

Influence of species of VA mycorrhizal fungi on cassava yield response to phosphorus fertilization

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Summary At three different sites with acid soils in Colombia field trials with cassava were monitored for frequency of VA mycorrhiza. Increasing levels of phosphorus (P) from 0 to 200 kg P/ha had been applied. The fields differed in the composition of species of VA mycorrhizal fungi. At all sites infections of the roots by the total mycorrhizal population decreased with increasing P fertilization, but at two sites the relative frequency and activity of one species, *Glomus manihotis*, increased with increasing P applications. This species was only present at two sites, and only in these sites a cassava yield response to up to 200 kg P was found. The differential activity of fungal species was confirmed in greenhouse trials, where *Entrophospora colombiana* was found to be most effective at 50 kg P and *G. manihotis* at 200 kg P.

Introduction

Cassava, *Manihot esculenta* Crantz, is a crop extremely well-adapted to acid-infertile soils with low phosphorus (P) content, so common in tropical America¹⁷. On these marginal soils cassava is grown as sole crop or as the last crop in the rotation before a fallow period³; however, without fertilization yields are generally very low. P deficiency is the major limitation for production. Yield responses to applications as high as 400 kg P/ha have been found, and levels of 100 to 150 kg P/ha are frequently recommended³. It is well known that cassava obligately depends on vesicular arbuscular (VA) mycorrhizal fungi for its P nutrition^{10,13,22}. An effective mycorrhizal association is required for P uptake and plant growth in nearly all soils¹¹. The VA mycorrhizal fungi take up P from the same sources which are also available to the plant roots¹⁴. Thus, theoretically, an increase of P into the labile pool by fertilization would be expected to increase the P uptake ability of the mycorrhizal root system of cassava. However, in many trials it was found that the cassava yield response to increasing P fertilization is highly dependent on the trial site^{2,7,9,15}. The present study was conducted to determine whether the site specificity for P response is related to the composition of the native mycorrhizal population.

Increasing P supply frequently inhibit the percentage of root length

infected by mycorrhizal fungi and therefore the beneficial activity of the endophytes may be decreased¹⁴. On the other hand, VA mycorrhizal fungi are known to consist in soils with very high P contents⁴, and thus, among the species of mycorrhizal fungi there may be differences in tolerance to low or high soil P conditions or to P fertilization.

Materials and methods

Field trials

Three established NPK trials with cassava were monitored for frequency of VA mycorrhiza. Two trials were located in the mountain region of the Cauca Department in South Colombia (at Agua Blanca with cassava cv M Col 113 and Barranquena, and at CIAT Quilichao with cv M Col 113 and M Col 1684). One trial was conducted in the Eastern Plains of Colombia in the Meta department (at Carimagua with cv M Ven 77 and M Col 638). The soils at the trial sites were acid clay loams with low nutrient availability and high Al saturation (Table 1). From the NPK trials only the following completely randomized treatments were considered: 100-0-100, 100-50-100, 100-100-100 and 100-200-100 kg/ha of N-P-K. Fertilizers were applied at time of planting in short bands at about 10 cm from the planting stake, N as urea, P as triple superphosphate and K as KCl. N and K were fractionated, half at planting and half at two months after planting. Fine roots were collected twice during the rainy seasons, except at Quilichao where the second sampling was conducted at the end of a dry season. At the first sampling roots were collected from 4 or 5 plants towards the center of each plot of each of the four replications at a distance of 5-20 cm from the plant's base and from 0-20 cm depth at 8 (Agua Blanca and Quilichao) or 5.5 (Carimagua) months after planting. The second sample was taken at harvest (14 months after planting at Agua Blanca and 12 months after planting at the other sites) by collecting fine roots from the soil after pulling up the whole plant for determination of yields. Representative subsamples of roots were processed and assessed for infection by VA mycorrhizal fungi, as reported elsewhere¹⁶. In addition to the infection of the roots by either hyphae, arbuscules and vesicles, the infection by vesicles alone was determined.

