

*Genetic Resources*

## The Chloroplast Genome of a Chlorophyll *a+c*-Containing Alga, *Odontella sinensis*

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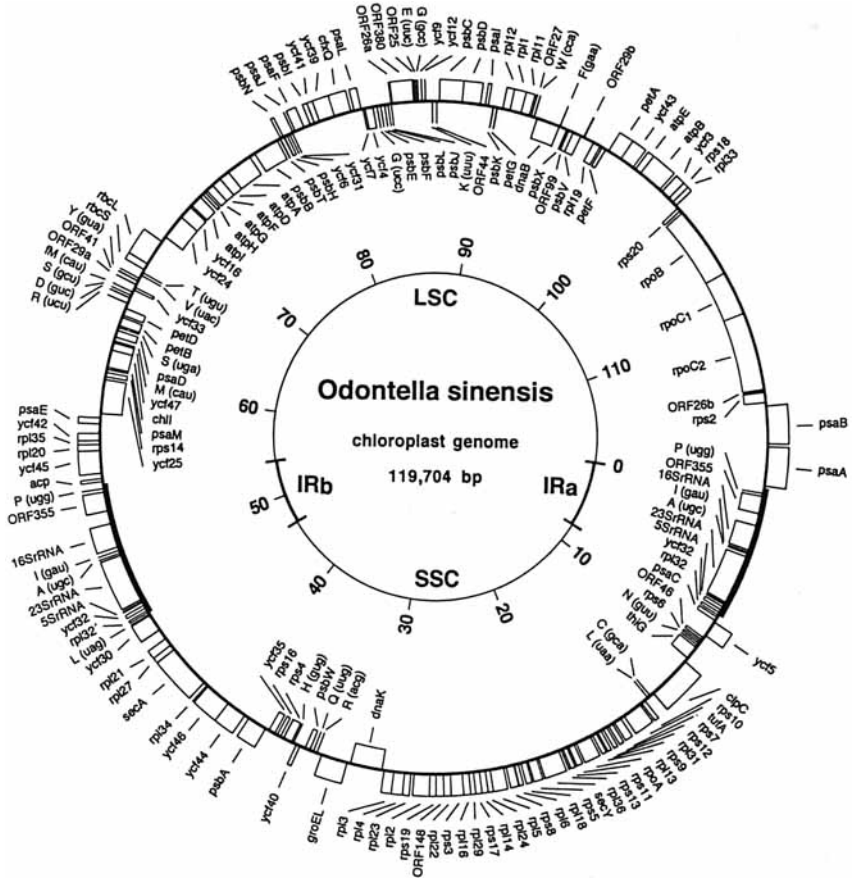
**Abstract:** The chloroplast genome of a marine centric diatom, *Odontella sinensis*, was cloned and sequenced. The circular genome is 119,704 bp in length (AC = Z67753;). It contains an inverted repeat sequence of 7,725 bp separating two single-copy regions of 38,908 and 65,346 bp, respectively, and 174 genes and open reading frames, of which nine are duplicated within the inverted repeat segments.

**C**hloroplast genomes from algae not belonging to the chlorophyll *a+b*-containing lineage have only recently been characterized by their entire nucleotide sequences. These include the plastid genome of *Porphyra purpurea* (191,028 bp; Reith & Munholland, 1995) and the cyanelle genome of the enigmatic unicellular flagellate *Cyanophora paradoxa* (135,599 bp; Stirewalt et al., 1995). To date, chloroplast genes from chlorophyll *a+c*-containing algae have been sequenced arbitrarily. They included genes that are known from land plant chloroplast genomes (e.g., *psaA*; *psbA*, -C, -D; *atpB*, -E, -I, -H, -F, -A; *rbcl*; *petB*, -D; cf. Kowallik, 1994), but also some that are novel or known to be nuclear in land plants (cf. Valentin et al., 1993).

In order to establish evolutionary traits among non-green plastids, the chloroplast genome of the centric diatom *Odontella sinensis* was cloned and sequenced. It contains 119,704 bp. An inverted repeat (IR) segment of 7,725 bp, containing the rRNA operon, separates a small single-copy (SSC) region of 38,908 bp from a large single-copy (LSC) region of 65,346

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**Abbreviations:** IR, inverted repeat; LSC, large single-copy region; nt, nucleotide; SSC, small single-copy region.



