

*Report*

## **Tumor proliferative activity and response to first-line chemotherapy in advanced breast carcinoma**

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### **Summary**

The relationship between tumor proliferative activity and response to first-line chemotherapy and survival was investigated in 76 advanced breast cancer patients. Proliferative activity was determined by means of Ki-67 immunohistologic staining on primary tumors (55 patients) or at the relapse site (21 patients), and was classified as low ( $\leq 25\%$  of stained cells) or high ( $> 25\%$  of stained cells). The usual WHO response criteria were used. The median duration of follow-up was 18 months (range 3–58).

Forty-seven patients (62%) had tumors with low, and 29 (38%) had tumors with a high rate of proliferative activity. The two groups were well balanced in terms of important variables such as disease-free survival, performance status, age, menopausal status, and the type of first-line chemotherapy (anthracycline-based regimens versus cyclophosphamide-methotrexate-5-fluorouracil). The estrogen receptor (ER) content, measured by means of immunohistochemical assay, was markedly different in the two groups, with 27/47 tumors with low proliferative activity (57%) and 6/29 with high-proliferative activity (21%) being ER positive ( $\geq 45\%$  of stained cells) ( $p = 0.003$ ). Moreover, a significant difference in the metastatic pattern was also evident, with a higher incidence of bone and a lower incidence of soft tissue metastases in the group of patients with tumors with low proliferative activity ( $p = 0.004$ ). Overall, 10/47 responses (21%: PR = 7, and CR = 3) were observed in the group with a low rate of proliferative activity, versus 14/29 (48%: PR = 9, and CR = 5) in the group with highly proliferative tumors, the difference being statistically significant ( $p = 0.03$ ). When a multivariate analysis was performed, the only factor that retained independent prognostic significance was the predominant site of disease, particularly soft tissues ( $p = 0.003$ ). Despite the difference in response rate, when survival analysis was performed according to the Kaplan-Meier method, no significant difference was observed in the two groups, but when the analysis was limited to responsive patients, the median survival observed in those with a low and those with a high rate of proliferation was 35 and 19 months respectively ( $p = 0.02$ ). The same results were obtained when multivariate survival analysis was carried out using Cox's regression model. These data suggest that there is a link between tumor proliferative activity and response to chemotherapy in advanced breast cancer, and may indicate the need to use more intensive treatments in selected patients with highly proliferative tumors.

## Introduction

Tumors contain both proliferating cells (i.e. cells actively progressing toward mitosis in each phase of the cell cycle) and non-proliferating or quiescent cells [1]. The ratio between proliferating cells and the total number of cells in a given tumor sample (growth fraction) [2], varies greatly from one tumor type to another, epithelial cancer usually having a lower growth fraction than embryonal tumors or non-Hodgkin's lymphomas [3].

The proliferative activity of tumors can be determined in a number of ways, including the counting of the number of mitoses on a histologic section, the incorporation of tritiated thymidine (thymidine labeling index, TLI) or 5-bromo-2-deoxyuridine, the cytometric flow evaluation of the proportion of cells in the S- or S-G2 phases, or the histologic staining of monoclonal antibodies which recognize the antigens expressed only by proliferating cells [4].

The Ki-67 monoclonal antibody reacts with a nuclear antigen expressed during all of the phases of the cell cycle except G0 [5, 6], and is regarded as a marker of cell proliferation [7]. A correlation between Ki-67 positivity and the cell proliferation data obtained using other techniques, such as TLI [8–10], S-phase fraction [11], and 5-bromo-2-deoxyuridine incorporation [12] has been reported.

Given that cytotoxic chemotherapy mainly acts by killing dividing cells, it might be expected to be more active against rapidly proliferating tumors, as suggested by the results obtained in high-grade non-Hodgkin's lymphoma, lymphoblastic leukemia, and germ-cell tumors.

In advanced breast carcinoma, there are no biological markers to indicate the likelihood of a response to systemic chemotherapy and so the patients to be treated in this way are usually selected by means of a process of exclusion [13]. The identification of a link between proliferative activity and the response to chemotherapy may help in selecting those patients who could benefit from this treatment modality. The present study was designed to examine the relationship between tumor proliferative activity (as assayed by Ki-67) and the response to first-line chemotherapy in advanced breast cancer.

## Patients and methods

### Patients

At our Institution, Ki-67 and estrogen receptor (ER) immunostainings are routinely performed on fresh breast cancer samples. All of the patients with histologically documented breast cancer who experienced a relapse between January 1989 and April 1994, and in whom the Ki-67 and ER content of the primary tumors (55 patients) or at the relapse site (21 patients) could be determined, were included in this study. The patients were considered eligible if they had measurable or evaluable disease; they were considered ineligible if the only manifestation of disease was a malignant effusion, a previously irradiated lesion, brain metastasis, or nuclide scan evidence of disease. Clinical staging was based upon a complete history, physical examination, a routine biochemical profile, a complete blood cell count and the results of imaging procedures for all patients before the beginning and after three cycles of chemotherapy. Response was evaluated according to standard WHO criteria [14].

