

Alternative mechanisms of action of anti-oestrogens

Anthony A. Colletta, John R. Benson, and Michael Baum

Section of Academic Surgery, Institute of Cancer Research, Royal Marsden Hospital, Fulham Road, London SW3 6JJ, UK

Key words: antiestrogens, cAMP phosphodiesterase, growth factors, insulin-like growth factors, protein kinase C, tamoxifen, transforming growth factor beta

Summary

The molecular mechanism of action of anti-oestrogens such as tamoxifen appears to be a complex mixture of antagonism of the mitogenic action of oestradiol at the level of the oestrogen receptor, plus a range of other activities from enzyme inhibition to growth factor modulation. This article will concentrate on two specific areas: 1) the inhibition of protein kinase C and calmodulin-dependent cAMP phosphodiesterase; and 2) the regulation by tamoxifen of peptide regulators of breast cancer epithelial cell growth such as insulin-like growth factor I (IGF I) and transforming growth factor beta (TGF- β). The elucidation of these mechanisms is potentially important in the treatment and chemoprevention of breast cancer — the quantitative contribution of each individual mechanism of the overall antineoplastic action of anti-oestrogens is central to developing new and possibly more effective anti-oestrogens and optimizing strategies for their use.

Oestrogens, like the other members of the steroid hormone family, elicit their biological activities through their cognate receptor as a ligand-activated transcription enhancer [1–3]. Anti-oestrogens, as their name implies, are antagonists of oestrogen function which serve to competitively interact with the oestrogen receptor (ER) in the nucleus of oestrogen responsive cells [4]. The net result of this interaction is to abrogate the biological function of oestradiol. There are a number of known anti-oestrogens of both steroidal and non-steroidal origin, but this review will focus on the prototype anti-oestrogen as far as breast cancer therapy is concerned, the triphenylethylene tamoxifen. There are a number of excellent reviews dealing with the structure and transcriptional activation of ER as well as the species, organ, and cell specific pharmacology of tamoxifen [5]. This article will concentrate on two

specific areas: i) the inhibition of protein kinase C and calmodulin-dependent cAMP phosphodiesterase; ii) the regulation by tamoxifen of peptide regulators of breast cancer epithelial cell growth such as insulin-like growth factor 1 and transforming growth factor beta.

The search for alternative mechanisms of action of anti-oestrogens was originally stimulated by the finding that the growth inhibition of human breast cancer cells *in vitro* with micromolar concentrations of tamoxifen and its active metabolites 4-hydroxytamoxifen and N-desmethyltamoxifen could not be completely reversed by competition with large concentrations of oestradiol [6, 7]. These findings suggested that pharmacological levels of these compounds may be acting at a site other than the conventional ER. Research in this area was stimulated by the discovery of a novel low affinity microsomal

binding protein specific for the triphenylethylene class of anti-oestrogens, termed the anti-oestrogen binding site (AEBS) [8]. However the universal distribution of AEBS, coupled with a failure to purify the protein/s to homogeneity, has led to the virtual abandonment of this line of research in recent years.

The original demonstration by O'Brian and colleagues that micromolar concentrations of tamoxifen inhibit protein kinase C activity were seen as being of great potential significance [9]. Protein kinase C (PKC) is a calcium and phospholipid-dependent protein kinase which can be activated by diverse agents such as tumour promoting phorbol esters and by diacylglycerol, a product of cellular inositol phospholipid hydrolysis [10]. Tamoxifen, 4-hydroxytamoxifen, and N-desmethyltamoxifen were shown to have IC₅₀ levels for the phosphotransferase activity of PKC of 8–25 μ M, as did the structurally related oestrogen agonist clomiphene [11, 12]. However, the calcium and phospholipid-independent phosphorylation of protamine sulphate also catalysed by PKC was not inhibited by any of the triphenylethylenes, suggesting that these compounds do not directly interact with the active site of the enzyme [9, 11].

Further experiments showed that the triphenylethylene-induced inhibition of PKC activity could be partially titrated by increasing the concentration of phospholipid in the phosphotransferase assays, suggesting that these drugs were interacting with the phospholipid rather than with PKC directly. The pharmacological importance of this enzyme inhibition was also demonstrated by the observation that the rank order for the growth inhibition of MCF-7 breast cancer cells by tamoxifen, 4-hydroxytamoxifen, and N-desmethyltamoxifen is the same as their potency in the inhibition of binding of radiolabelled phospholipid precursors in murine C3H/10T1/2 cells [9].

