REVIEW

Role of peroxisome proliferator‑activated receptors in non‑alcoholic fatty liver disease infammation

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Abstract

Overweight and obesity have been identifed as the most important risk factors for many diseases, including cardiovascular disease, type 2 diabetes and lipid disorders, such as non-alcoholic fatty liver disease (NAFLD). The metabolic changes associated with obesity are grouped to defne metabolic syndrome, which is one of the main causes of morbidity and mortality in industrialized countries. NAFLD is considered to be the hepatic manifestation of metabolic syndrome and is one of the most prevalent liver diseases worldwide. Infammation plays an important role in the development of numerous liver diseases, contributing to the progression to more severe stages, such as non-alcoholic steatohepatitis and hepatocellular carcinoma. Peroxisome proliferator-activated receptors (PPARs) are binder-activated nuclear receptors that are involved in the transcriptional regulation of lipid metabolism, energy balance, infammation and atherosclerosis. Three isotypes are known: PPAR- α , PPARδ/β and PPAR- γ . These isotypes play different roles in diverse tissues and cells, including the inflammatory process. In this review, we discuss current knowledge on the role PPARs in the hepatic infammatory process involved in NAFLD as well as new pharmacological strategies that target PPARs.

Keywords NAFLD · Infammation · PPARs

Introduction

Non-alcoholic fatty liver disease (NAFLD) is characterized by signifcant lipid deposition in hepatocytes with persistent changes in liver enzymes, such as AST and ALT [[1](#page-7-0)]. Overeating is the most common cause of the accumulation of excess lipids in the liver and it is estimated that over six hundred million overweight people worldwide will develop fatty liver disease [\[2](#page-7-1)]. In clinical practice, the initial diagnosis of hepatic steatosis is usually established using radiological imaging techniques and is based on the presence of liver

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fat accumulation $\geq 5\%$ in the absence of other recognized causes, such as alcohol, virus and medication [[3\]](#page-7-2).

NAFLD comprises a broad spectrum of liver damage, ranging from simple macrovesicular steatosis to non-alcoholic steatohepatitis (NASH), advanced fibrosis and cirrhosis [[4](#page-7-3), [5](#page-7-4)]. NAFLD is recognized as a cause of end-stage liver disease and is associated with increased rates of hepatocellular carcinoma, liver transplantation and death [\[6](#page-7-5)[–8](#page-7-6)]. Although not yet well understood, it is recognized that insulin resistance plays an important role in the pathogenesis of NAFLD. In healthy subjects, insulin stimulates hepatic and peripheral glucose absorption and suppresses hepatic glucose production [\[9](#page-7-7)]. In the fasting state, the liver becomes the main site of glucose production mediated by glycogenolysis and gluconeogenesis [[10,](#page-7-8) [11\]](#page-7-9). In patients with insulin resistance, hepatic autoregulation is altered and both gluconeogenesis and glycogenolysis are increased, resulting in the development of hyperglycemia [\[12](#page-7-10)].

According to the "multiple hit" theory, hyperinsulinemia in the context of insulin resistance frst leads to an increase in the release of free fatty acids from adipocytes, which then accumulate in the liver, resulting in steatosis. This initial step is followed by a series of complex interactions among hepatocytes,

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Kupffer cells, adipocytes, inflammatory mediators and oxygen radicals, the result of which is non-alcoholic steatohepatitis.

Infammation plays a central role in several acute and chronic liver diseases, contributing to the progression of liver damage to more severe stages, such as fbrosis, cirrhosis and hepatocellular carcinoma [[13](#page-7-11), [14](#page-7-12)]. In addition to hepatocytes, the liver also contains a complex repertoire of lymphoid and non-lymphoid cells that play key roles in hepatic and defense immunoregulation [[15](#page-7-13)]. The cells that are part of innate immunity in the liver include Kupfer cells, monocytes, neutrophils, dendritic cells, natural killer cells and natural killer T cells, which initiate and maintain hepatic infammation through the production of cytokines [\[16](#page-7-14)]. The dysregulated release of cytokines following liver damage may result in the excessive death of hepatocytes, which is a key event in several acute and chronic liver diseases [[17\]](#page-7-15).

