

Long-term adjuvant tamoxifen therapy for breast cancer

V. Craig Jordan

Department of Human Oncology, University of Wisconsin Clinical Cancer Center, Madison, Wisconsin 53792, USA

Key words: antiestrogen, tamoxifen metabolites, MCF-7 cells, adjuvant therapy, node-positive breast cancer

Abstract

Tamoxifen (ICI46,474) is a competitive inhibitor of estrogen action which has found ubiquitous application in the treatment of breast cancer. The drug is the front line endocrine therapy for breast cancer and is the proven treatment of choice for the adjuvant therapy of postmenopausal women with node-positive disease. Tamoxifen is available for the treatment of premenopausal patients with advanced disease, and is being evaluated in clinical trials as an adjuvant therapy for premenopausal patients with either node-positive or node-negative disease. Laboratory studies demonstrate that tamoxifen is a tumorstatic agent and long-term treatment strategies (chemosuppression) should be considered to apply the antiestrogen to its maximal therapeutic advantage. Optimal therapy with tamoxifen may also be achieved by treatment strategies to lower circulating estrogen levels in the premenopausal patient.

Tamoxifen is a well tolerated drug, and long-term therapy does not appear to induce metabolic tolerance. Concerns about premature osteoporosis or cardiovascular disease appear to be unfounded because tamoxifen has an appropriate level of target site-directed estrogenic activity. Isolated reports about the growth or appearance of endometrial carcinoma during long-term adjuvant tamoxifen therapy must be balanced against the risks of withholding treatment to patients with a fatal disease.

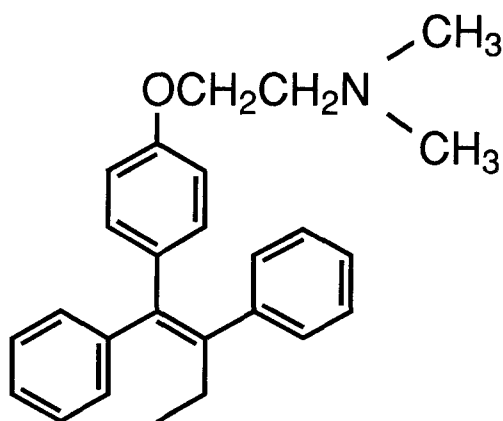
Introduction

Tamoxifen (ICI46,474, Fig. 1), a non-steroidal antiestrogen [1], was originally introduced for the treatment of advanced breast cancer in postmenopausal women [2–4]. However, the demonstrated efficacy and low incidence of side effects [5] has permitted the development of tamoxifen for the effective adjuvant treatment of node-positive breast cancer [6–8]. The drug is now considered to be the front line endocrine treatment for breast cancer.

The widespread acceptance of tamoxifen by the

medical community has naturally lead to the investigation of additional treatment strategies. Increasing numbers of premenopausal women with advanced breast cancer are receiving effective palliation with tamoxifen [9–11], and clinical trials are recruiting premenopausal women with both node-negative and node-positive disease [12, 13] to evaluate efficacy.

The aim of this article is to assess the progress being made to evaluate the strategy of long-term adjuvant tamoxifen therapy to suppress the recurrence of the disease following mastectomy. All the original clinical trials evaluated 1 or 2 years of



TAMOXIFEN (ICI 46,474)

Fig. 1. The structure of tamoxifen.

tamoxifen. Although this approach has merit [14], the laboratory data [15] points to the use of tamoxifen as a long-term chemosuppressive agent. The therapeutic value of this clinical approach will be balanced against perceived long-term side effects.

Mode of action of tamoxifen *in vitro*

Tamoxifen inhibits the binding of [³H]estradiol to estrogen receptors derived from human breast tumors [16]. Tamoxifen is a competitive inhibitor of estrogen binding *in vitro* [17], and increasing concentrations of estradiol will reverse the inhibitory action of tamoxifen on the growth of hormone-responsive MCF-7 breast cancer cells [18]. Nevertheless, large concentrations of antiestrogen will produce estrogen-irreversible effects on breast cancer cell growth *in vitro* [19, 20]. It remains speculative that estrogen receptor independent actions play a significant role in the antitumor actions of tamoxifen in patients.

Estrogen-stimulated growth has been the subject of intense investigation during the past 20 years. Progress has been facilitated by the development of estrogen-responsive breast cancer cell lines [21–23] and suitable culture conditions to study estrogen

action [24]. There is still considerable debate about the direct or indirect effects of estradiol. However, the concepts of estrogen-regulated growth are evolving with the application of autocrine and paracrine growth factor models [25] as a hypothesis to explain the ability of estrogen to shorten the breast cancer cell cycle.

Estrogen (Fig. 2) appears to be able to initiate a cascade of subcellular events, e.g. induction of progesterone receptor [26], increase in transforming growth factor α (TGF α) mRNA [27], and decrease in transforming growth factor β (TGF β) [28]. Antiestrogens like tamoxifen can block these actions (Fig. 3). In fact, it has been suggested that TGF β could act as a paracrine inhibitory factor on hormone independent breast cancer cells [28]. Estrogen receptor negative cells, unlike receptor positive cells, are particularly sensitive to the inhibitory actions of TGF β [29]. Therefore, in a breast tumor that is predominantly estrogen receptor positive, it could be possible that TGF β produced in the receptor positive cells will control the growth of estrogen receptor negative clones. Unfortunately this hypothesis has proved to be difficult to test successfully *in vivo* [30].

Mode of action of tamoxifen in animal models

Tamoxifen was initially evaluated for a whole range of clinical indications [31–33]. However, the efficacy in the treatment of breast cancer coupled with the parallel studies in animal models of breast cancer has established the drug as the 'gold standard' for endocrine therapy [34]. Two experimental systems, the dimethylbenzanthracene (DMBA)-induced rat mammary carcinoma model and the immune deficient (athymic) mouse implanted with breast cancer cell lines, have provided valuable information to support the use of long-term adjuvant tamoxifen therapy as a chemosuppressive strategy in breast cancer.

