

## Review Article

# Interleukin-4 and Breast Cancer

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IL-4 is a pleiotropic cytokine produced by T lymphocytes which acts on various cells of such as T and B lymphocytes, monocytes, fibroblast, endothelial cells, macrophages and some others. IL-4 was originally described as a B cell growth factor, and now known to provide potent anti-tumor activity against various tumors, including breast cancer. IL-4 can induce apoptosis in cultured breast cancer cells. In addition, it has been clarified that IL-4 plays an important role in the regulation of estrogen synthesis enzymes including 17  $\beta$ -HSD and 3  $\beta$ -HSD. These findings imply that IL-4 is a key enzyme not only for Th2 type immune reactions but also for tumor cell growth itself in human breast cancer.

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Interleukin-4 (IL-4) is a pleiotropic cytokine that exerts biological activities on various cells. It was first identified for its ability to induce B cell proliferation and differentiation<sup>1)</sup>. Important functions of IL-4 include B cell growth and immunoglobulin expression, T cell regulation, the generation of cytotoxic effector cells, and growth and differentiation effects on hematopoietic cells<sup>2-5)</sup>. Recent accumulated data have shown that IL-4 has potent anti-tumor activity against various types of carcinomas both *in vitro* and *in vivo*. This report summarizes the functions of IL-4, particularly its anti-tumoral activity, in human carcinomas including breast cancer.

### Source of IL-4

The main source of IL-4 is a subset of helper T cells (TH2) that also produce IL-5 and IL-6<sup>6)</sup>. This subset is clearly defined in mice and provides the most efficient helper activity for B cells. There is, however, no clear-cut evidence for human equivalents of the murine TH1 (IL-2 and IFN- $\gamma$

producing) and TH2 (IL-4 and IL-5 producing) T cell clones<sup>7)</sup>. IL-4 mRNA is detected in less than 5% of activated T cells<sup>8)</sup>. However, CD4+T cells infiltrating skin lesions of the patients with atopic dermatitis have been shown to produce high levels of IL-4<sup>9)</sup>. Cells other than T lymphocytes, in particular mast cell/basophils and bone marrow stromal cells also produce IL-4<sup>10)</sup>.

### Expression of IL-4 Receptor

IL-4 receptor (IL4-R) expression is detected on most T cells, B cells, mast cells, macrophages, cells of erythroid and myeloid lineage, fibroblasts, and most epithelial cells<sup>11-13)</sup>, but the receptor number per cell is generally low. The cDNA of IL-4R codes for a single chain transmembrane glycoprotein 140 KD polypeptide belonging to the hematopoietin receptor super family<sup>14-16)</sup>. IL-4R is also highly expressed in a wide variety of human tumors of epithelial origin including lung<sup>17)</sup>, ovary<sup>18)</sup>, kidney<sup>19, 20)</sup>, and breast<sup>18, 21)</sup> compared with normal cells. These findings indicate that increased expression of IL-4R is involved in the process of tumorigenesis. Approximately 30% of breast tumors have elevated expression of IL-4R<sup>22)</sup>. Subsequent analyses indicated that loss or down-regulation of IL-4R could contribute tumor progression via an effect on cell growth and differentiation<sup>23,24)</sup>. IL-4 signaling is inhibited by binding of IL-4 to the high-affinity IL-4 receptor alpha-chain and subsequent interaction with the common gamma-chain. Soluble forms of the extracellular domain of the alpha-chain (sIL-4R)

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### Abbreviation:

IL, Interleukin; TNF, Tumor necrosis factor; IFN, Interferon; TCR, T cell receptor; TGF, Transforming growth factor; IGF, Insulin-like growth factor; IRS, Insulin receptor substrate; ICAM, Intercellular adhesion molecule; VCAM, Vascular cell adhesion molecule; ELAM, E-selectin; VEGF, Vascular endothelial cell growth factor; HSD, Hydroxysteroid dehydrogenase

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were showed to be present in biological fluids and inhibit IL-4 activity by forming IL-4/sIL-4R complex<sup>25</sup>). Human recombinant sIL-4R induced the formation of IL-4/sIL-4R complex in the supernatant of activated T cells in a dose-dependent manner<sup>26</sup>). The production of human sIL-4R by T cells is regulated IL-4 and IFN-gamma and requires the activity of metalloproteinases<sup>27</sup>.