Peroxisome proliferator-activated receptors (PPARs) are nuclear receptors that are involved in the transcriptional regulation of lipid metabolism, energy balance, infammation and atherosclerosis. The three known isotypes are PPAR- α , PPAR-δ/β and PPAR-γ [[18,](#page-7-16) [19](#page-7-17)]. PPAR- α is the most commonly expressed in the liver and expressed in lower levels in the heart, skeletal muscle, intestine and kidneys, where it plays an important role in controlling the oxidation of fatty acids $[20]$ $[20]$. The activation of PPAR- α is related to the transcription of 80–100 genes involved in fatty acid oxidation, lipid metabolism and infammation [\[18](#page-7-16)]. Fibrates are the synthetic ligands of PPAR- α and are used to treat dyslipidemias in humans [[21](#page-7-19)]. PPAR-δ/β is present in a wide variety of cells, but its greatest expression occurs in tissues that have high metabolic activity, including hepatocytes, macrophages and adipocytes. When activated, PPAR-δ/β primarily regulates lipid metabolism, glucose homeostasis and inflammation $[22, 23]$ $[22, 23]$ $[22, 23]$ $[22, 23]$ $[22, 23]$. PPAR-γ is a transcription factor found in adipocytes, macrophages, monocytes, hepatocytes, muscle and endothelial cells and is the most studied of the three isotypes. It belongs to the superfamily of nuclear receptors that bind to specifc agonists, also known as ligands or activators of PPAR-γ [[24\]](#page-7-22). Thiazolidinediones (TZDs) are synthetic PPAR-γ agonists that act as insulin sensitizers and are used clinically to treat type 2 diabetes $[25]$. The three isotypes are involved in the inflammatory process in diferent tissues and cells. In this review, we focus on the role of PPARs in the infammatory process related to NAFLD and discuss new pharmacological strategies that target PPARs.

PPAR‑α

PPAR- α is expressed in many mammalian cells and tissues (Bishop-Bailey 2000; Braissant et al. 1996), including immune cells (e.g., macrophages). Consistent with this broad distribution, PPAR-α plays multiple regulatory functions, including the control of macrophage activity and infammation [\[26](#page-7-24), [27](#page-7-25)]. Specifcally in rodents, the mRNA expression of PPAR- α is highest in tissues characterized by a high rate of fatty acid oxidation, including brown adipose tissue, liver, kidneys and heart [\[28](#page-7-26)]. Considerable evidence indicates that PPAR- α serves as the master regulator of lipid metabolism in the liver, especially during fasting [[29\]](#page-7-27).

The activation of PPAR-α occurs after dimerization with the retinoid X receptor, forming a multi-protein complex with a variable set of protein coactivators. In its active form, PPAR- α binds to responsive elements in DNA, enhancing the transcription of various anti-infammatory proteins, such as the kB- α inhibitor [\[30](#page-7-28)]. PPAR- α negatively regulates the gene expression of pro-infammatory proteins by antagonizing the activities of other transcription factors, including nuclear factor kB (NF-kB), activator protein-1 (AP-1), the nuclear factor of activated T cells and signal transducers and activators of transcription proteins through direct protein–protein interactions in a process known as transrepression, which does not involve binding to typical receptor-specific PPAR response elements (PPRE), as occurs in transactivation [\[19,](#page-7-17) [31,](#page-8-0) [32\]](#page-8-1). Another PPREdependent model of transcriptional regulation has been proposed. Upon ligand activation, PPRE-bound PPAR-α directly interacts with p65 to abolish its binding to an NFκB response element in the complement C3 promoter [\[33\]](#page-8-2). The loss of PPAR-α-mediated gene transcription in PPAR-α null macrophages results in enhanced MAPK phosphorylation, leading to increased NF-κB $[26]$ $[26]$ $[26]$. PPAR-α ligands significantly reduce levels of pro-infammatory cytokines, such as interleukin-1 (IL-1), tumor necrosis factor-α (TNF-α), cyclooxygenase-2 (COX-2), and inducible nitric oxide synthase (iNOS) through the interference with AP-1 and NF-kB signaling pathways [\[34](#page-8-3)].

