Thomas N. Chase Justin D. Oh Spyridon Konitsiotis

Antiparkinsonian and antidyskinetic activity of drugs targeting central glutamatergic mechanisms

Abstract Motor dysfunction produced by the chronic non-physiological stimulation of dopaminergic receptors on striatal medium spiny neurons is associated with alterations in the sensitivity of glutamatergic receptors, including those of the Nmethyl-D-aspartate (NMDA) subtype. Functional characteristics of these ionotropic receptors are regulated by their phosphorylation state. Lesioning the nigrostriatal dopamine system of rats induces parkinsonian signs and increases the phosphorylation of striatal NMDA receptor subunits on serine and tyrosine residues. The intrastriatal administration of certain inhibitors of the kinases capable of phosphorylating NMDA receptors produces a dopaminomimetic motor response in these animals. Treating parkinsonian rats twice daily with levodopa induces many of the characteristic features of the human motor complication syndrome and further increases the serine and tyrosine phosphorylation of specific NMDA receptor subunits. Again, the intrastriatal administration of selective inhibitors of certain serine and tyrosine kinases alleviates the motor complications. NMDA receptor antagonists, including some non-competitive chan-

nel blockers, act both palliatively and prophylactically in rodent and primate models to reverse these levodopa-induced response alterations. Similarly, in clinical studies dextrorphan, dextromethorphan, and amantadine have been found to be efficacious against motor complications. Recent observations in animal models further indicate that certain amino-3-hydroxy-5-methyl-4-isoxazole proprionic acid (AMPA) antagonists alleviate, while others exacerbate, these complications. Thus, it appears that the denervation or intermittent stimulation of striatal dopaminergic receptors differentially activates signal transduction pathways in medium spiny neurons. These in turn modify the phosphorylation state of ionotropic glutamate receptors and consequently their sensitivity to cortical input. These striatal changes contribute to symptom production in Parkinson's disease, and their prevention or reversal could prove useful in the treatment of this disorder.

Key Words AMPA receptor · Medium spiny neuron · NMDA receptor · Phosphorylation · Signal transduction

Introduction

The cardinal signs of Parkinson's disease (PD) reflect alterations in striatal dopaminergic transmission. Tremor, rigid-

T. N. Chase $(\boxtimes) \cdot J. D. Oh \cdot S.$ Konitsiotis

Disorders and Stroke NIH, Building 10,

Room 5C211, 10 Center Drive, MSC/406,

Experimental Therapeutics Branch, National Institute of Neurological

Bethesda, MD 20892, USA

Fax: 001 301 496 6609

E-mail: Chase@helix.nigh.gov Tel.: 001 301 496 7993

> ity, and bradykinesia result from an insufficiency of intrasynaptic dopamine (DA) and become clinically manifest when striatal levels of the transmitter decline by about 50% [13, 33, 57]. These motor abnormalities initially respond well to drugs such as levodopa or DA agonists that

improve dopaminergic transmission. Later, however, the response to pharmacological agents of this type becomes increasingly less satisfactory, mainly due to the appearance of treatment-refractory signs (gait and balance disorders, freezing, dementia, and affective changes) and various adverse events, especially motor response complications [18, 55]. Most commonly, these latter changes include various types of response fluctuations and dyskinesias [55, 69]. As in the case of parkinsonian signs, these ultimately disabling adverse events probably reflect the non-physiological stimulation of striatal DA receptors. In contrast to the chronic hypostimulation produced by DA system degeneration, however, it is the periodic hyperstimulation associated with most dopaminomimetic therapies that appears to favor the appearance of motor response complications.

Non-physiological stimulation of dopamine receptors

While it is easy to understand why dopaminergic denervation might alter downstream mechanisms in the basal ganglia, the basis for the reactive changes associated with standard dopaminomimetic therapies has only recently become clear. DA-containing neurons comprising the nigrostriatal system characteristically manifest slow (about 4–5 Hz), single-spike activity which is occasionally interrupted by short bursts of faster (usually in the 15-20 Hz range) spiking in response to salient visual or auditory stimuli [29, 71]. Since postsynaptic receptor stimulation is roughly proportional to impulse activity at the presynaptic terminal, intrasynaptic DA concentrations normally remain fairly constant. Accordingly, the most physiological approach to dopamine replacement in PD would be to maintain stable normal intrasynaptic levels of the transmitter amine; however, with disease progression, this goal becomes progressively less attainable. As nigral dopaminergic neurons degenerate, the amount of exogenous levodopa entering striatal terminals diminishes. Instead, increasing amounts are taken up and converted to DA in other decarboxylase-containing cells, especially serotonergic neurons [52, 59, 84]. In the absence of appropriate mechanisms for storing or regulating the release of DA, the newly synthesized amine leaks into the extracellular compartment and diffuses into nearby DA receptors. Under such circumstances, intrasynaptic DA concentrations reflect the wide swings in cerebral levolopa levels that occur with standard precursor dosing regimens. Levodopa therapy in patients with advanced PD thus results, at best, in only episodic restoration of physiological dopamine levels [9]. For most of the dosing cycle, with the administration of short-acting DA agonists at any stage of Parkinson's disease, or of levodopa in patients with advanced disease, dopaminergic receptor stimulation remains at subthreshold levels, interrupted soon after each dose when it briefly rises into the physiological range. Indeed, postmortem determinations of striatal DA concentrations, as well as clinical measurements of spinal fluid homovanillic acid levels (the major metabolite of DA), suggest that transmitter levels in parkinsonian patients receiving standard levodopa therapy ordinarily peak well above the physiological range, presumably at levels approximating to those achieved in in vitro models exposed to high-intensity (tetanic) stimulation [17, 37, 58]. With a sufficient loss of DA terminals, dopaminergic transmission thus tends to be compromised whether or not dopaminomimetic treatment is initiated.