### Biological Effect of IL-4

IL-4 was first identified as a B cell growth factor (BCGF)<sup>1</sup>) because of its ability to promote B cell proliferation. In addition, IL-4 can induce resting B cells to increase their expression of class II major histocompatibility complex (MHC) molecules and low-affinity receptors for the Fc portion of IgE (Fc $\epsilon$ R). Moreover, IL-4 plays crucial roles in immunoglobulin isotype regulation, not only by inducing the production of the IgG1 isotype in mitogen and T cell stimulated cultures, but also by inducing high IgE levels<sup>2,5</sup>). IL-4 is well known to exert various functions on T cells and B cells. Both murine<sup>28</sup>) and human<sup>29</sup>) thymocytes proliferate in response to IL-4. The proliferative capability of these cells is independent of IL-2. IL-4 can also stimulate the generation of antigen-specific cytotoxic T cells<sup>30, 31</sup>). IL-4 is essential for in vitro growth of mucosal and connective tissue mast cells and may exert an autocrine effect, since it is produced by several transformed and non-trans-

formed mast cell lines<sup>32</sup>). IL-4 also enhances the antigen presenting ability and anti-tumor activity of macrophages and acts on many hemopoietic cell lineages, including immature erythroid, myelomonocytic and megakaryocytic precursors<sup>33</sup>. (Fig 1).

### Activity on Breast Cancer Cells

Transfection of the IL-4 gene in tumor cell lines<sup>34-36</sup>) and treatment of tumor cells with IL-4 have demonstrated that IL-4 has potent anti-tumor activity against various types of tumors including renal carcinoma, lung carcinoma, thyroid carcinoma, and breast carcinoma<sup>21, 37-40, 48</sup>). Tepper *et al*<sup>36</sup>). have shown that IL-4 transfected malignant tumor cell lines (plasmacytoma, breast adenocarcinoma) elicited a potent antitumor response when injected into mice. This antitumor response was observed in nu/nu mice, which are devoid of T cells, and bg/bg mice, which are devoid of NK cells, indicating that T and NK cells are not necessary for the effect of IL-4. Histologic examination of the subcutaneous inoculation site of IL-4 producing tumor cells showed infiltrating macrophages and eosinophils which are hypothesized to mediate the anti-tumor effect of IL-4. The action of IL-4 is clearly dose-dependent, with complete eradication of tumor requiring highly localized concentration of cytokines. The reversal of IL-4-induced tumor cytotoxicity by the in vivo administration of anti-IL-

