

# Molecular Evidence on the Evolutionary and Biogeographical Patterns of European Cyprinids

Rafael Zardoya, Ignacio Doadrio

Museo Nacional de Ciencias Naturales, CSIC, José Gutiérrez Abascal 2, 28006 Madrid, Spain

Received: 2 February 1999 / Accepted: 16 March 1999

**Abstract.** The phylogenetic relationships of 106 European cyprinid taxa were determined based on the complete nucleotide sequence (1140 bp) of the mitochondrial cytochrome *b* gene. The molecular phylogeny was used (1) to revise the current systematics of European cyprinids, (2) to establish the phylogenetic utility of traditional morphological characters that are widely used in Cyprinidae systematics, and (3) to discuss alternative hypotheses on the biogeography of the family in Europe. The age of the major lineages within European cyprinids was tentatively estimated with a molecular clock and showed full agreement with the fossil record of the group. Moreover, the results provided unambiguous evidence for a close phylogenetic affinity of some Caucasian and Greek endemic cyprinid taxa (e.g., *B. capito* and *B. brachycephalus* and *Leuciscus keadicus*, *Barbus graecus*, and *B. albanicus*, respectively) to Iberian and North African, but not Central European, cyprinids. The existence of such unexpected phylogenetic relationships refutes the classical hypothesis on the biogeography of European cyprinids, which assumes a dispersal of the cyprinid fauna from central Europe to southern Europe and northern Africa during the Miocene (and, hence, predicts a close phylogenetic relationship of all Caucasian, Greek, Iberian, and North African cyprinids to central European taxa). Instead, the existence of a Mediterranean realm independent of the central European route seems plausible based on the molecular evidence. It is likely that the new biogeographical scenario proposed here might apply to other primary freshwater European

animals with low dispersal abilities, including fish, amphibians, and invertebrates.

**Key words:** Molecular phylogeny — Cytochrome *b* — Biogeography — Cyprinids

## Introduction

Cyprinids, the largest and most successful family of primary freshwater fish in Eurasia, Africa, and northern America, are a good model for comprehending the evolutionary mechanisms driving the diversification and distribution of species. Primary freshwater fish are restricted to river and lake drainage systems and show little capacity for transwatershed dispersal. Thus, their distribution closely reflects their biogeographical history.

In particular, European cyprinids have an interesting pattern of distribution, in which numerous endemic species are found on the Iberian Peninsula and in southern Greece, with a relatively uniform fauna in Central Europe. Traditionally, it is believed that European cyprinids originated in eastern Asia and subsequently spread to Siberia during the Eocene (Banarescu 1989, 1992). However, because Europe was separated from Siberia by the shallow Ob Sea, their dispersal to Europe was impeded. During the Oligocene, cyprinids were able to colonize Europe because of the uplift of the Urals but ended once these mountains were formed. Three alternative hypotheses have been proposed to explain the further dispersion of cyprinids across Europe during the Miocene.

The first, classical, hypothesis proposes that cyprinids spread across central Europe via river connections to the

southern part of the continent and northern Africa (across the Gibraltar strait) until the Pliocene (Banarescu 1960, 1989, 1992; Almaça 1976, 1988). Subsequent isolation of the Iberian Peninsula and southern Greece from the rest of the continent would have been responsible for their rich endemic fauna. During most of the Miocene, southern Italy was below sea level (Steininger and Rögl 1984), and therefore, its cyprinid fauna would have a more recent origin (Plio-Pleistocene) (Bianco 1990). Furthermore, the uniform cyprinid fauna of central Europe was seriously depleted during the glacial periods and replaced mainly with Danubian cyprinid fauna during interglacial and postglacial periods (Banarescu 1989, 1992).

The second hypothesis (Doadrio 1990, 1994) is based on the close affinities of Asian, North African, southern Greek, and Iberian barbels. According to this hypothesis, the formation of the actual North African coast by land-mass movements across the Mediterranean Sea in the early Pliocene favored the South Mediterranean vicariance of barbels and the subsequent colonization of Northern Africa and the Iberian Peninsula (Doadrio 1990, 1994).

Finally, the third hypothesis (Bianco 1990) proposes that a major dispersal of cyprinids occurred around the circum-Mediterranean region during the Messinian salinity crisis (6–5 MYA), when the Mediterranean basin almost dried up and was subsequently refilled with fresh water from the Sarmatic Sea (Paratethys) (Hsü et al. 1977). According to this hypothesis, the so-called Lago Mare phase of the Mediterranean would have allowed the dispersal of freshwater fish across the Mediterranean and would be responsible for the actual high level of endemics on the Iberian Peninsula and in Southern Greece (Bianco 1990). However, the existence of the Lago Mare phase of the Mediterranean is highly controversial, and no fossil data support the complete desiccation of the Mediterranean basin (Steininger and Rögl 1984).

The above three hypotheses proposed to explain the colonization of Europe by cyprinids during the Miocene are based mainly on the actual distribution patterns, fossil record, and morphological similarities of European cyprinid taxa rather than on their phylogenetic relationships (but see Doadrio 1990, 1994). However, to discern alternative hypotheses on the biogeographical origin of European cyprinids, it is necessary to understand their phylogenetic relationships. The disjunct distribution of European cyprinids can be interpreted only within a phylogenetic framework that establishes monophyletic groups within the taxon.

In the present study we have examined the phylogenetic relationships of representatives of most of the cyprinid genera occurring in Europe to revise the systematics of the family, to evaluate the phylogenetic utility of the morphological characters that have traditionally been

used in inferring phylogenetic relationships within Cyprinidae, and to test among the three alternative biogeographical hypotheses. Complete mitochondrial cytochrome *b* sequences (1140 bp) of 89 cyprinid taxa from the Iberian Peninsula, Greece, and the Caucasus were determined (see Table 1) and analyzed with the currently used methods of phylogenetic inference. Cytochrome *b* sequences of 15 cyprinids from central Europe (Briolay et al. 1998) (see Table 1), *Cyprinus carpio* (Chang et al. 1994), and *Carassius auratus* (Zardoya and Doadrio 1998) were also included in the phylogenetic analyses.

## Materials and Methods

### Laboratory Procedures

Fish samples representing 89 taxa of European cyprinids (one specimen per taxon) were collected on the Iberian Peninsula (Zardoya and Doadrio 1998), in Greece (Zardoya et al. 1999), and in the Caucasus (this paper) (Table 1). Total cellular DNA was extracted from muscle following standard phenol/chloroform protocols (Townner 1991). Primers specifically designed for cyprinids (Schmidt and Gold 1993; Zardoya and Doadrio 1998) were used to amplify, via PCR, the entire cytochrome *b* gene. Thirty-five to forty cycles of PCR (denaturing at 94°C for 60 s, annealing at 45–50°C for 60 s, and extension at 72°C for 105–180 s) were performed in 25- $\mu$ l reactions containing 67 mM Tris-HCl, pH 8.3, 1.5 mM MgCl<sub>2</sub>, a 0.4 mM concentration of each dNTP, a 2.5  $\mu$ M concentration of each primer, template DNA (10–100 ng), and Taq DNA polymerase (1 U; Promega). PCR products were cloned using the pGEM-T vector (Promega) into *E. coli* JM109 and sequenced using the FS-Taq Dye Deoxy Terminator cycle-sequencing kit (Applied Biosystems Inc.) on an automated DNA sequencer (Applied Biosystems 377) following the manufacturer's instructions. DNA sequences of both strands were obtained using M13 universal (forward and reverse) sequencing primers.