Another key enzyme known to be inhibited by tamoxifen and structurally related analogues is calmodulin-dependent cAMP phosphodiesterase (CDP) [13]. A structure-activity relationship has been established for the inhibition of CDP, and has indicated that tamoxifen binds to, and interferes with the action of, calmodulin, rather than to the

phosphodiesterase itself [14, 15]. The oestrogen-irreversible cytotoxic effects of tamoxifen and a range of analogues correlates with the calmodulin inhibition seen in ER positive cell lines, but not in heterologous cell lines such as L1210 and Walker cells. This suggests that ER and calmodulin are involved in the same signalling pathway responsible for hormone-regulated cellular proliferation. This conclusion is supported by the finding that the calmodulin antagonist calmidazolium and tamoxifen block the MCF-7 cell cycle at the same point [16].

The notion that anti-oestrogens such as tamoxifen influenced the cellular microenvironment in a breast tumour via the stromal component of the tumour was stimulated by the counter intuitive clinical findings of the Nolvadex adjuvant tamoxifen trial (NATO) [17]. The results of this trial suggested that the clinical benefits of tamoxifen treatment as an adjuvant to surgical treatment were quite independent of the ER status of the primary tumour. The results of this trial were confirmed by the Scottish MRC trial [18] and the Early Breast Cancer Trialists' Collaborative Group [19]. Taken together these studies demonstrate that ER status does not predict a subgroup of breast cancer patients that will not respond to adjuvant tamoxifen treatment. At much the same time there were reports of pure mesenchymal tumours which showed spectacular clinical responses to tamoxifen and its chlorinated analogue toremifene [20].

In our early studies we used two human foetal fibroblast strains as an *in vitro* model for the stromal component of breast cancer [21]. These were chosen after a report demonstrating the foetal characteristics of fibroblasts taken from patients with a strong family history of breast cancer [22, 23]. These cell strains were found to lack ER by ligand binding, immunocytochemistry, and Northern analysis, although they responded to the addition of 0.5–1 μ M tamoxifen or toremifene with an increased secretion of biologically active transforming growth factor beta (TGF- β), particularly the prototype peptide TGF- β 1, with little or no involvement of TGF- β 2 [21]. Metabolic labelling experiments followed by immunoprecipitation of TGF- β 1 showed the *de novo* synthesis of this peptide in response to tamoxifen, although Northern analysis did not

demonstrate an increase in TGF- β 1 mRNA, suggesting a post-transcriptional mechanism of action of tamoxifen in these cells. Similar post-transcriptional mechanisms were also postulated for the induction of TGF- β 2 by retinoic acid in keratinocytes [24] and the induction of TGF- β 1 and TGF- β 2 by tamoxifen and synthetic progestins in breast cancer cells [25, 26].

However, such *in vitro* findings do not always correlate well with the situation *in vivo*, so we recently examined the regulation of TGF- β secretion by tamoxifen in breast cancer patients [27]. In this immunohistochemical study the secretion of each of the three mammalian TGF- β isoforms was examined in breast tumour sections taken from patients with histologically proven invasive ductal carcinoma prior to and after three months treatment with tamoxifen. These samples were obtained from the Royal Marsden Hospital primary medical therapy trial of tamoxifen [28]. Polyclonal antibodies to both the intracellular (LC) and extracellular (CC) forms of TGF- β 1 as well as antibodies to TGF- β 2 and TGF- β 3 were used, with pre- and post-treatment sections stained simultaneously to avoid any artefactual differences.

The data obtained from these sections demonstrated a consistent increase in the extracellular form of TGF- β 1, which is specifically recognised by the CC antibody. This staining was confined to the stromal extracellular matrix, with no evidence for immunoreactivity in or around the epithelial tumour foci [27]. Interestingly, the increase in CC-immunoreactive extracellular TGF- β 1 did not correlate with the expression of ER, as it was evident in both ER positive and negative tumours. There was very little staining for TGF- β 2 or TGF- β 3 in any of the tumour sections and they were unaffected by three months of tamoxifen treatment. The only evidence for LC-reactive intracellular TGF- β 1 was seen in the peritumoural fibroblasts after treatment with tamoxifen, suggesting these cells as the likely site of synthesis, and supporting our previous observations *in vitro*.

