

## Determination of macrocyclic trichothecenes in mouldy indoor materials by LC-MS/MS

C. Gottschalk, J. Bauer, K. Meyer

TU München, Chair of Animal Hygiene, Weihenstephaner Berg 3, 85354 Freising, Germany

### Abstract

*Stachybotrys* occurring in mouldy indoor environments is associated with the so called "sick building syndrome" in humans or cases of idiopathic pulmonary hemorrhages. Samples of mouldy materials from indoor environments (n=15) were analysed for the occurrence of this fungus and its secondary metabolites by a sensitive LC-MS/MS method. In four samples, *Stachybotrys* and macrocyclic trichothecenes have been detected. Maximum values for Satratoxin G and H in wallpaper were determined with 9.7 µg/cm<sup>2</sup> and 12.0 µg/cm<sup>2</sup>, respectively.

**Keywords:** macrocyclic trichothecenes, *Stachybotrys*, Satratoxin

### Introduction

Macrocyclic trichothecenes produced by *Stachybotrys* species are highly toxic compounds associated with the so called "sick building syndrome" in humans. As several case control studies have shown, the occurrence of *Stachybotrys* in indoor environments could be related to headaches, fatigue, nausea, vomiting, hemorrhages, depression, sleep disturbances, anxiety, vertigo, and amnesia. Also cases of pulmonary hemosiderosis in infants have been related to *Stachybotrys* growth (1, 2). In 1938, this mould was first identified as etiologic agent of stachybotryotoxicosis, a severe disease of horses, which has been caused by ingestion of mouldy straw. Later, macrocyclic trichothecenes were supposed to be the reason of this disease (3). Because of its high cellulolytic potential, *S. chartarum*, which is mainly responsible for toxin production, also grows on humid cellulose-containing materials like wallpaper or gypsum board, e.g. after water damages (4). In this study a sensitive LC-MS/MS method has been developed to determine Satratoxins and other macrocyclic trichothecenes in these matrices.

### Materials and Methods

#### Materials

15 samples were taken from mouldy indoor materials like wallpaper (n=3), wall (n=10) or gypsum board (n=2). Standard substances of Roridin A and Verrucaric acid were purchased from Sigma (Deisenhofen, Germany). Standards of Satratoxin G and H were kindly provided by Prof. Gareis (BfEL Kulmbach, Germany). Acetonitrile and methanol (HPLC-grade) were purchased from J.T. Baker (Griesheim, Germany). Water was purified by a milli-Q water purification system (Millipore GmbH, Schwalbach, Germany).

#### Mycological examination

For mould differentiation, smears of the mouldy area were plated on Sabouraud agar and incubated overnight at 37 °C. After an incubation time of one week at room temperature, micro-morphological characterisation was carried out by tape preparations and staining with lactophenol blue solution at 1000-fold magnification.

#### Sample preparation and recovery experiments

About 15 cm<sup>2</sup> wallpaper, wall or gypsum board was reduced to small pieces and extracted with 10 ml acetonitrile/water 84/16 (v/v) by shaking for 1 h using a horizontal shaker. The extract was centrifuged for 5 min at 5000 rpm. For analysis, an aliquot was diluted 1:5 with water and filtered through a 0.45 µm PTFE syringe filter (Supelco, Deisenhofen, Germany). For

---

Presented at the 28<sup>th</sup> Mykotoxin-Workshop, Bydgoszcz, Poland, May 29-31, 2006

Correspondence: Christoph Gottschalk, Lehrstuhl für Tierhygiene, Weihenstephaner Berg 3, 85354 Freising, Germany (christoph.gottschalk@wzw.tum.de)

testing the recovery, extracts of non-contaminated wallpaper were spiked with Satratoxin G and H at a level of 1 µg/cm<sup>2</sup> (n=3).

#### LC-MS/MS parameters

The LC-MS/MS system consisted of a HPLC apparatus Series 200 (Perkin Elmer, Rodgau-Jügesheim, Germany) with two pumps, a degasser, an autosampler and a column oven as well as an API 3200 triple quadrupole mass

spectrometer (Applied Biosystems, Darmstadt, Germany). As analytical column a Gemini 150x2 mm, 5 µm (Phenomenex, Aschaffenburg, Germany) was used. The oven temperature was set at 30 °C. The binary linear gradient consisted of eluent A (deionized water +5 mM ammonium formate) and eluent B (methanol +5 mM ammonium formate) with a flow rate of 400 µl/min: 0 min 10% B, 26 min 100% B, 30 min 100% B, 35 min 10% B.