The chemotherapy administered was the first for metastatic disease. Standard protocols were used: cyclophosphamide 600 mg/m<sup>2</sup>, methotrexate 40 mg/m<sup>2</sup>, 5-fluorouracil 600 mg/m<sup>2</sup> on day 1 (CMF = 43 patients); cyclophosphamide 500 mg/m<sup>2</sup>, doxorubicin 50 mg/m<sup>2</sup> (or epirubicin 70 mg/m<sup>2</sup>), 5-fluorouracil 750 mg/m<sup>2</sup> on day 1 (CAF or CEF = 27 patients); cisplatin 100 mg/m<sup>2</sup> on day 1, etoposide 80 mg/m<sup>2</sup> on day 1 through 3 (3 patients); vinorelbine 25 mg/m<sup>2</sup> (2 patients); carboplatin 50 mg/m<sup>2</sup> on day 1 through 3, 5-fluorouracil 375 mg/m<sup>2</sup> on day 1 through 5, folinic acid 250 mg/m<sup>2</sup> on day 1 through 5 (1 patient). All of the drugs were given intravenously in cycles with a 3 or 4 week interval, except for vinorelbine which was given weekly.

### Immunohistochemical Ki-67 and ER assay

Immunohistochemical staining for replicative fraction cells was performed using Ki-67 monoclonal antibody (DAKO-PC) [15]. Air-dried thin frozen sections mounted on glass slides were immersed in

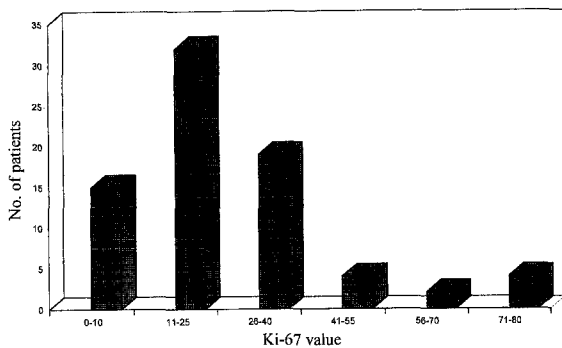


Fig. 1. Distribution of patients according to Ki-67 values.

acetone ( $-20^{\circ}\text{C}$ ) for 10 min and then, after rinsing in PBS, incubated for 1 hour, at room temperature, with anti-proliferation-associated antigen Ki-67 antibody (dilution 1:10) and immunostained using the APAAP technique. Counterstaining was performed using 3% methyl green. Field selection sought areas of highest Ki-67 expression evident by lower power scanning. Typically, the total cell count exceeded 1000 tumor cells. Specific staining was observed as red stained nuclei in Ki-67 positive cells. The results were expressed as the percentage of positive cells among the total number of cells.

The immunohistochemical ER staining procedure was performed using ERICA monoclonal kits (Abbott, Chicago, IL) [16]. At least 1000 tumor cells were examined, and the ER-positive cells were recognized by their brown stained nuclei. In a previous study [17], an ERICA threshold value of 45% gave the best level of sensitivity (0.810) and specificity (0.804) in comparison with the classical DCC (dextran coated charcoal) ER assay, and so the tumors were considered ER-negative if there was less than 45% of positive cells and ER-positive if there was 45% or more.

#### Statistical analysis

Crude and stratified analysis of the differences between groups were performed using the  $\chi^2$  statistics or Fisher's exact test [18]. Multivariate analysis, using unconditional logistic regression [19], was conducted in order to investigate the prognostic role of Ki-67 value and other explanatory covariates with

respect to the response rate, measured as a dichotomous variable. Overall survival (OS) estimates were obtained according to the Kaplan-Meier method [20] and the significance of the differences in survival time between the two groups was measured by the log-rank test [21]. In addition, a multivariate survival analysis was performed using the Cox's regression model [22]. Logistic regression was performed with SAS [23] and survival analysis was carried out using KMSURV [24] and COXSURV packages [22].

#### Results

The study involved 76 breast cancer patients who had had a relapse between January 1989 and April 1994, and for whom measurements of tumor proliferative activity by Ki-67 immunostaining and ER content were available. The stains were obtained either on the primary tumors (55 patients; 72%) or at the relapse site (21 patients; 28%). The distribution of Ki-67 at the two sites was similar, with 32/55 (58%) primary tumors and 15/21 (71%) metastases presenting a low proliferative rate ( $p = \text{n.s.}$ ).

