REVIEW



Dissection of pro-tumoral macrophage subtypes and immunosuppressive cells participating in M2 polarization

Onurcan Sezginer¹ · Nese Unver¹

Received: 25 April 2024 / Revised: 6 June 2024 / Accepted: 12 June 2024 / Published online: 27 June 2024 © The Author(s) 2024

Abstract

Alternatively activated macrophage (M2) polarization can result in one of four subtypes based on cytokines and signaling pathways associated with macrophage activation: M2a, M2b, M2c, and M2d macrophages. The majority of M2 subtypes are anti-inflammatory and pro-angiogenic, secreting growth factors (VEGF, PDGF) and matrix metalloproteinases (MMP2, MMP9) which boost tumor growth, metastasis, and invasion. M2-polarized macrophages are associated with immune suppressor cells harboring Myeloid derived suppressor cells, Regulatory T cells (Tregs), Regulatory B cells as well as alternatively activated (N2) neutrophils. Treg cells selectively support the metabolic stability, mitochondrial integrity, and survival rate of M2-like TAMs in an indirect environment. Also, the contribution of Breg cells influences macrophage polarization towards the M2 direction. TAM is activated when TAN levels in the tumor microenvironment are insufficient or vice versa, suggesting that macrophage and its polarization are fine-tuned. Understanding the functions of immune suppressive cells, mediators, and signaling pathways involved with M2 polarization will allow us to identify potential strategies for targeting the TAM repolarization phenotype for innovative immunotherapy approaches. In this review, we have highlighted the critical factors for M2 macrophage polarization, differential cytokine/chemokine profiles of M1 and M2 macrophage subtypes, and other immune cells' impact on the polarization within the immunosuppressive niche.

Highlights

- M2 macrophages are targetable immune cells for immunotherapy strategies.
- M2 type macrophages are more heterogeneous populations than M1 macrophages.
- M2 polarization is influenced by Tregs, Bregs, MDSCs, and N2 neutrophils.

Communicated by L Li.

Nese Unver nese.unver@hacettepe.edu.tr

¹ Department of Basic Oncology, Cancer Institute, Hacettepe University, Sihhiye, Ankara 06100, Türkiye

Graphical Abstract



Keywords Tumor-associated macrophage \cdot M2 macrophage \cdot Macrophage polarization \cdot Immunosuppressive niche \cdot Immunotherapy

Abbreviations

CSC	Cancer stem cells
CSF	Colony stimulating factor
DC	Dendritic cells
ECM	Extracellular matrix
EMT	Epithelial-mesenchymal transition
FDR	False Discovery Rate
FGF	Fibroblast growth factor
GAS6	Growth arrest-specific 6
HIF	Hypoxia inducible factor
IFN-γ	Interferon gamma
IL	Interleukin
LogFC	Logarithmic Fold Change
LPS	Lipopolysaccharides
MDSC	Myeloid-derived suppressor cells
MMP	Matrix metalloproteinase
MSC	Mesenchymal stem cells
NK	Natural killer cells
NO	Nitric oxide
OXPHOS	Oxidative phosphorylation
PDGF	Platelet derived growth factor
ROS	Reactive oxygen species
TAM	Tumor associated macrophage
TAN	Tumor associated neutrophils
TCA Cycle	The citric acid cycle
TGF-β	Transforming growth factor beta
TLR	Toll like receptor
TME	Tumor microenvironment
TMM	Trimmed Means of M-Values

TNF-α	Tumor necrosis factor alpha
VEGF	Vascular endothelial growth factor

Introduction

Tumor niche is a multifaceted complex that is host to a wide range of cellular and non-cellular actors [1]. Among these actors, due to their remarkable plasticity and range of functions, macrophages hold an important place. Albeit having anti-tumoral properties in early stages of tumor progression, macrophages termed as tumor-associated macrophages (TAMs) residing in the tumor niche have been associated with multiple types of malignancies, including tumorigenesis, vascularization, invasion and metastasis of tumor and drug resistance [2]. The functional differences of macrophages are directly linked with their polarization.

Macrophage polarization results in primarily two distinct subtypes, namely M1 or M2 macrophages. Among these subtypes, M2 macrophages are the primary culprit in supporting tumor growth and have been the target of anticancer therapies [3]. M2 macrophages have been reported to induce tumor progression through secretion of a wide variety of proliferation inducing and immunosuppressive cytokines and chemokines. In addition to their secretory profiles, they have been reported to have cell-cell interactions that hinder immune response against tumor cells. Many cells residing in the TME induce M2 like TAM polarization, including immunosuppressive cells and tumor associated stromal cells [4–6]. M2-like polarization of TAMs should be understood well for the generation of novel therapeutic strategies and improvement of foregoing therapeutic approaches.

This review discusses participation of immune suppressor cells in acquiring the M2 phenotype of macrophages at the cellular and molecular levels, in addition to underlining the importance of macrophage polarization in the tumor microenvironment. We have also emphasized the significance of secretory profiles and immunotherapeutic targeting of both M2 macrophage subtypes and their potential roles in the immunosuppressive niche.

Tumor associated macrophages (TAMs) in tumor microenvironment

The tumor microenvironment (TME) is a dynamic entity composed of cellular and non-cellular components that induce tumor growth and progression. The non-cellular components of the TME consists of the extracellular matrix (ECM), exosomes as well as cytokines and chemokines secreted by cellular components of the TME, which contain a vast variety of cells such as endothelial cells, fibroblasts, adipocytes, neuroendocrine cells, and immune cells (Fig. 1) [7].

Tumor cells attract circulating monocytes to the TME in response to several stimuli such as VEGF, CSF-1, TGF- β , GM-CSF, IL-6, IL-10, and chemokines. The differentiation and polarization of arriving monocytes into TAMs is influenced by cytokines, chemokines, and growth factors released by cellular components of TME (Fig. 1) [8].

The presence of TAMs is often associated with poor prognosis of patients with several types of cancer. Poor prognosis in gastric cancer [9], breast cancer [10], cervical cancer [11] are associated with multiple characteristics of TAMs, such as promoting angiogenesis, tumor migration and metastasis, and immune regulation.

TAMs, a subpopulation found in intratumoral hypoxic zones, play an important role in facilitating solid tumor growth and metastasis

Hypoxia, one of the hallmarks of TME, can affect the TAMs function as TAMs differentially express hypoxia-inducible factor- 1α (HIF1 α) and HIF2 α , which are the primary factors expressed in response to hypoxia [12, 13]. Through these factors, which leads to upregulation of angiogenic growth factors VEGF, fibroblast growth factor 2 (FGF2), platelet derived growth factor (PDGF), TAM driven vessel formation can occur in TME (Fig. 1).

Endothelial degradation primarily through matrix metalloproteinases MMP2, MMP7 and MMP9 is not only required for angiogenesis, but also promotes tumor migration and metastasis. Metalloproteinases, as well as serine proteases and cathepsins secreted by TAMs can degrade matrix membranes of endothelial cells surrounding tumor tissue to induce tumor dissociation, migration, and metastasis [14]. Thus, TAMs can directly, through release of angiogenic factors, or indirectly via endothelial degradation and upregulation of angiogenic modulators, achieve proangiogenic response to hypoxic conditions of the TME.