At the first sampling, representative soil samples from the zero fertilizer plots of the NPK trials were used to multiply in the greenhouse the native fungal species with *Pueraria phaseoloides* as host plant⁵. After one year, fungal spores were separated from these pots and the different species of VA mycorrhizal fungi were identified using keys^{18,21}. The ability of the species to form vesicles in the roots was determined in separate greenhouse trials with cassava cv CM 91-3 grown in sterilized soil from Quilichao and inoculated with pure isolates of the fungi.

Greenhouse trial

Soil from Quilichao was pasteurized with steam for 4 h and incubated with the equivalent of 0, 50, 100 and 200 kg P/ha as triple superphosphate. These applications resulted in extractable P (with 0.1N HCl + 0.03N NH₄F) of 2.9, 6.3, 11.2 and 21.3 µg P/g soil. In order to investigate solely the effect of mycorrhizal fungi on P uptake, each pot of 2 kg soil received a solution application of 285 mg NH₄NO₃, 190 mg KCl, 221 MgCl₂ · 6 H₂O and 10 mg ZnCl₂. Before planting of rooted tip cuttings of cassava cv M Ven 77 (one plant per pot), the whole pot content was mixed with infected soil from pure pot cultures with mycorrhizal fungi. The spore number in this infected soil was determined. Each of three fungal species, *Glomus manihotis*, *Entrophospora colombiana* and *Glomus occultum* was established with 1000 spores per pot; also there was a mixture of the three species with 350 spores of each species per pot. Each of the three mycorrhizal treatments was replicated four times at each P level. After two months of growth, the plant tops were harvested and the P content was analyzed; all roots were separated from the soil for measuring the total root length as well as the mycorrhizal root length per plant using a method described earlier⁶.

Table 1. Chemical and physical characteristics of soils at the three trial sites

Site	Organic matter (%)	P		meq/100 g soil				Soil classification*
		pH	($\mu\text{g/g}$ soil)	Al	Ca	Mg	K	
Agua Blanca**	6.2	4.4	0.8	8.5	0.95	0.46	0.16	Dystrandept
Quilichao	7.1	4.3	1.8	2.8	1.76	0.71	0.18	Dystropept
Carimagua	2.2	4.1	1.5	3.1	0.37	0.17	0.08	Haplustox

* After 'Instituto Geográfico Agustín Codazzi'¹².

** Soil samples at Agua Blanca were taken before application of 1 t/ha dolomitic lime; at other sites 15 days after application.

Table 2. Effect of increasing P application on fresh yields (t/ha) of different cassava cultivars grown at different sites in Colombia

Site	Cassava cultivar	Plant part	P application (kg P/ha)				LSD 5%
			0	50	100	200	
Agua Blanca	M Col 113	Roots	16.1	25.9	21.6	23.0	5.7
		Top growth	5.9	12.8	12.5	11.9	2.3
	Barranqueña	Roots	13.2	25.3	24.8	24.6	9.0
		Top growth	7.3	16.0	19.7	19.8	6.7
Quilichao	M Col 113	Roots	15.1	30.4	34.6	39.4	7.1
		Top growth	4.7	10.2	10.7	15.7	4.5
	M Col 1684	Roots	14.7	29.2	28.2	33.7	7.6
		Top growth	4.2	7.1	6.7	11.1	3.6
Carimagua	M Ven 77	Roots	9.2	17.0	19.7	21.8	4.8
		Top growth	4.9	9.8	12.0	13.3	2.2
	M Col 638	Roots	7.0	12.4	14.1	18.1	4.3
		Top growth	5.3	11.9	14.6	16.6	3.3

Results

Field trials

Whereas no yield response to more than 50 kg P/ha was found at Agua Blanca, both cassava cultivars responded to up to 200 kg P/ha at Quilichao and Carimagua (Table 2).

