

Special Article

Transforming Growth Factor Beta (TGF- β) in Inflammation: A Cause and a Cure

SHARON M. WAHL¹

Accepted: November 13, 1991

INTRODUCTION

Of the many polypeptide growth factors, transforming growth factor beta (TGF- β) appears to be relatively unique in its global effects on cell growth and differentiation (1). In addition to its complex growth regulatory activities on essentially all cell types, compelling evidence has accumulated over the past 5 years documenting the role of TGF- β as a potent immunomodulatory molecule (2). Quickly evident were its profound immunosuppressive actions and many diverse inhibitory effects *in vitro*. These observations were promptly followed by therapeutic trials of the peptide in experimental models of inflammation and immune-mediated disease. Although in many cases TGF- β has fulfilled its promise as an immunotherapeutic agent, continuing investigations have revealed the bifunctional nature of the peptide. Thus, as both an immunosuppressive and a potent proinflammatory molecule, TGF- β orchestrates events vital to the initiation, progression, and resolution of immune-mediated inflammatory responses. These opposing effects are likely controlled by the cytokine's selective production and latency, as well as receptor modulation and differential susceptibility of target cells at various stages of development, maturation, and activation

(Table I). As the seemingly contradictory nature of TGF- β continues to be unraveled and knowledge of its role in host defense continues to expand, we must remain cognizant that TGF- β is but one mediator, albeit an influential one, in the cytokine network that regulates the immune system.

TGF- β SYNTHESIS AND ACTIVATION

Synthesis

At least five homodimeric polypeptides, which share 70–80% homology and many biological activities (reviewed in Ref. 1), are represented in the highly conserved TGF- β family. Of these multiple forms, only TGF- β 1, - β 2, and - β 3 have been identified in mammalian species, and differences in cellular origin and actions between these isoforms are now being recognized. The gene for each of the isoforms is located on a different chromosome and differential expression may occur through distinct cell-specific regulatory mechanisms. The peptides are encoded as preproteins of 390–412 amino acids with signal peptides of 20–23 amino acids at the N terminus (3). Two major transcriptional start sites for TGF- β 1 mRNA have been identified, with two promoter regions containing binding sites for the transcription factors, NF-1, SP-1, and AP-1 (4–6).

TGF- β 1, - β 2, and - β 3 have a 4- or 5-amino acid processing site and, after signal peptidase and proteolytic cleavage, are processed to the 112-residue forms (12.5 kD), which have in common 9 cysteine

¹Cellular Immunology Section, National Institute of Dental Research, National Institutes of Health, Bethesda, Maryland 20892.

Table I. Factors Determining Biological Effects of TGF- β

| |
|-----------------------------------|
| Latent or active TGF- β |
| TGF- β isoform |
| Target-cell type |
| State of cell differentiation |
| Resting or activated cells |
| Receptor type and number |
| Influence of other growth factors |
| Binding proteins |
| Presence of antagonists |
| Extracellular matrix |

residues and the C-terminal sequence, Cys-Lys-Cys-Ser-COOH (3). The processed mature TGF- β homodimer (25 kD) is biologically inactive or latent due to its noncovalent association with the 75-kD glycosylated latency-associated protein (LAP), which is covalently linked to a 135-kD binding protein (1, 7).

Activation

The above complex requires activation to release the biologically active molecule, which then binds to the receptor to initiate signal transduction. The masking of TGF- β activity by its association with the LAP proregion dimer is important in protecting cells that make TGF- β from the molecule's potent regulatory effects and in controlling the pleiotropic paracrine activities. In all forms of TGF- β , except TGF- β 2, LAP contains the fibronectin cellular recognition (RGD) sequence (8), raising the possibility that the latent complex or, possibly, free LAP binds to cells through this domain or via an alternative heparin binding site (7). The ability of TGF- β to bind to matrix molecules may be pivotal in localizing, focusing, and concentrating its activities. The dynamic interaction between growth factors such as TGF- β and the extracellular matrix is likely essential to the evolution of inflammatory events (9).

Intense interest in the activation of the latent complex, a crucial regulatory step in TGF- β bioactivity (1, 10), has led to new insights into the physiologic mechanisms of activation of TGF- β . The LAP carbohydrates appear to be essential to maintaining latency since sialidase and glycosidases are effective activators of TGF- β (11). The latent complex reportedly binds, at least on bovine aortic endothelial and smooth muscle cells, to mannose 6-phosphate/insulin-like growth factor II receptors, where, once bound, it becomes susceptible to protease activation (12). These proteases, including plasmin and cathepsin, and transient exposure to

acidic conditions (6) partially activate the latent molecule (13). The activation of TGF- β is likely a multistep process, involving cells such as gamma interferon (γ IFN)-treated monocytes (14) and inflammatory macrophages (15) which generate sialidases and proteases within an acidic microenvironment (16, 17) to activate the latent complex. Once activated, TGF- β binds either to cellular receptors or to one of several binding proteins. Although the latent form of the molecule circulates with a half-life >90 min, active TGF- β is cleared from the circulation within minutes (18). Thus, latency confers stability, target-cell specificity, and a means for extracellular control of the broad spectrum activities of this ubiquitous molecule.

PROINFLAMMATORY ACTIVITIES OF TGF- β

Inflammatory-Cell Recruitment

The nearly immediate release of TGF- β by platelets (19) at sites of injury or immunologic challenge provides an important clue that TGF- β has an agonist role in the sequence of events constituting an inflammatory response. Platelets represent the most concentrated natural source of TGF- β , producing nearly 20 mg/kg (20). It is difficult to fathom that a molecule with only immunosuppressive potential would be one of the first mediators released in an inflammatory cascade. In fact, the notion of TGF- β as a promoter of the inflammatory process has been borne out in subsequent studies. Originally identified as the most potent chemoattractant for monocytes with activity in the femtomolar concentration range (21), TGF- β has more recently been found to have chemotactic activity for both neutrophils (22–24) and T lymphocytes (25). Moreover, the recruitment potential of TGF- β has been confirmed *in situ* following direct injection of the native or recombinant molecule intradermally (26) and intraarticularly (23, 27). Whereas locally applied TGF- β initiates leukocyte infiltration (28), this response is absent in monocytopenic animals (29).

In defining the underlying molecular signals responsible for this proinflammatory activity of TGF- β , evidence now indicates that TGF- β influences mononuclear phagocyte recruitment by four interrelated pathways: modulation of integrin expression, increased monocyte-matrix adhesion, enhancement of matrix-specific collagenase secretion, and chemotaxis (21, 30) (Table II). In order for circulating monocytes to exit the vascular bed at

Table II. Immunoregulation by TGF- β

| Proinflammatory | Immunosuppressive |
|--------------------|--------------------------|
| ↑ Integrins | ↓ ROI, NOI |
| ↑ Collagenase | ↓ TGF- β receptors |
| ↑ Chemotaxis | ↓ IL-1 receptor |
| ↑ Th 1 lymphocytes | ↓ IL-1ra |
| ↑ Cytokines | ↓ Hematopoiesis |
| ↑ Matrix | ↓ Proliferation |
| | ↓ LAK, NK, CTL |

sites of injury, they become enmeshed in a clot or adhere to the vascular wall, subsequently encountering the basement membrane, a barrier of type IV collagen, laminin, and fibronectin, which they must traverse. The signals regulating these events are not well delineated, although the cytokines interleukin 1 (IL-1), tumor necrosis factor (TNF), and γ IFN appear to be influential in certain aspects of this process (31–33). Considering that TGF- β has been shown to inhibit adhesion of leukocytes to endothelial cells *in vitro* (34), it is unclear how TGF- β induces leukocyte accumulation at injection sites *in vivo*. However, one mechanism whereby TGF- β may initiate the requisite cell–cell and cell–matrix interactions is by virtue of its ability to enhance cell surface integrin expression (30).

Integrins are heterodimers composed of an α and a β subunit (32, 33) which form transmembrane links between the extracellular matrix and the cellular cytoskeletal elements. Functionally, the β_2 heterodimers mediate cell–cell interactions, whereas the β_1 integrins (VLA) foster cell–matrix binding. In response to picomolar levels of TGF- β , monocyte $\alpha_3\beta_1$ receptors, which interact with cell binding domains of laminin, collagen, and fibronectin, increase. Besides $\alpha_3\beta_1$, monocyte fibronectin receptors ($\alpha_5\beta_1$) also increase as previously shown for mesenchymal cells (35). Augmentation of integrin receptor expression is reflected by increased mononuclear phagocyte adhesiveness to structural matrix substrates.

Negotiation of a path through the matrix to the site of inflammation requires leukocyte release of matrix-degrading enzymes. To this end, monocyte production of the 92- and 72-kDa gelatinase/type IV collagenases is augmented by TGF- β (30). This response is quite specific since circulating monocytes require an appropriate stimulus, such as TGF- β , to synthesize and secrete this enzymatic activity. Consequently, the rapid release of TGF- β , first by platelets and then by inflammatory cells themselves, can serve as a powerful recruitment

factor, mediating integrin expression, attachment, and collagenolytic activity in a finely tuned scenario enabling the leukocytes to marginate, undergo diapedesis, traverse the basement membrane, and commence extravascular directed migration.