Fig. 1 Immunosuppressive cells in cancer leading to the pro-tumoral (M2) macrophage polarization

TAMs have been shown to serve an important role in epithelial-mesenchymal transition (EMT) in addition to their role in tumor vascularization

Mesenchymal cancer cells can induce TAM-like phenotype in macrophages via secretion of GM-CSF which creates a positive feedback loop between TAMs and EMT [15]. In addition to aforementioned factors that induce EMT, IL-6, IL-8 and IL-10 secreted by TAMs also promote mesenchymal transition (Fig. 1).

TGF- β , a factor released by TAMs that plays an important role in EMT, can stimulate proliferation in later stages of tumor growth [16]. It leads to phosphorylation of SMAD2 and SMAD3 which combine with SMAD4 to form SMAD trimeric complex that translocates to the nucleus and induces transcription of EMT related genes δ EF1 and SIP1. δ EF1 and SIP1 in turn inhibits the expression of the alternative splicing factor ESRP to help cells go through EMT induced by TGF- β [17]. Thus, TGF- β has emerged to exert EMT triggering characteristics on tumor cells, and pro-tumoral TAM phenotype coincides with TGF- β [16].

Tumor necrosis factor-α (TNF-α) is another important cytokine that induces EMT. Endogenous secretion of TNF-α by TAMs and tumor cells induce EMT by inhibiting epithelial marker E-cadherin transcription and upregulating mesenchymal markers N-cadherin, fibronectin, and vimentin expression. TNF-α also induces MMP-9 expression which enhances tumor cells' ability to migrate and invade other tissues [18]. Along with their individual abilities to induce EMT, TNF-α and TGF-β can also work in concordance with one another to induce EMT. For example, in cervical cancer, TNF-α and TGF-β are shown to be working cooperatively to induce EMT and tumor stemness through NF-κB/Twist axis [19].

TAM-mediated IL-6 overexpression has been found to drive EMT in a β -catenin dependent manner [20]. Another cytokine, IL-8 acts to inhibit PI3K-Akt signaling and E-cadherin expression in tumor cells, which induces the acquisition of mesenchymal phenotype in tumor cells [21]. TAMs are also reported to induce EMT through Toll-like receptor 4 (TLR4)/IL-10 signaling by inducing mesenchymal markers Vimentin and Snail expression while simultaneously downregulating E-cadherin [22].

Tumor-associated macrophages modulate tumorigenicity of cancer stem cells

After acquiring a mesenchymal phenotype, cancer cells can enter a stem cell like state to become cancer stem cells (CSCs) with increased tumorigenic potential and chemoresistance [23]. CSCs as a concept can provide a better understanding of neoplastic progression, dissemination, and relapse of cancer [24]. A positive feedback loop occurs between CSCs and TAMs as CSCs through secretion of chemotactic factors CCL2, CCL3, CCL5, CCL8 and CXCL12 can recruit monocytes to TME. Recruited monocytes then gain a pro-tumoral phenotype through cytokines IL-4, IL-6, IL-10, IL13 and TGF-B; again, secreted by CSCs. Protumoral TAMs, in addition to induce EMT as mentioned before, can sustain CSCs' niche through secretion of cytokines (IL-6, IL-1 β , TNF- α and TGF- β) and chemokines (CCL2, CCL5 and CCL8) and inducing the expression of the transcription factor NANOG, SOX2, OCT3 and OCT4 [25]. In addition to secretory mechanisms, cell-to-cell interaction has been shown to trigger pathways critical for CSCs. For example, SHH pathway in colorectal cancer, as well as TGF-β/SMAD/NANOG pathway in pancreatic cancer have been reported to be upregulated by physical interactions between TAMs and CSCs [24, 25].

Among the factors that induce cancer stemness, glycoprotein NMB (GPNMB) is one of the most crucial proteins. GPNMB in TME is mainly expressed by TAMs, can be cleaved into a soluble form by ADAM10 which then binds to CD44 receptor in cancer cells to induce release several chemokines (CXCL1, CXCL2, CCL2, CCL5 and CCL7) as well as cytokines (IL-6, IL-11, and IL-33) [26]. IL-33 can bind to its receptor in TAMs, IL-1RL1, to induce TGF- β secretion by TAMs; while it can also bind to IL-1RL1 in other cancer cells to increase the population of CSCs through inducing the expression of stem cell genes (NANOG, NOTCH3, OCT3 and OCT4) as well as increase chemoresistance of cancer cells [27].

TAMs can both promote and suppress immunological responses against tumor cells

TAMs can induce immune response through activating $CD8^+$ T cells and natural killer (NK) cells, phagocytosis of tumor cells, and release of nitric oxide and reactive oxygen species [28]. Furthermore, TAMs can stimulate the activation of dendritic cells (DCs), which in turn can present tumor antigens to $CD8^+$ T cells and promote host immune response against tumors [29]. However, TAMs immunosuppressive features are far more common than its immuno-stimulant effects as they are known to secrete anti-inflammatory cytokines, primarily IL-10 and TGF- β [30].

In addition to cytokine production, TAMs can limit T cell activity by decreasing T cell receptor function through suppression of CD3- ζ chain expression via oxidative stress [31]. T regulatory cells (Tregs) can also be recruited to TME by TAMs via CCL22 secretion, which also serves to prevent T cell activation.

TAMs can upregulate PD-1 and PD-L1 expression, which leads to T cell exhaustion and dysfunction [32]. TAMs are

capable of inducing T cell exhaustion by preventing DC maturation. If not properly matured, DCs will have diminished expression of costimulatory molecules CD80 and CD86, and substantial expression of immunosuppressive PD-L1 expression, resulting in anergic T cells and impaired anti-tumoral host immune responses [33].

M1 and M2 macrophage subtypes are critical contributors to the tumor microenvironment

As evidenced by both cellular and molecular immunophenotypic studies, the role of TAMs in the TME is complex and multifaceted, and it can be either beneficial or detrimental to the tumor progression. This dichotomy stems from TAMs polarizing into two distinct functional subtypes: M1 and M2 macrophages (Fig. 2).

Macrophage polarization into M1 macrophages can occur through various signals (Fig. 2). Interferon- γ (IFN- γ) secretion by Type 1 T helper cells (T_h1), NK cells and CD8⁺ T cells contact with lipopolysaccharides (LPS), and GM-CSF can all promote M1 polarization [34]. M1 macrophages are intrinsically pro-inflammatory, characterized by their increased ability to secrete pro-inflammatory cytokines for instance TNF, IL-1β, IL-12 and IL-18.

Metabolic functions of M1 macrophages are also developed to sustain their pro-inflammatory function, as M1 macrophages, compared to other subtypes, hinge on glycolysis and pentose phosphate pathway to maintain their energy needs [35]. The presence of M1 macrophages is favorable in early stages of tumor development as they have shown to possess anti-tumor properties [36, 37].

M1 macrophages can attract CD8⁺ T lymphocytes and NK cells towards the TME by presenting antigens and secreting tumor-derived chemokines such as CXCL9, CXCL10, and CXCL11. Activated CD8⁺ T cells and NK

factors

cells in turn can secrete cytokines TNF-a, GM-CSF and IFN- γ , and chemokines CCL4, CCL5 and CCL23 to recruit even more immune cells to the TME to help suppress tumor growth [37].