It has now become increasingly clear that this non-physiological pattern of stimulation contributes to the appearance of the major motor complications associated with long-term levodopa or DA agonist administration. The nigrostriatal DA system terminates on the dendritic spines of the preponderant striatal nerve cell, the medium-sized spiny neuron [44]. Medium spiny neurons also receive glutamatergic axons descending from all areas of the cerebral cortex. In addition, they make synaptic contact with numerous other neuronal systems, both extrinsic (e.g., adrenergic and serotoninergic) and intrinsic (e.g., cholinergic and somatostatinergic) to the striatum. In turn, these GABAergic efferent neurons project, both directly and indirectly, to the major output nuclei of the basal ganglia, the internal segment of the globus pallidus and the pars reticulata of the substantia nigra [30, 73]. Medium spiny neurons, thus, serve as a major anatomical locus for the processing of cortical information through the basal ganglia. They also appear to contribute to certain plastic responses now associated with basal ganglia function [11, 16].

Increasing evidence suggests that the chronic non-physiological stimulation of DA receptors triggers adaptative responses in the striatum and other basal ganglionic structures. To date, these changes have largely been studied in animal models of PD. Rats rendered parkinsonian by the injection of 6-hydroxydopamine and then treated intermittently with levodopa (by twice-daily injection to simulate clinical conditions) develop progressive motor response alterations, which resemble the fluctuations occurring in similarly treated parkinsonian patients [67]. Daily measurements reveal a progressive shortening in response duration that becomes statistically significant within about 3 weeks. Thus, like parkinsonian patients, parkinsonian rats exhibit the wearing-off phenomenon. At the same time, they also show evidence of response alterations that mimic human on-off fluctuations. The frequency with which there is no response to an otherwise effective dose of levodopa rises, and the slope of the levodopa dose/motor response relation becomes steeper [49]. While parkinsonian rats develop motor fluctuations, under the conditions of these experiments they do not develop choreiform dyskinesias. In contrast to 6-hydroxydopamine-lesioned rats, cynomolgus monkeys lesioned with MPTP (1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine) develop typical choreiform and dystonic movements within a few weeks of daily levodopa treatment [8, 48].

Peptide cotransmitters

Levels of the peptide cotransmitters used by medium spiny neurons can serve as reliable markers of their functional state. Several of these neuropeptides have been found to undergo characteristic alterations in parkinsonian rodents and primates as a result of denervation and subsequent levodopa therapy [25, 27, 34, 35, 56, 68]. Spiny neurons that predominantly express the D2 DA receptor subtype mainly project to the internal segment of the globus pallidus via the external globus pallidus and subthalamic nucleus, and they contain the neuropeptides enkephalin and neurotensin. Spiny neurons that primarily express D1 DA receptors largely project directly to the internal segment of the globus pallidus/pars reticulata of the substantia nigra, and contain dynorphin and neurotensin [27, 45]. Lesioning the DA system of rats with 6-hydroxydopamine increases striatal concentrations of both enkephalin and neurotensin [25]. Subsequent intermittent levodopa treatment at a dose sufficient to induce motor response alterations leads to additional changes, most notably a striking rise in dynorphin and neurotensin levels [23, 25]. At the same time, the expression of messenger ribonucleic acids (mRNAs) encoding for these neuropeptides also increases, suggesting that their concentration changes may be the result of accelerated synthesis [27]. Related pharmacological studies indicate that these peptide modifications reflect alterations in spiny neuron output which can influence extrapyramidal motor function [22, 26, 48].