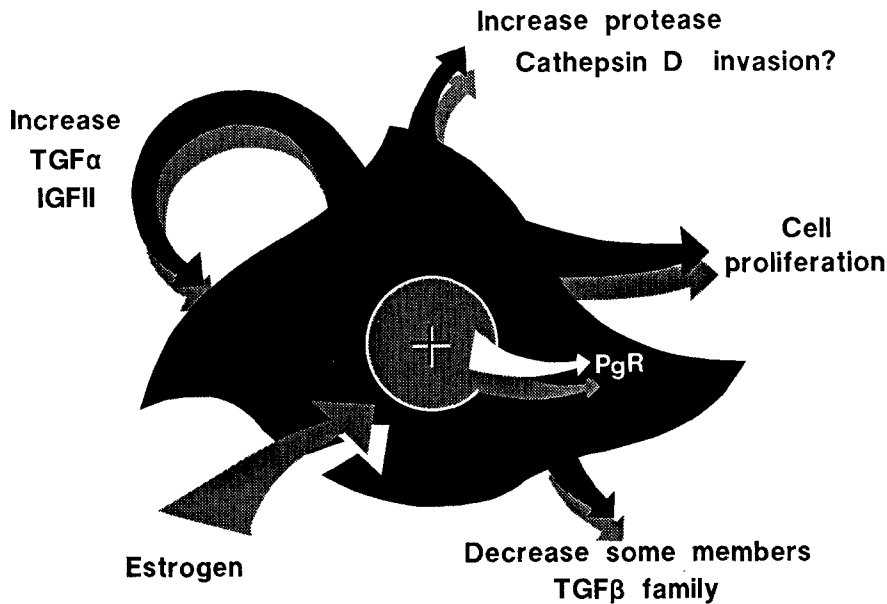


Fig. 2. The subcellular effects of estrogen in causing the replication of breast cancer cells (usually MCF-7) *in vitro*. Progesterone receptor (PgR), transforming growth factor α (TGF α), insulin-like growth factor II (IGFII), transforming growth factor β (TGF β).

Carcinogen-induced rat mammary carcinoma models

Mammary tumors can be induced by a single feeding (20 mg in 2 ml peanut oil) of DMBA to 50 day old female Sprague Dawley rats [35]. Palpable hormone responsive tumors appear about 3–6 months later. Tamoxifen inhibits the initiation [36, 37] and growth of established DMBA-induced tumors [38–42]. However, the value of this test system has been to determine whether short term tamoxifen therapy after the DMBA would produce a tumoricidal effect and cure the animals. The administration of tamoxifen at various doses or at various times after the carcinogenic insult [43, 44] only causes a delay in tumor appearance and, ultimately, the majority of animals have mammary tumors. The administration of continuous daily tamoxifen therapy after DMBA causes a complete suppression of tumorigenesis [45–47], but when treatment with tamoxifen is stopped tumor growth recurs [48].

These principles have been confirmed in the N-nitrosomethyl urea (NMU) model of rat mammary carcinogenesis [49]. A short course of tamoxifen therapy after NMU delays the appearance of mam-

mary tumors [50] but continuous therapy suppresses tumorigenesis [52]. Again, tumors appear if tamoxifen treatment is stopped [51].

Athymic animal models of hormone dependent breast cancer

Primary breast cancers can be transplanted into athymic (immune deficient) mice [51], but it has proved to be difficult to develop a reproducible model to study hormone-dependent primary breast cancer. Hormone-responsive breast cancer cell lines, which in the main have been established in culture from pleural effusions, can be grown into solid tumors in estrogen-supplemented ovariectomized athymic mice [53, 54]. The hormone-responsive tumors do not grow without estrogen treatment, while hormone-independent breast cancer cell lines do not require estrogen for growth. Tamoxifen and its metabolites will inhibit the estrogen-stimulated growth of MCF-7 tumors [55, 56], but once tamoxifen treatment is stopped tumor growth continues in the face of unopposed estrogen stimulation [55, 57].

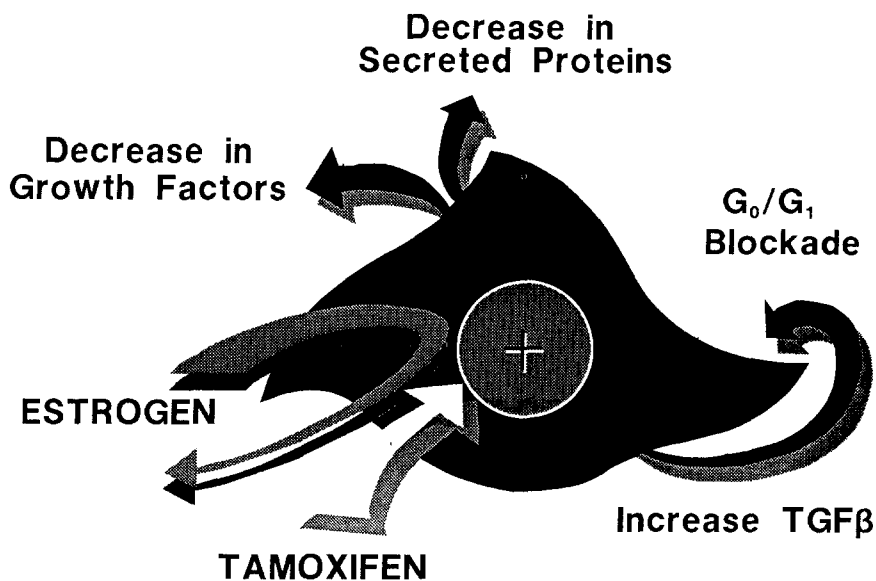


Fig. 3. The competitive action of tamoxifen to inhibit the binding of estrogen to nuclear estrogen receptors (+). The key to the figure is the same as in Figure 2.

The question can be posed whether the action of tamoxifen can be reversed with increasing levels of circulating estrogen. Indeed, these studies have just been completed in our laboratory, and the results demonstrate the competitive nature of the relationship of tamoxifen and estradiol *in vivo* [Fig. 4, 58]. Tamoxifen can inhibit estradiol-stimulated growth; however, if the level of circulating estrogen is increased 4–5 fold, the effect of tamoxifen as an antitumor agent is reversed.

Alternatively, the question can be asked whether, in the absence of circulating estradiol, tamoxifen can destroy implanted MCF-7 cells. Tamoxifen treatment of animals for up to six months does not result in the demonstration of a tumoricidal action of the drug. Estrogen can re-activate tumor growth after 6 months of tamoxifen therapy [59]. In fact, tumor growth may resume despite continuous tamoxifen therapy [60, 61]. Repeated transplantation of the growing tumors into tamoxifen-treated mice results in a tumor variant (MCF-7 TAM) that is facilitated to growth by tamoxifen. Growth is, however, reduced when tamoxifen treatment is stopped [62]. The tumors have estrogen receptors, and the tamoxifen-facilitated growth may be caused by its weak estrogen-like qualities [63].

Pure antiestrogens such as ICI164,384 [64] will inhibit tamoxifen-stimulated MCF-7 tumor growth [62].