Alternatively activated macrophage (M2) polarization shaped by tumor niche

In healthy individuals, alternatively activated macrophages, namely M2 macrophages, undergo polarization in response to fungal and parasitic infections, presence of apoptotic and necrotic cells, and downstream signaling from MCSF, IL-4, IL-13, IL-10 and TGF-β. The M2 subtype can be characterized primarily by the expression of pro-repair proteins Arginase-1 and Ym1 as well as surface marker CD206. In cancer, however, the M2 subtype is achieved through IL-4 and IL-13 secreted by T helper 2 cells [38]. Depending on cytokines and signaling pathways involved in macrophage activation, M2 polarization can result in one of four subtypes: M2a, M2b, M2c and M2d macrophages [6] (Fig. 2).

M2 macrophages act differently from M1 macrophages and differ in general through exhibiting lower glycolytic activity, enhanced aerobic citric acid cycle (TCA cycle) and oxidative phosphorylation (OXPHOS) reactions, and overall immunosuppressive effect [39].

M2 subtypes can be distinguished from each other by their metabolic signatures in addition to the deep omicbased profiling analysis. According to an analysis based on differentially expressed proteins conducted by Li et al. M2a macrophages play a role in the functions of phospholipids in phagocytosis and retinoic acid signaling; M2b participates in the transport of amino acids across the plasma membrane; M2c is engaged in the control of neutrophil chemotaxis; and M2d involves in the somatic recombination of



immunoglobulin gene segments [40]. Differences between M2 subtypes are not confined to these biological roles; omic studies of largely unexplored M2 subtypes will be directed by single cell analyses.

M2a-type TAMs are related to type II inflammation

M2a macrophages assist in tissue reconstruction and remodeling, immune regulation, and defense against parasites. M2a phenotype is achieved through IL-4 and/or IL-13 cytokine signaling, which are typically produced by T helper 2 (Th2) cells. M2a macrophages can be characterized through their high expression of the surface markers CD206, CD163, and scavenger receptor-A (SR-A) [41].

M2a macrophages in healthy individuals release a variety of cytokines and chemokines, such as IL-10, TGF-, CCL17, CCL18, and CCL22, which helps regulate the immune response and promote tissue repair and remodeling (Fig. 2) [11]. M2a macrophages, on the other hand, have been associated with tumor development because they generate growth factors and cytokines such as VEGF and PDGF, which can promote angiogenesis. In addition to promoting tumor growth, M2a macrophages also participate in tumor invasion and metastasis through production of a variety of proteases, such as MMPs and cathepsins, that degrade the ECM and facilitate tumor cell invasion and migration [42]. Furthermore, M2a macrophages can produce anti-inflammatory cytokines and chemokines, such as TGF- β and IL-10 (Fig. 2), that inhibit the activation and proliferation of T cells and NK cells, which are critical for host's anti-tumoral immune response.

Comparative analysis of cytokine (Fig. 3A) and chemokine (Fig. 3B) gene expression profiles between classically polarized M1, and alternatively polarized M2a and M2c macrophages were visualized in a heatmap. GEO dataset GSE227737 was used to analyze RNA-Seq data [43] of M1, M2a and M2c macrophages. Trimmed Means of M-Values (TMM) normalization was used to normalize the count values and edgeR package was used to determine differentially expressed genes across all groups. Genes with logarithmic fold change greater than 2 (logFC>2) and false discovery rate smaller than 0.05 (FDR < 0.05) were considered to have statistically significant variance across groups. Kyoto Encyclopedia of Genes and Genomes (KEGG) Brite database was used to evaluate the gene expression of human cytokines and chemokines.

Although extensive studies are yet to be carried out in terms of how M2 subtypes differ in their metabolism, it has been reported that the M2a macrophages display the most distinct metabolic functions compared to the other M2 subtypes. M2a macrophages have reduced energy demands indicated by decreased aerobic TCA cycle and OXPHOS activity, and lower cellular uptake of glucose compared to



Fig. 3 Comparative transcriptomic analysis of cytokine (A) and chemokine (B) gene expression profiles of classically polarized M1 and alternatively polarized M2a and M2c macrophages

other M2 subtypes, pointing towards a reduced level of glycolytic metabolism [44].

An inhibitory effect on the immune response by the substantial factors released by M2b polarized macrophages

M2b subtype is acquired upon activation of immune complexes and TLR signaling, as well as IL1R activation. Upon receiving signals from either one of these sources, NF- κ B p50 translocation into nucleus mediated by PI3K/Akt and MAPK signals is shown to be crucial in M2b polarization. In addition to NF- κ B p50, IRF3, a transcription factor expressed in macrophages, is reported to be activated during M2b polarization [45]. Although M2b cells can secrete pro-inflammatory cytokines such as IL-1 β , IL-6, TNF- α , CXCL1, CXCL2 and CXCL3, they are also found to be secreting significant amounts of IL-10 and low levels of IL-12 (Fig. 2) [45].

In addition to a plethora of pro- and anti- inflammatory cytokine secretion, CCL1 expression seems to be the hallmark of the M2b subtype of macrophages and essential for maintaining the M2b subtype [46]. CCL1 is a chemoattractant that recruits NK cells, immature B cells and DCs via interaction through cell surface CCR8. In M2b macrophages, though, autocrine production of CCL1 signaling helps cells survive without the need of exogenous growth factor. In tumor settings, this autocrine ability to proliferate helps M2b macrophages live longer than its counterparts and the prolonged presence of M2b macrophages have been reported to be linked with continuous inhibition of host antitumor immunity [47].

In contrast to M2a macrophages, M2b, M2c and M2d macrophages rely more on anaerobic glycolysis for ATP production than TCA cycle and OXPHOS. Furthermore, M2b macrophages can be distinguished from M2c and M2d in terms of TCA cycle intermediates such as enhanced intracellular glutamate, succinate and fumarate as well as increased extracellular lactate levels reflecting more active TCA cycle and glycolytic metabolism, respectively. Hence, supporting the concept that the M2b macrophages have the most ATP production amongst the M2 subtypes [44].

M2c-specific genes are involved in phagocytosis, angiogenesis, and tumor niche modification through the scavenging of pro-inflammatory factors

M2c macrophage subtype is another anti-inflammatory M2 subtype that is induced by glucocorticoids or IL-10 dependent M-CSF signaling. M2c macrophages aid efferocytosis through phagocytosis of apoptotic cells [48]. They achieve this by having a high level of mer tyrosine kinase (MerTK)

expression, which helps them identify apoptotic cells. In addition to elevated MerTK expression, they can also be characterized by comparatively high expression levels of CD14, CD163, CD169 and CD206, CCL16 and CCL18 [49, 50]. Furthermore, comparative transcriptomic analysis between macrophage subtypes has shown that the increased XCL2 and CXCL16 gene expression by M2c macrophages (Fig. 3B).