### Phylogenetic Analyses

The new sequences were aligned with the complete cytochrome *b* sequences of 15 French (Briolay et al. 1998) cyprinids, *Cyprinus carpio* (Chang et al. 1994), and *Carassius auratus* (Zardoya and Doadrio 1998). One Characidae (*Astyanax fasciatus*) (Zardoya and Doadrio 1998) and one Balitoridae (*Crossostoma lacustre*) (Tzeng et al. 1992) were used as outgroup taxa. Alignment was based on the inferred amino acid sequence. No ambiguous alignments were found and no gaps were postulated. All codon positions were included in the phylogenetic analyses.

Maximum-parsimony (MP) analyses [PAUP\* version d64 (Swofford 1998)] were performed using heuristic searches (TBR branch swapping; MULPARS option in effect) with 10 random stepwise additions of taxa. Transversions (Tv) were given 4 or 10 times the weight of transitions (Ts). The ancestral character-state reconstruction of the evolution of the number of pharyngeal tooth rows was performed using MacClade version 3.06 (Maddison and Maddison 1992). Neighbor-joining (NJ) (Saitou and Nei 1987) analyses based on HKY85 corrected distance matrices (using empirical Ts/Tv ratios and base frequencies) were performed with PAUP\* version d64 (Swofford 1998). The robustness of the inferred MP and NJ trees was tested by bootstrapping (Felsenstein 1985) (as implemented in PAUP\*, with 100 and 500 pseudoreplications, respectively). Maximum-likelihood (ML) analyses (based on the HKY85 model with empirical Ts/Tv ratios and base

**Table 1.** Cyprinid taxa, sampling drainages, and GenBank accession numbers

Taxa	Drainage	Genbank No.
Greece		
<i>Alburnoides bipunctatus ohridanus</i>	Aoos	AF090740
<i>Alburnoides b. ohridanus</i>	Prespa	AF090741
<i>Alburnoides b. strymonicus</i>	Strymon	AF090742
<i>Alburnus alburnus macedonicus</i>	Doirani	AF090743
<i>Alburnus a. strumicae</i>	Strymon	AF090745
<i>Alburnus a. thessalicus</i>	Pinios	AF090744
<i>Barbus albanicus</i>	Arachthos	AF090779
<i>Barbus barbus macedonicus</i>	Axios	AF090780
<i>Barbus b. thessalus</i>	Pinios	AF090781
<i>Barbus cyclolepis cyclolepis</i>	Evros	AF090782
<i>Barbus c. sperchiensis</i>	Sperchios	AF090783
<i>Barbus c. strumicae</i>	Agiaki	AF090784
<i>Barbus euboicus</i>	Manikiotiko	AF090785
<i>Barbus graecus</i>	Kifissos	AF090786
<i>Barbus peloponnesius</i>	Alphios	AF090787
<i>Barbus petenyi</i>	Aliakmon	AF090788
<i>Barbus petenyi</i>	Vegoritis	AF090789
<i>Barbus prespensis</i>	Prespa	AF090790
<i>Barbus rebeli</i>	Aoos	AF090791
<i>Chalcalburnus belvica</i>	Prespa	AF090746
<i>Chondrostoma prespensis</i>	Prespa	AF090747
<i>Chondrostoma vardarensis</i>	Aoos	AF090748
<i>Chondrostoma vardarensis</i>	Aoos	AF090749
<i>Gobio banarescui</i>	Aliakmon	AF090751
<i>Gobio gobio balcanicus</i>	Gallikos	AF090750
<i>Leuciscus cephalus macedonicus</i>	Nestos	AF090752
<i>Leuciscus c. prespensis</i>	Prespa	AF090753
<i>Leuciscus c. vardarensis</i>	Sperchios	AF090754
<i>Leuciscus cephalus</i>	Manikiotiko	AF090755
<i>Leuciscus peloponnesius</i>	Thyamis	AF090756
<i>Leuciscus peloponnesius</i>	Alphios	AF090757
<i>Leuciscus p. moreoticus</i>	Stymphalia	AF090758
<i>Leuciscus borysthenticus</i>	Fotolivos	AF090759
<i>Leuciscus keadicus</i>	Evrotas	AF090760
<i>Pachychilon macedonicus</i>	Axios	AF090761
<i>Pachychilon pictus</i>	Aoos	AF090762
<i>Phoxinellus prespensis</i>	Prespa	AF090763
<i>Pseudophoxinus stymphalicus</i>	Stymphalia	AF090766
<i>Pseudophoxinus stymphalicus</i>	Trichonis	AF090767
<i>Pseudophoxinus st. marathonicus</i>	Kifissos	AF090768
<i>Pseudophoxinus st. thesproticus</i>	Louros	AF090769
<i>Rutilus prespensis</i>	Prespa	AF090771
<i>Rutilus rutilus</i>	Strymon	AF090772
<i>Rutilus ylikiensis</i>	Kifissos	AF090773
<i>Rutilus ylikiensis</i>	Trichonis	AF090774
<i>Scardinius acarnanicus</i>	Trichonis	AF090775
<i>Telestes beoticus</i>	Kifissos	AF090770
<i>Telestes pleurobipunctatus</i>	Arachthos	AF090764
<i>Telestes p. alfiensis</i>	Alphios	AF090765
<i>Tropidophoxinellus hellenicus</i>	Pinios	AF090776
<i>Tropidophoxinellus spartiaticus</i>	Evrotas	AF090777
<i>Vimba melanops</i>	Strymon	AF090778
Portugal		
<i>Chondrostoma lusitanicum</i>	Arade	AF045986
<i>Chondrostoma macrolepidotus</i>	Mondego	AF045980
Spain		
<i>Anaocypris hispanica</i>	Guadiana	AF045978
<i>Barbus bocagei</i>	Duero	AF045969
<i>Barbus comizo</i>	Tajo	AF045967
<i>Barbus steindachneri</i>	Guadiana	AF045968
<i>Barbus graellsii</i>	Ebro	AF045973
<i>Barbus guiraonis</i>	Buyent	AF045972