At each trial site five or more species of mycorrhizal fungi were naturally present (Table 3). Of all species, a clear vesicle formation in the roots was found only with *G. manihotis* and *G. fasciculatum*. Thus each percentage vesicle infection of roots represents the infection with *G. fasciculatum* at Agua Blanca, with *G. manihotis* at Quilichao, and with both *G. manihotis* and *G. fasciculatum*, at Carimagua. The percentage infection by either hyphae, arbuscles or vesicles represent the root infection by the whole mycorrhizal population with its different species. Table 4 shows that at first root sampling with increasing P application the frequency of mycorrhiza decreased, *i.e.* infection by

Table 3. Species of VA mycorrhizal fungi at the three trial sites and information about vesicle formation inside roots of cassava cv CM-91-3

Agua Blanca		Quilichao		Carimagua	
Species	Vesicles formed	Species	Vesicles formed	Species	Vesicles formed
<i>Glomus microcarpum</i>	No	<i>Entrophospora colombiana</i>	No	<i>Entrophospora colombiana</i>	No
<i>Glomus fasciculatum</i>	Yes	<i>Glomus occultum</i>	No	<i>Acaulospora longula</i>	No
<i>Glomus occultum</i>	No	<i>Glomus manihotis</i>	Yes	<i>Gigaspora heterogama</i>	No
<i>Entrophospora colombiana</i>	No	<i>Acaulospora appendicula</i>	No	<i>Glomus occultum</i>	No
<i>Acaulospora appendicula</i>	No	<i>Gigaspora heterogama</i>	No	<i>Glomus fasciculatum</i>	Yes
				<i>Glomus manihotis</i>	Yes

all mycorrhizal fungi inclusive of *G. fasciculatum*. At Quilichao the frequency of *G. manihotis* increased more markedly than of all other fungi with application of 50 kg P/ha, and the decrease of this species was less marked at higher P rates. At Carimagua the root infection by *G. manihotis* and/or *G. fasciculatum* increased with increasing P fertilization while the infection by the whole mycorrhizal population decreased. From first to second root sampling the root infection by all non-vesicles forming fungi had increased at Agua Blanca and Carimagua, and decreased at Quilichao. Increasing P fertilization tended to affect the root infection in similar ways as at the first sampling. At second sampling, the root infection by the vesicles forming fungi was inconsistently influenced by increasing P applications. *G. manihotis* was present at low population at Quilichao. At both samplings, no differences in the frequency of all fungi in the roots were found between the two cassava cultivars at Agua Blanca. At Quilichao and Carimagua the frequency of mycorrhiza was lower in cv M Col 1684 and M Col 638, respectively, in comparison to the other cultivar.

Greenhouse experiment

Species of mycorrhizal fungi differed in their effectiveness to enhance growth and P uptake of cassava depending on the P level applied (Table 5). When the three fungal species were mixed in the inoculum, at the 50 kg P level the P uptake was similar to that of plants inoculated with *E. colombiana* alone; at the highest P level the P uptake was similar to those plants inoculated with *G. manihotis*. Total root length and infected root length of the plants had an optimum at 50 or 100 kg

Table 4. Effect of increasing P application on per cent mycorrhizal infection of different cassava cultivars grown at different sites in Colombia

Site	Sampling*	Cassava cultivar	Mycorrhizal frequency	P application (kg P/ha)				LSD 5%
				0	50	100	200	
Agua Blanca	1st	M Col 113	HAV**	38.6	22.7	16.6	23.0	4.1
			Ves**	15.2	4.3	4.7	7.0	2.0
		Barranqueña	HAV	31.8	12.9	17.3	17.6	3.7
			Ves	10.3	1.7	4.5	1.8	1.9
	2nd	M Col 113	HAV	50.2	50.4	37.5	40.2	4.9
			Ves	3.8	2.2	3.8	4.0	ns
		Barranqueña	HAV	36.6	52.7	36.3	45.1	3.4
			Ves	5.3	6.6	8.9	10.0	1.9
Quilichao	1st	M Col 113	HAV	66.4	79.3	50.9	47.1	5.9
			Ves	31.4	60.8	46.6	34.8	4.6
		M Col 1684	HAV	66.4	67.8	39.7	40.0	6.9
			Ves	27.0	51.6	31.0	24.6	5.9
	2nd	M Col 113	HAV	47.7	45.9	38.6	38.1	3.4
			Ves	2.3	5.3	2.9	3.3	1.0
		M Col 1684	HAV	39.2	47.6	23.5	20.7	3.8
			Ves	5.2	10.6	6.3	3.2	2.4
Carimagua	1st	M Ven 77	HAV	63.8	54.6	35.1	32.9	4.9
			Ves	0.0	6.8	7.1	12.9	2.1
		M Col 638	HAV	58.3	43.3	20.9	36.9	—***
			Ves	0.0	9.6	8.2	11.9	—
	2nd	M Ven 77	HAV	70.7	60.3	56.3	54.3	—
			Ves	8.7	6.0	6.8	11.0	—
		M Col 638	HAV	46.3	30.9	76.0	43.8	—
			Ves	2.1	2.7	36.5	4.5	—