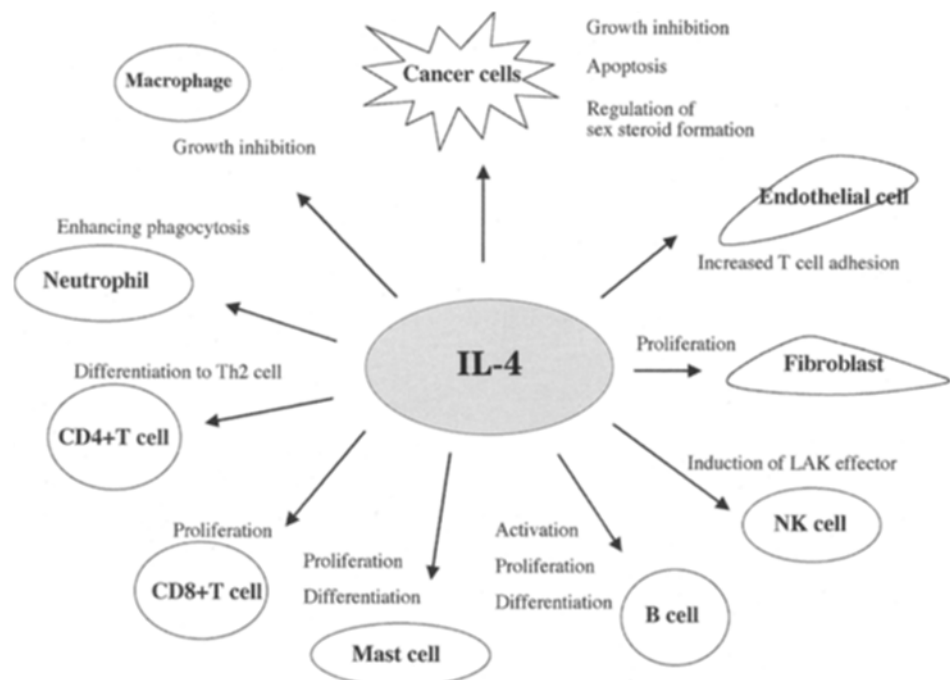


Fig 1. Biological activity of IL-4.

Table 1. IL-4 and Breast Cancer

|                 |                               | Cell lines                          | Remarks  |
|-----------------|-------------------------------|-------------------------------------|--|
| <i>in vitro</i> | Klara T ('91) <sup>40</sup>   | MDA-MB-330                          | Modulation of TNF-dependent anti-tumor response  |
|                 | Toi M ('92) <sup>21</sup>     | MCF-7 WT, MCF-7r<br>MDA-MB-231, 468 | Additive cell growth inhibition with TAM and TGF- $\beta$ 1                                |
|                 | Blais Y ('96) <sup>37</sup>   | ZR-75-1, T-47D                      | Inhibition of estrogen-induced breast cancer cell proliferation and induction of GCDFP-45  |
|                 | Gooch JL ('98) <sup>41</sup>  | MCF-7, MDA-MB-231                   | Growth inhibition induced by apoptosis   |
| <i>in vitro</i> | Tepper RI ('89) <sup>36</sup> | K485                                | Growth inhibition mediated by infiltrated inflammatory cells                               |
|                 | Pericle F ('94) <sup>43</sup> | TS/A-pc                             | Th2 memory in growth inhibition  |
|                 | Allione A ('94) <sup>44</sup> | TS/A-pc                             | Protection 80-100% of mice immunized with replicating tumor cells by the induction of IL-4 |
|                 | Musiani P ('96) <sup>45</sup> | TS/A                                | Rejection of IL-4 producing TSA cells by neutrophils and CD8 <sup>+</sup> lymphocytes      |
|                 | Maeda T ('99) <sup>46</sup>   | OCUB-M                              | No tumor formation in IL-4 producing cell <i>in vivo</i>                                   |
|                 | Pacor S ('99) <sup>47</sup>   | TS/A                                | Paracrine effect of IL-4 induced growth inhibition <i>in vivo</i>                          |

4 antibody provides further evidence for the action of IL-4 as a secreted factor. An additive antiproliferative effect of IL-4 with other cytokines has also been reported. The enhancement of the cytotoxic effect of tumor necrosis factor (TNF)- $\alpha$  by IL-4 was showed in breast carcinoma cells (MDA-MB-330) in a dose-dependent manner. This effect was also observed in human epidermoid carcinoma cells and human histiocytic lymphoma cells<sup>40</sup>. Combination treatment with IL-4 and other growth inhibitors, such as TGF- $\beta$  1 and tamoxifen, showed an additive growth inhibitory effect in breast carcinoma cell lines. IL-4 inhibited 90% of the 17 $\beta$ -estradiol-stimulated growth of MCF-7 WT cells without a change in estrogen receptor expression. Less inhibition of growth by IL-4 occurred when estrogens were absent. The hormone independent cell lines MCF-7r and MDA-MB-231 were inhibited by IL-4 and TGF- $\beta$  1<sup>21</sup>. Several lines of experimentation have provided evidence that the anti-tumor activity initiated by localized IL-4 secretion may be mainly due to the rapid influx of host effector cells capable of mediating tumor cytotoxicity directly or indirectly through other tumoricidal cytokines<sup>36</sup>. However, recently several investigators have elucidated another mechanism of the anti-tumor effect of IL-4, namely that IL-4 can induce apoptosis in breast cancer cell lines. Insulin-like growth factor (IGF)-1 and IL-4 share a common signaling pathway via the insulin receptor substrate (IRS)-1 molecule. IL-4 had no effect on IGF-1 stimulated proliferation,

