

Fig. 1. Kontrollkultur mit sehr ausgeprägter SDH-Aktivität, $\times 700$.

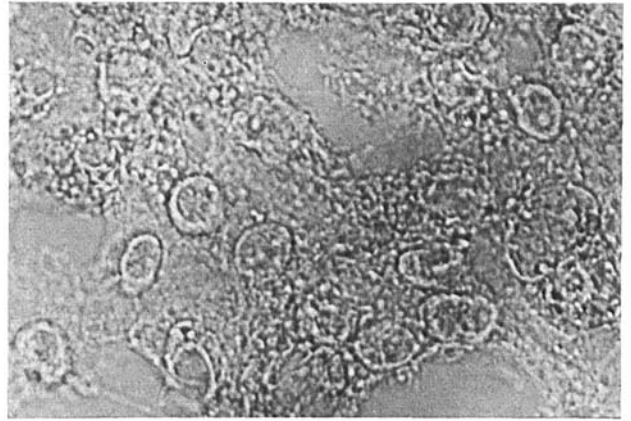


Fig. 2. Mit *Hepatitis-infectiosa*-Virus infizierte Kultur, die keine SDH-Aktivität aufweist, $\times 700$.

beobachtet, während sich die vollkommene Inhibition erst am Ende des 3. Tages nach der Infizierung zeigte.

Auf Grund dieser Ergebnisse kann man annehmen, dass die Hemmung der SDH-Aktivität nicht in direktem Zusammenhang mit der Biosynthese des Virus steht. Andererseits haben die elektronenoptischen Untersuchungen erwiesen⁴, dass in den von Viren infizierten Zellen die Mitochondrien tiefgehende Veränderungen erleiden. Es ist also wahrscheinlich, dass die Enzyminaktivierung den Veränderungen im Mitochondrialapparat zuzuschreiben ist. Da die mikroskopischen zytopathogenen Effekte der Virusinfektion erst 6 Tage nach der Infizierung festgestellt werden können, ist es für die frühzeitige Erkennung der Virusinfektion von besonderem Interesse, das Verhalten der SDH-Aktivität zu untersuchen.

Summary. The inhibitory action of *Hepatitis infectiosa* virus on the SDH activity of Detroit-6 (VA) cell lines was investigated. The full inhibition of the SDH activity took place at the end of the third day after the infection. As this phenomenon precedes the cytopathogenic effect

of viral infection, it may be of some help in the early detection of the infection.

F. WIENER, J. LÁSZLÓ
und K. SZÉKELY

*Institut für Biologie, Mikrobiologie und Virologie des
Medizinischen und Pharmazeutischen Instituts,
Tirgu-Mures (Rumänien), 2. August 1966.*

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A Polypeptide Antibacterial Agent Isolated from *Trichoderma viride*

Antibiotic U-22324 was discovered in the culture broth of the fungus *Trichoderma viride*. Fermentations were conducted under submerged culture conditions in a medium containing black strap molasses, 20 g/l; dextrin, 30 g/l; fish meal, 15 g/l; Pharmamedia (Traders Oil Mill Co., Fort Worth, Texas), 15 g/l; and tap water to make 1 l. The final pH of this medium was adjusted to 6.8 prior to sterilization. Antibiotic titers were measured by a microbiological disc-plate assay similar to the one described by HANKA et al.¹, using *Sarcina lutea* as the test organism. The log dose-response curve was linear over a concentration range from 1–5 mg U-22324/ml. Peak titers were usually obtained after 4–5 days' incubation at 25°C and amounted to 8–10 mg/ml. This represents an unusually

high yield for a single peptide to be synthesized by a microorganism.

Antibiotic U-22324 was isolated from a 1600 ml aliquot of filtered fermentation broth by precipitation of the active material at pH 4.6. The precipitate was collected with the aid of 1% (w/v) of Celite and the filter cake extracted with 400 ml of acetone. An oil, recovered by removal of the acetone at 30°C, crystallized upon refrigeration. This crude material (2.7 g) was purified by dissolving it in a minimum amount of absolute ethanol at room temperature, treatment with 200 mg of charcoal (Darco G60), concentration to 25 ml, and the slow addition of

- ¹ L. J. HANKA, D. J. MASON, and W. T. SOKOLSKI, *Antibiotics Chemother.* 11, 123 (1961).

25 ml diethyl ether to incipient crystallization. After filtration and drying the white crystals weighed 1.7 g.

The antibiotic exhibits a single pKa' value at 5.5, titrating as an acid, which, together with the fact that it does not react with ninhydrin, indicates it is a cyclic peptide. It exhibits no absorption above 220 nm and contains no sulfur.

Analysis of an hydrolysate of the antibiotic in a Beckman Spinco automatic amino acid analyzer² showed it to have the following composition: (Glu)₃(Pro)₂(Gly)₁(Ala)₂(Val)₂(Leu)₁ and an unknown amino acid which emerged at the position ordinarily occupied by cysteine. Amide nitrogen analysis indicated that 2 of the glutamic acid residues were present in their amide form. Sufficient material (2.5 g) was then hydrolyzed to permit isolation of the individual amino acids in larger quantities. The procedure of HIRS et al.³ was used for this purpose. Each

of the amino acids was obtained in crystalline form and in approximately 70% yield based upon the total quantitative ninhydrin method used to locate the various fractions⁴.