IL10 production by M2c macrophages is sustained by the production of Growth Arrest-Specific 6 (GAS6), which is a ligand for MerTK. As a result, the increase in IL10 production generates a positive loop in M2c macropahges [51]. In addition to GAS6, M2c macrophages were demonstrated to have a decreased phospholipid synthesis, which reduces the secretion metabolites required to produce pro-inflammatory cytokines such as IL1 β and induces production of IL4 and IL13 [44].

In tumorigenic conditions, M2c macrophages act similarly to M2a macrophages as they both release anti-inflammatory cytokines for instance IL10 and TGF- β (Fig. 2), as well as induce tumor invasion and metastasis through expression of VEGF and MMPs. In addition to their chemokine secretion, M2c macrophages possess increased levels of chemokine receptors CCR2 and CCR5 whose expression is induced by IL-10 signaling. However, it has been proposed that these chemokine receptors, when found in M2c macrophages, act as functional decoy receptors which scavenge proinflammatory chemokines in the TME [52].

M2d-type macrophages are differentiated by adenosine receptor agonists

Macrophage polarization is not a terminal fate for macrophages; in fact, polarized macrophages can switch from one subtype to another depending on the circumstances of their microenvironment. M2d macrophages are the prime example of this subtype switching as unlike other subtype mentioned above, M2d macrophages are derived from polarized M1 macrophages. Increasing levels of extracellular adenosine reported to be a key factor in M2d polarization; as M1 macrophages with high expression levels of adenosine receptors A2AR, respond to high extracellular adenosine levels by switching into anti-inflammatory and angiogenic M2d subtype of macrophages (Fig. 2) [53]. Adenosine production by apoptotic and necrotic cells can result in high extracellular adenosine levels. In addition to high extracellular adenosine levels, extracellular creatine, secreted by M2d macrophages can also induce M2 polarization and inhibit TLR expression to prevent inflammatory responsive niche in the TME. On the other hand, intracellular creatine concentration and uptake of creatine contribute to the maintenance of ATP homeostasis amidst phagocytosis [44].

M2 macrophage subtypes (M2a-d), while distinct, share similarities in their roles in tumor growth. M2 subtypes have characteristics that are mostly anti-inflammatory and pro-angiogenic, secreting growth factors (VEGF, PDGF) and matrix metalloproteinases (MMP2, MMP9) promoting tumor growth, metastasis, and invasion. Unlike their M1 counterparts, M2 macrophages secrete primarily immunosuppressive cytokines such as IL-4, IL-10 and IL-13 and are generally linked with tissue repair, cell proliferation and angiogenesis.

Thus, the M2 subpopulation of macrophages constitutes a much more heterogeneous group than M1 macrophages. Also, comparative transcriptomic analysis shows a remarkable distinction between M1 and M2 subtypes, and although similar in functional properties, M2 subtypes differ in their expression of cytokines and chemokines. (Fig. 3A-B).

Current targeting strategies for M2 macrophages in cancer immunotherapy

Higher proportion of M2 cells in the TME has been linked with poor prognosis and reduced immunotherapy response in several cancer types including ovarian cancer [54], nonsmall cell lung cancer [55], multiple myeloma [56] and colorectal cancer [57]. Hence, targeting M2 macrophages has been an ongoing focus for improving immunotherapy outcomes.

In a recent study using adenoviruses engineered to express IL-2 and TNF- α that reportedly reduces the number of M2 macrophages among other immunosuppressive cells and increases the efficiency of anti-PD1 therapy [58]. Another way of hampering M2 polarization is targeting IL-4R in M2 macrophages. It has been reported that the inhibition of IL-4 signaling by using IL-4 binding peptides resulted in down-regulation of M2 related markers and upregulation of M1 related genes [59].

In addition to cytokines, several metabolic pathways contribute to M2 polarization of macrophages. A recent study has shown that enhanced Zeb1 expression in THP-1 acute monocytic leukemia cell line resulted in increased expression of M2 markers such as CD206, Arg1 and IL-10, concluding that Zeb1 induced aerobic glycolysis contributes to M2 polarization of macrophages [60]. In addition, the Notch pathway has also been identified to be a key component of macrophage polarization, taking part in both M1-like and M2-like TAM generation [61]. Upon activation of the Notch signaling pathway, the resulting macrophage subtype reportedly depends on upstream ligand-receptor interaction. Macrophages, co-cultured with Delta Like Canonical Notch Ligand 4 (DDL4) expressing cells in vitro, have reported to have increased Neurogenic locus notch homolog protein 1 (NOTCH1) expression and a pro-inflammatory M1-like subtype and the blockage of DDL4 resulted in an M2-like subtype. However, further studies need to be performed to determine the extent to which the Notch signaling pathway modifies macrophage polarization in the TME [62].

Immunosuppressive cells in cancer contributing pro-tumoral (M2) macrophage polarization

TAMs primarily play an M2-like tumor-promoting role in the TME and influence multiple malignant processes such as immune suppression, angiogenesis, and tumor dissemination. M2 macrophages can be regulated by other immune cells, such as Treg cells, MDSCs and B cells This review emphasizes M2 macrophage polarization as well as the impact of other immune/stromal cells on polarization within the immunosuppressive niche. (Fig. 1). A research study in the literature supports this notion that when M2a macrophages cultured with the preconditioned-Mesenchymal stem cells (preMSC-CM) revealed increased expression of M2b/M2c-specific markers, implying that the secretome of preMSCs supports the repolarization of M2a-like macrophages to M2b/M2c subtypes [63].

Regulatory T cells (Tregs)

Malignant tumors with worse prognosis are related to elevated regulatory T cells and diminished M1 macrophages. M2 TAMs, as opposed to M1 macrophages, are more likely to encourage the recruitment of regulatory T cells to the TME [64–66].

There is a particular association between M2 macrophages and Tregs, a variety of solid tumors, as detected in colon cancer. High amounts of TGF-B, IL-10 and IL-32 are produced by Tregs, which further suppress the antitumor inflammatory response and trigger M2 macrophages to generate more cytokines and chemokines, allowing for the recruitment of more Tregs. Likewise, M2 macrophages and Tregs have been shown to have a synergistic impact in boosting ovarian cancer metastasis, tumor angiogenesis, and proliferation [4, 67–70]. The prevalence of M2 macrophages in Epstein-Barr virus-related Nasopharyngeal Carcinoma TME was associated with Treg, and both types of cells were related with tumor size and clinical stages [4]. In addition, co-infiltration of Tregs and M2 TAMs is significantly correlated with progression of premalignant lesions to oral squamous cell carcinoma as well [65].

The recruitment of CCR6⁺Foxp3⁺ regulatory T cells, a distinct effector/memory-like T cell subset, is dependent on the infiltration of TREM-1⁺ TAMs leading in

immunosuppression within the hypoxic niche of hepatocellular carcinoma for resistance to anti-PD-L1 [71, 72]. Consequently, addressing the specific subtype of TAMs should not be underestimated in malignancies that acquire immunotherapy resistance.

TAMs can be controlled indirectly by Treg cells as well. The primary cytokine that blocks M2-like TAM is IFN-g. Tregs possess the ability to block the release of IFN- γ by CD8⁺ T cells, which prevents the activation of fatty acid production in immunosuppressive M2-like TAM mediated by sterol regulatory element binding protein 1 (SREBP1). In conclusion, metabolic stability, mitochondrial integrity, and survival rate of M2-like TAMs are selectively maintained by Treg cells in an indirect context [2, 73].