**Table 1.** MRM-transitions, declustering potentials (DP) and collision energies (CE) for 8 macrocyclic trichothecenes

Analyte	Precursor Ion [M+NH <sub>4</sub> ] <sup>+</sup>		Fragment Ions				Retention time [min]
	Q1	DP [V]	Q3 (1)	CE [eV]	Q3 (2)	CE [eV]	
Roridin A	550.4	26	249.1	23	133.0	41	17.75
Roridin E *	532.4	31	231.1	21	249.1	25	19.90
Roridin L-2 *	548.3	31	249.1	21	231.1	21	15.38
Satratoxin F *	560.3	41	249.1	21	231.1	21	17.01
Satratoxin G	562.3	21	249.1	19	231.1	21	15.82
Satratoxin H	546.3	21	245.2	25	231.1	25	16.39
Verrucarin A	520.2	31	249.1	23	231.1	27	17.37
Verrucarin J *	502.4	30	231.1	25	249.1	25	19.58

\* Putative mass transitions as no standard substances available

**Table 2.** Identified fungi from mouldy indoor materials

Sample	<i>Alternaria</i> spp.	<i>Aspergillus</i> spp.	<i>Chaetomium</i> spp.	<i>Cladosporium</i> spp.	<i>Gliocladium</i> spp.	<i>Memnoniella</i> spp.	<i>Mucor</i> spp.	<i>Penicillium</i> spp.	<i>Stachybotrys</i> spp.	<i>Ulocladium</i> spp.
Gypsum board 1		x						x	x	
Gypsum board 2		x						x		
Wall 1		x				x		x		
Wall 2								x		
Wall 3		x			x		x			
Wall 4				x				x		
Wall 5		x						x		x
Wall 6				x				x		
Wall 7							x	x		
Wall 8										
Wall 9										x
Wall 10		x								
Wallpaper 1	x		x					x	x	
Wallpaper 2								x	x	
Wallpaper 3		x							x	

The MS/MS experiments were carried out in positive electrospray ionisation mode with an ion spray voltage of 4000 V and a source temperature of 300 °C. The nebulizer gas flow was set at 50 psi and the heating gas at 30 psi. As there were no standards available for Roridin E, Roridin L-2, Satratoxin F and Verrucarín J, probable mass transitions were assumed for these toxins. All substances showed best sensitivity as adduct-ions of ammonium  $[M+NH_4]^+$ . The MRM-transitions, declustering potentials and collision energies used as well as the retention times of the toxins are shown in Table 1.

## Results and Discussion

### Mould characterisation

*Stachybotrys* and its toxins could be detected on three wallpaper samples and one gypsum board sample. Other moulds like *Aspergillus* and *Penicillium* were also detected frequently (Table 2). From one wall sample no mould growth could be observed.

### LC-MS/MS analysis

Spiking experiments with Satratoxin G and H at a level of 1 µg/cm<sup>2</sup> (n=3) resulted in recovery rates of 92.9±0.2% (mean±RSD) for Satratoxin G and 107.4±2.1% for Satratoxin H.

All *Stachybotrys* positive samples (n=4) also contained macrocyclic trichothecenes. In the other samples none of the toxins analysed could be identified. From two wallpaper samples different pieces have been examined. The analysis gave varying results (Table 3). This indicates an unequal distribution of the toxins in the mouldy material. Satratoxin G and H showed maximum levels of 9.7 µg/cm<sup>2</sup> and 12.0 µg/cm<sup>2</sup>, respectively. Furthermore, substances with defined retention times and fragmentation patterns corresponding to those supposed for Roridin E, Roridin L-2, Satratoxin F and Verrucarín J could be identified (Figure 1). A co-occurrence of these toxins with Satratoxin G and H has been described formerly by Andersen *et al.* (5). However, a definite identification by the given methods was not possible due to lacking standard substances. Roridin A and Verrucarín A were not detected.

It can be concluded that the occurrence of Satratoxins in such high amounts implicates a severe contamination of indoor environments. However, these toxins were determined in indoor materials and not in indoor air or dust. Thus, a direct exposure of humans to these toxins is not yet proven. More information about the possible health risk will be gained in a further study, examining dust and/or air from the locations where the samples have been drawn.

