# Effect of Dietary Linolenic Acid and Docosahexaenoic Acid on Growth and Fatty Acid Composition of Rainbow Trout (Salmo gairdneri)<sup>1,2</sup>

T.C. YU and R.O. SINNHUBER, Department of Food Science and Technology, Oregon State University, Corvallis, Oregon 97331

## ABSTRACT

Methyl linolenate  $18:3\omega 3$  and docosahexaenoate  $22:6\omega 3$  were incorporated in semipurified diets at several levels and fed to trout previously maintained on a fatfree diet. After 14 weeks, the weight gain

<sup>1</sup>Presented in part at the AOCS Meeting, Atlantic City, October 1971.

<sup>2</sup>Technical paper no. 3247, Oregon Agricultural Experiment Station.

#### TABLE I

#### **Composition of Trout Diets**

Component	Weight %		
Casein	53.9		
Gelatin	9.7		
Dextrin	17.1		
Lipida	2.0		
Mineral mix <sup>b</sup>	4.0		
Carboxymethylcellulose	1.4		
Alpha-cellulose (Alphacel-NBC)	8.7		
Vitamin mix <sup>c</sup>	2.0		
Choline chloride	1.0		
Vitamin E concentrate (330 IU/g as			
dl-a-tocopheryl acetate)	0.2		

<sup>a</sup>Lipid composition listed in Table II.

<sup>b</sup>Bernhart-Tomerelli Salt Mix, modified by adding NaF and CaCl<sub>2</sub> at 0.002 and 0.02%, respectively.

<sup>c</sup>Supplied vitamins at following levels: (mg/kg) thiamine, 64; riboflavin, 144; niacinamide, 512; biotin, 1.6; Ca D-pantothenate, 288; pyridoxine, 48; folic acid, 19.2; menadione, 16; cobalamine, 0.159; i-ino-sitol (meso-), 2500; ascorbic, 1200; and *p*-aminoben-zoic acid, 400. Vitamins A and D were added at 25,000 and 4000 IU/kg, respectively.

and feed conversion of the fish on each diet were determined. The fatty acid composition of the lipid from each group of fish was analyzed by gas liquid chromatography. Both  $18:3\omega3$  and  $22:6\omega3$ fed at the 1% level supported maximum growth of the fish. The control group, which were fed no  $\omega$ 3 fatty acids, exhibited a shock syndrome, poor appetite and a very slow growth rate. Tissue fatty acid analysis revealed eicosatrienoic acid  $20:3\omega 9$  accumulated in the phospholipid fraction of this group. The  $20:3\omega 9$  level was lowered when either  $18:3\omega 3$  or 22:6 $\omega$ 3 was included in the diet. Analysis showed that the dietary  $18:3\omega 3$  was rapidly converted by the fish into  $22:6\omega 3$ with a high concentration in the phospholipid. However 22:6 $\omega$ 3 fed to the fish remained unchanged and little or no retroconversion of this fatty acid was observed.

#### INTRODUCTION

Recent studies have demonstrated that fatty acids of the linolenic family ( $\omega 3$ ) are essential for rainbow trout, whereas acids of the linoleic family ( $\omega 6$ ) do not seem to be required. Lee and coworkers (1) reported that corn oil in a semipurified diet led to poor growth and high mortality of rainbow trout. Replacing the corn oil with fish oil greatly stimulated growth and prevented further mortalities. Castell and coworkers (2,3) demonstrated that trout grew poorly on a diet containing 1% linoleic acid as

Diet no.	L	ipid compositi	on, %		Accumulated mortality	
	12:0	18:3 <b>ω</b> 3	22:6w3	Feed efficiency, <sup>a</sup> gain/feed		
1	2.00	0	0	0.32	17	
2	1.75	0	0.25	0.80	0	
3	1.50	0	0.50	0.88	1	
4	1.00	0	1.00	0.91	1	
5	1.50	0.50	0	0.84	3	
6	1.00	1.00	0	0.90	1	

#### TABLE II

Effect of Dietary Lipids on Feed Efficiency and Mortality of Rainbow Trout

<sup>a</sup>Feed efficiency is defined as units of weight gained per unit of dry feed consumed.

the only unsaturated fatty acid. The fish developed a shock syndrome, excessive liver mitochondrial swelling and other symptoms. On the other hand, Castell reported excellent growth of the fish with a diet containing 1% linolenic acid and recommended this level be included in diets for rainbow trout. Their results supported the findings of Richardson et al. (4), Brockerhoff and Hoyle (5), Brenner et al. (6) and Higashi et al. (7) that linolenic acid was nutritionally more important for fish than linoleic acid.