**Fig. 1. Gene map of the circular chloroplast genome of *Odontella sinensis*.** Genes transcribed clockwise are shown inside the circle, those transcribed counter-clockwise, are shown outside the circle. The inverted repeats are indicated by bold-type segments. Counting of the sequence starts at the right-hand inverted repeat (IRa) and continues clockwise.

bp (Fig. 1). The *Odontella* chloroplast genome reveals an overall coding capacity of 84.3 percent and has a (G+C)-content of 31.8 percent, which is equivalent to calculations based on the density of purified plastid DNA by equilibrium density sedimentation in gradients of CsCl (Kowallik, 1989).

To date, 174 genes and open reading frames (minimum of 25 amino acid residues) have been determined, of which nine are duplicated within the IR (Table I). No intron sequences or *ndh* genes have been found. Four pairs of genes show a 3'-5' overlap (*rpl4/rpl23*, 8 nt; *ycf24/ycf16*, 1 nt; *atpF/atpD*, 4 nt; *psbD/psbC*, 53 nt). All three stop codons are

**Table I. List of genes and ORFs of the *Odontella sinensis* chloroplast genome.** The gene list proceeds clockwise from the LSC boundary of the right-hand inverted repeat (IRA). Genes are listed together with their putative gene products. ORFs with a (G+C)-content less than 30% (ORFs 25, 26a, 26b, 27, 29a, 29b, 41, 44, 46) are designated (A+T)-rich and may be non-coding. Numerical positions include the first nucleotide of the translation initiation codon as well as the last nucleotide of the stop codon. Transcription initiation and termination sites of tRNAs and rRNAs were determined according to multiple alignments and secondary structure models. Genes marked "(–)" are on the complementary strand.

Gene	Gene Product	Coding Sites	
	<i>start IRA</i>		1
<i>trnP</i>	tRNA-Pro(ugg)	267	340
ORF355		478	1545
<i>rrn16</i>	16S ribosomal RNA	2210	3694
<i>trnI</i>	tRNA-Ile(gau)	3761	3835
<i>trnA</i>	tRNA-Ala(ugc)	3838	3910
<i>rrn23</i>	23S ribosomal RNA	3976	6866
<i>rrn5</i>	5S ribosomal RNA	6938	7063
<i>ycf32</i>	ORF36	7318	7428
<i>rpl32</i>	50S ribosomal protein L32	7563	7739
	<i>end IRA</i>	7725	
<i>psaC</i>	PSI, Fe-S polypeptide SU VII, 9 kDa	7825	8073
<i>ycf5</i>	ORF312 (–)	8139	9077
ORF46	(A+T)-rich	9287	9427
<i>rps6</i>	30S ribosomal protein S6	9482	9772
<i>trnN</i>	tRNA-Asn(guu)	9852	9922
<i>thiG</i>	involved in thiamine biosynthesis	9944	10729
<i>clpC</i>	Clp protease, caseinolytic-like (–)	10894	13551
<i>trnC</i>	tRNA-Cys(gca)	13780	13850
<i>trnL</i>	tRNA-Leu(uaa)	13921	14006
<i>rps10</i>	30S ribosomal protein S10 (–)	14207	14518
<i>tufA</i>	elongation factor Tu (–)	14531	15760
<i>rps7</i>	30S ribosomal protein S7 (–)	15831	16307
<i>rps12</i>	30S ribosomal protein S12 (–)	16341	16721
<i>rpl31</i>	50S ribosomal protein L31 (–)	16773	16991
<i>rps9</i>	30S ribosomal protein S9 (–)	17005	17421
<i>rpl13</i>	50S ribosomal protein L13 (–)	17445	17864
<i>rpoA</i>	RNA polymerase $\alpha$ -chain (–)	17893	18831
<i>rps11</i>	30S ribosomal protein S11 (–)	18883	19275
<i>rps13</i>	30S ribosomal protein S13 (–)	19320	19691
<i>rpl36</i>	50S ribosomal protein L36 (–)	19713	19826
<i>secY</i>	preprotein-translocase subunit Y (–)	19866	21143
<i>rps5</i>	30S ribosomal protein S5 (–)	21166	21723
<i>rpl18</i>	50S ribosomal protein L18 (–)	21765	22172
<i>rpl6</i>	50S ribosomal protein L6 (–)	22219	22758
<i>rps8</i>	30S ribosomal protein S8 (–)	22777	23175

