### Special articles

### Clinical significance of the sparteine/debrisoquine oxidation polymorphism

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Summary. The sparteine/debrisoquine oxidation polymorphism results from differences in the activity of one isozyme of cytochrome P450, the P450db1 (P450 IID1). The oxidation of more than 20 clinically useful drugs has now been shown to be under similar genetic control to that of sparteine/debrisoquine. The clinical significance of this polymorphism may be defined by the value of phenotyping patients before treatment. The clinical significance of such polymorphic elimination of a particular drug can be analyzed in three steps: first, does the kinetics of active principle of a drug depend significantly on P450db1?; second, is the resulting pharmacokinetic variability of any clinical importance?; and third, can the variation in response be assessed by direct clinical or paraclinical measurements? It is concluded from such an analysis that, in general, the sparteine/debrisoquine oxidation polymorphism is of significance in patient management only for those drugs for which plasma concentration measurements are considered useful and for which the elimination of the drug and/or its active metabolite is mainly determined by P450db1. At present, this applies to tricyclic antidepressants and to certain neuroleptics (e.g. perphenazine and thioridazine) and antiarrhythmics (e.g. propafenone and flecainide). Phenotyping should be introduced in to clinical routine under strictly controlled conditions to afford a better understanding of its potentials and limitations. The increasing knowledge of specific substrates and inhibitors of P450db1 allows precise predictions of drug-drug interactions. At present, the strong inhibitory effect of neuroleptics on the metabolism of tricyclic antidepressants represents the best clinically documented and most relevant example of such an interaction.

**Key words:** sparteine, debrisoquine, pharmacogenetics; oxidation polymorphism, clinical significance, oxidative drug metabolism, genetic control

Pharmacogenetics is concerned with the contribution of genetic factors to the variability of response to drugs. The impact of specific genetic factors is determined by the relative contribution of nongenetic constitutional factors, such as age, disease and environment.

To date, genetic factors affecting pharmacokinetics, in particular drug metabolism, have been the most extensively studied and best documented area of interest. The cDNAs for a number of important human drug receptors have recently been cloned (Frielle et al. 1987; Webb et al. 1987), and it should now be possible to study in a broader context the importance of genetic factors in pharmacodynamic variability.

Hepatic oxidation is by far the most important route of drug metabolism modifying the biological activity of drugs. The discovery of a common genetic polymorphism in the oxidation of debrisoquine and sparteine (Mahgoub et al. 1977; Eichelbaum et al. 1979a) created new interest in the role of pharmacogenetics in clinical pharmacotherapy. The population segregates into two phenotypes with respect to the oxidation of sparteine and debrisoquine. In Caucasians about 7% are classified as phenotypically poor metabolizers (PM) and the remainder as extensive metabolizers (EM) (Steiner et al. 1988b). The frequency of the PM phenotype appears to be markedly lower in non-Caucasian

