

Alignment Statistic for Identifying Related Protein Sequences

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Summary. Closely related proteins show an obvious kinship by having numerous matching amino acids in their aligned sequences. Kinship between anciently separated proteins requires a statistical evaluation to rule out fortuitous similarities. A simple statistic is developed which assumes equal probability for all codon pairs, and a table of critical values for amino acid sequence alignments of length 200 or less is presented. Applying this statistic to *V* and *C* regions of immunoglobulin chains, aligned on the basis of shared features of three-dimensional structure, provides evidence that the *V* and *C* sequences descended from a common ancestor. Similarly the distant evolutionary relationship of dehydrogenases, flavodoxin, and subtilisin, suggested by structural alignments, is verified. On the other hand, the statistic does not verify a common evolutionary origin for the heme binding pocket in globins and cytochrome *b₅*. Empirical evidence from the distribution of MMD values of amino acid pairs in comparisons of misaligned polypeptide chains and from Monte Carlo trials of sequences aligned with arbitrary gaps supports the validity of the statistic.

Key words: Structural alignments – Minimum mutation distance – Evolutionary relationship – Significance test – *V* and *C* immunoglobulin sequences – Dehydrogenases.

1. Introduction

Proteins recently separated from a common evolutionary progenitor show kinship by having numerous matching amino acids in their sequence structures. Anciently separated proteins usually do not reveal their kinship so easily, as many substitutions, insertions, and deletions of amino acids have accumulated in them. Sensitive criteria are needed to detect any sequence homologies which remain and to decide if they are

of sufficient scope to support the hypothesis of descent from a common ancestor. We develop here an alignment statistic which successfully identifies such distant relationships. We present this statistic in the form of a significance table for pairs of aligned amino acid sequence chains ranging from one to two hundred amino acid positions. We then use the table to illustrate a major application in the study of protein evolution. This application is to test sequence alignments proposed for functionally different protein chains, or portions of the chains, from structural similarities revealed by x-ray crystallography. In this regard, we find that the structural alignments proposed by Poljak et al. (1974) for the two kinds of regions in the antibody molecule named variable (*V*) and constant (*C*) show true sequence homology as judged by our significance table. This is also the case for stretches of dehydrogenases, flavodoxin, and subtilisin aligned on structural grounds by Rossman et al. (1974). In contrast the equivocal suggestion of homology on comparing the heme binding pocket in globins and cytochrome *b₅* (Rossman and Argos, 1975) is not supported by the alignment statistic.

Our test for common ancestry in an amino acid sequence alignment joins a number of others currently in the literature. Fitch (1970, 1975) and Jukes and Cantor (1969) determine the minimum mutation distance for each subalignment of a given length (say, 30) within the alignment as a whole, and compare this to expectations based upon random rearrangements of the same amino acid composition. Needleman and Wunsch (1970) determine the amino acid difference (i.e., number of nonidentical amino acids) in an entire alignment, and compare this to expectation. Sankoff (1972) and Barker and Dayhoff (1972) use modified versions of the Needleman and Wunsch method. Haber and Koshland (1970) employ a functional approach: they determine the number of identical amino acid pairs and electrochemically similar amino acid pairs in an alignment, and compare this to random expectation. McLachlan (1971, 1972) has further developed this functional approach. Our method employs the minimum mutation distance for the alignment as a whole, and compares this to expectations based upon an equal and independent probability of each nucleotide pair.

2. The Alignment Statistic

In the significance test to determine whether an aligned pair of sequences of length = *n* (hereinafter denoted, "*n*-alignment") share a common ancestry, we wish to distinguish between the *null hypothesis* (absence of common ancestry) and the most plausible *alternative hypothesis* (presence of common ancestry). If an *n*-alignment satisfies the null hypothesis, then we expect its *minimum mutation distance* (Fitch and Margoliash, 1967) or MMD, to belong to a random collection of *n*-alignments. Alternatively, if the *n*-alignment has a sufficiently recent common ancestry, then its MMD would be smaller than expected from the null hypothesis, because of the inherited matched alignment positions. The null hypothesis is rejected for an *n*-alignment if the probability that that *n*-alignment was generated randomly is less than an arbitrary cutoff point α ($\alpha = 0.05, 0.01, \text{etc.}$).

