Mycoflora of the toxic feeds associated with equine leukoencephalomalacia (ELEM) outbreaks in Brazil

M.C.A. Meireles,¹ B. Corrêa,² O. Fischman,³ W. Gambale,² C.R. Paula,² N.O. Chacon-Reche² & C.R. Pozzi⁴

¹Faculdade de Veterinária, Universidade Federal de Pelotas (UFPel), Pelotas, RS, Brasil; ²Seção de Micologia, Departamento de Microbiologia, Instituto de Ciências Biomédicas, Universidade de São Paulo (ICB-USP), São Paulo, SP, Brasil; ³Setor de Micologia, Disciplina de Biologia Celular, Escola Paulista de Medicina (EPM), São Paulo, SP, Brasil; ⁴Instituto de Zootecnia (IZ), Secretaria de Agricultura do Estado de São Paulo, Nova Odessa, SP, Brasil

Received 12 October 1993; accepted in revised form 4 June 1994

Abstract. The mycoflora of 39 feed samples associated with 29 Equine Leukoencephalomalacia (ELEM) outbreaks was studied from 1988 to 1990, in Brazil. Microbiological examination indicated *Fusarium* spp. as the most frequent mold which occurred in 97.4% of samples followed by *Penicillium* spp. in 61.5% and *Aspergillus* spp. in 35.9%. The moisture content of feed implicated in death of horses was above 15% which can favor the development of *Fusarium* spp. From the genus, *F. moniliforme* was the predominant species with an occurrence of 82.0%. Two additional species, not commonly associated with animal toxicosis, were isolated in low frequency, *F. proliferatum* (12.8%) and *F. subglutinans* (2.6%). It is important to emphasize that the isolation of *F. proliferatum* and *F. subglutinans* from feed obtained from the epizootic areas has not been documented previously in Brazil.

Key words: Equine, Feeds, Fusarium proliferatum, Fusarium subglutinans, Leukoencephalomalacia, Mycoflora

Introduction

Equine leukoencephalomalacia (ELEM), is a neurotoxic disease of equidae, characterized by multifocal liquefactive necrosis of the white matter in one or both cerebral hemispheres. It is a seasonal mycotoxicosis, most common when a hot and dry season is followed by a wet and cold period [1]. The syndrome has been associated with the consumption of feed contamined with fumonisins (FB₁ and FB₂), produced especially by *Fusarium moniliforme* and probably by other species of the genus [2–4]. *F. moniliforme* has been the predominant fungus isolated from moldy feed causing outbreaks of ELEM in the USA [5–7], Republic of South Africa [8–10], Egypt [11], New Caledonia [12], Argentina [13], China [14] and

Part of the thesis of M.C.A. Meireles to get the degree of Doctor 'Leucoencefalomalácia Equina (LEME) no Brasil: Aspectos epizootiológicos, microbiológicos e micotoxicológicos', Preliminary communication in 1989, Congresso Brasileiro de Microbiologia, Rev Microbiol (São Paulo) 20 (Supl. 1): 359 (F 181). Brazil [15–19] and has also been noticed in Greece and probably in Germany [5].

In 1971, Wilson & Maronpot [20] experimentally reproduced the typical disease and established toxigenic *F. moniliforme* as the fungal agent responsable for the mycotoxicosis. ELEM was induced by the oral administration or intravenous injection of FB₁ produced by *F. moniliforme* to horses [21–23]. Other species of the genus, *F. proliferatum* and *F. sub-glutinans*, have been also associated with outbreaks of ELEM and Porcine Pulmonary Edema (PPE) [3, 4, 24].

This paper reports the mycoflora of 39 feed samples associated with 29 ELEM outbeaks from four Brazilian states.