Role of NMDA receptors in Parkinson's disease

The effects of non-physiological stimulation on synaptic plasticity have been extensively studied in various animal models of learning and behavior [62]. Early investigative attention focused largely on the ability of repetitive highfrequency (tetanic) stimulation to evoke long-term potentiation (LTP) in the hippocampus [38, 51, 85]. More recently, LTP-like phenomena have been found in cortical and subcortical areas which influence motor behavior [2, 12, 14]. In these model systems, considerable evidence suggests that a rise in the sensitivity of glutamatergic receptors, especially those of the N-methyl-D-aspartate (NMDA) subtype, contributes to the persisting, activity-dependent changes in neuronal responses [10, 60]. Since NMDA and DA receptors are co-expressed in close proximity along the distal dendrites of medium spiny neurons [43, 73], the foregoing preclinical observations prompted evaluations of the possibility that the non-physiological stimulation of DA receptors on these striatal neurons might enhance NMDA receptor sensitivity in ways that favor the clinical appearance of parkinsonism and long-lasting motor complications [15, 24, 65].

NMDA receptors are heteroligomers assembled to form ligand-gated ion channels from one or two NR1 subunits,

expressed in eight currently recognized splice variants (ah), and two or three NR2 subunits composed of four homologous isoforms (A-D) [64, 87]. In rat striatum, medium spiny neurons express NR1 variants along with NR2B and, to a lesser extent, NR2A subunits [19]. Protein phosphorylation serves as a major regulatory mechanism for these receptors [32, 75]. The phosphorylation of tyrosine residues has been reported to modulate channel characteristics, including opening probability [86, 88], while serine/threonine phosphorylation by calcium/phospholipidstimulated or cAMP-stimulated protein kinases appears to affect their subcellular distribution and anchoring to plasma membranes [36, 79]. Recent studies of rat striatal NMDA receptors have revealed changes in both tyrosine and serine phosphorylation that are associated with the development of parkinsonism following nigrostriatal system destruction, as well as with the appearance of motor response alterations following intermittent levodopa therapy.

The phosphorylation of rat striatal NMDA receptors at tyrosine residues increases when 6-hydroxydopamine-induced parkinsonism becomes evident [53, 62], and to an even greater extent when the altered motor responses to levodopa appear [62]. Both nigrostriatal denervation and subsequent levodopa administration mainly affect striatal NR2B subunits, although levodopa treatment also augments NR2A subunit tyrosine phosphorylation. In agreement with previous observations [80], we have also found that lesioning with 6-hydroxydopamine selectively increases striatal NR2A protein expression. Subsequent intermittent levodopa administration normalized levels of NR2A subunits, but had no effect on NR2B expression [62]. In relation to serine phosphorylation, there are increases on striatal NR2A subunits when rats are lesioned with 6-hydroxydopamine, and these are further augmented when the animals show altered motor responses to levodopa [63]. Neither nigrostriatal pathway destruction nor levodopa administration affect the expression or serine phosphorylation of NR2B subunits. Since receptor channel function, and therefore calcium ion fluxes, at the NMDA receptor complexes reflect their phosphorylation state, it is not unreasonable to assume that the observed enhancement in tyrosine and serine phosphorylation contributes to their heightened sensitivity and thus to the motor dysfunction which accompanies dopaminergic denervation and dopaminomimetic therapy [63].

Phosphorylation changes affecting NMDA receptors on striatal spiny neurons presumably depend on intracellular signaling cascades linking them with nearby dopaminergic receptors. Both the nigral dopaminergic and cortical glutamatergic systems innervate the distal dendrites of medium spiny neurons [43, 73]. Glutamatergic excitatory projections terminate at the spine heads, where all three of the major glutamatergic receptor subtypes [NMDA, α amino-3-hydroxy-5-methyl-4-isoxazole proprionic acid (AMPA) and kainate] are expressed [1, 40]. Dopaminergic terminals make synaptic contact on the spine necks, within a micron of the glutamate receptors, as well as at more proximal sites [72]. This intimate anatomical arrangement affords the potential for close physiological interactions between dopaminergic and glutamatergic receptor-mediated mechanisms. Indeed, recent observations suggest that the non-physiological stimulation of rat DA receptors activates striatal kinases believed to be capable of directly phosphorylating NMDA receptor subunits [61-63]. These include serine kinases, such as cyclic AMP-protein kinase A (PKA) and calcium/calmodulin-dependent protein kinase II (CaMKII), as well as tyrosine kinases, which are as yet unidentified but are most likely to be members of the src or fyn families [42, 61–63, 76, 77, 88]. Intrastriatal injection of the PKA inhibitor, Rp-cAMPS, or the CaMKII inhibitor, KN-93, for example, reverses the motor response changes produced by intermittent levodopa therapy [61, 63]. At the same time, KN-93 has been shown to attenuate the enhanced serine phosphorylation of NMDA receptors produced by levodopa therapy [63]. With respect to tyrosine phosphorylation, striatal infusion of the tyrosine kinase inhibitor, genistein, normalizes both the tyrosine phosphorylation increases and the motor response changes associated with levodopa treatment. Conversely, the tyrosine phosphatase inhibitor, okadaic acid, potentiates these alterations [62]. The foregoing results support the possibility that sensitization of NMDA receptors on striatal spiny neurons resulting, at least in part, from heightened subunit phosphorylation contributes to the onset of parkinsonian signs as a consequence of dopaminergic denervation, as well as to the appearance of the response modifications associated with intermittent levodopa treatment. In either case, alterations in cortical glutamatergic input to the striatum presumably modify striatal output in ways that influence motor function.