Biotransformation	Consequence for poor metabolizers	Drug	Reference
$D \Rightarrow M_a$	1. Accumulation of drug, D	Beta-blockers	
↓ Mb		Metoprolol Timolol Bufuralol	Lennard et al. 1983 Lewis et al. 1985 Dayer et al. 1985
		Antiarrhythmics	
		Sparteine N-propylajmaline Propafenone Flecainide	Eichelbaum et al. 1979b Zekorn et al. 1985 Siddoway et al. 1987 Beçkmann et al. 1988
		Tricyclic antidepressants	
		Nortriptyline Desipramine Clomipramine	Bertilsson et al. 1980 Bertilsson and Åberg-Wistedt 1983 Balant-Gorgia et al. 1986**
		Neuroleptics	
		Perphenazine Thioridazine	Dahl-Puustinen et al. (in press) von Bahr et al. (in press)
		Miscellaneous	
		Debrisoquine 4-Hydroxyamphetamine Phenformin Amiflamine Perhexiline Dextromethorphan Guanoxan Indoramin Methoxyphenamine CGP 1S 210G	Sloan et al. 1983 Kitchen et al. 1979** Oates et al. 1983 Alvan et al. 1984 Cooper et al. 1987 Schmid et al. 1985** Sloan et al. 1978** Pierce et al. 1987 Roy et al. 1985 Gleiter et al. 1985
	2. Reduced formation of active metabolite, $M_a$	Encainide (active metabolite O-desmethylencainide)	Wang et al. 1984
	3. Increased formation of toxic metabolite, M <sub>b</sub>	No known examples	
$D \Rightarrow M_a$	4. Reduced formation of	Codeine	Dayer et al. 1988
*↓ Mb	active metabolite $M_b$	(active metabolite: morphine)	
$\begin{array}{l} D \Rightarrow M_a \\ \downarrow^* & \downarrow^* \\ M_b & M_c \end{array}$	5. Accumulation of active metabolite, $M_a$	Imipramine (active metabolite: desipramine)	Brøsen et al. 1986 a + b
$\begin{array}{ccc} D & \stackrel{*}{\Rightarrow} & M_a \\ \stackrel{*}{\downarrow} & \stackrel{\downarrow}{\downarrow} & \\ M_b & M_c \end{array}$	6. Accumulation of drug, D, and active metabolite, $M_a$	Amitriptyline (active metabolite: nortriptyline)	Mellström et al. 1983 Baumann et al. 1986

Table 1. Pharmacokinetic consequences of polymorphism for drugs eliminated via the sparteine/debrisoquine oxygenase, P450db1

 $\Rightarrow$ : important pathway;  $\rightarrow$ : less important pathway; \*: pathway catalyzed by the P450 which oxidizes sparteine and debrisoquine; \*\*: only urine data available

populations (Nakamura et al. 1985; Lou et al. 1987; Iyun et al. 1986). PM are homozygous for an autosomal recessive allele, and EM comprise heterozygotes and homozygous dominants (Evans et al. 1980; Steiner et al. 1985). It has recently been shown that the so-called recessive allele exists in several forms (Gonzalez et al. 1988). It is not yet clear if the existence of multiple mutated alleles can explain some of the pronounced variability in drug clearance seen within the EM phenotype (Brøsen et al. 1986a). The sparteine/debrisoquine oxidation polymorphism is a reflection of differences in the activity of a specific isozyme of cytochrome P-450, P450db1 (or P450IID1, Nebert et al. 1987; or P450bufI, Zanger et al. 1988). Recent studies strongly suggest that P450db1 is absent altogether in PM, but other P450s may contribute to a minor extent to the oxidation of at least some compounds that are mainly dependent on P450db1 for elimination (Boobis et al. 1983; Zanger et al. 1988; Gonzalez et al. 1988).

The sparteine/debrisoquine oxidation polymorphism has a strong impact on the metabolism of several drugs in patients, notably the tricyclic antidepressants (Bertilsson et al. 1980, Mellström et al. 1983; Bertilsson and Åberg-Wistedt 1983; Brøsen et al. 1986a, b), metoprolol (Lennard et al. 1983), propafenone (Siddoway et al. 1987), flecainide (Beckmann et al. 1988), and certain neuroleptics (Dahl-Puustinen et al., in press; von Bahr et al., in press).

Another drug oxidation polymorphism that has been revealed in studies of the 4-hydroxylation of S-mephenytoin (Küpfer and Preisig 1984) is related to the activity of a different isozyme of P450 (P450meph, Meyer et al. 1986). P450meph also oxidizes mephobarbital (Küpfer and Branch 1985), hexobarbital (Knodell et al. 1988) and is partially responsible for the elimination of diazepam and N-desmethyldiazepam (Bertilsson et al., in press). The broader clinical significance of the mephenytoin polymorphism has yet to be established.

The discovery of genetic polymorphism of drug oxidation has had a considerable impact on experimental and biochemical research on drug metabolism. In contrast, its impact on the clinical use of therapeutics is still limited. The clinical perspective of the sparteine/debrisoquine oxidation polymorphism is reviewed here.

