

α and β diversity of fishes in relation to a gradient of habitat structural complexity supports the role of environmental filtering in community assembly

Eduardo R. Cunha^{1,2} D [·](http://orcid.org/0000-0001-7858-2133) Kirk O. Winemiller² · João C. B. da Silva¹ · Taise M. Lopes¹ · Luiz C. Gomes¹ · **Sidinei M. Thomaz¹ · Angelo A. Agostinho1**

Received: 27 August 2018 / Accepted: 15 March 2019 / Published online: 23 March 2019 © Springer Nature Switzerland AG 2019

Abstract

α-diversity often responds to habitat structural complexity as a unimodal function. In aquatic systems, increasing density of aquatic vegetation creates more habitat structural complexity for fishes, but only up to a certain threshold, beyond which fish abundance and diversity are restricted by reduced space. As a result, species turnover and nestedness should be observed over habitat structural complexity gradients, reflecting the sorting of species according to aspects of their environment. We investigated the relationship of fish α and β diversity along gradients of habitat structural complexity created by aquatic vegetation in the floodplain of Upper Paraná River. We collected a total of 1832 fishes (24 species) along vegetation density gradients. Our results revealed that α diversity peaked at intermediate levels of habitat structural complexity where interstitial spaces were numerous but no so small as to limit occupancy by most fishes. Low α diversity was associated with lower habitat structural complexity, as commonly reported, and this may result from the influence of predation mortality or threat where there is less physical structure that provides refuge from predators and interference with predator lines of sight for prey detection. Fish diversity is low in patches with high habitat structural complexity because small interstitial spaces restrict fish size and dissolved oxygen concentration sometimes is low. Aquatic vegetation density in floodplain habitats therefore functions as a strong environmental filter influencing spatial patterns of fish α and β diversity.

Keywords Brazil · Community structure · Floodplain · Habitat structure · Paraná River · Species nestedness · Species replacement · Species turnover · Freshwater

Introduction

Understanding the mechanisms that yield biodiversity patterns is one of the most intriguing and longest-standing challenges in ecology (Whittaker [1960,](#page-11-0) [1972\)](#page-11-1). Different factors can influence local diversity (α diversity) and its variation

Electronic supplementary material The online version of this article [\(https://doi.org/10.1007/s00027-019-0634-3\)](https://doi.org/10.1007/s00027-019-0634-3) contains supplementary material, which is available to authorized users. (β diversity) within regional species pools (ɣ diversity) at different scales of space and time (Fukami [2004\)](#page-9-0). Investigations that simultaneously consider various components of diversity and analyze their variation in response to environmental factors can reveal processes shaping ecological communities (Leibold and Mikkelson [2002;](#page-10-0) Fukami [2004;](#page-9-0) Kraft et al. [2011;](#page-10-1) Mittelbach and Schemske [2015](#page-10-2)). An important environmental influence on species diversity is habitat structural complexity (MacArthur and MacArthur [1961](#page-10-3); Tews et al. [2004;](#page-11-2) Kovalenko et al. [2012;](#page-10-4) Stein and Kreft [2015](#page-11-3)). Habitat structural complexity is commonly characterized as the density or diversity of physical structures that influence habitat suitability for organisms (MacArthur and MacArthur [1961;](#page-10-3) McCoy and Bell [1991;](#page-10-5) Tokeshi and Arakaki [2012\)](#page-11-4). Habitat structural complexity can enhance fitness by increasing availability of food resources and/or refuges from predation (Gause [1936](#page-9-1); Huffaker [1958;](#page-10-6) MacArthur and MacArthur [1961;](#page-10-3) Sánchez-Botero et al. [2007\)](#page-10-7), which

 \boxtimes Eduardo R. Cunha edurcunha@gmail.com

¹ Programa de Pós-Graduação em Ecologia de Ambientes Aquáticos Continentais, Universidade Estadual de Maringá, Maringá, PR 87020-900, Brazil

² Department of Wildlife and Fisheries Sciences, Texas A&M University, College Station, TX 77843-2258, USA

should result in a positive relationship between α diversity and habitat structural complexity. In some habitats, however, greater structural complexity may reduce useable space for large organisms (Gibb and Parr [2010](#page-9-2); Strayer and Findlay [2010](#page-11-5); Yeager and Hovel [2017\)](#page-11-6). In these cases, α diversity should reveal a negative relationship with habitat structural complexity, or a hump-shaped pattern that derives from the influence of two opposing mechanisms (Crowder and Cooper [1979;](#page-9-3) Heck and Orth [1980;](#page-10-8) Diehl and Kornijow [1998](#page-9-4); Tokeshi and Arakaki [2012](#page-11-4); Paxton et al. [2017](#page-10-9)).

Gradients of habitat structural complexity are common in many landscapes (August [1983](#page-9-5); Fahr and Kalko [2011](#page-9-6)). A single lake, for example, can show patches with different levels of structural complexity that range from bare sediment to dense beds of submerged vegetation (St. Pierre and Kovalenko [2014](#page-11-7)). Because species respond differentially to levels of structural complexity, patterns of coexistence may be clumped with species turnover along complexity gradients (Leibold and Mikkelson [2002](#page-10-0); Leibold et al. [2004;](#page-10-10) Strayer and Findlay [2010](#page-11-5)). Limiting factors at lowest and highest levels of habitat structural complexity, such as predation risk (low complexity) and lack of space for movement (high complexity), would further influence assemblage nestedness (Leibold and Mikkelson [2002](#page-10-0); Leibold et al. [2004](#page-10-10); Hylander et al. [2005](#page-10-11)).

Whereas variation in α diversity may reveal insights about mechanisms for coexistence of species at local scales, β diversity provides insight about community assembly in response to environmental gradients (Hylander et al. [2005](#page-10-11); Petsch et al. [2017\)](#page-10-12). Although these diversity components are complementary, most studies investigating changes in assemblage structure in relation to habitat structural complexity have only examined α diversity (e.g., Newman et al. [2015](#page-10-13)). Few studies have attempted to analyze variation in β diversity across gradients of structural complexity, and studies are needed that jointly investigate both diversity components (Petsch et al. [2017](#page-10-12)). Moreover, analysis of species turnover and nestedness can provide further insights about mechanisms of community assembly (Hylander et al. [2005](#page-10-11); Fahr and Kalko [2011\)](#page-9-6).

Here, we conducted a field survey to investigate the role of structural complexity as a function of vegetation density regulating small fish α and β diversity in lakes of a floodplain in Brazil. Several studies investigating the effects of habitat structural complexity on diversity have been performed in tropical aquatic ecosystems (Arrington et al. [2005](#page-9-7); Willis et al. [2005](#page-11-8); Dibble and Pelicice [2010\)](#page-9-8). Fishes in general, and freshwater fishes in particular, respond strongly to habitat structural complexity (Strayer and Findlay [2010](#page-11-5)), with many species relying on submerged structures (e.g., aquatic vegetation) for feeding, refuge from predation, spawning or nesting (Rozas and Odum [1988](#page-10-14); Rossier et al. [1996;](#page-10-15) Santos et al. [2011](#page-11-9); Yeager and Hovel [2017](#page-11-6)). In the tropics, diverse fishes of small body size take

refuge in beds of aquatic vegetation (Lopes et al. [2015](#page-10-16)). In tropical floodplains, the density of aquatic vegetation generally varies along water depth gradients, with higher plant density and smaller and more numerous interstices in shallow water near shore (Lopes et al. [2015\)](#page-10-16). In addition, dissolved oxygen concentration often varies as a function of plant density and distance from shore, with dense patches in shallow water sometimes experiencing aquatic hypoxia during the night (Miranda and Hodges [2000;](#page-10-17) Miranda et al. [2000](#page-10-18)). Therefore, aquatic vegetation in floodplain habitats creates environmental gradients of both structural complexity and environmental stress.