There was a substantial range of Ki-67 expression in our patients (0% to 80%), with a median value of 25% (mean  $\pm$  SD = 25%  $\pm$  17%). The distribution of the patients according to Ki-67 status is shown in Fig. 1.

The correlation between the proliferative activity revealed by Ki-67 and the response to chemo-

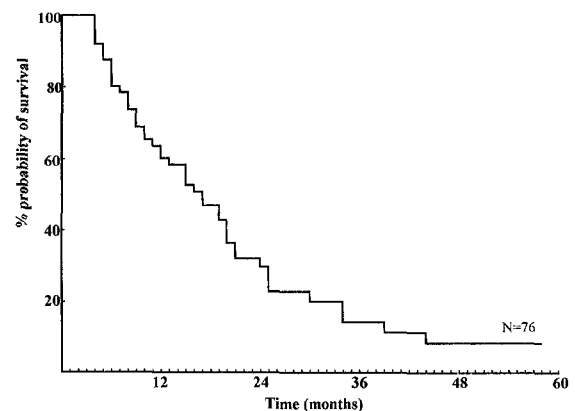


Fig. 2. Overall survival of patients.

therapy made it possible to identify the median value of 25% as a cut-off level separating two groups with a significant difference in response rate. The overall response rate was 32% (24 out of 76: partial responses [PR] = 16, and complete responses [CR] = 8; 95% confidence interval [CI], 22%–42%). The median duration of PR was 8 months (range 4–24), as was that of CR (range 2–17). Figure 2 shows the overall survival of all of the patients; the median duration of survival was 17 months.

When the response to chemotherapy was related to the predominant site of disease, regardless of the Ki-67 content, CR and PR were observed in 12/22 soft tissue (55%), 8/34 visceral (24%), and 4/20 bone metastases (20%) ( $p = 0.02$ ).

In the 47 patients with slowly proliferating tumors ( $\leq 25\%$  positive cells), 10 responses were observed (PR = 7, CR = 3) (21%; 95% CI, 9%–33%); in the 29 patients with highly proliferative tumors

(> 25% positive cells) there were 14 responses (PR = 9, CR = 5) (48%; 95% CI, 30%–66%). This difference was significant ( $p = 0.03$ ). The two groups were well balanced in terms of a number of important variables that may have influenced outcome, such as disease-free survival, performance status, age, menopausal status, and the type of first-line chemotherapy. The ER content differed markedly between the two groups, with 27/47 slowly proliferating tumors (57%) and 6/29 highly proliferative tumors (21%) being ER-positive ( $p = 0.003$ ). Moreover, there was also a significant difference in the metastatic pattern, with a higher incidence of bone and a lower incidence of soft tissue metastases in the group with slowly proliferating tumors ( $p = 0.004$ ) (Table 1).

The highest response rate (83%) was observed in the patients with a Ki-67 value of 50% or more. However, this cut-off point created a disproportion

Table 1. Patient characteristics according to Ki-67 status

Characteristic	Low Ki-67 (n = 47)		High Ki-67 (n = 29)		p value
	No.	%	No.	%	
Median age (range) (years)	57 (25–73)		54 (30–71)		n.s.
Performance status					
0	28	60	16	55	
1–2	19	40	13	45	n.s.
Menopausal status					
pre- or peri-	11	23	7	24	
post	36	77	22	76	n.s.
Estrogen receptor					
negative	20	43	23	79	
positive	27	57	6	21	0.003
Dominant disease site					
soft tissues	9	19	13	45	
viscera	20	43	14	48	
bone	18	38	2	7	0.004
Median DFS (range) (months)	20 (0–120)		16 (0–180)		n.s.
Adjuvant chemotherapy					
CMF	10	21	9	31	
FAC	4	9	4	14	n.s.
Prior hormonotherapy					
Adjuvant	13	28	3	10	
Metastatic	25	53	8	28	
Type of first-line chemotherapy					
CMF	29	62	13	45	
Anthracycline-based	13	28	14	48	
Other	5	10	2	7	n.s.
Median follow-up (range) (months)	17 (3–58)		17 (4–28)		

Table 2. Number of responses (CR-PR) according to the predominant disease site and Ki-67 status

Predominant disease site	(No. of patients)	Low Ki-67	High Ki-67
		CR-PR/No. of patients (%)	CR-PR/No. of patients (%)
Soft tissues	(22)	3/ 9 (33%)	9/13 (69%)
Viscera	(34)	3/20 (15%)	5/14 (36%)
Bone	(20)	4/18 (22%)	0/ 2

in the sample, with only six patients in the highly proliferative group.