<i>rpl5</i>	50S ribosomal protein L5 (-)	23209	23925
<i>rpl24</i>	50S ribosomal protein L24 (-)	23957	24190
<i>rpl14</i>	50S ribosomal protein L14 (-)	24191	24556
<i>rps17</i>	30S ribosomal protein S17 (-)	24569	24823
<i>rpl29</i>	50S ribosomal protein L29 (-)	24868	25137
<i>rpl16</i>	50S ribosomal protein L16 (-)	25144	25557
<i>rps3</i>	30S ribosomal protein S3 (-)	25609	26253
<i>rpl22</i>	50S ribosomal protein L22 (-)	26295	26642
<i>ORF148</i>	(-)	26691	27137
<i>rps19</i>	30S ribosomal protein S19 (-)	27149	27427
<i>rpl2</i>	50S ribosomal protein L2 (-)	27472	28299
<i>rpl23</i>	50S ribosomal protein L23 (-)	28324	28632
<i>rpl4</i>	50S ribosomal protein L4 (-)	28625	29272
<i>rpl3</i>	50S ribosomal protein L3 (-)	29320	29910
<i>dnaK</i>	Hsp70-type chaperone	30213	32057
<i>groEL</i>	chaperonin, 60 kDa (-)	32125	33711
<i>trnR</i>	tRNA-Arg(acg)	33891	33964
<i>trnQ</i>	tRNA-Gln(uug)	34020	34089
<i>psbW</i>	PSII, protein W, 13 kDa	34326	34673
<i>ycf40</i>	ORF73, homologous to <i>Porphyra</i> ORF71 (-)	34997	35218
<i>trnH</i>	tRNA-His(gug)	35294	35368
<i>rps4</i>	30S ribosomal protein S4	35435	35955
<i>rps16</i>	30S ribosomal protein S16	36238	36477
<i>ycf35</i>	ORF128	36607	36993
<i>psbA</i>	PSII, D1 reaction-center protein (-)	37339	38421
<i>ycf44</i>	ORF382, homologous to <i>Porphyra</i> ORF437 (-)	38642	39790
<i>ycf46</i>	ORF497, homologous to <i>Porphyra</i> ORF491 (-)	39799	41292
<i>rpl34</i>	50S ribosomal protein L34 (-)	41404	41550
<i>secA</i>	preprotein-translocase subunit $\alpha$ (-)	41583	44249
<i>rpl27</i>	50S ribosomal protein L27 (-)	44272	44523
<i>rpl21</i>	50S ribosomal protein L21 (-)	44546	44863
<i>ycf30</i>	former <i>trsE</i> ( <i>rbcR</i> homolog) (-)	45509	46438
<i>trnL</i>	tRNA-Leu(uag) (-)	46484	46564
	<i>end IRb</i>	46634	
<i>rpl32'</i>	50S ribosomal protein L32' (-)	46632	46796
<i>ycf32</i>	ORF36 (-)	46931	47041
<i>rrn5</i>	5S ribosomal RNA (-)	47296	47421
<i>rrn23</i>	23S ribosomal RNA (-)	47493	50383
<i>trnA</i>	tRNA-Ala(ugc) (-)	50449	50521
<i>trnI</i>	tRNA-Ile(gau) (-)	50524	50598
<i>rrn16</i>	16S ribosomal RNA (-)	50665	52149
<i>ORF355</i>	(-)	52814	53881
<i>trnP</i>	tRNA-Pro(ugg) (-)	54019	54092
	<i>start IRb</i>		54358
<i>acp</i>	acyl carrier protein (-)	54453	54695
<i>ycf45</i>	ORF455, homologous to <i>Porphyra</i> ORF565 (-)	54953	56320
<i>rpl20</i>	50S ribosomal protein L20 (-)	56561	56905

