SHORT NOTE

C.H. Robinson • O.B. Borisova • T.V. Callaghan J.A. Lee

Fungai hyphal length in litter of *Dryas octopetala* **in a high-Arctic polar semi-desert, Svalbard**

Received: 21 February 1995/Accepted 27 March 1995

Abstract Mean fungal hyphal length was estimated in the surface horizons of a lithosol under *Dryas octopetala* in a polar semi-desert ecosystem at Ny-Alesund, Svalbard (78°56'N, 11°50'E). This site is the most northerly from which such values have been collected and provides the only record from polar semi-desert sites in the Eurasian high Arctic. Although mean $(+ SE)$ fungal hyphal length was the lowest recorded in surface horizons of northern tundra $(23 \pm 1 \text{ m g}^{-1})$, it was of the same order as results from a Canadian high-Arctic soil and Antarctic soils of high pH and low moisture and organic matter content, with little vegetation cover.

Introduction

Tundra ecosystems contain 11% of the world's soil carbon pool (Mellilo et al. 1990), and in these, soils hold 95% of the organically bound plant nutrients (Jonasson 1983). Even so, because of slow decomposition rates in cold, wet or dry soil environments, plant growth is often limited by nutrient availability (e.g. Wookey et al. 1994). Since saprotrophic fungi are primary decomposers of organic matter, and mycorrhizas of polar dwarf shrubs may facilitate plant nutrient uptake, fungal biomass¹, which can be estimated from fungal hyphal length, is a measure of a potentially major influence on the nutrition of (Arctic) plants through uptake, immobilisation and mineralisation of

¹ On etymological grounds, 'biomass' should be restricted to living matter, but here, as in many mycological studies, total mass is the intended meaning (Frankland et al. 1990)

C.H. Robinson (\boxtimes) \cdot O.B. Borisova \cdot T.V. Callaghan \cdot J.A. Lee Sheffield Centre for Arctic Ecology, Department of Animal and Plant Science, University of Sheffield, Sheffield, S10 2UQ, UK

nutrients (Frankland et al. 1990). Total fungal hyphal length is a measure of absolute fungal presence, and has been widely used in soil perturbation studies (e.g. Bardgett 1991) and site comparisons (review by Kjoller and Struwe 1982).

Despite the excellent synthesis of hyphal length values from various tundra ecosystems during the International Biological Programme (Dowding and Widden 1974; Holding 1981), such measurements were obtained from only one polar semi-desert site in the northern hemisphere, Truelove Lowland in Canada's Devon Island (Widden 1977), although polar desert and semi-desert tundra are characteristic of 93% of the high-Arctic land area (Bliss and Matveyeva 1992). In this study, we aimed to quantify fungal hyphal length in the surface soil horizons of one of the most northerly Eurasian polar semi-desert ecosystems, in order to compare these values with those from surface horizons of other tundra soils.

Materials and methods

The polar semi-desert site in situated within the *Dryas octopetala* L. zone of Svalbard (Brattbakk 1986), 3 km to the west of Ny-Alesund in northwestern Spitsbergen at an elevation of 22 m a.s.l.. Approximately 70% of the ground surface is bare of vegetation; the soil is a lithosol developed over limestone, and organic matter is mostly present under mats of *Dryas octopetala,* a winter-green prostrate dwarf shrub. The area is a well-drained interfluve with little sign of frost heave or sorting. Full details of the research site are provided in Wookey et al. (1993) and Robinson et al. (1995). Replicate plots, each measuring $1.5 \text{ m} \times 1.5 \text{ m}$, were allocated randomly in 1991. Three cores (51-mm diameter, *ca.* 10-g fresh weight) were taken randomly from the surface to 30 mm depth $($ = standing dead plus fallen litter) of 15 plots from under *D. octopetala* on 14 July 1994. At this site, in common with many previous tundra studies (Dowding and Widden 1974), only a 'summer' sampling was made due to practical difficulties in winter sampling and minimal microbiological facilities. No more than three soil samples were available per plot due to the lack of vegetation cover and organic soil on the plots. The soil samples were placed in plastic bags and stored at 2°C for up to 3 days. The gravimetric moisture content of each core was determined by oven-