Mononuclear-Cell Activation

Once at the site of injury or inflammation, the recruited leukocytes are subject to regulation by higher concentrations of a plethora of potential mediators and cytokines, including TGF- β . For example, levels in excess of 20 ng/ml of active TGF- β have been reported in inflamed synovial fluids (15, 36, 37). Understanding the mechanisms regulating the exuberant production of this peptide may elucidate the immunopathology of diseases such as rheumatoid arthritis. Within inflammatory sites, several types of cells are capable of producing TGF- β , including activated lymphocytes, macrophages, neutrophils, and synovial fibroblasts (15, 37–43). Moreover, TGF- β has been shown to augment its own secretion in many cells, setting in motion both autocrine and paracrine circuits of potentially cyclic release. The autoinduction of TGF- β 1 transcription is mediated by AP-1 (Jun-Fos) binding sites in the TGF- β 1 promoter (6). Monocytes, responding to picomolar concentrations of TGF- β , demonstrate increased mRNA expression and secretion of both TGF- β 1 and β 2 peptides (40). Activated T cells also transcribe and translate both TGF- β 1 and TGF- β 2 (42). Accordingly, expression of TGF- β mRNA appears most abundant during active inflammation and cell-mediated immune reactions (44).

Apart from its capacity to promote cell recruitment, TGF- β is a potent regulator of other critical cell functions. At picomolar concentrations, TGF- β increases resting monocyte mRNA levels for IL-1, TNF, platelet-derived growth factor (PDGF), basic fibroblast growth factor (bFGF), and IL-6 (21, 27, 39, 40–47). The secretion of IL-1, TNF, and IL-6 (40, 45, 47) can trigger a cytokine cascade in which each of these molecules plays a part. For example, induction of IL-1 stimulates T cells posttranscriptionally to produce elevated levels of active TGF- β (48), which can further increase IL-1 synthesis (40) in a positive feedback loop. In addition to cytokines, TGF- β induces Fc γ RIII (CD16) on newly recruited monocytes (49, 50), which together with increased adhesion molecules, promote phagocytic activity and the release of toxic oxygen radicals *in*

vitro (49) and *in vivo* (51, 52). Thus, one of the principal actions of TGF- β in monocytes may be to control gene transcription, albeit transiently (53), for a series of biologically active molecules that control many of the subsequent events in the inflammatory process.

The effect of TGF- β on T lymphocytes, originally considered to be exclusively inhibitory, now appears to be more complex. TGF- β has recently been shown to function in a costimulatory capacity to regulate T-cell proliferation triggered through the T-cell receptor/CD3 complex (54). Moreover, exposure of CD4⁺ murine T cells to TGF- β results in the rapid conversion of putative precursors (CD45RB high, Pgp-1 low) to mature memory CD4 cells as characterized by loss of CD45RB, increased expression of Pgp-1 (CD44), increased proliferative capacity, and secretion of lymphokines (IL-2), all traits consistent with the T-helper cell type I phenotype (54, 55). The differentiation state of the cell appears to be particularly critical in determining how the cell will respond to TGF- β , with naive T cells induced to proliferate and the growth of mature T cells (CD45RB low, CD44 high) subject to inhibition. At sites of immunologic challenge the initial rapid release and accumulation of TGF- β would be conducive to the generation of TH₁ cells, which are essential to the evolution of delayed-type hypersensitivity and antigenic memory. Localization of this subset of cells within the lesion may be facilitated by the TGF- β regulated increase in CD44 (33).

The identification of TGF- β at sites of inflammation (15, 36, 37, 43, 56), coupled with the consequences observed following local injection of TGF- β (23, 26, 27), implicates the peptide as playing a critical role in the early phases of an inflammatory response (Table II). While functioning to promote inflammatory events, aberrant expression of TGF- β may have pathologic consequences. In this context, locally administered TGF- β exacerbates an ongoing inflammatory lesion as demonstrated in an experimental model of synovial inflammation and tissue destruction (57).

Inhibition of TGF- β -Mediated Recruitment and Activation

If TGF- β is, in fact, contributing to the chronicity of arthritis and other lesions, then specific antagonists of TGF- β should be able to block the progression of disease or, possibly, induce regression. In

animals receiving an arthropathic dose of bacterial cell walls, a single injection of anti-TGF- β intraarticularly reversed the sequelae of events culminating in joint destruction. Whereas the isotype control antibody had no appreciable effect, anti-TGF- β administered to the contralateral joint suppressed both the acute neutrophil-mediated synovitis and the chronic T cell-dependent and monocyte-mediated destructive phase of the arthritis. These data establish a very strong association between the enhanced TGF- β identified in synovial fluids and tissues of arthritis patients (15, 36, 37, 43) and the pathophysiology of this disease. In another disease model, antibodies to TGF- β also suppressed the progression of experimental glomerulonephritis (59), indicating that abnormal expression of this otherwise normal chemotactic and growth factor can be pathologic.

TISSUE REPAIR AND FIBROSIS

Wound Healing

After the initial inflammatory reaction, TGF- β plays a critical role in resolution of the response and in tissue repair and fibrosis. The response initiated solely by TGF- β is self-limited and reversible (26, 27), likely due to the short half-life of TGF- β (18), down-regulation of TGF- β receptors (49, 60), and the fact that TGF- β cannot prevent leukocyte programmed cell death (PCD) (61). However, in concert with other inflammatory mediators as well as with matrix components, TGF- β regulates many processes including angiogenesis, chemotaxis, fibroblast proliferation and controlled synthesis and degradation of the extracellular matrix, necessary for tissue repair (1). Fibroblasts recruited by TGF- β (27, 62) are indirectly stimulated by this cytokine to proliferate through the induction of PDGF (63) and PDGF receptors (64). Subsequently, the production of fibroblast collagen, fibronectin, and glycosaminoglycan is directly upregulated by TGF- β (1, 26, 65). Moreover, many of the effects of TGF- β on connective tissue regeneration and neovascularization (26, 27, 66) may well be secondary to the induction of mediators by the infiltrating mononuclear phagocytes. Compounding the TGF- β -mediated matrix regeneration is the TGF- β enhanced production of protease inhibitors and suppression of proteolytic enzymes (67, 68). The net effect of this differential regulation of matrix proteins, enzymes, and enzyme inhibitors is the accumulation of matrix proteins

Table III. Potential Therapeutic Applications of TGF- β

| |
|--------------------------------------|
| Local administration |
| Soft tissue repair |
| Wound healing |
| Decubitus, diabetic, vascular ulcers |
| Burns |
| Periodontal lesions |
| Retinal tears |
| Hard tissue repair: fractures |
| Psoriasis |
| Systemic administration |
| Antiinflammatory agent |
| Autoimmune diseases |
| Organ transplants |
| Osteoporosis |
| Malignancies |

and scar formation (26), which can be taken advantage of clinically to promote deficient wound healing (69, 70) (Table III).

Fibroproliferative Disorders

Beyond the normal regulation of inflammation and scar formation, aberrations in persistence, production, or regulation of TGF- β may be important in excessive matrix generation. TGF- β apparently must be present in a regulated amount, since insufficient levels of the peptide may impede inflammatory and healing processes, and excess TGF- β potentiates chronic inflammatory and fibrotic lesions. In this regard, TGF- β is a potent upregulator of collagen gene expression (71–73), indicating its potential role in fibrotic disorders of the liver (56, 74, 75), lung (76, 77), and skin (78). Moreover, a spectrum of connective tissue disorders including rheumatoid arthritis, scleroderma, myelofibrosis, and hepatic, intraocular, and pulmonary fibrosis exhibits substantial TGF- β mRNA and peptide synthesis (15, 36, 76, 78–82). These observations showing an increase in TGF- β , especially TGF- β_2 (78, 81), are consistent with a role for overexpression of the molecule in pathologic processes. Whether this abundance of TGF- β is the consequence of increased synthesis or absence of regulation is unclear.

Unregulated TGF- β may result from inadequate levels of binding proteins and clearance (83, 84) or lack of antagonists such as IL-4 (85). *In vitro*, both γ IFN and TNF counteract the profibrotic activity of TGF- β (86), and importantly, patients with chronic hepatitis C show decreases in both liver TGF- β_1 mRNA and procollagen type 1 mRNA following α IFN therapy (79). Thus, regulation of production

and bioactivity of this cytokine influence the outcome of an inflammatory response.

REGULATION OF RECEPTORS

The ability of cells and the local milieu to generate TGF- β in a form that binds to cellular TGF- β receptors is crucial to the progression of inflammatory events. Once dissociated from the latency-associated peptide, the active form of the TGF- β interacts with its specific cell surface receptors to mediate its multiple biologic activities. With few exceptions, cells express TGF- β receptors, and coupled with the diversity of its effects, this peptide appears somewhat unique in its broad spectrum of effects on normal as well as pathologic immune responses. Thus, the study of TGF- β receptors and their modulation is of prime importance for understanding the mechanisms by which TGF- β elicits its biologic effects and for identifying mechanisms to control the effects of this pleiotropic peptide.

