ORIGINAL RESEARCH



A DFT and QSAR study of the role of hydroxyl group, charge and unpaired-electron distribution in anthocyanidin radical stabilization and antioxidant activity

Hussein M. Ali¹ · Isra H. Ali²

Received: 11 March 2017 / Accepted: 19 June 2017 / Published online: 7 July 2017 © Springer Science+Business Media, LLC 2017

Abstract Density function theory study was conducted to examine the structure, planarity, and charge and unpaired electron delocalization of some anthocyanidins and monohydroxy flavylium ions as well as all their possible phenoxyl radicals. Bond dissociation energy of O-H bond was in the order of 3-<5-<4'-<7-OH bond except for in malvidin where 4'-OH has less bond dissociation energy than 5-OH bond because of the extra stability provided by the two o-OMe groups. Stabilization by both charge and unpaired electron resonances were suggested for each radical. The present work revealed that the 3-anthocyanidin radicals were stabilized by three types of resonance through 5-, 7- or 4'-oxygen atoms while 5-, 7- or 4'-radicals were stabilized only by resonance through pyrylium or 3-oxygen atom. Positive charge was found delocalized on carbons 2, 5, 7, 9, and 4' by resonance and moves towards the radical oxygen upon radical formation. In all radicals, radical oxygen carries the highest spin density and least negative charge therefore, its charge, spin density and the least bond dissociation energy (O-H) in each anthocyanidin were well

Electronic supplementary material The online version of this article (doi:10.1007/s00044-017-1964-0) contains supplementary material, which is available to authorized users.

Results were partially presented in Food Quality-2016 Conference, Dubai, 5-6 Dec 2016.

Hussein M. Ali Hussein_galaleldeen@agr.asu.edu.eg

- ¹ Agricultural Biochemistry Department, Faculty of Agriculture, Ain Shams University, Shoubra El-Kheima, Cairo, Egypt
- ² Nanomaterials Lab, Center of Material Science (CMS), Zewail City of Science and Technology, Giza 12588, Egypt

quantitatively correlated with the experimental radical scavenging activity.

Keywords Anthocyanidins · Anthocyanins · Antioxidants · Radical delocalization · BDE · Resonance stabilization

Introduction

Anthocyanins are an important class of flavonoids; they are glycosides of anthocyanidins, polyphenols of 2phenylbenzopyrylium ion (C6-C3-C6). Anthocyanins provide the nutritive value to many functional food and exhibit strong antioxidant activity (Azevedo et al. 2010), where some of them are more potent antioxidants than vitamins E (Kong et al. 2003) and C (Bagchi et al. 1998), which could explain their high biological activities and protection effects against many chronic diseases (Bagchi et al. 1998; Kong et al. 2003; Ma et al. 2016). The high activity of anthocyanins was attributed to their cationic structure and low oxidation potential compared to other classes of flavonoids (van Acker et al. 1996). In addition, they give many flowers, fruits and vegetables their characteristic purple (e.g., cauliflower, grapes, and corn) and red (e.g., cabbage, beet, and sorghum) colors and responsible for most colors of berry family fruits.

Hundreds of natural anthocyanins have been reported (Andersen and Jordheim 2006); all of them share in having hydroxyl groups in positions 5, 7, and 4' while most of them have additional hydroxyl group in position 3. Other hydroxyl groups may also occur in positions 6, 3', and 5'. Hydroxyl groups can be free, methylated or glycosylated. The 4'-OH group is not methylated in all natural anthocyanins; interestingly, its methylation (Rahman et al. 2006) or glycosylation (Zhao et al. 2014) reduces the antioxidant activity. Glycosylation can be found in positions 3, 5, 7, 3',

and 5' but more frequently in positions 3 and 5 (Kähkönen and Heinonen 2003). Therefore, anthocyanidins differ mainly in the number and position of the hydroxyl and methoxy groups on ring B and the presence of 3-hydroxyl group on ring C; furthermore, anthocyanins vary in the number, type and site of glycosylation (Zhao et al. 2014). Anthocyanidin structure is also characterized by the aromaticity of the three rings and by the presence of an electron deficiency with a delocalized positive charge, which distinguish anthocyanidins from other classes of flavonoids. The structures of the anthocyanidins examined in the present work are presented in Fig. 1.

The antioxidant activity of simple phenols and flavonoids is increased by the presence of a catechol moiety and electron donating groups e.g., hydroxyl and methoxy groups (Kähkönen and Heinonen 2003; Ali et al. 2013). The 3-hydroxyl group also enhances the antioxidant activity of anthocyanins and other flavonoids (Amić et al. 2007; Ali et al. 2016). However, anthocyanidins either lacking 3-free or glycocylated hydroxyl group (3-deoxyanthocyanidins) e.g., luteolinidin or missing both a 3-OH group and a catechol structure e.g., apigeninidin still exhibit good antioxidant activity (Boveris et al. 2001; Carbonneau et al. 2014).