We formulated a series of questions and conducted a deductive approach to test a hypothesis about the effects of habitat structural complexity regulating small fish α and β diversity. Our questions included: Do α and β diversity of fish assemblages differ from diversity derived from a random partitioning of regional diversity? If yes, how is habitat complexity related to species diversity components? What is the role of oxygen concentration in explaining the relationship between diversity components of fish assemblages and habitat structural complexity? Considering previous theory on habitat structural complexity, we hypothesized that α diversity is related to habitat structural complexity in a hump-shaped fashion, while habitat structural complexity leads to changes in β diversity considering both turnover and nestedness components. We further predicted that (1) α and β diversity patterns are different from those expected in random assemblages; (2) along a gradient of habitat structural complexity, α diversity increases until an optimum level but decreases after that; and (3) a gradient of habitat structural complexity leads to changes in β diversity accompanied by increases in species turnover and nestedness at lowest and highest levels of habitat structural complexity. We also investigated the potential influence of fish size and species tolerance to hypoxic condition on α and β diversity patterns in relation to gradients of structural complexity and dissolved oxygen concentration. Aspects of β diversity were investigated using the concepts extended to abundance data described by Podani et al. (2013) , who refer to (1) β diversity as differences in abundance of particular species along ecological gradients (hereafter total β diversity), (2) turnover as the replacement of abundances of particular species with the same abundances of different species along ecological gradients, and (3) nestedness as reductions in the abundances of particular species that yields assemblage subsets along ecological gradients.

Materials and methods

Study area

Surveys were performed in natural lakes in the floodplain of the Baía River, a black-water tributary of the Upper Paraná River in Brazil. The Baía and Paraná rivers meander over broad alluvial plains within an area of $28,106 \text{ km}^2$ (Stevaux [1994](#page-11-10); Orfeo and Stevaux [2002](#page-10-20); Agostinho et al. [2004c\)](#page-9-9) and have seasonal patterns of discharge (Agostinho et al. [2000,](#page-9-10) [2004c\)](#page-9-9). Geology of the region is dominated by sedimentary and volcanic rocks (Stevaux [1994](#page-11-10); Orfeo and Stevaux [2002](#page-10-20)), and the floodplains contain numerous secondary channels, lakes, and wetlands (Stevaux [1994](#page-11-10); Agostinho et al. [2004c](#page-9-9); Stevaux and Souza [2004](#page-11-11)). Within the Baía floodplain, lakes and flooded areas are highly connected during the wet season. Most lakes are shallow [depth<3 m (Stevaux and Souza [2004](#page-11-11))] with extensive littoral zones containing extensive beds of aquatic vegetation. Dominant species of aquatic vegetation are *Eichhornia azurea* (Sw.) Kunth and *Eichhornia crassipes* (Mart.) Solms. These floating vegetation have similar morphology consisting of networks of submerged stems and roots, with emergent stems and leaves that increase habitat complexity for many small fish species (Delariva et al. [1994;](#page-9-11) Gomes et al. [2012](#page-10-21)).

The regional fish fauna has $>$ 270 species (Agostinho et al. [2004a](#page-9-12), [2007a\)](#page-9-13), with the Baía River generally having greater fish densities and diversity compared to similar habitats in flowing reaches of the nearby Paraná River (Fernandes et al. [2009](#page-9-14)). Several piscivorous species are common, including *Pseudoplatystoma corruscans* (Spix and Agassiz 1829), *Cichla kelberi* Kullander and Ferreira, 2006, *Hemisorubim platyrhynchos* (Valenciennes 1840), *Rhaphiodon vulpinus* Spix and Agassiz 1829 and *Astronotus crassipinnis* (Heckel 1840) (Agostinho et al. [2004b](#page-9-15), [2007a;](#page-9-13) Gomes et al. [2012](#page-10-21)). These relatively large species are highly mobile and move among habitats throughout the landscape. Many small species (≤4 cm total length; e.g., *Hyphessobrycon, Moenkhausia* and *Serrapinus* spp.) and juveniles of larger species are largely restricted to stands of submerged vegetation where they feed on aquatic invertebrates and benthic microalgae while also obtaining cover that reduces exposure to predators. Given their small size and vulnerability to piscivorous fishes, dispersal by these small fishes is limited compared to that by larger fishes (Delariva et al. [1994;](#page-9-11) Agostinho et al. [2007b](#page-9-16); Gomes et al. [2012;](#page-10-21) Lopes et al. [2015](#page-10-16)).

Sampling procedure

In December 2011, field surveys were performed at 48 locations distributed in a nested design within four lakes in

the floodplain of the Baía River. In each lake, we randomly selected two vegetation mats dominated by mixtures of *E. azurea* and *E. crassipes*, with the requirement that there was variation in the density of vegetation between and within stands. In general, the area of each vegetation stand was ≥ 400 m², and stands were separated from each other by at least 100 m. We sampled six locations along 15-m transects within each stand. Each transect began at the stand margin with open water and ran towards the shoreline, with collecting locations positioned 3 m apart. Variation in vegetation density was observed both within and between transects, so that sample collection spanned a range of levels of structural complexity as a function of vegetation density.

For sampling small fishes, we used $0.3 \text{ m} \times 0.3 \text{ m} \times 0.3 \text{ m}$ box-trap made with plexiglass [details appear in Dibble and Pelicice [\(2010](#page-9-8)); Lopes et al. ([2015\)](#page-10-16)]. These traps have floats that keep them near the water surface and embedded within the aquatic vegetation. This device captures fishes with lengths up to c.a. 10 cm and have minimal impact to habitat structure during deployment (Ribeiro and Zuanon [2006](#page-10-22)). Fishes living within plant structures are small (median: 3.1 and 95% CI 2.3–6.2 cm); larger species are sometimes encountered in vegetated areas, but these generally occupy open-water spaces where their movements are not impeded. To test for potential bias associated with gear selectivity, fish capture data were compared for the traps and surveys conducted using seine nets in floating vegetation stands in the same lakes. Fish community structure or body length distributions were not significantly different for samples obtained by the two methods (details of this analysis appear in the supplementary materials). Traps were deployed in each survey site at 0700 h and then inspected for fish after 12 and 24 h. Fishes from each trap survey were counted, identified to species level with the aid of taxonomic keys (Graça and Pavanelli [2007;](#page-10-23) Ota et al. [2018](#page-10-24)). α diversity at each site along a transect was recorded as species richness per trap day.

To characterize physical and chemical aspects of habitat at each trap location, we measured the following parameters at 0.4 m depth using a YSI probe and digital meter: dissolved oxygen (mg L^{-1}), pH, conductivity (μS m⁻¹), and temperature (°C). Water depth (m) was measured using a meter stick. These measures were taken twice at each trap location on the date of the survey (morning and evening) and averaged. Although averaging obscures minimum and maximum values that could have ecological implications beyond the average (e.g., minimum dissolved oxygen), averages nonetheless effectively revealed spatial gradients for environmental factors. Because dissolved oxygen concentration sometimes has an influence on fish distributions within vegetation stands (Miranda and Hodges [2000](#page-10-17)), we included this environmental factor as an explanatory variable in our

statistical analysis. Plant density was estimated at each trap location. After each trap was removed, all plant material within a 0.5×0.5 m quadrat adjacent to the trap location was collected, separated into emergent and submerged components, and each component was dried (60 °C) until a constant dry weight (DW) was achieved. To provide an estimate of habitat structural complexity, submerged plant DW was converted to DW per unit volume of habitat (Kg DW L^{-1}), with habitat volume calculated from measurements of water depth and the area sampled.