Material and methods

Equine feed samples. Each sample associated with ELEM outbreaks was packaged in paper sacs, approximately 1 kg of each one, identified and transport-

Type of feed	Geographic location (states)				
	R.G. do Sul	São Paulo	S. Catarina	Minas Gerais	
Corn	15	2	2	1	20
Ground corn & straw	3	4	_	1	8
Ground corn	4	2		-	6
Commercial pellets	a	2	_	~	2
Oats	1	_	_	-	1
Straw	_	1	<u> </u>	-	1
Rice grass	1	-	-	~	1
Total	24	11	2	2	39

Table 1. Geographic location and type of 39 feed samples

^aSample not taken.

ed to the laboratory. The 39 samples were obtained from farms in four Brazilian states (1988–1990): Rio Grande do Sul (24), São Paulo (11), Santa Catarina (2) and Minas Gerais (2) (Table 1).

Determination of mycoflora according to Busta et al. [25]. Fungi were determined by blending a 10 g portion of each sample in 90 ml of phosphate buffered saline (PBS). Serial dilutions, until a 10^{-5} concentration, was made from each material: 1 ml of each dilution was spread on each of two Sabouraud glucose agar (pH 5.6) plates, containing chloramphenicol (100 μ g/ml) and sodium azide (300 μ g/ml). The plates were incubated for 7 days at 25 °C and observed daily. Fungal colonies were selected for subculturing and identified according to the methods for each genus [26–28].

Results and discussion

From 1988 to 1990, 29 ELEM outbreaks were studied in four Brazilian states: Rio Grande do Sul (13), São Paulo (12), Santa Catarina (2) and Minas Gerais (2) (Table 2 and Fig. 1). Approximately a hundred purebred horses died, most within 12 hours after clinical signs appeared. The syndrome was characterized clinically by acute death with neurological signs. All of the necropsied horses showed focal to diffuse uni or bilateral areas of liquefative necrosis of cerebral white matter. The animals had been maintained in confinement.

Outbreaks of ELEM are usually seasonal. Those reported here occurred from late fall through early

spring and were most common in winter (June, July, August and September months). (Table 2 and Fig. 1).

In Brazil, most authors [15, 17, 18] correlate ELEM with low temperatures without taking into consideration other climatic factors except Barros *et al.* [16] emphazised the role of pre-harvest rainfall. Moisture is important to the growth of *Fusarium* spp. and colonization is recognized by a pink to reddish brown color overlying the grain. In 75% of the cases, the moisture content of the corn kernels was above 15% which can favor the development of *Fusarium* spp. Corrêa *et al.* [29] reported moisture content higher than 16% in stored maize, contaminated with *Fusarium* spp. in Brazil.

Mycological examination of 39 feed samples indicated the presence of 3 genera of filamentous fungi shown in Table 3 and Fig. 2. *Fusarium* spp. was the most frequent mold which occurred in 97.4% of samples followed by *Penicillium* spp. in 61.5% and *Aspergillus* spp. in 35.9%. Colony counts of *Fusarium* spp. ranged from 8×10^3 to 6.7×10^9 colonyforming units (CFU) per g. These are high values when they are compared with those of the International Commission of Microbiological Specifications for Commission Foods.

Maize is frequently colonized by *Fusarium* spp. Inadequately stored grains, moisture content above 15%, and low temperatures after harvesting are probably the ideal conditions for mold colonization [5, 30-32].

F. moniliforme was the most frequent mold (82.0%) (Table 4 and Fig. 3) which agrees with foreign and Brazilian authors [2, 3, 6, 7, 17, 18, 29, 32]. This fungus colonized all the 39 samples, except one.

Month	R. (G. do Sul	São	Paulo	S. C	atarina	Min	as Gerais	Tota	1
	N	(%)	N	(%)	N	(%)	N	(%)	N	(%)
Jan	_a	_	-	_	_	-	-			-
Feb	_	_		-	_		-	-	-	-
Mar	-	-	-	-	_	-		-	-	-
Apr	_	-		-			-		-	-
May	-	-		_	_	-			-	
Jun	1	7.7	3	25.0	-		1	50.0	5	17.2
Jul	6	46.2	4	33.3	-	-	-		10	34.5
Aug	3	23.1	-	-	1	50.0	1	50.0	5	17.2
Sep	3	23.1	2	16.7	1	50.0	-	-	6	20.7
Oct		-	2	16.7		-	-		2	6.9
Nov	_	-	1	8.3	-		_	-	1	3.5
Dec		-	-	-	-	-	-	_	-	-
Total	13	44.8	12	41.4	2	6.9	2	6.9	29	100.0

Table 2. Monthly distribution of 29 ELEM outbreaks, from 1988 to 1990, in four Brazilian states

^a- No outbreaks.