**Table 1.** Continued

Taxa	Drainage	Genbank No.
<i>Barbus haasi</i>	Ebro	AF045976
<i>Barbus meridionalis</i>	Tordera	AF045977
<i>Barbus microcephalus</i>	Guadiana	AF045971
<i>Barbus sclateri</i>	Guadalquivir	AF045970
<i>Chondrostoma arcasii</i>	Duero	AF045979
<i>Chondrostoma lemmingii</i>	Guadiana	AF045987
<i>Chondrostoma lemmingii</i>	Guadiana	AF045988
<i>Chondrostoma lemmingii</i>	Guadalquivir	AF045989
<i>Chondrostoma polylepis</i>	Tajo	AF045982
<i>Chondrostoma p. duriensis</i>	Duero	AF045983
<i>Chondrostoma p. willkommii</i>	Guadalquivir	AF045984
<i>Chondrostoma toxostoma</i>	Ebro	AF045985
<i>Gobio gobio</i>	Tajo	AF045996
<i>Iberocypris palaciosi</i>	Guadalquivir	AF045990
<i>Leuciscus carolitertii</i>	Duero	AF045994
<i>Leuciscus cephalus</i>	Ebro	AF045995
<i>Leuciscus alburnoides</i>	Guadiana	AF045992
<i>Leuciscus pyrenaicus</i>	Guadiana	AF045991
Algeria		
<i>Barbus callensis</i>	Kebir	AF045974
France		
<i>Abramis brama</i>	Saone	Y10441
<i>Alburnoides bipunctatus</i>	Saone	Y10445
<i>Alburnus alburnus</i>	Rhone	Y10443
<i>Barbus barbus barbus</i>	Durance	Y10450
<i>Blicca bjoerkna</i>	Saone	Y10442
<i>Chondrostoma nasus</i>	Rhone	Z75109
<i>Gobio gobio</i>	Rhone	Y10452
<i>Leucaspis delineatus</i>	Rhone	Y10447
<i>Leuciscus cephalus</i>	Rhone	Y10446
<i>Phoxinus phoxinus</i>	Rhone	Y10448
<i>Pseudorasbora parva</i>	Rhone	Y10453
<i>Rutilus rutilus</i>	Saone	Y10440
<i>Scardinius erythrophthalmus</i>	Rhone	Y10444
<i>Telestes souffia</i>	Saone	Y10439
<i>Tinca tinca</i>	Saone	Y10451
The Caucasus		
<i>Alburnus filippii</i>	Samur	AF095602
<i>Barbus brachycephalus</i>	Terek	AF095603
<i>Barbus capito</i>	Terek	AF045975
<i>Barbus ciscaucasicus</i>	Kuma	AF095604
<i>Barbus tauricus</i>	Kuban	AF095605
<i>Chondrostoma oxyrhynchum</i>	Samur	AF095606
<i>Gobio ciscaucasicus</i>	Uluchaj	AF095607
<i>Hemiculter leucisculus</i>	Sulak	AF095608
<i>Leuciscus cephalus orientalis</i>	Rubas	AF095609
<i>Rutilus caspicus</i>	Samur	AF095610

frequencies) were performed using PUZZLE version 4.0 (Strimmer and von Haeseler 1996) and 10,000 pseudo-replications.

A rate constancy test (two-cluster test) was performed with LINTREE (Takezaki et al. 1995) using the HKY85 distance. Those taxa that showed significantly different substitution rates (at the 5% level) were excluded from further analyses. The remaining taxa were reanalyzed in PUZZLE version 4.0 (Strimmer and von Haeseler 1996) with ML using the clock-like option to obtain a clock-constrained tree (in which all root-to-tip distances have equal value). ML analyses were based on the HKY85 model with empirical Ts/Tv ratios and base frequencies.

European cyprinid biogeography was analyzed in a cladistic framework using Diva 1.1. (Ronquist 1997). To establish a putative biogeographical scenario, the inferred molecular phylogeny as well as the



actual distribution of the taxa was analyzed, and ancestral distributions were optimally reconstructed taking into account both vicariant and dispersal events.

## Results

### *Phylogenetic Relationships of European Cyprinids*

A total of 1140 positions was analyzed, of which 513 were constant sites and 500 were phylogenetically informative sites using the parsimony criterion. An overall Ts/Tv ratio of 4.6 was estimated for this data set. Pair-wise sequence divergence between taxa varied from 0.3 to 25%. Variability among sequences was detected mainly in third codon positions. Substitutions showed some level of saturation in third codon positions (between 25 and 55% sequence divergence) but not in first and second codon positions (not shown).

The phylogenetic analysis of the European cyprinid data set with NJ, using *Astyanax fasciatus* (Characidae) (Zardoya and Doadrio 1998) and *Crossostoma lacustre* (Balitoridae) (Tzeng et al. 1992) as outgroup taxa, recovered the tree shown in Fig. 1. The robustness of this tree was confirmed by bootstrapping (Felsenstein 1985) (Fig. 1). Two main clades corresponding to the subfamilies Cyprininae and Leuciscinae (Cavender and Coburn 1992) were found (Fig. 1). Moreover, at least two lineages of *Barbus* (*Barbus* s.s. and *Luciobarbus*), one of *Hemiculter*, one of *Tinca*, one of *Gobio*, one of *Phoxinus*, one of *Pseudophoxinus* + *Phoxinellus*, and nine of Leuciscinids (including *Pachychilon*, *Pseudophoxinus*, *Scardinius*, *Tropidophoxinellus*, *Abramis* + *Vimba* + *Blicca*, *Anaocypris* + *Leucaspius* + *Chalcalburnus* + *Alburnus*, *Alburnoides*, *Leuciscus*, *Rutilus* s.str., and *Telestes* s.l. + *Chondrostoma*) were identified (Fig. 1). Interestingly, *Hemiculter*, *Tinca*, *Gobio*, *Phoxinus*, and *Alburnus* (and relatives), which have been traditionally classified in their own subfamilies (for a review see Howes 1991), are recovered as distinct, more or less basal lineages within Leuciscinae (Fig. 1). Moreover, at least two origins for the actual European cyprinid fauna were found. Some taxa, Cyprininae as well as Leuciscinae, clearly revealed a central European origin, whereas others (e.g., *Barbus graecus*, *B. albanicus*, *B. brachycephalus*, Iberian barbels, Iberian *Leuciscus*, and *L. keadicus*) were found to have Mediterranean ties (Fig. 1). MP analyses using Ti:Tv ratios of 4:1 (12 MP trees of 8513 steps) and 10:1 (12 MP trees of 14,564 steps) recovered similar and congruent topologies (not shown).

### *Phylogenetic Utility of Morphological Traits*

Cyprinids have traditionally been diagnosed by a protrusile mouth, toothless jaws, and pharyngeal teeth (Howes 1991). However, these characters have recently been considered plesiomorphic (see Cavender and Coburn

1992; Fink and Fink 1996). Some Cyprinids, mainly cyprinins (with the exception of *Carassius*), are also characterized by the presence of barbels. The original (non-cladistic) diagnoses of cyprinid subfamilies were based largely on the number of pharyngeal tooth rows and the presence or absence of barbels (Howes 1991). To evaluate the phylogenetic utility of such morphological traits, they were mapped onto the molecular phylogeny based on cytochrome *b* nucleotide sequence data. The number of rows of pharyngeal teeth (Rutte 1962) was found to be a fairly good phylogenetic character at the genus level, in which different states could be unambiguously associated (with few exceptions) with different monophyletic assemblages (Fig. 2). On the other hand, the presence or absence of barbels was found to be a poor phylogenetic trait (not shown).