 72

É

 $\frac{1}{2}$.
بدا

J $\frac{1}{2}$

ť $\frac{1}{2}$ \ddot{a}

J

j

٠ť. Í

ć.

drying a subsample at 105°C for 48 h. Fungal hyphal lengths were determined in a field laboratory, using the membrane filtration method (after Hanssen et al. 1974, modified by Sundman and Sivelä 1978). A sample of known weight *(ca.* 5-g wet weight) was homogenised in 100 ml of 60 mM phosphate buffer for 30 s. Ten millilitres of the homogenate was taken, diluted to 50 ml and a 5-ml sample to which one drop of 0.1% phenolic aniline blue had been added was filtered through a membrane filter (1.2-gm pore size). Each filter was blotted dry and mounted in immersion oil. Five filters were prepared per core, and total hyphal lengths were estimated in five microscope fields of view on each filter by a grid intersection method (Olson 1950) using \times 500 magnification. Fungal diameter was estimated for each hyphal fragment in one randomly selected field of view from each filter to give a mean diameter for 100 hyphae per plot. Mycelial biomass was estimated from hyphal length as in Frankland et al. (1978). The pH of a soil subsampte from each core was estimated after Allen (1989).

Results and Discussion

The range of values obtained was between 13 and 32 m g^{-1} , with a median of 23 m g^{-1} , and a mean $(\pm \text{SE})$ per plot of 23 \pm 1 m g⁻¹ (n = 15). These values have been estimated, we believe, from the most northerly latitude to date (see Table 1) and are extremely low compared to those from other biomes. For example, estimates of 1900-4432 m g^{-1} have been recorded for the litter layer of temperate woodland soils (review by Kjøller and Struwe 1982). The mean value is also considered to be the lowest reported from the surface horizons of tundra in the northern hemisphere, but is, however, of the same order as those at 30-cm horizontal distance from a mat of *Dryas integrifolia* M. Vahl. in a raised beach soil on Devon Island (39 m g^{-1}) , from a new moraine and a marble schist soil on Signy Island, Antarctica and from Mt. Allen in Canada at 2800 m (Table 1). Data available for Signy and Devon Islands showed that the above soils were dry, of high pH and contained low amounts of organic matter, similar to the polar semi-desert soils in the present study (Table 1). Fungal hyphal length in tundra soils has been shown to increase with soil temperature and moisture, together with decreasing soil pH (Dowding and Widden 1974). The summer soil temperature was relatively high at the Ny-Alesund site (Table 1), and thus, the low hyphal length could have been due to the relatively low moisture content and high pH of the soils present. Additionally, in polar semi-desert communities the major factor inhibiting saprotrophic fungal development could be a lack of organic matter rather than physical conditions (Widden and Parkinson 1979), or simply the lack of plant cover itself, which would reduce the possibility of including the ectomycorrhizal hyphae often associated with polar dwarf shrubs (Miller and Laursen 1974). The extremely small value in the present study may also represent an atypical seasonally low length, however, the maximum value (136 m g^{-1}) in Truelove Lowland was approximately only twofold higher than the minimum (Widden et al.

1972), and is still very low compared to non-lithosol estimates (Table 1). Using a mean (\pm SE) hyphal diameter of 2.99 (\pm 0.07) µm, the mean (\pm SE) fungal biomass value for the Ny-Ålesund site was 4.89×10^{-5} $(\pm 0.323 \times 10^{-5})$ g oven-dry weight mycelium per g oven-dry litter. With an estimated soil bulk density of 0.13 g cm^{-3} [calculated by the method of Harrison and Bocock (1981), using 79% loss-on-ignition determined from these soils] the fungal biomass was equivalent to 0.1 g m^{-2} which, although very low in comparison with other Arctic sites and other biomes (Kjøller and Struwe 1982; Syzova and Panikov 1995), is similar to the values obtained from standing dead material on a polygon rim (0.3 g m^{-2}) and from the litter $(0-20 \text{ mm})$ of a dry grass-covered polygon (0.5 g m^{-2}) in tundra at Barrow, Alaska (in Kjøller and Struwe 1982).

