

Concurrence of cortex and platelet serotonin₂ receptor binding characteristics in the individual and the putative regulation by serotonin*

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Summary. Changes in the central and peripheral serotonergic receptor activity have been reported to be involved in depression and suicidality. To elucidate the interdependence between central and peripheral receptor sites and their regulation by serotonin, we estimated intra-individual serotonin₂ receptor binding characteristics in porcine cortex synaptosomes and in platelet membranes using ³H-LSD as ligand and ketanserin as competitor and quantified the relevant serotonin concentrations. A positive correlation between the apparent half maximal saturation concentration, K_D , of the receptor in cortex synaptosomes and platelet membranes ($r = 0.65$, $p = 0.0046$, $n = 18$), and between the apparent maximal binding capacity, B_{max} , of the receptor in cortex synaptosomes and platelets ($r = 0.52$, $p = 0.027$, $n = 18$) was observed. The blood serotonin concentrations correlated negatively with the maximal binding capacity, B_{max} , in platelets ($r = -0.77$, $p = 0.0002$, $n = 18$). These results suggest that the binding characteristics of the central and peripheral serotonin₂ receptor are similar, and that the platelet receptor activity may be regulated by blood levels of serotonin.

Keywords: Serotonin₂ receptor binding characteristics, serotonin concentrations, brain cortex synaptosomes, platelet membranes, intra-individual correlation, pig.

Introduction

The serotonin₂ receptor has been pharmacologically characterized in the mammalian frontal cortex and blood platelets. The turnover of serotonin by platelets

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is considered to serve as peripheral model of central serotonergic neurons (Sneddon, 1973; Stahl and Meltzer, 1978). Alterations of the activity of the serotonin₂ receptor in both tissues have been implicated in depression and aggressive behaviour (Arora and Meltzer, 1989; Aprison et al., 1978; Mann et al., 1986; McKeith et al., 1987; Pandey et al., 1990; Stanley and Mann, 1983; Yates et al., 1990). However, little is known with respect to in vivo finding concerning the regulatory influence of the endogenous concentrations of the ligand serotonin on serotonin receptors.

Despite the postulate that platelets serve as a peripheral model for brain serotonergic neuron function intra-individual studies using an identical protocol to compare serotonin₂ receptor binding characteristics in both tissues have not been carried out. To examine these issues we studied intra-individual brain and platelet serotonin₂ receptor binding characteristics and related these to their respective blood serotonin concentrations.

Materials and methods

The present study was carried out with brain specimen and blood samples obtained from 18 German female pigs from the local abattoir immediately after sacrifice. Wherever applicable, the procedures were carried out using an identical protocol.

Brain cortex synaptosomes

Porcine (Deutsche Landrasse) brain (n = 18) was transported on ice from the local abattoir to the laboratory. The frontal cortex (3–5 g) was dissected ventrally of the chiasma opticum on the right hemisphere. All procedures were carried out at 4 °C. The synaptosomes were prepared according to Creese and Snyder (1977). The brain cortical tissue was cut into small pieces, suspended and homogenized in 80 ml of buffer I (50 mmol/L Tris-HCl, pH 7.7). The homogenate was sonicated in ice-water for 45 s with a Braun sonicator (Sonic 300 S, Braun, Melsungen, Germany) and filtered through 45 µ nylon gauze. The filtrate was centrifuged for 20 min at 20 000 g (MSE Superspeed 75, Measuring Scientific Equipment LTD, London, England). The sediment was suspended in buffer I, homogenized and centrifuged twice as described above. The final pellet was resuspended and homogenized in buffer II (50 mmol/L Tris-HCl, pH 7.4, containing 120 mmol/L NaCl, 5 mmol/L KCl, 1 mmol/L MgCl₂ and 0.05% ascorbic acid), resulting in 200 mg wet weight/ml. The synaptosomes were stored at –80 °C until assayed for receptor studies.

Platelet membranes

We collected also from each animal 100 ml blood into siliconized glass tubes containing 5 ml 5.5 mmol/L EDTA. All subsequent procedures were carried out at 4 °C. The blood was centrifuged after 45 min for 15 min at 200 g (Minifuge 2, Heraeus Christ, Osterode, Germany) to obtain platelet-rich plasma. The supernatant was centrifuged for 10 min at 4000 g to prepare platelet-poor plasma and platelets. Platelet membranes were prepared by the method of Pandey et al. (1990). In brief, the platelets were suspended in buffer III (5 mmol/L Tris-HCl, containing 0.1% EDTA, pH 7.5) and centrifuged for 10 min at 4000 g. The sediment was resuspended in 4 ml of buffer III and stored at –80 °C until used for binding studies. Prior to the assay the platelet membranes were thawed on ice, homogenized in a loose-fitting Potter Elvehjem glass homogenizer (Braun, Melsungen, Germany) and centrifuged for 20 min at 30 000 g. The membrane pellet was suspended in buffer III,

homogenized as described above and centrifuged for 20 min at 30 000 g. The final membrane pellet was suspended in 3.5 ml of buffer II, homogenized and sonicated for 25 s in ice-water.