Receptor Characterization

Based on size, three classes of TGF- β receptors have been characterized: type I 53-kDa binding glycoprotein, type II 85- to 110-kDa glycoprotein, and type III dimer composed of 250- to 350-kDa proteoglycan subunits (87, 88). Of these different forms, types I and II appear to have signal transducing capabilities (89–91), whereas type III receptors, which consist of the glycosaminoglycan residues heparan sulfate and chondroitin sulfate attached to a 100- to 140-kDa core binding protein, appear to function in a ligand storage capacity and/or to interact with extracellular matrix constituents (88). Soluble forms of the type III receptor have been detected which bind TGF- β similarly to the membrane bound form (92). Although not identified on hematopoietic cells, a type IV (60-kD) and a type V (400-kD) glycoprotein receptor have recently been reported (93, 94) but their functional characteristics have not been established. No signaling occurs following ligand binding to the type III proteoglycan (89). However, signal transduction pathways initiated via ligand binding to the types I and II receptors, which may number only a few hundred, have not been fully elucidated, with no demonstrated tyrosine kinase activity (95, 96), but possible involvement of G proteins (97). After cloning of the genes for these TGF- β receptors has been accomplished, a better understanding of receptor

function in transmitting signals to the cell interior will be forthcoming.

Receptor Modulation

The number of TGF- β receptors on cell surfaces varies with the cell type and with the level of maturation or activation, providing a basis for differential sensitivity and regulation of cellular functions in response to TGF- β (Table I). This differential regulation may provide a crucial mechanism for reversing the potentially cyclic protagonist activities of TGF- β . For example, resting human monocytes freshly isolated from peripheral blood possess ~400 TGF- β binding sites (binding constant, <10 pM), all type I receptors. In these circumstances, the monocytes are exquisitely sensitive to stimulation by TGF- β (21, 53). As these cells become activated by inflammatory stimuli including γ IFN or bacterial-derived lipopolysaccharide, the expression of TGF- β receptors is down-regulated. Consequently, these cells lose their sensitivity to TGF- β stimulation (21, 53), even though they are not globally suppressed since they may remain responsive to stimulation by other agents.

In addition to activation-induced loss of TGF- β receptors, the type I receptors are acutely down-regulated or internalized ($T_{1/2} = 5$ min) following exposure to TGF- β as the receptor-ligand complexes are internalized (60). This measurable ligand-induced loss of TGF- β receptors is somewhat unique to monocytes, probably because these cells express only one class of receptors, whereas many other types of cells express two or three receptor classes, masking receptor fluctuation. Subsequent receptor regeneration in monocytes is slow (60), and the incomplete regeneration of receptors, together with the loss of TGF- β receptors induced by inflammatory stimuli, is responsible for the reduced sensitivity of activated monocytes to TGF- β . This loss of sensitivity to TGF- β by activated monocytes may be a turning point in reversing an inflammatory response.

Conversely, but with similar immunosuppressive consequences, T lymphocytes increase their expression of types I and II TGF- β receptors and also express the proteoglycan binding sites for TGF- β as they become activated (42, 89, 98). This dichotomy in receptor expression by these two cells central in an immune response may subserve the same function, namely, down-regulation of the response. Once T cells become activated, expressing

increased TGF- β receptors, exposure to picomolar levels of TGF- β may compromise the ability of these cells to proliferate and secrete certain cytokines (42, 55, 98–100). It is remarkable that TGF- β stimulates naive CD4 cells to become memory cells and then ultimately blocks their further clonal expansion in a negative feedback loop. Escape from suppression has been associated with loss of TGF- β receptors (101). Thus, whether the effects of TGF- β on a cell are inhibitory or stimulatory depends in large part on the context (9).

IMMUNE SUPPRESSION BY TGF- β

Growth Inhibition

In several models of inflammatory disease, including adjuvant and bacterial cell wall-induced arthritis (102, 103), experimental allergic encephalomyelitis (103–105), graft rejection (106), and reperfusion-induced injury (107), systemically administered TGF- β down-modulates organ inflammation. How TGF- β promotes or inhibits immune processes remains elusive, yet the evidence appears solid that systemic, in contrast to local, administration favors suppression (Table II). Significantly, a striking absence of generalized immune suppression was noted in experimental animals receiving TGF- β systemically (102, 105), despite pronounced suppression of the localized immune responses. These observations are consistent with the resistance of resting lymphocytes, but increased susceptibility of activated T lymphocytes, to suppression. Probably the single most important determinant of the biological consequences of TGF- β on a particular target cell is related to its state of differentiation (Table I). Such opposing effects of TGF- β on cells at different times in their life cycle suggests the following sequence of events: TGF- β is released early and functions in many capacities to promote inflammation based on its stimulating effects on immature monocytes and T cells. Once these cells differentiate and become activated, they become susceptible to phenotypic modulation and functional inhibition by TGF- β . Under normal conditions, the transition from pro- to antiinflammatory responsiveness occurs with minimal disruption of host cell and tissue homeostasis.

In T-lymphocyte populations susceptible to growth arrest, TGF- β directly inhibits mitosis in the late G₁ phase of the cell cycle (99, 109) by an ill-defined mechanism. Inhibition involves regula-

tion of transcription of the early growth factor-dependent *c-myc* gene and of the transferrin receptor (110, 111). Moreover, in addition to these critical regulatory targets, growth inhibition may involve accumulation of the underphosphorylated retinoblastoma (*Rb*) gene product (111, 112). In addition to direct suppression of clonal expansion, TGF- β may also suppress these events indirectly by inhibiting IL-1 receptor expression (113, 114) and by inducing synthesis of IL-1 receptor antagonist (IL-1ra) (115, 116). IL-1ra, a 22-kDa protein which binds to the IL-1 receptor, has no agonist properties and blocks IL-1 binding and signal transduction, thereby aborting IL-1-dependent responses (117).

The reported differential susceptibility of CD4⁺ and CD8⁺ lymphocytes to TGF- β inhibition of proliferation may further contribute to resolution of an inflammatory response, since inhibition of mature CD4⁺ cells responsible for cell-mediated immune functions (44, 118) is near-complete, whereas TGF- β is considerably less inhibitory (119) and may even promote (54) CD8⁺ lymphocyte growth. This resistant CD8⁺ population synthesizes products including IL-4, IL-5, and IL-10 which have additional immunosuppressive actions (118). Under the appropriate conditions, TGF- β appears to foster the generation of inhibitory feedback molecules and apparently works in concert with them to reverse immune processes.

Inhibition of Hematopoiesis

Another contributing factor to the immunosuppression observed following systemic administration of TGF- β may be related to suppression of leukocytosis (102). A number of studies have shown that TGF- β is an effective inhibitor of hematopoietic events, acting primarily on immature precursor populations (120–124), consistent, once again, with the concept that the effects of TGF- β are linked to differentiation. While these *in vitro* studies have been useful for evaluating the direct effects of TGF- β in defined systems where, dependent on dose and duration, inhibition of essentially all lineages has been observed, the *in vivo* administration of TGF- β more closely reflects the physiologic consequences of this peptide based on multisystem responses (102, 125). In animal models, exogenously delivered TGF- β may not influence normal hematopoietic events to the same extent as that observed in animals undergoing intense hematopoietic activity. This conclusion is based on studies

showing that the peripheral white blood-cell count of animals receiving only TGF- β was not markedly suppressed, whereas animals with active inflammation and exhibiting leukocytosis had markedly reduced leukocyte numbers after systemic TGF- β treatment (102). Similarly, in animals subjected to a lethal dose of ionizing radiation, in which induction of IL-1 and TNF is radioprotective, TGF- β , albeit at very high doses (10 μ g/mouse), reversed the host's ability to recover (126). Thus, TGF- β 's ability to inhibit early progenitor bone marrow cell proliferation may include direct suppression of growth, but also reflect its capacity to antagonize IL-1.

At a more physiologic level, recent evidence indicates that basal autocrine production of TGF- β by a subpopulation of early progenitors negatively regulates their cycling status and this may occur through interaction with the *Rb* gene product (127). In these studies, the addition of antisense, but not sense, TGF- β oligonucleotides enhanced colony formation of all lineages of early progenitors without any effect on late erythroid progenitors or macrophage colonies. Not only did antisense TGF- β release hematopoietic progenitors from a quiescent growth factor-unresponsive state, but exogenously added TGF- β completely blocked early multipotential progenitor colony formation (127). The control of hematopoiesis by basal autocrine production of TGF- β may, in turn, be subject to retinoid control, since normal systemic levels of retinoids are believed to regulate functional TGF- β expression in responsive cells (128). Any augmentation of the basal TGF- β levels or exogenous administration of TGF- β may then become immunosuppressive, at least in part, by limiting the number of available leukocytes.