* 1st sampling at 5.5 MAP (Carimagua) or 8 MAP (others); 2nd at harvest.

** Frequency of either hyphae, arbuscules or vesicles (HAV) or of vesicles alone (Ves).

*** No statistical analysis was possible as observations were from bulked samples of all 4 replications.

Table 5. Effect of increasing P application on shoot dry matter and P uptake of cassava cv M Ven 77 inoculated with different species of mycorrhizal fungi

Species of mycorrhizal fungi	Dry matter (g/plant)				P uptake (mg/plant)			
	P application (kg/ha)				P application (kg/ha)			
	0	50	100	200	0	50	100	200
<i>G. manihotis</i>	1.55	9.02	10.93	13.73	2.01	13.31	22.61	31.91
<i>G. occultum</i>	2.05	7.49	9.32	11.21	2.77	10.64	13.41	18.23
<i>E. colombiana</i>	2.77	14.44	15.13	14.21	2.87	17.64	21.14	27.91
Mixture*	2.53	15.52	14.20	14.89	2.68	18.09	26.78	30.91
LSD 5%			2.40				3.96	

* Mixture of the three species

Table 6. Effect of increasing P application on root length, mycorrhizal root length and mycorrhizal frequency in roots of cassava cv M Ven 77 inoculated with different species of mycorrhizal fungi

Species of mycorrhizal fungi	P application (kg/ha)	Total root length (m/plant)	Root length with mycorrhiza (m/plant)	Root length with vesicles (m/plant)	Mycorrhizal frequency (%)	
					HAV*	Vesicles
<i>G. manihotis</i>	0	5.8	3.3	1.2	56.9	20.7
	50	15.5	13.9	11.6	89.7	74.8
	100	20.2	18.3	12.8	90.6	63.4
	200	14.5	12.6	7.2	86.9	49.7
	LSD 5%	4.6	3.9	4.6	17.6	16.2
<i>G. occultum</i>	0	7.7	1.5	0	19.5	0
	50	25.6	3.2	0	12.4	0
	100	22.9	1.9	0	8.3	0
	200	19.7	1.3	0	6.6	0
	LSD 5%	9.9	n.s.*	—	n.s.	—
<i>E. colombiana</i>	0	7.9	3.1	0	39.2	0
	50	12.4	7.8	0	62.9	0
	100	11.0	5.7	0	51.8	0
	200	7.9	5.6	0	70.9	0
	LSD 5%	3.5	2.7	—	n.s.	—
Mixture of three species	0	6.0	5.4	0.9	90.0	15.0
	50	11.9	10.5	6.2	88.2	52.1
	100	8.2	7.3	4.0	89.0	48.8
	200	11.1	8.0	4.1	72.1	36.9
	LSD 5%	4.0	4.3	2.1	n.s.	16.2

* Percentage infection with either hyphae, arbuscules or vesicles.

** n.s. = not significant ($P = 0.05$).

P/ha depending on the fungal species (Table 6). The percentage mycorrhizal frequency only increased with P applications in plants inoculated with *G. manihotis*. With this species, inoculated alone or in mixture the frequency of vesicles had an optimum at 50 kg P.