Tamoxifen-stimulated tumor growth has not been described routinely in the clinics, but it is possible that this may occur after many years of adjuvant therapy. An interesting feature of the animal model is the finding that the MCF-7 tumor will also grow with estrogen treatment [62]. Therefore, if this form of tamoxifen drug resistance should occur in the clinics, it will not be sufficient to withdraw tamoxifen alone; the patient's own estrogen could support tumor growth. Nonestrogenic antiestrogens are currently being tested for clinical application. These new drugs may be of value as second line therapies [62, 64]. Indeed, the principle may already have been demonstrated with the finding that patients with advanced disease who respond, but then fail tamoxifen therapy, can respond to aminoglutethimide therapy [65]. The reverse is uncommon. If it is possible for a tumor to be dependent on tamoxifen for growth then a withdrawal response should be observed, similar to that documented for high-dose estrogen therapy [66]. The fact that this is rarely observed is probably a

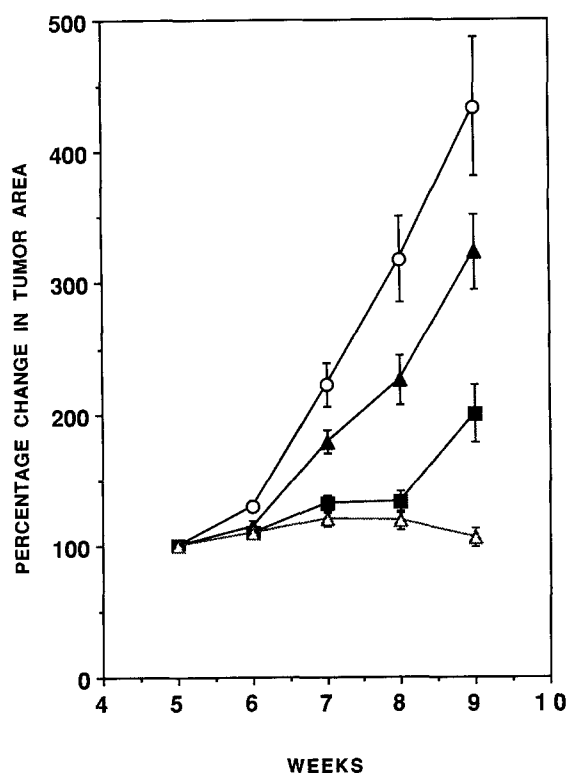


Fig. 4. The reversal of the action of tamoxifen on estradiol-stimulated MCF-7 tumor growth in athymic mice. Breast tumors were grown to approx. 0.5 cm² with a 1 cm silastic implant of estradiol before tamoxifen (2 cm silastic implant producing tamoxifen levels of 50 ng/ml) was implanted. Tamoxifen inhibits the growth of tumors. However, the implantation of delivery systems that produce higher circulating levels of estradiol causes increased tumor growth. The groups are: △ tamoxifen alone (no E₂), 9 ± 4 pg/ml; ○, E₂ alone, 755 ± 152 pg/ml; ■ tamoxifen + E₂, 543 ± 42 pg/ml; ▲ tamoxifen + E₂, 1950 ± 558 pg/ml.

reflection of the prolonged biological half life of tamoxifen.

Clinical pharmacology

Tamoxifen is readily absorbed following oral administration. The recommended daily dose (depending upon the country) is 10 or 20 mg bid or 10 mg tid. However, the long plasma half-life (7 days) at steady state (achieved after 30 days of administration) probably makes it unnecessary to use divided daily doses.

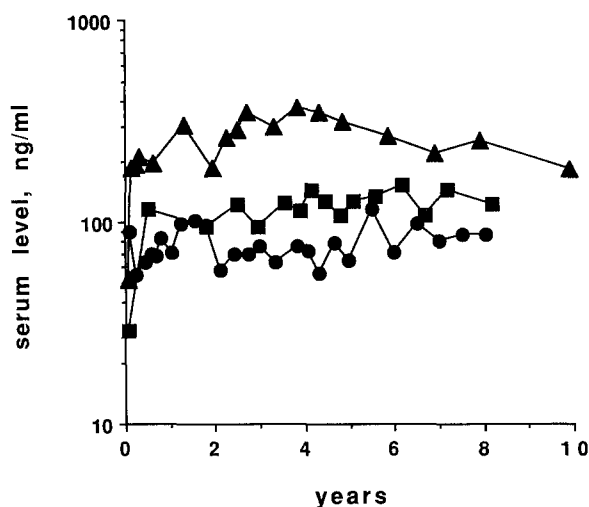


Fig. 5. The serum tamoxifen levels in patients receiving 10 mg bid for the long-term adjuvant treatment of node-positive breast cancer. Three patients (■, ▲, ●) have been treated for up to 10 years with tamoxifen.

The range of tamoxifen serum levels is extremely large (50–300 ng/ml), and this variability is important if patient compliance becomes an issue during long-term adjuvant tamoxifen therapy. A single determination from a patient can only really provide information about whether the patient is taking the drug or not. Several determinations must be made over a period of months to establish the steady state level for the individual.

In conjunction with Dr. Douglass Tormey we have monitored the serum levels of patients receiving adjuvant tamoxifen therapy (10 mg bid) to prevent the recurrence of node-positive breast cancer. Serum levels are stable for the ten years of therapy (Fig. 5) and there are no unusual estrogenic metabolites. We have monitored 10 patients for up to 10 years and we cannot detect any evidence of metabolic tolerance.

Tamoxifen [67] is extensively metabolized in patients (Fig. 6). The principal metabolite is N-desmethyltamoxifen [68] which may be further converted to Metabolite Y [69]. A minor metabolite is 4-hydroxytamoxifen; however, this compound has a binding affinity for the estrogen receptor 20–30x that of tamoxifen [70]. Figure 7 shows the relative levels of various metabolites during the long-term adjuvant treatment with tamoxifen for node-posi-

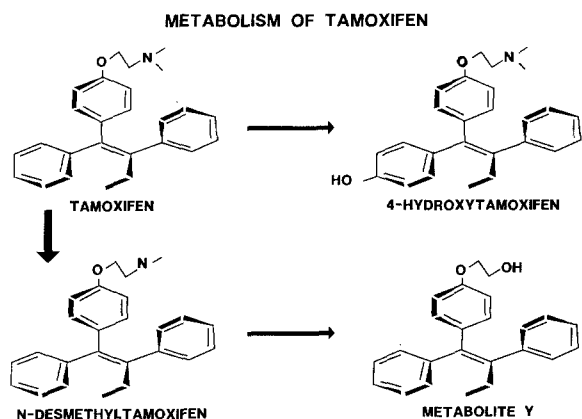


Fig. 6. The metabolism of tamoxifen in patients.

tive breast cancer. Each of these metabolites exhibits antiestrogenic properties which probably contribute to the overall efficacy of tamoxifen as an antitumor agent [71, 72].