Statistical analysis

Additive partitioning of species diversity

We partitioned the total diversity of samples into α and β components and used null models to assess if the observed partition is structurally different from random expectations. Nonrandom species distributions would suggest that ecological factors have a significant influence on diversity patterns (Crist et al. [2003\)](#page-9-17). These results were used to test our hypothesis that habitat structural complexity and perhaps dissolved oxygen concentrations influence spatial patterns of fish diversity in vegetation stands. Diversity partitioning was performed using the additive framework and randomization method of Crist et al. ([2003\)](#page-9-17). Null models were derived from 9999 randomizations and compared with observed values. Analyses were performed in the vegan package (Oksanen et al. [2019](#page-10-25)) in R.

α diversity and explanatory variables' associations

We used Linear Mixed-Effect Models (LMM) to test the hypothesis that α diversity is related to habitat structural complexity in a hump-shaped fashion, and to test for a potential influence of dissolved oxygen concentration in the relationship. Fixed effects were vegetation density, [vegetation density] 2 (to enable evaluation of hump-shaped relationships of diversity with structural complexity) and dissolved oxygen concentration. We used a hierarchical random structure to account for the nested structure of samples and to ensure the assumption of independence among samples (see the work of Millar and Anderson [\(2004\)](#page-10-26) for a discussion on nested designs in fisheries field experiments). Random effects included nested effects of lake identity and transect identity, in addition to the distance from the survey location to the margin of the vegetation patch with open water. We tested for multicollinearity by estimating a variance inflation factor (VIF), and all VIFs were $<$ 3 (betweenvariable $|r| < 0.65$), indicating no collinearity (Zuur et al. [2010](#page-11-12)). Model selection based on Akaike Information Criterion (AIC_C, corrected second-order criterion) was done on data subsets with different combinations of explanatory

variables. Parsimonious models were those with Δ_i AIC_{*C*} < 2 (Burnham and Anderson [2002\)](#page-9-18). Model selection only supported inclusion of the second-order term for vegetation density when the model also included the first-order term for vegetation density. We used Wald χ^2 tests and conditional and marginal \mathbb{R}^2 for validating model fitting. Analyses were performed in R using vegan, MuMIn and lme4 packages (Bartón [2018;](#page-9-19) Bates et al. [2014](#page-9-20)).

β diversity and explanatory variables' associations

Pairwise measurements of $β$ diversity were estimated using dissimilarity indexes based on species abundance data. β diversity was expressed as the Ružička dissimilarity index (quantitatively equivalent to Jaccard) that measures changes in assemblage composition between two sites with particular emphasis on differences in species abundance. The Ružička dissimilarity index was calculated as $(B+C)/(A+B+C)$, where A is the sum of abundances for each species that are shared by sites 1 and 2, B is the sum of abundances for each species that are exclusive to site 1, and C is the sum of abundances for each species that are exclusive to site 2. Because this measure of β diversity incorporates variation due to species turnover and nestedness, we used the decomposition of Ružička dissimilarity index to investigate variation in these components. In general, two methods for decomposing β diversity have been promoted (see Legendre [2014\)](#page-10-27), and the method used here is the one proposed in Podani et al. ([2013](#page-10-19)). Their method provides a robust measure of nestedness that differs fundamentally from other popular methods (e.g., Baselga [2010\)](#page-9-21) that may bias interpretations of nestedness (Schmera and Podani [2011\)](#page-11-13). Species turnover was calculated as the relativized abundance replacement, which measures the replacement of abundances of particular species in one site by the same abundances of completely different species in another site (Podani et al. [2013\)](#page-10-19). Relativized abundance replacement is measured as $2 \text{ min}(B, C)/(A + B + C)$, and the maximum relativized abundance replacement is achieved when two sites show the same total abundance but share no species. Nestedness was calculated using relativized nestedness that measures the extent to which species abundances at one site are a subset of species abundances at another site (Podani et al. [2013](#page-10-19)). Relativized nestedness is measured as $(A+|B-C|)/(A+B+C)$ when $A>0$, otherwise it equals to 0. Relativized nestedness yields its maximum value when abundances for every species at one site do not exceed their respective abundances at the other site.

We used distance-based Redundancy Analysis to test the hypothesis that habitat structural complexity leads to changes in β diversity considering both turnover and nestedness components, and to test for potential influence of dissolved oxygen concentration on the relationship. Explanatory variables were vegetation density, [vegetation density]², and dissolved oxygen concentration. We incorporated nested structure of sampling design by conditioning the distance from the survey location to open water and by using null models that constrain permutations under a hierarchical design for significance tests (Winkler et al. [2015\)](#page-11-14). Similar to α diversity analysis, model selection was conducted considering Akaike Information Criterion (AIC_C, corrected second-order criterion). Model selection was done on data subsets with different combinations of explanatory variables, and only supported inclusion of the second-order term for vegetation density when the model also included the firstorder term for vegetation density. The most parsimonious model was that with the smallest AIC*C*. To provide further support for interpretations, we calculated the index for local contribution to β diversity (LCBD), an estimate of the ecological uniqueness of a particular local assemblage (Legendre and De Caceres [2013\)](#page-10-28). LCBD measures the proportion of variation in β diversity attributed to a particular local assemblage, and can be obtained by decomposing the sums of squares of a dissimilarity matrix. We calculated LCBD from β diversity measurements and its components and used them as a basis to weight the sizes of circles representing local assemblages in ordination plots. Analyses were performed using the vegan package (Oksanen et al. [2019\)](#page-10-25) in R, including functions provided by Pierre Legendre [\(2014](#page-10-27)).

Biological features

Because one of our hypotheses predicts that α diversity declines beyond an optimum level of habitat structural complexity due to reduction of interstitial spaces, we evaluated relationships between fish size and vegetation density. We used the median fish standard length and the variation in fish standard length in each sample to characterize fish size. Not all individuals were measured; therefore, bootstrapping was performed to estimate the median and the variation in fish standard length within local assemblages. Bootstraps involved resampling standard length measurements for the species abundances in a sample from pools of species standard length measurements. Pools of species standard length measurements considered individuals measured in the whole data set. Estimates for the median and the variation in fish standard length were obtained by median, minimum and maximum values after 999 simulations. We used Linear Mixed-Effect Models to test whether median and variation in fish standard length were constrained by habitat structural complexity. Vegetation density was treated as a fixed effect, and random effects included nested effects of lake identity and transect identity, in addition to the distance from the open-water end of the transect. Wald χ^2 tests and conditional and marginal $R²$ were used to assess model fit. Power function and logarithmic transformations were applied to vegetation

density in order to linearize relationships. Analyses were performed in R using lme4 and MuMIn packages (Bartón [2018;](#page-9-19) Bates et al. [2014\)](#page-9-20).