For a random 1-alignment (consisting of a single codon pair), there are 61 possible non-terminating codons for one sequence and 61 for the other, $61 \times 61 = 3721$ possible codon pairs all told. We are not assuming that nucleotide pair alignments are equiprobable in DNA known to share a common ancestry, and indeed such has been called into question by the empirical studies of Barker and Dayhoff (1972). Here we are setting up the *null hypothesis*, for the case in which common ancestry is assumed to be *absent*. For a random n -alignment, there are 3721^n possible sequence pairs. In generating a random distribution we assume that each of the 3721^n n -alignments has equal probability of occurrence, namely $\frac{1}{3721^n}$ apiece. Some of the n -alignments

are perfectly matched, some have a single nucleotide difference, . . . , and some are perfectly mismatched (i.e., with 3^n nucleotide differences). The probability that an n -alignment has exactly i observed nucleotide differences is denoted $p(i, n)$.

Under those conditions, we find in the 1-alignment problem, 61 occurrences of perfectly matched alignments, 526 occurrences of alignments with a single nucleotide difference, 1568 occurrences with 2 differences, and 1566 occurrences with 3 differences; $61 + 526 + 1568 + 1566 = 3721$ occurrences all told. We say that the probability of actual difference = 0 is $\frac{61}{3721} = 0.01639$, probability of 1 is $\frac{526}{3721} = 0.14136$, probability of 2 is $\frac{1568}{3721} = 0.42139$, and probability of 3 is $\frac{1566}{3721} = 0.42085$.

Since our observations consist of amino acid pairs, we will tend to underestimate the true nucleotide difference. For example, codons AUU and AUA have an actual distance = 1, but since both codons specify the same amino acid (isoleucine), at the amino acid level the observed distance between them is 0. The observed difference between a pair of codons is obtained by mapping each codon into its corresponding amino acid by means of the genetic code, and then finding the *minimum mutation distance* (MMD) for that pair of amino acids (Table 1). The MMD for a pair of amino acids is the minimum number of nucleotide steps necessary to convert one amino acid into the other. By examining the 3721 possible 1-alignments in light of *observed differences*, we find the probability of observed difference = 0 is $\frac{235}{3721} = 0.06316$, proba-

Table 1. The observed difference between a pair of codons is obtained by mapping each non-terminating codon into its corresponding amino acid, and then finding the MMD for that pair of amino acids.

	Codon Pair		Amino Acid Pair		Actual Difference	Observed Difference
1.	AAA	AAA	LYS	LYS	0	0
2.	AAA	AAC	LYS	ASN	1	1
3.	AAA	AAG	LYS	LYS	1	0
4.	AAA	AAU	LYS	ASN	1	1
5.	AAA	ACA	LYS	THR	1	1
6.	AAA	ACC	LYS	THR	2	1
.						
.						
3721.	UUU	UUU	PHE	PHE	0	0

bility of 1 is $\frac{1706}{3721} = 0.45848$, probability of 2 is $\frac{1698}{3721} = 0.45633$, and probability of 3 is $\frac{82}{3721} = 0.02204$. Note that we are not altering the actual probability, say, of a 0-nucleotide difference (it remains at 61/3721); we are merely stating that an additional 174 codon comparisons which actually have higher difference values are *observed* as having 0 differences. In other words, our test compensates for the fact that the full nucleotide difference implicit in certain amino acid pairs is not actually seen in the data. This blunting effect of the MMD statistic tends to render our test more conservative – some alignments which are actually similar by reason of common ancestry will be missed by our statistic.