Additional Mechanisms of Immune Suppression

In addition to inhibition of T-lymphocyte proliferation and hematopoiesis, other pathways may contribute to the impressive immunosuppression induced by TGF- β in certain disease conditions. One of the earliest effects of elevated systemic TGF- β might be elimination of lesion-centered chemotactic concentration gradients required for directed extravascular migration. In animal models of inflammation, exogenously administered systemic TGF- β impairs inflammatory cell movement into the site of immunologic challenge (102). Moreover, elevations in circulating levels of TGF- β in

the autoimmune mouse strain MRL/lpr are associated with defective neutrophil migratory responses and phagocytic function (129). Another contributing factor to the suppressed host response may be the ability of TGF- β to diminish rapidly and completely IL-1- or TNF-induced hepatocyte acute-phase protein synthesis (130).

In other *in vitro* systems, TGF- β antagonizes the activity of IL-1 and TNF, and this antagonism extends to other cytokines including IL-2, IL-3, and γ IFN (42, 99, 131–134). Besides antagonizing cytokine activity, TGF- β may also inhibit the production of IL-1 and TNF (46, 135). Inhibition requires pretreatment of mononuclear phagocytes with TGF- β (37, 136), and when TGF- β is administered after stimulation, TGF- β is less or no longer inhibitory (137), which may reflect the modulation of TGF- β receptors (53). Although the inhibition of IL-1 was initially reported to occur posttranscriptionally (46), it appears that IL-1 production is not inhibited but that TGF- β coinduces IL-1 receptor antagonist (IL-1ra), thereby masking IL-1 bioactivity (115, 116). The induction of IL-1ra, by virtue of its ability to block the plethora of IL-1-dependent inflammatory and immune phenomena, may be a key event in TGF- β -mediated immune suppression.

Whether by cytokine blockade or by other molecular mechanisms, TGF- β also suppresses the development, activity, and/or differentiation of cytotoxic T, LAK, and natural killer cells (108, 133, 138–143), effectively compromising cytolytic functions. The ability of TGF- β to inhibit expression of the cytolytic pore-forming protein by CD8⁺ T cells likely contributes mechanistically to the overall decreased cytolytic activity (144).

Another susceptible lymphoid target for TGF- β is represented by cells of B lymphocyte lineage. Besides blocking their proliferative response, TGF- β inhibits IgG and IgM mRNA synthesis and protein secretion (145, 146). Inhibition of the switch from the membrane forms of μ and γ -H chain mRNA to the secreted forms in stimulated B-cell cultures occurs without altering expression of Oct-2 or NF- κ B (146). As a consequence of this decrease in membrane Ig expression and the capacity to secrete Ig in response to stimulation, B-cell responsiveness to antigens and antigen presentation are compromised. Related to its ability to inhibit B-cell growth, TGF- β promotes switching of IgM- or IgD-bearing B cells to cells bearing the IgA isotype (147–149). By inhibiting enhancement of Fc ϵ R₂ (CD23) expression, TGF- β can also be considered a negative

regulatory molecule in IgE-mediated immune responses (150). Thus, by a series of interconnecting pathways, TGF- β suppresses immune cell functions, pivotal in termination of the immune response but with the potential, if unchecked, to cause pathology.

ACQUIRED IMMUNODEFICIENCY SYNDROME

Because of the association of TGF- β with immune suppression, it was not surprising that augmented production of this cytokine was reported in patients with acquired immune deficiency syndrome (AIDS), the prototypic disease of immune suppression. Moreover, *in vitro* infection of mononuclear cells with HIV-1 causes the cells to express and secrete a peptide identified as TGF- β (52, 151–153). Mechanistically, the HIV-1 tat protein appears to up-regulate TGF- β production (152), and typical of the self-perpetuating cyclic nature of cytokine regulation, TGF- β induced by HIV-1 stimulates IL-6 production, and then IL-6 up-regulates TGF- β (154, 155). Crucial to the manifestations of HIV-1 infection, TGF- β enhances HIV-1 replication in cells of monocytic lineage and in PHA-stimulated peripheral blood mononuclear cells (156–158), although decreased viral replication occurs in the chronically infected U1 cell line (156). These data provide fairly compelling reasons to suspect that TGF- β may play an important role in the spread of infection and/or disease progression by amplifying the existing viral load. Such an hypothesis is compatible with the recent observations documenting increased TGF- β in the central nervous system of AIDS patients, a tissue highly susceptible to HIV-1 infection (153).

In addition to modulating virus expression, the increased production of TGF- β likely contributes to impaired lymphocyte proliferative responses characteristic of AIDS; this impaired proliferation can be restored in the presence of antibodies to TGF- β (152). Increased levels of TGF- β may contribute to the HIV-induced CD4/CD8 inversion by promoting naive CD8⁺ T-cell growth and suppressing mature CD4⁺ T-cell proliferation. The inability to confine TGF- β to localized areas of infection (53) may influence cellular functions elsewhere and accelerate the spread of opportunistic pathogens.

That TGF- β can suppress the expression of activated macrophage effector functions, including the generation of toxic oxygen and nitrogen species (136, 137, 159), may facilitate pathogen survival in

the tissues. In experimental models, systemically administered TGF- β facilitates parasite survival and the development of fatal infections. Not only did TGF- β exacerbate infection in susceptible animals, but TGF- β treatment of animals otherwise resistant to *Trypanosoma cruzi* caused a striking increase in parasitemia and mortality (160). Elevated levels of endogenous TGF- β have similar consequences since spleen cells of rodents infected with *Trypanosoma cruzi*, lymphocyte choriomeningitis virus (LCMV) or exposed to bacterial products produce sharply elevated quantities of TGF- β which progressively suppress lymphocyte growth and macrophage oxidative metabolism (100, 160, 161). Thus, if bacterium- or virus-infected cells manifest increased generation of TGF- β as has been shown not only for HIV-1 and LCMV (108), but also for CMV (162), EBV (163), and hepatitis C (79), this cytokine may contribute to diminished phagocyte functions, as well as other immune-dependent sequelae associated with such infections. These observations support the hypothesis that augmented TGF- β levels may exert global negative regulatory effects in conditions such as AIDS and provide impetus for identifying relevant antagonists which might alter the course of infections and immune function in these patients.

SUMMARY

As we continue to explore the biology of TGF- β in the network of cells and mediators contributing to host defense, the mechanisms controlling whether the pro- or antiinflammatory effects of this peptide prevail will be unraveled. Understanding these basic mechanisms may offer new approaches for identifying agonists and/or antagonists and in which circumstances their use might be appropriate. The striking differences between local and systemic administration of this cytokine reaffirm that the functional consequences of any biologic mediator must be considered in context (9) and, furthermore, suggest avenues of therapeutic application (Table III). In summary, the central role of TGF- β in normal and aberrant host defense has become indisputable.

ACKNOWLEDGMENTS

The author is indebted to Ms. Chiquita Odinma for her excellent secretarial assistance.

REFERENCES

1. Roberts AB, Sporn MB: The transforming growth factor β s. *Handbk Exp Pharm* 95:419–458, 1990
2. Wahl SM, McCartney-Francis N, Mergenhagen SE: Inflammatory and immunomodulatory role of transforming growth factor beta. *Immunol Today* 10:258–261, 1989
3. Derynck R, Jarett JA, Chen EY, Eaton DH, Bell JR, Assoian K, Roberts AB, Sporn MB, Goeddel DV: Human transforming growth factor- β complementary DNA sequence and expression in normal and transformed cells. *Nature* 316:701, 1985
4. Kim SJ, Denhez F, Kim KY, Holt JT, Sporn MB, Roberts AB: Activation of the second promoter of the transforming growth factor β 1 gene by transforming growth factor β 1 and phorbol ester occurs through the same target sequences. *J Biol Chem* 264:19373–19378, 1989
5. Kim SJ, Jeang KT, Glick A, Sporn MB, Roberts AB: Promoter sequences of the human transforming growth factor- β 1 gene responsive to transforming growth factor- β 1 autoinduction. *J Biol Chem* 264:7041–7045, 1989
6. Kim SJ, Angel P, Lafyatis R, Hattori K, Kim KY, Sporn MB, Karin M, Roberts AB: Autoinduction of transforming growth factor β 1 is mediated by the AP-1 complex. *Mol Cell Biol* 10:1492–1497, 1990
7. Wakefield LM, Smith DM, Falanders KC, Sporn MB: Latent transforming growth factor- β from human platelets. A high molecular weight complex containing precursor sequences. *J Biol Chem* 263:7646, 1988
8. Ruoslahti E, Pierschbacher MD: Arg-Gly-Asp: A versatile cell recognition signal. *Cell* 44:517–578, 1986
9. Nathan C, Sporn M: Cytokines in context: *J Cell Biol* 113:981–986, 1991
10. Wakefield LM, Smith DM, Masui T, Harris CC, Sporn MB: Distribution and modulation of the cellular receptor for transforming growth factor-beta. *J Cell Biol* 105:965, 1987
11. Miyazano K, Heldin CH: Interaction between TGF- β 1 and carbohydrate structures in its precursor renders TGF- β 1 latent. *Nature* 338:158–160, 1989
12. Dennis PA, Rifkin DB: Cellular activation of latent transforming growth factor β requires binding to the cation-independent mannose 6-phosphate/insulin-like growth factor type II receptor. *Cell Biol* 88:580–584, 1991
13. Lyons RM, Keski-Oja J, Moses HL: Proteolytic activation of latent transforming growth factor- β from fibroblast-conditioned medium. *J Cell Biol* 106:1659–1665, 1988
14. Twardzik DR, Mikovits JA, Ranchalis JE, Purchio AF, Ellingsworth L, Ruscetti FW: γ -Interferon-induced activation of latent transforming growth factor- β by human monocytes. In *Transforming Growth Factor- β s: Chemistry, Biology, and Therapeutics*, KA Piez, MB Sporn (eds). New York, New York Academy of Sciences, 1990, p 276
15. Wahl SM, Allen JB, Wong HL, Dougherty SF, Ellingsworth LR: Antagonistic and agonistic effects of transforming growth factor- β and IL-1 in rheumatoid synovium. *J Immunol* 145:2514–2519, 1990
16. Pilatte Y, Bignon J, Lambre CR: Lysosomal and cytosolic sialidases in rabbit alveolar macrophages: Demonstration of increased lysosomal activity after in vivo activation with bacillus Calmette Guerin. *Biochim Biophys Acta* 923:150, 1987