### *Cladogenic Events Within European Cyprinids*

A total of 49 taxa representing the main European cyprinid lineages was subjected to ML analyses. Rate constancy among these taxa was assessed with the two-cluster test (Takezaki et al. 1995) using HKY85 distances. Three taxa, namely, *Pseudophoxinus stymphalicus maratonicus*, *Phoxinellus prespensis*, and *Alburnus filippi*, showed a substitution rate significantly faster (at a 5% level) than the average. These taxa were excluded from subsequent analyses. A clock-constrained tree was constructed by ML based on HKY85 distances (Fig. 3), in which branch lengths can be used to estimate tentative divergence dates.

The molecular clock was calibrated (0.76% per MY) using two important and well-dated geological events: the formation of the strait of Korinthos in the late Pliocene (2.5 MYA), which separates the Peloponnesus from the mainland (Dermitzakis 1990), and the opening of the Gibraltar Strait after the Messinian salinity crisis (5 MYA), which separates North Africa from the Iberian Peninsula. *L. peloponensis* from the Thyamis and Alphios rivers (4.5% nucleotide sequence divergence) and *Telestes pleurobipunctatus* from the Arachthos River and *T. p. alfiensis* from the Alphios River (4.6%) are cyprinid taxa that were separated by the formation of the Strait of Korinthos. On the other hand, the flooding of the Gibraltar Strait prompted the separation of North African barbs such as *Barbus callensis* from Iberian barbs such as *B. bocagei* (7.4%), *B. graellsii* (8.3%), *B. guiraonis*, (8.3%), *B. steindachnerii* (7.5%), *B. comizo* (7.6%), *B. microcephalus* (8.2%), and *B. sclateri* (8.5%). Tentative minimum divergence dates for the main cladogenic events affecting European cyprinids are shown in Table 2. The standard errors associated with these estimates were calculated using the clock-constrained ML tree. According to the results, cyprinids may have originated in the Eocene (Fig. 3 and Table 2). The two main subfamilies, i.e., Cyprininae and Leuciscinae, may have appeared



□ Iberian peninsula    ▨ Greece    ■ Caucasus    ▤ central Europe    ▥ Algeria    ▧ eastern Asia    ▩ Palearctic

**Fig. 1.** Phylogenetic relationships of European cyprinids based on cytochrome *b* sequence data. A 50% majority-rule consensus bootstrap tree obtained with NJ (using HKY85 distances using empirical base composition and transition/transversion ratio), based on 500 pseudo-

replications, is depicted. *Astyanax fasciatus* (Characidae) and *Crossostoma lacustre* (Balitoridae) were used as outgroup. Nodes with bootstrap values below 50% were forced to collapse and yield polytomies. MP analyses arrived at similar and congruent trees (see text).

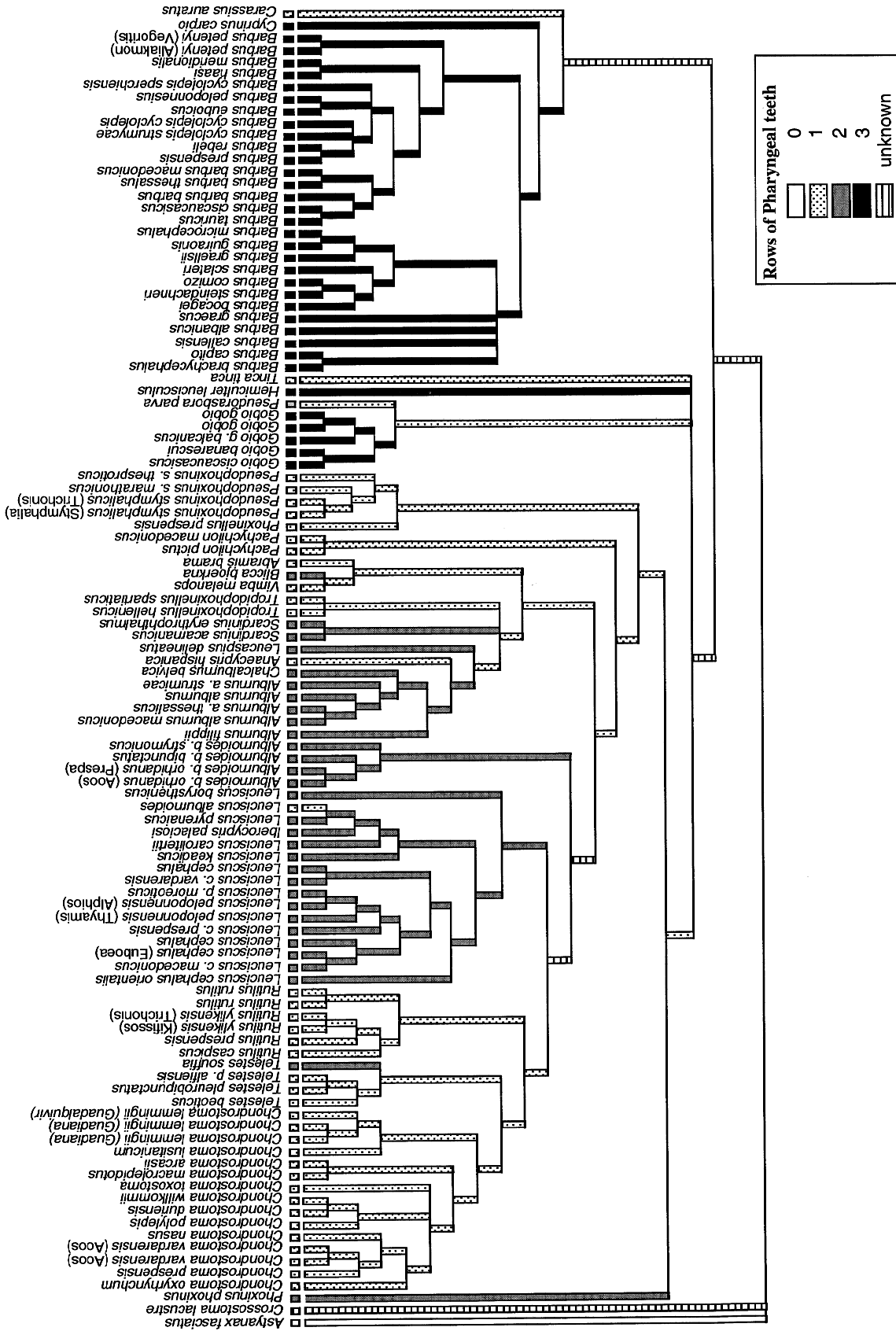
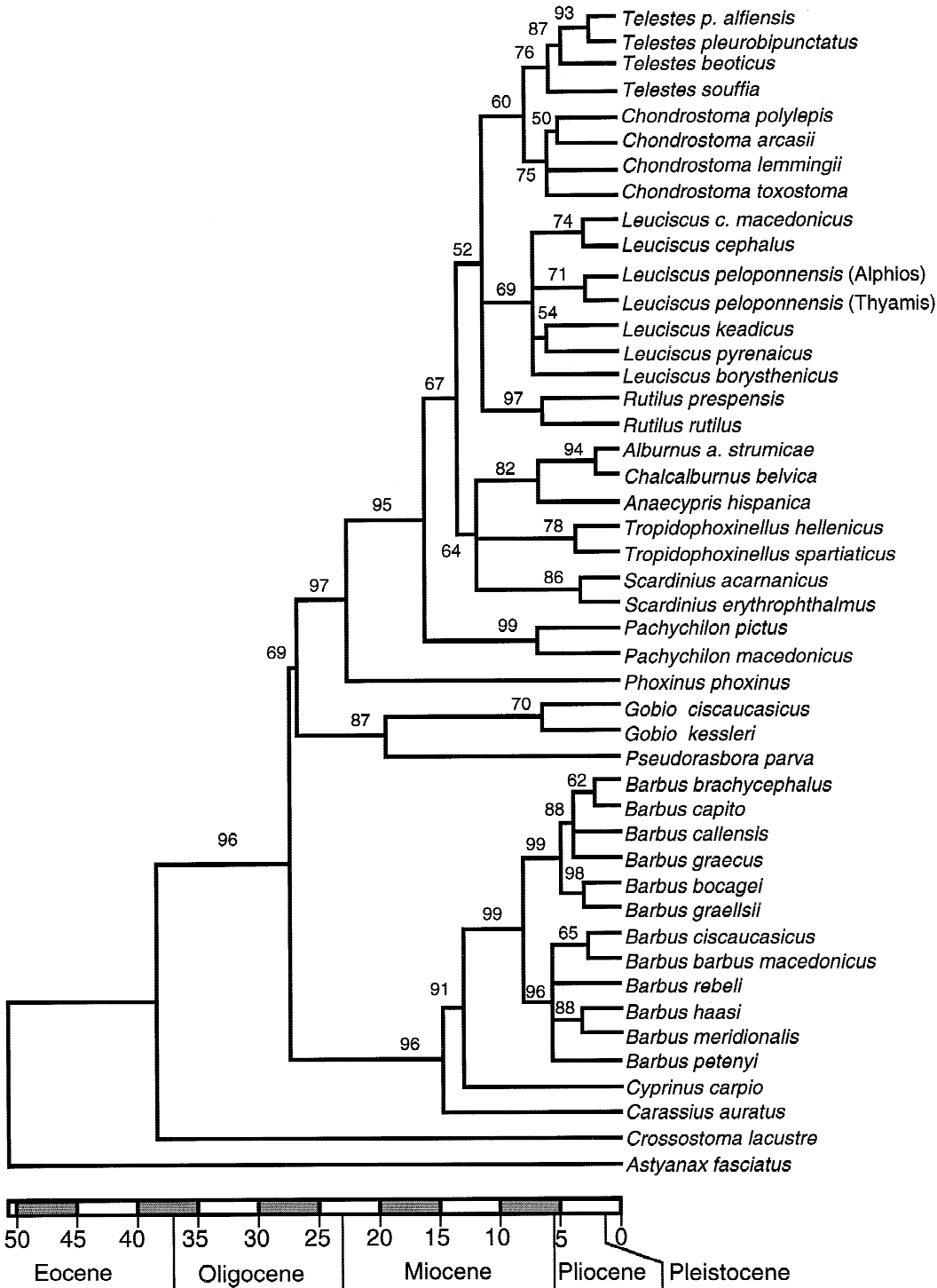