For $n = 1$, we have already shown that $p(0,1) = 0.06316$, $p(1,1) = 0.45848$, $p(2,1) = 0.45633$, and $p(3,1) = 0.02204$. For $n = 2$, it is possible to calculate $p(j,2)$ by listing out all possible 2-alignments ($3721^2 = 13,845,841$ all told), and tabulating how many have observed difference = 0, how many have observed difference = 1, etc. There is a *recursion formula* which allows us to build up a table of p 's in a stepwise fashion. In a recursion formula, if we know the answer for $n = 1$ (which we do), we can get the answer for $n = 2$; if we know $n = 2$, we can get $n = 3$; etc. To calculate $p(i, n)$, we separate the n -alignment into a *first part* of length = $n - 1$ and a *second part* of length = 1. There are only four possible ways that the n -alignment could have observed difference = i : (a) if the first part has difference i and the second part has difference 0; (b) first part $i - 1$, second part 1; (c) first part $i - 2$, second part 2; (d) first part $i - 3$, second part 3. The second part cannot have length greater than 3 because it is a single codon pair. Thus we calculate (Table 2); $p(i, n) = 0.06316 \cdot p(i, n - 1) + 0.45848 \cdot p(i - 1, n - 1) + 0.45633 \cdot p(i - 2, n - 1) + 0.02204 \cdot p(i - 3, n - 1)$. For example, the probability of obtaining 3 differences from a 2-alignment ($i = 3, n = 2$) is $p(3,2) = 0.06316 \cdot 0.02204 + 0.45848 \cdot 0.45633 + 0.45633 \cdot 0.45848 + 0.02204 \cdot 0.06316 = 0.42122$.

Each row in Table 2 can be expressed as a histogram, which establishes the cutoff point for the significance test. Figure 1a shows the histogram for $n = 1$; Figure 1b shows the histogram for $n = 4$. When we state that the 5% significance level for a 4-alignment is 3, we mean that there is *less than* a 5% chance (precisely, a 4.035% chance) that a 4-alignment will have 3 or fewer nucleotide differences. The cutoff point 3 is established by cutting the histogram for the 4-alignment (Fig. 1b) at the right most point such that the left tail of the histogram sums to less than 5%. For any significance level α and any n -alignment, the cutoff point m is established by

Table 2. Recursive evaluation of $p(i, n)$ by the formula

$$p(i, n) = 0.06316 \cdot p(i, n - 1) + 0.45848 \cdot p(i - 1, n - 1) + 0.45633 \cdot p(i - 2, n - 1) + 0.02204 \cdot p(i - 3, n - 1).$$

$p(i, n)$ is the probability that an n -alignment has exactly i observed nucleotide differences

n	$p(0, n)$	$p(1, n)$	$p(2, n)$	$p(3, n)$	$p(4, n)$	$p(5, n)$	$p(6, n)$
1	0.06316	0.45848	0.45633	0.02204	—	—	—
2	0.00399	0.05791	0.26784	0.42122	0.22844	0.02011	0.00049
3	0.00025	0.00549	0.04529	0.17592	0.33105	0.30412	0.12278 etc.
4	0.00002	0.00046	0.00549	0.03438	0.12235	0.25226	0.30213 etc.

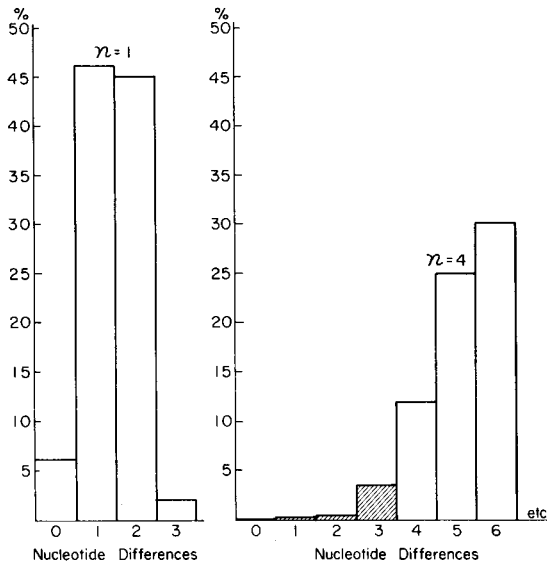


Fig. 1. a The significance histogram for $n = 1$. b The significance histogram for $n = 4$. The hatched area in this histogram represents the maximum part of the left tail of the histogram which sums to less than 5 %

cutting the n -alignment histogram at the rightmost (i.e., maximum) point such that the left tail of the histogram (i.e., all histogram bars up to and including m) sums to less than α . This is achieved by maximizing m , where

$$\sum_{i=0}^m p(i, n) < \alpha$$

Table 3 shows the maximum observed difference for n between 1 and 200 at selected significance levels ($\alpha = 0.95, 0.50, 0.05, 0.01, 0.001, 0.0001$).