So far only linolenic acid has been used to study the  $\omega 3$  fatty acid requirements of fish. The objective of this experiment was to isolate the long chain highly unsaturated docosahexaenoic acid 22:6 $\omega 3$  from fish oil and to compare the effect of this fatty acid with that of linolenic acid on growth and fatty acid composition of rainbow trout.

#### MATERIALS AND METHODS

Methyl linolenate  $18:3\omega 3$  and methyl laurate 12:0, both 99% pure, were obtained from the Hormel Institute. The source of docosahexaenoic acid  $22:6\omega 3$  was Pacific Coho salmon roe which had been freeze dried and the oil extracted with chloroform-methanol 2:1. The egg oil was converted to fatty acid methyl esters by use of sodium methoxide (8). The resulting esters were dissolved in acetone and subjected to low temperature crystallization at -80 C. After removal of the precipitated esters, the 22:6 $\omega$ 3 content increased from 21.0% to 30.6% as estimated by gas liquid chromatography (GLC). The 22:6 $\omega$ 3 was isolated from the ester mixture by argentation chromatography using a column packed with silicic acid containing 20% AgNO<sub>3</sub> (9). The elution solvents were similar to those described by Stein and Slawson (10) with the addition of equal parts of the antioxidants, butylated hydroxyanisole and butylated hydroxytoluene (0.001%) to prevent autoxidation of the un-

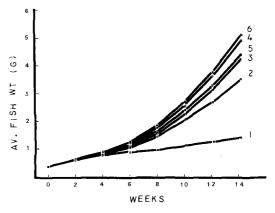


FIG. 1. Effect of dietary fatty acids on growth rate of rainbow trout. Curve 1, control trout diet containing no  $\omega 3$  fatty acid. Curves 2, 3 and 4, diets containing 0.25, 0.5 and 1.0% 22:6 $\omega 3$ , respectively. Curves 5 and 6, diets containing 0.5 and 1.0% linolenic acid, respectively.

saturated esters. The saturated and monounsaturated esters were eluted with diethyl etherpentane 1:1 and more highly unsaturated esters were eluted with diethyl ether-pentane-cyclohexane 5:3:2. Finally the ester of 22:6 $\omega$ 3 was eluted with diethyl ether-cyclohexane 1:1. Fractions containing high percentages of 22:6 $\omega$ 3 were pooled. The final product, as analyzed by GLC, contained 97.0% 22:6 $\omega$ 3; 2.5% 20:5 $\omega$ 3, and was free from 18:3 $\omega$ 3 and  $\omega$ 6 fatty acids. The isolated 22:6 $\omega$ 3 was finally put through a silicic acid column to remove the antioxidants (11).

Rainbow trout (Salmo gairdneri) was used as the experimental animal. Eggs from the brood stock of this laboratory were hatched and the fry fed a fat-free diet for one month and then randomly divided into lots of 50 fish. Each experimental diet was fed to duplicate lots of fish. The composition of the diets is shown in Table I. The casein, gelatin, dextrin and cellulose used in the diet had been extracted with warm isopropanol to remove trace lipids. The

Diet		Average fish wt (g) in weeks <sup>a</sup>							
	0	2	4	6	8	10	12	14	
1	0.53	0.76	0.97	0.97	1.03	1.21	1.35	1.56	
2	0.53	0.85	0.90	1.23	1.56	2.15	2.73	3.58	
3	0.50	0.78	0.92	1.19	1.63	2.39	3.23	4.32	
4	0.51	0.76	0.98	1.37	1.84	2.62	3.70	5.00	
5	0.53	0.79	0.94	1.28	1.75	2.51	3.37	4.47	
6	0.52	0.84	1.00	1.39	1.93	2.84	3.88	5.20	

Effect of Dietary Linide on Growth of Rainbow Trout

TABLE III

<sup>a</sup>Average of duplicate tanks.