# *Clinical significance of the sparteine/debrisoquine oxidation polymorphism*

Interindividual variation in drug metabolism is inevitable, but is often inconvenient from a clinical point of view. Genetic polymorphism is a special case, in which most variability is due to genetic inheritance (Steiner et al. 1985). This should not be regarded as a particular disadvantage, since it should be possible to predict the optimal treatment or dosage from the patient's phenotype. The significance of the sparteine/debrisoquine oxidation polymorphism in clinical use of a particular drug would be demonstrated if the value were accepted of routinely phenotyping patients prior to treatment. The clinical significance of polymorphic drug oxidation in relation to a particular drug would require, first, that the kinetics of the drug was significantly dependent on P450db1, second, that the resulting pharmacokinetic variability was of clinical importance and, third, that the interindividual variability in response could not be determined by direct clinical or paraclinical measurements. The latter two points are in fact characteristics of drugs for which plasma level monitoring is also considered useful.

**Table 2.** Drugs which have been shown not to be eliminated via the sparteine/debrisoquine oxygenase (P450db1)

Drug	Reference
Amobarbital	Inaba et al. 1980
Antipyrine	Danhof et al. 1981
Metaqualone	Oram et al 1982
Ethinyloestradiol	Back et al. 1984
Theophylline	Dahlqvist et al. 1984
Propranolol <sup>a</sup>	Lennard et al. 1984
Mephenytoin	Küpfer and Preisig 1984
Maprotiline	Gabris et al. 1985
Carbamazepine	Eichelbaum et al. 1985
Tolbutamide	Miners et al. 1985
Pinacidil	Shaheen et al. 1986
Quinidine Nifedipine Phenytoin	Mikus et al. 1986 Beerahee et al. 1987 Steiner et al. 1987
Prazosin	Lennard et al. 1988
Haloperidol	Gram et al. 1988
Codeine <sup>b</sup>	Dayer et al. 1988
Cyclosporine	Kronbach et al. 1988 <sup>c</sup>
Diazepam N-desmethyldiazepam	Bertilsson et al. (in press)
Midazolam	Kronbach et al. (personal communication) <sup>c</sup>

<sup>a</sup> 4'-hydroxylation of propranolol, a minor pathway, is impaired in poor metabolizers

<sup>b</sup> O-demethylation, a minor but important pathway is impaired in poor metabolizers (see text)

<sup>2</sup> Demonstrated in human liver microsome preparations in vitro

### Drug elimination and P450db1

Sparteine, debrisoquine or dextromethorphan (Schmid et al. 1985) may be used for the phenotyping procedure. A PM subject is defined as one in whom the metabolic ratio (MR = amount of parent compound/amount of metabolite) in an 8-12 h urine sample is >20 for sparteine (Eichelbaum et al. 1982), >12.6 for debrisoquine (Evans et al. 1980) or >0.3 for dextromethorphan (Schmid et al. 1985).

The pharmacokinetic significance of P450db1 depends on its relative contribution to the overall elimination (clearance) of the drug (Jackson et al. 1986). The phenotype can predict the overall elimination of a drug or active metabolite when they are substrates for P450db1, and elimination via alternative isozymes of P450, non-oxidative metabolism or excretion unchanged does not occur to any significant extent in the EM. These criteria are largely met for the drugs listed in Table 1. For the drugs listed in Table 2 no such difference in total clearance between EM and PM has been shown. It may be inferred, therefore, that these drugs are oxidized by dif-



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Fig. 1. Apparent clearance of imipramine and desipramine after a single oral dose of 100 mg of both drugs on separate occasions. 100 mg in 6 rapid extensive metabolizers  $(\bullet)$ , 6 slow extensive metabolizers  $(\bullet)$ , 6 slow extensive metabolizers  $(\bullet)$ , 10 and 6 poor metabolizers  $(\bullet)$ . (Data from Brøsen et al. 1986a)

ferent isozymes of P450, and, in consequence, the sparteine/debrisoquine oxidation polymorphism should not be expected to be of any significance for their clinical use. Codeine, however, is an exception to this statement; in EM a maximum of 10% of codeine is O-demethylated to morphine, which is believed to be the active analgesic component. O-Demethylation is catalyzed by P450db1 (Dayer et al. 1988), and this explains why codeine has no analgesic effect in PM (P.Dayer, personal communication).