Computational methods have frequently been used to predict and explain the antioxidant activity of anthocyanidins and other flavonoids (Pop et al. 2012; Lu et al. 2014). According to the hydrogen atom transfer mechanism, the antioxidant activity of anthocyanidins in aprotic solvents depends mainly on the stability of the resulting phenoxyl radicals and the spread of the unpaired electron over the entire molecule (Leopoldini et al. 2011; Foti 2012). The resonance stabilization of 3-, 5-, 7- and 4'-phenoxyl radicals of anthocyanidins has been suggested (Ali et al. 2016); however, the effects of detailed structural features of anthocyanins on their radical stabilization and potent

Fig. 1 Resonance, charge distribution (positive in *green* and negative in *red*) and selected bond lengths (*black*); atoms carrying the highest positive charge and involved in charge delocatization are marked by a *filled circle* in Pn model (color figure online)

antioxidant activity have not been fully studied; besides, there are still some variations in the literature in the order of $BDE_{(O-H)}$ of anthocyanidins (Pop et al. 2012; Lu et al. 2014; Foti 2012) and absence of quantitative correlations between the anthocyanidin antioxidant activity and their BDE(O-H) and charge and unpaired electron distribution. Since this information may aid in explaining the observed high antioxidant activity of anthocyanidins and the biological activity of many functional food as well as predicting the antioxidant activity of new compounds; the present study involves a density function theory (DFT) calculation of some anthocyanidin structures and the possible resonance pathways caused by the delocalization of both the charge and unpaired electron. In addition, the experimental radical scavenging activity of anthocyanidins is quantitatively correlated with their charge and unpaired election distribution as well as their least O-H bond dissociation free energy. The studied compounds were 3-, 5-, 7- and 4'hydroxyflavylium ions and four anthocynidins bearing no catechol structure (apigeninidin (Ap), pelargonidin (Pg), peonidin (Pn), and malvidin (Mv)) along with all their possible phenoxyl radicals. Anthocyanidins with a catechol structure were excluded from this study to avoid the extra unpaired electron delocalization resulted from the formation of semiquinone radicals (Castaňeda-Ovando et al. 2009; Ali and Ali 2015).

Computation methods

DFT calculations

All calculations were performed using Gaussian 09 program package at the level of DFT/B3LYP with a basis set 6-31-G (d,p) while CambridgeSoft Chem 3D package of



ChemOffice 12 was used for model visualization and finding the bond lengths and the dihedral angles between the benzopyrylium ring system and each of the 2-phenyl plane (ϕ O1-C2-C1'-C2') and the 3-O-H bond (ϕ C2-C3-O3-H3). Radicals were generated by abstracting a hydrogen atom from the desired hydroxyl group. Calculations of parent compounds were executed at restricted closed shell level while calculations of radicals were accomplished at unrestricted open shell level. Harmonic vibrational frequencies were computed at the same level of theory to characterize saddle points and find the zero-point energies (ZPE) for thermochemical correction of electronic thermal enthalpies of all species. Calculations were performed in the gas-phase at temperature 298.15 K and pressure 1 atmosphere. For all compounds and radicals, energy was minimized and the lowest energy structure was used in all calculations without any geometry or symmetry constraints. Charge of each atom was calculated by natural population analysis with the sum of charges on all atoms is 1.000 (net charge). Spin density was calculated by frequency calculation; spin densities of hydrogen atoms are summed into those of heavy atoms. Sum of spin densities of all atoms is one. Positive charges on carbon atoms and negative or positive charges on oxygen atoms as well as spin densities were placed on the models.

Bond dissociation energy (BDE_{O-H}) of all hydroxyl groups in molecules was calculated by subtracting the electronic thermal enthalpy of the parent compound (H_p) from that of the resulted radical (H_R) and hydrogen atom

 $(H_{\rm H})$ according to the following equation:

$$BDE_{O-H} = [(H_{R} + H_{H}) - H_{P}]$$

×Correction factor × Conversion Factor

Where correction factor suggested by the program manual to eliminate systematic error in thermal energies of frequency calculations is 0.9804 and the conversion factor from Hartree to Kcal/mol unit is 627.51 whereas hydrogen atom enthalpy was -0.4979 Hartree.

Statistical analysis

Regression analyses were performed using SPSS software version 16. The validity of models was assessed by Pearson correlation coefficient (R), standard error of the estimate (SE), the number of data point (N), the least significant difference (p), and the 95% confidence intervals (in parentheses) for each regression coefficient.