Pharmacological evaluations of this possibility were initially based on the premise that if an increase in striatal NMDA receptor sensitivity played a role in the production of symptoms associated with dopaminergic denervation or levodopa treatment, then pharmacological blockade of these receptors should ameliorate the motor dysfunction. The results of early studies in parkinsonian rats appeared consistent with this possibility, since NMDA receptor antagonists, such as MK801, were found to act both palliatively and prophylactically to decrease response alterations [24, 49, 65]. Subsequent observations in parkinsonian primates provided additional support for this hypothesis. Co-administration of certain NMDA antagonists to these animals substantially reduced the dyskinesiogenic effects of levodopa [4, 7, 28, 66]. Similarly, studies in parkinsonian patients given NMDA receptor antagonists, such as dextrorphan, dextromethorphan or amantadine, indicated that drugs of this type can alleviate motor fluctuations as well as peak dose dyskinesias [6, 81–83]. Regarding the clinical appearance of parkinsonian signs, results from 6hydroxydopamine-lesioned rodents [76] as well as MPTPlesioned primates [5, 31, 54] indicate that some intrastriatally- or systemically administered NMDA antagonists possess antiparkinsonian activity. Similarly, the well-established symptomatic benefit conferred to mildly afflicted parkinsonian patients by amantadine suggests that NMDA receptor antagonists can act as clinically effective pallia-

tives [21, 74]. The foregoing preclinical and clinical observations support the possibility that striatal NMDA receptor sensitization contributes to the characteristic motor dysfunction occurring with both dopaminergic denervation and levodopa therapy.

Role of AMPA receptors in Parkinson's disease

Recent observations suggest that functional alterations in glutamate receptors other than those of the NMDA type may also contribute to symptom production in PD. For example, the administration of the competitive AMPA receptor antagonist, NBQX, to parkinsonian rats or monkeys reportedly has little or no effect on motor function but potentiates the antiparkinsonian action of levodopa [41, 46, 47]. In rats, we have found that NBQX acts to reverse levodopa-associated motor response alterations [48]. In primates, a selective, non-competitive antagonist of the AMPA allosteric modulation site (LY 300164) alone did not modify the severity of parkinsonian signs, but did attenuate levodopa-induced dyskinesias. Conversely, a selective AMPA agonist (CX516) by itself had no antiparkinsonian activity, but potentiated levodopa-associated dyskinesias [43]. These animal model results suggest that alterations in AMPA receptor-mediated mechanisms contribute to the motor dysfunction associated with dopaminergic denervation and subsequent dopaminomimetic treatment. The same may be true for glutamate receptors in the metabotropic family. All three currently identified subtypes of these Gprotein-coupled receptors are known to be expressed in the striatum [87]. Both group I and III metabotropic agonists have now been reported to induce contralateral rotation, similar to dopaminomimetics, in 6-hydroxydopamine lesioned rats [70, 39].

A complex series of events involving glutamate receptor-mediated mechanisms in the basal ganglia participate in the plastic changes in motor function that characteristically arise in parkinsonian patients, initially as a result of the loss of striatal dopaminergic innervation and later due to the intermittent high-intensity stimulation produced by most currently available dopaminomimetic therapies. Although drugs that prevent or reverse these changes might be expected to confer symptomatic benefit, considerable additional investigative effort will be required to determine the optimal sites of pharmaceutical intervention. In part, this will involve the precise elucidation of the relative contribution of factors, such as the activation of particular signal transduction cascades, the differential phosphorylation of glutamate receptor subunits, and the sensitization of glutamatergic receptor subtypes to their natural

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ligand. It may also entail improving our understanding of the modulatory role played by other neuronal systems that make synaptic contact with critical neurons in the striatum and in downstream basal ganglionic structures. Clearly, the need for improved palliative treatments for PD continues to be a crucial goal for pharmacological discovery. Of particular importance in this regard is exploration of the therapeutic potential of currently available drugs that selectively interact with central glutamatergic systems. Conceivably, the precise targeting of glutamatergic mechanisms in the basal ganglia could provide the safest and most effective therapy for all stages of PD. In view of the neuroprotective potential of NMDA and AMPA antagonists and metabotropic agonists, intensifying the evaluation of their palliative utility for those suffering from PD seems eminently worthwhile.