Competition of species at different P levels

The ability of the vesicles forming fungi, *i.e.* *G. manihotis* and *G. fasciculatum* to compete with the other fungi for infecting the roots at increasing P application levels was calculated after Sieverding¹⁹ and is shown in Table 7. The competitive ability of *G. manihotis* increased with increasing P level in the mixture of fungi used in the greenhouse trial. In the field, at Agua Blanca the competitive ability of

Table 7. Relation* of the frequency of vesicles to the frequency of either hyphae arbuscles or vesicles; greenhouse trial and field trials (first observation of mycorrhizal frequency)

Trial	Cassava cultivar	Mycorrhizal fungi	P application (kg P/ha)			
			0	50	100	200
Greenhouse	M Ven 77	<i>G. manihotis</i>	0.36	0.83	0.70	0.57
		<i>G. occultum</i>	0	0	0	0
		<i>E. colombiana</i>	0	0	0	0
		Mixture**	0.17	0.59	0.55	0.51
Agua Blanca	M Col 113	Native species	0.39	0.19	0.28	0.30
	Barranqueña	Native species	0.32	0.13	0.26	0.10
Quilichao	M Col 113	Native species	0.47	0.77	0.92	0.74
	M Col 1684	Native species	0.41	0.76	0.78	0.62
Carimagua	M Ven 77	Native species	0	0.12	0.20	0.39
	M Col 638	Native species	0	0.22	0.39	0.32

* Calculated as:
$$\frac{\text{Mycorrhizal frequency of vesicles (\%)}}{\text{Mycorrhizal frequency of either hyphae, arbuscules or vesicles (\%)}}$$

** Mixture of the three species used in the greenhouse trial.

G. fasciculatum decreased by P application. At Quilichao, *G. manihotis* competed strongly with all other fungi in the range of 50–200 kg P. At Carimagua, both or one of the vesicles forming fungi intensified competitive ability with increasing P application: it may be assumed that it was *G. manihotis*, because *G. fasciculatum* was negatively affected by P fertilization at Agua Blanca.

Discussion

In this report we used the relative incidence of vesicles in the roots as an indication of the presence of *G. manihotis* and *G. fasciculatum*. This is feasible because anatomical structures of VA mycorrhizal fungi in the roots serve as such indicators¹. We concede that the calculations might be an underestimation of the presence of these fungi. From the vesicles forming fungi, *G. manihotis* is known to be one of the most effective species for increasing cassava growth on acid soils; *G. fasciculatum* is of low to moderate effectiveness¹¹. As shown by the greenhouse trials, species of VA mycorrhizal fungi may differ in their effectiveness with increasing P fertilization. From the literature there are also some indications that fungal species might differ in their adaptation to certain soil chemical conditions^{8,20}.

The field trials showed that all naturally occurring endophytes were able to make efficient use of P dressings of 50 kg P/ha added to those very low P containing soils. However, a growth response to higher P applications was not found in Agua Blanca since none of the

endophytes was adapted to those P levels. At the other two sites the P tolerant *G. manihotis* was natively present and there a growth response up to 200 kg P was found. The extent of the response to increasing P application depended on the relative enhancement of this endophyte in the roots, as was clearly visible when the sites are compared (Tables 2, 4 and 7).

The change in mycorrhizal incidence in the roots during the growth period of cassava may be partly due to the different development stages of the plant¹⁴; however, with cassava the main effect was due to the rainy and dry season¹⁹.

From the results we may conclude that the composition of the native VA mycorrhizal population determines the level of response to P application on acid low P soils. However, site specific recommendations of higher P applications will only be possible if a methodology is developed to determine rapidly the population of P tolerant species of VA mycorrhizal fungi in a specific field sample.