Results and discussion

Structure and planarity

The dihedral angle (O1, C2, C1' and C2') of the examined flavyliums (Fv) and anthocyanidins as well as their phenoxyl radicals is listed in Table 1. The angle determines the

 ΔS^{b} ΔS^{b} Flavylium and BDE φ O1-C2φ C2- ΔG^a Flavylium and BDE φ O1φ C2- ΔG^a anthocyanidin C1 - C2' C3-O3nthocyanidin C2-C1-C3-O3-(O-H) (O-H) Η C2'Η 3-OHFv^c 18.5 4[\]-RPg 22.3 30.2 90.128 21.1 81.762 28.060 3-RFv^c 83.484 0.0 74.933 28.680 5-RPg 82.500 26.7 19.7 74.502 26.825 _ 4[\]-OHFv 0.0 23.5 82.561 27.858 7-RPg 90.867 19.1 _ $4^{-}RFv$ 91.851 0.0 83.423 28.268 Peonidin Pn 24.6 273 5-OHFv 0.0 3-RPn 74.748 0.0 66.504 27.651 _ _ 73.887 26.205 4[\]-RPn 5-RFv 81.700 -0.1_ 87.483 22.2 23.4 79.116 28.063 7-OHFv 0.1 5-RPn 27.0 19.4 74.194 26.413 82.069 7-RFv 92.651 84.899 26.034 7-RPn 90.067 24.0 81.762 27.855 0.0 18.6 Apigeninidin Ap 0.0 _ Malvidin Mv 25.3 24.0 4[\]-RAp 90.805 0.0 _ 82.315 28.476 3-RMv 74.994 1.2 66.627 28.0631 _ 5-RAp 83.607 0.0 75.917 25.792 4[\]-RMv 79.547 23.0 71.611 26.617 _ 20.2 7-RAp 92.958 _ 84.776 27.443 5-RMv 81.823 27.3 17.2 73.948 26.413 1.1 Pelargonidin Pg 24.3 27.4 7-RMv 89.636 24.3 17.0 81.331 27.855 3-RPg 76.594 0.0 _ 68.227 28.063

Table 1 Thermodynamic parameters of O-H dissociation and dihedral angles (ϕ) of parent compounds and their radicals

^a Kcal/mol

^b cal/mol K

^c Fv and R stand for flavylium ion and radical respectively

coplanarity of ring B with respect to the benzopyrylium ring required for maximum overlap of the *p*-orbitals of ring atoms and thus increasing the stability of the resulted phenoxyl radicals leading to better antioxidant activity of parent compounds (Leopoldini et al. 2011). There is still controversy concerning the dihedral angle of anthocyanidins depending on the used computational method and the examined compounds. The planar and non-planar conformations of a number of anthocyanidins were examined and found that the former conformer is slightly more stable by 3.0 Kcal/mol for cyanidin and 3.4 Kcal/mol for delphinidin; the dihedral angle of the optimized non-planar conformer was 23.6° in pelargonidin and 25.3° in cyanidin as calculated by DFT/ B3LYP methods (Sakata et al. 2006). Semi-empirical method at AM1 level predicted planar structure of pelargonidin with dihedral angel 0.3° (Rastelli et al. 1993). AM1 calculation predicted also a complete planar structure of 4'-hydroxy flavylium but little deviation from the planarity was found in flavylium ion (9.9°) and some mono and dihydroxylated flavyliums on ring B (Pereira et al. 1996, 1997). The same authors found also by using Ab initio methods at the level of HF/3-21 G that flavylium and 4'-hydroxy flavylium are completely planar while pelargonidin has a dihedral angle 15.3°. The difference in planarity among compounds was attributed, by the authors, to the attraction between O1 and H-atom on C2' in one way and the repulsion between H-atoms on C3 and C6' in another way.

DFT/B3LYP calculations gave also contradicted results (Lu et al. 2014) where some reports suggested planar structure of some anthocyanidins e.g., peonidin, malvidin, petunidin, and cyanidin (Guzmán et al. 2009; Lu et al. 2014), others predicted non-planarity of the same compounds with dihedral angle -10.70, -7.91, -28.14 (Woodford 2005) and 5.9° (Woodford 2005; Liu 2008), respectively. The 3-deoxyflavons apigenin and luteolin were predicted planar by DFT/B3LYP calculation (Leopoldini et al. 2004). The limited number of reported X-ray crystallography structures reveled non-planarity of some anthocyanidins where the dihedral angle was found 4.14° in apigeninidin (Woodford 2005), 3.8° in pelargonidin bromide and 10.1° in cyanidin bromide (Sakata et al. 2006).