Biotransformation of the drugs listed in Table 1 is markedly reduced in PM, and total clearance typically is about five times lower in PM than in EM (e.g. desipramine; Fig. 1). the difference is less pronounced when the drug is only partly oxidized by P450db1 (e.g. imipramine; Fig. 1).

Steiner (1985) showed that genetic inheritance accounted for as much as 80% of the total variability in debrisoquine MR. This is an unusually high degree of genetic contribution, but it is important to realize that a number of confounding factors in the individual patient may alter the relative importance of P450db1 in the overall elimination of a given drug (Table 3).

Selective induction of P450db1 in EM (1.1, Table 3) will exaggerate the difference in metabolic clearance between phenotypes. However, enzyme inducers have only a limited effect on the elimination of drugs that are largely dependent on P450db1 for their clearance (Eichelbaum et al. 1986). Conversely, induction of alternative drug metabolising enzymes will reduce the relative contribution of P450db1, and so may obscure the effect of the genetic polymorphism (2.1, Table 3). This has been suggested as an explanation for the effect of cigarette smoking on amitriptyline demethylation (Mellström et al. 1983).

The phenotypic difference may also be reduced or abolished by functional impairment of P450db1 in EM (Table 3); for example, quinidine is a very potent competitive inhibitor of sparteine oxidation in human liver microsomes in vitro (Otton et al. 1984) and the in vivo metabolism of sparteine and debrisoquine is almost completely abolished in patients treated with quinidine (Brinn et al. 1986; Speirs et al. 1986; Brøsen et al. 1987). Hence, an EM would appear to change to phenotypical PM, and the underlying genetic variation in metabolism would be completely lost (2.2, Table 3). Similar findings have been reported during treatment with thioridazine, levomepromazine, and propafenone (Syvählahti et al. 1986; Siddoway et al. 1987). Phenotyping during treatment with a potent inhibitor is uninformative.

The activity of P450db1 is also impaired by tricyclic antidepressants and some neuroleptics, but not to such an extent that the EM phenotype appears to change to PM (Nordin et al. 1985; Brøsen et al. 1986b; Gram et al., 1989). Such weak inhibitors may still contribute to the pharmacokinetic variability found in EM subjects.

Another functional characteristic of P450db1 appears to be non-linear kinetics for some drugs in EM but linear kinetics in PM (2.3, Table 3). With oral dosing this probably reflects saturation of P450db1 during the first pass through the liver (Brøsen and Gram 1988). The apparently much higher Michaelisconstant,  $K_m$ , for bufuralol-oxidation found in PM than EM (Dayer et al. 1984) probably reflects the activity of a different isozyme of P450, for which saturation would not be a problem at the drug concentrations or doses yielding saturation in EM. As a consequence, the difference in clearance between phenotypes is reduced but not extinguished when

**Table 3.** Pharmacokinetic factors that theoretically can alter the relative significance of the sparteine/debrisoquine oxygenase (P450db1) for the elimination of a particular drug

1. Increased significance of P450db1

- 1.1. Selective induction of P450db1 in extensive metabolizers (not important)
- 2. Decreased significance of P450db1
- 2.1. Induction of alternative P450's
- 2.2. Inhibition of P450db1
- 2.3. Saturation of P450db1

the dose is increased, as has been shown for propafenone (Fig. 2; Siddoway et al. 1987).

Stereoselective metabolism is another important characteristic of P450db1. Metoprolol is administered as the racemate, and in EM the inactive R-enantiomer is metabolized faster than the active S-enantiomer, whereas metabolism is not stereoselective in PM (Lennard et al. 1983). Accordingly, the difference in plasma concentrations between phenotypes is smaller for the active S-enantiomer than for the inactive R-enantiomer (Fig. 3).

For most of the drugs listed in Table 1 it is well documented that their oxidation co-segregates with that of sparteine and debrisoquine. However, the demonstration of a genetic polymorphism in drug oxidation requires both family and population studies, which so far have been carried out only with a limited number of these compounds.