17. Silver IA, Murrills RJ, Etherington DJ: Microelectrode studies on the acid microenvironment beneath adherent macrophages and osteoclasts. *Exp Cell Res* 175:266, 1988
18. Wakefield LM, Winokur TS, Hollands RS, Christopherson K, Levinson AD, Sporn MB: Recombinant latent transforming growth factor β 1 has a longer plasma half-life in rats than active transforming growth factor β 1, and a different tissue distribution. *J Clin Invest* 86:1976-1984, 1991
19. Assoian RK, Komoriya A, Meyers CA, Miller DM, Sporn MB: Transforming growth factor-beta in human platelets. *J Biol Chem* 258:7155-7160, 1983
20. Van den Eijnden-van Raaij AJM, Koornneef I, van Zoelen EJJ: A new method for high yield purification of type beta transforming growth factor from human platelets. *Biochem Biophys Res Commun* 157:16-23, 1988
21. Wahl SM, Hunt DA, Wakefield L, McCartney-Francis N, Wahl LM, Roberts AB, Sporn MB: Transforming growth factor beta (TGF- β) induces monocyte chemotaxis and growth factor production. *Proc Natl Acad Sci* 84:5788-5792, 1987
22. Brandes ME, Mai UEH, Ohura K, Wahl SM: Human neutrophils express type I TGF- β receptors and chemotax to TGF- β . *J Immunol* 147:1600-1606, 1991
23. Fava RA, Olsen NJ, Postlethwaite AE, Broadley KN, Davidson JM, Nanney LB, Lucas C, Townes AS: Transforming growth factor β 1 (TGF- β 1) induced neutrophil recruitment to synovial tissues: Implications for TGF- β -driven synovial inflammation and hyperplasia. *J Exp Med* 173:1121-1132, 1991
24. Reibman J, Meisler S, Lee TC, Gold LI, Cronstein BN, Haines KA, Kolasinski SL, Weissmann GW: Transforming growth factor β 1, a potent chemoattractant for human neutrophils, bypasses classic signal-transduction pathways. *Proc Natl Acad Sci* 85:6805-6809, 1991
25. Adams DH, Hathaway M, Shaw J, Burnett D, Elias E, Strain AJ: Transforming growth factor- β induces human T lymphocyte migration in vitro. *J Immunol* 147:609-612, 1991
26. Roberts AB, Sporn MB, Assoian RK, Smith JM, Roche NS, Wakefield LM, Heine UI, Liotta LA, Falanga V, Kehrl JH: Transforming growth factor type β : rapid induction of fibrosis and angiogenesis in vivo and stimulation of collagen formation in vitro. *Proc Natl Acad Sci* 83:4167-4171, 1986
27. Allen JB, Manthey CL, Hand A, Ohura K, Ellingsworth L, Wahl SM: Rapid onset synovial inflammation and hyperplasia induced by transforming growth factor β . *J Exp Med* 171:231-247, 1990
28. Lynch SE, Colvin RB, Antoniades HN: Growth factors in wound healing. *J Clin Invest* 84:640-646, 1989
29. Pierce GF, Mustoe TA, Lingelbach J, Masakowski VR, Gramates P, Deuel TF: Transforming growth factor β reverses the glucocorticoid-induced wound-healing deficit in rats: Possible regulation in macrophages by platelet-derived growth factor. *Cell Biol* 86:2229-2233, 1989
30. Wahl SM, Allen JB, Weeks BS, Wong HL, Klotman PE: TGF- β enhances integrin expression and Type IV collagenase secretion in human monocytes (submitted for publication), 1992
31. Wahl SM: Recruitment. In *Encyclopedia of Immunology*, JM Roitt, PJ Delves (eds). London, WB Saunders, pp 1304-1307, 1991
32. Ruoslahti E: Integrins. *J Clin Invest* 81:1-5, 1991
33. Springer TA: Adhesion receptors of the immune system. *Nature* 346:425-434, 1990
34. Gamble JR, Vadas MA: Endothelial adhesiveness for blood neutrophils is inhibited by transforming growth factor- β . *Science* 242:97, 1988
35. Ignatz R, Heino J, Massague J: Regulation of cell adhesion receptors by transforming growth factor β : Regulation of vitronectin receptor and LFA-1. *J Biol Chem* 264:389, 1989
36. Fava R, Olsen N, Keski-Oja J, Moses H, Pincus T: Active and latent forms of transforming growth factor β activity in synovial effusions. *J Exp Med* 169:292, 1989
37. Brennan FM, Chantry D, Turner M, Foxwell B, Maini R, Feldmann M: Detection of transforming growth factor-beta in rheumatoid arthritis synovial tissue: Lack of effect on spontaneous cytokine production in joint cell cultures. *Clin Exp Immunol* 81:278-285, 1990
38. Assoian RK, Fleurdelys BE, Stevenson HC, Miller PJ, Madtes DK, Raines EW, Ross R, Sporn MB: Expression and secretion of type β transforming growth factor by activated human macrophages. *Proc Natl Acad Sci* 84:6020, 1987
39. Wahl SM, McCartney-Francis N, Allen JB, Dougherty EB, Dougherty SF: Macrophage production of TGF- β and regulation by TGF- β . *Ann NY Acad Sci* 593:188-196, 1990
40. McCartney-Francis N, Mizel D, Wong H, Wahl LM, Wahl SM: TGF- β regulates production of growth factors and TGF- β by human peripheral blood monocytes. *Growth Factors* 4:27-35, 1990
41. Grotendorst GR, Smale G, Pencev D: Production of transforming growth factor beta by human peripheral blood monocytes and neutrophils. *J Cell Physiol* 140:396-402, 1989
42. Kehrl JH, Wakefield LM, Roberts AB, Jakowlew S, Alvarez-Mon M, Derynck R, Sporn MB, Fauci AS: Production of transforming growth factor- β by human T lymphocytes and its potential role in the regulation of T cell growth. *J Exp Med* 163:1037, 1986
43. Lafyatis R, Thompson NL, Remmers EF, Flanders KC, Roche NS, Kim SJ, Case JP, Sporn MB, Roberts AB, Wilder RL: Transforming growth factor- β production by synovial tissues from rheumatoid patients and streptococcal cell wall arthritic rats. *J Immunol* 143:1142-1148, 1989
44. Yamamura M, Uyemura K, Deans RJ, Weinberg K, Rea TH, Bloom BR, Modlin RL: Defining protective responses to pathogens: Cytokine profiles in Leprosy lesions. *Science* 254:277-279, 1991
45. Wiseman DM, Polverini PJ, Kamp DW, Leibovich SJ: Transforming growth factor-beta (TGF- β) is chemotactic for human monocytes and induces their expression of angiogenic activity. *Biochem. Biophys Res Commun* 157:793, 1988
46. Chantry D, Turner M, Abney E, Feldmann M: Modulation of cytokine production by transforming growth factor- β ¹. *J Immunol* 142:4295-4300, 1989
47. Turner M, Chantry D, Feldmann M: Transforming growth factor β induces the production of interleukin 6 by human peripheral blood mononuclear cells. *Cytokine* 2:211, 1990
48. Bristol LA, Ruscetti FW, Brody DT, Durum SK: IL-1 α induces expression of active transforming growth factor- β in nonproliferating T cells via a post-transcriptional mechanism. *J Immunol* 145:4108-4114, 1990