Figure 1 shows the structure of the examined anthocyanidins and the structural model of peonidin; models of other parent compounds are placed in Supplementary material (S1) while their 3-, 4'-, 5- and 7-radicals are presented in Figs. 2 and 3 and S2–S5. In peonidin, the conformation with methoxy group toward the 3-OH group is preferred over the one with the methoxy group toward the pyrylium oxygen (Fig. 1) by stabilization energy (ΔG) 5.30 Kcal/mol. Models illustrate the charge distribution and selected bond lengths (C2-C1' and C–O bonds) in addition to the spin density distribution of radicals. Table 1 lists the dihedral angles (ϕ O1-C2-C1'-C2') and (ϕ C2-C3-O3-H3) of the parent compounds and the resulting phenoxyl radicals;

Fig. 2 Resonance, charge (positive in *green* and negative in *red*), spin density (*blue*) and selected bond lengths (*black*) of 3- and 4'-radicals (atoms carrying the unpaired electron in the main resonance are marked by a *filled circle* in 3-RPg and 4'-Mv models) (color figure online)



Fig. 3 Resonance, charge (positive in *green* and negative in *red*), spin density (*blue*) and selected bond lengths (*black*) of 5- and 7-radicals (atoms carrying the unpaired electron in the main resonance are marked by a filled circlel in 5-RPg and 7-RPg models) (color figure online)



while the former angle examines the direction of ring B (i.e., the molecular planarity), the later angle shows the orientation of the 3-O-H bond, both directions are in respect to the benzopyrylium ring. It is believed that the orientation of the 3-OH bond is responsible for ring B direction and thus the planarity of the molecule (Sakata et al. 2006; Lu et al. 2014). In previous DFT calculation, the 3-O-H bond was located away from ring B and thus Pg, Pn, and Mv were found planar (Guzmán et al. 2009). However, in the present study, 3-oxyflavylium and anthocyanidins (Pg, Pn, and Mv) as well as their 5-, 7- or 4'-radicals showed the 3-O-H bond directed towards the B ring with a dihedral angle (ϕ C2-C3-O3-H3) 17.2–27.4° causing ring B to rotate by a dihedral angle (\$\$\phi\$ O1-C2-C1'-C2'\$) 22.2-30.2°. In addition, in all 3-radicals (Figs. 2, S2), the strain between the 3-O-H group and ring B was removed; therefore, the 3-radicals were all found planar with dihedral angle (\$\$\phi\$\$ O1-C2-C1'-C2') 0.0-1.2°. The planar structure of 3-radicals is also manifested by the π -bond characters of C2-C1' bond where the bond length decreased from (1.442–1.454 Å) in the parent compounds (Figs. 1, S1) to (1.417–1.429 Å) in their 3-radicals (Figs. 2, S2). Moreover, the C2-C1' bond in 3radicals is shorter than in other corresponding radicals; for example, the bond length in 3-RPg is 1.418 Å while in 4'-RPg, 5-RPg, and 7-RPg are 1.440, 1.444, and 1.438 Å, respectively. In consequence, all compounds lacking 3-OH group (5-, 7- and 4'-OHFv and Ap) and their radicals were also found planar with dihedral angle (ϕ O1-C2-C1'-C2') $-0.1-1.1^{\circ}$.

Bond lengths of all parent compounds (Figs. 1, S1) and resulting radicals (Figs. 2–3 and S2–S5) showed that the pyrylium oxygen bonds (O1-C2 and O1-C9, where the fused bond carbons of the benzopyrylium ring are

numbered consequently 9 and 10 in the present article) are single bonds with the former is shorter showing some double bond character (1.333-1.356 Å) than the later (1.356-1.377 Å) consistent with previous AM1 and ab intio calculations of flavylium cation (Pereira et al. 1996). The carbonyl formation of the radical oxygen is expressed by shortening the C–O bond length from (1.335-1.358 Å) in the parent compounds to (1.230-1.248 Å) in all radicals.

Charge resonance and distribution

The positive charge in parent compounds (Figs. 1, S1) is located on ring carbons next to oxygen atoms (pyrylium, hydroxyl, or methoxy oxygen atoms) which could stabilize positive charge by the π -electron donating ability of oxygen atoms. The highest positive charge (0.364-0.482) is located on carbons 2, 5, 7, 9, and 4' while carbon atoms next to 3-OH, 3'-OMe or 5'-OMe (in 3-OHFv, Pg, Pn, and Mv) carry the least positive charge (0.258-0.270). The pyrylium oxygen carries the least negative charge i.e., more positive character than other oxygen atoms which is consistent with the previous DFT calculation (Guzmán et al. 2009). The increased resonance through ring B and localization of charge on C4' was observed in 4'-hydroxy flavylium compared to the unsubstituted flavylium (Pereira et al. 1997; Sakata et al. 2006). The proposed resonance (Fig. 1) explains the higher positive charge delocalization on carbons 2, 5, 7, 9, and 4' while the lower charge on C3, C3', or C5' in 3-OHFv, Pg, Pn, and Mv could be due to only the inductive effect of the neighboring oxygen atoms (Seyoum et al. 2006).