TAMs and Tregs tend to communicate in the opposite directions. Tregs can be recruited to TME by CCL1 secreting M2b macrophages [51], as well as TGF- β and IL-10 producing M2c macrophages, thus creating a loop that supports anti-inflammatory phenotype in TME maintained by both M2 TAMs and Tregs [74].

As a result, assessing Tregs and M2 macrophages combined will assist in predicting the cancer progression. Taking Treg/M2 TAM status into consideration will also contribute to immunotherapeutic approaches regarding the prognosis of the disease.

Regulatory B cells (Bregs)

In addition to monocytes, which are their primary source, tumor-associated macrophages (TAM) can also develop from B-cell precursors. They differ from monocyte-derived TAM in inducing FoxP3⁺ Tregs in these macrophages, suppressing proliferation of T cells, and phagocytizing apoptotic cells more efficiently. Since they are derived from B cell precursors (pre-B and immature B cells into TAM), they are called B-MF [75] and contribute to escape mechanisms in cancer.

IL-10 released from B reg cells exerts anti-inflammatory effects in mice and humans and suppresses Th1 and Th17 responses. It leads to the production of CD4⁺CD25⁺FoxP3⁺ Treg cells and the suppression of macrophage and monocytederived inflammatory cytokines [76]. CD1d^{hi}CD5⁺ Breg cells producing IL-10 diminish TNF α and NO production. It diminishes the expression of markers associated with monocyte activation. Even a small number of CD1d^{hi}CD5⁺Breg causes antibody-mediated depletion of lymphoma cells due to the cytotoxic activity of monocytes and macrophages [77, 78]. Therefore, the contribution of Breg cells to macrophage polarization towards the M2 direction in different cancer types [76] should also be evaluated.

Myeloid-derived suppressor cells (MDSCs)

Myeloid-derived suppressor cells (MDSC) are detected in many cancer patients as well as in vivo cancer models and they serve to suppress the immune system by preventing CD4 and CD8 T cell activation. Communication between macrophages and MDSCs further subverts tumor immunity by enhancing production of IL-10 by MDSC and diminishing IL-10 production derived from macrophages in spontaneously metastatic 4T1 mouse mammary cancer [78]. The elevated generation of IL-10 originated from MDSCs also reshapes helper T cell differentiation towards the Th2 phenotype, impacting cytotoxic T lymphocyte development.

Th2 cells also generate a large amount of IL-4, which contributes to the formation of TAMs, as well. Cross-talk with MDSC may also influence macrophage MHC II expression, while IL10 has been shown to boost the expression of March 1, a ubiquitin ligase family member necessary for the ubiquitination of the cytoplasmic tail of MHC II molecules in monocytes [79, 80]. This indicates that MDSC cells potentially influence antigen-presenting capabilities of TAMs. Levels of MDSCs and CD163⁺ M2 macrophages are found to be elevated correlatively in NSCLC patients after they receive ionizing radiation therapy, pointing towards potential biomarkers during NSCLC diagnosis and favorable therapeutic response [81].

In a mouse breast cancer model, breast tumor initiation is linked to the shift of tumor infiltrating MDSCs to TAMs driven by the hypoxic TME [82]. HIF1 α is a key regulator of MDSC development and function in the TME. HIF1 supports the differentiation of M-MDSCs into tumor-associated macrophages via a process involving CD45 tyrosine phosphatase activity and STAT3 downregulation [83–85].

Alternatively activated (N2) neutrophils

TAMs and tumor associated neutrophils (TANs) are polarized by the TME to become pro-tumoral and favor tumor growth and development, invasiveness as well as matrix remodeling, angiogenesis, and metastasis, while blocking anti-tumoral immune surveillance [86]. The findings of the KrasLSL-G12D/+; p53fl/fl mouse model has demonstrated that the spleen mobilizes immature myeloid cells, and that these cells augment TAM and TAN responses compromised in the tumor progression. With regard to this, a high number of TAM and TAN precursors physically migrated from the spleen to the tumor stroma, and CCR2 signaling is involved in the recruitment of tumor-promoting spleen-derived TAMs [87]. Further research into the mechanism behind all this mutual exclusion is warranted.

Triple negative mouse breast cancer models have demonstrated that the tumors did not attract TANs and TAMs

Purpose of the study	Cancer type	Study status	Clinical trial identification
The evaluation of correlation between M1/M2 phenotype and clinical outcome.	Lung cancer	Unknown	NCT00690261
Investigaton of correlation between M1/M2 macrophage polarization and [18 F] DPA-714 PET/CT binding.	Triple Negative Breast	Completed	NCT04320030
Characterization of TAMs under combined effect of metformin and durvalumab.	Head and Neck Squamous Cell Carcinoma Oral Cavity Squamous Cell Carcinoma Oropharyngeal Squamous Cell Carcinoma	Active	NCT03618654

Table 1 The clinical trials focusing macrophage polarization in different types of solid cancer

uniformly. Even though they are the same breast cancer subtype, they might be immuno-subtyped as neutrophilenriched (NES, CD11b⁺ Ly6C^{mid} Ly6G⁺) or macrophageenriched (MES, CD11b⁺ Ly6G⁻ Ly6C⁻ F4/80⁺) based on their predisposition for recruiting TANs and TAMs which shown as mutually exclusive. TAM was triggered when TAN was depleted, or vice versa.

Conclusion and future perspective

The molecular goals have been to convert M2-polarized macrophages into M1-like macrophages via manipulating signaling pathway ligands (such as TLR), monoclonal antibodies, and miRNA/siRNA-based gene therapy [88]. Table 1 provides a summary of completed/ongoing clinical studies evaluating macrophage polarization in multiple types of solid cancer.

As a result of the extensive crosstalk between tumorassociated macrophages (M2-like macrophages) associated with poor prognosis in the tumor niche, personalized multiparametric therapeutic strategies should be taken into consideration and collectively analyzed. Thus, the remarkable improvement can be achieved relevant to therapeutic strategies, especially in immunotherapy approaches.

Combined therapeutic regimens including conventional, and immunotherapy should also be rigorously developed based on tumor niche characteristics to provide synergistic action while avoiding major side effects. Besides, insightful regulatory mechanisms between M2 macrophages and other immunosuppressive immune cells will pay the way for the cancer drug discovery in terms of targeting tumor microenvironment and insightful immunotherapy approaches.

Author contributions O.S. and N.U. wrote the manuscript and prepared the figures.

Funding This work did not receive any specific grant from funding agencies.

Open access funding provided by the Scientific and Technological Research Council of Türkiye (TÜBİTAK).

Data availability GEO dataset GSE227737 was used to analyze

RNA-Seq data and stated within the manuscript.

Declarations

Ethical approval Not applicable.

Competing interests The authors declare no competing interests.

Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http://creativecommons.org/licenses/by/4.0/.