Murine models have demonstrated that PPAR-α expression is also regulated by natural compounds, such as Rutin, which is a plant-derived flavonoid commonly found in fruit, tea, wine $\left[35\right]$ $\left[35\right]$ $\left[35\right]$ curcumin and vitamin E $\left[36\right]$ $\left[36\right]$ $\left[36\right]$ as well as highprotein diets and fsh oil [\[37\]](#page-8-6). However, the anti-infammatory mechanisms have not been evaluated. In contrast, hydroxytyrosol, which is a polyphenol with cytoprotective efects, demonstrated anti-infammatory properties when evaluated in a high-fat diet (HFD) model. HFD-fed mice exhibited infammation, together with the downregulation of PPAR-α and Nrf2 and the up-regulation of NF-κB. Hydroxytyrosol supplementation attenuated the metabolic changes produced by the high-fat diet, normalizing the activity of Nrf2 and PPAR-α and attenuating the gene expression of the pro-infammatory mediators NF-κB, TNF-α, IL-6, and IL-1β [\[38](#page-8-7)].

Astaxanthin is a natural xanthophyll carotenoid that is abundant in marine organisms, such as microalgae and salmon [\[39\]](#page-8-8). It is also a PPAR-α agonist and PPAR- $γ$ antagonist that ameliorates lipid accumulation in HepG2 cells by regulating genes involved in lipid metabolism [\[40](#page-8-9)]. Another study by the same group showed that astaxanthin administration reduced the hepatic pro-infammatory cytokines TNF- α and IL-6 through the activation of PPAR- α in HFDfed C57BL/6J mice [\[41](#page-8-10)].

Fibrates are widely used to ameliorate the macrovascular and microvascular risks associated with metabolic syndrome and are considered PPAR- α agonists [[42\]](#page-8-11). This class includes clofbrate, gemfbrozil, fenofbrate, bezafbrate and ciprofbrate. Clofbrate was developed in the 1960s as the frst member of this class and is no longer available due to its adverse efects. Gemfbrozil and fenofbrate are available in the United States, whereas gemfbrozil, fenofbrate, bezafbrate and ciprofbrate are available in Europe. Although the main mechanism of action of fbrates is related to lipid reduction, these substances also have been shown to contribute to a reduction in infammation. In APOE2 knockout mice fed a western diet containing high levels of sucrose and cholesterol to induce steatohepatitis, treatment with fenofibrate decreased hepatic steatosis, hepatic macrophage accumulation, infammatory gene expression and the up-regulation of genes involved in beta oxidation [\[43](#page-8-12)]. Similar results were found in rats fed a high fat and fructose diet [[44](#page-8-13), [45](#page-8-14)].

In a small study involving 16 human patients with biopsyconfrmed NASH, 12 months of clofbrate treatment resulted in no changes in ALT or histological fndings in comparison with baseline [\[46\]](#page-8-15). Similarly, a small pilot study involving 16 NAFLD patients treated with fenofbrate for 48 weeks showed a lower concentration of plasma ALT concentration, but no signifcant improvement in histological fndings compared to baseline liver biopsies [\[47](#page-8-16)]. Studying 90 NAFLD patients for 24 weeks, El-Haggar and Mostafa [[48\]](#page-8-17) observed that fenofbrate improved infammation by reducing plasma levels of TNF-α and acting on fbrotic markers, such as TGFβ. Thus, given the available clinical data, PPAR-α agonists may lower the plasma concentration of ALT and reduce infammation, but do not appear to produce histological improvements in NASH in humans.