We also evaluated the hypothesis that interspecific differences in adaptation for resistance to aquatic hypoxia yields a pattern of species filtering along gradients of dissolved oxygen. We tested the proportional contribution of species possessing auxiliary respiratory adaptations or documented tolerance to hypoxia to α diversity and total fish abundance in relation to dissolved oxygen gradients. Species were classified based on previous published literature reporting adaptations such as presence of dermal lip protuberances that enhance aquatic surface respiration, aerial respiration using the gut or swim bladder for gas exchange, capacity for anaerobic respiration and hypoxia tolerance. We used Generalized Linear Mixed-Effect Models to test the relationship between the percentage of hypoxia-tolerant fishes in the local assemblage and the dissolved oxygen concentration. The fixed effect was oxygen concentration, and random effects included nested effects of lake identity, transect identity and distance from the open-water end of the transect. Residuals were modeled following a binomial distribution using a log-link function to handle the percentage data. Wald χ^2 tests and conditional and marginal R^2 were used to assess model fit. Analyses were performed in R using lme4 and MuMIn packages (Bartón [2018\)](#page-9-19).

Results

A total of 1832 small fishes belonging to 24 species was collected during the study. Most species were Characiformes, and the most abundant species were *Moenkhausia bonita* Benine, Castro & Sabino 2004, *Hyphessobrycon eques, Serrapinnus heterodon* (Steindachner 1882) and *Serrapinnus notomelas* (Eigenmann 1915) (210–351 individuals per species) (Fig. [1](#page-5-0)). More than half of the collected species lack known adaptations for resisting aquatic hypoxia, except for aquatic surface respiration during extreme hypoxia, a nearly universal behavior in teleost fishes. The number of fish caught per trap varied from to 1 to 137 individuals day⁻¹, except for 2 traps from which no individuals were captured (Table [1](#page-5-1)). Percent richness and abundance of hypoxia-tolerant species varied between 0 and 100%. Median standard length was 3.2 cm (minimum: 2.8; maximum: 7.20 cm), and mean SL range was c.a. 3.9 cm (minimum: 0.0; maximum: 5.4). Aquatic vegetation density, our proxy for habitat structural complexity, varied from 0 to 2.5 10^{-3} Kg DW L⁻¹. Dissolved oxygen concentration varied from nearly 0 to 6 mg L^{-1} .

Fig. 1 Relative abundance (diamonds) of fish species at survey locations across a gradient of aquatic vegetation density. Reciprocal averaging was used for ranking species and sites in order to emphasize the gradient changes in species composition among sites. Arrows in the y-axis indicate direction of changes in vegetation density

ordered species																				
																				sites ordered
Erythrinus erythrinus Hoplerythrinus unitaeniatus	Serrapinnus sp. 2 Acestrorhynchus lacustris	Serrapinnus notomelas Satanoperca sp.	Leporinus lacustris	Serrapinnus heterodon Cichlasoma paranaense	Cichla kelberi	Moenkhausia sanctaefilomenae	Hoplias spp.	Aphyocharax anisitsi	Roeboides descalvadensis	Crenicichla britskii	Psellogrammus kennedyi	Apistogramma commbrae	Hyphessobrycon eques	Laetacara araguaiae	Moenkhausia bonita	Characidium sp.	Serrasalmus marginatus	Pyrrhulina australis	Astyanax lacustris Species relative abundance	

Table 1 Fish assemblage attributes and habitat environmental variables measured to investigate α and β diversity in relation to a gradient of aquatic vegetation density

Measurements are summarized according to quartiles (minimum, first quartile, median, third quartile and maximum values)

Additive partitioning of species diversity

As expected, patterns of both α and β diversity differed significantly from random ($p < 0.001$ for both α and β diversity; Fig. [2a](#page-6-0)), suggesting that one or more non-random mechanisms drive diversity patterns. Whereas α diversity was lower than expected at random $(8.1 \text{ vs. } 10.6 \text{ species})$, β diversity was greater (18.9 vs. 16.4 species). Decomposition of the β diversity according to spatial scales revealed significant differences at trap $(p = 0.001)$ and transect scales ($p = 0.003$) but not among lakes ($p = 0.43$), indicating that variation in β diversity was mostly associated with small spatial scales.

α diversity and explanatory variable associations

The best model for fish α diversity (Δ AIC_C < 2, ω_i = 0.69) explained 46% of the variation (conditional $R^2 = 0.46$), with fixed effects accounting for 30% of the variation (marginal $R² = 0.30$ $R² = 0.30$ $R² = 0.30$; Fig. 3). Only vegetation density was included in the best model, with diversity having a significant hump-shaped response to vegetation density ($\chi^2 = 21.46$; $p < 0.001$; Fig. [3](#page-6-1)).

β diversity and explanatory variable associations

Variation in total β diversity was most strongly related to vegetation density, [vegetation density] 2 and oxygen

Fig. 2 Additive partitioning of α and β diversity of fishes inhabiting vegetation stands. Observed α and β are contrasted with expected values obtained by randomizations. Analyses were performed with (**a**) and without considering nested sampling structure (**b**) (i.e., traps and transects within lakes). *Significant differences from the null model

Fig. 3 Partial residuals of the relationship between fish α diversity and vegetation density for fishes inhabiting vegetation stands

content ($F = 2.39$; $p < 0.001$), suggesting that habitat structural complexity and dissolved oxygen concentration regulate spatial variation in species occurrence (Fig. [4](#page-6-2)a). Analysis of individual components of β diversity showed that species turnover was most strongly correlated with vegetation density and [vegetation density]² ($F = 2.53$; $p < 0.001$). Complementary to this result, LCBD_{repl} values tended to increase towards lower and upper limits of vegetation density, suggesting that turnover rates are even greater towards the highest and the lowest levels of habitat structural complexity (Fig. [4b](#page-6-2)). For the nestedness

Fig. 4 Distance-based Redundancy Analyses (dbRDA) assessing the relationship of variation in aspects of fish β diversity with vegetation density and dissolved oxygen concentration. **a** Total β diversity, **b** species turnover, **c** nestedness, VegDen vegetation density, VegDen² [vegetation density]²

component of β diversity, dissolved oxygen was the only correlated variable ($F=1.95$; $p=0.04$; Fig. [4](#page-6-2)c).

Biological features

Vegetation density was significantly and negatively correlated with fish standard length (χ^2 = 35.72; p < 0.001; Fig. [5a](#page-7-0)), explaining 44% of variation (conditional $R^2 =$

Fig. 5 Relationships of the median (**a**) and the range (**b**) of fish standard length with vegetation density for fishes inhabiting vegeation stands **Fig. ⁶**Relationships between the percentage of abundance (**a**) and

0.48; marginal $R^2 = 0.44$). Vegetation density also significantly correlated with the variation in fish standard length, but with the variation in fish standard length having a significant hump-shaped response to vegetation density (χ^2 = 6.35; p = 0.01; conditional R^2 = 0.47; marginal $R^2 = 0.11$; Fig. [5](#page-7-0)b). Together, fish standard length and variation in fish standard length suggests that the size of interstitial spaces within highly complex habitats constrains fish body sizes.