Fig. 2. Evolution of the number of rows of pharyngeal teeth in European cyprinids. The number of rows (zero to three) of pharyngeal teeth of each taxa were mapped onto the cytochrome *b* molecular phylogeny. Ancestral states were reconstructed with the maximum-parsimony method using MacClade (Maddison and Maddison 1992).



**Fig. 3.** Clock-constrained ML tree showing the major cladogenetic events in the history of European cyprinids. The tree was constructed on the assumption of a homogeneous rate of sequence divergence among taxa [as tested by the two-cluster test (Takezaki et al. 1995)]. The scale bar below the tree shows the time scale resulting from a

calibration of the molecular clock (0.76% per MY) based on the formation of the Strait of Korinthos and the opening of the Gibraltar Strait (see text). Numbers in nodes are quartet puzzling (Strimmer and von Haeseler 1996) support values based on 10,000 pseudo-replications.

in the mid-Oligocene and radiated during the late Oligocene and Miocene. Major speciation events resulting in the current European cyprinid fauna occurred largely during the Pliocene (Fig. 3 and Table 2).

**Discussion**

The phylogenetic analysis of the cytochrome *b* nucleotide sequence data supports the traditional subdivision of

**Table 2.** Estimated dates of separation of the major European cyprinid lineages

Cladogenetic event	Date (MYA)	(±SE)
Separation of North African, Greek, and Caucasian <i>Luciobarbus</i>	3.8	(0.3)
Separation of Iberian <i>Luciobarbus</i>	5.0	(0.3)
Separation of <i>L. keadicus</i> from <i>L. pyrenaicus</i>	6.0	(0.4)
Separation of <i>Anaocypris</i>	6.8	(0.6)
Radiation of <i>Leuciscus</i> ( <i>Squalius</i> )	7.2	(0.3)
Separation of Iberian <i>Chondrostoma</i>	7.8	(0.3)
Separation of <i>Barbus</i> and <i>Luciobarbus</i>	8.1	(0.4)
Origin of <i>Leuciscus</i> ( <i>Squalius</i> )	11.4	(0.4)
Major radiation within leuciscins	13.6	(0.5)
Separation of <i>Pachychilon</i>	16.4	(0.8)
Separation of phoxinins	23.0	(1.0)
Separation of gobionins (including <i>Pseudorasbora</i> )	27.2	(0.9)
Separation of Cyprininae and Leuciscinae	27.7	(0.9)
Origin of cyprinids	38.9	(2.5)

European Cyprinidae into two subfamilies: Cyprininae (including barbines) and Leuciscinae (including cultrins, tincins, gobionins, phoxinins, and alburnins + leuciscins) (Zardoya and Doadrio 1998; Zardoya et al. 1999). This result is in agreement with previous phylogenies based on osteological data (Chen et al. 1984; Cavender and Coburn 1992). Two recent molecular phylogenies based on complete cytochrome *b* (Briolay et al. 1998) and partial 16S rRNA and cytochrome *b* (Gilles et al. 1998) nucleotide sequence data have also shown that alburnins should be included within leuciscins and that phoxinins are the sister group of alburnins + leuciscins (as in Fig. 1). However, in both studies, the relative positions of gobionins and tincins were unresolved due to low bootstrap support (Briolay et al. 1998; Gilles et al. 1998). In general, the phylogenetic relationships reported by Briolay et al. (1998), although supported by low bootstrap values, are in agreement with ours (Zardoya and Doadrio 1998; Zardoya et al. 1999; this paper). On the other hand, the phylogenetic relationships (particularly within Leuciscinae) recovered by Gilles et al. (1998) are not supported by our results. As Gilles et al. (1998) point out, the lack of resolution of their analyses is due mainly to the design of the data set (too high a number of taxa with respect to the number of informative characters) and the unfortunate selection of hybrid specimens as representatives of some of the species (with the associated artificial results due to introgression processes).