In all radicals, the positive charge is still found on the same carbon atoms as in parent compounds indicating that

radicals undergo the paired-electron resonance similar to that described for parent compounds to delocalize the positive charge. In contrary to the parent compounds, the oxygen lost the hydrogen atom (the radical oxygen) carries either positive charge (7-RFv and 7-Ap, S5) or least negative charge among oxygen atoms i.e., able to carry more positive character while the pyrylium oxygen is the second least negatively charged oxygen after the radical oxygen. The low negative charge density observed on radical oxygen atom, despite its high electronegativity reported in simple phenols (Foti et al. 2010), could be attributed, in anthocyanidin radicals, to the different types of resonance and high electron delocalization over the three rings presented in Figs. 1-3. The same observation of increasing positive charge where the radical is formed was also previously reported in other flavonoid systems (Sadasivam and Kumaresan 2011). In addition, atoms in the ring at which the radical is formed carries more positive character than the same ring in other radicals; for example, atoms of ring B in 4'-RPg carry more positive charge than the same ring in 3-RPg, 5-RPg, or 7-RPg while ring A in 5-RPg or 7-RPg carries more positive charge than in 3-RPg or 4'-RPg suggesting a radical stabilization by vicinity to a charge. Accordingly, the more positive (or less negative) charge on radical oxygen should reflect the radical stabilization and the antioxidant activity. Therefore, charge on radical oxygen (of lowest BDE), as shown in Figs. 2, S2, S3 and S4, of 4'-hydroxyflavylium ion (4'-R, -0.012), apigeninidin (5-R, -0.011), pelargonidin (3-R, -0.078), peonidin (3-R, -0.098) and malvidin (3-R, -0.104) was found correlated with scavenging percentage of DPPH (7.12, 10.20, 36.43, 53.49, and 68.64% respectively) and hydroxyl radicals (11.19, 33.33, 47.46, 65.82, and 66.78% respectively) reported previously (Ali et al. 2016). Correlations can be presented by the following equations:

% scavenging DPPH = $0.86 (\pm 6.03) - 566.35 (\pm 82.37)$ charge on radical oxygen

R = 0.970, SE =7.55, N = 5, and p = 0.006

% scavenging OH = $16.17 (\pm 7.99) - 474.44 (\pm 109.20)$ charge on radical oxygen

R = 0.929, SE = 10.01, N = 5, and p = 0.023

These correlations reflect the role of charge distribution in stabilizing the anthocyanidin radicals and thus their potent antioxidant activity.

Unpaired electron resonance and distribution

All O-H bond dissociation energy (BDE) of the examined compounds were calculated and listed in Table 1. BDE is frequently used, depending on the hydrogen atom transfer mechanism, to identify the best hydroxyl group as hydrogen atom donor (Guzmán et al. 2009; Foti et al. 2010). In the three tested 3-oxyanthocyanidins (Pg, Pn, and Mv), the 3-O-H bond is the easiest to break relative to other hydroxyl groups in each molecule with BDE 74.748-76.594 Kcal/ mol which could be derived by the steric relive of 3-O-H bond and the stability of the resulted planar radicals discussed above; this finding rationalize the previous suggestion that 3-OH group in anthocyanidins plays an important role in their antioxidant activity (Wang et al. 1997; Lu et al. 2014). There are still some variations in the literature in the order of BDE_(O-H) of anthocyanidins. For examples, the order in malvidin was reported as 3 > 4' > 5 > 7 (Lu et al. 2014; Guzmán et al. 2009) and 3 > 5 -> 7 > 4' (Pop et al. 2012) while in peonidin the reported orders were 3 > 5 > 7>4' (Guzmán et al. 2009) and 3>4'->5>7 (Lu et al. 2014). The present calculation showed that the order of $BDE_{(O-H)}$ in Ap, Pg and Pn is 3-<5-<4'-<7-OH; however, in Mv, the 4'-O-H bond has lower BDE than that of 5-O-H bond where the order is 3 - 4' - 5 - 7-OH. It can be observed that the BDE of 4'-O-H bond decreases in the order of Pg > Pn > Mv (90.128, 87.483 and 79.547 Kcal/ mol respectively) which matches the order of their antioxidant activity (Rahman et al. 2006; Ali et al. 2016) and the order of increasing number of neighboring methoxy groups (at 3' and 5') which stabilizes the 4'-radical and explains the obtained order of BDE(O-H) in Mv. Malvidin also exhibited high scavenging activity against singlet oxygen even larger than cyanidin that contains a catechol moiety which was attributed to the role of 4'-OH and the electron donating o-OMe groups in favoring the interaction with the singlet oxygen electrophile (De Rosso et al. 2008). These results indicate the important of the BDE of both 3and 4'-OH in determining the antioxidant activity of anthocyanidins. The order of Gibbs free energy (ΔG) of the O-H dissociation matches that of the corresponding BDE in all the examined flavyliums and anthocyanidins while the entropy change (ΔS) changed only within 2.6 cal/mol K indicating that the process is mainly governed by the dissociation enthalpy. Although BDE of hydroxyl groups is frequently correlated with the antioxidant activity of phenolic compounds, no such quantitative correlation was reported between the antioxidant activity of anthocyanidins and their lowest $BDE_{(O-H)}$ (or $\Delta G_{(O-H)}$). Therefore, this correlation not only shows the dependence of antioxidant activity on the BDE(O-H) but also validates the choice of the eligible hydroxyl group for each anthocyanidin. Correlations of radical scavenging activity (Ali et al. 2016) with the least $BDE_{(O-H)}$ in each compound are presented by the following equations.