The percentage of fish specimens in assemblages that were categorized as hypoxia-tolerant was negatively correlated with dissolved oxygen concentration (χ^2 = 6.84; $p < 0.001$; conditional $R^2 = 0.23$; marginal $R^2 = 0.23$; Fig. [6](#page-7-1)a). Similarly, the percentage of fish species categorized as hypoxia-tolerant was negatively correlated with oxygen concentration (χ^2 = 3.90; p = 0.04; conditional R^2 $= 0.19$; marginal $R² = 0.16$; Fig. [6](#page-7-1)b). The fact that relative abundance and richness of hypoxia-tolerant fishes were greater when dissolved oxygen was lower strongly implies that dissolved oxygen gradients influenced fish distributions and thereby function as a strong environmental filter during local community assembly.

percentage of species richness (**b**) of hypoxia-tolerant fish species within a location with dissolved oxygen concentration (mg L^{-1}) in aquatic vegetation stands

Discussion

Habitat structural complexity has been recognized as an environmental factor that regulates species α diversity, but its relationship with β diversity is poorly explored. Our results on additive partitioning of fish species diversity indicates diversity components significantly differ from random across gradients of habitat structural complexity (Fig. [2](#page-6-0)), suggesting that deterministic mechanisms might be responsible for regulating both α and β (Chase et al. [2011](#page-9-22)). In addition, our findings demonstrate that both α and β components of fish diversity were strongly associated with a gradient of vegetation density (Figs. [1](#page-5-0), [2,](#page-6-0) [3\)](#page-6-1). Whereas α diversity tended to have highest values at intermediate levels of vegetation density, β diversity (particularly species turnover) was significant across the entire gradient (Figs. [2](#page-6-0), [3,](#page-6-1) [4,](#page-6-2) [5](#page-7-0)). Such findings provide empirical support for the predicted hypothesis that multiple components of species diversity vary in relation to gradients of vegetation complexity in floodplain systems, and habitat structural complexity influences local community assembly.

Results for additive partitioning of diversity indicate that fish α diversity was lower than expected by chance, whereas $β$ diversity was greater, suggesting that fish diversity across structural complexity gradients was strongly associated with species-specific responses to environmental conditions. Higher β and lower α diversity than expected by chance can indicate intraspecific aggregation within habitat patches (Crist et al. [2003](#page-9-17); Fahr and Kalko [2011;](#page-9-6) Myers et al. [2013\)](#page-10-29), resulting in clumped species turnover along environmental gradients. Our surveys focused on a gradient of vegetation density, and we infer that species turnover was strongly influenced by levels of structural complexity. Intraspecific aggregations have been reported for Pacific reef fishes and inferred to be associated with resource partitioning and restricted availability of physical habitat (Rodríguez-Zaragoza et al. [2011\)](#page-10-30), mechanisms that also may apply to small fishes that inhabit structurally complex habitats in tropical floodplain systems (Arrington et al. [2005](#page-9-7); Willis et al. [2005](#page-11-8)).

In agreement with additive partitioning of diversity, we found empirical support for species turnover along the gradient of vegetation density. The gradient of vegetation density ranges from sparse stands with relatively few submerged stems and roots and large interstitial spaces, to dense stands with high structural complexity and numerous small interstices. Large species generally require larger open spaces and would be excluded from dense stands of aquatic vegetation (Fahr and Kalko [2011;](#page-9-6) Yeager and Hovel [2017](#page-11-6)). Accordingly, our analysis of fish size revealed a significant reduction in fish size with increasing vegetation density, and only very small characids, such as *Serrapinus notomelas* and *Serrapinnus heterodon*, were captured from areas of dense vegetation (Fig. [1\)](#page-5-0). Similarly, a recent study conducted in eelgrass habitat demonstrated that structural complexity regulates the size distribution of juvenile giant kelpfish, inferring that dense vegetation constrains the foraging efficiency of large fish (Yeager and Hovel [2017](#page-11-6)).

Another potential mechanism contributing to species turnover along gradients of habitat structural complexity is interspecific differences in susceptibility to predation. Predation has been shown to be a strong regulator of fish assemblage structure in freshwater systems (Jackson and Harvey [1989;](#page-10-31) Layman and Winemiller [2004\)](#page-10-32), and predation mortality generally is greater when habitat structural complexity is low (Gause [1936;](#page-9-1) Huffaker [1958;](#page-10-6) Rozas and Odum [1988](#page-10-14); Santos et al. [2009](#page-10-33)). Thus, species turnover along gradients of habitat structural complexity could indicate that predationmediated colonization and extinction of patches is selective and species specific. For example, the characid *Moenkhausia bonita* has a relatively elongated body that should enhance swimming speed but reduce lateral maneuverability. In areas with sparse vegetation with large spaces of open water, this morphology likely enhances predator escape by fleeing rather than hiding. The idea that habitat structural complexity differentially influences species vulnerability to predation is consistent with Poff's [\(1997](#page-10-34)) assertion that habitat structural complexity is an important environmental filter that sorts species according to functional traits and associated survival strategies (Poff [1997](#page-10-34)).

We also found that α diversity attained a maximum at intermediate levels of vegetation density, suggesting tradeoffs among mechanisms influencing species distributions along a structural complexity gradient. Theoretical models predict that α diversity of a given taxonomic group responds to environmental gradients according to a hump-shaped function, with each species being more or less normally distributed over a limited interval of the gradient (Crowder and Cooper [1979](#page-9-3); Diehl and Kornijow [1998](#page-9-4); Tokeshi and Arakaki [2012\)](#page-11-4). This hump-shaped function can derive from the interaction of mechanisms that facilitate colonization and persistence in habitats (e.g., connectivity, availability of refuges and resources) and those that hamper them (e.g., isolation, space limitation, competition, predation). Structural complexity has been shown to reduce exposure to predators and enhance α diversity (Rozas and Odum [1988;](#page-10-14) Sánchez-Botero et al. [2007\)](#page-10-7), however, excessive habitat structural complexity may limit space available for movement and foraging (Yeager and Hovel [2017](#page-11-6)). A recent study of reef fishes (Paxton et al. [2017](#page-10-9)) also found a hump-shaped relationship of α diversity along a gradient of structural complexity, suggesting that this pattern could be common in freshwater and marine systems.

Contrary to our prediction, species nestedness was not significantly associated with vegetation density, although it was significantly associated with dissolved oxygen concentration. Our fish surveys of aquatic vegetation stands included samples collected at night, when hypoxia may occur in dense patches, as well as samples taken during daytime when the water column tended to be well oxygenated. This allowed us to test for separate effects of vegetation density and oxygen concentration, two factors that have been found to influence fish abundance (Miranda and Hodges [2000\)](#page-10-17). Greater assemblage nestedness and occurrence of hypoxia-tolerant fishes were associated with low dissolved oxygen, which suggests that dissolved oxygen functions as an environmental filter in concert with structural complexity of aquatic habitats (Keddy [1992;](#page-10-35) Poff [1997\)](#page-10-34). Adaptation for tolerance of hypoxic conditions is relatively common among fishes inhabiting tropical floodplains. In our samples, for example, characids with auxiliary respiratory adaptations, such as *Aphyocharax anisitsi* (presence of dermal lip protuberances) and *Hoplias* spp. (anaerobic respiration), contributed to high proportional abundance and α diversity of hypoxia-tolerant species in dense vegetation. Periodic occurrence of extreme hypoxic conditions has been invoked as a major determinant of fish spatial distributions and local

population dynamics in tropical and subtropical floodplains (Lowe-McConnell [1964;](#page-10-36) Winemiller [1989;](#page-11-15) Miranda and Hodges [2000;](#page-10-17) Miranda et al. [2000\)](#page-10-18), and the present results support the importance of low dissolved oxygen concentration as an environmental filter structuring fish communities (De Macedo-Soares et al. [2010;](#page-9-23) Arantes et al. [2018](#page-9-24)).