3. Structural Alignments

V and C Sequences. Each different kind of immunoglobulin chain can be divided into V and C regions which are thought to share a remote common ancestry. Strong sequence homology exists among the different V regions, also among the different C regions (Jukes and Cantor, 1969; Barker and Dayhoff, 1972). Homology between V and C sequences, however, has not been obvious. The assumption of a remote ancestry between them rests on such features as similar location of disulfide-bonded cysteine residues and similar chain folding patterns. Poljak et al. (1974) have now used the common three-dimensional structural features revealed by x-ray crystallography at 2.0\AA resolutions for two V and two C regions in a particularly well studied human myeloma immunoglobulin to align these V and C sequences. We find significant homology between them by our alignment statistic as shown in Table 4. To our knowledge this is the first clear statistical evidence from sequence data for the hypothesis of common genetic ancestry of V and C regions.

An immunoglobulin molecule of the IgG class contains two identical light chains, each about 220 amino acids long, and two identical heavy chains, each about 440 amino

Table 3. Critical values for n -alignments where $n \leq 200$. At each significance level, the greatest observed MMD at which the null hypothesis of no common ancestry can still be rejected is given. For example, for a 14-alignment, the null hypothesis can be rejected at the 5% level for an observed MMD of 15 or less.

N	.01%	.1%	1%	5%	10%	50%	95%	N	.01%	.1%	1%	5%	10%	50%	95%
1	-1	-1	-1	-1	0	1	1	101	118	124	129	133	136	144	155
2	-1	-1	0	0	1	2	2	102	120	125	130	135	137	144	156
3	-1	0	0	1	2	3	3	103	121	127	132	136	139	147	158
4	0	1	2	3	3	4	4	104	122	128	133	138	140	148	159
5	1	2	3	4	4	5	5	105	124	129	134	139	141	150	161
6	2	3	4	5	5	6	6	106	125	131	136	140	143	151	162
7	3	4	5	6	6	7	7	107	126	132	137	142	144	153	164
8	4	5	6	7	7	8	8	108	127	133	139	143	146	154	165
9	5	6	7	8	8	9	9	109	129	135	140	145	147	156	167
10	6	7	8	9	10	11	11	110	130	136	141	146	148	157	168
11	7	8	10	11	12	13	13	111	131	137	143	147	150	159	170
12	8	9	11	13	13	14	14	112	133	139	144	149	151	160	171
13	9	10	12	14	15	16	15	113	134	140	145	150	153	161	172
14	10	12	13	15	16	17	16	114	135	141	147	151	154	163	174
15	11	13	15	16	17	18	17	115	137	143	148	153	155	164	176
16	12	14	16	18	19	20	18	116	138	144	149	154	157	166	177
17	13	15	17	19	20	21	19	117	139	145	151	156	158	167	179
18	14	16	18	20	21	22	20	118	140	147	152	157	160	169	180
19	16	17	20	22	23	24	21	119	142	148	154	158	161	170	182
20	17	19	21	23	24	25	22	120	143	149	155	160	162	171	183
21	18	20	22	24	25	26	23	121	144	151	156	161	164	173	185
22	19	21	24	26	27	28	24	122	146	152	158	163	165	174	186
23	20	22	25	27	28	29	25	123	147	153	159	164	167	176	188
24	21	24	26	28	29	30	26	124	148	155	160	165	168	177	189
25	23	25	27	30	31	32	27	125	150	156	162	167	169	179	190
26	24	26	29	31	32	33	28	126	151	157	163	168	171	180	192
27	25	27	30	32	34	35	29	127	152	159	165	170	172	182	193
28	26	29	31	34	35	36	30	128	153	160	166	171	174	183	195
29	27	30	33	35	36	37	31	129	155	161	167	172	175	184	196
30	29	31	34	36	38	39	32	130	156	163	169	174	176	186	198
31	30	32	35	38	39	40	33	131	157	164	170	175	178	187	199
32	31	34	36	39	40	41	34	132	159	165	171	176	179	189	201
33	32	35	38	40	42	43	35	133	160	167	173	178	181	190	202
34	33	36	39	42	43	44	36	134	161	168	174	179	182	192	204
35	35	37	40	43	44	45	37	135	163	170	175	181	183	193	205