References

- Bożyk A, Wojas-Krawczyk K, Krawczyk P, Milanowski J. Tumor Microenvironment—A Short Review of Cellular and Interaction Diversity. Biology. 2022;11:929.
- 2. Pan Y, Yu Y, Wang X, Zhang T. Tumor-Associated macrophages in Tumor Immunity. Front Immunol. 2020;11:583084.
- 3. Yao Y, Xu X-H, Jin L. Macrophage polarization in physiological and pathological pregnancy. Front Immunol. 2019;10:792.
- Aliyah S, Ardiyan Y, Mardhiyah I, Herdini camelia, Dwianingsih E, Aning S, et al. The distribution of M2 macrophage and Treg in Nasopharyngeal Carcinoma Tumor Tissue and the correlation with TNM Status and Clinical Stage. Asian Pac J Cancer Prev. 2021;22:3447–53.
- He H, Xu J, Warren CM, Duan D, Li X, Wu L, et al. Endothelial cells provide an instructive niche for the differentiation and functional polarization of M2-like macrophages. Blood. 2012;120:3152–62.
- Yunna C, Mengru H, Lei W, Weidong C. Macrophage M1/M2 polarization. Eur J Pharmacol. 2020;877:173090.
- 7. Wang M, Zhao J, Zhang L, Wei F, Lian Y, Wu Y, et al. Role of tumor microenvironment in tumorigenesis. J Cancer. 2017;8:761–73.
- 8. Qian B-Z, Pollard JW. Macrophage diversity enhances Tumor Progression and Metastasis. Cell. 2010;141:39–51.
- Wu H, Xu J-B, He Y-L, Peng J-J, Zhang X-H, Chen C-Q, et al. Tumor-associated macrophages promote angiogenesis and lymphangiogenesis of gastric cancer. J Surg Oncol. 2012;106:462–8.

- Yuan Z-Y, Luo R-Z, Peng R-J, Wang S-S, Xue C. High infiltration of tumor-associated macrophages in triple-negative breast cancer is associated with a higher risk of distant metastasis. OncoTargets Ther. 2014;1475.
- Wang Q, Sudan K, Schmoeckel E, Kost BP, Kuhn C, Vattai A, et al. CCL22-Polarized TAMs to M2a macrophages in Cervical Cancer in Vitro Model. Cells. 2022;11:2027.
- Semenza GL, Oxygen Sensing. Hypoxia-inducible factors, and Disease Pathophysiology. Annu Rev Pathol Mech Dis. 2014;9:47–71.
- Takeda N, O'Dea EL, Doedens A, Kim J, Weidemann A, Stockmann C, et al. Differential activation and antagonistic function of HIF-α isoforms in macrophages are essential for NO homeostasis. Genes Dev. 2010;24:491–501.
- 14. Lewis CE, Pollard JW. Distinct role of macrophages in different tumor microenvironments. Cancer Res. 2006;66:605–12.
- Su S, Liu Q, Chen J, Chen J, Chen F, He C, et al. A positive Feedback Loop between Mesenchymal-Like Cancer cells and macrophages is essential to breast Cancer metastasis. Cancer Cell. 2014;25:605–20.
- Shen W, Tao G, Zhang Y, Cai B, Sun J, Tian Z. TGF-β in pancreatic cancer initiation and progression: two sides of the same coin. Cell Biosci. 2017;7:39.
- Horiguchi K, Sakamoto K, Koinuma D, Semba K, Inoue A, Inoue S, et al. TGF-β drives epithelial-mesenchymal transition through δEF1-mediated downregulation of ESRP. Oncogene. 2012;31:3190–201.
- Cruceriu D, Baldasici O, Balacescu O, Berindan-Neagoe I. The dual role of tumor necrosis factor-alpha (TNF-α) in breast cancer: molecular insights and therapeutic approaches. Cell Oncol. 2020;43:1–18.
- Dong W, Sun S, Cao X, Cui Y, Chen A, Li X, et al. Exposure to TNF-α combined with TGF-β induces carcinogenesis in vitro via NF-κB/Twist axis. Oncol Rep. 2017;37:1873–82.
- Che D, Zhang S, Jing Z, Shang L, Jin S, Liu F, et al. Macrophages induce EMT to promote invasion of lung cancer cells through the IL-6-mediated COX-2/PGE 2 /β-catenin signalling pathway. Mol Immunol. 2017;90:197–210.
- Deng F, Weng Y, Li X, Wang T, Fan M, Shi Q. Overexpression of IL-8 promotes cell migration via PI3K-Akt signaling pathway and EMT in triple-negative breast cancer. Pathol - Res Pract. 2021;223:152824.
- Liu C-Y, Xu J-Y, Shi X-Y, Huang W, Ruan T-Y, Xie P, et al. M2-polarized tumor-associated macrophages promoted epithelial-mesenchymal transition in pancreatic cancer cells, partially through TLR4/IL-10 signaling pathway. Lab Invest. 2013;93:844–54.
- Borovski T, De Sousa E, Melo F, Vermeulen L, Medema JP. Cancer Stem Cell Niche: the place to be. Cancer Res. 2011;71:634–9.
- Jinushi M, Chiba S, Yoshiyama H, Masutomi K, Kinoshita I, Dosaka-Akita H, et al. Tumor-associated macrophages regulate tumorigenicity and anticancer drug responses of cancer stem/initiating cells. Proc Natl Acad Sci. 2011;108:12425–30.
- Zhang B, Ye H, Ren X, Zheng S, Zhou Q, Chen C, et al. Macrophage-expressed CD51 promotes cancer stem cell properties via the TGF-β1/smad2/3 axis in pancreatic cancer. Cancer Lett. 2019;459:204–15.
- Liguori M, Digifico E, Vacchini A, Avigni R, Colombo FS, Borroni EM, et al. The soluble glycoprotein NMB (GPNMB) produced by macrophages induces cancer stemness and metastasis via CD44 and IL-33. Cell Mol Immunol. 2021;18:711–22.
- Lin L, Li Y, Liu M, Li Q, Liu Q, Li R. The Interleukin-33/ST2 axis promotes glioma mesenchymal transition, stemness and TMZ resistance via JNK activation. Aging. 2020;12:1685–703.
- Ma P-F, Gao C-C, Yi J, Zhao J-L, Liang S-Q, Zhao Y, et al. Cytotherapy with M1-polarized macrophages ameliorates liver

fibrosis by modulating immune microenvironment in mice. J Hepatol. 2017;67:770–9.