However, fibrates are weak PPAR-α agonists and have limited efficacy due to dose-related adverse events. To address this problem, a new generation of PPAR-αspecific agonists, known as selective PPAR- α modulators $(SPPARM\alpha)$, have been developed to maximize the receptormediated efects and diminish the side efects. Pemafbrate is a novel SPPARMα designed to have highly selective and tissue-specifc activity without the unwanted side efects of currently used fbrates and was developed for the treatment of dyslipidemia. Recent studies demonstrate that pemafbrate is a more potent PPAR-α-agonist than fenofbrate. In a phase-2 study, pemafbrate reduced plasma concentrations of liver enzymes in patients with dyslipidemia [[49\]](#page-8-18). Moreover, pemafbrate in db/db mice fed an MCD and AMLN diet (exhibiting the three stages of NAFLD: steatosis, steatohepatitis with fbrosis and cirrhosis) inhibited the expression of pro-inflammatory and pro-fibrotic genes ($F4/80$, TNF α , collagen 1α), indicating that this new drug is a promising therapeutic agent for NAFLD/NASH [[50](#page-8-19)].

Recently, Takei et al. [[51\]](#page-8-20) evaluated the effects of K-877 (another selective PPAR-α modulator) on metabolism and infammation in vitro and in vivo, and compared the results to those achieved with classic PPAR-α agonists. K-877 was found to be associated with benefcial changes in liver disease markers, suggesting the potential of this novel agent in the treatment of NASH/NAFLD, possibly related to PPAR-α pathway activation.

Other PPAR- α agonists have emerged with potential antiinfammatory activity in the liver. LPSF/GQ-02 is a molecule developed as a thiazolidine derivative. According to da Costa Leite et al. [[52\]](#page-8-21), the molecular structure of LPSF/ GQ-02 indicates a possible role as a PPAR-γ agonist. However, animal model studies have shown increased PPAR-α expression after LPSF/GQ-02 administration [[53\]](#page-8-22). Soares e Silva et al. [\[54](#page-8-23)] found that LPSF/GQ-02 was efective at improving the hepatic architecture, decreasing fat accumulation, reducing the amount of collagen, decreasing infammation by reducing IL-6, iNOS, COX-2 and F4/80 as well as increasing the protein expression of $I \kappa B\alpha$, cytoplasmic NFKB-65, eNOS and IRS-1 in LDLR⁻/[−] mice fed a high-fat diet. These results suggest direct action on factors that afect infammation, insulin resistance and fat accumulation. More studies are needed to characterize this receptor better and further clarify the possible mechanisms of action related to the infammatory process in the liver so that more selective and effective novel molecules can be developed (Fig. [1\)](#page-3-0).

PPAR‑β/δ

PPAR- $β/δ$ is the least studied among the PPAR isotypes, although it has signifcant expression in tissues responsible for controlling lipid metabolism, such as adipocytes, the small intestine, heart, skeletal muscle, liver and macrophages [\[55](#page-8-24)]. The removal of the gene encoding the PPARβ/δ isoform is lethal for the initial development of nearly all embryos due to a defect in the formation of the placenta [\[56](#page-8-25)]. Therefore, this isoform plays an important role not only in the regulation of metabolism, but also in the development of the organism [[57\]](#page-8-26). There are strong indications that fatty acids, triglycerides and prostacyclin are endogenous PPAR-β/δ activators. Experimental evidence suggests that PPAR-β/δ activation may have therapeutic value in the treatment of metabolic syndrome [\[58](#page-8-27)[–60\]](#page-8-28). Studies indicate that PPAR-β/δ activation exerts regulatory efects on fatty acid

Fig. 1 A schematic summarizing the anti-infammatory activity of PPAR-α in the liver. Green arrows indicate activation, and red bar-headed line indicates inhibition (see text for details)

catabolism, reverse cholesterol transport and energy metabolism [\[61](#page-9-0)]. Moreover, some PPAR-β/δ agonist substances were able to reduce insulin resistance and plasma glucose in an animal model of type 2 diabetes [[62](#page-9-1), [63](#page-9-2)].