The response rate of each predominant site of disease is shown in Table 2, according to the proliferative rate of the tumors. The highly proliferative tumors showed a higher response rate among soft tissues and visceral metastases, whereas the tumors with a low proliferative rate showed a higher response rate in bone metastases (only two tumors with a high level of Ki-67 staining had metastasized predominantly to the bone). However, according to the multivariate analysis, the only factor that retained independent prognostic significance across

the different fitted models was the predominant site of disease, particularly soft tissues ( $p = 0.003$ ), whereas borderline significance was observed for Ki-67 value ( $p = 0.05$ ).

A better response rate in high proliferative tumors was observed either in patients receiving CMF ( $n = 13$ ; CR-PR = 5, 38%) or an anthracycline-based regimen ( $n = 14$ ; CR-PR = 8, 57%); however, this difference was not statistically significant.

Table 3 shows the characteristics of the patients who responded to chemotherapy, according to their Ki-67 status.

Since an inverse relationship between the prolif-

Table 3. Characteristics of patients achieving an objective response according to Ki-67 status

Characteristic	Low Ki-67 (n = 47)	High Ki-67 (n = 29)
	No.	No.
Partial response	7	9
Complete response	3	5
Objective response	10 (21%)	14 (48%)
Performance status		
0	6	10
1-2	4	4
Menopausal status		
pre- or peri-	3	5
post	7	9
Estrogen receptor		
negative	4	13
positive	6	1
Dominant disease site		
soft tissues	3	9
viscera	3	5
bone	4	0
Median DFS (range) (months)	19 (0-52)	10 (0-32)
Type of first-line chemotherapy		
CMF	6	5
Anthracycline-based	4	8
Other	0	1
Median follow-up (range) (months)	18 (4-46)	9 (4-24)

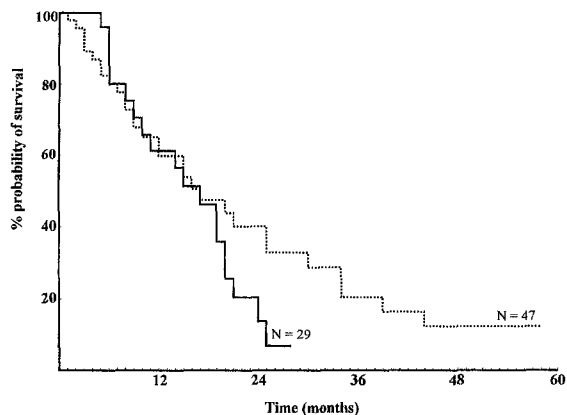


Fig. 3. Overall survival of patients according to Ki-67 values. (---) low Ki-67 values; (—) high Ki-67 values ( $p = ns$ ).

erative activity revealed by Ki-67 and ER content was observed, the correlation between response to chemotherapy and ER was evaluated. In the 43 ER-negative tumors, 17 responses were observed (39%) (95% CI, 24%–54%); in the 33 ER-positive tumors there were 7 responses (21%) (95% CI, 7%–35%) ( $p = ns$ ).

However, despite the better response rate observed in the tumors with a high level of Ki-67 staining, the survival curves of the two groups were not significantly different when the Kaplan-Meier method was applied (Fig. 3).

In the subset of responsive patients, a median survival of 35 and of 20 months was observed in those with slowly and rapidly proliferating tumors, respectively ( $p = 0.02$ ) (Fig. 4). The longer survival in low Ki-67 tumor patients achieving an objective response to first-line chemotherapy is mainly due to a more effective control of the disease played by the endocrine treatment administered later in the evolution of the disease. In fact, a hormonal treatment prescribed to 8/10 patients with low-proliferative tumors resulted, in 5 patients, in a stabilization of the tumor ranging from 6 to 12 months; in high proliferative tumors, mainly because of the aggressive course of the cancer, a hormonal treatment was given only to 2 patients, without observing any response or stabilization.

Finally, when multivariate survival analysis was performed using the Cox's regression model, the results were comparable to those obtained with uni-

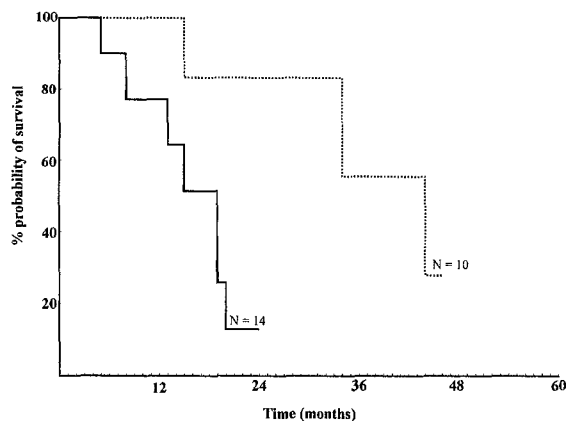


Fig. 4. Overall survival of responsive patients according to Ki-67 values. (---) low Ki-67 values; (—) high Ki-67 values ( $p = 0.02$ ).

variate analysis. More specifically, the independent prognostic role of Ki-67 value was confirmed in the subset of responsive patients ( $p = 0.03$ ).