- Li H, Shi B. Tolerogenic dendritic cells and their applications in transplantation. Cell Mol Immunol. 2015;12:24–30.
- Solinas G, Schiarea S, Liguori M, Fabbri M, Pesce S, Zammataro L, et al. Tumor-conditioned macrophages Secrete Migrationstimulating factor: a new marker for M2-Polarization, influencing Tumor Cell Motility. J Immunol. 2010;185:642–52.
- Otsuji M, Kimura Y, Aoe T, Okamoto Y, Saito T. Oxidative stress by tumor-derived macrophages suppresses the expression of CD3 ζ chain of T-cell receptor complex and antigen-specific T-cell responses. Proc Natl Acad Sci. 1996;93:13119–24.
- Yin C, Han Q, Xu D, Zheng B, Zhao X, Zhang J. SALL4-mediated upregulation of exosomal miR-146a-5p drives T-cell exhaustion by M2 tumor-associated macrophages in HCC. OncoImmunology. 2019;8:e1601479.
- del Rio M-L, Buhler L, Gibbons C, Tian J, Rodriguez-Barbosa J-I. PD-1/PD-L1, PD-1/PD-L2, and other co-inhibitory signaling pathways in transplantation. Transpl Int off J Eur Soc Organ Transpl. 2008;21:1015–28.
- Andrade MR, Amaral EP, Ribeiro SC, Almeida FM, Peres TV, Lanes V, et al. Pathogenic Mycobacterium bovis strains differ in their ability to modulate the proinflammatory activation phenotype of macrophages. BMC Microbiol. 2012;12:166.
- Kiseleva V, Vishnyakova P, Elchaninov A, Fatkhudinov T, Sukhikh G. Biochemical and molecular inducers and modulators of M2 macrophage polarization in clinical perspective. Int Immunopharmacol. 2023;122:110583.
- Bernsmeier C, van der Merwe S, Périanin A. Innate immune cells in cirrhosis. J Hepatol. 2020;73:186–201.
- Dungan LS, McGuinness NC, Boon L, Lynch MA, Mills KHG. Innate IFN-γ promotes development of experimental autoimmune encephalomyelitis: a role for NK cells and M1 macrophages. Eur J Immunol. 2014;44:2903–17.
- Anderson NR, Minutolo NG, Gill S, Klichinsky M. Macrophagebased approaches for Cancer Immunotherapy. Cancer Res. 2021;81:1201–8.
- Viola A, Munari F, Sánchez-Rodríguez R, Scolaro T, Castegna A. The metabolic signature of macrophage responses. Front Immunol. 2019;10:1462.
- Li P, Ma C, Li J, You S, Dang L, Wu J, et al. Proteomic characterization of four subtypes of M2 macrophages derived from human THP-1 cells. J Zhejiang Univ-Sci B. 2022;23:407–22.
- Pepe G, Calderazzi G, De Maglie M, Villa AM, Vegeto E. Heterogeneous induction of microglia M2a phenotype by central administration of interleukin-4. J Neuroinflammation. 2014;11:211.
- Mantovani A, Biswas SK, Galdiero MR, Sica A, Locati M. Macrophage plasticity and polarization in tissue repair and remodelling: macrophage plasticity and polarization in tissue repair and remodelling. J Pathol. 2013;229:176–85.
- 43. Gunalp S, Helvaci DG, Oner A, Bursalı A, Conforte A, Güner H et al. TRAIL promotes the polarization of human macrophages toward a proinflammatory M1 phenotype and is associated with increased survival in cancer patients with high tumor macrophage content. Front Immunol [Internet]. 2023;14. https://www.frontiersin.org/articles/https://doi.org/10.3389/fimmu.2023.1209249.
- Fuchs AL, Costello SM, Schiller SM, Tripet BP, Copié V. Primary human M2 macrophage subtypes are distinguishable by Aqueous Metabolite profiles. Int J Mol Sci. 2024;25:2407.
- Li Y, Cai L, Wang H, Wu P, Gu W, Chen Y, et al. Pleiotropic regulation of macrophage polarization and tumorigenesis by formyl peptide receptor-2. Oncogene. 2011;30:3887–99.
- Asai A, Nakamura K, Kobayashi M, Herndon DN, Suzuki F. CCL1 released from M2b macrophages is essentially required for the maintenance of their properties. J Leukoc Biol. 2012;92:859–67.

- Asai A, Tsuchimoto Y, Ohama H, Fukunishi S, Tsuda Y, Kobayashi M, et al. Host antitumor resistance improved by the macrophage polarization in a chimera model of patients with HCC. OncoImmunology. 2017;6:e1299301.
- Lai Y-S, Putra RBDS, Aui S-P, Chang K-T. M2C polarization by Baicalin enhances efferocytosis via Upregulation of MERTK receptor. Am J Chin Med. 2018;46:1899–914.
- 49. Cappello P, Caorsi C, Bosticardo M, De Angelis S, Novelli F, Forni G, et al. CCL16/LEC powerfully triggers effector and antigen-presenting functions of macrophages and enhances T cell cytotoxicity. J Leukoc Biol. 2004;75:135–42.
- Zizzo G, Hilliard BA, Monestier M, Cohen PL. Efficient clearance of early apoptotic cells by human macrophages requires M2c polarization and MerTK induction. J Immunol. 2012;189:3508–20.
- 51. Zhang Q, Sioud M. Tumor-Associated Macrophage subsets: shaping polarization and targeting. Int J Mol Sci. 2023;24:7493.
- Mantovani A, Sica A, Sozzani S, Allavena P, Vecchi A, Locati M. The chemokine system in diverse forms of macrophage activation and polarization. Trends Immunol. 2004;25:677–86.
- 53. Ferrante CJ, Pinhal-Enfield G, Elson G, Cronstein BN, Hasko G, Outram S, et al. The Adenosine-Dependent Angiogenic switch of macrophages to an M2-Like phenotype is Independent of Interleukin-4 receptor alpha (IL-4Rα) signaling. Inflammation. 2013;36:921–31.
- Macciò A, Gramignano G, Cherchi MC, Tanca L, Melis L, Madeddu C. Role of M1-polarized tumor-associated macrophages in the prognosis of advanced ovarian cancer patients. Sci Rep. 2020;10:6096.
- Sumitomo R, Hirai T, Fujita M, Murakami H, Otake Y, Huang C. M2 tumor–associated macrophages promote tumor progression in non–small–cell lung cancer. Exp Ther Med. 2019;18:4490–8.
- 56. Chen X, Chen J, Zhang W, Sun R, Liu T, Zheng Y et al. Prognostic value of diametrically polarized tumor-associated macrophages in multiple myeloma. Oncotarget Vol 8 No 68 [Internet]. 2017 [cited 2017 Jan 1]; https://www.oncotarget.com/article/22340/ text/.
- Herrera M, Herrera A, Domínguez G, Silva J, García V, García JM, et al. Cancer-associated fibroblast and M2 macrophage markers together predict outcome in colorectal cancer patients. Cancer Sci. 2013;104:437–44.
- Cervera-Carrascon V, Quixabeira DCA, Santos JM, Havunen R, Milenova I, Verhoeff J et al. Adenovirus Armed With TNFa and IL2 Added to aPD-1 Regimen Mediates Antitumor Efficacy in Tumors Refractory to aPD-1. Front Immunol [Internet]. 2021;12. https://www.frontiersin.org/articles/https://doi.org/10.3389/ fimmu.2021.706517.
- 59. Gunassekaran GR, Poongkavithai Vadevoo SM, Baek M-C, Lee B. M1 macrophage exosomes engineered to foster M1 polarization and target the IL-4 receptor inhibit tumor growth by reprogramming tumor-associated macrophages into M1-like macrophages. Biomaterials. 2021;278:121137.
- Jiang H, Wei H, Wang H, Wang Z, Li J, Ou Y, et al. Zeb1-induced metabolic reprogramming of glycolysis is essential for macrophage polarization in breast cancer. Cell Death Dis. 2022;13:206.
- Palaga T, Wongchana W, Kueanjinda P. Notch Signaling in macrophages in the Context of Cancer Immunity. Front Immunol. 2018;9:652.
- 62. Liu H, Wang J, Zhang M, Xuan Q, Wang Z, Lian X, et al. Jagged1 promotes aromatase inhibitor resistance by modulating tumorassociated macrophage differentiation in breast cancer patients. Breast Cancer Res Treat. 2017;166:95–107.
- 63. Holthaus M, Santhakumar N, Wahlers T, Paunel-Görgülü A. The secretome of preconditioned mesenchymal stem cells drives polarization and reprogramming of M2a macrophages toward an IL-10-Producing phenotype. Int J Mol Sci. 2022;23:4104.