Specifcally in the liver, PPAR-β/δ expression has been identified in many cell types, such as hepatocytes, Kupffer cells and hepatic stellate cells. Although the main efects of PPAR-β/δ in the liver are related to fatty acid and glucose metabolism, evidence suggests that PPAR-β/δ also controls anti-infammatory mechanisms in the liver. More specifcally, PPARβ/δ is a key regulator of the alternative activation of Kupfer cells towards anti-infammatory activity in the presence of IL4 and IL13 stimulation [[64](#page-9-3)]. PPAR-β/δ activation in HepG2 cells attenuates interleukin (IL)-6-induced inflammation and insulin resistance. These effects were mediated via the PPARβ/δ inhibition of IL-6-induced signal transducer and activator of transcription (STAT)-3, resulting in the restoration of normal insulin signaling [\[65](#page-9-4)]. Kupfer cell-specifc deletion of PPAR-β/δ in mice resulted in impaired hepatic Akt phosphorylation coupled with increased hepatic infammation [[64\]](#page-9-3). Together, these studies demonstrate that the activation of hepatic PPAR-β/δ attenuates infammation and contributes to improved hepatic insulin sensitivity, since the increase in infammation is directly related to hepatic insulin resistance.

Many unsaturated fatty acids can bind to PPARβ/δ in a pattern closely resembling the binding to PPAR-α. In relation to the synthetic agonists, the compound GW501516 [\[66\]](#page-9-5), which is the most potent and specifc, has become the reference PPARβ/δ agonist [[67\]](#page-9-6) Despite promising early

results, however, the further investigation and development of GW501516 was discontinued after observations in animal studies of its association with the rapid induction of cancer in several organs (liver, stomach, tongue, skin, bladder, ovaries, womb and testes). Nonetheless, it is commercially available for non-human research purposes, together with GW0742, which is also a highly selective PPARβ/δ agonist $[66]$ $[66]$ $[66]$. Lee et al. $[68]$ $[68]$ concluded that GW501516 was able to suppress the activation of infammasome and reduce IL-1 β levels, possibly through the modulation of AMPK phosphorylation, and decreased the production of radical oxygen species in both in vivo and in vitro studies, associating the anti-infammatory efect with the improvement of hepatic steatosis in mice. In a mouse model of NASH, PPARβ/δ improved hepatic steatosis and infammation by the regulation of lipid metabolism and the inhibition of the infammatory response activation of AMPK [[69\]](#page-9-8).

Using a type 2 diabetic rat model (in vivo study) as well as HepG2 and Raw264.7 cells (in vitro study), Lee et al. [\[70\]](#page-9-9) demonstrated that the PPARβ/δ agonist GW0742 attenuated hyperglycemia and fat accumulation in the liver. These changes were possibly due to the suppression of infammatory cytokines, such as TNF- α and MCP-1, suggesting that this PPAR-β/δ agonist has beneficial effects against NAFLD. In another study using GW0742, the activation of PPAR-β/δ was found to inhibit CCl4-induced liver toxicity through a mechanism involving the PPAR-β/δ-dependent downregulation of pro-infammatory signaling through interactions between PPAR-β/δ and NF-κB [[71\]](#page-9-10).

In several respects, PPAR-β/δ is comparable to PPAR-α, since both are important regulators of the beta oxidation of fatty acids. Sanderson et al. [[72](#page-9-11)] used microarray analysis to compare the hepatic transcriptome of PPAR-α-null mice and PPARβ/δ-null mice, and found little overlap between the PPAR-α-dependent and PPAR-β/δ-dependent regulation of genes. The authors also strengthened observations that PPAR-β/δ governs glucose use and the metabolism of lipoproteins and plays an important anti-infammatory role in the liver.

The ability of PPAR-β/δ to regulate several important pathways in the liver, such as lipid homeostasis, infammation and insulin resistance, demonstrates that this transcription factor plays an important role in metabolic regulation (Fig. [2\)](#page-4-0).