LIPIDS, VOL. 7, NO. 7

#### TABLE IV

Percentage	Fatty	Acid	Composition	of	Phospholipids <sup>a</sup>
------------	-------	------	-------------	----	----------------------------

Fatty acids				Diet			
	Fat-free, 1 month	1	2	3	4	5	6
12:0		0.5	0.5	0.6	0.4	0.6	0.5
14:0	0.9	2.4	2.2	1.8	1.7	2.2	2.0
16:0	14.2	16.1	15.9	16.8	18.1	11.7	16.6
<b>16</b> :1 $\omega$ 7	9.7	14.1	12.7	11.9	10.5	12.1	10.5
18:0	7.2	5.1	4.6	5.3	5.7	4.8	5.0
18:1ω9	30.8	35.8	32.7	29.1	27.2	31.9	29.2
18:2w6 <sup>b</sup>	3.2	4.3	3.2	2.7	2.3	2.8	2.1
18:3 <i>w</i> 3						1.6	3.8
20:1ω11 <sup>b</sup>	3.5	2.9	2.6	2.5	1.8	2.3	2.0
18:4 <b>ω</b> 3						1.1	2.3
<b>20:2ω9</b>	2.4	2.5	2.5	2.3	2.1	2.4	1.4
20:3w9	4.2	8.6	5.9	4.8	3.0	3.6	1.5
20:4 <b>ω</b> 6	1.1	1.2	0.6	0.5	Trace	0.8	0.6
20:4ω3							0.6
20:5w3	Trace		Trace	0.9	0.8	1.7	1.5
22:5w6	Trace	0.6					
22:5w3	Trace					Trace	Trace
22:6 <b>ω</b> 3	23.0	5.7	16.8	21.0	26.7	16.1	20.9

<sup>a</sup>Average of analysis of duplicate tanks.

<sup>b</sup>Other isomers may be present.

quantity of the lipids in the diets was adjusted to 2% by varying the amount of the added 12:0 and all diets were isocaloric. The lipids added to each diet are listed in Table II.

The fish were held in 75 liter fiberglass tanks. The water temperature was 11.5 C, and flow rate was ca. 8 liters/min. The fish were fed three times daily. Food was offered only as long as the fish continued to feed.

Feed consumption and mortality were recorded. The fish were weighed every 2 weeks and the experiment was terminated at the end of 14 weeks.

The fatty acid composition of the pooled samples of each lot was determined. The lipid was extracted from the fish by the method of Folch et al. (12) and further separated into phospholipid and neutral lipid fractions as described previously (1). Methyl esters were prepared from these lipids by transesterification with boron-trifluoride in anhydrous methanol (13). Separation and identification of the component fatty acids was carried out as described by Lee et al. (1).

# **RESULTS AND DISCUSSION**

The growth rate was extremely slow for the group of fish receiving diet 1 (see Table I) which was devoid of  $\omega$ 3 fatty acids. The fish also exhibited poor appetite and a shock syndrome as described by Sinnhuber (14). Addition of  $\omega$ 3 fatty acids, even at a low level

(diet 2), to the diet vastly improved the condition of the fish and increased the growth rate (Fig. 1). None of the fish on supplemental diets exhibited the shock syndrome.

The accumulated mortality was high in the group of fish on diet 1, without  $\omega_3$  acids. Incorporation of  $\omega_3$  fatty acids in the diets effectively lowered the mortality (Table II). The feed efficiency also increased with increasing levels of  $\omega_3$  fatty acids as shown in Table II. These results indicated that trout responded equally well to dietary 18:3 $\omega_3$  and 22:6 $\omega_3$ .

The growth of fish on diet 6, containing 1.0% 18:3 $\omega$ 3, was similar to that reported by Castell et al. (2,3). Their results showed that the growth rate of trout approached a maximum when 1% 18:3 $\omega$ 3 was added in the diet. The growth rate was considerably lower if only 0.5% of either 22:6 or 18:3 was incorporated in the diet (Table III).