N, Number of outbreaks; (%): Relative frequency.



Fig. 1. Seasonal distribution of 29 ELEM outbreaks in Brazil from 1988 to 1990. Brazilian states: RS, Rio Grande do Sul; SP, São Paulo; SC, Santa Catarina; MG, Minas Gerais.

Table 3. Frequency of filamentous fungi isolated from 39 feed samples associated with 29 ELEM outbreaks in Brazil, from 1988 to 1990

Filamentous fungi	Absolute frequency	Relative frequency (%)
Fusarium spp.	38	97.4
Penicillium spp.	24	61.5
Aspergillus spp.	14	35.9



Fig. 2. Distribution of mycoflora in 39 feed samples associated with 29 ELEM outbreaks in Brazil from 1998 to 1990.



Fig. 3. Frequency of Fusarium spp. isolated from 39 feed samples associated with 29 ELEM outbreaks in Brazil.

Table 4. Frequency of Fusarium species, isolated from 39 feed samples associated with 29 ELEM outbreaks in Brazil, from 1988 to 1990

Fusarium species Isolated	Absolute frequency (F)	Relative frequency (%)		
F. moniliforme	32	82.0		
F. proliferatum	5	12.8		
F. subglutinans	1	2.6		
Absence of Fusarium spp.	1*	2.6		
Total	39	100.0		

*Rice grass sample.

The rice grass (*Echinochlo* sp.) sample was colonized only by *Aspergillus* sp. Other *Fusarium* species in the samples were *F. proliferatum* (12.8%) and *F. subglutinans* (2.6%) (Table 4 and Fig. 3), which are species not as commonly associated with the toxicosis [2–4, 24]. Besides the *F. moniliforme*, *F. proliferatum* is an important fumonisin producer [3, 4, 24]. One *F. subglutinans* isolated from a culture collection did not produce any detectable fumonisin [4].

Fusarium toxins are normally produced at low temperatures. The termal shock is apparently necessary to induce biosynthesis [34, 35]. *F. moniliforme* and *F. proliferatum* produced FB₁ and FB₂ in vitro, when cultivated on sterile maize. After the initial growth at 25–27 °C, fumonisin production was induced with a thermal shock of 15 °C [3, 36].

In Brazil only *F. moniliforme* isolated from feed has been associated with ELEM [15–18, 37]. The isolation of *F. proliferatum* and *F. subglutinans* has not been previously documented in Brazil from feed samples implicated as the causative fungi of the ELEM.

Acknowledgements

The authors wish to acknowledge Dr Paul E. Nelson, Department of Plant Pathology, College of Agriculture, Fusarium Research Center, The Pennsylvania State University (USA), for help in classification of the isolates of Fusarium species. This work was supported in part by CNPq (Conselho Nacional de Desenvolvimento Científico e Tecnológico), CAPES (Coordenação e Aperfeiçoamento de Pessoal de Nivel Superior) and FAPESP (Fundação de Amparo á Pesquisa do Estado de São Paulo).

References

- McCue PM. Equine leukoencephalomalacia. The Compendium (Equine) 1989; 11: 646–51.
- Meireles MCA, Corrêa B, Purchio A, Fischman O, Gambale W, Paula CR. Microbiota fúngica em alimentos envolvidos com quadro de leucoencefalomalácia eqüina (LEME), ocorridos no ano de 1988. Rev Microbiol (São Paulo) 1989; 29(Supl. 1): 359.
- Ross PF, Nelson PE, Richard JL, Osweiler GD, Rice LG, Plattner RD, Wilson TM. Production of fumonisins by *Fusarium* moniliforme and *Fusarium proliferatum* isolates associated with equine leukoencephalomalacia and a pulmonary edema syndrome in swine. Appl Environ Microbiol 1990; 56: 3225– 6.
- 4. Thiel PG, Marasas WFO, Sydenham EW, Shephard GS, Gelderblom WCA, Nieuwenhuis JJ. Survey of fumonisin pro-

duction by *Fusarium* species. Appl Environ Microbiol 1991; 57: 1089–93.