49. Welch G, Wong H, Wahl SM: Selective induction of Fc γ RIII on human monocytes by transforming growth factor- β . *J Immunol* 144:344–3448, 1990
50. Phillips JH, Chang C, Lanier LL: Platelet-induced expression of Fc γ RIII (CD16) on human monocytes. *Eur J Immunol* 21:895–899, 1991
51. Wahl SM, Allen JB, Welch G, Wong H: Transforming growth factor β in synovial fluids modulates Fc γ RIII (CD16) expression on mononuclear phagocytes. *J Immunol* 148:485–490, 1992
52. Allen JB, Wong HL, Guyre P, Simon G, Wahl SM: Circulating Fc γ RIII positive monocytes in AIDS patients. Induction by transforming growth factor beta. *J Clin Invest* 87:1773–1779, 1991
53. Brandes ME, Wakefield LM, Wahl SM: Modulation of monocyte type 1 TGF- β receptors by inflammatory stimuli. *J Biol Chem* 266:19697–19703, 1991
54. Lee HM, Rich S: Co-stimulation of T cell proliferation by transforming growth factor- β 1. *J Immunol* 147:1127–1133, 1991
55. Swain SL, Huston G, Tonkonogy S, Weinberg A: Transforming growth factor- β and IL-4 cause helper T cell precursors to develop into disting effector helper cells that differ in lymphokine secretion pattern and cell surface phenotype. *J Immunol* 147:2991–3000, 1991
56. Manthey CL, Allen JB, Ellinsworth LR, Wahl SM: In situ expression of transforming growth factor beta in streptococcal cell wall-induced granulomatous inflammation and hepatic fibrosis. *Growth Factors* 4:17–26, 1990
57. Wahl AM, Allen JB, Brandes ME: Cytokine modulation of bacterial cell wall-induced arthritis. *In Progress in Inflammation Research and Therapy*. Birkhäuser Verlag, Basel pp 29–34, 1991
58. Wahl SM, Allen JB, Wong H, Dasch J: Reversal of synovial inflammation by antibodies to TGF- β (submitted for publication), 1992
59. Border WA, Okuda S, Languino LR, Sporn MB, Ruoslahti E: Suppression of experimental glomerulonephritis by antiserum against transforming growth factor beta 1. *Nature* 346:6282, 1990
60. Brandes ME, Wahl SM: Acute and chronic downregulation of type I TGF- β receptors by TGF- β (submitted), 1992
61. Mangan DF, Wahl SM: Differential regulation of monocyte programmed cell death (apoptosis) by chemotactic factors and inflammatory cytokines. *J Immunol* 147:3408–3412, 1991
62. Postlethwaite AE, Keski-Oja J, Moses HL, Kang AH: Stimulation of the chemotactic migration of human fibroblasts by transforming growth factor beta. *J Exp Med* 165:251–256, 1987
63. Leof EB, Proper JA, Goustin AS, Shipley GD, DiCorleto PE, Moses HC: Induction of c-sis mRNA and platelet-derived growth factor-like activity by transforming growth factor type- β : A proposed model for indirect mitogenesis involving autocrine activity. *Proc Natl Acad Sci* 83:2453–2457, 1986
64. Ishikawa O, LeRoy EC, Trojanowska M. Mitogenic effect of transforming growth factor β 1 on human fibroblasts involves the induction of platelet-derived growth factor α receptors. *J Cell Physiol* 145:181–186, 1990
65. Igotz RA, Endo T, Massagué J: Regulation of fibronectin and type I collagen mRNA levels by transforming growth factor- β *J Biol Chem* 262:6443–6446, 1987
66. Yang EY, Moses HL: Transforming growth factor beta 1-induced changes in cell migration, proliferation, and angiogenesis in the chicken chorioallantoic membrane. *J Cell Biol* 111:731–471, 1990
67. Overall CM, Wrana JL, Sodek J: Independent regulation of collagenase, 72-kDa progelatinase, and metalloendoproteinase inhibitor expression in human fibroblasts by transforming growth factor- β . *J Biol Chem* 264:1860–1869, 1989
68. Edwards DR, Murphy G, Reynolds JJ, Whitman SE, Docherty AJP, Angel P, Heath JK: Transforming growth factor beta modulates the expression of collagenase and metalloproteinase inhibitor. *EMBO J* 6:1899–1904, 1987
69. Mustoe TA, Pierce GF, Thomason A, Gramates P, Sporn MB, Deuel TF: Transforming growth factor type beta induces accelerated healing of incisional wounds in rats. *Science* 237:1333–1336, 1987
70. Smiddy WE, Glaser BM, Green R, Connor TB, Roberts AB, Lucas R, Sporn MB: Transforming growth factor beta—a biologic chorioretinal glue. *Arch Ophthalmol* 107:577–580, 1989
71. Raghow R, Postlethwaite AE, Keski-Oja J, Moses HL, Kang AH: Transforming growth factor- β increases steady state levels of type I procollagen, and fibronectin messenger RNAs posttranscriptionally in cultured human dermal fibroblasts. *J Clin Invest* 79:1285–1288, 1987
72. Varga J, Rosenbloom J, Jimenez SA: Transforming growth factor β (TGF- β) causes a persistent increase in steady-state amount of type I and type III collagen and fibronectin mRNAs in normal human dermal fibroblasts. *Biochem J* 247:597–604, 1987
73. Rossi P, Roberts AB, Roche NS, Karsenty B, Sporn MB, de Crombrughe B: A nuclear factor 1 binding site mediates the transcriptional activation of a type 1 collagen promoter by transforming growth factor- β . *Cell* 52:405–414, 1988
74. Armendariz-Borunda J, Seyer JM, Kang AH, Raghow R: Regulation of TGF- β gene expression in rat liver intoxicated with carbon tetrachloride. *FASEB J* 4:215–221, 1990
75. Czaja MJ, Weiner FR, Flanders KC, Giambone MA, Wind R, Biempica L, Zern MA: In vitro and in vivo association of transforming growth factor- β 1 with hepatic fibrosis. *J Cell Biol* 108:2477–2482, 1989
76. Khalil N, Bereznyay O, Sporn M, Greenberg AH: Macrophage production of transforming growth factor β and fibroblast collagen synthesis in chronic pulmonary inflammation. *J Exp Med* 170:727–737, 1989
77. Perkett EA, Lyons RM, Moses HL, Brigham KL, Meyrick B: Transforming growth factor β activity in sheep lung lymph during the development of pulmonary hypertension. *J Clin Invest* 86:1459–1464, 1990
78. Kulozik M, Hogg A, Lankat-Buttgereit B, Kreig T: Colocalization of transforming growth factor β 2 with α 1 (I) procollagen mRNA in tissue sections of patients with systemic sclerosis. *J Clin Invest* 86:917–922, 1990
79. Castilla A, Prieto J, Fausto N: Transforming growth factors β 1 and α in chronic liver disease. Effects of interferon alfa therapy. *New Engl J Med* 324:933–940, 1991
80. Peltonen J, Kahari L, Jaakkola S, Kahari V-M, Varga J, Uitto J, Jimenez SA: Evaluation of transforming growth factor β and type I procollagen gene expression in fibrotic