The final protein concentration of the synaptosomal and platelet membrane preparations was adjusted to 1 mg protein/ml.

Platelets were counted in a Neubauer chamber; the results were validated by external quality control.

Binding assay

The serotonin₂ receptor binding assays were carried out in triplicate in siliconized glass tubes, with 150 μ l buffer II and 100 μ l membrane suspension, corresponding tubes to 100 μ g protein. The reaction was started with the addition of 50 μ l ³H-LSD in 6 different final concentrations ranging from 0.08 to 5.0 nmol/L corresponding to 1700–100 000 cpm. The final volume of the incubation was 300 μ l. Nonspecific binding was determined by replacing 50 μ l buffer II with a non-radioactive competitor, the serotonin₂ receptor antagonist, ketanserin in buffer II in a final concentration of 1 μ mol/L. Specific binding was the difference between the binding in the absence and presence of ketanserin. Incubations were performed for 3 h at 37 °C. The incubation was terminated by rapid filtration through Whatman GF/C filters; the filters were washed twice with ice-cold buffer II and counted in 5 ml Aquasafe 300 (Zinsser Analytic, Frankfurt, Germany) in a liquid scintillation beta-counter (Packard, Frankfurt, Germany).

Protein determination

Protein was determined with a modified method of Lowry et al. (1951) using bovine serum albumin as standard.

Quantification of serotonin

Serotonin was measured by high pressure liquid chromatography using electrochemical detection, in blood, platelet rich plasma, and frontal cortex homogenate (Rao and Fels, 1987). The intra- and interassay coefficients of variation were 2.9 and 4.4%, respectively. The lower limit of detection was 5 nmol/L.

Statistics

All experiments were carried out in triplicate. The average coefficient of variation over the entire concentration range of the binding experiment was $4.9 \pm 2.6\%$ (mean \pm SD). Binding parameters were analyzed according to Scatchard (1949) using linear regression analysis; the average correlation coefficient r was -0.95 ± 0.05 and -0.95 ± 0.04 , ($n = 18$) for the binding experiments with synaptosomes and platelet membranes, respectively. Linear regression analysis (Pearson's linear correlation coefficient) was performed to assure correlations.

Results

Serotonin₂ receptor binding in porcine frontal cortex synaptosomes and platelet membranes was saturable and possessed high affinity (Table 1). From the time and temperature curve we inferred that the specific binding was highest at 37 °C, the equilibrium was attained after 3 h at 37 °C. The Scatchard plots of the specific binding were linear, indicating the presence of a single binding site in both tissue preparations in the concentration range between 0.08 to 5 nmol/L ³H-LSD (Fig. 1 A and B).

Table 1. Binding characteristics of the serotonin₂ receptor and serotonin concentrations (n = 18)

Parameter	Mean ± SD	Range
<i>Brain cortex</i>		
Serotonin ₂ receptor, synaptosomes		
B _{max} (fmol/mg protein)	209.1 ± 60.6	122.7 – 322.5
K _D (nmol/L)	0.99 ± 0.64	0.44 – 2.54
Serotonin, homogenate (nmol/g wet weight)	0.832 ± 0.53	0.344 – 1.86
<i>Blood</i>		
Serotonin ₂ receptor, platelet membranes		
B _{max} (fmol/mg protein)	77.8 ± 32.9	43.6 – 159.4
K _D (nmol/L)	0.56 ± 0.37	0.29 – 1.7
Serotonin, blood (μmol/L)	3.65 ± 1.26	1.25 – 6.05
Serotonin (nmol/10 ⁹ platelets)	14.35 ± 3.90	7.85 – 19.95
Platelet count (μl ⁻¹)	253000 ± 54088	119000 – 331000

At a concentration of 0.2 nmol/L ³H-LSD the specific binding of the receptor in brain cortex synaptosomes was 67 ± 9% (n = 18) of the total binding and 76 ± 12% (n = 18) using platelet membranes. At the lowest concentration used, i.e., at 0.08 nmol/L ³H-LSD, the total binding amounted to 9.8 ± 3.9% and 6.3 ± 2.7% of the radioactivity incubated with brain cortex synaptosomes and platelet membranes, respectively.

The apparent maximal binding capacity, B_{max}, and the apparent half maximal saturation concentration, K_D, of the receptor in brain cortex synaptosomes and platelet membranes displayed a bimodal distribution in that both parameter estimates were higher in the brain than in platelet membranes. There was no interdependence between the apparent maximal binding capacity and the half maximal saturation concentration of the serotonin₂ receptor in either tissue.