Long-term adjuvant tamoxifen therapy

The majority of the clinical trials that were established to test the efficacy of tamoxifen therapy chose to use one or two years of treatment. This strategy was the result of several factors: 1) the short period of therapy might produce a tumoricidal effect before the onset of drug resistance; 2) there was a general belief that extended tamoxifen therapy would only encourage the outgrowth of aggressive receptor-negative disease; and 3) there were uncertainties about the long-term toxicological effects of tamoxifen.

The recent overview of randomized clinical trials has established that tamoxifen therapy for up to two years can result in an increase in the survival of node-positive postmenopausal patients [14]. The situation with regard to premenopausal women is less encouraging, as tamoxifen does not seem to provide benefit over and above combination chemotherapy (most trials use tamoxifen with chemotherapy in premenopausal patients).

Swayed by the encouraging clinical trials data and the laboratory studies [15], most clinical trials organizations are focused upon an evaluation of 5 or 10 years of adjuvant tamoxifen therapy for node-

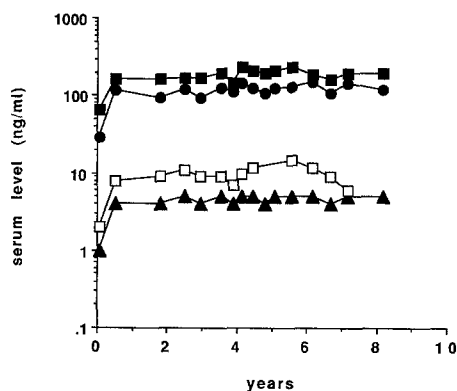


Fig. 7. The steady state serum levels of tamoxifen and its metabolites during the long-term adjuvant treatment of node-positive breast cancer. \blacktriangle 4-hydroxytamoxifen, \square Metabolite Y, \bullet tamoxifen, \blacksquare N-desmethyl-tamoxifen.

positive (and in some cases, node-negative) disease.

In 1977, based upon the encouraging laboratory data [43, 45], Dr. Douglass Tormey organized a pilot (nonrandomized) clinical study to evaluate the toxicology and efficacy of long-term tamoxifen in node-positive breast cancer patients. Three groups of women were selected (pre- and postmenopausal women with no bias towards receptor status): chemotherapy, chemotherapy plus tamoxifen, or chemotherapy and then tamoxifen alone up to five years [73]. The results demonstrated the safety of tamoxifen and provided the preliminary data to establish the ECOG trials EST 4181 and EST 5181 to evaluate the efficacy of long-term adjuvant tamoxifen therapy in randomized clinical trials [74].

The NSABP has compared 5-FU, L-PAM, and tamoxifen as an adjuvant for two years with a registration arm of the same treatment but with an additional year of tamoxifen alone. The results are encouraging and support the view that long-term tamoxifen therapy benefits the patient [75].

Several clinical trials have evaluated the efficacy of tamoxifen alone. Delozier and coworkers [76] compared three years of adjuvant tamoxifen therapy vs no treatment in a small randomized clinical study. The tamoxifen-treated patients who were receptor-positive had a survival advantage when

compared with either receptor-negative patients or those not receiving tamoxifen.

A much larger randomized clinical study has evaluated five years of adjuvant tamoxifen therapy against a control arm that received tamoxifen therapy upon first recurrence [77]. The Scottish trial has demonstrated a survival advantage for patients receiving adjuvant tamoxifen therapy, thereby dispelling the fears that early resistance to tamoxifen will occur during an extended treatment schedule. The patient population for the Scottish trial contains a significant number of premenopausal women and node-negative patients. The NSABP has taken the view that the logical direction for their clinical trials is to evaluate the efficacy of long-term adjuvant tamoxifen therapy in pre- and postmenopausal node-negative patients who had ER-positive primary tumors. The preliminary results of their investigation are encouraging [13], as tamoxifen produces an increase in disease-free survival. Nevertheless, the large potential patient population (up to half a million women could be taking tamoxifen forever, in the US alone, by the turn of the century), and the improved survival, requires an evaluation of the perceived risks of long-term tamoxifen therapy to be balanced against the benefits in delaying recurrence of a fatal disease.

Antiestrogenic side effects of tamoxifen

Estrogen is required for the maintenance of bone and to provide a beneficial lipid profile in postmenopausal patients. Osteoporosis and developing cardiovascular disease are serious complications of the advancing years following menopause. Estrogen replacement therapy has provided a clear benefit for the long-term health and wellbeing of women [78]. Obviously the long-term administration of an 'antiestrogen' would be expected to cause premature osteoporosis, atherosclerosis, and an increase in myocardial infarction.

Tamoxifen is an antiestrogen [6] but there appears to be a target site specificity to its action. Studies in laboratory animals have demonstrated that tamoxifen has an estrogen-like effect upon bone [79, 80], and in fact tamoxifen can inhibit

estrogen-stimulated increases in ovariectomized rat uterine wet weight but produce an additive estrogenic effect on bone density [81]. These encouraging laboratory results suggest that tamoxifen may in fact have a beneficial effect on patient bone density. This appears to be true from the results of preliminary clinical studies [82, 83].

Similarly, tamoxifen appears to produce an estrogen-like effect upon the lipid profile in postmenopausal women [84]. The LDL cholesterol fraction has a significant decrease during tamoxifen adjuvant therapy [85, 86]. Whether these positive effects produced by tamoxifen in women will be identified as the overall cause of the apparent increased survival of patients (regardless of survival from breast cancer) remains to be established [87].

Estrogenic side effects of tamoxifen

The estrogen-like actions of tamoxifen are well documented in postmenopausal patients. There is a partial decrease in circulating gonadotrophins [88], an increase in sex hormone binding globulin [89], an estrogen-like change in circulating proteins [90], and an estrogen-like effect on vaginal cytology [91].

There are, however, two principal concerns, liver carcinogenesis and endometrial carcinoma, which may involve the estrogenic component of tamoxifen's action. Animal studies have adequately documented the role of estrogen as a promoter of liver carcinogenesis. The estrogens in oral contraceptives are extremely potent promoters [92]. In contrast, only very large (200 mg/kg) daily doses of tamoxifen can promote liver carcinogenesis in rats [93]. To date, no evidence of an increase in tamoxifen-induced hepatomas has emerged from randomized clinical trials.