In summary, we conclude that the density and associated structural complexity of aquatic vegetation stands in tropical floodplain lakes affects multiple diversity components of fish assemblages. Our findings provide empirical evidence that intermediate levels of vegetation density maximize α diversity of fish assemblages, suggesting tradeoffs among structuring mechanisms along the habitat structural complexity gradient. High β diversity along the gradient of vegetation density reveals species turnover and suggests species-specific responses to levels of structural complexity and other environmental factors. We obtained evidence that dissolved oxygen concentration is another significant environmental filter structuring these fish metacommunities, and infer that species colonization and persistence at locations along environmental gradients are likely influenced by multiple factors and mechanisms that operate on different spatiotemporal scales.

Acknowledgements We thank the Coordenação de Aperfeiçoamento de Pessoal de Nível Superior (CAPES/Proex) for their financial support. ERC thanks the Itaipu Binacional and Fundação Araucária (an organization of the Government of Paraná state, Brazil) for providing scholarships. SMT, AAA and LCG thanks the Brazilian Council of Research (CNPq) for continuous funding through a Research Productivity Grant.

References

 $\circled{2}$ Springer

- Agostinho AA, Thomaz SM, Minte-Vera CV, Winemiller KO (2000) Biodiversity in the high Paraná River floodplain. In: Gopal B, Junk WJ, Davis JA (eds) Biodiversity in wetlands: assessment, function and conservation. Backhuys Publishers, Leiden, pp 89–118
- Agostinho AA, Bini LM, Gomes LC, Júlio HF Jr, Pavanelli CS, Agostinho CA (2004a) Fish assemblages. In: Thomaz SM, Agostinho AA, Hahn NS (eds) The Upper Paraná River and its floodplain physical aspects, ecology and conservation. Backhuys Publishers, Leiden, pp 223–246
- Agostinho AA, Gomes LC, Veríssimo S, Okada EK (2004b) Flood regime, dam regulation and fish in the Upper Paraná River: effects on assemblage attributes, reproduction and recruitment. Rev Fish Biol Fish 14:11–19.<https://doi.org/10.1007/s11160-004-3551-y>
- Agostinho AA, Thomaz SM, Gomes LC (2004c) Threats for biodiversity in the floodplain of the Upper Paraná River: effects of hydrological regulation by dams. Ecohydrol Hydrobiol 4:255–256
- Agostinho AA, Pelicice FM, Petry AC, Gomes LC, Júlio HF Jr (2007a) Fish diversity in the upper Paraná River basin: habitats, fisheries, management and conservation. Aquat Ecosyst Heal Manag 10:174–186.<https://doi.org/10.1080/14634980701341719>
- Agostinho AA, Thomaz SM, Gomes LC, Baltar SLSMA (2007b) Influence of the macrophyte *Eichhornia azurea* on fish assemblage of the Upper Paraná River floodplain (Brazil). Aquat Ecol 41:611– 619.<https://doi.org/10.1007/s10452-007-9122-2>
- Arantes CC, Winemiller KO, Petrere M, Castello L, Hess LL, Freitas CE (2018) Relationships between forest cover and fish diversity in the Amazon River floodplain. J Appl Ecol 55:386–395. [https](https://doi.org/10.1111/1365-2664.12967) [://doi.org/10.1111/1365-2664.12967](https://doi.org/10.1111/1365-2664.12967)
- Arrington DA, Winemiller KO, Layman CA (2005) Community assembly at the patch scale in a species rich tropical river. Oecologia 144:157–167.<https://doi.org/10.1007/s00442-005-0014-7>
- August PV (1983) The role of habitat complexity and heterogeneity in structuring tropical mammal communities. Ecology 64:1495– 1507.<https://doi.org/10.2307/1937504>
- Bartón K (2018) MuMIn: Multi-model inference. R package version 1.42.1. <http://CRAN.R-project.org/package=MuMIn>
- Baselga A (2010) Partitioning the turnover and nestedness components of beta diversity. Global Ecol Biogeogr 19:134–143. [https](https://doi.org/10.1111/j.1466-8238.2009.00490.x) [://doi.org/10.1111/j.1466-8238.2009.00490.x](https://doi.org/10.1111/j.1466-8238.2009.00490.x)
- Bates D, Maechler M, Bolker B, Walker S (2014) lme4: linear mixed-effects models using Eigen and S4. R package version $1(7):1-23$
- Burnham KP, Anderson DR (2002) Model selection and multimodel inference: a practical information—theoretic approach. Springer, New York, 454 p
- Chase JM, Kraft NJB, Smith KG, Vellend M, Inouye BD (2011) Using null models to disentangle variation in community dissimilarity from variation in α-diversity. Ecosphere 2:1–11. [https://doi.](https://doi.org/10.1890/ES10-00117.1) [org/10.1890/ES10-00117.1](https://doi.org/10.1890/ES10-00117.1)
- Crist TO, Veech JA, Gering JC, Summerville KS (2003) Partitioning species diversity across landscapes and regions: a hierarchical analysis of α, $β$, and $γ$ diversity. Am Nat 162:734–743
- Crowder LB, Cooper WE (1979) Structural complexity and fish-prey interactions in ponds: a point of view. In: Johson DL, Stein RA (eds) Response of fish to habitat structure in standing waters. North Central Division American Fisheries Society Special publication, Ohio, pp 2–10
- De Macedo-Soares PHM, Petry AC, Farjalla VF, Caramaschi EP (2010) Hydrological connectivity in coastal inland systems: lessons from a Neotropical fish metacommunity. Ecol Freshw Fish 19:7–18. <https://doi.org/10.1111/j.1600-0633.2009.00384.x>
- Delariva RL, Agostinho AA, Nakatani K, Baumgartner G, (1994) Ichthyofauna associated to aquatic macrophytes in the upper Parana river floodplain. Rev Unimar 16:41–60
- Dibble ED, Pelicice FM (2010) Influence of aquatic plant-specific habitat on an assemblage of small neotropical floodplain fishes. Ecol Freshw Fish 19:381–389. [https://doi.org/10.111](https://doi.org/10.1111/j.1600-0633.2010.00420.x) [1/j.1600-0633.2010.00420.x](https://doi.org/10.1111/j.1600-0633.2010.00420.x)
- Diehl S, Kornijow R (1998) Influence of submerged macrophytes on trophic interactions among fish and macroinvertebrates. In: Jeppesen E, Sondergaard M, Christoffersen K (eds) Structuring role of subbmerged macrophytes in lakes. Springer, New York, pp 24–46
- Fahr J, Kalko EKV (2011) Biome transitions as centres of diversity: habitat heterogeneity and diversity patterns of West African bat assemblages across spatial scales. Ecography 34:177–195. [https](https://doi.org/10.1111/j.1600-0587.2010.05510.x) [://doi.org/10.1111/j.1600-0587.2010.05510.x](https://doi.org/10.1111/j.1600-0587.2010.05510.x)
- Fernandes R, Gomes LC, Pelicice FM, Agostinho AA (2009) Temporal organization of fish assemblages in floodplain lagoons: the role of hydrological connectivity. Environ Biol Fishes 85:99–108. [https](https://doi.org/10.1007/s10641-009-9466-7) [://doi.org/10.1007/s10641-009-9466-7](https://doi.org/10.1007/s10641-009-9466-7)
- Fukami T (2004) Community assembly along a species pool gradient: implications for multiple-scale patterns of species diversity. Popul Ecol 46:137–147.<https://doi.org/10.1007/s10144-004-0182-z>
- Gause GF (1936) The struggle for existence. Soil Sci 41:159. [https://](https://doi.org/10.1097/00010694-193602000-00018) doi.org/10.1097/00010694-193602000-00018
- Gibb H, Parr CL (2010) How does habitat complexity affect ant foraging success? A test using functional measures on three continents. Oecologia 164:1061–1073. [https://doi.org/10.1007/s0044](https://doi.org/10.1007/s00442-010-1703-4) [2-010-1703-4](https://doi.org/10.1007/s00442-010-1703-4)
- Gomes LC, Bulla CK, Agostinho AA, Vasconcelos LP, Miranda LE (2012) Fish assemblage dynamics in a neotropical floodplain relative to aquatic macrophytes and the homogenizing effect of a flood pulse. Hydrobiologia 685:97–107. [https://doi.org/10.1007/s1075](https://doi.org/10.1007/s10750-011-0870-6) [0-011-0870-6](https://doi.org/10.1007/s10750-011-0870-6)
- Graça WJ, Pavanelli CS (2007) Peixes da planície de inundação do alto rio Paraná e áreas adjacentes. EDUEM, Maringá
- Heck KL Jr, Orth RJ (1980) Seagrass habitats: the roles of habitat complexity, competition and predation in the structuring associated fish and motile macroinvertebrate assemblages. In: Kennedy VS (ed) Estuarine perspectives. Academic press, New York, pp 449–464