The maximal binding capacity, B_{max}, of the serotonin₂ receptor in brain cortex synaptosomes and in platelet membranes correlated (r = 0.52, p = 0.027, n = 18); this applied also to the receptor's half maximal saturation concentration, K_D, in both tissues (r = 0.65, p = 0.0046, n = 18). Blood and cortex homogenate serotonin concentrations showed an inhomogeneous correlation (r = -0.49, p = 0.04, n = 18). The cortex homogenate serotonin concentrations did not correlate with the synaptosomal receptor's maximal binding capacity. However, we noted a linear correlation between blood serotonin levels and the

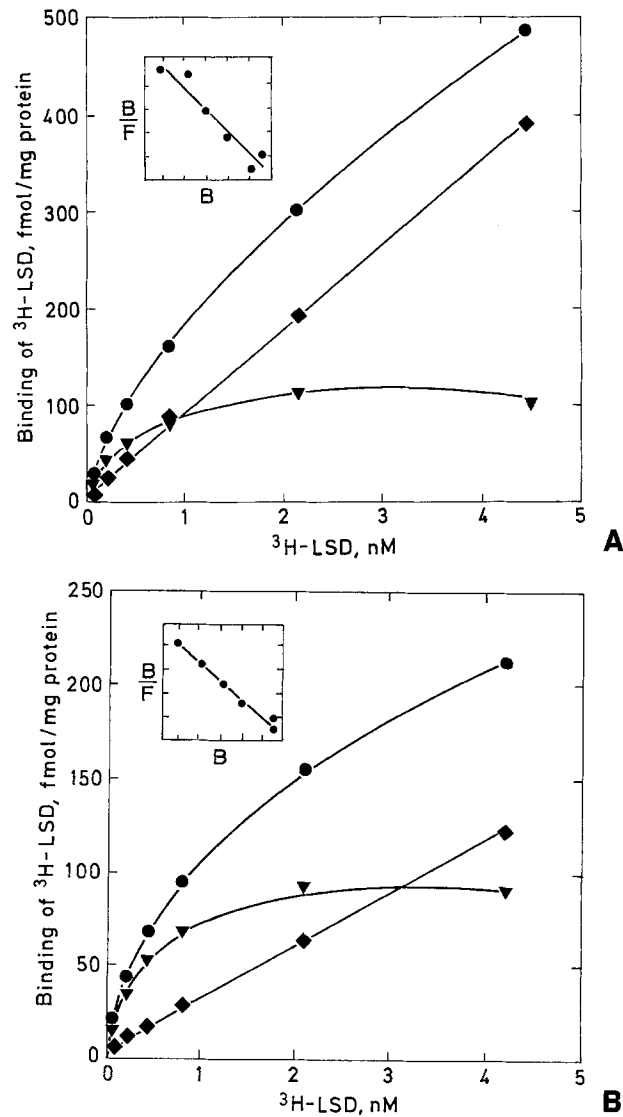


Fig. 1. Binding of ³H-LSD to serotonin₂ receptors in **A** porcine frontal cortex synaptosomes and **B** porcine platelet membranes. ● Total binding; ◆ unspecific binding; ▼ specific binding

maximal binding capacity of the receptor of platelet membranes ($r = -0.77$, $p = 0.0002$, $n = 18$, Fig. 2).

Discussion

The mammalian frontal cortex comprises the highest serotonin₂ receptor density in the brain (Leysen et al., 1981; Engel et al., 1984; Hoyer et al., 1986; Cheetham et al., 1988). Serotonin₂ receptors on serotonergic neurons and platelets are coupled to the phosphoinositol pathway (Conn and Sanders-Bush, 1986; Mikuni

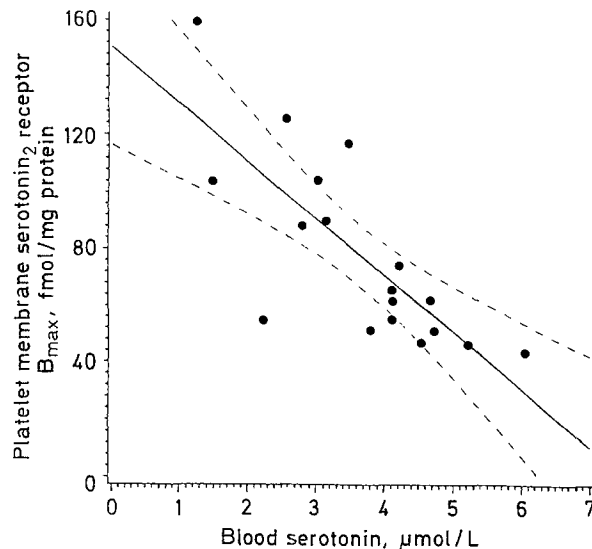


Fig. 2. Correlation of the apparent maximal binding capacity, B_{max} , of the serotonin₂ receptor in platelet membranes with blood serotonin concentrations. Dashed lines indicate the 95% confidence interval

et al., 1991). It has been shown that ^{125}I -LSD binds to human cortex ($K_D = 0.35 \pm 0.02 \text{ nM}$) and platelets ($K_D = 0.37 \pm 0.03 \text{ nM}$) with similar affinity. Moreover, the inhibition profiles correlated between these compartments (Elliott and Kent, 1989), but these studies were not carried out within an intra-individual set-up.