<i>rpl35</i>	50S ribosomal protein L35 (-)	56915	57109
<i>ycf42</i>	ORF204, homologous to <i>Porphyra</i> ORF199 (-)	57112	57726
<i>psaE</i>	PSI, subunit IV (-)	57835	58050
<i>ycf25</i>	ORF 644, homologous to <i>E. coli</i> cell-division protein FtsH	58339	60273
<i>rps14</i>	30S ribosomal protein S14	60428	60730
<i>psaM</i>	PSI, protein M	60884	60976
<i>chlI</i>	chlorophyll biosynthesis, probable magnesium-chelatase subunit	61054	62115
<i>ycf47</i>	ORF74, homologous to <i>Porphyra</i> ORF71	62143	62367
<i>trnM</i>	tRNA-Met(cau)	62586	62659
<i>psaD</i>	PSI, ferredoxin-binding protein II	62744	63163
<i>trnS</i>	tRNA-Ser(uga)	63199	63286
<i>petB</i>	cytochrome b6	63341	63988
<i>petD</i>	cytochrome b6/f complex, subunit IV	64035	64517
<i>trnR</i>	tRNA-Arg(ucu) (-)	64774	64845
<i>trnD</i>	tRNA-Asp(guc) (-)	65093	65167
<i>trnS</i>	tRNA-Ser(gcu) (-)	65253	65340
<i>trnFM</i>	tRNA-fMet(cau) (-)	65380	65452
<i>ycf33</i>	ORF64	65614	65808
<i>ORF29a</i>	(A+T)-rich (-)	65920	66009
<i>ORF41</i>	(A+T)-rich (-)	66057	66182
<i>trnY</i>	tRNA-Tyr(gua) (-)	66350	66431
<i>trnV</i>	tRNA-Val(uac)	66555	66626
<i>trnT</i>	tRNA-Thr(ugu)	66647	66717
<i>rbcS</i>	Rubisco, small subunit (-)	66864	67283
<i>rbcL</i>	Rubisco, large subunit (-)	67323	68795
<i>ycf24</i>	ORF486	69086	70546
<i>ycf16</i>	ORF251	70546	71301
<i>atpI</i>	ATP synthase CF <sub>O</sub> subunit IV	71429	72157
<i>atpH</i>	ATP synthase CF <sub>O</sub> subunit III	72227	72475
<i>atpG</i>	ATP synthase CF <sub>O</sub> subunit II	72587	73057
<i>atpF</i>	ATP synthase CF <sub>O</sub> subunit I	73121	73660
<i>atpD</i>	ATP synthase CF <sub>1</sub> subunit $\delta$	73657	74220
<i>atpA</i>	ATP synthase CF <sub>1</sub> subunit $\alpha$	74268	75779
<i>psbB</i>	PSII, CP47 chlorophyll apoprotein	76048	77577
<i>psbT</i>	PSII, protein T, 3 kDa ( <i>ycf8</i> )	77629	77727
<i>psbN</i>	PSII, protein N (-)	77752	77883
<i>psbH</i>	PSII, phosphoprotein, 10 kDa	77966	78169
<i>ycf6</i>	ORF29	78255	78344
<i>ycf31</i>	ORF42	78384	78512
<i>psaJ</i>	PSI, subunit IX, 5 kDa (-)	78575	78700
<i>psaF</i>	PSI, subunit III, plastocyanin-binding (-)	78732	79289
<i>psbI</i>	PSII, protein I, 4.8 kDa (-)	79654	79770
<i>ycf41</i>	ORF113, homologous to <i>Porphyra</i> ORF111 (-)	79855	80196
<i>ycf39</i>	ORF319 (-)	80212	81171
<i>cfxQ</i>	involved in Rubisco-expression (-)	81201	82076
<i>psaL</i>	PSI, subunit XI (-)	82171	82623
<i>ycf7</i>	ORF31	82800	82895