36	36	38	41	44	45	46	38	136	164	171	177	182	185	194	207
37	37	40	43	46	47	48	39	137	165	172	178	183	187	196	208
38	38	41	44	47	49	50	40	138	167	174	180	185	188	197	210
39	40	42	46	48	50	51	41	139	168	175	181	186	189	198	211
40	41	44	47	50	51	52	42	140	169	176	182	188	190	200	213
41	42	45	48	51	53	54	43	141	170	178	184	189	192	202	214
42	43	46	50	52	54	55	44	142	172	179	185	190	193	203	216
43	44	48	51	54	55	56	45	143	173	180	186	192	195	205	217
44	46	49	52	55	57	58	46	144	174	182	188	193	196	206	219
45	47	50	54	57	58	59	47	145	176	183	189	195	197	207	220
46	48	51	55	58	59	60	48	146	177	184	191	196	199	209	222
47	49	53	56	59	61	62	49	147	178	186	192	197	200	210	223
48	51	54	58	61	62	63	50	148	180	187	193	199	202	212	225
49	52	55	59	62	64	65	51	149	181	188	195	200	203	213	226
50	53	57	60	63	65	66	52	150	182	190	196	202	204	215	228
51	54	58	61	65	66	67	53	151	184	191	197	203	206	216	229
52	56	59	63	66	68	69	54	152	185	193	199	204	207	217	230
53	57	60	64	67	69	70	55	153	186	194	200	206	209	219	232
54	58	62	65	69	71	72	56	154	187	195	202	207	210	220	233
55	59	63	67	70	72	73	57	155	189	197	203	209	211	222	235
56	61	64	68	72	73	74	58	156	190	198	204	210	213	223	236
57	62	66	70	73	75	76	59	157	191	199	206	211	214	225	238
58	63	67	71	74	76	77	60	158	193	201	207	213	216	226	239
59	65	68	72	76	77	78	61	159	194	202	208	214	217	228	241
60	66	70	74	77	79	80	62	160	195	203	210	215	218	229	242
61	67	71	75	78	80	81	63	161	197	205	211	217	220	230	244
62	68	72	76	80	82	83	64	162	198	206	212	218	221	232	246
63	70	73	78	81	83	84	65	163	199	207	214	220	223	233	247
64	71	75	79	82	84	85	66	164	200	209	215	221	224	235	248
65	72	76	80	84	86	87	67	165	202	210	217	222	226	236	250
66	73	77	82	85	87	88	68	166	203	212	218	224	227	238	251
67	75	79	83	87	89	90	69	167	204	213	220	225	228	239	253
68	76	80	84	88	90	91	70	168	206	214	221	227	230	240	254
69	77	81	86	89	91	92	71	169	207	216	222	228	231	242	256
70	78	83	87	91	93	94	72	170	208	217	224	229	233	243	257
71	80	84	88	92	94	95	73	171	210	218	225	231	234	245	259
72	81	85	90	93	95	96	74	172	211	220	226	232	235	246	260
73	82	87	91	95	97	98	75	173	212	221	228	234	237	248	262
74	84	88	92	96	98	99	76	174	213	222	229	235	238	249	263
75	85	89	94	98	100	101	77	175	215	224	231	236	240	251	265
76	86	91	95	99	101	102	78	176	216	225	232	238	241	252	266
77	87	92	96	100	102	103	79	177	217	226	233	239	242	253	267
78	89	93	98	102	104	105	80	178	219	228	235	241	244	255	269
79	90	95	99	103	105	106	81	179	220	229	236	242	245	256	270
80	91	96	100	104	107	108	82	180	221	231	237	243	247	258	272
81	92	97	102	106	108	109	83	181	222	232	239	245	248	259	273
82	94	99	103	107	109	110	84	182	224	233	240	246	249	261	275
83	95	101	105	109	111	112	85	183	225	235	242	248	251	262	276
84	96	103	106	110	112	113	86	184	226	236	243	249	252	263	278
85	98	103	107	111	114	115	87	185	228	237	244	250	254	265	279
86	99	104	109	113	115	116	88	186	229	239	246	252	255	266	281
87	100	105	110	114	116	117	89	187	230	240	247	253	256	267	282
88	101	107	111	115	118	119	90	188	231	241	248	255	258	268	284
89	103	108	113	117	119	120	91	189	233	243	250	256	259	271	285
90	104	109	114	118	120	121	92	190	234	244	251	257	261	272	287
91	105	110	115	120	122	123	93	191	235	246	253	259	262	274	288
92	107	112	117	121	123	124	94	192	237	247	254	260	263	275	290
93	108	113	118	122	125	126	95	193	238	248	255	262	265	276	291
94	109	114	119	124	126	127	96	194	239	250	257	263	266	278	293
95	111	116	121	125	127	128	97	195	240	251	258	264	268	279	294
96	112	117	122	127	129	130	98	196	242	252	260	266			

