, J = 8.32 Hz, 2H), 7.58 (d,  $J = 8.32$  Hz, 2H), 7.46 (t,  $J = 2.44$  Hz, 1H), 7.34 (m, 2H), 7.16 (s, 1H), 6.72 (d,  $J = 5.12$  Hz, 1H), 6.58 (s, 1H), 4.02  $(broad s, 2H), 3.54 (t, J = 6.28 Hz, 2H), 3.44 (t, J = 1 Hz, 2H),$ 3.08 (broad s, 2H), 2.98 (broad s, 2H), 2.86 (t,  $J = 7.4$  Hz, 2H), 2.29–2.25 (m, 2H), 1.79–1.75 (m, 2H), 1.64–1.61 (m, 2H); MS (LC/MS,  $M + H^{+}$ ): 503.60.



1-(3-cyanophenyl)-1,4-diazepane (14a) was prepared as described in WO2018089493 [\[16](#page-15-0)].



Preparation of 1-(naphthalen-1-yl)-1,4-diazepane (14g): 1-Bromonaphthalene (2.89 ml, 20.65 mmol) was stirred with homopiperazine (2.08 g, 20.65 mmol) in the presence of tris(dibenzylideneacetone)dipalladium(0) (0.05 g), sodium tert-butoxide  $(2.77 \text{ g}, 28.92 \text{ mmol}, 1.4 \text{ eq})$ , and  $(\pm)$ -2,2<sup>'</sup>-Bis(diphenylphosphino)-1,1'-binaphthalene

 $(0.05 \text{ g})$  in anhydrous toluene  $(100 \text{ ml})$ . The reaction mixture was refluxed at 110 °C for 48 h. After 48 h, the reaction mixture was cooled and filtered with a celite pad to obtain a filtrate. The filtrate was purified by normal phase chromatography (20% methanol in ethyl acetate) to provide 1- (naphthalen-1-yl)-1,4-diazepane  $(2.91 \text{ g}, 62\% \text{ yield})$ : <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  8.23–8.25 (d, J = 8 Hz, 1H), 7.88–7.85 (d, J = 7.76 Hz, 1H), 7.66–7.64 (d, J = 8.2 Hz, 1H),  $7.58-7.50$  (m, 2H),  $7.45-7.41$  (t,  $J = 8.08$ ,  $7.52$  Hz, 1H),  $7.24 - 7.22$  (dd,  $J = 6.56$ , 0.88, 0.84 Hz, 1H), 3.68–3.64  $(m, 4H), 3.59-3.57$   $(m, 2H), 3.41-3.38$   $(t, J = 6, 5.92$  Hz, 2H), 2..44–2.39 (quint,  $J = 5.88$ , 5.72, 5.68 Hz, 2H); MS  $(LC/MS, M + H^{+})$ : 227.90.



Preparation of 1-(3-(trifluoromethyl)phenyl)-1,4-diazepane (14b): The title compound was prepared according to the procedure for 1-(naphthalen-1-yl)-1,4-diazepane, except 1 bromo-3-(trifluoromethyl)benzene was substituted for 1-bromonaphthalene. The reaction was purified by normal phase chromatography (20% methanol in ethyl acetate) to provide 1-(3-(trifluoromethyl)phenyl)-1,4-diazepane (53% yield): <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.17 (td, J = 7.96, 0.68, 0.64, 0.44 Hz, 1H), 6.75 (m, 3H), 3.47 (t,  $J = 6.12$  Hz, 2H), 3.43 (t, J = 5.48 Hz, 2H), 2.91 (t, J = 5.44 Hz, 2H), 2.71 (t, J  $= 5.68$  Hz, 2H), 1.78 (m, 3H); MS (LC/MS, M + H<sup>+</sup>): 245.85.



Preparation of 2-(1,4-diazepan-1-yl)benzonitrile (14c): The title compound was prepared according to the procedure for 1- (naphthalen-1-yl)-1,4-diazepane, except 2-bromobenzonitrile

was substituted for 1-bromonaphthalene. The reaction was purified by normal phase chromatography (10% methanol in dichloromethane) to provide 2-(1,4-diazepan-1-yl)benzonitrile (47% yield): <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.49 (dd, J = 6.08, 1.68 Hz, 1H), 7.37 (tdtd,  $J = 7.04$ , 1.76, 1.6 Hz, 1H), 6.90 (d, J  $= 8.6$  Hz, 1H), 6.78 (tdd, J = 7.28, 0.88 Hz, 1H), 3.66 (m, 4H), 3.15 (m, 2H), 2.99 (t,  $J = 5.64$  Hz, 2H), 2.01 (quint 2H); MS (LC/MS,  $M + H^{+}$ ): 202.90