- Huffaker C (1958) Experimental studies on predation: dispersion factors and predator-prey oscillations. Hilgardia 27:343–383
- Hylander K, Nilsson C, Jonsson BG, Göthner T (2005) Differences in habitat quality explain nestedness in a land snail meta-community. Oikos. <https://doi.org/10.1111/j.0030-1299.2005.13400.x>
- Jackson DA, Harvey HH (1989) Biogeographic associations in fish assemblages: local vs. regional processes. Ecology 70:1472–1484
- Keddy PA (1992) Assembly and response rules: two goals for predictive community ecology. J Veg Sci 3:157–164. [https://doi.](https://doi.org/10.2307/3235676) [org/10.2307/3235676](https://doi.org/10.2307/3235676)
- Kovalenko KE, Thomaz SM, Warfe DM (2012) Habitat complexity: approaches and future directions. Hydrobiologia 685:1–17. [https](https://doi.org/10.1007/s10750-011-0974-z) [://doi.org/10.1007/s10750-011-0974-z](https://doi.org/10.1007/s10750-011-0974-z)
- Kraft NJB, Comita LS, Chase JM, Sanders NJ, Swenson NG, Crist TO, Stegen JC, Vellend M, Boyle B, Anderson MJ, Cornell HV (2011) Disentangling the drivers of β diversity along latitudinal and elevational gradients. Science 333:1755–1758. [https://doi.](https://doi.org/10.1126/science.1208584) [org/10.1126/science.1208584](https://doi.org/10.1126/science.1208584)
- Layman CA, Winemiller KO (2004) Size-based responses of prey to piscivore exclusion in a species-rich neotropical river. Ecology 85:1311–1320. <https://doi.org/10.1890/02-0758>
- Legendre P (2014) Interpreting the replacement and richness difference components of beta diversity. Glob Ecol Biogeogr 23:1324–1334. <https://doi.org/10.1111/geb.12207>
- Legendre P, De Caceres M (2013) Beta diversity as the variance of community data: dissimilarity coefficients and partitioning. Ecol Lett 16:951–963.<https://doi.org/10.1111/ele.12141>
- Leibold MA, Mikkelson GM (2002) Coherence, species turnover, and boundary clumping: elements of meta-community structure. Oikos 97:237–250.<https://doi.org/10.1034/j.1600-0706.2002.970210.x>
- Leibold MA, Holyoak M, Mouquet N et al (2004) The metacommunity concept: a framework for multi-scale community ecology. Ecol Lett 7:601–613. [https://doi.org/10.1111/j.1461-0248.2004.00608](https://doi.org/10.1111/j.1461-0248.2004.00608.x) [.x](https://doi.org/10.1111/j.1461-0248.2004.00608.x)
- Lopes TM, Cunha ER, Silva JCB, Behrend RD, Gomes LC (2015) Dense macrophytes influence the horizontal distribution of fish in floodplain lakes. Environ Biol Fishes 98:1741–1755. [https://doi.](https://doi.org/10.1007/s10641-015-0394-4) [org/10.1007/s10641-015-0394-4](https://doi.org/10.1007/s10641-015-0394-4)
- Lowe-McConnell RH (1964) The fishes of the Rupununi savanna district of British Guiana, South America. Part 1. Ecological groupings of fish species and effects of the seasonal cycle on the fish. J Linn Soc 45:103–144
- MacArthur R, MacArthur JW (1961) On bird species-diversity. Ecology 42:594–598.<https://doi.org/10.2307/1932254>
- McCoy ED, Bell SS (1991) Habitat structure: the evolution and diversification of a complex topic. In: Bell SS, McCoy ED, Mushinsky HR (eds) Habitat structure: the physical arrangement of objects in space. Chapman and Hall, London, pp 3–27
- Millar RB, Anderson MJ (2004) Remedies for pseudoreplication. Fish Res 70:397–407.<https://doi.org/10.1016/j.fishres.2004.08.016>
- Miranda LE, Hodges KB (2000) Role of aquatic vegetation coverage on hypoxia and sunfish abundance in bays of a eutrophic reservoir. Hydrobiologia 427:51–57. [https://doi.org/10.1023/a:10039](https://doi.org/10.1023/a:1003999929094) [99929094](https://doi.org/10.1023/a:1003999929094)
- Miranda LE, Driscoll MP, Allen MS (2000) Transient physicochemical microhabitats facilitate fish survival in inhospitable aquatic plant stands. Freshw Biol 44:617–628. [https://doi.org/10.104](https://doi.org/10.1046/j.1365-2427.2000.00606.x) [6/j.1365-2427.2000.00606.x](https://doi.org/10.1046/j.1365-2427.2000.00606.x)
- Mittelbach GG, Schemske DW (2015) Ecological and evolutionary perspectives on community assembly. Trends Ecol Evol 30:241–247. <https://doi.org/10.1016/j.tree.2015.02.008>
- Myers JA, Chase JM, Jiménez I, Jørgensen PM, Araujo-Murakami A, Paniagua-Zambrana N, Seidel R (2013) Beta-diversity in temperate and tropical forests reflects dissimilar mechanisms of community assembly. Ecol Lett 16:151–157. [https://doi.org/10.1111/](https://doi.org/10.1111/ele.12021) [ele.12021](https://doi.org/10.1111/ele.12021)
- Newman SP, Meesters EH, Dryden CS, Williams SM, Sanchez C, Mumby PJ, Polunin NV (2015) Reef flattening effects on total richness and species responses in the Caribbean. J Anim Ecol 84:1678–1689.<https://doi.org/10.1111/1365-2656.12429>
- Oksanen J, Blanchet FG, Friendly M, Kindt R, Legendre P, McGlinn D, Minchin PR, O'Hara RB, Simpson GL, Solymos P, Stevens MHH, Szoecs E, Wagner H (2019) vegan: Community Ecology Package. R package version 2.5-4. [http://CRAN.R-project.org/](http://CRAN.R-project.org/package=vegan) [package=vegan](http://CRAN.R-project.org/package=vegan)
- Orfeo O, Stevaux J (2002) Hydraulic and morphological characteristics of middle and upper reaches of the Parana River (Argentina and Brazil). Geomorphology 44:309–322. [https://doi.org/10.1016/](https://doi.org/10.1016/s0169-555x(01)00180-5) [s0169-555x\(01\)00180-5](https://doi.org/10.1016/s0169-555x(01)00180-5)
- Ota RR, Deprá GD, Graça WJ, Pavanelli CS (2018) Peixes da planície de inundação do alto rio Paraná e áreas adjacentes: revised, annotated and updated. Neotrop Ichthyol 16:e170094. [https://doi.](https://doi.org/10.1590/1982-0224-20170094) [org/10.1590/1982-0224-20170094](https://doi.org/10.1590/1982-0224-20170094)
- Paxton AB, Pickering EA, Adler AM, Taylor JC, Peterson CH (2017) Flat and complex temperate reefs provide similar support for fish: evidence for a unimodal species-habitat relationship. PLoS One 12:1–22. <https://doi.org/10.1371/journal.pone.0183906>
- Petsch DK, Schneck F, Melo AS (2017) Substratum simplification reduces beta diversity of stream algal communities. Freshw Biol 62:205–213.<https://doi.org/10.1111/fwb.12863>
- Podani J, Ricotta C, Schmera D (2013) A general framework for analyzing beta diversity, nestedness and related community-level phenomena based on abundance data. Ecol Complex 15:52–61. <https://doi.org/10.1016/j.ecocom.2013.03.002>
- Poff NL (1997) Landscape filters and species traits: towards mechanistic understanding and prediction in stream ecology. J N Am Benthol Soc 16:391–409
- Ribeiro OM, Zuanon J (2006) Comparação da eficiência de dois métodos de coleta de peixes em igarapés de terra firme da Amazônia Central. Acta Amaz 36:389–394. [https://doi.org/10.1590/s0044](https://doi.org/10.1590/s0044-59672006000300017) [-59672006000300017](https://doi.org/10.1590/s0044-59672006000300017)
- Rodríguez-Zaragoza FA, Cupul-Magaña AL, Galván-Villa CM et al (2011) Additive partitioning of reef fish diversity variation: a promising marine biodiversity management tool. Biodivers Conserv 20:1655–1675
- Rossier O, Castella E, Lachavanne JB (1996) Influence of submerged aquatic vegetation on size class distribution of perch (*Perca fluviatilis*) and roach (*Rutilus rutilus*) in the littoral zone of Lake Geneva (Switzerland). Aquat Sci 58:1–14. [https://doi.org/10.1007/](https://doi.org/10.1007/BF00877636) [BF00877636](https://doi.org/10.1007/BF00877636)
- Rozas LP, Odum WE (1988) Occupation of submerged aquatic vegetation by fishes—testing the roles of food and refuge. Oecologia 77:101–106.<https://doi.org/10.1007/bf00380932>
- Sánchez-Botero JI, Leitão RP, Caramaschi EP, Garcez DS (2007) The aquatic macrophytes as refuge, nursery and feeding habitats for freshwater fish from Cabiúnas Lagoon, Restinga de Jurubatiba National Park, Rio de Janeiro, Brazil. Acta Limnol Bras 19:143–153
- Santos AFGN, Santos LN, García-Berthou E, Hayashi C (2009) Could native predators help to control invasive fishes? Microcosm