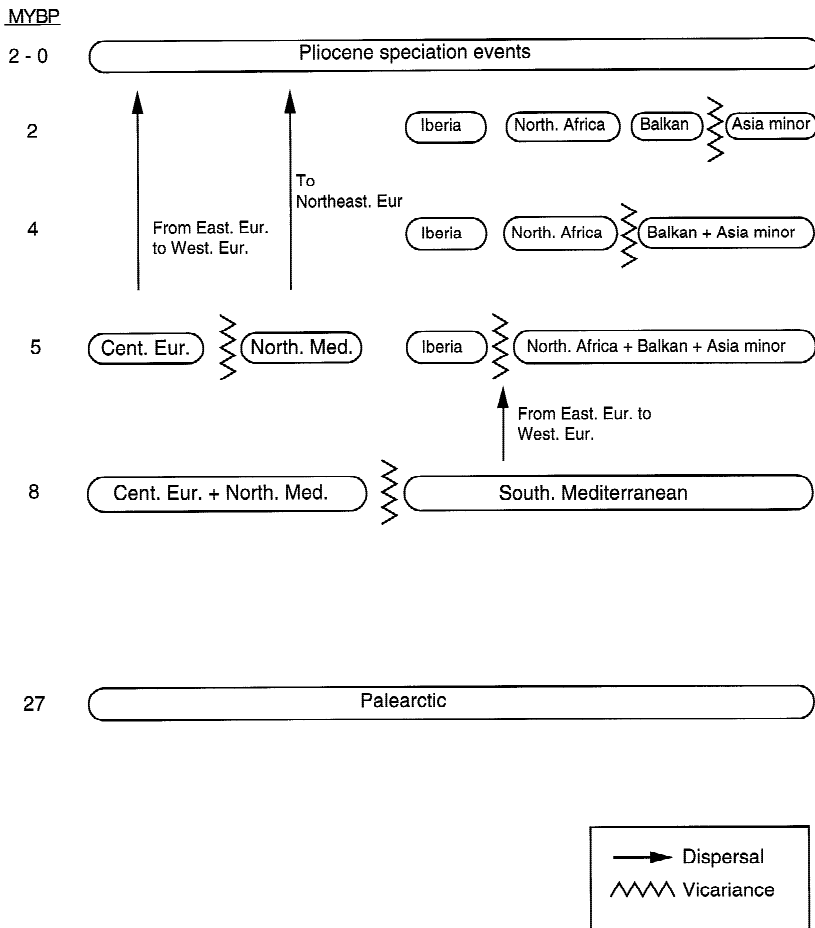
The cyprinid groupings reported here imply either that the absence of barbels is an ancestral cyprinid state, and, hence, that tincin and gobionin barbels are not homologous to those of barbines (Fink and Fink 1996), or that the presence of barbels is the ancestral cyprinid state and that they were lost by the common ancestor of phoxinins, alburnins, and leuciscins (Howes 1991). Taking into ac-

count that the barbel structure and pattern of innervation of *Cyprinus* and *Barbus* are more complex than those of *Tinca* and *Gobio* (Howes 1991), we consider the first hypothesis to be more reliable. In any case, the phylogenetic utility of this trait is rather limited. Conversely, the number of rows of pharyngeal teeth showed reduced homoplasy and proved to be a more reliable phylogenetic marker (at least at the genus level) when mapped onto the cytochrome *b* molecular phylogeny (Fig. 2). According to the mapping of this trait, the common ancestor of leuciscins, phoxinins, and alburnins had a single row of pharyngeal teeth. The acquisition of two rows of pharyngeal teeth seems to have occurred independently in *Phoxinus*, *Scardinius*, *Blicca*, *Alburnus*, *Alburnoides*, *Telestes*, and *Leuciscus* [with the exception of *Leuciscus alburnoides*, a hybridogenetic species (Carmona et al. 1997), which has reverted to the ancestral state], whereas *Pseudophoxinus*, *Pachychilon*, *Tropidophoxinellus*, *Rutilus*, and *Chondrostoma* retained the ancestral state (Fig. 2). *Anaocypris hispanica* exhibits a single row of pharyngeal teeth, whereas *Leucaspis* and *Alburnus* spp. (in the same monophyletic group) show two rows. Hence, according to the NJ phylogeny, it seems that *A. hispanica* has reverted to the ancestral character. However, in the MP analyses, the position of *A. hispanica* was basal to that of *Leucaspis* and *Alburnus* spp. (Zardoya et al. 1999), suggesting that, in fact, the single row of *A. hispanica* is a retention of the ancestral state (Fig. 2). *Barbus* and *Cyprinus* are characterized by three rows of pharyngeal teeth. This character state is shared with *Gobio* and *Hemiculter* (Fig. 2).

The calibration of the molecular clock to 0.76 %/MY is highly congruent with the widely accepted rate of substitution of 0.7% per MY for mitochondrial DNA of poikilothermic vertebrates (Martin and Palumbi 1993; see also Johns and Avise 1998). The estimated divergence dates are in full agreement with the fossil record of Eurasia (Cavender 1991) (although slightly underestimated due to the inherent biases associated to the calibration of a molecular clock). However, caution should be taken with the oldest estimates, e.g., the cypriniform-characiform split, in which the error is considerably higher due to saturation in the cytochrome *b*. According to the estimated dates (Fig. 3 and Table 2), cyprinids likely originated in the Eocene (about 39 MYA). This tentative dating supports the hypothesis, based on the fossil record, of an Asian origin of the family (Banareescu 1989; Cavender 1991).