The fatty acid composition of phospholipids and of neutral lipids is shown in Tables IV and V. The lipid extracted from the whole fish, after receiving a fat-free diet for one month was 1.14% and showed a high percentage of  $22:6\omega 3$ (23.0%) in the phospholipid fraction. The neutral lipid fraction contained ca. 10.0%  $22:6\omega 3$ . These values represent the carry-over from the egg. Calculation of the total amount of  $22:6\omega 3$  remaining in the fish after 14 weeks on diet 1 showed that fish had conserved ca. 70% of the original  $22:6\omega 3$ .

Fatty acids	Diet								
	Fat-free, 1 month	1	2	3	4	5	6		
12:0		4.9	9,4	9.1	5.2	7.5	5.5		
14:0	2.6	4.8	4.1	3.3	2.9	3.4	3.0		
16:0	15.8	14.7	14.6	15.9	16.5	16.0	17.5		
16:1 <b>ω</b> 7	11.5	13.6	12.7	13.2	14.5	13.4	12.6		
18:0	4.6	4.9	4.6	6.3	4.2	5.0	4.2		
$18:1\omega 9$	42.6	48.6	43.5	38.0	41.0	40.7	37.4		
18:2w6 <sup>b</sup>	3.6	2.1	1.8	2.9	2.3	1.8	1.6		
18:3w3					Trace	2.5	8.5		
20:1w11 <sup>b</sup>	3.3	3.8	3.8	3.3	2.5	2.9	2.8		
18:4 <b>ω</b> 3						0.7	1.5		
20:2ω9	1.2	1.8	1.8	2.0	2.1	1.5	1.4		
20:3 <i>w</i> 9	1.8	1.5	1.2	1.2	1.0	0.8	1.2		
<b>20:4ω6</b>	1.4								
20:5 <b>ω</b> 3	1.9			Trace	1.0				
22:5w3	Trace								
22:6w3	10.0		2.9	5.2	7.1	4.0	3.9		

TABLE V

Percentage Fatty Acid Composition of Neutral Lipids<sup>a</sup>

<sup>a</sup>Average of analysis of duplicate tanks.

<sup>b</sup>Other isomers may be present.

The acid 20:3 $\omega$ 9 was biosynthesized and found at high concentration in the phospholipids of fish which received only lauric acid (diet 1). The concentration of  $20:3\omega 9$  was lowered when dietary 18:3ω3 or 22:6ω3 increased (Table IV). Linolenic acid appeared to be somewhat more efficient than  $22:6\omega 3$  in reducing the level of  $20:3\omega 9$ . This is in agreement with the experiment of Brenner and Jose (15) which showed that the  $20:3\omega 9$  in liver and heart tissues of the fat deficient rats was lowered when  $18:3\omega3$  or  $22:6\omega3$  was included in their diet. They suggested that 22:6 $\omega$ 3 inhibited the incorporation of 20:3 $\omega$ 9 into the B position of the glycero phospholipids. As a result, further formation of the 20:3 $\omega$ 9 was inhibited. 18:3 $\omega$ 3 competitively inhibited the enzymatic desaturation and chain elongation reaction of  $18:1\omega 9$ .

The formation of  $18:4\omega 3$ ,  $20:4\omega 3$ ,  $20:5\omega 3$ ,  $22:5\omega 3$  and  $22:6\omega 3$  was observed in the phospholipids of the fish receiving dietary  $18:3\omega 3$  (diets 5 and 6). The concentration of  $22:6\omega 3$  was especially high. These results are quite similar to the conversion of linolenic acid into docosahexaenoic acid in kelp bass as reported by Kayama et al. (16). In the groups of fish placed on diets containing  $22:6\omega 3$  (diets 2-4), there was a very high concentration of  $22:6\omega 3$  in the phospholipids; while only a low concentration of  $20:5\omega 3$  was observed.