Unopposed estrogen administration to postmenopausal women causes an increase in endometrial carcinoma [94]. At present, there is really no information concerning the long-term effects of tamoxifen therapy on the human endometrium. Nevertheless, tamoxifen has been used successfully to treat endometrial carcinoma [95], so that its action is unlikely to be as a full estrogen.

The concern about the estrogen-like actions of tamoxifen to support the growth of occult endometrial cancer comes from the observation that a human, hormone-responsive endometrial carcinoma will grow more rapidly in athymic mice during tamoxifen treatment [96]. A number of anecdotal reports [97–99] have linked tamoxifen with an increase in endometrial carcinoma. However, this is only to be expected, as tamoxifen could only inhibit the growth of about $\frac{1}{3}$ of the tumors that might occur in patients with breast cancer. To date, only one randomized clinical trial has reported an increase in endometrial carcinoma during long-term tamoxifen therapy [100]. No increase in endometrial carcinoma has been found in the Scottish trial [101] or been reported by the ECOG or NSABP clinical trials organizations in the United States.

The Swedish study [100] is particularly interesting because tamoxifen causes a significant decrease in the numbers of second primary breast cancers. This again illustrates the target site specificity of the drug observed in animals; tamoxifen causes human endometrial carcinomas to grow but prevents the growth of bitransplanted breast tumors (MCF-7) in the athymic mouse [102]. Overall, it should be obvious that the benefits of tamoxifen to inhibit the recurrence (or prevent the appearance of primary breast tumors) of a fatal disease outweigh the potential concerns about endometrial carcinoma, a curable disease, in node-positive breast cancer.

The application of tamoxifen to prevent the recurrence of node-negative disease should be monitored carefully by the physician, and suspicious bleeding and discharge should be investigated immediately.

Ovarian actions of tamoxifen

The trend towards treating large populations of premenopausal, node-negative women with long-term adjuvant tamoxifen therapy [13, 77] must take into account the known effects of tamoxifen upon ovarian steroidogenesis. Tamoxifen induces ovulation in premenopausal patients [32, 33], and there is a known effect of tamoxifen to increase ovarian

steroidogenesis [103–105]. Clearly, patients must be counseled to use barrier contraceptives, as tamoxifen therapy is not recommended during pregnancy. The increases in ovarian estrogen production could possibly reduce the efficacy of tamoxifen either if the patient is non-compliant or if the drug is rapidly cleared (reflected by low serum concentrations). However, the antitumor action of tamoxifen is not easily reversed in the athymic animal model [58]. Nevertheless, for optimal efficacy, a low-estrogen environment would probably be beneficial for tamoxifen. Physicians could consider ovariectomy or administration of sustained release preparations of synthetic luteinizing hormone-releasing hormone [106] to prevent rises in ovarian estrogen and to prevent the long-term effects of ovarian stimulation (which are as yet unknown).

Conclusions

Tamoxifen has proved to be the front line endocrine therapy for breast cancer. The drug has proven efficacy to improve survival in postmenopausal, node-positive patients (approx. 2 years of adjuvant therapy), and long-term treatment (5–10 years) schedules are being evaluated in clinical trial. Overall, the drug is remarkably nontoxic (compared with other cancer therapeutic agents), and very few side effects of significance have been noted that should deter the physician from treating node-positive postmenopausal disease. There is a growing clinical experience with premenopausal patients, especially those that are node-negative. Physicians must use their clinical judgement whether to offer their patients tamoxifen to prevent the recurrence of an invariably fatal disease or to withhold a relatively nontoxic therapy (because of possible long-term side effects) until recurrence.

Acknowledgements

I would like to thank Dr Y. Iino, Dr Douglass C. Tormey, and Susan Langan-Fahey for their contributions to these studies, and to thank the contributors to the Eileen Henrich memorial fund at the

UWCCC for their generosity in providing equipment for the measurement of tamoxifen in patients. The studies were supported by P30-CA 14520, P01-CA 20432, and ICI Americas.