- Wilson BJ, Maronpot RR, Hildebrandt PK. Equine leukoencephalomalacia. J Am Vet Med Assoc 1973; 163: 1293–5.
- Vesonder RF, Haliburton J, Golinski P. Toxicity of field samples and *Fusarium moniliforme* from feed associated with equine-leukoencephalomalacia. Arch Environ Contam Toxicol 1989; 18: 439–42.
- Wilson TM, Nelson PE, Marasas WFO, Thiel PG, Shephard GS, Sydenham EW, Nelson HA, Ross PF. A mycological evaluation and in vivo toxicity evaluation of feed from 41 farms with equine leukoencephalomalacia. J Vet Diagn Invest 1990; 2: 352–4.
- Kellerman TS, Marasas WFO, Pienaar JG, Naudé TW. A mycotoxicosis of equidae caused by *Fusarium moniliforme* Sheldon: A preliminary communication. Onderstepoort J Vet Res 1972; 39: 205–8.
- Pienaar JG, Kellerman TS, Marasas WFO. Field outbreaks of leukoencephalomalacia in horses consuming maize infected by *Fusarium verticillioides (Fusarium monilifome)* in South Africa. J South African Vet Assoc 1981; 52: 21–4.
- Kellerman TS, Coetzer JAW, Naudé TW. Plant poisonings and mycotoxicoses of livestock in Southern Africa, 2. ed. Cape Town: Oxford University Press, 1990: 75–82.
- Badiali L, Abou-Youssef MH, Radwan AI, Harndy FM, Hildebrandt PK. Moldy corn poisoning as the major cause of an encephalomalacia syndrome in Egyptian equidae. Am J Vet Res 1968; 29: 2029–35.
- Laurent D, Platzer N, Kohler F, Sauviat MP, Pellegrin F. Macrofusine et micromoniline: Deux nouvelles mycotoxines isolées de maïz infesté par *Fusarium moniliforme* Sheld. Microbiol Aliment Nutr 1989; 7: 9–16.
- Monina MI, Mascotena EA, Ruager J, Idiart JR, Reinoso EH, Muro A, Nosetto EO, Pons ER Leucoencefalomalacia equina: Casos registrados en el país. Rev Mil Vet 1981; 28: 13–17.
- Xin DY. Study on the aetiology of equine mouldy corn poisoning. Acta Vet Zootech Sinica 1987; 18: 125–8.
- Riet-Correa F, Meireles MCA, Soares JM, Machado JJ, Zambrano AFH. Leucoencefalomalácia em eqüinos associado à ingestão de milho mofado. Pesq Vet Bras 1982; 2: 27–30.
- Barros CSL, Barros SS, Dos Santos MN, Souza MA. Leucoencefalomalácia em equinos no Rio Grande do Sul. Pesq Vet Bras 1984; 4: 101–7.
- Hirooka EY, Viotti NMA, Marochi MA, Ishii K, Ueno Y. Leucoencefalomalácia em eqüinos no norte do Paraná. Rev Microbiol (São Paulo) 1990; 21: 223–7.
- Xavier JG, Brunner CHM, Sakamoto M, Corrêa B, Fernandes WR, Dias JLC. Equine leukoencephalomalacia: Report of five cases. Braz J Vet Res Anim Sci (São Paulo) 1991; 28: 185–9.
- Sydenham EW, Marasas WFO, Shephard GS, Thiel PG, Hirooka EY. Fumonisins concentrations in Brazilian feeds associated with field outbreaks of animal mycotoxicoses. Agric Food Chem. 1992; 40: 994–7.
- Wilson BJ, Maronpot RR. Causative fungus agent of leucoencephalomalacia in equine animals. Vet Rec 1971; 88: 484–6.
- Marasas WFO, Kellerman TS, Gelderblom WCA, Coetzer JAW, Thiel PG, Van Der Lugt JJ. Leukoencephalomalacia in a horse induced by fumonisin B₁ isolated from *Fusarium moniliforme*. Onderstepoort J Vet Res 1988; 55: 197–203.
- Kellerman TS, Marasas WFO, Thiel PG, Gelderblom WCA, Cawood M, Coetzer JAW. Leukoencephalomalacia in two horses induced by oral dosing of fumonisin B₁. Onderstepoort J Vet Res 1990; 57: 269–75.