## Discussion

In breast cancer, tumor proliferative activity, as revealed by Ki-67 immunostaining [25–28], TLI [29, 30], and S-phase fraction by DNA flow-cytometry [31–33], is a well defined prognostic factor, with highly proliferative tumors being associated with shorter disease-free and overall survival. However, while these data underline the more aggressive biological behavior of rapidly proliferating tumors, they are only marginally informative about the possible role of tumor proliferative activity as a predictor of response to chemotherapy. In early breast cancer, the majority of available data suggest a significant correlation between the pre-treatment tumor proliferative state measured by means of DNA flow-cytometry [34, 35] and TLI [36], and the response to preoperative chemotherapy. In advanced breast cancer, only one article dealing with the possible role of tumor proliferation (by TLI) in the response to chemotherapy has been published [37]. In this study of 25 patients the response to chemotherapy was significantly higher in tumors expressing a higher TLI.

In tumors characterized by a higher response rate to chemotherapy, such as non-Hodgkin's lympho-

mas, the relationship between tumor proliferation and response to chemotherapy is more complex and the results are conflicting. In general, there is a correlation between the rate of proliferation and the grade of lymphoma, with low-grade lymphomas expressing a low growth fraction and vice-versa [38–40]. In this disease, the impact of proliferation on survival is generally reported as being negative [41–44], and only rarely it is positively associated with response [45].

In the present study, a Ki-67 value of 25% allowed the identification of two groups of patients with a different probability of responding to chemotherapy. The only imbalance between the two groups was represented by the ER content and the dominant site of disease, with an excess of soft tissue involvement in the group with highly proliferative tumors and an excess of bone involvement in those with slowly proliferating tumors. This particular pattern of diffusion is probably due to a selectivity in the metastatic process related to differences in phenotypes. The importance of the predominant site of disease in determining the response to chemotherapy is well known, and it seems that the response rate of soft tissue and visceral metastases (the sites that most often respond to chemotherapy) is related to proliferative activity. The lack of statistical significance in our study could be due to the limited size of the sample.

The significant inverse relationship between Ki-67 expression and ER content is in agreement with a number of already published data relating tumor proliferation to ER content [46–50].

The response rate among highly proliferative tumors was significantly higher than in the group with slowly proliferating tumors. However, only 48% of the rapidly proliferating tumors responded to chemotherapy, thus making this index insuitable as a predictor of response; furthermore, although the response rate in the group with slowly proliferating tumors was lower (21%), it was certainly not negligible. Both a stable [51], but more often an increasing TLI-revealed proliferation rate from primary to methachronous lesions have been reported [52, 53]. In the present study, there was a temporal (and biological) gap between the time at which proliferative activity was determined (at cancer diagnosis, in

72% of the patients) and the beginning of chemotherapy for advanced disease, and this may have weakened the association between the two variables.

The survival curves of patients with rapidly or slowly proliferating tumors did not differ significantly. In the whole group, median survival was 17 months, which compares well with data from studies reporting a higher response rate [54]. Of interest is the analysis of the survival curves of responsive patients, which shows better survival in the group with slowly proliferating tumors. This fact is probably related to the differential effect of hormone-therapy in the two groups of patients.

The cell cycle is controlled by a number of factors: oncogenes substituting growth factors (e.g. *jun*, *fos*, *mos*) or promoting cell survival (*bcl-2*), and tumor-suppressor genes monitoring progression through the G<sub>1</sub> phase (*Rb*, *P53*) etc. [55], and these may play a role in the response to chemotherapy [56]. Furthermore, resistance to chemotherapy has been linked to the expression of membrane proteins, small cytoplasmic peptides (glutathione), enzymes, and many other factors [57], although their relationship to the proliferative state of the tumors has not yet been characterized. It is likely that only the simultaneous study of some of these factors would increase the possibility of predicting responsiveness to chemotherapy.

In conclusion, our data suggest the importance of studying the relationship between tumor proliferation and both the response rate to chemotherapy and patient survival. The higher response rate in rapidly proliferating tumors does not lead to better survival, thus confirming the view that malignancies are not curable by chemotherapy because they proliferate rapidly [58] and perhaps indicating the need for an intensification of treatment on the basis of the rate of tumor proliferation.