## *Role of the sparteine/debrisoquine oxidation polymorphism in drug therapy*

Most studies in this field have established an association between a polymorphic drug oxidation phenotype and some pharmacokinetic measurements for a given drug. Before the sparteine/debrisoquine oxidation polymorphism can be considered to be of clinical significance in drug therapy, however, several additional characteristics of the drug should be considered.

Several of the drugs in Table 1 with established polymorphic metabolism are either obsolete or unimportant. As a general rule, a genetic polymorphism will have clinical relevance only for drugs that are widely available.

Second, the pharmacodynamic properties of the drug must be considered. Some drugs have a wide therapeutic plasma concentration range (i.e. a large therapeutic index), and may therefore be prescribed in standard doses without problems, evens if genetic polymorphism in their metabolism results in significant pharmacokinetic variability. Some drugs may have a small therapeutic index, but their dose is easily titrated on the basis of clinical or paraclinical effects. In both cases neither phenotyping nor direct assessment of the pharmacokinetic variability is necessary. The beta-adrenoceptor antagonists, metoprolol and timolol may be considered to fulfil both these requirements (Table 4), although it is not yet known whether a chronically elevated plasma level of a beta-adrenoceptor antagonist during long-term treatment may harm the PM patient.

The potential clinical importance of the sparteine/debrisoquine oxidation polymorphism is asso-



Fig. 2. Relationship between daily proparenone dose and trough plasma concentration during steady-state in 13 extensive metabolizers ( $\bigcirc$ ) and 4 poor metabolizers ( $\bigcirc$ ). After Siddoway et al. 1987 (with permission from author and publisher)  $\delta \times \pm SEM$ 

ciated with those drugs for which assessment of pharmacokinetic variability (by drug level monitoring) is important. Such drugs are characterized by a low therapeutic index and the lack of means to titrate the dose on the basis of immediate measures of effect. For drugs with a metabolism that cosegrate with the sparteine/debrisoquine oxidation polymorphism, this applies to tricyclic antidepressants (Gram et al. 1984), neuroleptics (Dahl 1986), and possibly to flecainide and propafenone (Table 4).

For tricyclic antidepressants there are two patient groups at risk (Fig. 4). One group comprises PM and slow EM subjects. If given the standard recommended doses these patients will develop toxic plasma concentrations (Fig. 4). The other group comprises rapid EM, who suffer therapeutic failure because the attained plasma concentrations are far too low (Fig. 4). At least 20–30% of patients belong to these risk groups (Brøsen et al. 1986b). Side effects during the therapeutic use of tricyclic antidepressants are rarely, if ever, life-threatening, but they are often so unpleasant (dry mouth, sedation and tremor) that the treatment is stopped. Toxic reactions may be misinterpreted as symptoms of depression or



Steady-state concentration

Fig. 4. Theoretical distribution of steady-state concentration during treatment with a fixed dose of a drug of which the metabolism is exclusively dependent on sparteine/debrisoquine oxygenase (P450db1). Higher mode poor metabolizers; lower mode extensive metabolizers.  $\Box$ , therapeutic failure;  $\Box$ , improvement;  $\Box$ , toxicity

may occur as clear CNS-toxicity at drug levels only 50–100% above the therapeutic plasma concentration (Preskorn, in press). The clinical picture both in the rapidly and slowly metabolizing patients is there-

fore much the same: recurrent depressive episodes not responding to treatment due to lack of dose individualization (Bertilsson et al. 1985; Sjöqvist and Bertilsson 1986).

In a recent study, the dose of imipramine required to produce a therapeutic plasma concentration (imipramine plus desipramine: 700-900 nM) ranged from 50 to 400 mg per day in 32 EM patients, and in two PM it was 50 mg per day (Brøsen et al. 1986b). The therapeutic dose did not exhibit distinct bimodality but the PM subjects represented the lower extreme. The correlation between the dose and the sparteine or debrisoquine MR was relatively weak, although statistically significant. Phenotyping was useful mainly in separating EM and PM, but it coult not replace drug plasma concentration monitoring to achieve the target concentration. However, dosing according to phenotype or metabolic ratio would have yielded less variability in steady state level than use of a standard dosing scheme. The weak correlation was due to the activity of alternative isozymes of P450 for demethylation, and possibly for hydroxylation, and confounding factors such as dose-dependent kinetics (Table 4) and nonmetabolic variation in MR.