We believe the origin of the  $20:5\omega 3$  is from the small amount (2.5%) which is present in the  $22:6\omega 3$  supplement and is not the result of retroconversion. A positive answer will require further experimentation. No formation of 20:4 $\omega$ 3, 18:4 $\omega$ 3 and 18:3 $\omega$ 3 was detected in the lipids of the fish. Rat experiments conducted by Verdino and coworkers (17) and Schlenk et al. (18) showed that a large quantity of 20:4 $\omega$ 6 (essential to rats) was formed by retroconversion by fat deficient animals when placed on diets containing 22:5 $\omega$ 6. Only a trace of 18:2 $\omega$ 6 was formed. Furthermore the conversion of arachidonic acid to 18:2 $\omega$ 6 in rats was insignificant. It seems that 22:6 $\omega$ 3 in trout has a behavior similar to 20:4 $\omega$ 6 in rats.

Fish oils are generally low in  $18:3\omega3$  but rich in other  $\omega3$  fatty acids: namely,  $18:4\omega3$ ,  $20:5\omega3$ ,  $22:5\omega3$  and  $22:6\omega3$ . The total  $\omega3$ fatty acid content in fish oil is ca. 20-30%. This experiment reaffirmed the ability of fish to synthesize  $22:6\omega3$  from lower  $\omega3$  fatty acid and the capability of both  $18:3\omega3$  and  $22:6\omega3$ to alleviate essential fatty acid deficiency symptoms, increase feed efficiency and promote rapid growth.

This experiment stressed the importance of  $\omega 3$  fatty acids in trout nutrition but did not rule out a possible requirement for  $\omega 6$  fatty acids. A long term feeding experiment is in progress to determine whether trout can survive and reproduce without dietary  $\omega 6$  fatty acids.

#### ACKNOWLEDGMENTS

T. Will and R. Foster gave technical assistance. This investigation was supported in part by the National Oceanic and Atmospheric Administration Institutional Sea Grant GH 97.

## REFERENCES

- 1. Lee, D.J., J.N. Roehm, T.C. Yu and R.O. Sinnhuber, J. Nutr. 92:93 (1967).
- 2. Castell, J.D., R.O. Sinnhuber, J.H. Wales and D.J. Lee, Ibid. 102:77 (1972).
- 3. Castell, J.D., R.O. Sinnhuber, J.H. Wales and D.J. Lee, Ibid. 102:87 (1972).
- 4. Richardson, T., A.L. Tappel and E.H. Gruger, Jr., Arch. Biochem. Biophys. 94:1 (1961).
- 5. Brockerhoff, H., and R.J. Hoyle, Ibid. 102:452 (1963).
- 6. Brenner, R.R., D.V. Vazza and M.E. DeTomas, J. Lipid Res. 4:341 (1963).
- Higashi, H., T. Kaneko, S. Ishii, M. Ushiyama and T. Sugihasi, J. Vitaminol. 12:74 (1966).
  Nadenicek, J.D., and O.S. Privett, Chem. Phys.
- Lipids 2:409 (1968).
- 9. Kishimoto, Y., and N.S. Radin, J. Lipid Res. 4:437 (1963).
- 10. Stein, R.A., and V. Slawson, Anal. Chem.

40:2017 (1968). 11. Hardy, R., and J.N. Keay, J. Chromatogr. 27:474 (1967).

- 12. Folch, J., M. Lees and G.H.S. Stanley, J. Biol. Chem. 226:497 (1957).
- 13. Morrison, W.R., and L.M. Smith, J. Lipid Res. 5:600 (1964).
- 14. Sinnhuber, R.O., "Fish in Research," Edited by O.W. Neuhaus and J. Halver, Academic Press, New York, 1969.
- 15. Brenner, R.R., and P. Jose, J. Nutr. 85:196 (1965).
- 16. Kayama, M., Y. Tsuchiya, J.C. Nevenzel, A. Fulco and J.F. Mead, JAOCS 40:499 (1963).
- 17. Verdino, B., M.L. Blank, O.S. Privett and W.O. Lundberg, J. Nutr. 83:234 (1964).
- 18. Schlenk, H., J.L. Gellerman and D.M. Sand, Biochem. Biophys. Acta 137:420 (1967).

[Received January 19, 1972]