<i>ycf4</i>	ORF181	82917	83462
<i>trnG</i>	tRNA-Gly(ucc)	83508	83578
<i>psbE</i>	cytochrome b559 $\alpha$ -chain	83634	83888
<i>psbF</i>	cytochrome b559 $\beta$ -chain	83901	84032
<i>psbL</i>	PSII, protein L	84068	84184
<i>psbJ</i>	PSII, protein J	84209	84328
<i>ORF26a</i>	(A+T)-rich (-)	84426	84506
<i>ORF380</i>		84538	85680
<i>ORF25</i>	(A+T)-rich (-)	85745	85822
<i>trnE</i>	tRNA-Glu(uuc) (-)	85882	85953
<i>trnG</i>	tRNA-Gly(gcc) (-)	85995	86066
<i>ycf9</i>	ORF61 (-)	86186	86371
<i>ycf12</i>	ORF34 (-)	86419	86523
<i>trnK</i>	tRNA-Lys(uuu)	86730	86801
<i>ORF44</i>	(A+T)-rich	86932	87066
<i>psbC</i>	PSII, CP43 chlorophyll apoprotein (-)	87163	88578
<i>psbD</i>	PSII, D2 reaction-center protein (-)	88526	89581
<i>psaI</i>	PSI, subunit VIII (-)	89824	89940
<i>psbK</i>	PSII, protein K	90146	90280
<i>petG</i>	cytochrome b6/f complex, subunit V	90417	90530
<i>rpl12</i>	50S ribosomal protein L12 (-)	90778	91161
<i>rpl1</i>	50S ribosomal protein L1 (-)	91234	91926
<i>rpl11</i>	50S ribosomal protein L11 (-)	91949	92374
<i>ORF27</i>	(A+T)-rich (-)	92456	92539
<i>trnW</i>	tRNA-Trp(cca) (-)	92660	92732
<i>dnaB</i>	DNA-replication helicase	92815	94182
<i>trnF</i>	tRNA-Phe(gaa) (-)	94184	94256
<i>psbX</i>	PSII, protein X, 4.1 kDa	94593	94709
<i>ORF99</i>		94783	95082
<i>psbV</i>	PSII, cytochrome c <sub>550</sub> ( <i>petK</i> )	95139	95630
<i>ORF29b</i>	(A+T)-rich (-)	95659	95748
<i>rpl19</i>	50S ribosomal protein L19	96389	96751
<i>petF</i>	ferredoxin	96895	97194
<i>petA</i>	apocytochrome f (-)	97278	98222
<i>ycf43</i>	ORF263, homologous to <i>Porphyra</i> ORF254 (-)	98256	99047
<i>atpE</i>	ATP synthase CF <sub>1</sub> subunit $\epsilon$ (-)	99151	99552
<i>atpB</i>	ATP synthase CF <sub>1</sub> subunit $\beta$ (-)	99566	100993
<i>ycf3</i>	ORF179 (-)	101162	101701
<i>rps18</i>	30S ribosomal protein S18 (-)	101792	102010
<i>rpl33</i>	50S ribosomal protein L33 (-)	102014	102208
<i>rps20</i>	30S ribosomal protein S20	102420	102701
<i>rpoB</i>	RNA polymerase $\beta$ -chain	102865	107004
<i>rpoC1</i>	RNA polymerase $\beta'$ -chain	107028	109559
<i>rpoC2</i>	RNA polymerase $\beta''$ -chain	109599	114044
<i>ORF26b</i>	(A+T)-rich	114058	114138
<i>rps2</i>	30S ribosomal protein S2	114159	114848
<i>psaB</i>	PSI, P700 apoprotein A2 (-)	114970	117171
<i>psaA</i>	PSI, P700 apoprotein A1 (-)	117351	119609

used, with a preference for TAA. Four genes start with GTG instead of ATG (*rps3*, *rps13*, *rpl23*, *rbcS*).

Identified genes are named according to Hallick and Bottomley (1983) and Hallick (1989). Open reading frames shared by homologous sequences of other chloroplast genomes are designated as *ycf* (Hallick and Bairoch, 1994), genes for tRNAs are labelled using the one-letter code of their respective amino acids. Open reading frames unique to *Odontella* are designated as ORF, followed by the number of encoded amino acid residues. Genes shared with the *Porphyra purpurea* chloroplast genome and the cyanelle genome of *Cyanophora paradoxa*, but unknown from land-plant chloroplast genomes, include *acp*, *atpD*, *atpG*, *dnaK*, *groEL*, *petF*, *psaE*, *psaF*, *psbV*, *psbW*, *psbX*, *rbcS*, *tufA*, *secY*, and *ycf16*, -24, -30, -31, -32, -33, -35, -39. In addition, several genes for ribosomal subunits (*rpl1*, -3, -6, -11, -12, -18, -34, -35, and *rps5*, -6, -10, -13, -17, 20) appear to be unique among plastid genomes of non-green algae. Whereas eight open reading frames (*ycf40-47*) have counterparts in the *Porphyra* chloroplast genome, none of these ORFs were found in the cyanelle genome.

A detailed description of the sequence and its phylogenetic implications will appear elsewhere.

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