References

- Albin RL, Makowiec RL, Hollingsworth ZR, Dure LS 4th, Penney JB, Young AB (1992) Excitatory amino acid binding sites in the basal ganglia of the rat: a quantitative autoradiographic study. Neuroscience 46:35–48
- Asanuma H, Pavlides C (1997) Neurobiological basis of motor learning in mammals. Neuroreport 8:i–vi
- 3. Bernheimer H, Birkmayer W, Hornykiewicz O, Jellinger K, Seitelberger F (1973) Brain dopamine and the syndromes of Parkinson and Huntington. Clinical, morphological and neurochemical correlations. J Neurol Sci 20:415–455
- Blanchet PJ, Konitsiotis S, Chase TN (1998) Amantadine reduces levodopainduced dyskinesias in parkinsonian monkeys. Mov Disord 13:798–802
- Blanchet PJ, Konitsiotis S, Whitmore N, Woodward R, Chase TN (1999) Different effects of subunit specific NMDA antagonists in parkinsonian monkeys. J Pharmacol Exp Ther 290(3):1034–1040
- Blanchet PJ, Metman LV, Mouradian MM, Chase TN (1996) Acute pharmacologic blockade of dyskinesias in Parkinson's disease. Mov Disord 11: 580–581
- 7. Blanchet PJ, Papa SM, Metman LV, Mouradian MM, Chase TN (1997) Modulation of levodopa-induced motor response complications by NMDA antagonists in Parkinson's disease. Neurosci Biobehav Rev 21:447–453
- Boyce S, Clarke CE, Luquin R, Peggs D, Robertson RG, Mitchell IJ, Sambrook MA, Crossman AR (1990) Induction of chorea and dystonia in parkinsonian primates. Mov Disord 5:3–7
- 9. Bravi D, Mouradian MM, Roberts JW, Davis TL, Sohn YH, Chase TN (1994) Wearing-off fluctuations in Parkinson's disease: contribution of postsynaptic mechanisms. Ann Neurol 36: 27–31
- Cain DP (1997) LTP, NMDA, genes and learning. Curr Opin Neurobiol 7: 235–242

- 11. Calabresi P, Pisani A, Mercuri NB, Bernardi G (1996) The corticostriatal projection: from synaptic plasticity to basal ganglionic disorders. Trends Neurosci 19:19–24
- 12. Calabresi P, Saiardi A, Pisani A, Baik JH, Centonze D, Mercuri NB, Bernardi G, Borrelli E (1997) Abnormal synaptic plasticity in the striatum of mice lacking dopamine D2 receptors. J Neurosci 7:4536–4544
- 13. Carroll CB, Holloway V, Brotchie JM, Mitchell IJ (1995) Neurochemical and behavioural investigations of the NMDA receptor-associated glycine site in the rat striatum: functional implications for treatment of parkinsonian symptoms. Psychopharmacology (Berl) 119:55–65
- 14. Castro-Alamancos MA, Donoghue JP, Connors BW (1995) Different forms of synaptic plasticity in somatosensory and motor areas of the neocortex. J Neurosci 15:5324–5333
- Cepeda C, Levine MS (1998) Dopamine and N-methyl-D-aspartate receptor interactions in the neostriatum. Dev Neurosci 20:1–18
- 16. Cervo L, Samanin R (1996) Effects of dopaminergic and glutamatergic receptor antagonists on the establishment and expression of conditioned locomotion to cocaine in rats. Brain Res 731: 31–38
- 17. Chase TN, Oh JD (1999) Striatal mechanisms contributing to the pathogenesis of parkinsonian signs and levodopa-associated motor complications. Ann Neurol (in press)
- Chase TN (1970) Cerebrospinal fluid monoamine metabolites and peripheral decarboxylase inhibitors in parkinsonism. Neurology 20(Suppl):36–40
- Chen Q, Reiner A (1996) Cellular distribution of the NMDA receptor NR2A/2B subunits in the rat striatum. Brain Res 743:346–352
- 20. Clarke CE, Sambrook MA, Mitchell IJ, Crossman ARH (1987) Levodopa-induced dyskinesia and response fluctuations in primates rendered parkinsonian with 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine (MPTP). J Neurol Sci 78:273–280