Preparation of 1-(2-(trifluoromethyl)phenyl)-1,4-diazepane (14d): The title compound was prepared according to the procedure for 1-(naphthalen-1-yl)-1,4-diazepane, except 1-bromo-2-(trifluoromethyl)benzene was substituted for 1-bromonaphthalene. The reaction was purified by normal phase chromatography (20% methanol in ethyl acetate) to provide 1-(2-(trifluoromethyl)phenyl)-1,4-diazepane (61% yield): <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.66 (broad s, 1H), 7.55–7.53 (d, J = 6.96 Hz, 1H), 7.50–7.46 (t, J = 7.76, 7.6 Hz, 1H), 7.33–7.31 (d,  $J = 7.96$  Hz, 1H), 7.22–7.18 (t, J  $= 7.64$  Hz, 1H), 3.46–3.43 (t, J  $= 5.68$ , 5.56 Hz, 2H), 3.39–3.35 (m, 2H), 3.30–3.27 (m, 2H), 3.15–3.12 (t,  $J =$ 6.36 Hz, 2H), 2.21–2.15 (quint,  $J = 6.32$ , 6.24, 5.72, 5.64 Hz, 2H); MS (LC/MS,  $M + H^{+}$ ): 246.10



Preparation of 1-(2,4-dichlorophenyl)-1,4-diazepane (14e): The title compound was prepared according to the procedure for 1-(naphthalen-1-yl)-1,4-diazepane, except 1 bromo-2,4-dichlorobenzene was substituted for 1-bromonaphthalene. The reaction was purified by normal phase chromatography (20% methanol in ethyl acetate) to provide 1-(2,4-dichlorophenyl)-1,4-diazepane (38% yield): <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.33 (broad s, 1H), 7.14–7.11 (m, 1H), 7.02–7.00 (d,  $J = 7.8$  Hz, 1H),  $3.27 - 3.22$  (quart,  $J = 6.44$ , 6 Hz, 4H),  $3.09 - 3.05$  (quart,  $J =$ 6.24, 5.56 Hz, 4H),  $1.98-1.92$  (quint,  $J = 6$ , 5.84, 5.76 Hz, 2H); MS (LC/MS,  $M + H^{+}$ ): 245.90.



Preparation of 1-(3,5-dichlorophenyl)-1,4-diazepane (14f): The title compound was prepared according to the procedure for 1-(naphthalen-1-yl)-1,4-diazepane, except 1-bromo-3,5 dichlorobenzene was substituted for 1-bromonaphthalene. The reaction was purified by normal phase chromatography (20% methanol in ethyl acetate) to provide 1-(3,5-dichlorophenyl)- 1,4-diazepane (35% yield): <sup>1</sup> H NMR (400 MHz, MeOD-d4) δ 6.68–6.66 (d, J = 6.92 Hz, 3H), 3.75 (broad s, 2H), 3.55–3.52  $(t, J = 5.92 \text{ Hz}, 2H), 3.33 \text{ (borad s, 2H)}, 3.22 \text{ (broad s, 2H)},$ 2.21 (broad s, 2H); MS (LC/MS,  $M + H^{+}$ ): 245.90



Preparation of 1-(4-(trifluoromethyl)pyridin–2-yl)-1,4 diazepane  $(14i)$ : A solution of homopiperazine  $(0.1 g,$ 0.998 mmol, 1.1 eq) and 2-chloro-4-(trifluoro-methyl)pyridine (0.23 ml, 0.908 mmol) in 1-butanol was refluxed at 117 °C for 16 h. After 16 h, the reaction mixture was cooled and basified and was stirred with concentrated sodium hydroxide (20 ml) overnight. After that, the mixture was further purified by solid loading in normal phase chromatography (10% methanol with 1% ammonia hydroxide in dichloromethane) to give 1-(4-(trifluoromethyl)pyridin-2 yl)-1,4-diazepane (169 mg, 69%): <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 8.18 (d, J = 5.16 Hz, 1H), 6.63 (d, J = 5.12 Hz, 1H), 6.57 (s, 1H), 3.76 (t,  $J = 6.28$  Hz, 2H), 3.66 (t,  $J =$ 6.16 Hz, 2H), 3.45 (s, 3H), 3.01 (t, 8 Hz, 2H), 2.84 (t,  $J =$ 5.6 Hz, 2H), 1.92 (pent,  $J = 6.12$ , 5.88, 5.84 Hz, 2H); MS  $(LCMS, M + H^{+})$ : 245.85.