As well as these observed increases in negative growth regulators after tamoxifen treatment, there is now compelling evidence to suggest a tamoxifen-mediated reduction in positive growth regulators,

such as insulin-like growth factors I and II. These are potent mitogens for breast cancer cells [29–31] which express type 1 IGF receptors for which both IGF I and II are functional ligands [32]. Levels of this receptor correlate loosely with levels of oestrogen receptor (ER) in cells [33], but ER negative cells still express significant levels of IGF receptor [34]. However, breast cancer cells *in vitro* generally do not express IGF I or IGF II mRNA, suggesting that these cells may participate in a paracrine rather than an autocrine pathway [35, 36]. This is supported by evidence that fibroblasts derived from breast tumour tissue express high levels of IGF I and II mRNA, with fibroblasts from benign tumours overexpressing IGF I mRNA whilst those from malignant tumours overexpress IGF II [37]. Therefore IGF's produced by fibroblasts may act upon neighbouring epithelial cells in a positive paracrine manner. Pollak and co-workers have demonstrated that tamoxifen treatment *in vivo* is associated with a reduction in serum IGF I levels [38] and proposed that as IGF I is not only a mitogen for breast cancer cells, but also stimulates the motility and metastatic potential of malignant cells [39, 40], the reduction in circulating IGF I might be an important component of tamoxifen's mode of action *in vivo*. Other experiments have suggested that tamoxifen may mediate this effect through a reduction in growth hormone secretion [41].

A recent *in vivo* study has addressed the relevance of the growth hormone reduction on circulating IGF I levels by examining IGF I expression in tissues which are common sites for breast cancer metastases (liver and lung). By using hypophysectomised rats with and without recombinant growth hormone replacement therapy, these authors demonstrated that the induction of IGF I was partly independent of pituitary function as significant reductions in IGF I expression after tamoxifen treatment were still seen in the hypophysectomised animals receiving growth hormone replacement [42]. This study did not allow any conclusions to be drawn as to whether the reduction in IGF I expression occurred in the parenchymal or stromal elements of these tissues.

The molecular mechanism of action of anti-oestrogens such as tamoxifen appears to be a complex

mixture of antagonism of the mitogenic action of oestradiol at the level of ER as well as a range of other activities from enzyme inhibition to growth factor modulation. The elucidation of these mechanisms is of potential importance in the treatment and chemoprevention of breast cancer. The move towards developing pure steroidal anti-oestrogens, devoid of any of the other beneficial effects of tamoxifen, continues apace, and determination of the quantitative contribution of each individual mechanism to the overall antineoplastic actions of anti-oestrogens is central to development of optimal pharmacological strategies.