References

1. Harper MJK, Walpole AL: A new derivative of triphenylethylene: Effect on implantation and mode of action in rats. *J Reprod Fertil* 13: 101-119, 1967
2. Cole MP, Jones CTA, Todd IDH: A new antioestrogenic agent in late breast cancer: An early clinical appraisal of ICI46,474. *Br J Cancer* 25: 270-275, 1971
3. Ward HWC: Antioestrogen therapy for breast cancer: A trial of tamoxifen at two dose levels. *Br Med J* i: 13-14, 1973
4. O'Halloran MJ, Maddock PG: ICI 46,474 in breast cancer. *J Irish Med Assoc* 67: 38-39, 1974
5. Legha SS, Carter SK: Antiestrogens in the treatment of breast cancer. *Cancer Treat Rev* 3: 205-216, 1976
6. Furr BJA, Jordan VC: The pharmacology and clinical uses of tamoxifen. *Pharmacol Ther* 25: 127-205, 1984
7. Legha SS: Tamoxifen in the treatment of breast cancer. *Ann Int Med* 109: 219-228, 1988
8. Love RR: Tamoxifen therapy in primary breast cancer: Biology, efficacy and side effects. *J Clin Oncol* 7: 803-815, 1989
9. Sawka CA, Pritchard KI, Paterson DJA, Thomson DB, Skelley WE, Myers RE, Mobbs BG, Malkin A, Meakin JW: Role and mechanism of action of tamoxifen in premenopausal women with metastatic breast cancer. *Cancer Res* 46: 3152-3156, 1986
10. Ingle JN, Krook JE, Green SJ, Kubista TP, Everson LK, Ahman DL, Chang MN, Bisel HF, Windschitl HE, Twito DI, Pfeifle DM: Randomized trial of bilateral oophorectomy versus tamoxifen in premenopausal women with metastatic breast cancer. *J Clin Oncol* 4: 4178-4185, 1986
11. Buchanan RB, Blamey RW, Durrent KR, Howell A, Paterson AG, Preece PE, Smith DC, Williams CJ, Wilson RG: A randomized comparison of tamoxifen with surgical oophorectomy in premenopausal patients with advanced breast cancer. *J Clin Oncol* 4: 1326-1330, 1986
12. Cancer Research Campaign Adjuvant Breast Trial Working Party: Cyclophosphamide and tamoxifen as adjuvant therapy in the management of breast cancer. *Br J Cancer* 57: 604-607, 1988
13. Fisher B, Constantino J, Redmond C, and other members of the NSABP: A randomized clinical trial evaluating tamoxifen in the treatment of patients with node-negative breast cancer who have estrogen receptor-positive tumors. *N Engl J Med* 320: 479-484, 1989
14. Early Breast Cancer Trialists' Collaborative Group: Effect of adjuvant tamoxifen and of cytotoxic therapy on mortality in early breast cancer. *N Engl J Med* 319: 1681-1692, 1988
15. Jordan VC: Long-term tamoxifen therapy for breast cancer. *In: DeVita VT, Hellman S, Rosenberg S (eds): Important Advances in Oncology.* Lippincott, Philadelphia, pp 179-192, 1989
16. Jordan VC, Koerner S: Tamoxifen (ICI46,474) and the human carcinoma 8S estrogen receptor. *Eur J Cancer* 11: 205-206, 1975
17. Skidmore JR, Walpole AL, Woodburn J: Effect of some triphenylethylenes on oestradiol binding *in vitro* to macromolecules from uterus and anterior pituitary. *J Endocrinol* 52: 289-298, 1972
18. Lippman ME, Bolan G: Oestrogen-responsive human breast cancer in long-term tissue culture. *Nature (Lond)* 256: 592-595, 1975
19. Reddel RR, Murphy LC, Sutherland RL: Effects of biologically active metabolites of tamoxifen on the proliferation kinetics of MCF-7 human breast cancer cells *in vitro*. *Cancer Res* 43: 4618-4624, 1983
20. Taylor CM, Blanchard B, Zava DT: Estrogen receptor mediated and cytotoxic effects of the antiestrogens tamoxifen and 4-hydroxytamoxifen. *Cancer Res* 44: 1409-1414, 1984
21. Soule HD, Vazquez J, Long A, Albert S, Brennan M: A human cell line from a pleural effusion derived from a breast carcinoma. *J Natl Cancer Inst* 51: 1409-1416, 1973
22. Engel LW, Young NA, Tralka TS, Lippman ME, O'Brien SJ, Joyce MJ: Establishment and characterization of three new continuous cell lines derived from human breast carcinomas. *Cancer Res* 38: 3352-3364, 1978
23. Keydar I, Chen L, Karby S, Weiss FR, Delarea J, Radu M, Chaitik S, Brenner HJ: Establishment and characterization of a cell line of human breast carcinoma origin. *Eur J Cancer* 15: 659-670, 1979
24. Berthois Y, Katzenellenbogen JA, Katzenellenbogen BS: Phenol red in tissue culture media is a weak estrogen: Implications concerning the study of estrogen-responsive cells in culture. *Proc Natl Acad Sci USA* 83: 2496-2500, 1986
25. Sporn MB, Todaro GJ: Autocrine secretion and malignant transformation of cells. *N Engl J Med* 303: 878-880, 1980
26. Horwitz KB, Koseki Y, McGuire WL: Estrogen control of progesterone receptor in human breast cancer: Role of estradiol and antiestrogen. *Endocrinology* 103: 1742-1751, 1978
27. Bates SE, Davidson NE, Vilverius EM, Freter CE, Dickson RB, Tam JP, Kudlow JE, Lippman ME, Saloman DS: Expression of transforming growth factor α and its messenger ribonucleic acid in human breast cancer: its regulation by estrogen and its possible functional significance. *Mol Endocrinol* 2: 543-555, 1988
28. Knabbe C, Lippman ME, Wakefield LM, Flanders KC, Kasid A, Derynck R, Dickson RB: Evidence that transforming growth factor- β is a hormonally regulated nega-