Table 4. Comparison of *V* and *C* Regions of Fab' New for Sequence Homology by the Alignment Statistic

Sequence Pairs ^a	Number of Compared Residue Positions	MMD Values	Significance Level
$V_L - V_H$	102	98	< 0.01 %
Misaligned $V_L - V_H$	99	139	> 10 % < 50 %
$V_L - C_L$	84	97	> 0.1 % < 0.1 %
Misaligned $V_L - C_L$	82	116	> 10 % < 50 %
$V_L - C_{H1}$	82	100	> 0.1 % < 1 %
Misaligned $V_L - C_{H1}$	83	121	> 50 % < 95 %
V_L -Myoglobin	103	149	> 50 % < 95 %
$V_H - C_L$	91	111	> 0.1 % < 1 %
$V_H - C_{H1}$	89	114	> 1 % < 5 %
Misaligned $C_{H1} - V_H$	89	123	> 10 % < 50 %
V_H -Myoglobin	117	174	> 50 % < 95 %
$C_L - C_{H1}$	101	92	< 0.01 %
Misaligned $C_{H1} - C_L$	92	134	> 50 % < 95 %
C_L -Myoglobin	105	167	> 95 %
C_{H1} -Myoglobin	103	155	> 50 % < 95 %

a When V_L was used as a misaligned sequence in the comparisons its alignment was placed out of register by shifting each of its residues one position over to the right. When C_{H1} was used as a misaligned sequence, its alignment was placed out of register by shifting each of its residues two positions over to the right. Sequences were also compared to an unrelated protein, dolphin myoglobin.

acids long. The *V* region of each light chain (V_L) consist of the N-terminal half of the chain and the *C* region (C_L) consists of the C-terminal half. The *V* region of each IgG heavy chain (V_H) consists of the N-terminal quarter of the chain and is thus about the same lengths as V_L . Moreover the remaining 3 quarters of the chain can be divided into three homology domains C_{H1} , C_{H2} , and C_{H3} , each showing clear genetic relationship to C_L and each at about the same length as a *V* region. A fragment of IgG, called Fab' because it is the antigen binding fragment, consists of the two complete light chains and the N-terminal half ($V_H + C_{H1}$) of the heavy chains. The amino-acid sequences aligned by Poljak et al. (1974) were from the Fab' fragment from the IgG myeloma immunoglobulin NEW. Thus the homologies tested in Table 4 are for V_L , V_H , C_L , and C_{H1} regions of IgG.

The table shows that the probability that these sequences belong to a random collection is very low for the $V_L - V_H$ and $C_L - C_{H1}$ pairs, less than 0.01 %. While not that low for the several *V - C* pairs it is still small, falling between 1% and 5% for $V_H - C_{H1}$, between 0.1 % and 1 % for $V_L - C_{H1}$, and also for $V_H - C_L$ and falling between 0.01 % and 0.1 % for $V_L - C_L$.

Thus if we take the usual cutoff point for rejecting a null hypothesis in statistical tests, 5 % probability, we find that in all cases significant homology exists among these *V* and *C* region sequences. Moreover the probabilities are all greater than 10 % and cluster about 50 %, supporting the null hypothesis, when the sequences are misaligned or when they are compared to myoglobin, an unrelated protein.