% scavenging DPPH = $290.83 (\pm 81.83) - 3.18 (\pm 1.02)$ BDE R = 0.875, N = 5, SE = 14.95, p = 0.052

% scavenging OH = $292.73 (\pm 35.87) - 3.08 (\pm 0.45)$ BDE

R = 0.970, N = 5, SE = 6.55, p = 0.006

The unpaired electron distribution is modeled by the spin density as illustrated in models of flavylium and anthocyanidin radicals presented in Figs. 2-3 and S2-S5. The 3radicals can be stabilized by resonance through 5-, 7 or 4'oxygen atom (Ali et al. 2016) as presented in Fig. 2. These resonance pathways are manifested by the location of spin density (>0.025) on the three pathway atoms (O3, C4, C9, C4)C7, C5, and O5), (O3, C4, C9, C7, and O7) and (O3, C4, C9, C2, C2', C4', and O4') respectively. On the other hand, O1 can not be unpaired electron acceptor in any of these resonance pathways and thus does not bear any spin density in all the examined 3-oxyanthocyanidin radicals. It can also note the similarity between the unpaired electron distribution of 3-radicals and charge distribution mentioned above. The planarity and the wide distribution of the unpaired electron of 3-radicals could explain the lower BDE of the 3-OH group and the impotence of this group, discussed earlier, in determining the antioxidant activity of 3-oxyanthocyanidins.

The 4'-RFv radical can be stabilized by resonance through the pyrylium oxygen, O1 (Fig. 2) where alternative atoms in that route (O4', C3', C5', C1', and O1) carry the highest spin density (0.054 (O1)-0.385 (O4')) followed by delocalization of the unpaired electron over ring C then ring A at lesser extent. Similar behavior of higher spin density on alternative atoms of ring B than on atoms of rings A and C can be observed in all other 4'-radicals. Considerable higher spin density is located on the methoxy oxygen of Pn (0.090) and Mv (0.060 and 0.108) than on hydroxyl oxygen atoms (≤ 0.017) indicating the involvement of the *ortho* methoxy oxygen in the unpaired electron resonance as shown in Figs. 2, S3. In addition, the 4'-radical oxygen carries lower spin density in Pn (0.270) and Mv (0.269) than in Pg (0.376) radicals; besides, the C4'-O bond in Pn and Mv has more π -bond character (bond length 1.230 and 1.231 Å, respectively) than in Pg (1.243 Å) which all indicate the strong effect of the ortho electron donating methoxy groups in delocalizing the unpaired electron¹⁶ and explains the lower BDE of 4'-O-H than that of 5-O-H in Mv, in contrary to other tested anthocyanidins.

Radicals of 5-OH and 7-OH groups are characteristic by high spin density on alternative atoms of rings A and C i.e.,

O5 or O7 respectively in addition to C6, C8, C10, C3, and O1 suggesting a resonance through the pyrylium oxygen as illustrated in Figs. 3, S4, S5. The resonance can also be extended to O3 in 3-oxyanthocyanidins instead of O1 as indicated by the existence of spin density on O3. On the other hand, the absence of possible resonance between the *meta* 5- and 7-oxygen atoms led to existence of unpaired electron on only either one of them. The resonance stabilization of 5- and 7-anthocyanidin radicals supports the reported significant higher antioxidant activity of Ap than that of 4'-OHFv (Ali et al. 2016). In addition, the location of the double bonds of the benzipyrylium ring (rings A and C) in all 5- and 7-anthocyanidin radicals matches exactly that of the suggested resonance.

It can be observed that the spin density is distributed over the three rings in all radicals even those showed nonplanarity indicating that the planarity discrepancy does not prevent the through resonance among the rings; however, the movement of the unpaired electron from the benzopyrylium ring to ring B is higher in the planar 3-radicals than in the corresponding 5- or 7-radicals; for example, the sum spin density of ring B is higher in planar 3-RPg radical (Fig. 2) than in the non-planar 5-RPg or 7-RPg radicals (Fig. 3). In all radicals, the radical oxygen retains the highest spin density consistent with previous calculations (Lu et al. 2014). All radicals also form a π -bond character between radical oxygen and neighboring carbon atom (C=O) revealing the initiation of a single-electron resonance for delocalization of the unpaired electron. It is well known that more delocalized unpaired electron reflected by more spin density distribution results in more stable radical and lower BDE (Sadasivam and Kumaresan 2011). In addition, the presence of electron donating hydroxyl groups on carbon atoms of a positive spin density increases radical stabilization (Seyoum et al. 2006). Therefore, the decrease of spin density on the radical oxygen is a measure of the increased resonance and thus can be used as indicator of radical stabilization and antioxidant activity. Correlating the spin density on the radical oxygen (of the lowest BDE_{OH}) with the previously reported (Ali et al. 2016) scavenging of DPPH and hydroxyl radicals gave good correlations.