We chose the pig as animal model since porcine and human brain display similar pharmacological profiles of the serotonin₂ receptors (Hoyer et al., 1986). Since it is controversial whether lateralisation of the receptor exists (Yates et al., 1991; Arató et al., 1991), we decided to use the right hemisphere. On account of the size of porcine brain, the cortex synaptosomes of an individual can be prepared in sufficient quantity to estimate the binding characteristics of the serotonin₂ receptor. Moreover, from the same animal adequate amounts of blood can be obtained to prepare platelet membranes; this permits the intra-individual comparison of the serotonin₂ receptor's activity from frontal cortex and platelet membranes.

The salient findings of this study on central and peripheral serotonin₂ receptors are the positive correlations observed between the apparent half maximal saturation concentration, K_D , of the serotonin₂ receptor in cortex synaptosomes and platelet membranes, and the apparent maximal binding capacity, B_{max} , of brain cortex synaptosomes and platelet membranes. In addition we noted that blood serotonin concentrations correlate negatively with the B_{max} -values of the serotonin₂ receptor of the platelet membranes. This suggests that serotonin is a regulatory factor in that it causes a down-regulation of the serotonin₂ receptor at high endogenous concentrations in the periphery. A similar interdependence

between the serotonin concentration and the change in maximal binding capacity of the serotonin₂ receptor was not found in the frontal cortex. This could be due to the inhomogeneous distribution of serotonin in brain tissue, since for technical reasons the brain tissue used for the determination of serotonin and for the isolation of the synaptosomes was not identical.

On the other hand, the regulation of the serotonin₂ receptor in mammalian brain seems to be unique, in that denervation or chronic blockade do not provoke receptor supersensitivity or up-regulation in mammalian brain (Leysen et al., 1983, 1986; Conn and Sanders-Bush, 1987). Moreover, a down-regulation of the serotonin₂ receptor is perceptible after antagonist treatment, i.e., with ketanserin (Leysen und Pauwels, 1990) as well as after agonist (methoxyphenethylamine) treatment (McKenna et al., 1989). Taken together these results do not appear to be consistent with the generally accepted hypothesis on receptor regulation according to which for example β -adrenergic and cholinergic muscarinergic receptors are supersensitive following stimulus deprivation and are desensitized after agonist treatment (Leysen, 1990). Leysen (1990) speculated that under physiological conditions the serotonin₂ receptor might receive little stimulation and hence exists in a state of supersensitivity that permits solely a one-way down-regulation in emergencies.

Serotonin₂ receptor regulation may proceed disparately during distinct stages of receptor synthesis and receptor life. There is evidence, that alterations in the receptor level are due to changes on the transcriptional level, i.e., increased mRNA levels (Roth et al., 1990 a). On the other hand, following mianserin-treatment, a dissociation between receptor and mRNA levels was noted which points to post-transcriptional regulation. In addition post-translational regulatory mechanisms may occur such as receptor internalization and degradation, as well as modifications on account of receptor phosphorylation, glycosylation or proteolysis (Roth et al., 1990 b). Overall changes in membrane fluidity may also be responsible for alterations in the maximal binding capacity (Heron et al., 1980). In our study, increasing blood serotonin concentrations seem to down-regulate the maximal binding capacity, B_{max} , of the serotonin₂ receptor on platelets. The underlying mechanism is not clear; the receptor might be internalized when endogenous serotonin levels are high, but any of the above mentioned mechanisms could be of importance.

We interpret our findings of similar receptor binding characteristics in brain and on peripheral blood cells in that the expression of the serotonin₂ receptor might be determined by similar – yet unknown – transcriptional, post-transcriptional or post-translational events in the brain cortex and in platelets. The fact that the maximal binding capacity and the half maximal saturation concentration of the serotonin₂ receptor in frontal cortex and in platelets show bimodal distribution might indicate that there are also independent regulatory mechanisms. The intra-individual correlation between the B_{max} of the synaptosomal and corresponding platelet receptors and between the K_D s supports the notion that platelets serve as an experimental model to assess the function

of central serotonergic neurons. Thus, findings of alterations of the platelet serotonin₂ receptors in psychiatric diseases might provide information on the function of frontal cortex serotonin₂ receptor.

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