Measurements of fungal hyphal length recorded in Table 1, including ours, were not all carried out using the membrane filter technique, but all refer to total (living and dead) length. Ba $\hat{\text{a}}$ th and Söderström (1980) showed that hyphal length values were 1.1-3.9 times higher using the Jones and Mollison (1948) agar film technique than the membrane filter method. Even if the higher correction factor was used, the value obtained in our study (90 m g^{-1}) would be of the same order as those from the Devon Island beach ridge and the Signy Island old and new moraine soils, together with the marble schist soils, and many orders different to values from other tundra biome sites (Table 1).

This study has shown that, although the mean fungal hyphal length at this polar semi-desert site is very low compared to temperate and most other tundra values, it is similar to those from analogous lithosols in both Arctic and Antarctic regions.

Acknowledgements This study was funded by the Natural Environment Research Council (NERC) as part of the Arctic Terrestrial Ecology Special Topic Programme, We thank the staff of the Norsk Polarinstitutt and the Kings Bay Kull Compani at Ny-Alesund and N.I. Cox, manager of the NERC Arctic Research Station, for helping to provide facilities and logistical support, also Prof. Ian Alexander, University of Aberdeen, for use of the microscope. We thank Dr. Juliet C. Frankland, Institute of Terrestrial Ecology, Merlewood Research Station, for invaluable advice throughout this study. TVC acknowledges NERC for financial support.