The two subfamilies, Cyprininae and Leuciscinae, which may have originated in the mid-Oligocene (around 27 MYA) (Fig. 3 and Table 2), show different evolutionary and biogeographical patterns. The data suggest a very early radiation of Leuciscinae and a later radiation of Cyprininae. However, the latter result is probably an artifact due to the absence of key basal lineages within the Cyprininae data set. Cyprininae includes at least two





**Fig. 4.** Paleogeographical hypothesis of European Cyprininae evolution. Major dispersal and vicariance events of the Cyprininae are shown. Cyprininae may have crossed from Asia to Europe in the mid-Oligocene (27 MYA) while the Urals were forming. The Alpine orogenesis (10–8 MYA) may lead to a vicariant split of barbins into two subgenera, *Barbus* and *Luciobarbus*, which inhabited central Europe + northern Mediterranean and southern Mediterranean, respectively. In the late Miocene, cladogenesis in *Luciobarbus* followed the successive plate tectonic movements that occurred in the southern Mediterranean region. The subgenus *Barbus* was likely divided into two subgroups (*B. meridionalis* and related species, *B. barbus* and related species) during the regression of the Sarmatic Sea, in the late Miocene (5 MYA). In the Pliocene (2 MYA), the establishment of the actual drainage systems led to a major speciation event in both *Barbus* and *Luciobarbus*.

monophyletic groups (subgenera *Barbus* and *Luciobarbus*) with unrelated allopatric patterns (Figs. 1 and 3). Actually, *Luciobarbus* (Doadrio 1990) occurs on the Iberian Peninsula and in northern Africa, southwestern Greece, and Asia Minor (up to the Caucasus), whereas *Barbus* is distributed in central Europe, north Mediterranean, and the Caucasus. Therefore, our results are in clear disagreement with the classical hypothesis of a unique origin of European barbels and further dispersal through central Europe during the Oligocene and Miocene (Banarescu 1960; Almaça 1976, 1988; Banarescu 1989, 1992). Based on the dating of the main cladogenetic events of European Cyprininae (Fig. 3 and Table 2), and their actual distribution patterns, it is possible to postulate a paleobiogeographical scenario that reconstructs the vicariant and dispersal events that participated in the evolution of these taxa (summarized in Fig. 4) using a dispersal–vicariance analysis (Ronquist 1997) (not shown). According to this scenario, the lineage leading to modern European *Barbus* originated in the mid-Miocene (around 13 MYA) (Fig. 3 and Table 2). The oldest fossil record of *Barbus* in Europe is also mid-Miocene (15–11 MYA) (Quenstedt 1852), supporting our dating. Moreover, the major cladogenetic event in the *Barbus* lineage was the separation of the South Mediterranean barbins from the central European and North

Mediterranean taxa (about 8 MYA) (Fig. 4) (Doadrio 1990, 1994). This event might be directly related to the Alpine orogenesis in the late Miocene (10–8 MYA), which created a stretch of mountain chains from the Alps through the Dinarides and the Hellenides to Anatolia (Maldonado 1985), separating the above-mentioned regions.

On the other hand, and according to the dispersal–vicariance analysis (Ronquist 1997), European Leuciscinae have suffered numerous and complex cladogenetic events which are more difficult to interpretate and correlate with paleogeographical events. According to the fossil record, by the late Oligocene–early Miocene, leuciscins had reached the Iberian Peninsula (Cabrera and Gaudant 1985). Around that time (about 16 MYA), the *Pachychilon* lineage may have originated (Fig. 3 and Table 2). Hence, the Greek endemic *Pachychilon* may be a relict genus reminiscent of the fossil leuciscin fauna that inhabited Europe in the early to mid-Miocene [e.g., “*Rutilus antiquus*” and “*Rutilus pachecoi*” (Cabrera and Gaudant 1985), *Palaeoleuciscus* (Gaudant 1977)]. To test this hypothesis further, it would be interesting to find apomorphies between *Pachychilon* and the above-mentioned fossil taxa. The first major radiation within leuciscins occurred in the mid-Miocene (13.6 MYA), and extant lineages such as those leading to, e.g., *Scardi-*

*nius*, *Rutilus*, *Leuciscus*, and *Alburnus* were originated (Cavender 1991). The well-known fossil record of *Leuciscus* on the Iberian Peninsula dates back to the middle Miocene (De la Peña 1995), supporting these datings. Finally, extant species of European Leuciscinae originated mostly during the Pliocene (2.5–1.8 MYA), when the configuration of the actual European drainages was set up (Banarescu 1989).

In conclusion, our data provide a molecular phylogenetic framework which turns out to be very useful in revising the systematics of European cyprinids and the phylogenetic utility of morphological characters currently used to infer phylogenetic relationships within Cyprinidae. Furthermore, molecular evidence strongly supports biogeographical hypotheses (Bianco 1990; Doadrio 1990, 1994) that highlight the importance of the southern Mediterranean realm in the evolution of some European cyprinid taxa. The distribution of primary freshwater fish is directly related to paleobiogeography. Therefore, it is expected that the relationships reported here may also be found in other freshwater animals such as fish [e.g., *Valencia* and *Cobitis* (Banarescu 1989), gobies (Penzo et al. 1998)], amphibians [e.g., *Rana* and *Triturus* (Oosterbroek and Arntzen 1992; Beerli et al. 1996)], *Salamandra* (Veith et al. 1998), and invertebrates [e.g., decapods (Albrecht 1982), mollusks] living in southern Europe.

**Acknowledgments.** Annie Machordom, José Ambrosio Carmona, Anabel Perdices, Paloma Garzón, Yiannis Karakousis, Panos S. Economidis, Alex Mironovski, and Sacha Golubstov collaborated in the fish sampling. Ehab Abouheif, Kai Erik Witte, and an anonymous reviewer provided helpful suggestions on an early version of the manuscript. Lourdes Alcaraz assisted in the DNA extraction and cloning. David Swofford granted permission to publish results based on the test version of his PAUP\* program. R.Z. was sponsored by a postdoctoral contract from the Ministerio de Educacion y Ciencia of Spain. This work received partial financial support from Ministerio de Educación y Ciencia Grant PB-920025 to I.D.