- 21. Danysz W, Parsons CG, Kornhuber J, Schmidt WJ, Quack G (1997) Aminoadamantanes as NMDA receptor antagonists and antiparkinsonian agents-preclinical studies. Neurosci Biobehav Rev 21:455–468
- 22. Engber TM, Boldry RC, Chase TN (1991) The kappa-opioid receptor agonist spiradoline differentially alters the rotational response to dopamine D1 and D2 agonists. Eur J Pharmacol 200: 171–173
- 23. Engber TM, Boldry RC, Kuo S, Chase TN (1992) Dopaminergic modulation of striatal neuropeptides: differential effects of D1 and D2 receptor stimulation on somatostatin, neuropeptide Y, neurotensin, dynorphin and enkephalin. Brain Res 581:261–268
- 24. Engber TM, Papa SM, Boldry RC, Chase TN (1994) NMDA receptor blockade reverses motor response alterations induced by levodopa. Neuroreport 5:2586–2588
- 25. Engber TM, Susel Z, Kuo S, Gerfen CR, Chase TN (1991) Levodopa replacement therapy alters enzyme activities in striatum and neuropeptide content in striatal output regions of 6-hydroxydopamine lesioned rats. Brain Res 552:113–118
- 26. Ferraro L, Antonelli T, O'Connor WT, Fuxe K, Soubrie P, Tanganelli S (1998) The striatal neurotensin receptor modulates striatal and pallidal glutamate and GABA release: functional evidence for a pallidal glutamate-GABA interaction via the pallidal-subthalamic nucleus loop. J Neurosci 18: 6977–6989
- 27. Gerfen CR, Engber TM, Mahan LC, Susel Z, Chase TN, Monsma FJ Jr, Sibley DR (1990) D1 and D2 dopamine receptor-regulated gene expression of striatonigral and striatopallidal neurons. Science 250:1429–1432
- 28. Gomez-Mancilla B, Bedard PJ (1993) Effect of nondopaminergic drugs on Ldopa-induced dyskinesias in MPTPtreated monkeys. Clin Neuropharmacol 16:418–427

- 29. Grace AA, Bunney BS (1984) The control of firing pattern in nigral dopamine neurons: single spike firing. J Neurosci 4:2866–2876
- 30. Graybiel AM, Aosaki T, Flaherty AW, Kimura M (1994) The basal ganglia and adaptive motor control. Science 265:1826–1831
- Greenamyre JT, O'Brien CF (1991) Nmethyl-D-aspartate antagonists in the treatment of Parkinson's disease. Arch Neurol 48:977–981
- 32. Gurd JW (1997) Protein tyrosine phosphorylation: implications for synaptic function. Neurochem Int 31:635–649
- 33. Guttman M, Burkholder J, Kish SJ, Hussey D, Wilson A, DaSilva J, Houle S (1997) [11C]RTI-32 PET studies of the dopamine transporter in early dopanaive Parkinson's disease: implications for the symptomatic threshold. Neurology 48:1578–1583
- 34. Henry B, Crossman AR, Brotchie JM (1999) Effect of repeated L-DOPA, bromocriptine, or lisuride administration on preproenkephalin-A and preproenkephalin-B mRNA levels in the striatum of the 6-hydroxydopamine-lesioned rat. Exp Neurol 155:204–220
- 35. Herrero MT, Augood SJ, Hirsch EC, Javoy-Agid F, Luquin MR, Agid Y, Obeso JA, Emson PC (1995) Effects of L-DOPA on preproenkephalin and preprotachykinin gene expression in the MPTP-treated monkey striatum. Neuroscience 68:1189–1198
- 36. Hisatsune C, Umemori H, Inoue T, Michikawa T, Kohda K, Mikoshiba K, Yamamoto T (1997) Phosphorylationdependent regulation of N-methyl-Daspartate receptors by calmodulin. J Biol Chem 272:20805–20810
- 37. Hornykiewicz O (1998) Biochemical aspects of Parkinson's disease. Neurology 51 [Suppl 2]:S2–S9
- 38. Izquierdo I, Medina JH (1997) Memory formation: the sequence of biochemical events in the hippocampus and its connection to activity in other brain structures. Neurobiol Learn Mem 68:285–316
- 39. Kearney JA, Becker JB, Frey KA, Albin RL (1998) The role of nigrostriatal dopamine in metabotropic glutamate agonist-induced rotation. Neuroscience 87:881–891
- 40. Kita H (1996) Glutamatergic and GABAergic postsynaptic responses of striatal spiny neurons to intrastriatal and cortical stimulation recorded in slice preparations. Neuroscience 70: 925–940
- 41. Klockgether T, Turski L, Honore T, Zhang Z, Gash DM, Kurlan R, Greenamyre JT (1991) The AMPA receptor antagonist NBQX has antiparkinsonian effects in monoaminedepleted rats and MPTP-treated monkeys. Ann Neurol 30:717–723