which is known to inhibit apoptosis, in MCF-7 and MDA-MB-231 cell lines. IGF-1 reversed IL-4 induced growth inhibition, suggesting that the mechanism of IL-4 induced growth inhibition in these cells is the induction of apoptosis<sup>41</sup>. These findings underlie the importance of the balance between IGF-1 and IL-4 for the induction of apoptosis in breast cancer cells. For instance, breast cancer cells that have become hormone resistant are likely to enhance IGF-1 production. Although little is yet known about IL-4 expression in hormone resistant tumors an IGF-1 dominant, IL-4 recessive status might be achieved in such tumors. It is known that E2 and insulin induce polyamine transport in breast cancer, such as the ZR-75-1 cell lines<sup>42</sup>. IL-4 is also characterized to be a potent inducer of polyamine transport. IL-4 caused a marked increase of uptake of radiolabeled spermidine with 80 pg/ml ED50. Interferon (IFN)- $\gamma$  is reported to mimic the IL-4 induced spermidine uptake. This is an interesting aspect of IL-4 considering its biological function. *In vivo* and *in vitro* studies of breast cancer are shown in Table 1.

#### **Effects on the Endothelium**

IL-4 mediating expression of adhesion molecules in endothelial cells may be an important mechanism by which IL-4 promotes the selective transmigration of eosinophils into tissues. IL-4 has been shown to stimulate, alone or synergistically with IL1 or TNF- $\alpha$ , the induction of VCAM-1

Table 2. Phase II Clinical Trial of rIL-4 in Patients with Solid Tumor

|                     | SOG <sup>54)</sup>   | UCCRC <sup>55)</sup>   | UC <sup>56)</sup>  | CHNMC <sup>57)</sup>   |
|---------------------|--|--|--|--|
| Tumor Type          | advanced melanoma  | NSCLC  | advanced RCC   | melanoma advanced RCC  |
| No. of Pt           | 36   | 63   | 19   | 49   |
| Dose                | 5 $\mu$ g/kg   | 0.25 or 1.0 $\mu$ g/kg   | 1.0 $\mu$ g/kg   |  |
| Route and frequency | s.c., day 1-28<br>35 d cycle   | s.c.<br>3 times/week   | s.c.<br>3 times/week   | i.v.   |
| Response            | CR 1, NC 2   | PR 1, NC 9   | no response  | CR 1   |
| Toxicity            | nausea, vomiting,<br>diarrhea, malaise, fatigue,<br>edema, headache,<br>myalgias, arthralgias,<br>fever, chill, liver<br>dysfunction | fatigue, fever,<br>vomiting, dyspnea,<br>anorexia,<br>duodenal ulcer | fever, fatigue,<br>myalgias,<br>arthralgias,<br>nausea, anorexia<br>gastrointestinal<br>bleeding | fever, malaise, nasal<br>congestion,<br>diarrhea,<br>cardiac ischemia,<br>gastrointestinal<br>bleeding |

SOG, Southwest Oncology Group; UCCRC, University of Chicago Cancer Research Center; UC, University of Chicago; CHNMC; City of Hope National Medical Center; NSCLC, Non-small cell lung cancer; RCC, Renal cell carcinoma.

on human endothelial cells, while ICAM-1 and ELAM-1, which promote the binding of all granulocyte subtypes to endothelium<sup>48)</sup>, are unaffected by IL-4<sup>49)</sup>. IL-4 is a potent regulator of angiogenesis. Interestingly, IL-4 behave bimodally with regard to endothelial cell growth. In lower concentrations at picomolar levels, IL-4 stimulates proliferation and migration of endothelial cells and also can promote in vivo neovascularization. On the other hand, in higher nanomolar concentrations, IL-4 inhibits endothelial cell growth and migration. IL-4 blocked the induction of corneal neovascularization in rat by basic fibroblast growth factor<sup>50)</sup>. Furthermore, it was reported that IL-4 down-regulates the expression of one VEGF receptor, VEGF-R2, in an experiment using mouse endothelial cells. The local anti-angiogenic activity of IL-4 could account for the decrease in tumor vessel density observed in IL-4 secreting gliomas<sup>51)</sup>. On the other hand, in our clinical analysis of primary breast cancer, we failed to demonstrate a significant correlation between IL-4 levels in tumor tissues and angiogenic grade, which might reflect the bi-functional role of IL-4.