PPAR‑γ

PPAR- $γ$ is more highly expressed in adipose tissue and plays a crucial role in the diferentiation of adipocytes, inducing the expression of important markers involved in lipid metabolism, such as fatty acid-binding protein (aP2) [[73\]](#page-9-12), phosphoenolpyruvate carboxykinase [[74](#page-9-13)] and lipoprotein lipase [\[75](#page-9-14)]. It also controls the expression of fatty acid transporter protein 1 and CD-36 [\[76](#page-9-15)], both of which are involved in lipid uptake by adipocytes. The importance of PPAR-γ in adipogenesis has been demonstrated in several studies. For example, PPAR-γ is induced during the diferentiation of pre-adipocytes in vitro and its ectopic expression in non-adipogenic fbroblasts stimulates adipogenesis in the presence of PPAR-γ ligands [[77\]](#page-9-16).

It should also be stressed that increased PPAR-γ expression is a feature of the steatotic liver and several studies attribute a causal role to PPAR-γ in the development of steatosis through mechanisms involving the activation of lipogenic genes and de novo lipogenesis [\[78,](#page-9-17) [79\]](#page-9-18). Accordingly, the targeted deletion of PPAR-γ from hepatocytes and macrophages protects mice against diet-induced hepatic steatosis [\[80](#page-9-19)]. Moreover, hepatic PPAR-γ expression is robustly induced in NAFLD patients and experimental models [[79,](#page-9-18) [81](#page-9-20)[–83](#page-9-21)], suggesting a possible pro-steatotic role of PPAR-γ in both parenchymal and non-parenchymal cells.

Since it is a transcription factor, PPAR-γ also afects other important pathways in the organism. It has potent anti-infammatory properties that modulate the immune infammatory response [[84\]](#page-9-22). Similarly to PPAR-α, PPAR-γ activation alleviates the infammatory response through the negative interference in the transcriptional repression of genes, including NF-κB and STAT [[85](#page-9-23), [86\]](#page-9-24). PPAR-γ also regulates genes related to infammation, especially in macrophages [\[87](#page-9-25)].

Macrophages are essential components of innate immunity and play a central role in infammation and host defense [[88](#page-9-26), [89\]](#page-9-27). Kupffer cells, which are hepatic resident macrophages, represent the largest group of fxed macrophages in the body and account for 20–25% of non-parenchymal cells in the liver. Moreover, there is increasing evidence that Kupfer cells critically contribute to the pathogenesis of NAFLD [\[90\]](#page-9-28). Macrophage polarization is an important

Fig. 2 A schematic summarizing the anti-infammatory activity of PPAR-β/δ in the liver. Green arrows indicate activation, and red bar-headed line indicates inhibition (see text for details)

mechanism for regulating the infammatory response and is classically characterized by two subtypes: M1 macrophages, which are pro-infammatory and induced by TLR ligands, such as LPS and IFNγ, and M2 macrophages, which are anti-infammatory and activated by IL-4/IL-13 [[91](#page-9-29)]. PPAR-γ activation has been found to play an important role in macrophage polarization [[87,](#page-9-25) [92\]](#page-9-30). In one study, the disruption of PPAR-γ impaired alternative M2 macrophage activation and predisposed mice to obesity and insulin resistance [\[64](#page-9-3)]. Using animal model of NASH, Zhong and Liu [[93](#page-10-0)] confrmed that the activation of PPAR-γ regulates the polarization of the macrophages to M2, leading to the prevention of the development of NAFLD.

A number of natural ligands may activate PPAR-γ, including unsaturated fatty acids, eicosanoids and components of oxidized LDLs. However, the receptor affinity for many of these ligands is low and, in some cases, the physiological relevance of the ligand has not yet been determined [[94\]](#page-10-1).

PPAR-γ is the molecular target of a class of synthetic linkers known as thiazolidinediones (TZDs). The main characteristic of these molecules is the presence of a dione ring. Troglitazone was the frst PPAR-γ agonist approved by the US Food and Drug Administration (FDA) for the treatment of type 2 diabetes in 1997, but was withdrawn from the market after confrmation of severe hepatotoxicity and death [\[95\]](#page-10-2). Rosiglitazone and pioglitazone, which were approved for use by the FDA in 1999, are considered the second generation of PPAR-γ agonists.