- tive growth factor in human breast cancer cells. *Cell* 48: 417–428, 1987
29. Arteaga CL, Tandon AK, Von Hoff DD, Osborne CK: Transforming growth factor- β : Potential autocrine growth inhibitor of estrogen receptor-negative human breast cancer cells. *Cancer Res* 48: 3898–3904, 1988
 30. Robinson SP, Jordan VC: Antiestrogenic action of toremifene on hormone dependent, independent and heterogeneous breast tumor growth in the athymic mouse. *Cancer Res* 49: 1758–1762, 1989
 31. El-Sheikha Z, Klopper A, Beck JS: Treatment of menometrorrhagia with an antioestrogen: *Clin Endocrinol* 1: 275–282, 1972
 32. Klopper A, Hall M: New synthetic agent for the induction of ovulation: Preliminary trial in women. *Brit Med J* i: 152–154, 1971
 33. Williamson JG, Ellis JD: The induction of ovulation by tamoxifen. *J Obstet Gynaecol Brit Comm* 80: 844–847, 1973
 34. Jordan VC: The development of tamoxifen for breast cancer therapy: A tribute to the late Arthur L. Walpole. *Breast Cancer Res Treat* 11: 197–209, 1988
 35. Huggins C, Grand LC, Brillantes P: Mammary cancer induced by a single feeding of polynuclear hydrocarbons and its suppression. *Nature (Lond)* 189: 204–207, 1961
 36. Jordan VC: Antitumor activity of the antioestrogen ICI46,474 (tamoxifen) in the dimethylbenzanthracene (DMBA)-induced rat mammary carcinoma model. *J Steroid Biochem* 5: 354, 1974
 37. Jordan VC: Effect of tamoxifen (ICI46,474) on initiation and growth of DMBA-induced rat mammary carcinomata. *Eur J Cancer* 12: 419–424, 1976
 38. Jordan VC: The antitumor effect of tamoxifen in the dimethylbenzanthracene-induced mammary carcinoma model. *Proc Symposium on the Hormonal Control of Breast Cancer*. Alderley Park, pp 11–17, ICI Pharmaceuticals Division PLC, Macclesfield, 1975
 39. Nicholson RI, Golder MP: The effect of synthetic antioestrogens on the growth and biochemistry of rat mammary tumours. *Eur J Cancer* 11: 571–579, 1975
 40. Jordan VC, Dowse LJ: Tamoxifen as an antitumour agent: Effect on oestrogen binding. *J Endocrinol* 68: 297–303, 1976
 41. Jordan VC, Koerner S: Tamoxifen as an antitumour agent: Role of oestradiol and prolactin. *J Endocrinol* 68: 305–310, 1976
 42. Jordan VC, Jaspan T: Tamoxifen as an antitumour agent: Oestrogen binding as a predictive test for tumour response. *J Endocrinol* 68: 453–460, 1976
 43. Jordan VC, Dix CJ, Allen KE: The effectiveness of long-term treatment in a laboratory model for adjuvant hormone therapy of breast cancer. *In*, Salmon SE, Jones SE (eds) *Adjuvant Therapy of Cancer II*. Grune and Stratton, New York, pp 19–26, 1979
 44. Jordan VC, Allen KE: Evaluation of the antitumour activity of the non-steroidal antioestrogen monohydroxytamoxifen in the DMBA-induced rat mammary carcinoma model. *Eur J Cancer* 16: 239–251, 1980
 45. Jordan VC: Use of the DMBA-induced rat mammary carcinoma system for the evaluation of tamoxifen treatment as a potential adjuvant therapy. *Rev Endocr Rel Cancer (Oct. Suppl)*: 49–55, 1978
 46. Jordan VC, Allen KE, Dix CJ: The pharmacology of tamoxifen in laboratory animals. *Cancer Treat Rep* 64: 745–759, 1980
 47. Robinson SP, Jordan VC: The reversal of the antitumor effect of tamoxifen by progesterone in the 7,12-dimethylbenzanthracene-induced rat mammary carcinoma model. *Cancer Res* 47: 5386–5390, 1987
 48. Robinson SP, Mauel DA, Jordan VC: Antitumor actions of toremifene in the 7,12-dimethylbenzanthracene (DMBA)-induced rat mammary tumor model. *Eur J Cancer Clin Oncol* 24: 1817–1821, 1988
 49. Gullino PM, Pettigrew HM, Grantham FH: N-Nitrosomethylurea as a mammary carcinogen in rats. *J Natl Cancer Inst* 54: 401–414, 1975
 50. Wilson AJ, Tehrani F, Baum M: Adjuvant tamoxifen therapy for early breast cancer: An experimental study with reference to oestrogen and progesterone receptors. *Br J Surg* 69: 121–125, 1982
 51. Gottardis MM, Jordan VC: The antitumor actions of keoxifene and tamoxifen in the N-nitrosomethylurea-induced rat mammary carcinoma model. *Cancer Res* 47: 4020–4024, 1987
 52. Giovanella BC, Stehlin JS, Williams LJ, Lee SS, Shepard RC: Heterotransplantation of human cancers into nude mice. *Cancer* 42: 2269–2281, 1978
 53. Soule HD, McGrath CM: Estrogen proliferation of clonal human breast carcinoma cells in athymic mice. *Cancer Lett* 10: 177–189, 1980
 54. Shafie SM, Grantham FH: Role of hormones in the growth and regression of human breast cancer cells (MCF-7) transplanted into athymic mice. *J Natl Cancer Inst* 67: 51–56, 1981
 55. Osborne CK, Hobbs K, Clark GM: Effect of estrogens and antiestrogens on growth of human breast cancer cells in athymic mice. *Cancer Res* 45: 584–590, 1985
 56. Gottardis MM, Robinson SP, Jordan VC: Estradiol-stimulated growth of MCF-7 tumors implanted in athymic mice: A model to study the tumoristic action of tamoxifen. *J Steroid Biochem* 30: 311–314, 1988
 57. Jordan VC, Gottardis MM, Robinson SP, Friedl A: Immune-deficient animals to study 'hormone-dependent' breast and endometrial cancer. *J Steroid Biochem*, 1989
 58. Jordan VC, Iino Y, Langan-Fabey S, Ricchio M, Wolf DM: Reversible inhibition of oestradiol-stimulated breast cancer growth by the non-steroidal antioestrogen tamoxifen. *Proc Brit Pharm Soc C127*, London, Jan 3–5, 1990
 59. Jordan VC: Chemosuppression of breast cancer with tamoxifen: Laboratory evidence and future clinical investigations. *Cancer Invest* 6: 5–11, 1988
 60. Osborne CK, Coronado EB, Robinson JP: Human breast