## Drug-drug interactions related to selective inhibition of the P450db1 isozyme

The testing of different drugs for their ability competitively to inhibit the oxidation of sparteine, debrisoquine, desipramine or bufuralol in vitro has proven to be a useful primary screening procedure (Boobis et al. 1983; Otton et al. 1984; Inaba et al. 1985; von Bahr et al. 1985; Fonne-Pfister and Meyer 1988). A compound not inhibiting oxidation of the model drug can be excluded as a substrate for P450db1, whereas a compound that does competitively inhibit it may be a substrate of P450db1. Some of these inhibitors, e.g. quinidine, propafenone and certain neuroleptics (Table 5) are very potent, with inhibitor constant values (K<sub>i</sub>) in the nanomolar or low micromolar range.

Thus, the P450db1 may be the site for important and predictable drug-drug interactions. This aspect may turn out to be as important clinically as the genetic polymorphism. Several studies in vitro and in vivo have shown that quinidine is a selective inhibitor of the P450db1 (Table 5). Since P450db1 is absent from the livers of PM (Zanger et al. 1988), it is understandable that the relative impairment of oxidation is more pronounced in EM than in PM (Brinn et al. 1986; Steiner et al. 1988a). The picture may become even more complicated if a potent inhibitor like propafenone (Tables 1 and 5; Fig. 2) is also eliminated by P450db1, because PM then develop much higher inhibitor concentrations than EM when the same dose is given. At higher inhibitor concentrations there may be significant inhibition of other P450 isozymes as well. It is likely that this could explain the drug concentration-related adverse reactions in a PM during combined treatment with metoprolol and propafenone (Wagner et al. 1987b).

When potent inhibitors like quinidine or propafenone are added to treatment with a polymorphically

 
 Table 4. Non-pharmacokinetic factors determining the clinical significance of the sparteine/debrisoquine oxidation polymorphism for drug known to be eliminated via P450db1

Drug	Important and widely available	Plasma concentration monitoring recommended?	Conclusion: possible clinical significance
Metoprolol	Yes	No	No <sup>a</sup>
Timolol	Yes	No	No
Bufuralol	No	No	No
Sparteine	No	No	No
N-Propy- lajmaline	No	Yes <sup>b</sup>	No
Propafenone	Yes	Yes <sup>c</sup>	Yes
Flecainide	Yes	Yes	Yes
Nortriptyline	Yes	Yes <sup>d</sup>	Yes
Desipramine	Yes	Yes <sup>d</sup>	Yes
Clomipramine	Yes	Yes <sup>d</sup>	Yes
Imipramine	Yes	Yes <sup>d</sup>	Yes
Amitriptyline	Yes	Yes <sup>d</sup>	Yes
Debrisoquine	No	No	No
4-Hydroxy-			
amphetamine	No	No	No
Phenformin	No	No	No
Amiflamine	No	No	No
Perhexilline	No	No	No
Dextro-			
methorphan	Yes	No	No
Guanoxan	No	No	No
Indoramin	No	No	No
Methoxy-	2.5	<b>N</b> 7	• •
phenamine	No	No	No
CGP 1S 210G	No	No	No
Encainide	No	No	No
Perphenazine	Yes	Yes <sup>e</sup>	Yes
Thioridazine	Yes	Yes <sup>e</sup>	Yes

<sup>a</sup> see Clark et al. 1984; <sup>b</sup> for discussion, see Zekorn et al. 1985; <sup>c</sup> see Siddoway et al. 1987; <sup>d</sup> see Gram et al. 1984; <sup>e</sup> see Dahl 1986

oxidized drug (Table 1), the pharmacokinetics are indistinguishable from those observed in PM, which means in practice that the plasma concentration may increase about five-fold (Wagner et al. 1987a; Brøsen and Gram, accepted).