TZDs primarily sensitize the action of insulin directly on adipocytes and indirectly on the release of adipocytokines. In the direct efect, TZDs promote the uptake and storage of fatty acids in adipose tissue, increasing the mass of this tissue and sparing other insulin-sensitive tissues, such as skeletal muscle and the liver [\[96\]](#page-10-3). In other words, TZDs promote the distribution of fat from liver and skeletal muscle cells to adipocytes.

Several studies in the literature point out that the pharmacological activation of PPAR-γ has favorable effects on the liver. The adenovirus-mediated overexpression of PPAR-γ reduced hepatic steatosis, infammation and fbrosis in a model of steatohepatitis [\[97](#page-10-4)]. Similarly, the use of rosiglitazone prevented the development of NASH in an animal model induced with a choline–methionine defcient diet [[98\]](#page-10-5). Recently, Deng et al. [[99](#page-10-6)] demonstrated that the PPAR-γ agonist pioglitazone exerts its anti-infammatory and anti-fibrotic effects by repressing the expression level of platelet-derived growth factor and tissue inhibitor of metalloproteinase-2 in an animal model of NAFLD. van der Veen et al. [\[100\]](#page-10-7) found that infammatory and fbrotic markers were reduced after the use of pioglitazone in a model of phosphatidylethanolamine *N*-methyltransferase-defcient mice, even without altering steatosis levels. In a model of acute hepatic injury in rats, rosiglitazone was able to reduce levels of infammatory markers, such as TNF-α, IL-6 and COX-2, through the downregulation of NFκB [[101\]](#page-10-8).

In humans, pioglitazone has been shown to improve hepatic steatosis, infammation and fbrosis in a meta-analysis including both diabetic and non-diabetic patients with NASH [[102](#page-10-9)]. In the PIVENS trial involving 247 non-diabetic adults with NASH randomized to receive pioglitazone, vitamin E or a placebo for 96 weeks, pioglitazone did not meet its primary endpoint, although it demonstrated a reduction in hepatic steatosis, lobular infammation as well as serum alanine and aspartate aminotransferase levels. However, subjects receiving pioglitazone gained more weight than those who received vitamin E or the placebo $[103]$ $[103]$ $[103]$, which is a side effect seen in several other studies. Although many clinical studies report the risk of adverse efects with regard to congestive cardiac failure, bladder cancer and osteoporosis [[104](#page-10-11)[–106](#page-10-12)], the European and American Associations for the Study of the Liver recommend pioglitazone for the treatment of NASH, but with important considerations regarding the long-term risks [[107,](#page-10-13) [108](#page-10-14)].

Controversially, some experimental studies indicate that increased PPAR-γ expression is a characteristic of the steatotic liver and several studies attribute a causal role of PPAR-γ in the development of steatosis through mechanisms involving the activation of genes from the pathways of lipogenesis and de novo lipogenesis [[80](#page-9-19), [100,](#page-10-7) [109](#page-10-15)]. PPAR-γ deletion in hepatocytes and macrophages protected mice against diet-induced hepatic steatosis [[80](#page-9-19)], suggesting a pro-steatotic role of PPAR-γ in both parenchymal and non-parenchymal cells. Moreover, treatment of ob/ob mice with rosiglitazone did not reverse the histological changes of NAFLD, but increased oxidative stress and steatosis [[109\]](#page-10-15). Consistent with these fndings, Soares e Silva et al. [[54\]](#page-8-23) found that pioglitazone increased levels of hepatic triglycerides in a mouse model of NAFLD, with a consequent increase in steatosis associated with elevated infammatory markers (IL-6, COX-2, F4/80 and iNOS).

Based on all of these findings, it is difficult to confirm whether PPAR- γ is the causal factor or a consequence of the accumulation of fat in the liver. These limitations highlight the need for novel approaches, such as more selective PPAR-γ agonists or drugs that effectively activate downstream targets (Fig. [3](#page-6-0)).