- 42. Kohr G, Seeburg PH (1996) Subtypespecific regulation of recombinant NMDA receptor-channels by protein tyrosine kinases of the src family. J Physiol (Lond) 492:445–452
- 43. Konitsiotis S, Blanchet PJ, Lamers E, Verhagen L, Chase TN (1999) The effects of AMPA receptor modulation on parkinsonian symptoms and levodopainduced dyskinesias in MPTP monkeys. Neurology (in press)
- 44. Kotter R (1994) Postsynaptic integration of glutamatergic and dopaminergic signals in the striatum. Prog Neurobiol 44:163–196
- 45. Le Moine C, Bloch B (1995) D1 and D2 dopamine receptor gene expression in the rat striatum: sensitive cRNA probes demonstrate prominent segregation of D1 and D2 mRNAs in distinct neuronal populations of the dorsal and ventral striatum. J Comp Neurol 355: 418–426
- 46. Loschmann PA, Lange KW, Kunow M, Rettig KJ, Jahnig P, Honore T, Turski L, Wachtel H, Jenner P, Marsden CD (1991) Synergism of the AMPA-antagonist NBQX and the NMDA-antagonist CPP with L-dopa in models of Parkinson's disease. J Neural Transm [P-D Sect] 3:203–213
- 47. Luquin MR, Obeso JA, Laguna J, Guillen J, Martinez-Lage JM (1993) The AMPA receptor antagonist NBQX does not alter the motor response induced by selective dopamine agonists in MPTP-treated monkeys, Eur J Pharmacol 235:297–300
- 48. Marin C, Engber TM, Chaudhuri P, Peppe A, Chase TN (1996) Effects of kappa receptor agonists on D1 and D2 dopamine agonist and antagonist-induced behaviors. Psychopharmacology (Berl) 123:215–221
- 49. Marin C, Papa SM, Engber TM, Bonastre M, Tolosa E, Chase TN (1996) MK801 prevents levodopa-induced motor response alterations in parkinsonian rats. Brain Res 736:202–205
- 50. Marin C, Tolosa E, Chase TN (1999) NMDA and Non NMDA antagonist effects on levodopa associated response alterations in parkinsonian rats. Synapse (in press)
- 51. McEachern JC, Shaw CA (1996) An alternative to the LTP orthodoxy: a plasticity-pathology continuum model. Brain Res Brain Res Rev 22:51–92
- 52. Melamed E, Hefti F, Wurtman RJ (1980) Nonaminergic striatal neurons convert exogenous L-dopa to dopamine in parkinsonism. Ann Neurol 8:558–563
- 53. Menegoz M, Lau LF, Herve D, Huganir RL, Girault JA (1995) Tyrosine phosphorylation of NMDA receptor in rat striatum: effects of 6-OHdopamine lesions. Neuroreport 7:125– 128

- 54. Mitchell IJ, Carroll CB (1997) Reversal of parkinsonian symptoms in primates by antagonism of excitatory amino acid transmission: potential mechanisms of action. Neurosci Biobehav Rev 21:469–475
- 55. Miyawaki E, Lyons K, Pahwa R, Troster AI, Hubble J, Smith D, Busenbark K, McGuire D, Michalek D, Koller WC (1997) Motor complications of chronic levodopa therapy in Parkinson's disease. Clin Neuropharmacol 20:523–530
- 56. Morissette M, Goulet M, Soghomonian JJ, Blanchet PJ, Calon F, Bedard PJ, DiPaolo T (1997) Preproenkephalin mRNA expression in the caudate-putamen of MPTP monkeys after chronic treatment with the D2 agonist U91356A in continuous or intermittent mode of administration: comparison with L-DOPA therapy. Brain Res Mol Brain Res 49:55–62
- 57. Morrish PK, Rakshi JS, Bailey DL, Sawle GV, Brooks DJ (1998) Measuring the rate of progression and estimating the preclinical period of Parkinson's disease with [18F]dopa PET. J Neurol Neurosurg Psychiatry 64 (March):314–319
- 58. Mouradian MM, Heuser IJ, Baronti F, Fabbrini G, Juncos JL, Chase TN (1989) Pathogenesis of dyskinesias in Parkinson's disease. Ann Neurol 25: 523–526
- 59. Ng LK, Chase TN, Colburn RW, Kopin IJ (1972) L-dopa in Parkinsonism. A possible mechanism of action. Neurology 22:688–696
- Nicoll RA, Malenka RC (1995) Contrasting properties of two forms of long-term potentiation in the hippocampus. Nature 377:115–118
- 61. Oh JD, Del Dotto P, Chase TN (1997) Protein kinase A inhibitor attenuates levodopa-induced motor response alterations in the hemi-parkinsonian rat. Neurosci Lett 228:5–8
- 62. Oh JD, Russell D, Vaughan CL, Chase TN (1998) Enhanced tyrosine phosphorylation of striatal NMDA receptor subunits: effect of dopaminergic denervation and levodopa administration. Brain Res 813:150–159
- 63. Oh JD, Vaughan CL, Chase TN (1999) Effect of dopamine denervation and dopamine agonist administration on serine phosphorylation of striatal NMDA receptor subunits. Brain Res 813:433–442
- 64. Ozawa S, Kamiya H, Tsuzuki K (1998) Glutamate receptors in the mammalian central nervous system. Prog Neurobiol 54:581–618
- 65. Papa SM, Boldry RC, Engber TM, Kask AM, Chase TN (1995) Reversal of levodopa-induced motor fluctuations in experimental parkinsonism by NMDA receptor blockade. Brain Res 701:13–18