References

1. Evans RM: The steroid and thyroid hormone receptor superfamily. *Science* 240: 889–895, 1988
2. Ham J, Parker MG: Regulation of gene expression by nuclear hormone receptors. *Curr Opin Cell Biol* 1: 503–511, 1989
3. Green S, Chambon P: Nuclear receptors enhance our understanding of transcriptional regulation. *Trends Genet* 4: 309–314, 1988
4. Coezy E, Borgna JL, Rochefort H: Tamoxifen and metabolites in MCF-7 cells: correlation between binding to oestrogen receptor and inhibition of cell growth. *Cancer Res* 42: 317–323, 1982
5. Green S, Chambon P: The oestrogen receptor: from perception to mechanism. In: Parker MG (ed) *Nuclear Hormone Receptors 2*. Academic Press, London, 1991, pp 15–34
6. Sutherland RL, Hall RE, Taylor IW: Cell proliferation kinetics of MCF-7 human mammary carcinoma cells in culture and effect of tamoxifen on exponentially growing and plateau phase cells. *Cancer Res* 43: 3998–4006, 1983
7. Sutherland RL, Watts CKW, Reunitz PC: Definition of two distinct mechanisms of action of antioestrogens on human breast cancer cell proliferation using hydroxytriphenylethylenes with high affinity for the oestrogen receptor. *Biochem Biophys Res Comm* 140: 523–529, 1986
8. Sutherland RL, Murphy LC, Ming San Foo, Green MD, Whybourne AN: High affinity anti-oestrogen binding site distinct from the oestrogen receptor. *Nature* 288: 273–275, 1980
9. O'Brian CA, Liskamp RM, Solomon DH, Weinstein IB: Inhibition of protein kinase C by tamoxifen. *Cancer Res* 45: 2462–2465, 1985
10. Berridge MJ, Irving RF: Inositol triphosphate, a novel second messenger in cellular signal transduction. *Nature* 312: 315–321, 1984
11. O'Brian CA, Liskamp RM, Solomon DH, Weinstein IB: Triphenylethylenes: A new class of Protein Kinase C inhibitors. *J Natl Cancer Inst* 76: 1243–1246, 1986
12. Su H-D, Mazzei GJ, Vogler WR, Kuo JF: Effect of tamoxifen, a non-steroidal antioestrogen, on phospholipid/calcium dependent protein kinase and phosphorylation of its endogenous substrate proteins from the rat brain ovary. *Biochem Pharmacol* 34: 3649–3653, 1985
13. Lam H-YP: Tamoxifen is a calmodulin antagonist in the activation of a cAMP phosphodiesterase. *Biochem Biophys Res Comm* 118: 27–32, 1984
14. Rowland MG, Parr IB, MacCague R, Jarman M, Goddard PM: Variation of the inhibition of calmodulin dependent cyclic AMP phospho-diesterase among analogues of tamoxifen: Correlation with cytotoxicity. *Biochem Pharmacol* 40: 283–289, 1990
15. Gulino A, Barrera G, Vacca A, Farina A *et al.*: Calmodulin antagonism and growth inhibitory activity of triphenylethylene antioestrogens in MCF-7 human breast cancer cells. *Cancer Res* 46: 6274–6278, 1986
16. Musgrove EA, Wakeling AE, Sutherland RL: Points of action of oestrogen antagonists and a calmodulin antagonist within the MCF-7 human breast cancer cell cycle. *Cancer Res* 49: 2398–2404, 1989
17. Nolvadex Adjuvant Trial Organisation: Controlled trial of tamoxifen as a single adjuvant agent in the management of early breast cancer. *Br J Cancer* 54: 608–611, 1987
18. Medical Research Council Scottish Trials Office: Adjuvant tamoxifen in the management of operable breast cancer. *Lancet* ii: 171–175, 1988
19. Early Breast Cancer Trialists Collaborative Group: Systemic treatment of early breast cancer by hormonal, cytotoxic or immune therapy. 133 randomised trials involving 31,000 recurrences and 24,000 deaths among 75,000 women. *Lancet* 339: 1–15 and 71–75, 1992
20. Brooks MD, Ebbs SR, Colletta AA, Baum M: Desmoid tumours treated with triphenylethylenes. *Eur J Cancer* 28: 1014–1018, 1992
21. Colletta AA, Wakefield LM, Howell FV, Roozendaal KEP, Danielpour D, Ebbs SR, Sporn MB, Baum M: Anti-oestrogens induce the secretion of active transforming growth factor beta from human foetal fibroblasts. *Br J Cancer* 62:405–409, 1990
22. Haggie JA, Sellwood RA, Howell A, Birch JM, Schor SL: Fibroblasts from relatives of patients with hereditary breast cancer show foetal-like behaviour *in vitro*. *Lancet* i: 1455–1457, 1987
23. Shor SL, Haggie JA, Durning P, Howell A, Smith L, Sellwood RA, Crowther D: Occurrence of a foetal fibroblast phenotype in familial breast cancer. *Int J Cancer* 37: 831–836, 1986
24. Glick AB, Danielpour D, Morgan D, Sporn MB, Yuspa SH: Induction and autocrine receptor binding of TGF- β 2 during terminal differentiation of primary mouse keratinocytes. *Mol Endocrinology* 4: 46–52, 1990
25. Knabbe C, Lippman ME, Wakefield LM, Flanders KC, Kasid A, Derynck R, Dickson RB: Evidence that transforming