In order to align the *V* and *C* regions against one another on the basis of matched features of three-dimensional structure, Poljak et al. (1974) interspersed the alignments with gaps and insertions. Such a procedure reduces the number of compared residues

between sequences. This actually makes it harder for these residues to pass the significance test for homology in that with a decreasing n -alignment the MMD value must decrease to a proportionately greater extent to support the hypothesis of common ancestry. For example, as Table 3 shows, at n -alignment of 100 an MMD value 1.32 times n achieves the 5 % significance level, but at an n -alignment of 10 the MMD value must be reduced to 1.0 times n to have this significance level. The structural alignment for V and C regions loses only about 15 % of its total number of residues to insertions and we see in Table 3 the n -alignments which remain easily pass the test for homology. We have also restricted our n -alignments to stretches of contiguous amino acids between gaps or insertions as the case may be, and these too pass the significance test for homology, although usually at a somewhat borderline level. For example, V_L and C_L were compared over three such uninterrupted stretches starting at V_L residue positions 1, 15, and 18 with n -alignments of 14, 17 and 24 respectively. The first of these n -alignments has MMD of 13, which achieves the 1 % significance level, but when moved out of register, or misaligned (each residue of V_L was moved one position to the left), it has an MMD value of 19, or 50 % probability of being random. The 18 n -alignment has an MMD of 19, between a 1 % and 5 % probability of being random, but misaligned its MMD is 28, a probability of randomness between 50 % and 95 %. The 24 n -alignment has an MMD of 27, a probability between 1 % and 5 %, but misaligned its MMD is 38, probability between 50 % and 95 %. We consider such results further evidence that aligned V and C region sequences in fact do show a real evolutionary relationship.

Dehydrogenases, Flavodoxin, and Subtilisin. Analogous work to that on immunoglobulin V and C regions has been done by Rossmann et al. (1974) on dehydrogenases, flavodoxin, and subtilisin. The dehydrogenases share a common structural domain whose function is to bind nicotinamide adenine dinucleotide (NAD). The same structure is utilized to bind flavin mononucleotide in flavodoxin, and it is also similar to an aromatic pocket of subtilisin. This permitted Rossmann et al. (1974) to recognize corresponding amino acids when sequence comparisons alone would fail. Their best alignment was found by comparing a particular stretch of 37 residues (residues 22–58) representing the most conserved part of the adenine binding pocket in lactate dehydrogenase (LDH) to corresponding residues in the other proteins, glyceraldehyde-3-phosphate dehydrogenase (GAPDH), liver alcohol dehydrogenase (LADH), glutamate dehydrogenase (GluDH), flavodoxin, and subtilisin. As can be seen in Table 5 these comparisons pass the alignment statistic test for homology. This supports the claim of Rossmann et al. that the conserved structures in these proteins are evolutionarily related.

Heme-Binding Pocket in Globins and Cytochrome b_5 . Rossmann and Argos (1975) have also used their method of comparing the three-dimensional structures of folded polypeptide chains to investigate the possible evolutionary derivation of cytochrome b_5 and the globins from a common primordial heme-binding protein. They found that up to 52 residues of 85 three-dimensionally characterized residues in calf liver cytochrome b_5 are structurally and topologically equivalent to the globin fold in horse oxyhemoglobin. On determining minimum base changes (or MMD values in our terminology) for the numbers of "equivalenced" residues in their best "fits" between

Table 5. Comparison of the most conserved part of LDH to corresponding residues on other dehydrogenases, flavodoxin, and subtilisin.