- 66. Papa SM, Chase TN (1996) Levodopainduced dyskinesias improved by a glutamate antagonist in Parkinsonian monkeys. Ann Neurol 39:574–578
- 67. Papa SM, Engber TM, Kask AM, Chase TN (1994) Motor fluctuations in levodopa treated parkinsonian rats: relation to lesion extent and treatment duration. Brain Res 662:69–74
- 68. Parent A, Asselin MC, Cote PY (1996) Dopaminergic regulation of peptide gene expression in the striatum of normal and parkinsonian monkeys. Adv Neurol 69:73–77
- Quinn NP (1998) Classification of fluctuations in patients with Parkinson's disease. Neurology 51 [Suppl 2]:S25–S29
- 70. Sacaan AI, Bymaster FP, Schoepp DD (1992) Metabotropic glutamate receptor activation produces extrapyramidal motor system activation that is mediated by striatal dopamine. J Neurochem 59:245–251
- 71. Schultz W (1994) Behavior-related activity of primate dopamine neurons. Rev Neurol (Paris) 150:634–639
- 72. Seasack SR, Aoki C, Pickel VM (1994) Ultrastructural localization of D2 receptor-like immunoreactivity in midbrain dopamine neurons and their striatal targets. J Neurosci 14:88–106
- 73. Smith AD, Bolam JP (1990) The neural network of the basal ganglia as revealed by the study of synaptic connections of identified neurones. Trends Neurosci 13:259–265
- 74. Stoof JC, Booij J, Drukarch B, Wolters EC (1992) The anti-parkinsonian drug amantadine inhibits the N-methyl-Daspartic acid-evoked release of acetylcholine from rat neostriatum in a noncompetitive way. Eur J Pharmacol 213: 439–443

- 75. Suen PC, Wu K, Xu JL, Lin SY, Levine ES, Black IB (1998) NMDA receptor subunits in the postsynaptic density of rat brain: expression and phosphorylation by endogenous protein kinases. Brain Res Mol Brain Res 59: 215–228
- 76. Suzuki T, Okumura-Noji K (1995) NMDA receptor subunits epsilon 1 (NR2A) and epsilon 2 (NR2B) are substrates for Fyn in the postsynaptic density fraction isolated from the rat brain. Biochem Biophys Res Commun 216: 582–588
- 77. Tan SE, Chen SS (1997) The activation of calcium/calmodulin-dependent protein-kinase II after glutamate or potassium stimulation in hippocampal slices. Brain Res Bull 43:269–273
- 78. Testa CM, Standaert DG, Young AB, Penney JB Jr (1994) Metabotropic glutamate receptor mRNA expression in the basal ganglia of the rat. J Neurosci 14:3005–3018
- 79. Tingley WG, Ehlers MD, Kameyama K, Doherty C, Ptak JB, Riley CT, Huganir RL (1997) Characterization of protein kinase A and protein kinase C phosphorylation of the N-methyl-D-aspartate receptor NR1 sub-unit using phosphorylation site-specific antibodies. J Biol Chem 72:5157–5166
- Ulas J, Cotman C (1996) Dopaminergic denervatiom of striatum results in elevated expression of NR2A subunit. Neuroreport 7:1789–1793
- 81. Verhagen Metman L, Blanchet PJ, van den Munckhof P, Del Dotto P, Natte R, Chase TN (1998) A trial of dextromethorphan in parkinsonian patients with motor response complications. Mov Disord 13:414–417

- 82. Verhagen Metman L, Del Dotto P, Natte R, van den Munckhof P, Chase TN (1998) Dextromethorphan improves levodopa-induced dyskinesias in Parkinson's disease. Neurology 51: 203–206
- 83. Verhagen Metman L, Del Dotto P, van den Munckhof P, Fang J, Mouradian MM, Chase TN (1998) Amantadine as treatment for dyskinesias and motor fluctuations in Parkinson's disease. Neurology 50:1323–1326
- 84. Wachtel SR, Abercrombie ED (1994) L-3,4-dihydroxyphenylalanine-induced dopamine release in the striatum of intact and 6-hydroxydopamine-treated rats: differential effects of monoamine oxidase A and B inhibitors. J Neurochem 63:108–117
- 85. Wang JH, Ko GY, Kelly PT (1997) Cellular and molecular bases of memory: synaptic and neuronal plasticity. J Clin Neurophysiol 14:264–293
- 86. Wang YT, Salter MW (1994) Regulation of NMDA receptors by tyrosine kinases and phosphatases. Nature 369: 233–235
- 87. Wollmuth LP, Kuner T, Seeburg PH, Sakmann B (1996) Differential contribution of the NR1- and NR2A-subunits to the selectivity filter of recombinant NMDA receptor channels. J Physiol (Lond) 491:779–797
- 88. Yu XM, Askalan R, Keil GJ 2nd, Salter MW (1997). NMDA channel regulation by channel-associated protein tyrosine kinase Src. Science 275: 674–678