- Davidsson S, Fiorentino M, Giunchi F, Eriksson M, Erlandsson A, Sundqvist P, et al. Infiltration of M2 Macrophages and Regulatory T Cells Plays a role in recurrence of renal cell carcinoma. Eur Urol Open Sci. 2020;20:62–71.
- 65. Kouketsu A, Sato I, Oikawa M, Shimizu Y, Saito H, Tashiro K, et al. Regulatory T cells and M2-polarized tumour-associated macrophages are associated with the oncogenesis and progression of oral squamous cell carcinoma. Int J Oral Maxillofac Surg. 2019;48:1279–88.
- 66. Sun W, Wei F-Q, Li W-J, Wei J-W, Zhong H, Wen Y-H, et al. A positive-feedback loop between tumour infiltrating activated Treg cells and type 2-skewed macrophages is essential for progression of laryngeal squamous cell carcinoma. Br J Cancer. 2017;117:1631–43.
- Cao Q, Wang Y, Zheng D, Sun Y, Wang Y, Lee VWS, et al. IL-10/TGF-β–Modified Macrophages Induce Regulatory T Cells and protect against Adriamycin Nephrosis. J Am Soc Nephrol. 2010;21:933–42.
- Chen Y, Gao Y, Ma X, Wang Y, Liu J, Yang C, et al. A study on the correlation between M2 macrophages and regulatory T cells in the progression of colorectal cancer. Int J Biol Markers. 2022;37:412–20.
- Ris MM, Deitrich RA, Von Wartburg JP. Inhibition of aldehyde reductase isoenzymes in human and rat brain. Biochem Pharmacol. 1975;24:1865–9.
- Tiemessen MM, Jagger AL, Evans HG, van Herwijnen MJC, John S, Taams LS. CD4 ⁺ CD25 ⁺ Foxp3 ⁺ regulatory T cells induce alternative activation of human monocytes/macrophages. Proc Natl Acad Sci. 2007;104:19446–51.
- Wu Q, Zhou W, Yin S, Zhou Y, Chen T, Qian J, et al. Blocking triggering receptor expressed on myeloid cells-1-Positive Tumor-Associated macrophages Induced by Hypoxia reverses immunosuppression and anti-programmed cell death Ligand 1 resistance in Liver Cancer. Hepatology. 2019;70:198–214.
- 72. Yu S, Wang Y, Hou J, Li W, Wang X, Xiang L et al. E Wang editor 2020 Tumor-infiltrating immune cells in hepatocellular carcinoma: Tregs is correlated with poor overall survival. PLoS ONE 15 e0231003.
- Liu C, Chikina M, Deshpande R, Menk AV, Wang T, Tabib T, et al. Treg cells promote the SREBP1-Dependent metabolic fitness of Tumor-promoting macrophages via repression of CD8 + T cellderived Interferon-γ. Immunity. 2019;51:381–e3976.
- Vidyarthi A, Agnihotri T, Khan N, Singh S, Tewari MK, Radotra BD, et al. Predominance of M2 macrophages in gliomas leads to the suppression of local and systemic immunity. Cancer Immunol Immunother. 2019;68:1995–2004.
- Chen C, Park B, Ragonnaud E, Bodogai M, Wang X, Zong L, et al. Cancer co-opts differentiation of B-cell precursors into macrophage-like cells. Nat Commun. 2022;13:5376.
- Schwartz M, Zhang Y, Rosenblatt JD. B cell regulation of the anti-tumor response and role in carcinogenesis. J Immunother Cancer. 2016;4:40.
- Horikawa M, Minard-Colin V, Matsushita T, Tedder TF. Regulatory B cell production of IL-10 inhibits lymphoma depletion during CD20 immunotherapy in mice. J Clin Invest. 2011;121:4268–80.
- Sinha P, Clements VK, Bunt SK, Albelda SM, Ostrand-Rosenberg S. Cross-talk between myeloid-derived suppressor cells and macrophages subverts Tumor immunity toward a type 2 response. J Immunol. 2007;179:977–83.
- DeNardo DG, Barreto JB, Andreu P, Vasquez L, Tawfik D, Kolhatkar N, et al. CD4+T cells regulate pulmonary metastasis of mammary carcinomas by enhancing Protumor properties of macrophages. Cancer Cell. 2009;16:91–102.
- Thibodeau J, Bourgeois-Daigneault M-C, Huppé G, Tremblay J, Aumont A, Houde M, et al. Interleukin-10-induced MARCH1

mediates intracellular sequestration of MHC class II in monocytes. Eur J Immunol. 2008;38:1225–30.

- 81. Lv M, Zhuang X, Shao S, Li X, Cheng Y, Wu D et al. Myeloid-Derived Suppressor Cells and CD68+CD163+M2-Like Macrophages as Therapeutic Response Biomarkers Are Associated with Plasma Inflammatory Cytokines: A Preliminary Study for Non-Small Cell Lung Cancer Patients in Radiotherapy. Ortega E, editor. J Immunol Res. 2022;2022:1–16.
- Fang Z, Wen C, Chen X, Yin R, Zhang C, Wang X, et al. Myeloidderived suppressor cell and macrophage exert distinct angiogenic and immunosuppressive effects in breast cancer. Oncotarget. 2017;8:54173–86.
- Corzo CA, Condamine T, Lu L, Cotter MJ, Youn J-I, Cheng P, et al. HIF-1α regulates function and differentiation of myeloidderived suppressor cells in the tumor microenvironment. J Exp Med. 2010;207:2439–53.
- Kumar V, Cheng P, Condamine T, Mony S, Languino LR, McCaffrey JC, et al. CD45 Phosphatase inhibits STAT3 transcription

factor activity in myeloid cells and promotes Tumor-Associated Macrophage differentiation. Immunity. 2016;44:303–15.

- Liu G, Bi Y, Shen B, Yang H, Zhang Y, Wang X, et al. SIRT1 limits the function and fate of myeloid-derived suppressor cells in tumors by orchestrating HIF-1 α –Dependent glycolysis. Cancer Res. 2014;74:727–37.
- Wu L, Zhang XH-F. Tumor-Associated neutrophils and macrophages—heterogenous but not chaotic. Front Immunol. 2020;11:553967.
- Cortez-Retamozo V, Etzrodt M, Newton A, Rauch PJ, Chudnovskiy A, Berger C, et al. Origins of tumor-associated macrophages and neutrophils. Proc Natl Acad Sci. 2012;109:2491–6.
- Poh AR, Ernst M. Targeting macrophages in Cancer: from bench to Bedside. Front Oncol. 2018;8:49.

Publisher's Note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.