The unknown constituent amino acid was identified by IR- and NMR-spectroscopy as 2-methylalanine (or α -amino isobutyric acid). 8M are present per M of polypeptide. To our knowledge this amino acid has been isolated only once before⁵, and from an antibiotic. Positive identification of the other amino acids was obtained by comparison of their IR-spectra with authentic samples. The optical activities of the amino acids were determined and all the active ones proved to have the L-configuration. A cyclic polypeptide with the composition (GluN)₂(Glu)₁(Pro)₂(Gly)₁(Ala)₂(Dimethyl ala)₈(Val)₂(Leu)₁ has the theoretical elementary composition for C₈₁H₁₃₅N₂₁O₂₃ of C, 54.90; H, 7.68; N, 16.6. Found: C, 54.89; H, 7.74; N, 15.7. While the observed nitrogen value is more in accord with N₂₀, which would indicate a single glutamine residue, direct amide nitrogen analysis yielded 1.43%; theory for 2 glutamine residues, 1.58%.

The in vitro antibacterial spectrum of antibiotic U-22324 (Table I) indicates that the compound inhibits G⁺ bacteria exclusively when tested in the 2-fold broth dilution assay⁶.

The maximum tolerated dose of U-22324 in mice was 40 mg/kg/day when administered s.c. and 80 mg/kg/day orally. The antibiotic was inactive in mice experimentally infected with *S. hemolyticus* when administered s.c. at the maximum tolerated dose. No blood levels were demonstrated in mice dosed s.c. with 80 mg/kg as determined by the method of LEWIS et al.⁶.

Of 14 pathogenic fungi tested in vitro, the antibiotic inhibited *Blastomyces dermatitidis* at a concentration of 100 μ g/ml and *Nocardia asteroides*, *Hormodendrum compactum*, *Histoplasma capsulatum*, *Trichophyton mentagrophytes* UC-4800 and *Coccidioides immitis* at concentrations of 1000 μ g/ml as shown in Table II.

Antibiotic U-22324 decreases the surface tension of water extensively and might, therefore, have some utility as a detergent. A solution containing 100 μ g/ml of the antibiotic in deionized water had a surface tension of 41.6 dynes/cm at 25°C as compared to 71.8 dynes/cm for deionized water and 52.7 dynes/cm for a 100 μ g/ml solution of sodium lauryl sulfate (Duponol) respectively⁷.

Zusammenfassung. Antibiotikum U-22324 ist ein zyklisches Peptid, das aus Kulturflüssigkeit von *Trichoderma viride* isoliert wurde. Das Peptid hat die Aminosäurezusammensetzung (GluN)₂(Glu)₁(Pro)₂(Gly)₁(Ala)₂(Dimethyl ala)₈(Val)₂(Leu)₁.

C. E. MEYER and F. REUSSER

Department of Microbiology, The Upjohn Company, Kalamazoo (Michigan, USA), September 12, 1966.

Table I. In vitro antibacterial activity of antibiotic U-22324 in brain heart infusion broth

Test organism	Minimal inhibitory concentration μ g/ml
<i>E. coli</i> ATCC-26	> 1000
<i>K. pneumoniae</i> PCI-602	> 1000
<i>P. vulgaris</i> ATCC-8427	> 1000
<i>Ps. aeruginosa</i> ATCC-9027	> 1000
<i>S. paratyphi</i> UC-263	> 1000
<i>S. pullorum</i> MSDH-75	> 1000
<i>S. typhosa</i> UC-215	> 1000
<i>S. aureus</i> UC-76	62.5
<i>S. aureus</i> UC-70	125
<i>S. faecalis</i> ATCC-6057	31
<i>S. hemolyticus</i> C-203	31
<i>S. viridans</i> UC-155	31

Table II. In vitro antifungal activity of antibiotic U-22324 in Agar dilution test

Test organisms	Minimal inhibitory concentration μ g/ml
<i>Nocardia asteroides</i>	1000
<i>Blastomyces dermatitidis</i>	100
<i>Coccidioides immitis</i>	1000
<i>Geotrichum sp.</i>	> 1000
<i>Hormodendrum compactum</i>	1000
<i>Cryptococcus neoformans</i>	> 1000
<i>Histoplasma capsulatum</i>	1000
<i>Sporotrichum schenckii</i>	> 1000
<i>Monosporium apiospermum</i>	> 1000
<i>Trichophyton rubrum</i>	> 1000
<i>Trichophyton interdigitale</i>	> 1000
<i>Candida albicans</i>	> 1000
<i>Trichophyton violaceum</i>	> 1000
<i>Trichophyton asteroides</i>	> 1000
<i>Trichophyton mentagrophytes</i> UC-4797	> 1000
<i>Trichophyton mentagrophytes</i> UC-4860	1000

The antifungal tests were done on agar plates. Different concentrations (1000, 100, 10 or 1 μ g/ml) of the antibiotic were dissolved in the agar and the plates were inoculated by the cross-streak technique. Results are expressed as the minimal inhibitory concentration of antibiotic yielding total inhibition of fungal growth. Endomycin inhibits these organisms at 1 or 10 μ g/ml.

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³ C. H. W. HIRS, S. MOORE, and W. H. STEIN, *J. Am. chem. Soc.* **76**, 6063 (1954).

⁴ S. MOORE and W. H. STEIN, *J. biol. Chem.* **211**, 907 (1954).

⁵ G. W. KENNER and R. C. SHEPPARD, *Nature* **181**, 48 (1958).

⁶ C. LEWIS, H. W. CLAPP, and J. E. GRADY, *Antimicrobial Agents and Chemotherapy* (Ed.: J. C. Sylvester, 1962), p. 570.

⁷ The performance of the surface tension measurements by R. C. ANDERSON, the antifungal tests by A. DIETZ, analyses on the automatic amino acid analyzer by S. H. EPPSTEIN and the in vivo studies by C. LEWIS are gratefully acknowledged.