Dual and pan agonists

The involvement of PPARs in various biological processes, especially in the modulation of lipid metabolism and infammatory response, makes these nuclear receptors an important target for the development of new molecules. In recent years, agonists of PPARs have arisen with affinity for binding to multiple isoforms, known as dual agonists or PAN agonists.

Fig. 3 A schematic summarizing the anti-infammatory activity of PPAR-γ in the liver. Green arrows indicate activation, and red bar-headed line indicates inhibition (see text for details)

The first developed dual agonists, called glitazars (PPAR α/γ), have shown interesting results by reducing important infammatory markers in mice [[110–](#page-10-16)[112](#page-10-17)], but these studies were later abandoned because of serious adverse efects [[113,](#page-10-18) [114](#page-10-19)]. Recently, a new dual agonist $PPAR\alpha/\gamma$, saroglitazar, has presented interesting and promising results in murine model [[115\]](#page-10-20) and there was an approval for its use in India in a prospective, randomized, and registered clinical trial comparing saroglitazone with pioglitazone in patients with NAFLD (GLAZED) ([https://clini](https://clinicaltrials.gov/ct2/show/NCT02265276) [caltrials.gov/ct2/show/NCT02265276\)](https://clinicaltrials.gov/ct2/show/NCT02265276). This study began in October 2014 and was scheduled to end in September 2015, but no published results are currently available [\[42](#page-8-11)].

The GFT505, also known as Elafbranor, is PPAR-α/δ agonist and it has presented interesting results in reducing parameters of the metabolic syndrome that are related to the development of NAFLD. Short-term phase II studies demonstrated that GFT505 improves several metabolic parameters in patients with dyslipidemia and/or prediabetes [\[116](#page-10-21), [117](#page-10-22)]. Staels and cols [\[118\]](#page-10-23) evaluated the action of GTF505 on animal models of NAFLD (WD in hApoE2 KI mice, MCD diet in db/db mice and fbrosis in rat induced for CCl4), and observed that the dual agonist reduced infammatory parameters (IL-1 β , TNF- α and F4/80) associated with decreased hepatic lipid accumulation and fbrosis. More recent, Ratziu and cols [[119\]](#page-10-24), evaluated the action of Elafbranor in an international, randomized, double-blind placebo-controlled trial of patients with non-alcoholic steatohepatitis (NASH). The patients received Elafbranor 80 mg or 120 mg each day for 52 weeks. Liver enzymes, lipids, glucose profles, and markers of systemic infammation were signifcantly reduced including improvement of the cardiometabolic risk profle.

In this context, IVA337 is classifed as a new agonist generation of PPARs, having the ability to activate in a moderate and balanced way the three isoforms of PPAR $(α, β/δ)$ and γ). In a recent study, the authors investigated the effects of IVA337 on several preclinical models reproducing the main metabolic and hepatic features associated with NASH. As a result, they found that this novel compound was able to reduce infammasome genes (NLRP3, ASC, Caspase-1, IL-1β and IL-18) and important inflammatory cascade genes (CCL5, CCR2, NF-κB) as well as important markers of lipid and fbrotic metabolism, representing a potential future drug for the treatment of patients with NASH [[120](#page-10-25)].

Conclusion

Non-alcoholic fatty liver disease (NAFLD) is the most common pathology of liver and is considered part of metabolic syndrome. The evolution of NAFLD is related to the development and progression of infammation and many studies show that the reduction of infammation is a key point for the improvement of this pathology. The action potential of PPARs in NAFLD has been evaluated for several years and it is undeniable that these transcription factors regulate many infammatory processes in the liver. The bulk of these studies demonstrate the considerable anti-infammatory potential of PPARs. However, since these receptors are transcription factors and are involved in a complex metabolic network,

further studies must be conducted to explore and clarify the signaling pathways involved in the hepatic anti-infammatory process.

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Author contributions The two authors contributed equally in the elaboration of the manuscript.

Compliance with ethical standards

Conflict of interest The authors who have taken part in this study declared that they do not have anything to disclose regarding funding or confict of interest with respect to this manuscript.

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