- Wilson T M, Ross PF, Owens DL, Rice LG, Green SA, Jenkins SJ, Nelson HA. Experimental reproduction of ELEM: A study to determine the minimum toxic dose in ponies. Mycopathologia 1992; 117: 115–20.
- Ross PF, Rice LG, Osweiler GD, Nelson PE, Richard JL, Wilson TM. A review and update of animal toxicoses associated with fumonisins-contaminated feeds and production of fumonisins by *Fusarium* isolates. Mycopathologia 1992; 117: 109–14.
- Busta FF, Petterson EH, Adams DM, Johnson MG. Colony count method. In: Speck ML. ed. Compendium of methods for the microbiological examination of foods. Washington, DC: American Public Health Association, 1984: 62–77.
- 26. Raper KB, Fennel DI. The genus Aspergillus. Baltimore: Williams & Wilkins, 1965.
- 27. Barnet HL, Hunter BB. Illustrated genera of imperfect fungi, 3rd ed., Minneapolis: Burgess, 1972.
- Nelson PE, Toussoun TA, Marasas WFO. *Fusarium* species: An illustrated manual for identification. University Park, PA: The Pennsylvania State University Press, 1983.
- Corrêa B, Pozzi CR, Gambale W, Paula CR, Chacon-Reche NO. Postharvest and stored corn in Brazil: Mycoflora interaction abiotic factors, and mycotoxins occurrence. In: International IUPAC Symposium on Mycotoxins and Phycotoxins, XIII. Mexico City, 1992: p. 123 (Abstract No. 75).
- Lillehoj EB, Zuber MS. Distribution of toxin-producing fungi in mature maize kernels from diverse environments. Trop Sci 1988; 19–24.
- Moss MO. The environmental factors controlling mycotoxins formation. In: Smith JE, Henderson R. eds. Mycotoxins and animal foods. Boca Raton, FL: CRC Press, 1991: 37–56.

- Wilson TM, Nelson PE, Ryan TB, Rouse CD, Pittman CW, Neal TP, Portereld ML, Saunders GK. Linking leukoencephalomalacia to commercial horse rations. Vet Med 1985; 80: 63–9.
- Gabal MA, Awad YL, Morcos MB, Barakat AM, Maliki G. Fusariotoxicoses of farm animals and mycotoxic leucoencephalomalacia of the equine associated with the finding of trichothecenes in feedstuffs. Vet Human Toxicol 1986; 28: 207–12.
- 34. Mirocha CJ, Pathen SV, Christensen CM. Chemistry of *Fusarium* toxin and stachybotrys mycotoxins. In: Wyllie TD, Morehouse LG. ed. Mycotoxic fungi, mycotoxins, mycotoxicoses: Mycotoxic fungi and chemistry of mycotoxins. New York: Marcel Dekker, 1978: 365–420.
- Ueno Y. Trichothecenes: Chemical, biological and toxicological aspects. New York: Elsevier, 1983: 61–71.
- Sydenham EW, Gelderblom WCA, Thiel PG, Marasas WFO. Evidence for the natural occurrence of fumonisin B₁, a mycotoxin produced by *Fusarium moniliforme*, in corn. J Agric Food Chem 1990; 38: 285–90.
- Martins VMV, Martins E, Neves DS, Riet-Correa F, Meireles MCA. Leucoencefalomalácia em eqüinos, no Estado de Santa Catarina. In: Ciclo de Atualização em Medicina Veterinária (CAMEV), 6. Lajes-SC, 1988 (Resumo p. 237).

Address for correspondence: Dr Benedito Corrêa, Departamento de Microbiologia, Instituto de Ciências Biomédicas, Universidade de São Paulo, Av. Prof. Lineu Prestes, 1374-2°. andar, Cidade Universitária, 05508-900, São Paulo, SP, Brasil Fax: (55) 11813 0845