### **Hormone and IL-4**

There is evidence suggesting that local intracrine formation of sex steroids from inactive precursors secreted by the adrenal gland plays a crucial role in the regulation of growth and function of peripheral target tissues, including

breast.  $17\beta$ -HSD and  $3\beta$ -HSD activities represent critical sites of control and regulation of sex steroid formation and/or inactivation in human breast cancer cells. Turgeon C *et al*<sup>52)</sup> investigated the potential effects of IL-4 and IL-6 on  $17\beta$ -HSD activity in ZR-75-1 and T-47D human breast cancer cell lines, and showed that IL-4 and IL-6 regulate the  $17\beta$ -HSD activity, whereas IL-4, but not IL-6, may induce  $3\beta$ -HSD activity in these two cell lines. Interestingly, IL-4 increased oxidative  $17\beta$ -HSD activity, but significantly decreased reductive  $17\beta$ -HSD activity in low concentrations (20 pM-100 pM) in ZR-75-1 cells. This enzymatic modulation elicited a marked decrease of E2 formation. In contrast, in T-47D cells, IL-4 increased the formation of E2, which suggests that the regulatory mechanism of  $17\beta$ -HSD activity by IL-4 differs among cell lines, and perhaps among individual tumors. Genetic expression of  $3\beta$ -HSD type 1, which is essential for the biosynthesis of active estrogens and androgens, was markedly induced by picomolar levels of IL-4 in various types of cultured breast cancer cells including ZR-75-1, T-47D, and ER-negative MDA-MB-231 cells, and normal mammary epithelium. It was further found that IL-4 can enhance the DNA-binding activity of Stat6 in these cells, and the Stat6 activated by IL-4 can bind to the promoter region of the  $3\beta$ -HSD type 1 gene<sup>53)</sup>. According to our recent analysis of the intra-factorial correlation among cytokines including IL-4, chemokines and

growth factors in human breast tumor tissues, IL-4 is frequently overexpressed and its expression is significantly associated with other Th-2 type cytokines including IL-10. In addition, intratumoral IL-4 levels tend to be negatively correlated with the level of hormone receptor, which seems to indicate that IL-4 plays important roles in hormone-dependent and -independent growth of breast cancer.

### Therapeutic Approaches

Accumulated data supports a potential clinical use of IL-4. Phase I/II clinical trials of IL-4 therapy in patients with solid tumors are already underway. Some of these studies are summarized in Table 2<sup>54-57</sup>. In some studies, IL-4 is being given subcutaneously to patients with renal cell carcinoma, melanoma, and non-small cell lung carcinoma. The doses range from 0.25-5.0  $\mu$ g/kg. Clinical trials in patients with breast cancer have not been reported yet. IL-4 can induce serious adverse effects, including fever, nausea, diarrhea, anorexia, fatigue, cardiac ischemia and gastrointestinal bleeding. Although no significant response against solid tumors has been reported to date, several novel approaches focusing on IL-4 related cell growth inhibition and apoptosis are now under investigation.

