# Effects of the prostaglandin analogue misoprostol on inflammatory mediator release by human monocytes

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## Abstract

The effect of misoprostol (M) on IL-1 $\beta$ , TNF- $\alpha$ , and lipid mediator release (assessed by RIA) by adherent (assessed by electron microscopy) human monocytes were studied *in vitro*. Human monocytes stimulated with E. Coli-derived lipopolysaccharide showed an increase in both IL-1 $\beta$  and TNF- $\alpha$  release. Incubation of the monocytes with LPS and M (18 hrs.), resulted in a reduction of both IL-1 $\beta$  and TNF- $\alpha$  levels. Leukotriene B<sub>4</sub> levels did not increase in response to LPS or M. LPS also caused an increase in thromboxane (TXB<sub>2</sub>). *M* decreased TXB<sub>2</sub> levels. 6-keto PGF1 $\alpha$  (6KP). Incubation with LPS and M stimulated release. LPS caused an increase in PGE<sub>2</sub> levels. M (100  $\mu$ M) caused an increase in PGE<sub>2</sub> levels, M (1  $\mu$ M) had no effect on PGE<sub>2</sub>. These data suggest a possible immunomodulatory role for misoprostol in inflammatory diseases.

#### Introduction

Monocytes produce a variety of proinflammatory mediators in response to LPS [1]. These mediators, such as IL-1 $\beta$  and tumor necrosis factor- $\alpha$ , may be involved in inflammatory bowel disease (ulcerative colitis and Crohn's disease). Inhibition of these mediators could be of therapeutic value in IBD. Misoprostol [( $\pm$ )-methyl 11a, 16-hydroxy-16methyl-9-oxoprost-13E-1-oate] is a PGE<sub>1</sub> analogue reported to have immunomodulatory activity. Therefore, we studied its effects on the production of inflammatory mediators by human monocytes.

#### Materials and methods

A mixed population of human monocytes and lymphocytes was obtained by a discontinuous

metrizamide gradient method [2]. Cells were pooled and resuspended in an  $\alpha$ -modified Eagle's medium (a-MEM, Gibco) supplemented with 20 mM 3-(N-morpholino) propanesulfonic acid, 13.3 mM NaHCO<sub>3</sub>, 2 mM glutamine, 50 µM 2mercaptoethanol, 70 µg of streptomycin sulfate per ml and 1% fetal calf serum. Sterile plastic 16 mm wells were preincubated with media for 30 minutes at  $37^{\circ}$ C in a 5% CO<sub>2</sub> incubator.  $2.5 \times 10^6$  cells were added in a volume of one ml to each well and incubated for one hour at 37 °C in a CO<sub>2</sub> incubator. Three washes with cold phosphate buffered saline removed loosely adherent cells. One ml of media was added to each well. Lipopolysaccharide (LPS) from Escherichia coli 0111: B4 (Sigma) was added to a final concentration of 100 ng/ml together with the appropriate concentration of misoprostol. The cells (90% pure) were then incubated for 18 hours at 37°C in a CO<sub>2</sub> incubator. After 18 hours the supernatant

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Table 1	
Effect of LPS and Misoprostol (Miso) on inflammatory mediator release by human monocytes.	

Mediator	n	Control	LPS 100 ng/ml	MISO 1 µM	MISO 100 μ <i>M</i>
IL-1 $\beta$ (fmol)	16-21	$1.35 \pm 0.62$	3.94+0.82***	3.19+0.48**	1.26+0.24***
TXB, (ng/ml)	16-24	$1.146 \pm 0.18$	$3.55 \pm 0.86 ***$	$1.35 \pm 0.125 ***$	$0.135 \pm 0.134 ***$
6KP (ng/ml)	16-24	$0.035 \pm 0.005$	$0.139 \pm 0.01 ***$	$0.172 \pm 0.01 *$	$0.277 \pm 0.01 ***$
$TNF\alpha$ (pg/ml)	16-24	$161.3 \pm 20$	$340 \pm 52.4 *$	131.5 + 17.2*	120.3 + 12.8 ***
$PGE_2$ (µg/ml)	16-23	$0.24 \pm 0.145$	$8.58 \pm 4.11$	$5.885 \pm 1.83$	$1669.5 \pm 268.5 ***$

Values are means  $\pm$ SE \* p < 0.05, \*\* p < 0.01, \*\*\* p < 0.01.

were removed, centrifuged  $(170 \times g \text{ for } 15 \text{ min})$ and stored at -20°C until assayed [3]. Interleukin- $1\beta$  and tumor necrosis factor- $\alpha$  levels were determined by validated radioimmunoassays (Amersham & Genzyme, respectively). 6KP, PGE<sub>2</sub>, LTB<sub>4</sub> and TXB<sub>2</sub> levels were also determined by validated radioimmunoassays (NEN). Statistical assessment was by analysis of variance for each comparison, with a factor for treatment and a factor for donor. P values less than 0.05 were considered significant.

### Results

Incubation of human monocytes with LPS caused a significant increase in IL-1 $\beta$  (p<0.001), TXB<sub>2</sub> (p < 0.001), and 6KP (p < 0.001) levels (Table 1). Misoprostol  $(1 \mu M)$  reduced IL-1 $\beta$  and TXB<sub>2</sub> (p < 0.001) production. Misoprostol  $(1 \mu M)$ caused a significant increase (p < 0.05) in 6KP release. At a higher concentration (100  $\mu M$ ), misoprostol caused a significant decrease in IL-1 $\beta$ (p < 0.001) and TXB<sub>2</sub> (p < 0.001) release, but 6KP levels increased significantly (p < 0.001). These effects of misoprostol were concentration related. LPS significantly (p < 0.05) increased TNF- $\alpha$  release. Both  $1 \mu M$  and  $100 \mu M$  of misoprostol caused a significant (p < 0.05 and p < 0.001 respectively) reduction in TNF- $\alpha$  release. PGE<sub>2</sub> levels were increased by LPS, but not by misoprostol  $(1 \ \mu M)$ . Misoprostol  $(100 \ \mu M)$ significantly (p < 0.01) stimulated PGE<sub>2</sub> production. LTB<sub>4</sub> levels were unaffected by LPS as all values were below sensitivity (0.625 ng/ml) of the RIA (data not shown).

# Discussion

Misoprostol inhibited release of IL-1 $\beta$  from human monocytes in vitro, in contrast to previously re-

ported data using naturally occurring prostaglandins [4]. TNF- $\alpha$  and thromboxane release were also inhibited by incubation with misoprostol. This prostaglandin also stimulated 6KP, which is indicative of enhanced prostacyclin activity. Misoprostol also dramatically increased PGE<sub>2</sub> levels. PGE's are believed to have potent anti-inflammatory properties [5] and have been shown to reduce lymphokine production in activated macrophages [6]. Furthermore, misoprostol has improved renal function in cyclosporine-treated renal transplant recipients and reduced the frequency of acute transplant rejection [7]. These results suggest that misoprostol may have a potential beneficial immunomodulatory role in inflammatory diseases.

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