- cancer in the athymic nude mouse: Cytostatic effects of long-term antiestrogen therapy. *Eur J Cancer Clin Oncol* 23: 1189–1196, 1987
61. Gottardis MM, Jordan VC: Development of tamoxifen-stimulated growth of MCF-7 tumors in athymic mice after long-term antiestrogen administration. *Cancer Res* 48: 5183–5187, 1988
 62. Gottardis MM, Jiang SY, Jeng MH, Jordan VC: Inhibition of tamoxifen-stimulated growth of an MCF-7 tumor variant in athymic mice by novel steroidal antiestrogens. *Cancer Res* 49: 4090–4093, 1989
 63. Gottardis MM, Wagner RJ, Borden EC, Jordan VC: Differential ability of antiestrogens to stimulate breast cancer cell (MCF-7) growth *in vivo* and *in vitro*. *Cancer Res* 49: 4765–4769, 1989
 64. Wakeling AE, Bowler J: Steroidal pure antioestrogens. *J Endocrinol* 112: R7–R10, 1987
 65. Murray RML, Pitt P: Aminoglutethimide in tamoxifen-resistant patients: The Melbourne experience. *Cancer Res* 42 (Suppl): 3437s–3441s, 1982
 66. Stoll BA: Palliation by castration or by hormone administration. *In: Stoll BA (ed) Breast Cancer Management Early and Late*. W. Heinemann Medical Books Ltd, London, pp 133–146, 1977
 67. Jordan VC: Metabolites of tamoxifen in animals and man: Identification, pharmacology and significance. *Breast Cancer Res Treat* 2: 123–138, 1982
 68. Adam HK, Douglas EJ, Kemp JV: The metabolism of tamoxifen in humans. *Biochem Pharmacol* 27: 145–147, 1979
 69. Jordan VC, Bain RR, Brown RR, Gosden B, Santos MA: Determination and pharmacology of a new hydroxylated metabolite of tamoxifen observed in patient sera during therapy for advanced breast cancer. *Cancer Res* 43: 1446–1450, 1983
 70. Jordan VC, Collins MM, Rowsby L, Prestwich G: A monohydroxylated metabolite of tamoxifen with potent antioestrogenic activity. *J Endocrinol* 75: 305–316, 1977
 71. Jordan VC, Dix CJ, Naylor KE, Prestwich G, Rowsby L: Non-steroidal antioestrogens: Their biological effects and potential mechanisms of action. *J Toxicol Environ Health* 4: 364–390, 1978
 72. Coezy E, Borgna JL, Rochefort H: Tamoxifen and metabolites in MCF-7 cells and correlation between binding to estrogen receptor and inhibition of cell growth. *Cancer Res* 42: 317–323, 1982
 73. Tormey DC, Jordan VC: Long-term tamoxifen adjuvant therapy in node-positive breast cancer: A metabolic and pilot clinical study. *Breast Cancer Res Treat* 4: 297–302, 1984
 74. Falkson HC, Gray R, Wolberg WM: Adjuvant therapy of postmenopausal women with breast cancer – an ECOG Phase III study. Abstract 67, ASCO San Francisco, May, 1989
 75. Fisher B, and other NSABP Investigators: Prolonging tamoxifen for primary breast cancer: Findings from the National Surgical Adjuvant Breast and Bowel Project Clinical Trial. *Ann Int Med* 106: 649–654, 1987
 76. Delozier T, Julien JP, Juret P, Veyret C, Couette JE, Grai Y, Olliver JM, deRanieri E: Adjuvant tamoxifen in postmenopausal breast cancer: preliminary results of a randomized trial. *Breast Cancer Res Treat* 7: 105–110, 1986
 77. Breast Cancer Trials Committee, Scottish Cancer Trials Office (MRC): Adjuvant tamoxifen in the management of operable breast cancer: The Scottish trial. *Lancet* ii: 171–175, 1987
 78. Whitehead MI, Fraser D: Controversies concerning the safety of estrogen replacement therapy. *Am J Obstet Gynecol* 156: 1313–1322, 1987
 79. Turner RT, Wakeley GK, Hannon KS, Bell NA: Tamoxifen prevents the skeletal effects of ovarian hormone deficiency in rats. *J Bone Min Res* 2: 449–456, 1987
 80. Turner RT, Wakeley GK, Hannon KS, Bell NH: Tamoxifen inhibits osteoclast mediated resorption of trabecular bone in ovarian hormone deficient rats. *Endocrinology* 122: 1146–1150, 1988
 81. Jordan VC, Phelps E, Lingren JU: Effects of antiestrogens on bone in castrated and intact female rats. *Breast Cancer Res Treat* 10: 31–35, 1987
 82. Love RR, Mazess RB, Tormey DC, Barden HS, Newcomb PA, Jordan VC: Bone mineral density in women with breast cancer treated for at least two years with tamoxifen. *Breast Cancer Res Treat* 12: 297–301, 1988
 83. Turken S, Siris E, Seldin E, Seldin D, Flaster E, Hyman G, Lindsay R: Effects of tamoxifen on spinal bone density in women with breast cancer. *J Natl Cancer Inst* 81: 1086–1088, 1989
 84. Rossner S, Wallgren A: Serum lipoproteins and proteins after breast cancer surgery and effects of tamoxifen. *Atherosclerosis* 52: 339–346, 1984
 85. Bertelli G, Pronzoto P, Amoroso D: Adjuvant tamoxifen in primary breast cancer: Influence on plasma lipids and antithrombin III levels. *Breast Cancer Res Treat* 12: 307–310, 1988
 86. Love RR, Newcomb PA, Wiebe DA, Surawicz TS, Jordan VC, Carbone PP, DeMets DC: Lipid and lipoprotein effects of tamoxifen therapy in postmenopausal women with node negative breast cancer. *Breast Cancer Res Treat* abstract 204, 1989
 87. Smith I: Adjuvant tamoxifen for early breast cancer. *Br J Cancer* 57: 527–528, 1988
 88. Jordan VC, Fritz NF, Tormey DC: Endocrine effects of adjuvant chemotherapy and long-term tamoxifen administration on node positive patients with breast cancer. *Cancer Res* 47: 624–630, 1987
 89. Jordan VC, Fritz NF, Tormey DC: Long-term adjuvant therapy with tamoxifen: Effects on sex hormone binding globulin and antithrombin III. *Cancer Res* 47: 4517–4519, 1987
 90. Fex G, Adielson G, Mattson W: Oestrogen-like effects of tamoxifen on the concentration of proteins in plasma. *Acta Endocrinol Copen* 97: 109–113, 1981

91. Boccardo F, Buzzi L, Ruboyotti A, Nicolo G, Rosso R: Oestrogen-like action of tamoxifen on vaginal epithelium in breast cancer patients. *Oncology* 38: 281–285, 1981
92. Yager JD, Yager R: Oral contraceptive steroids as promoters of hepatocarcinogenesis in female Sprague-Dawley rats. *Cancer Res* 40: 3680–3685, 1980
93. Gau T. Stuart Pharmaceuticals, A division of ICI Americas, Wilmington, Delaware. Open letter to all US medical oncologists describing the toxicological findings in rats with high dose tamoxifen treatment, 1986
94. Fox H: Endometrial carcinogenesis and its relation to oestrogens. *Path Res Pract* 179: 13–19, 1984
95. Swenerton KD: Treatment of advanced endometrial adenocarcinoma with tamoxifen. *Cancer Treat Rep* 64: 805–811, 1980
96. Satyaswaroop PG, Zaino RJ, Mortel R: Estrogen-like effects of tamoxifen on human endometrial carcinoma transplanted into nude mice. *Cancer Res* 44: 4006–4010, 1984
97. Killackey MA, Hakes TB, Pierce VK: Endometrial adenocarcinoma in breast cancer patients receiving tamoxifen. *Cancer Treat Rep* 69: 237–238, 1985
98. Hardell L: Tamoxifen as risk factor for carcinoma of corpus uteri. *Lancet* ii: 563, 1988
99. Hardell L: Pelvic irradiation and tamoxifen as risk factors for carcinoma of corpus uteri. *Lancet* ii: 1432, 1988
100. Fornander T, Rutqvist LE, Cedermark B, Glas U, Mattson A, Silversward JC, Skoog L, Somell A, Theve T, Wilking N, Askergren J, Hjolmar ML: Adjuvant tamoxifen in early breast cancer: Occurrence of new primary cancers. *Lancet* i: 117–120, 1989
101. Stewart HJ, Knight GM: Tamoxifen and the uterus and endometrium. *Lancet* i: 375–376, 1989
102. Gottardis MM, Robinson SP, Satyaswaroop PG, Jordan VC: Contrasting actions of tamoxifen on endometrial and breast tumor growth in the athymic mouse. *Cancer Res* 48: 812–815, 1988
103. Groom GV, Griffiths K: Effect of the antioestrogen tamoxifen on plasma levels of luteinizing hormone, follicle-stimulating hormone, prolactin, oestradiol and progesterone in normal premenopausal women. *J Endocrinol* 70: 421–428, 1976
104. Tajima C, Fukushima T: Endocrine profile in tamoxifen-induced ovulatory cycles. *Fert Steril* 40: 23–27, 1983
105. Ravdin PM, Fritz NF, Tormey DC, Jordan VC: Endocrine status of premenopausal node positive breast cancer patients following adjuvant chemotherapy and long-term tamoxifen. *Cancer Res* 48: 1026–1029, 1988
106. Nicholson RI, Walker KJ, Turkes A, Turkes AO, Dyas J, Blamey RW, Cambell FC, Robinson MRG, Griffiths K: Therapeutic significance and the mechanism of action of the LH-RH agonist ICI 118,630 in breast cancer and prostatic cancer. *J Steroid Biochem* 20: 129–135, 1984