experiments with the neotropical characid, *Brycon orbignyanus*. Ecol Freshw Fish 18:491–499. [https://doi.org/10.111](https://doi.org/10.1111/j.1600-0633.2009.00366.x) [1/j.1600-0633.2009.00366.x](https://doi.org/10.1111/j.1600-0633.2009.00366.x)

- Santos LN, Agostinho AA, Alcaraz C, Carol J, Santos AF, Tedesco P, García-Berthou E (2011) Artificial macrophytes as fish habitat in a Mediterranean reservoir subjected to seasonal water level disturbances. Aquat Sci 73:43–52. [https://doi.org/10.1007/s0002](https://doi.org/10.1007/s00027-010-0158-3) [7-010-0158-3](https://doi.org/10.1007/s00027-010-0158-3)
- Schmera D, Podani J (2011) Comments on separating components of beta diversity. Comm Ecol 12:153–160. [https://doi.org/10.1556/](https://doi.org/10.1556/ComEc.12.2011.2.2) [ComEc.12.2011.2.2](https://doi.org/10.1556/ComEc.12.2011.2.2)
- St. Pierre JI, Kovalenko KE (2014) Effect of habitat complexity attributes on species richness. Ecosphere 5:art22. [https://doi.](https://doi.org/10.1890/ES13-00323.1) [org/10.1890/ES13-00323.1](https://doi.org/10.1890/ES13-00323.1)
- Stein A, Kreft H (2015) Terminology and quantification of environmental heterogeneity in species-richness research. Biol Rev 90:815–836.<https://doi.org/10.1111/brv.12135>
- Stevaux JC (1994) The upper Paraná river (Brazil): geomorphology, sedimentology and paleoclimatology. Quat Int 21:143-161. [https](https://doi.org/10.1016/1040-6182(94)90028-0) [://doi.org/10.1016/1040-6182\(94\)90028-0](https://doi.org/10.1016/1040-6182(94)90028-0)
- Stevaux JC, Souza IA (2004) Floodplain construction in an anastomosed river. Quat Int 114:55–65. [https://doi.org/10.1016/s1040](https://doi.org/10.1016/s1040-6182(03)00042-9) [-6182\(03\)00042-9](https://doi.org/10.1016/s1040-6182(03)00042-9)
- Strayer DL, Findlay SEG (2010) Ecology of freshwater shore zones. Aquat Sci 72:127–163. [https://doi.org/10.1007/s0002](https://doi.org/10.1007/s00027-010-0128-9) [7-010-0128-9](https://doi.org/10.1007/s00027-010-0128-9)
- Tews J, Brose U, Grimm V, Tielbörger K, Wichmann MC, Schwager M, Jeltsch F (2004) Animal species diversity driven by habitat heterogeneity/diversity: the importance of keystone structures. J Biogeogr 31:79–92
- Tokeshi M, Arakaki S (2012) Habitat complexity in aquatic systems: fractals and beyond. Hydrobiologia 685:27–47. [https://doi.](https://doi.org/10.1007/s10750-011-0832-z) [org/10.1007/s10750-011-0832-z](https://doi.org/10.1007/s10750-011-0832-z)
- Whittaker RH (1960) Vegetation of the Siskiyou Mountains, Oregon and California. Ecol Monogr 30:279–338
- Whittaker RH (1972) Evolution and measurement of species diversity. Taxon 1:213–251
- Willis SC, Winemiller KO, Lopez-Fernandez H (2005) Habitat structural complexity and morphological diversity of fish assemblages in a neotropical floodplain river. Oecologia 142:284–295. [https://](https://doi.org/10.1007/s00442-004-1723-z) doi.org/10.1007/s00442-004-1723-z
- Winemiller KO (1989) Development of dermal lip protuberances for aquatic surface respiration in South American characid fishes. Copeia 1989:382–390.<https://doi.org/10.2307/1445434>
- Winkler AM, Webster MA, Vidaurre D, Nichols TE, Smith SM (2015) Multi-level block permutation. Neuroimage 123:253–268. [https://](https://doi.org/10.1016/j.neuroimage.2015.05.092) doi.org/10.1016/j.neuroimage.2015.05.092
- Yeager ME, Hovel KA (2017) Structural complexity and fish body size interactively affect habitat optimality. Oecologia 185:257–267. <https://doi.org/10.1007/s00442-017-3932-2>
- Zuur AF, Ieno EN, Elphick CS (2010) A protocol for data exploration to avoid common statistical problems. Methods Ecol Evol 1:3–14. <https://doi.org/10.1111/j.2041-210X.2009.00001.x>

Publisher's Note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.