### Reference

- 1) Howard M, Farrar J, Hilfiker M, *et al*: Identification of a T cell-derived B cell stimulatory factor distinct from IL-2. *J Exp Med* 155:914-923, 1982.
- 2) Yokota T, Arai N, de Vries J, *et al*: Molecular biology of interleukin-4 and interleukin-5 genes and biology of their products that stimulate B cells, T cells and haematopoietic cells. *Immuno Rev* 102:137-138, 1988.
- 3) Coffman RL, Seymour B, Lebman DA, *et al*: The role of helper T cell products in mouse B cell differentiation and isotype regulation. *Immuno Rev* 102:5-28, 1988.
- 4) Snpper CM, Finkelman FD, Paul WE: Regulation of IgG1 and IgE production by interleukin 4. *Immunol Rev* 102:29-56, 1988.
- 5) Noma Y, Sidras P, Naito T, *et al*: Cloning for cDNA encoding the murine IgG 1 induction factor by a novel strategy using SP6 promoter. *Nature* 319:640-644, 1986.
- 6) Mosmann TR, Coffman RL: TH1 and TH2 cells: difference patterns of lymphokine secretion lead to different functional properties. *Ann Rev Immunol* 7:145-173, 1989.
- 7) Paliard X, De Waal MR, Yssel H, *et al*: Simultaneous production of IL-2, IL-4 and IFN-gamma by activated human CD4+ and CD4+ T cell clones. *J Immunol* 141: 849-855, 1988.
- 8) Lewis DB, Prickett KS, Larsen A *et al*: Restricted production of interleukin-4 by activated human T cells. *Proc Natl Acad Sci USA* 85:9743-9747, 1988.
- 9) van der Heijden FL, Wierenga EA, Bos JD, *et al*: High frequency of IL-4 producing CD4+ allergen-specific T lymphocytes in atopic dermatitis lesional skin. *J Invest Dermatol* 97:389-394, 1991.
- 10) Banchereau J: Interleukin 4. *Nucl Med Biol* 17:619-623, 1990.
- 11) Lownthall JW, Castle BE, Christiansen J, *et al*: Expression of high affinity receptors for murine interleukin 4 (BSF-1) on haematopoietic and non-haematopoietic cells. *J Immunol* 140:456-464, 1988.
- 12) Park LS, Friend D, Sassenfeld HM, *et al*: Characterization of the high affinity cell surface receptor for murine B cell stimulatory factor-1. *Proc Natl Acad Sci USA* 84:1669-1673, 1987.
- 13) Cabrillat H, Galizzi JP, Djossou O, *et al*: High affinity binding of human interleukin 4 to cell lines. *Biochem Biophys Res Commun.* 149:995-1001, 1987.
- 14) Idzerda RL, March CJ, Mosley B, *et al*: Human interleukin 4 receptor confers biological responsiveness and defines a novel receptor superfamily. *J Exp Med* 171:861-873, 1990.
- 15) Galizzi J-P, Zuber CE, Harada N, *et al*: Molecular cloning of a cDNA encoding the human IL-4 receptor. *Int Immunol* 2:669-675, 1990.
- 16) Bazan J F: A novel family of growth factor receptors: a common binding domain in the growth hormone, prolactin, erythropoietin and IL-6 receptors and the p75IL-2 receptor beta-chain. *Biochemical and Biophys Res Commun.* 164:788-795, 1989.
- 17) Topp MS, Koenigsman M, Anthony M, *et al*: Recombinant human interleukin-4 inhibits growth of some human lung tumor cell lines in vitro and in vivo. *Blood* 82:2837-2844, 1993.
- 18) Obiri NI, Siegel JP, Varricchio F, *et al*: Expression of high-affinity IL-4 receptors on human melanoma, ovarian and breast carcinoma cells. *Clin Exp Immunol* 85:148-155, 1994.
- 19) Obiri NI, Hillman GG, Haas GP, *et al*: Expression of high-affinity interleukin-4 receptors on human renal cell carcinoma cells and inhibition of tumor cell growth in vitro by interleukin-4. *J Clin Invest* 91:88-93, 1993.
- 20) Varricchio F, Obiri NI, Haas GP, *et al*: Immunostaining of interleukin-4 receptors on human renal cell carcinoma. *Lymphokine Cytokine Res* 12:465-469, 1993.
- 21) Toi M, Bicknell R, Harris AL: Inhibition of colon and breast carcinoma cell growth by interleukin-4. *Cancer Res* 52:275-279, 1992.
- 22) Mat I, Larche M, Melcher D: Tumor associated upregulation of IL-4 receptor complex. *Br J Cancer.* 62(supple x):96-98, 1990.
- 23) Kaklamanis L, Koukourakis. MI, Leek R: Loss of interleukin-4 receptor-associated molecule gp200-MR6 in human breast cancer; Prognostic significance. *Br J Cancer* 74:1627-1631, 1996.
- 24) Tubly A, Luqmani YA, Shousha S, *et al*: Differential expression of gp200-MR6 molecule in benign hyperplasia and down-regulation in invasive carcinoma of the breast. *Br J Cancer* 74:1005-1011, 1996.
- 25) Jung T, Bews JP, Enssle KH, *et al*: Detection of and discrimination between total and free human interleukin 4 and free soluble interleukin 4 receptor by ELISA. *J Immunol Method* 217:41-50, 1998.