- growth factor-beta is a hormonally regulated negative growth factor in human breast cancer. *Cell* 48: 417-428, 1987
26. Colletta AA, Wakefield LM, Howell FV, Danielpour D, Baum M, Sporn MB: The growth inhibition of human breast cancer cells by a novel synthetic progestin involves the induction of transforming growth factor beta. *J Clin Invest* 87: 277-283, 1991
 27. Butta A, MacLennan K, Flanders KC, Sacks NPM, Smith I, MacKinna A, Dowsett M, Wakefield LM, Sporn MB, Baum M, Colletta AA: Induction of transforming growth factor beta in human breast cancer *in vivo* following tamoxifen treatment. *Cancer Res* 52: 4261-4262, 1992
 28. Mansi JL, Smith IE, Walsh G, A'Hern RP, Harmer C, Sinnett HD, Trott PA, Fisher C, McKinna JA: Primary medical therapy for operable breast cancer. *Eur J Clin Oncol* 25: 1623-1627, 1989
 29. Myall Y, Shiu RPC, Bhaumick B: Receptor binding and growth promoting activity of insulin-like growth factors in human breast cancer cells (T47D) in culture. *Cancer Res* 44: 5486-5490, 1984
 30. Karey KP, Sirbasku DA: Differential responsiveness of human breast cancer cell lines MCF-7 and T47D to growth factors and 17β -oestradiol. *Cancer Res* 48: 4083-4092, 1988
 31. Cullen KJ, Lippman ME, Chow D, Hill S, Rosen N, Zwiebel JA: Insulin-like growth factor-II over-expression in MCF-7 cells induces phenotypic changes associated with malignant progression. *Mol Endocrinology* 6: 91-100, 1992
 32. Cullen KJ, Yee D, Sly WS *et al.*: Insulin-like growth factor receptor expression and function in human breast cancer. *Cancer Res* 50: 48-53, 1990
 33. Pekonen F, Paranen S, Makinen T, Rutanen E-M: Receptors for epidermal growth factor and insulin-like growth factor I and their relation to steroid receptors in human breast cancer. *Cancer Res* 48: 1343-1347, 1988
 34. Peyrat JP, Bonneterre J, Beuscart R, Djianne J, Demaille A: Insulin-like growth factor I receptors in human breast cancer and their relation to oestradiol and progesterone receptors. *Cancer Res* 48: 6429-6433, 1988
 35. Yee D, Cullen KJ, Paik S, Perdue JF, Hampton B, Schwartz A, Lippman ME, Rosen N: Insulin-like growth factor II mRNA expression in human breast cancer. *Cancer Res* 48: 6691-6696, 1988
 36. Yee D, Paik S, Lebovic G, Marcus R, Favoni R, Cullen KJ, Lippman ME, Rosen N: Analysis of IGF I gene expression in malignancy. Evidence for a paracrine role in human breast cancer. *Mol Endocrinol* 3: 509-517, 1989
 37. Cullen KJ, Smith HS, Hill S, Rosen N, Lippman ME: Growth factor messenger RNA expression by human breast fibroblasts from benign and malignant lesions. *Cancer Res* 51: 4978-4985, 1991
 38. Pollak M, Huynh HT, Pratt Lefebvre S: Tamoxifen reduces serum insulin-like growth factor I (IGF I). *Breast Cancer Research and Treatment* 22: 91-100, 1992
 39. Stracke ML, Kohn EC, Aznavoorian SA, Wilson LL, Salomon D, Krutzsch HC, Liotta LA, Schiffmann E: Insulin-like growth factors stimulate chemotaxis in human melanoma cells. *Biochem Biophys Res Comm* 153: 1076-1083, 1988
 40. Pollak M, Sem AW, Richard M, Tetenes E, Bell R: Inhibition of metastatic behaviour of murine osteosarcoma by hypophysectomy. *J Natl Cancer Inst* 84: 966-971, 1992
 41. Tannenbaum GS, Gurd W, Lapointe M, Pollak M: Tamoxifen attenuates pulsatile growth hormone secretion: Mediated in part by somatostatin. *Endocrinology* 130: 3395-3402, 1992
 42. Huynh HT, Tetenes E, Wallace L, Pollak M: *In vivo* inhibition of insulin-like growth factor I gene expression by tamoxifen. *Cancer Res* 53: 1727-1730, 1993