The clinical significance of such interactions depends on the frequency of the drug combination, the role of P450db1 in overall elimination of the drug, the potency of the inhibition, the clinical consequences of a raised plasma drug level (therapeutic index), and the feasibility of clinical dose titration.

It was reported several years ago that neuroleptics strongly inhibited, the metabolism of tricyclic antidepressants (Gram and Overø 1972; Gram 1975) and this has subsequently been confirmed by several

Inhibitor	Inhibition					
	In vitro			In vivo		
	Substrate	K <sub>i</sub> nM	Reference	Substrate	Reference	
Antiarrhythmics:						
Quinidine	Sparteine Desipramine Bufuralol Codeine	60 270 150 <sup>a</sup> 15	Otton et al. 1984 von Bahr et al. 1985 Zanger et al. 1988 Dayer et al. 1988	Metoprolol Sparteine Debrisoquine Desipramine	Leeman et al. 1986 Brinn et al. 1986 Brøsen et al. 1987 Steiner et al. 1988	
Propafenone	Debrisoquine Bufuralol	700 50 <sup>a</sup>	Siddoway et al. 1987 Kroemer et al. 1989	Debrisoquine Metoprolol	Siddoway et al. 1987 Wagner et al. 1987a	
Neuroleptics:						
Chlorpromazine Haloperidol	Sparteine Sparteine	7000 1000	Inaba et al. 1985 Inaba et al. 1985	Nortriptyline Imipramine Sparteine	Gram and Overø 1972 Gram et al. 1989	
Thioridazine	Desipramine	750	von Bahr et al. 1985	Desipramine Debrisoquine	Hirschowitz et al. 1983 Syvählahti et al. 1986	
Fluphenazine	Bufuralol	1030 <sup>a</sup>	Fonne-Pfister and Meyer 1988	Imipramine	Siris et al. 1982	

 Table 5. Drugs known to be potent inhibitors of P450db1 function in human liver in vitro and in vivo

<sup>a</sup> Kinetics based on a cumene hydroperoxide-mediated reaction

groups (review by Gram and Brøsen, in press). Neuroleptics are frequently added to antidepressant treatment, in particular in elderly, delusional depressed patients, and the interaction appears to be one of the most clinically important drug-drug interactions in relation to the P450db1 isozyme (Gram et al. 1984). Codeine is a special case (Table 2), because no morphine will be formed in EM during combined treatment with quinidine and presumably other potent inhibitors (P.Dayer, personal communication).

#### Conclusion

The sparteine/debrisoquine oxidation polymorphism is a determinant of the extremes in pharmacokinetic variability of several clinically useful drugs. This monogenically determined variability becomes clinically significant only in those cases where pharmacokinetic variability is considered important relative to the efficacy and clinical safety of the drug. This mean in essence that it is applicable to those drugs for which plasma concentration monitoring is considered useful, and at present the list includes tricyclic antidepressants, certain neuroleptics and possibly flecainide and propafenone. Codeine and possibly other prodrugs, which depend on P450db1 for their conversion to an active metabolite, are exceptions to this general rule. The mere demonstration of genetic polymorphism in the oxidation of a new drug should not of itself lead to discontinuation of its development (Balant et al., in press), nor should detection of genetic polymorphism in the oxidation of a drug which is already on the market lead to discontinuation of its clinical use.

Phenotyping with sparteine, debrisoquine or dextromethorphan before treatment with a tricyclic antidepressant may serve as a safe and simple test to enable clinicians, to select an appropriate initial dose and to reduce interindividual variability in drug concentrations. The test result (MR) is usually not sufficiently precise to replace plasma concentration monitoring, but the optimal strategy for blood sampling (time to reach steady-state, dose-dependent kinetics and drug interactions) is different in EM and PM.

Since the test result will apply to several tricyclic antidepressants and phenotyping has to be performed only once in a life-time, the approach should be particularly useful in patients with recurrent depression. Analogous considerations may apply to antipsychotic treatment with neuroleptics, and antiarrhythmic treatment with propafenone and flecainide. As for any clinical test, its introduction into routine use should be implemented under continuous surveillance in a follow-up programme that would ultimately enable our understanding of its potentials and limitations to be refined.