- 26) Jung T, Wagner K, Neumann C, *et al*: Enhancement of human IL-4 activity by soluble IL-4 receptors *in vitro*. *Eur J Immunol* 29:864-871, 1999.
- 27) Jung T, Schrader N, Hellwing M, *et al*: Soluble human interleukin 4 receptor is produced by activated T cells under the control of metalloproteinases. *Int Arch Allergy Immunol* 119:23-30, 1999.
- 28) Zlotnick A, Ransom J, Frank G, *et al*: Interleukin 4 is a growth factor for activated thymocytes: Possible role in T cell ontogeny. *Proc Natl Acad Sci USA* 84: 3856-3860, 1987.
- 29) Spits H, Yssel H, Paliard X, *et al*: IL-4 inhibits IL-2 mediated induction of human lymphokine-activated killer cells, but not the generation of antigen specific cytotoxic T lymphocytes in mixed lymphocyte cultures. *J Immunol* 141:29-36, 1988.
- 30) Brown M, Hu-Li J, Paul WE: IL-4/B cell stimulatory factor-1 stimulates T cell growth by an IL-2 independent mechanism. *J Immunol* 141:504-511, 1988.
- 31) Widmer MB, Grabstein KH: Regulation of cytolytic T lymphocytes generation by B cell stimulatory factors. *Nature* 326:795-798, 1987.
- 32) Lee F, Yokota T, Otsuka T, *et al*: Isolation and characterization of a mouse interleukin cDNA clone that express B-cell stimulatory factor 1 activities and T-cell and mast-cell stimulating activities. *Proc Natl Acad Sci USA* 83:2061-2065, 1986.
- 33) Paul WE, Ohara J: B-cell stimulatory factor-1/interleukin 4. *Ann Rev Immunol* 5:429-459, 1987.
- 34) Santin AD, Ioli GR, Hiserodt JC, *et al*: Development and characterization of an IL-4 secreting human ovarian carcinoma cell line. *Gynecol Oncol* 58:230-239, 1995.
- 35) Golumbek PT, Lazenby AJ, Levitsky HI, *et al*: Treatment of established renal cancer by tumor cells engineered to secrete interleukin 4. *Science* 254:713-716, 1991.
- 36) Tepper RI, Pittengale PK, Leder P: Murine interleukin4 displays potent anti-tumor activity *in vivo*. *Cell* 57:503-512, 1989.
- 37) Blais Y, Gingras S, Haagensen DE, *et al*: Interleukin-4 interleukin-13 inhibit estrogen-induced breast cancer cell proliferation and stimulate GCDFP-15 expression in human breast cancer cells. *Mol Cell Endocrinol* 121; 11-18, 1996.
- 38) Topp MS, Papadimitriou CA, Eitelbach F, *et al*: Recombinant human interleukin 4 has antiproliferative activity on human tumor cell lines derived from epithelial and nonepithelial histologies. *Cancer Res*, 55:2173-2176, 1995.
- 39) Cressent M, Pidoux E, Cohen R, *et al*: Interleukin-2 and interleukin-4 display potent antitumor activity on rat medullary thyroid carcinoma cells. *Euro J Cancer* 31A:2379-2384, 1995.
- 40) Klara T, Bharat BA: Interleukin4 potentiates the antiproliferative effect of tumor necrosis factor on various tumor cell lines. *Cancer Res* 51:4266-4270, 1991.
- 41) Gooch JL, Lee AV, Yee D: Interleukin4 inhibit growth and induce apoptosis in human breast cancer cells. *Cancer Res* 58:4199-4205, 1998.
- 42) Blais Y, Zhao C, Huber M, *et al*: Growth-independent induction of spermidine transport by IL-4 and IL-13 in human cancer cells. *Int J Cancer* 67:532-538, 1996.