% scavenging DPPH = $257.91 (\pm 41.68)$ -624.95 (±116.34) spin density

R = 0.952, N = 5, SE = 9.49, p = 0.013

% scavenging OH = $216.58 (\pm 64.67)$ -481.65 (± 180.51) spin density

R = 0.839, N = 5, SE = 14.72, p = 0.076

Finally, the 3-radical oxygen of each anthocyanidin carries less spin density and higher negative charge (less positive character) than other radical oxygen atoms; for example, compare spin density (0.347) and charge (-0.078) of Pg (Fig. 2) the 3-radical oxygen with those of other radicals (Figs. 3, S3). Therefore, it could be stated that the less positive charge and spin density on radical oxygen, the higher antioxidant activity since the charge and unpaired electron are then more delocalized over the rings.

Conclusion

A DFT study of the anthocyanidins apigeninidin, pelargonidin, peonidin and malvidin, plus the 3-, 4'-, 5- or 7 flavylium ions as well as their phenoxyl radicals revealed some points concerning their molecular planarity, charge distribution and the resonance stability of their radicals. The existence and direction of the 3-OH bond affects ring B orientation and compound planarity where compounds and radicals lacking 3-O-H bond were found planar. Bond dissociation energy of O-H bond in anthocyanidins was in the order of 3 - < 5 - < 4' - < 7-OH bond except for in malvidin where 4'-OH has less BDE than 5-OH bond because of the extra stability provided by the two o-OMe groups. Positive charge was stabilized by resonance on carbons 2, 5, 7, 9 and 4'. The 3-anthocyanidin radicals were stabilized by unpaired electron resonance through 5-, 7- or 4'-oxygen atom while 5-, 7- or 4'-radicals were stabilized by resonance through pyrylium or 3-oxygen atom. In all radicals, radical oxygen retains the highest spin density and forms a carbonyl group (C=O) with neighboring carbon atom. In addition, delocalization of the unpaired electron was observed through the three rings but was higher in planar radicals. Experimental radical scavenging activity was found well correlated with both charge and unpaired electron delocalization expressed by charge and unpaired electron density on radical oxygen.

Compliance with ethical standards

Conflict of interest The authors declare that they have no competing interests.

References

- Ali HM, Abo-Shady A, Sharaf Eldeen HA, Soror HA, Shousha WG, Abdel-Barry OA, Saleh AM (2013) Structural features, kinetics and SAR study of radical scavenging and antioxidant activities of phenolic and anilinic compounds. Chem Cent J 7:53–61. doi:10. 1186/1752-153X-7-53
- Ali HM, Ali IH (2015) QSAR and mechanisms of radical scavenging activity of phenolic and anilinic compounds using structural, electronic, kinetic, and thermodynamic parameters. Med Chem Res 24:987–998. doi:10.1007/s00044-014-1174-y