## References

- Albrecht H (1982) On the origin of the Mediterranean crayfishes. *Quaderni del Laboratorio di Tecnologia della Pesca, Ancona* 3: 355–362
- Almaça C (1976) Zoogeografia e especiação dos ciprinídeos da Península Ibérica. *Natura Lisboa* 4:3–28
- Almaça C (1988) Remarks on the biogeography of Euro-Mediterranean *Barbus* (Cyprinidae, Pisces). *Bull Ecol* 19:159–162
- Banarescu P (1960) Einige Fragen zur Herkunft und Verbreitung der Süßwasserfischfauna der europäisch-mediterranen Unterregion. *Arch Hydrobiol* 57:16–134
- Banarescu P (1989) Zoogeography and history of the freshwater fish fauna of Europe. In: Holeik J (ed) *The freshwater fishes of Europe, Vol 1*. AULA-Verlag, Wiesbaden, pp 88–107
- Banarescu P (1992) Zoogeography of fresh waters: Distribution and dispersal of freshwater animals in North America and Eurasia, Vol 2. AULA-Verlag, Wiesbaden
- Beerli P, Hotz H, Uzzell T (1996) Geologically dated sea barriers calibrate a protein clock for Aegean water frogs. *Evolution* 50: 1676–1687
- Bianco P (1990) Potential role of the paleohistory of the Mediterranean and Paratethys basins on the early dispersal of Euro-Mediterranean freshwater fishes. *Ichthyol Explor Freshwaters* 1:167–184
- Briolay J, Galtier N, Brito RM, Bouvet Y (1998) Molecular phylogeny of Cyprinidae inferred from cytochrome *b* DNA sequences. *Mol Phylogenet Evol* 9:100–108
- Cabrera L, Gaudant J (1985) Los ciprinídeos (Pisces) del sistema lacustre Oligocénico-Miocénico de los Monegros (Sector SE de la cuenca del Ebro, provincias de Lleida, Tarragona, Huesca y Zaragoza). *Acta Geol Hispanica* 20:219–226
- Carmona JA, Sanjur O, Doadrio I, Machordom A, Vrijenhoek R (1997) Hybridogenetic reproduction and maternal ancestry of polyploid European fish: The *Tropidophoxinellus alburnoides* complex. *Genetics* 146:983–993
- Cavender TM (1991) The fossil record of the Cyprinidae. In: Winfield IJ, Nelson JS (eds) *Cyprinid fishes: Systematics, biology and exploitation*. Chapman & Hall, London, pp 34–54
- Cavender TM, Coburn M (1992) Phylogenetic relationships of North American Cyprinidae. In: Mayden R (ed) *Systematics, historical ecology, and North American freshwater fishes*. Stanford University Press, Stanford, CA, pp 293–327
- Chang YS, Huang FL, Lo TB (1994) The complete nucleotide sequence and gene organization of carp (*Cyprinus carpio*) mitochondrial genome. *J Mol Evol* 38:138–155
- Chen XL, Yue PQ, Lin RD (1984) Major groups within the family Cyprinidae and their phylogenetic relationships. *Acta Zootaxon Sin* 9:424–440
- De la Peña A (1995) Tertiary fishes from the Iberian continental basins: History and fossil record. *Coloq Paleontol* 47:25–46
- Dermitzakis MD (1990) The evolution of the Aegeis during the Late Cenozoic. *Geol Balcanica* 20:3–16
- Doadrio I (1990) Phylogenetic relationships and classification of west palaearctic species of the genus *Barbus* (Osteichthyes, Cyprinidae). *Aquat Living Resource* 3:265–282
- Doadrio I (1994) Freshwater fish fauna of North Africa and its biogeography. *Ann Mus R Afr Centr Zool* 275:21–34
- Felsenstein J (1985) Confidence limits on phylogenies: An approach using the bootstrap. *Evolution* 39:783–791
- Fink SV, Fink WL (1996) Interrelationships of Ostariophysan fishes. In: Stiassny MLJ, Parenti LR, Johnson GD (eds) *Interrelationships of fishes*. Academic Press, San Diego, pp 209–250
- Gaudant J (1977) Nouvelles observations sur l'ichthyofaune stampinienne d'Oberdorf (Canton de Soleure). *Ecol Geol Helv* 70:789–809
- Gilles A, Lecointre G, Faure E, Chappaz R, Brun G (1998) Mitochondrial phylogeny of European cyprinids: Implications for their systematics, reticulate evolution, and colonization time. *Mol Phylogenet Evol* 10:132–143
- Howes GJ (1991) Systematics and biogeography: An overview. In: Winfield IJ, Nelson JS (eds) *Cyprinid fishes. Systematics, biology and exploitation*. Chapman & Hall, London, pp 1–33
- Hsü K, Montadert L, Bernouilli D, et al. (1977) History of the Mediterranean salinity crisis. *Nature* 267:399–403
- Johns GC, Avise JC (1998) A comparative summary of genetic distances in the vertebrates from the mitochondrial cytochrome *b* gene. *Mol Biol Evol* 15:1481–1490
- Maddison WP, Maddison DR (1992) *MacClade: Analysis of phylogeny and character evolution*. Sinauer Associates, Sunderland, MA
- Maldonado A (1985) Evolution of the Mediterranean basins and a detailed reconstruction of the Cenozoic paleoceanography. In: Margaleff R (ed) *Key environments, western Mediterranean*. Pergamon Press, Oxford, pp 17–59
- Martin AP, Palumbi SR (1993) Body size, metabolic rate, generation time and the molecular clock. *Proc Natl Acad Sci USA* 90:4087–4091
- Oosterbroek P, Arntzen JW (1992) Area-cladograms of Circum-Mediterranean taxa in relation to Mediterranean paleogeography. *J Biogeogr* 19:3–20
- Penzo E, Gandolfi G, Bargelloni L, Colombo L, Patarnello T (1998)

- Messinian salinity crisis and the origin of the freshwater lifestyle in Western Mediterranean gobies. *Mol Biol Evol* 15:1472–1480
- Quenstedt FA (1852) *Handbuch der petrefaktenkunde*, Vol 2 (Tafeln). Laupp, Tübingen
- Ronquist F (1997) Dispersal-vicariance analysis: A new approach to the quantification of historical biogeography. *Syst Biol* 46:195–203
- Rutte E (1962) Schlundzähne von Süßwasserfischen. *Paleontographica Abt A* 120:165–212
- Saitou N, Nei M (1987) The neighbor-joining method: A new method for reconstructing phylogenetic trees. *Mol Biol Evol* 4:406–425
- Schmidt TR, Gold JR (1993) Complete sequence of the mitochondrial cytochrome *b* gene in the cherryfin shiner *Lythrurus roseipinnis* (Teleostei: Cyprinidae). *Copeia* 1993:880–883
- Steininger FF, Rögl F (1984) Paleography and palinspastic reconstruction of the Neogene of the Mediterranean and Paratethys. In: Dixon JE, Robertson AH (eds) *The geological evolution of the Eastern Mediterranean*. Geological Society Special Publication No 17. Blackwell, Oxford, pp 659–668
- Strimmer K, von Haeseler A (1996) Quartet puzzling: A quartet maximum-likelihood method for reconstructing tree topologies. *Mol Biol Evol* 13:964–969
- Swofford DL (1998) PAUP\*: Phylogenetic analysis using parsimony (\* and other methods), Version 4.0, Sinauer Associates, Sunderland, MA
- Takezaki N, Rzhetsky A, Nei M (1995) Phylogenetic test of the molecular clock and linearized trees. *Mol Biol Evol* 12:823–833
- Towner P (1991) Purification of DNA. In: Brown TA (ed) *Essential molecular biology. A practical approach*. Oxford University Press, Oxford, pp 47–68
- Tzeng CS, Hui CF, Shen SC, Huang PC (1992) The complete nucleotide sequence of the *Crossostoma lacustre* mitochondrial genome: Conservation and variations among vertebrates. *Nucleic Acids Res* 20:4853–4858
- Veith M, Steinfartz S, Zardoya R, Seitz A, Meyer A (1998) A molecular phylogeny of true salamanders (family Salamandridae) and the evolution of terrestriality of reproductive modes. *J Zool Syst Evol Res* 36:7–16
- Zardoya R, Doadrio I (1998) Phylogenetic relationships of Iberian cyprinids: Systematic and biogeographical implications. *Proc R Soc Lond B* 265:1365–1372
- Zardoya R, Economidis PS, Doadrio I (1999) Phylogenetic relationships of Greek Cyprinidae: Molecular evidence for the independent origin of the Greek cyprinid fauna. *Mol Phylogenet Evol* (in press)