References

- Allen SE (ed) (1989) Chemical analysis of ecological materials. Blackwell, Oxford
- Baath E, Söderström B (1980) Comparison of the agar-film and membrane-filter methods for the estimation of hyphal lengths in soil, with particular reference to the effect of magnification. Soil Biol Biochem 12:385-387
- Bailey AD, Wynn-Williams DD (t982) Soil microbiological studies at Signy Island, South Orkney Islands. Br Antarct Surv Bull 51:167-191
- Bardgett RD (1991) The use of the membrane filter technique for comparative measurements of hyphal lengths in different grassland sites. Agric Ecosystems Environ 34:115-119
- Bliss LC, Matveyeva NV (1992) Circumpolar arctic vegetation. In: Chapin FS, Jefferies RL, Reynolds JF, Shaver GR, Svoboda J (eds) Arctic ecosystems in a changing climate: an ecophysiological perspective. Academic Press, San Diego, pp *59-89*
- Brattbakk I (1986) Vegetasjonsregioner Svalbard og Jan Mayen. Nasjonalatlas for Norge, Kartblad 4.1.3
- Brown J, Veum AK (1974) Soil properties of the IBP sites. In: Holding AJ, Heal OW, MacLean SF, Flanagan PW (eds) Soil organisms and decomposition in tundra. Tundra Biome Steering Committe, Stockholm, pp 27-48
- Dowding P, Widenn P (1974) Some relationships between fungi and their environment in tundra regions. In: Holding AJ, Heal OW, MacLean SF, Flangan PW (eds) Soil organisms and decomposition in tundra. Tundra Biome Steering Committee, Stockholm, pp 123-150
- Frankland JC, Lindley DK, Swift MJ (1978) A comparison of two methods for the estimation of mycelial biomass in leaf litter. Soil Biol Biochem 10:323-333
- Frankland JC, Dighton J, Boddy L (1990) Methods for studying tungi in soil and forest litter. In: Grigorova R, Norris JR (eds) Methods in microbiology, 22. Academic Press, London, pp 343-404
- Hanssen JF, Goksoyr J (1975) Biomass and production of soil and litter fungi at Scandinavian tundra sites. In: Wielgolaski FE (ed) Fennoscandian tundra ecosystems. Part 1. Plants and microorganisms. Ecological Studies 16, Springer, Berlin Heidelberg New York, pp 239-243
- Hanssen JF, Thingstad TF, Goksoyr J (1974) Evolution of the hyphal length and fungal biomass in soil by a membrane filter technique. Oikos 25:102-107
- Harrison AF, Bocock KL (1981) Estimation of soil bulk-density from loss-on-ignition values. J Appl Ecol 18:919-927
- Hayes AJ (1973) Studies on the microfungi occurring at Stordalen and Njulla, 1972. Swedish IBP Tundra Project, Tech Rep 15
- Heat OW, Flanagan PW, French DD, MacLean SF (1981) Decomposition and accumulation of organic matter. In: Bliss LC, Heal OW, Moore JJ (eds) Tundra ecosystems: a comparative analysis. IBJ 25, Cambridge University Press, Cambridge, pp 587-633
- Holding AJ (1981) The microflora of tundra. In: Bliss LC, Heal OW, Moore JJ (eds) Tundra ecosystems: a comparative analysis. IBP 25, Cambridge University Press, Cambridge, pp 561-585
- Jonasson S (1983) Nutrient content and dynamics in north Swedish shrub tundra areas. Holarct Ecol 6:295-304
- Jones PTC, Mollison JE (1948) A technique for the quantitative estimation of microorganisms. J Gen Microbiol 2:54-69
- Kjøller A, Struwe S (1982) Microfungi in ecosystems: fungal occurrence and activity in soil. Oikos 39:389-422
- Mellilo J, Callaghan TV, Woodward FI, Salati E, Sinha SK (1990) Effects on ecosystems. In: Houghton JT, Jenkins GT, Ephraums JJ (eds) Climate change, the IPCC scientific assessment. Cambridge University Press, Cambridge, pp 282-310
- Miller OK, Laursen GA (1974) Belowground fungal biomass on US tundra biome site at Barrow, Alaska. In: Holding AJ, Heal OW, MacLean SF, Flanagan PW (eds) Soil organisms and decomposition in tundra. Tundra Biome Steering Committee, Stockholm, pp 151-158
- Olson FCW (1950) Quantitative estimates of filamentous algae. Trans Am Microsc Soc 59:272-279
- Rheinberg P (1974) Studies on the microfungi occurring at Stordalen and Njulla, 1973. Swedish IBP Tundra Project, Tech Rep 17
- Robinson CH, Wookey PA, Parsons AN, Potter JA, Callaghan TV, Lee JA, Press MC, Welker JM (1995) Response of plant litter decomposition, and nitrogen mineralisation to simulated environmental change in a high arctic polar semi-desert and a subarctic dwarf shrub heath. Oikos.
- Sundman V, Sivelä SA (1978) A comment on the membrane filter technique for estimation of length of fungal hyphae in the soil. Soil Biol Biochem 10:399-401
- Syzova MV, Panikov NS (1995) Biomass and composition of microbial communities in soils of Northern Russia. In: Callaghan TV, Oechel WC, Gilmanov T, Holten JI, Maxwell B, Molau U, Sveinbjörnsson B, Tyson M (eds) Global change and arctic terrestrial ecosystems. Commission of the European Communities Ecosystem Research Report. Brussels
- Widden P (1977) Microbiology and decomposition on Truelove Lowland. In: Bliss LC (ed) Truelove Lowland, Devon Island, Canada, a high arctic ecosystem. University of Alberta Press, Alberta, pp 505-530
- Widden P, Parkinson D (1979) Populations of fungi in a high arctic ecosystem. Can J Bot 21:2408-2417
- Widden P, Newell T, Parkinson D (1972) Decomposition and microbial populations of Truelove Lowland, Devon Island. In Bliss LC (ed) Devon Island IBP project high arctic ecosystem. University of Alberta Press, Alberta, pp 341-368
- Wookey PA, Parsons AN, Welker JM, Potter JA, Callaghan TV, Lee JA, Press MC (1993) Comparative responses of phenology and reproductive development to simulated environmental change in sub-arctic and high arctic plants. Oikos 67:490-502
- Wookey PA, Welker JM, Parsons AN, Press MC, Callaghan TV, Lee JA (1994) Differential growth, allocation and photosynthetic responses of *Polygonum viviparum* to simulated environmental change at a high arctic polar semi-desert. Oikos 70:131-139