## References

1. Tubiana M: Tumor cell proliferation kinetics and tumor growth rate. *Acta Oncol* 28: 113–121, 1988
2. Mendelsohn ML: The growth fraction: a new concept applied to tumors. *Science* 132: 1496, 1960

3. Tubiana M, Malaise EP: Comparison of cell proliferation kinetics in human and experimental tumors: response to irradiation. *Cancer Treat Rep* 60: 1887–1895, 1976
4. Quinn CM, Wright NA: The clinical assessment of proliferation and growth in human tumours: evaluation of methods and applications as prognostic variables. *J Pathol* 160: 93–102, 1990
5. Gerdes J, Schwab U, Lemke H, Stein H: Production of a mouse monoclonal antibody reactive with a human nuclear antigen associated with cell proliferation. *Int J Cancer* 31: 13–20, 1983
6. Gerdes J, Lemke H, Baisch H, Wacker HH, Schwab U, Stein H: Cell cycle analysis of a cell proliferation-associated human nuclear antigen defined by the monoclonal antibody Ki-67. *J Immunol* 133: 1710–1715, 1984
7. Gerdes J, Dallenbach F, Lennert K, Lemke H, Stein H: Growth fractions in malignant non-Hodgkin lymphomas (NHL) as determined *in situ* with the monoclonal antibody Ki-67. *Hemathol Oncol* 2: 365–371, 1984
8. Kamel OW, Franklin WA, Ringus JC, Meyer JS: Thymidine labeling index and Ki-67 growth fraction in lesions of the breast. *Am J Pathol* 134: 107–113, 1989
9. Bouzubar N, Walker KJ, Griffiths K, Ellis IO, Elston CW, Robertson JF, Blamey RW, Nicholson RI: Ki-67 immunostaining in primary breast cancer: pathological and clinical associations. *Br J Cancer* 59: 943–947, 1989
10. Deshmukh P, Ramsey L, Garewal HS: Ki-67 labeling index is a more reliable measure of solid tumor proliferative activity than tritiated thymidine labeling. *Am J Clin Pathol* 94: 192–195, 1990
11. Vielh P, Chevillard S, Mosseri V, Donatini B, Magdelenat H: Ki-67 index and S-phase fraction in human breast carcinomas. Comparison and correlation with prognostic factors. *Am J Pathol* 94: 681–686, 1990
12. Gaglia P, Bernardi A, Venesio T, Caldarola B, Lauro D, Cappa APM, Calderini P, Liscia DS: Cell proliferation of breast cancer evaluated by anti-BrdU and anti-Ki-67 antibodies: its prognostic value on short-term recurrences. *Eur J Cancer* 29A: 1509–1513, 1993
13. Henderson IC, Garber JE, Breitmeyer JB, Hayes DF, Harris JR: Comprehensive management of disseminated breast cancer. *Cancer* 66: 1439–1448, 1990
14. Miller AB, Hoogstraten B, Staquet M, Winkler A: Reporting results of cancer treatment. *Cancer* 47: 207–214, 1981
15. Gerdes J, Pickartz H, Brotherton J, Hammerstein J, Weitzel H, Stein H: Growth fractions and estrogen receptors in human breast cancers as determined *in situ* with monoclonal antibodies. *Am J Pathol* 129: 486–492, 1987
16. Pertschuk LP, Eisenberg KB, Carter AC, Feldman JG: Immunohistologic localization of estrogen receptors in breast cancer with monoclonal antibodies. *Cancer* 55: 1513–1518, 1985