**Table 3.** Results of LC-MS/MS analysis for 8 macrocyclic trichothecenes (ng/cm<sup>2</sup>)

	Roridin A	Roridin E*	Roridin L-2*	Satratoxin F*	Satratoxin G	Satratoxin H	Verrucarín A	Verrucarín J*
Sample								
Gypsum board 1	-	-	-	-	18	22	-	-
Wallpaper 1a**	-	-	-	-	27	20	-	-
Wallpaper 1b**	-	+	-	-	740	740	-	-
Wallpaper 2a**	-	+++	-	-	1560	3260	-	-
Wallpaper 2b**	-	++	++	+	9650	12000	-	++
Wallpaper 3	-	+++	++	++	260	360	-	++

\* Quantitation not possible lacking standard substances \*\* a/b: Different spots of one sampled area

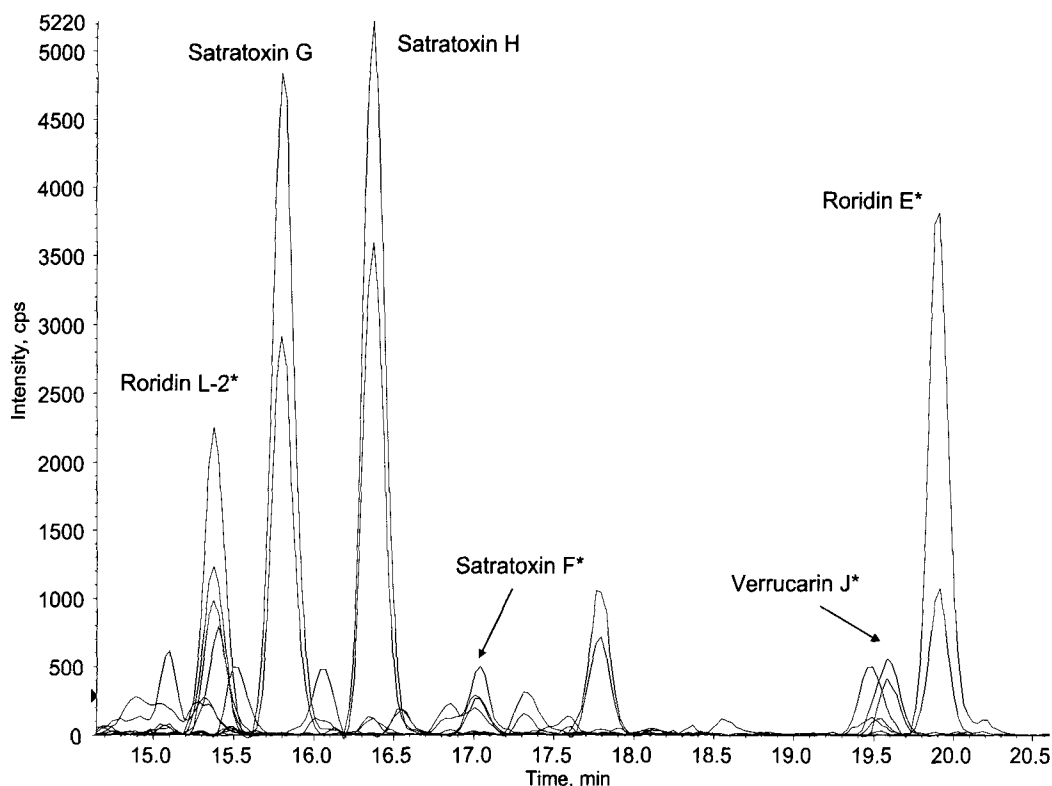


Figure 1. Extracted ion chromatogram (LC-MS/MS) of a wallpaper-extract

## References

- Johanning E, Biagini R, Hull D, Morey P, Jarvis B, Landsbergis P (1996) Health and immunology study following exposure to toxigenic fungi (*Stachybotrys chartarum*) in a water-damaged office environment. *Int Arch Occup Environ Health* 68: 207-218
- Jarvis BB, Sorenson WG, Hintikka EL, Nikulin M, Zhou Y, Jiang J, Wang S, Hinkley S, Etsel RA, Dearborn D (1998) Study of toxin production by isolates of *Stachybotrys chartarum* and *Memmoniella echinata* isolated during a study of pulmonary hemosiderosis in infants. *Appl Env Microb* 64: 3620-3625
- Jarvis BB, Lee Y-W, Comezoglu SN, Yatawara CS (1986) Trichothecenes produced by *Stachybotrys atra* from Eastern Europe. *Appl Env Microb* 51: 915-918
- Gravesen S, Nielsen PA, Iversen R, Nielsen KF (1999) Microfungal contamination of damp buildings – examples of risk constructions and risk materials. *Env Health Persp* 107 (Supplement 3): 505-508
- Andersen B, Nielsen KF, Thrane U, Szaro T, Taylor JW, Jarvis BB (2003) Molecular and phenotypic descriptions of *Stachybotrys chlorohalonata* sp. nov. and two chemotypes of *Stachybotrys chartarum* found in water-damaged buildings. *Mycologia* 95 (6): 1227-1238