- skin diseases by in situ hybridization. *J Invest Dermatol* 94:365-371, 1990
81. Connor TB, Roberts AB, Sporn MB, Danielpour D, Dart LL, Michels RG, de Bustros S, Enger C, Kato H, Lansing M, Hayashi H, Glaser BM: Correlation of fibrosis and transforming growth factor- β type 2 levels in the eye. *J Clin Invest* 83:1661-1666, 1989
 82. Terui T, Niitsu Y, Mahara K, Fujisaki Y, Urushizaki Y, Mogi, Y, Kohgo Y, Watanabe N, Ogura M, Saito H: The production of transforming growth factor- β in acute megakaryoblastic leukemia and its possible implication in myelofibrosis. *Blood* 75:1540-1548, 1990
 83. Gentry LE, Nash BW: The pro domain of pre-pro-transforming growth factor β 1 when independently expressed is a functional binding protein for the mature growth factor. *Biochemistry* 29:6851-6857, 1990
 84. Danielpour D, Sporn MB. Differential inhibition of transforming growth factor β 1 and β 2 activity by α -₂-macroglobulin. *J Biol Chem* 265:6973-6977, 1990
 85. Wong HL, Welch GR, Brandes ME, Wahl SM: Interleukin-4 (IL-4) antagonizes induction of Fc γ RIII (CD16) expression by TGF- β on human monocytes. *J Immunol* 147:1843-1848, 1991
 86. Kahari VM, Qiu-Chen Y, Ming WS, Ramirez F, Uitto J: Tumor necrosis factor- α and interferon- γ suppress the activation of human type I collagen gene expression by transforming growth factor- β 1. *J Clin Invest* 86:1489-1495, 1990
 87. Massagué J, Cheifetz S, Boyd FT, Andres JL: TGF- β receptors and TGF- β binding proteoglycans: Recent progress in identifying their functional properties. *Ann NY Acad Sci* 593:59-72, 1990
 88. Segarini P: A system of transforming growth factor- β receptors. *Am J Resp Cell Mol Biol* 4:395-396, 1991
 89. Segarini P, Rosen DM, Seyedin SM: Binding of transforming growth factor- β to cell surface proteins varies with cell type. *Mol Endocrinol* 3:261-272, 1989
 90. Laiho M, Weis FMB, Massagué J: Concomitant loss of transforming growth factor (TGF)- β receptor types I and II in TGF- β resistant cell mutants implicates both receptor types in signal transduction. *J Biol Chem* 265:18518-18524, 1990
 91. Kumar A, Rogers T, Maizel A, Sharma S: Loss of transforming growth factor β 1 receptors and its effects on the growth of EBV-transformed human B cells. *J Clin Invest* 147:998-1006, 1991
 92. Andres JL, Stanley K, Cheifetz S, Massagué J: Membrane-anchored and soluble forms of betaglycan, a polymorphic proteoglycan that binds transforming growth factor- β . *J Cell Biol* 109:3137-3145, 1989
 93. Chieftz S, Ling N, Guillemin R, Massagué J: A surface component on GH₃ pituitary cells that recognizes transforming growth factor- β , activin, and inhibin. *J Biol Chem* 263:17225-17228, 1988
 94. O'Grady P, Kuo MD, Baldassare JJ, Huang SS, Huang JS: Purification of a new type high molecular weight receptor (Type V receptor) of transforming growth factor β (TGF- β) from bovine liver. *J Biol Chem* 266:8583-8589, 1991
 95. Fanger BO, Wakefield LM, Sporn MB: Structure and properties of the cellular receptor for transforming growth factor type beta. *Biochemistry* 25:3083-3091, 1986
 96. Libby J, Martinez R, Weber MJ: Tyrosine phosphorylation in cells treated with transforming growth factor- β . *J Cell Physiol* 129:159-166, 1986
 97. Murthy US, Anzano MA, Stadel JM, Greig R: Coupling of TGF- β -induced mitogenesis to G-protein activatin in AKR-2B cells. *Biochem Biophys Res Commun* 152:1228-1235, 1988
 98. Ellingsworth L, Nakayama D, Segarini P, Dasch J, Carrillo P, Waegell W: Transforming growth factor- β s are equipotent growth inhibitors of interleukin-1-induced thymocyte proliferation. *Cell Immunol* 114:41-54, 1988
 99. Wahl SM, Hunt DA, Wong HL, Dougherty S, McCartney-Frances N, Wahl LM, Ellingsworth L, Schmidt JA, Hall G, Roberts AB, Sporn MB: Transforming growth factor beta is a potent immunosuppressive agent which inhibits interleukin-1 dependent lymphocyte proliferation. *J Immunol* 140:3026-3032, 1988
 100. Wahl SM, Hunt DA, Bansal G, McCartney-Francis N, Ellingsworth L, Allen JB: Bacterial cell wall induced immunosuppression: Role of transforming growth factor beta. *J Exp. Med.* 168:1403-1417, 1988
 101. Siepl C, Malipiero UV, Fontana A: Transforming growth factor β (TGF- β) resistant helper T lymphocyte clones show a concomitant loss of all three types of TGF- β receptors. *J Immunol* 146:3063-3067, 1991
 102. Brandes ME, Allen JB, Ogawa Y, Wahl SM: TGF- β 1 suppress leukocyte recruitment and synovial inflammation in experimental arthritis. *J Clin Invest* 87:1108-1113, 1991
 103. Kuruvilla AP, Shah R, Hochwald GM, Liggitt HD, Palladino MA, Thorbecke GJ: Protective effect of transforming growth factor β 1 on experimental autoimmune diseases in mice. *Proc Natl Acad Sci* 88:2918-2921, 1991
 104. Racke MK, Dhib-Jalbut S, Cannella B, Albert PS, Raine CS, McFarlin DE: Prevention and treatment of chronic relapsing experimental allergic encephalomyelitis by transforming growth factor- β . *J Immunol* 146:3012-3017, 1991
 105. Johns LD, Flanders KC, Ranges GE, Sriram S: Successful treatment of experimental allergic encephalomyelitis with transforming growth factor- β 1. *J Immunol* 147:1792-1796, 1991
 106. Wallick SC, Figari IS, Morris RE, Levinson AD, Palladino MA: Immunoregulatory role of transforming growth factor β (TGF- β) in development of killer cells: Comparison of active and latent TGF- β 1. *J Exp Med* 172:1777-1784, 1990
 107. Lefer AM, Tsao P, Aoki N, Palladino MA Jr: Mediation of cardioprotection by transforming growth factor- β . *Science* 249:61-64, 1990
 108. Su HC, Leite-Morris KA, Braun L, Biron CA: A role for transforming growth factor- β 1 in regulating natural killer cell and T lymphocyte proliferative responses during acute infection with lymphocytic choriomeningitis virus. *J Immunol* 147:2712-2727, 1991
 109. Siepl C, Bodmer S, MacDonald HR, Martin RD, Hofer R, Fontana A: The glioblastoma-derived T cell suppressor factor/transforming growth factor- β 2 inhibits T cell growth without affecting the interaction of interleukin 2 with its receptor. *Eur J Immunol* 18:593-600, 1988
 110. Ruegemer JJ, Ho SN, Augustine JA, Schlager JW, Bell MP, McKean DJ, Abraham RT: Regulatory effects of transforming growth factor- β on IL-2- and IL-4-dependent T cell-cycle progression. *J Immunol* 144:1767-1776, 1990

111. Pietenpol JA, Holt JT, Stein RW, Moses HL: Transforming growth factor β 1 suppression of c-myc gene transcription: Role in inhibition of keratinocyte proliferation. *Proc Natl Acad Sci USA* 87:3758–3762, 1990
112. Laiho M, DeCaprio JA, Ludlow JW, Livingston DM, Massagué J: Growth inhibition by TGF- β linked to suppression of retinoblastoma protein phosphorylation. *Cell* 62:175, 1990
113. Dubois CM, Ruscetti FW, Palaszynski EW, Falk LA, Oppenheim JJ, Keller JR: Transforming growth factor β is a potent inhibitor of interleukin 1 (IL-1) receptor expression: Proposed mechanism of inhibition of IL-1 action. *J Exp Med* 172:737–744
114. Harvey AK, Hrubey PS, Chandrasekhar S: Transforming growth factor- β inhibition of interleukin-1 activity involves down-regulation of interleukin-1 receptors on chondrocytes. *Exp Cell Res* 195:376–385, 1991
115. Turner M, Chantry D, Katsikis P, Berger A, Brennan GM, Feldman M: Induction of the interleukin 1 receptor antagonist protein by transforming growth factor- β . *J Immunol* 21:1635–1639, 1991
116. Wahl SM, Costa G, Corcoran M, Wahl LM, Berger AE: Sequential induction of interleukin 1 and interleukin 1 receptor antagonist (IL-1ra) in human monocytes by transforming growth factor β (submitted), 1992
117. Dripps DJ, Brandhuber BJ, Thompson RC, Eisenberg SP: Interleukin-1 (IL-1) receptor antagonist binds to the 80-kDa IL-1 receptor but does not initiate IL-1 signal transduction. *J Biol Chem* 266:10331–10336, 1991
118. Salgame P, Abrams JS, Clayberger C, Goldstein H, Convit J, Modlin RL, Bloom BR: Differing lymphokine profiles of functional subsets of human CD4 and CD8 T cell clones. *Science* 254:279–282, 1991
119. Lotz M, Kekow J, Carson DA: Transforming growth factor- β and cellular immune responses in synovial fluids. *J Immunol* 144:4189–4194, 1990
120. Keller JR, Mantel C, Sing GK, Ellingsworth LR, Ruscetti SK, Ruscetti FW: Transforming growth factor β 1 regulates early murine hematopoietic progenitors and inhibits the growth of interleukin 3-dependent myeloid leukemia cell lines. *J Exp Med* 168:737, 1988
121. Ottman OG, Pelus LM: Differential proliferative effects of transforming growth factor- β on human hematopoietic progenitor cells. *J Immunol* 140:2661, 1988
122. Sing GK, Keller JR, Ellingsworth LR, Ruscetti FW: Transforming growth factor β selectively inhibits normal and leukemic human bone marrow cell growth *in vitro*. *Blood* 72:1504, 1988
123. Ishibashi T, Miller SL, Burstein SA: Type β transforming growth factor is a potent inhibitor of murine megakaryocytopoiesis *in vitro*. *Blood* 69:1737–1741, 1987
124. Strassman G, Cole MD, Newman W: Regulation of colony-stimulating factor 1-dependent macrophage precursor proliferation by type β transforming growth factor. *J Immunol* 140:2645, 1988
125. Goey H, Keller JR, Back T, Longo DL, Ruscetti FW, Wiltrout RH: Inhibition of early murine hemopoietic progenitor cell proliferation after *in vivo* locoregional administration of transforming growth factor- β 1. *J Immunol* 143:877–880, 1989
126. Neta R, Oppenheim JJ, Schreiber RD, Chizzonite R, Ledney GD, MacVittie TJ: Role of cytokines (interleukin 1, tumor necrosis factor, and transforming growth factor β) in natural and lipopolysaccharide-enhanced radioresistance. *J Exp Med* 173:1177–1182, 1991
127. Hatzfeld J, Li ML, Brown EL, Sookdeo H, Levesque JP, O'Toole T, Gurney C, Clark SC, Hatzfeld A: 1991. Release of early human hematopoietic progenitors from quiescence by antisense transforming growth factor β 1 or Rb oligonucleotides. *J Exp Med* 174:925–929, 1991
128. Glick AB, Flanders KC, Danielpour D, Yuspa SH, Sporn MB: Retinoic acid induces transforming growth factor β 2 in cultured keratinocytes and mouse epidermis. *Cell Regulat* 1:87–97, 1989
129. Gresham HD, Ray CJ, O'Sullivan FX: Defective neutrophil function in the autoimmune mouse strain MRL/lpr. *J Immunol* 146:3911–3921, 1991
130. Taylor AW, Ku NO, Mortensen RF: Regulation of cytokine-induced human C-reactive protein production by transforming growth factor- β 1. *J Immunol* 145:2507–2513, 1990
131. Czarniecki CW, Chiu HH, Wong GHW, McCabe SM, Palladino MA: Transforming growth factor- β 1 modulates the expression of class II histocompatibility antigens on human cells. *J Immunol* 140:4217, 1988
132. Ohta M, Greenberger JS, Anklesaria P, Bassols A, Massagué J: Two forms of transforming growth factor- β distinguished by multipotential hematopoietic progenitor cells. *Nature (Lond)* 329:539–541, 1987
133. Ranges GE, Figari IS, Espevik T, Palladino MA: Inhibition of cytotoxic T cell development by transforming growth factor β and reversal by recombinant tumor necrosis factor α . *J Exp Med* 166:991, 1987
134. Musso T, Epionza-Delgado I, Pulkki K, Gusella GL, Longo DL, Varesio L: Transforming growth factor β downregulates interleukin-1 (IL-1) induced IL-6 production by human monocytes. *Blood* 76:2466–2469, 1990
135. Espevik T, Figari IS, Shalaby MR, Lackides GA, Lewis GD, Shepard HM, Palladino Jr, MA: Inhibition of cytokine production by cyclosporin A and transforming growth factor β . *J Exp Med* 166:571–576, 1987
136. Tsunawaki S, Sporn M, Ding A, Nathan C: Deactivation of macrophages by transforming growth factor- β . *Nature* 334:260, 1988
137. Ding A, Nathan CF, Graycar J, Derynck R, Stuehr DL, Srimal S: Macrophage deactivating factor and transforming growth factor- β 1, - β 2, and - β 3 inhibit induction of macrophage nitrogen oxide synthesis by IFN- γ 1. *J Immunol* 145:940–944, 1990
138. Rook AH, Kehrl JH, Wakefield LM, Roberts AB, Sporn MB, Burlington DB, Lane HC, Fauci AS: Effects of transforming growth factor β on the functions of natural killer cells: Depressed cytolytic activity and blunting of interferon responsiveness. *J Immunol* 136:3916, 1986
139. Mulé JJ, Schwarz SL, Roberts AB, Sporn MB, Rosenberg SA: Transforming growth factor-beta inhibits the *in vitro* generation of lymphokine-activated killer cells and cytotoxic T cells. *Cancer Immunol Immunother* 26:95, 1988
140. Fontana A, Frei K, Bodmer S, Hofer E, Schreider MH, Pallino Jr, MA, Zindernagel, RM: Transforming growth factor- β inhibits the generation of cytotoxic T cells in virus-infected mice. *J Immunol* 143:3230–3234, 1989
141. Espevik T, Figari IS, Ranges GE, Palladino MA: Transforming growth factor- β 1 (TGF- β 1) and recombinant human