17. Molino A, Micciolo R, Turazza M, Bonetti F, Piubello Q, Corgnati A, Sperotto L, Martignoni G, Bonetti A, Nortilli R, Castelli P, Rodella S, Capelli P, Manfrin E, Pelosi G, Cetto GL: Estrogen receptors in 699 primary breast cancers: a comparison of immunohistochemical and biochemical methods. *Breast Cancer Res Treat* 34: 221–228, 1995
18. Fleiss LJ: Assessing significance in a fourfold table. In: Fleiss LJ (ed) *Statistical Methods for Rates and Proportions*. John Wiley & Sons, New York, 1981, pp 19–32
19. Hosmer DW, Lemeshow S: *Applied Logistic Regression*. John Wiley & Sons, New York, 1989
20. Kaplan EL, Meier P: Non-parametric estimation from incomplete observations. *J Am Stat Assoc* 53: 457–461, 1958
21. Mantel N: Evaluation of survival data and two new rank order statistics arising in its consideration. *Cancer Chemother Rep* 50: 163–170, 1966
22. Campos-Filho N, Franco EL: COXSURV: a microcomputer program for multiple regression by Cox proportional hazards model. *Comp Meth Prog Biomed* 31: 81–87, 1990
23. SAS Institute Inc. *SAS Guide for Personal Computers*, version 6.03. SAS Institute Inc., Cary, NC
24. Campos-Filho N, Franco EL: KMSURV: a microcomputer program for univariate survival data analysis. *Comp Meth Prog Biomed* 27: 223–228, 1988
25. Veronese SM, Gambacorta M, Gottardi O, Scanzi F, Ferrari M, Lampertico P: Proliferation index as a prognostic marker in breast cancer. *Cancer* 71: 3926–3931, 1993
26. Wintzer HO, Zipfel I, Schulte-Mönting J, Hellerich U, von Kleist S: Ki-67 immunostaining in human breast tumors and its relationship to prognosis. *Cancer* 67: 421–428, 1991
27. Sahin AA, Jungsil R, Jae YR, Blick MB, El-Naggar AK, Ordonez NG, Fritsche HA, Smith TL, Hortobagyi GN, Ayala AG: Ki-67 immunostaining in node-negative stage I/II breast carcinoma. *Cancer* 68: 549–557, 1991
28. Weikel W, Beck T, Mitze M, Knapstein PG: Immunohistochemical evaluation of growth fractions in human breast cancers using monoclonal antibody Ki-67. *Breast Cancer Res Treat* 18: 149–154, 1991
29. Silvestrini R, Daidone MG, Valagussa P, Di Fronzo G, Mezzanotte G, Bonadonna G: Cell kinetics as a prognostic indicator in node-negative breast cancer. *Eur J Cancer Clin Oncol* 25: 1165–1171, 1989
30. Silvestrini R, Daidone MG, Valagussa P, Di Fronzo G, Mezzanotte G, Mariani L, Bonadonna G: <sup>3</sup>H-thymidine-labeling index as a prognostic indicator in node-positive breast cancer. *J Clin Oncol* 8: 1321–1326, 1990
31. Muss HB, Kute TE, Case D, Smith LR, Booher C, Long R, Kammire L, Gregory B, Brockschmidt JK: The relationship of flow cytometry to clinical and biologic characteristics in women with node-negative primary breast cancer. *Cancer* 64: 1894–1900, 1989
32. O'Reilly SM, Camplejohn DM, Barnes DM, Millis RR, Rubens RD, Richards MA: Node-negative breast cancer: prognostic subgroups defined by tumor size and flow cytometry. *J Clin Oncol* 8: 2040–2046, 1990
33. Clark GM, Dressler LG, Owens MR, Pounds G, Oldaker T, McGuire WL: Prediction of relapse or survival in patients with node negative breast cancer by DNA flow cytometry. *N Engl J Med* 320: 627–633, 1989
34. Remvikos Y, Beuzebec P, Zajdela A, Voillemot N, Magdel-