- Ali HM, Almagribi W, Al-Rashidi MN (2016) Structure-activity relationship, kinetics And synthesis of anthocyanidins And anthocyanins as radical scavengers and reductants. Food Chem 194:1275–1282. doi:10.1016/j.foodchem.2015.09.003
- Amić D, Amić DD, Bešlo D, Rastija V, Lučić B, Trinajstić N (2007) SAR and QSAR of the Antioxidant Activity of Flavonoids. Curr Med Chem 14:827–845
- Andersen OM, Jordheim M (2006) The anthocyanins. In: Andersen OM, Markham KR (Eds) Flavenoids, chemistry, biochemistry and applications. Taylor & Francis Group, Boca Raton, p 471–551
- Azevedo J, Fernandes I, Faria A, Oliveira J, Fernandes A, de Freitas V, Mateus N (2010) Antioxidant properties of anthocyanidins, anthocyanidin-3- glucosides and respective portisins. Food Chem 119:518–523
- Bagchi D, Garg A, Krohn RL, Bagchi M, Bagchi BJ, Balmoori J, Stohs SJ (1998) Protective effects of grape seed proanthocyanidins and selected antioxidants against TPA-induced hepatic and brain lipid peroxidation and DNA fragmentation, and peritoneal macrophage activation in mice. Gen Pharmacol 30:771–776
- Boveris AD, Galatro A, Sambrotta L, Ricco R, Gurni AA, Puntarulo S (2001) Antioxidant capacity of a 3-deoxyanthocyanidin from soybean. Phytochemistry 58:1097–1105
- Carbonneau M-A, Cisse M, Mora-Soumille N, Dairi S, Rosa M, Michel F, Lauret C, Cristol J-P, Dangles O (2014) Antioxidant properties of 3-deoxyanthocyanidins and polyphenolic extracts from Côte d'Ivoire's red and white sorghums assessed by ORAC and in vitro LDL oxidisability tests. Food Chem 145:701–709
- Castaňeda-Ovando A, Pacheco-Hernández ML, Páez-Hernández ME, Rodríguez JA, Galán-Vidal CA (2009) Chemical studies of anthocyanins: a review. Food Chem 113:859–871
- De Rosso VV, Vieyra FEM, Mercadante AZ, Borsarelli CD (2008) Singlet oxygen quenching by anthocyanin's flavylium cations. Free Radic Res 42:885–891
- Foti MC (2012) Solvent effects on the activation parameters of the reaction between an α-tocopherol analogue and dpph•: the role of H-bonded complexes. Int J Chem Kinet 44:524–531
- Foti MC, Amorati R, Pedulli GF, Daquino C, Pratt DA, Ingold KU (2010) Influence of "Remote" intramolecular hydrogen bonds on the stabilities of phenoxyl radicals and benzyl cations. J Org Chem 75:4434–4440
- Guzmán R, Santiago C, Sánchez M (2009) A density functional study of antioxidant properties on anthocyanidins. J Mol Struct 935:110–114
- Kähkönen MP, Heinonen MH (2003) Antioxidant activity of anthocyanins and their aglycons. J Agric Food Chem 51:628–633
- Kong J-M, Chia L-S, Goh N-K, Chia T-F, Brouillard R (2003) Analysis and biological activities of anthocyanins. Phytochemistry 64:923–933
- Leopoldini M, Pitarch IP, Russo N, Toscano M (2004) Structure, conformation, and electronic properties of apigenin, luteolin, and taxifolin antioxidants. A first principle theoretical study. J Phys Chem A 108:92–96
- Leopoldini M, Russo N, Toscano M (2011) The molecular basis of working mechanism of natural polyphenolic antioxidants. Food Chem 125:288–306
- Liu Z (2008) Theoretical studies of natural pigments relevant to dyesensitized solar cells. J Mol Struct 862:44–48
- Lu L, Qiang M, Li F, Zhang H, Zhang S (2014) Theoretical investigation on the antioxidative activity of anthocyanidins: a DFT/ B3LYP study. Dyes Pigments 103:175–182
- Ma T, Hu N, Ding C, Zhang Q, Li W, Suo Y, Wang H, Bia B, Ding C (2016) In vitro and in vivo biological activities of anthocyanins from *Nitraria tangutorun* Bobr. Fruits. Food Chem 194:296–303
- Pereira GK, Donate PM, Galembeck SE (1996) Electronic structure of hydroxylated flavylium cation. J Mol Struct 363:87–96

- Pereira GK, Donate PM, Galembeck SE (1997) Effects of substitution for hydroxyl in the B-ring of the flavylium cation. J Mol Struct 392:169–179
- Pop R, Ştefănuţ MN, Căta A, Tănasie C, Medeleanu M (2012) Ab initio study regarding the evaluation of the antioxidant character of cyanidin, delphinidin and malvidin. Cent Eur J Chem 10:180–186
- Rahman MM, Ichiyanagi T, Komiyama T, Hatano Y, Konishi T (2006) Superoxide radical- and peroxynitrite-scavenging activity of anthocyanins; structure–activity relationship and their synergism. Free Radic Res 40:993–1002
- Rastelli G, Costantino L, Albasini A (1993) Physico-chemical properties of anthocyanidins. Part 1. Theoretical evaluation of the stability of the neutral and anionic tautomeric forms. J Mol Struct 279:157–166
- Sadasivam K, Kumaresan R (2011) Theoretical investigation on the antioxidant behavior of chrysoeriol and hispidulin flavonoid compounds—a DFT study. Comput Theor Chem 963:227–235

- Sakata K, Saito N, Honda T (2006) Ab initio study of molecular structures and excited states in anthocyanidins. Tetrahedron 62:3721–3731
- Seyoum A, Asres K, El-Fiky FK (2006) Structure–radical scavenging activity relationships of flavonoids. Phytochemistry 67:2058–2070
- van Acker SBE, van den Berg D-J, Tromp MNJL, Griffioen DH, van Bennekom WP, van der Vijgh WJF, Bast A (1996) Structural aspects of antioxidant activity of flavonoids. Free Radic BioMed 20:331–342
- Wang H, Cao G, Prior RL (1997) Oxygen radical absorbing capacity of anthocyanins. J Agric Food Chem 45:304–309
- Woodford JN (2005) A DFT investigation of anthocyanidins. Chem Phys Lett 410:182–187
- Zhao CL, Chen ZJ, Bai XS, Ding C, Long TJ, Wei FG, Miao KR (2014) Structure–activity relationships of anthocyanidin glycosylation. Mol Divers 18:687–700