Preparation of 2-(1,4-diazepan-1-yl)isonicotinonitrile (14h): The title compound was prepared according to the procedure for 1-(4-(trifluoromethyl)pyridin-2-yl)-1,4-diazepane, except 2-bromoisonicotinonitrile was substituted for 2-chloro-4-(trifluoromethyl)pyridine. The mixture was purified by normal phase chromatography (10% methanol with 1% ammonia hydroxide in dichloromethane) to provide 2- (1,4-diazepan-1-yl)isonicotinonitrile (72% yield): <sup>1</sup>H NMR  $(400 \text{ MHz}, \text{ MeOD-}d4)$  δ 6.99 (d, J = 4 Hz, 1H), 5.78 (s, 1H), 5.58 (dd,  $J = 4.08$ , 0.96 Hz, 1H), 2.76 (t,  $J = 5.4$  Hz, 2H), 2.47 (t, J = 6.16 Hz, 2H) 2.09-1.98 (m, 4H), 0.91 (pent,  $J = 5.68$  Hz, 2H); MS (LC/MS,  $M + H^{+}$ ): 203.



Preparation of 2-(4-(trifluoromethyl)pyridin-2-yl)-2,6 diazaspiro[3.3]heptane (18b): A solution of 2-chloro-4- (trifluoromethyl)pyridine (0.08 g. 0.411 mmol), tertbutyl 2,6-diazaspiro[3.3]heptane-2-carboxylate (0.1 g, 0.411 mmol) and triethylamine (0.1 ml) in 1-butanol (6 ml) was refluxed at  $117^{\circ}$ C for 16 h. After 16 h, the reaction was cooled and purified by normal phase chromatography (10% methanol in dichloromethane) to provide an intermediate, tert-butyl 6-(4-(trifluoromethyl) pyridin-2-yl)-2,6-diazaspiro[3.3]heptane-2-carboxylate: MS (LC/MS,  $M + H^{+}$ ): 344.10. The intermediate was then dissolved in trifluoroacetic acid and stirred for 16 h. After 16 h, the reaction was concentrated to give an oil residue. The residue was dissolved in methanol (20 ml) and stirred with Amberlite™ IRN-78 ion-exchange resin, OH-form at room temperature for 16 h. The basified reaction was then filtered to remove the resin concentrated to give a product, 2-(4-(trifluoromethyl)pyridin-2-yl)-2,6-diazaspiro[3.3]heptane, used for the next step without chromatographic purification. (94.5 mg, 77% yield): <sup>1</sup>H NMR (400 MHz, MeOD) δ 8.18 (d, J = 5.16 Hz, 1H), 6.63 (d,  $J = 5.12$  Hz, 1H), 6.57 (s, 1H), 3.62 (s, 4H), 3.35 (s, 4H). (LC/MS,  $M + H^{+}$ ): 244.



Preparation of 2-(2,6-diazaspiro[3.3]heptan-2-yl)isonicotinonitrile (18a): The title compound was prepared according to the procedure for 2-(4-(trifluoromethyl)pyridin-2 yl)-2,6-diazaspiro[3.3]heptane, except 2-chloroisonicotinonitrile was substituted for 2-chloro-4-(trifluoromethyl)pyridine. tert-Butyl 6-(4-cyanopyridin-2-yl)-2,6-diazaspiro[3.3]heptane-2 carboxylate: MS (LC/MS,  $M + H^+$ ): 302.10. The intermediate was then deprotected with trifluoroacetic acid and basified with Amberlite™ IRN-78 ion-exchange resin to give 2-(2,6-diazaspiro[3.3]heptan-2-yl)isonicotinonitrile (68% yield). <sup>1</sup>H NMR  $(400 \text{ MHz}, \text{ MeOD})$  δ 8.12 (d, J = 5.04 Hz, 1H), 6.78 (d, J = 5.18 Hz, 1H), 6.49 (s, 1H), 3.72 (s, 4H), 3.41 (s, 4H), (LC/MS,  $M + H^{+}$ ): 201.



Preparation of N-(4-(6-(4-cyanopyridin-2-yl)-2,6-diazaspiro[3.3]heptan-2-yl)butyl)-4-(thiophen-3-yl)benzamide (9j): The title compound was prepared according to the procedure for 4-(thiophen-3-yl)-N-(4-(6-(4-(trifluoro-methyl)pyridin-2 yl)-2,6-diazaspiro[3.3]heptan-2-yl)butyl) benzamide, except 2- (2,6-diazaspiro[3.3]heptan-2-yl)isonicotinonitrile was substituted for 2-(4-(trifluoro-methyl)pyridin-2-yl)-2,6-diazaspiro [3.3] heptane. The reaction mixture was first purified by normal phase chromatography (10% methanol in dichloromethane) and reverse phase chromatography (10% acetonitrile/0.1% formic acid in water) to provide N-(4-(6- (4-cyanopyridin-2-yl)-2,6-diazaspiro[3.3]heptan-2-yl)butyl)-4- (thiophen-3-yl)benzamide  $(47\%$  yield): <sup>1</sup>H NMR  $(400$  MHz, CDCl<sub>3</sub>)  $\delta$  8.16-8.15 (dd, J = 4.48, 0.6 Hz, 1H), 7.81-7.79 (d, J  $= 8.32$  Hz, 2H), 7.59-7.57 (d, J  $= 8.32$  Hz, 2H), 7.47-7.46 (t,  $J = 2.12$  Hz, 1H), 7.34 (d,  $J = 2.16$  Hz, 2H), 7.09 (broad s, 1H),  $6.72-6.71$  (dd,  $J = 3.96$ , 1.2, 1.16 Hz, 1H), 6.37 (s, 1H), 4.12 (s, 4H), 3.96 (s, 4H), 3.40 (broad s, 2H), 2..93 (broad s, 2H), 1.62 (broad s, 4H); MS (LC/MS,  $M + H^{+}$ ): 458.