- 43) Pericle F, Giovarelli M, Colombo MP: An efficient Th2-type memory follows CD8+ lymphocyte-derived and eosinophil-mediated rejection of a spontaneous mammary adenocarcinoma engineered to release IL-4. *J Immunol* 153:5659-5673, 1994.
- 44) Allione A, Consalvo M, Nanni P, *et al*: Immunizing and curative potential of replicating and nonreplicating murine mammary adenocarcinoma cells engineered with IL-2, IL-4, IL6, IL-7, IL-10, tumor necrosis factor alpha, granulocyte-macrophage colony-stimulating factor, and gamma-interferon gene or admixed with conventional adjuvants. *Cancer Res* 54:6022-6026, 1994.
- 45) Musiani P, Allione A, Modica A, *et al*: Role of neutrophils and lymphocytes in inhibition of a mouse mammary adenocarcinoma engineered to release IL-2, IL-4, IL-7, IL10, IFN-alpha, IFN-gamma, and TNF-alpha. *Lab Inves* 74: 146-157, 1996.
- 46) Maeda T, Matsubara H, Sugaya M: Loss of tumorigenicity of human breast cancer cells engineer to produce IL-2, IL-4 or GM-CSF in nude mice. *Int J Oncol* 15:943-7, 1999.
- 47) Pacor S, Gagliardi R, Spessotto P: Paracrine effects of IL-4 transfection on TS/A adenocarcinoma cells mediate reduced *in vivo* growth. *Pathol Oncol Res* 5: 110-16, 1999.
- 48) Butcher EC: Leukocyte-endothelial cell recognition: three or more steps to specificity and diversity. *Cell* 67:1033-1036, 1991.
- 49) Schleimer RP, Sterbinsky SA, Kaiser J, *et al*: IL-4 induces adherence of human eosinophils and basophils but not neutrophils to endothelium. *J Immunol* 148:1086-1092, 1992.
- 50) Volpert OV, Fong T, Koch AE, *et al*: Inhibition of angiogenesis by interleukin 4. *J Exp Med* 188:1039-1046, 1998.
- 51) Saleh M, Davis ID, Wilks AF: The paracrine role of tumor-derived mIL-4 on tumor-associated endothelium. *Int J Cancer* 72:664-672, 1997.
- 52) Turgeon C, Gingras S, Carriere MC, *et al*: Regulation of sex steroid formation by interleukin 4 and interleukin 6 in breast cancer cells. *J Steroid Mole Biol* 65:151-162, 1998.
- 53) Gingras S, Moriggi R, Groner B, *et al*: Induction of 3  $\beta$ -hydroxysteroid dehydrogenase/delta5-delta4 isomerase type 1 gene transcription in human breast cancer cell lines and in normal mammary epithelial cells by interleukin-4 and interleukin-13. *Mol Endocrinol* 13:66-81, 1999.
- 54) Whitehead RP, Unger JM, Goodwin JW, *et al*: Phase II trial of recombinant human interleukin 4 in patients with disseminated malignant melanoma; A South west Oncology Group study. *J Immunol* 21:440-446, 1998.
- 55) Vokes EE, Figlin R, Hochster H: A phase II study of recombinant human interleukin 4 for advanced or recurrent non-small cell lung cancer. *Cancer J Sci Am* 4:46-51, 1998.
- 56) Stadler WM, Rybak ME, Vogelzang NJ: A phase II study of subcutaneous recombinant human interleukin 4 in metastatic renal cell carcinoma. *Cancer* 76:1629-1633, 1995.
- 57) Margolin K, Aronson FR, Sznol M, *et al*: Phase II studies of recombinant human interleukin 4 in advanced renal cancer and malignant melanoma. *J Immunolther Emphasis Tumor Immunol* 15:147-53, 1994.