- enat H, Pouillart P: Correlation of pretreatment proliferative activity of breast cancer with the response to cytotoxic chemotherapy. *J Natl Cancer Inst* 81: 1383-1387, 1989
35. O'Reilly SM, Camplejohn RS, Rubens RD, Richard MA: DNA flow cytometry and response to preoperative chemotherapy for primary breast cancer. *Eur J Cancer* 28: 681-683, 1992
  36. Gardin G, Alama A, Rosso R, Campora E, Repetto L, Pronzato P, Merlini L, Naso C, Camoriano A, Meazza R, Barbieri F, Baldini E, Giannessi PG, Conte PF: Relationship of variations in cell kinetics induced by primary chemotherapy to tumor regression and prognosis in locally advanced breast cancer. *Breast Cancer Res Treat* 32: 311-318, 1994
  37. Sulkes A, Livingston RB, Murphy WK: Tritiated thymidine labeling index and response in human breast cancer. *J Natl Cancer Inst* 62: 513-515, 1979
  38. Weiss LM, Strickler JG, Medeiros LJ, Gerdes J, Stein H, Warnke RA: Proliferative rates of non-Hodgkin lymphomas as assessed by Ki-67 antibody. *Hum Pathol* 18: 1155-1159, 1987
  39. Diamond LW, Bharat NN, Rappaport H: Flow cytometry in the diagnosis and classification of malignant lymphoma and leukemia. *Cancer* 50: 1122-1135, 1982
  40. Korkolopoulou P, Patsouris E, Pangalis G, Tsenga A, Elemenoglou J, Thomas-Tsangali E, Spandidos D, Kittas C: A comparative assessment of proliferating cell nuclear antigen, c-myc p62, and nucleolar organizer region staining in non-Hodgkin lymphomas: a histochemical and immunohistochemical study of 200 cases. *Human Pathol* 24: 371-377, 1993
  41. Cowan RA, Harris M, Jones M, Crowther D: DNA content in high and intermediate grade non-Hodgkin's lymphoma - prognostic significance and clinico-pathological correlations. *Br J Cancer* 60: 904-910, 1989
  42. Grierson HL, Wooldridge TN, Purtilo DT, Pierson J, Bast M, Wooldridge L, Armitage JO, Weisenburger DD: Low proliferative activity is associated with a favorable prognosis in peripheral T-cell lymphoma. *Cancer Res* 50: 4845-4848, 1990
  43. Silvestrini S, Costa A, Boracchi P, Giardini R, Rilke F: Cell proliferation as a long-term prognostic factor in diffuse large-cell lymphomas. *Int J Cancer* 54: 231-236, 1993
  44. Miller TP, Grogan TM, Dahlberg S, Spier MC, Brazier RM, Banks PM, Foucar K, Kjeldsberg CR, Levy N, Nathwani BN, Schnitzer B, Tubbs RR, Gaynor ER, Fisher RI: Prognostic significance of the Ki-67-associated proliferative antigen in aggressive non-Hodgkin lymphomas: a prospective Southwest Oncology Group trial. *Blood* 83: 1460-1466, 1994
  45. Williamson JMS, Grigor I, Smith MEF, Holgate CS, O'Brien CJ, Morgan DR, Quirke P, Alison DL, Child JA, Bird CC: Ploidy, proliferative activity, cluster differentiation, antigen expression and clinical remission in high-grade non-Hodgkin lymphoma. *Histopathology* 11: 1043-1054, 1987
  46. Silvestrini R, Daidone MG, Bertuzzi A, Di Fronzo G: Relationship between estrogen receptors and cellular proliferation. *Recent Results Cancer Res* 91: 163-168, 1984
  47. Gasparini G, Dal Fior S, Pozza F, Bevilacqua P: Correlation of growth fraction by Ki-67 immunohistochemistry with histologic factors and hormone receptors in operable breast carcinoma. *Breast Cancer Res Treat* 14: 329-336, 1989
  48. Silvestrini R, Daidone MG, Del Bino G, Mastore M, Luisi A, Di Fronzo G, Boracchi P: Prognostic significance of proliferative activity and ploidy in node-negative breast cancers. *Ann Oncol* 4: 213-219, 1993
  49. Di Stefano D, Mingazzini P, Scucchi L, Donnetti M, Marinuzzi V: A comparative study of histopathology, hormone receptors, peanut lectin binding, Ki-67 immunostaining and nucleolar organizer region-associated proteins in human breast cancer. *Cancer* 67: 463-471, 1991
  50. Veronese SM, Gambacorta M: Detection of Ki-67 proliferation rate in breast cancer. Correlation with clinical and pathologic features. *Am J Clin Pathol* 95: 30-34, 1991
  51. Meyer JS, McDivitt RW: Reliability and stability of the thymidine labeling index of breast carcinoma. *Lab Invest* 54: 160-164, 1986
  52. Meyer JS, Lee JY: Relationships of S-phase fraction of breast carcinoma in relapse to duration of remission, estrogen receptor content, therapeutic responsiveness, and duration of survival. *Cancer Res* 40: 1890-1896, 1980
  53. Silvestrini R, Valentinis B, Daidone MG, Di Fronzo G, Coradini D, Salvadori B: Biological characterisation of primary and methachronous lesions in breast cancer. *Eur J Cancer* 28A: 2006-2010, 1992
  54. Harris JR, Morrow M, Bonadonna G: Cancer of the breast. In: De Vita VT Jr, Hellman S, Rosenberg SA (eds) *Cancer: Principles and Practice of Oncology* (ed 4). Lippincott, Philadelphia, 1993, pp 1264-1332
  55. Smets LA: Programmed cell death (apoptosis) and response to anti-cancer drugs. *Anti-Cancer Drugs* 5: 3-9, 1994
  56. Fisher TC, Milner AE, Gregory CD, Jackman AL, Aherne GW, Hartley JA, Dive C, Hickman J: bcl-2 modulation of apoptosis induced by anticancer drugs: resistance to thymidylate stress is independent of classical resistance pathways. *Cancer Res* 53: 3321-3326, 1993
  57. Goldie JH: Drug resistance. In: Perry MC (ed) *The Chemotherapy Source Book*. Williams & Wilkins, Baltimore, 1992, pp 54-66
  58. Armitage JO: Tumor proliferative rate and response to chemotherapy. *Ann Int Med* 116: 771-773, 1992