Preparation of 4-(thiophen-3-yl)-N-(4-(6-(4-(trifluoromethyl)pyridin-2-yl)-2,6-diaza-spiro[3.3] heptan-2-yl) butyl)benzamide (9k): A solution of 2-(4-(trifluoromethyl) pyridin-2-yl)-2,6-diazaspiro[3.3]heptane (0.1 g, 0.411 mmol), N-(4-bromobutyl)-4-(thiophen-3-yl)benzamide (0.14 g, 0.411 mmol) and triethylamine (0.3 ml, 2.12 mmol) in tetrahydrofuran (6 ml) was refluxed at 66 °C for 48 h. After 48 h, the reaction was cooled and purified by normal phase chromatography (10% methanol in dichloromethane) and reverse phase chromatography (40% acetonitrile/0.1% formic acid in water) to provide 4-(thiophen-

<span id="page-14-0"></span>3-yl)-N-(4-(6-(4-(trifluoromethyl)pyridin-2-yl)-2,6-diazaspiro[3.3]heptan-2-yl)butyl)benzamide (89.1 mg, 43% yield): <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  8.25 (d, J = 5.16 Hz,  $J = 1H$ ), 7.88 (d,  $J = 7.92$  Hz, 2H), 7.64 (d,  $J = 8$  Hz, 2H), 7.53 (s, 1H), 7.40-7.37 (s, 2H), 6.82-6.81 (d,  $J = 5.08$  Hz, 1H), 6.44 (s, 1H), 4.21 (broad s, 4H), 4.07 (broad s, 4H), 3.47 (broad s, 2H), 3.03 (broad s, 2H), 1.68 (broad s, 4H); MS (LC/MS,  $M + H^{+}$ ): 501.

Computational values: TPSA and cLogP values were calculated using the Dotmatics software suite (Dotmatics LLC The Old Monastery, Windhill Bishops, Stortford Herts, CW23 2ND UK).

### Competitive radioligand-binding studies

For competitive binding studies, transfected HEK293 cell homogenates were suspended in homogenization buffer and incubated with radioligand [125I]IABN, in the presence or absence of inhibitor at 37 °C for 60 min with  $\int_1^{125}$  [IABN (total volume  $= 150$  ul) as previously described [\[17\]](#page-15-0). Competitive radioligand studies were performed to determine the concentration of inhibitor that inhibits 50% of the specific binding of the radioligand  $(IC_{50}$  value). The final radioligand concentration was approximately equal to the  $K_d$  value for the binding of the radioligand. For each competition curve, triplicates were performed using two concentrations of inhibitor per decade over five orders of magnitude. Binding was terminated by the addition of cold wash buffer (10 mM Tris– HCl/150 mM NaCl,  $pH = 7.5$ ) and filtration over a glass-fiber filter (Pall A/B filters, #66198). A Packard Cobra Gamma Counter was used to measure the radioactivity of  $[^{125}$ I]IABN.

The competition curves were modeled for a single binding site using

## $Bs = Bo - ((Bo + L)/(IC50 + L))$

where Bs is the amount of ligand bound to receptor and Bo is the amount of ligand bound to receptor in the absence of competitive inhibitor. L is the concentration of the competitive inhibitor. The  $IC_{50}$  value is the concentration of competitive inhibitor that inhibits 50% of the total specific binding.  $IC_{50}$  values were determined using nonlinear regression analysis with Table Curve 2D v 5.01 (Jandel, SYSTAT, Systat Software, Inc., San Jose, CA, USA). The values for Bs and Bo were constrained using experimentally derived values. The IC50 values were converted to equilibrium dissociation constants  $(K_i)$  using the Cheng and Prusoff (1973) correction. Mean  $K_i$  values are reported for at least three independent experiments.

#### Aqueous solubility assay

Compounds were assessed for their solubility at pH 7.4 using the commercially available Millipore MultiScreenTM Solubility filter system (Millipore, Billerica, MA). Analysis was performed by liquid chromatography tandem mass spectrometry (LC/MS/MS).

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#### Compliance with ethical standards

Conflict of interest The authors declare no competing interests.

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