References

- 1 Abbott L K 1982 Comparative anatomy of vesicular-arbuscular mycorrhizas formed on subterranean clover. *Aust. J. Bot.* 30, 485–489.
- 2 CIAT 1985 Annual Report of the Cassava Program for 1982 and 1983. Centro Internacional de Agricultura Tropical, Cali, Colombia.
- 3 Cock J H 1985 *Cassava: New Potential for a Neglected Crop*. Westview Press, Boulder, Colorado. 191 pp.
- 4 Davis E A, Young J L and Rose S L 1984 Detection of high-phosphorus tolerant VAM-fungi colonizing hops and peppermint. *Plant and Soil* 81, 29–36.
- 5 Gerdemann J W and Trappe J M 1974 *The Endogonaceae in the Pacific Northwest*. Mycologia Memoir Nr. 5.
- 6 Giovannetti M and Mosse B 1980 An evaluation of techniques for measuring vesicular-arbuscular mycorrhizal infection in roots. *New Phytol.* 84, 489–500.
- 7 Gomez J and Howeler R H 1980 Cassava production in low fertility soils. *In Cassava Cultural Practices: Proceedings of a Workshop held in Salvador, Bahia, Brazil.*, 18–21. March 1980. Eds. E H Weber, M J C Toro and M Graham. International Development Research Centre, Ottawa, Canada, pp 93–102.
- 8 Hayman D S 1982 Influence of soils and fertility on activity and survival of vesicular-arbuscular mycorrhizal fungi. *Phytopathology* 72, 1119–1125.
- 9 Howeler R H 1980 Soil related cultural practices for cassava. *In Cassava Cultural Practices: Proceedings of a Workshop held in Salvador, Bahia, Brazil.*, 18–21 March 1980. Eds. E J Weber, M J C Toro and M Graham. International Development Research Centre, Ottawa, Canada, pp 59–69.
- 10 Howeler R H, Cadavid L F and Burckhardt E 1982 Response of cassava to VA mycorrhizal inoculation and phosphorus application in greenhouse and field experiments. *Plant and Soil* 69, 327–339.
- 11 Howeler R H and Sieverding E 1983 Potentials and limitations of mycorrhizal inoculation illustrated by experiments with field grown cassava. *Plant and Soil* 75, 245–261.
- 12 Instituto Geografico 'Agustin Codazzi' 1976 *Estudio General de Suelos de los Municipios de Santander de Quilichao, Piendamó, Morales, Buenos Aires, Cajibío y Caldono*. Volumen XII, No. 4, 466 pp.

- 13 Kang B T, Islam R, Sanders F E and Ayanaba A 1980 Effect of phosphate fertilization and inoculation with VA-mycorrhizal fungi on performance of cassava (*Manihot esculenta* Crantz) grown on an Alfisol. *Field Crop Research* 3, 83–94.
- 14 Mosse B 1981 Vesicular-Arbuscular Mycorrhizal Research for Tropical Agriculture. Hawaii Institute of Tropical Agriculture and Human Resources, Research Bull. 194, 82 p.
- 15 Okeke J E and Kang B T 1981 Evaluation of some major soils from Southern Nigeria for cassava production. *In Tropical Root Crops: Research Strategies for the 1980's*. Eds. E R Terry, K A Oduro and F Caveness. International Development Research Centre, Ottawa, Canada, pp 99–103.
- 16 Salinas J G, Sanz J I and Sieverding E 1985 Importance of VA mycorrhizae for phosphorus supply to pasture plants in tropical Oxisols. *Plant and Soil* (*In press*).
- 17 Sanchez P A and Salinas J G 1980 Low-input technology for managing Oxisols and Ultisols in Tropical America. *Adv. Agron.* 34, 279–406.
- 18 Schenck N C, Spain J L, Sieverding E and Howeler R H 1984 Several new and unreported vesicular-arbuscular mycorrhizal fungi (Endogonaceae) from Colombia. *Mycologia* 76, 685–699.
- 19 Sieverding E 1985 Influence of method of VA mycorrhizal inoculum placement on the spread of root infection in field-grown cassava. *J. Agric. Crop Sci.* 154, (*In press*).
- 20 Sylvia D M and Schenck N C 1983 Application of superphosphate to mycorrhizal plants stimulates sporulation of phosphorus-tolerant vesicular-arbuscular mycorrhizal fungi. *New Phytol.* 95, 655–661.
- 21 Trappe J M 1982 Synoptic keys to the genera and species of zygomycetous mycorrhizal fungi. *Phytopathology* 72, 1102–1108.
- 22 Zaag P van der, Fox R L, Pena R S and Yost R S 1979 Phosphorus nutrition of cassava including mycorrhizal effects on P, K, Zn and Ca uptake. *Field Crop Res.* 2, 253–263.