Sequence Pair ^a	Number of Compared Residue Positions	MMD Value	Significance Level	
Dogfish LDH-Yeast GAPDH	37	40	0.1 %	
Misaligned Dogfish LDH-Yeast GAPDH	36	50	> 10 %	< 50 %
Dogfish LDH-Horse LADH	36	38	> 0.01 %	< 0.1 %
Misaligned Dogfish LDH-Horse LADH	35	49	> 0 %	< 50 %
Dogfish LDH-Bovine GluDH	36	43	> 1 %	< 5 %
Misaligned Dogfish LDH-Bovine GluDH	35	46	> 10 %	< 50 %
Dogfish LDH- <i>Clostridium</i> flavodoxin	33	39	> 1 %	< 5 %
Misaligned Dogfish LDH- <i>Clostridium</i> flavodoxin	32	48	> 50 %	< 95 %
Dogfish LDH-subtilisin	36	41	> 0.1 %	< 1 %
Misaligned Dogfish LDH-subtilisin	35	46	> 10 %	< 50 %

a When dogfish LDH was used as a misaligned sequence in these comparisons, its alignment was placed out of register by shifting each of its residues one position over to the left

the two proteins, they felt that these mutation values were in the range expected for evolutionarily related proteins, but still not low enough to prove it.

In the six best structural fits, recorded in Table 1 of Rossman and Argos (1975), there were 29, 52, 51, 40, 46 and 48 equivalenced residues yielding MMD values of 36, 67, 70, 50, 60, and 62 respectively. According to our alignment statistic these MMD values are indicative of the match between evolutionarily unrelated or random polypeptide chains. Only the N-alignment of 40 equivalenced residues with an MMD value of 50 achieves the 5 % significance level, i.e. borderline evidence for significant amino acid homology. Otherwise the N-alignments show probabilities of randomness of 10 % or greater. Thus a better case might be made that the similarity the three-dimensional structure between the heme binding pocket in globins and cytochrome *b₅* was produced by evolutionary convergence rather than derived by common inheritance from some primordial heme binding protein. It is worth emphasizing that our alignment statistic is apparently quite discriminating in providing evidence for evolutionary homology when applied to proposed structural alignments.

4. Further Empirical Support for the Statistic

It is unlikely that all conceivable *n*-alignments are equally probable, because, regardless of ancestry, certain conceivable but structurally ill-conditioned polypeptide chains could never be expressed in a living organism. Thus the derivation of our alignment statistic is not strictly valid. On the other hand, it appears that the *proportions* of 0-differences, 1-differences, 2-differences, and 3-differences (6 %, 46 %, 46 %, and 2 %, respectively)

respectively) match the proportions seen in observed, non-aligned sequences. For example, when V_L and C_L chains are one position out of alignment, we obtain difference proportions of 9 %, 45 %, 42 % and 5 % for the resulting 82 amino acid pairs. As another example, when dogfish LDH and yeast GAPDH, at the most conserved part of the adenine binding pocket, are one position out of alignment we obtain difference proportions of 8 %, 44 %, and 3 % for the resulting 36 pairs. If we combine the data on these two examples, the difference proportions are 8 %, 45 %, 42 %, and 4 % for the resulting 118 amino acid pairs.

The assumption of *independence* of each alignment position to its neighbor is also unlikely on the basis of structural considerations, but again the predicted and actual *proportions* may not be very dissimilar. If we consider sequential *pairs* of residues in the previous examples, we obtain 0-difference, 1-difference, . . . , and 6-difference values of 4 %, 7 %, 27 %, 38 %, 20 %, 4 %, and 0 % respectively for the resulting 55 amino acid pairs. The predicted values (Table 2) are 0 %, 6 %, 27 %, 42 %, 23 %, 2 %, and 0 %, respectively.

Another criticism of the derivation of our alignment statistic is its assumption that no systematic bias enters in the selection of an alignment. In many insertion/deletion problems, the investigator may slide the amino acid residue gap in an effort to minimize the observed MMD value. Obviously, the introduction of an arbitrary number of arbitrary-sized deletions and insertions can be used to lower the observed MMD to any desired value, thus destroying the sense of the test. A simulated distribution of MMD values was generated for every n -alignment with $n \leq 12$ and every gap-length $n - 1$, where a *single gap* was permitted to slide along the alignment and settle in position with lowest MMD value. In 5000 Monte Carlo trials apiece, the simulated significance table was never more than one mutation less than predicted in Table 3. Simulation experiments are open to the criticism that they are not exhaustive trials, but our experience suggests that manipulation of insertions and deletions in moderation may not substantially bias the evaluation of significance levels.

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