- tumor necrosis factor- α reciprocally regulate the generation of lymphokine-activated killer cell activity: Comparison between natural porcine platelet-derived TGF- β_1 and TGF- β_2 , and recombinant human TGF- β_1 . *J Immunol* 140:2312, 1988
142. Ortaldo JR, Mason AT, O'Shea JJ, Smyth MJ, Falk LA, Kennedy ICS, Longo DL, Ruscetti FW: Mechanistic studies of transforming growth factor β inhibition of IL-2 dependent activation of CD3-large granular lymphocyte functions. *J Immunol* 146:3791-3798, 1991
 143. Koo GC, Manyak CL, Dasch J, Ellingsworth L, Shultz LD: Suppressive effects of monocytic cells and transforming growth factor- β on natural killer cell differentiation in autoimmune viable motheaten mutant mice. *J Immunol* 147:1194-1200, 1991
 144. Smyth MJ, Strobl SL, Young HA, Ortaldo JR, Ochoa AC: Regulation of lymphokine-activated killer activity and perforating protein gene expression in human peripheral blood CD8⁺ T lymphocytes. *J Immunol* 146:3289-3297, 1991
 145. Kehrl JH, Taylor AS, Delsing GA, Roberts AB, Sporn MB, Fauci AS: Further studies of the role of transforming growth factor- β in human B cell function. *J Immunol* 143:1868-1874, 1989
 146. Kehrl JH, Thevenin C, Rieckmann P, Fauci A: Transforming growth factor- β suppresses human B lymphocyte Ig production by inhibiting synthesis and the switch from the membrane form to the secreted form of Ig mRNA. *J Immunol* 146:4016-4023, 1991
 147. Sonoda E, Matsumoto R, Hitoshi Y, Ishii T, Sugimoto M, Araki S, Tominaga A, Yamaguchi N, Takatsu K: Transforming growth factor β induces IgA production and acts additively with interleukin 5 for IgA production. *J Exp Med* 170:1415, 1989
 148. Kim PH, Kagnoff MF: Transforming growth factor β 1 increases IgA isotype switching at the clonal level. *J Immunol* 145:3773-3778, 1990
 149. Coffman RL, Lebman DA, Shrader B: Transforming growth factor β specifically enhances IgA production by lipopolysaccharide-stimulated murine B lymphocytes. *J Exp Med* 170:1039, 1989
 150. Tanaka M, Lee K, Yodoi J, Saito H, Iwai Y, Kim KM, Morita M, Mayumi M, Mikawa H: Regulation of Fc ϵ receptor 2 (CD23) expression on a human eosinophilic cell line EoL3 and a human monocytic cell line U937 by transforming growth factor β . *Cell Immunol* 122:96, 1989
 151. Kekow J, Wachsman W, McCutchan JA, Cronin M, Carson DA, Lotz M: Transforming growth factor β and non-cytopathic mechanisms of immunodeficiency in human immunodeficiency virus infection. *Proc Natl Acad Sci USA* 87:8321, 1990
 152. Lotz M, Kekow J, Cronin MT, McCutchan JA, Clark-Lewis I, Carson DA, Wachsman W: Induction of transforming growth factor- β (TGF- β) by HIV-1 tat: A noncytopathic pathway of immunodeficiency in HIV infection. *FASEB J* 4:A2014, 1990
 153. Wahl SM, Allen JB, McCartney-Francis N, Morganti-Kossmann MC, Kossmann T, Ellingsworth L, Mergenhagen SE, Orenstein JM: Transforming growth factor beta. A potential macrophage and astrocyte-derived mediator of CNS dysfunction in AIDS. *J Exp Med* 173:891-899, 1991
 154. Nakajima K, Martinez-Maza O, Hirano T, Breen EC, Nishanian PG, Salazar-Gonzalez JF, Fahey JL, Kishimoto T: Induction of IL-6 (B cell stimulatory factor-2/IFN- β 2) production by HIV. *J Immunol* 142:531-536, 1989
 155. Zhou D, Munster A, Winchurch RA: Pathologic concentrations of interleukin 6 inhibit T cell responses via induction of activation of TGF- β . *FASEB J* 5:2582-2585, 1991
 156. Poli G, Kinter AL, Justement JS, Bressler P, Kehrl JH, Fauci AS: Transforming growth factor β suppresses human immunodeficiency virus expression and replication in infected cells of the monocyte/macrophage lineage. *J Exp Med* 173:589, 1991
 157. Lazdins JK, Klimkait T, Woods-Cook K, Walker M, Alteri E, Cox D, Cerletti N, Shipman R, Bilbe G, McMaster G: In vitro effect of transforming growth factor- β on progression of HIV-1 infection in primary mononuclear phagocytes. *J Immunol* 147:1201-1207, 1991
 158. Peterson PK, Gekker G, Chao CC, Schut R, Molitor TW, Balfour HH Jr: Cocaine potentiates HIV-1 replication in human peripheral blood mononuclear cell cocultures. *J Immunol* 146:81-84, 1991
 159. Nelson BJ, Ralph P, Green SJ, Nacy CA: Differential susceptibility of activated macrophage cytotoxic effector reactions to the suppressive effects of transforming growth factor- β 1. *J Immunol* 146:1849-1857, 1991
 160. Silva JS, Twardzik DR, Reed S: Regulation of Trypanosoma cruzi infections in vitro and in vivo by transforming growth factor β (TGF- β). *J Exp Med* 174:539-545, 1991
 161. Su HC, Leite-Morris KA, Braun L, Biron CA: A role for transforming growth factor- β 1 in regulating natural killer cell and T lymphocyte proliferative responses during acute infection with lymphocytic choriomeningitis virus. *J Immunol* 147:2717-2727, 1991
 162. Smith PD, Wahl SM: Cytokines. *In* Natural Immunity, DS. Nelson (ed). New York, Academic Press, 1989, pp 241-283
 163. Chao CC, Janoff EN, Hu S, Thomas K, Gallagher M, Tsang M, Peterson PP: Altered cytokine release in peripheral blood mononuclear cell cultures from patients with chronic fatigue syndrome. *Cytokine* 3:292-298, 1991