The relative substrate specificity of different cytochrome P450 isozymes affords an opening for new and more rational research on drug-drug interactions. Studies in vitro with microsomal preparations from human liver may be particularly useful in initial screening for interactions of possible clinical importance.

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#### References

- Alván G, Grind M, Graffner C, Sjöqvist F (1984) Relationship of N-demethylation of amiflamine and its metabolite to debrisoquine hydroxylation polymorphism. Clin Pharmacol Ther 36: 515-519
- Back DJ, Maggs JL, Purba HS, Newby S, Park BK (1984) 2-Hydroxylation of ethinyloestradiol in relation to the oxidation of sparteine and antipyrine. Br J Clin Pharmacol 18: 603-607
- von Bahr C, Spina E, Birgersson C (1985) Inhibition of desmethylimipramine 2-hydroxolation by drugs in human liver microsomes. Biochem Pharmacol 14: 2501–2505
- von Bahr C, Guengerich FP, Morin G, Nordin C (1989) The use of human liver banks in pharmacogenetic research. In: Dahl SG, Gram LF (eds) Clinical pharmacology in psychiatry: From molecular studies to clinical reality. Springer, Berlin Heidelberg New York Tokyo (in press)
- Balant LP, Gundert-Remy U, Boobis AR, von Bahr C (1989) Genetic polymorphism in drug metabolism: Relevance for the development of new drugs. Eur J Clin Pharmacol (in press)
- Balant-Gorgia AE, Balant LP, Genet C, Dayer P, Aeschlimann JM, Garrone G (1986) Importance of oxidative polymorphism and levomepromazine treatment on the steady-state blood concentrations of clomipramine and its major metabolites. Eur J Clin Pharmacol 31: 449-455
- Baumann P, Jonzier-Perey M, Koeb L, Küpfer A, Tinguely D, Schöpf J (1986) Amitriptyline pharmacokinetics and clinical response: II. Metabolic polymorphism assessed by hydroxylation of debrisoquine and mephenytoin. Int Clin Psychopharmacol 1: 102–112
- Beckmann J, Hertrampf R, Gundert-Remy U, Mikus G, Gross AS, Eichelbaum M (1988) Is there a genetic factor in flecainide toxicity? Br Med J 297: 1316
- Beerahee M, Wilkins MR, Jack DB, Beevers DG, Kendall MJ (1987) Twelve hour (through) plasma nifedipine concentrations during chronic treatment with nifedipine retard. Eur J Clin Pharmacol 32: 347–349
- Bertilsson L, Eichelbaum M, Mellström B, Säwe J, Schulz H-U, Sjöqvist F (1980) Nortriptyline and antipyrine clearance in relation to debrisoquine hydroxylation in man. Life Sci 27: 1673-1677
- Bertilsson L, Åberg-Wistedt A (1983) The debrisoquine hydroxylation test predicts steady-state plasma levels of desipramine. Br J Clin Pharmacol 15: 388-390
- Bertilsson L, Åberg-Wistedt A, Gustafsson LL, Nordin C (1985) Extremely rapid hydroxylation of debrisoquine: A case report with implication for treatment with nortriptyline and other tricyclic antidepressants. Ther Drug Monit 7: 478-480
- Bertilsson L, Henthorn TK, Sanch E, Tybring G, Säwe J, Villén T (1989) Importance of genetic factors in the regulation of diazepam metabolism: Relationship to S-mephenytoin, but not debrisoquine hydroxylation phenotype. Clin Pharmacol Ther (in press)

- Boobis AR, Murray S, Kahn GC, Robertz GM, Davies DS (1983) Substrate specificity of the form of cytochrome P-450 catalyzing the 4-hydroxylation of debrisoquine in man. Mol Pharmacol 23: 474-481
- Brinn R, Brøsen K, Gram LF, Haghfelt T, Otton SV (1986) Sparteine oxidation is practically abolished in quinidine-treated patients. Br J Clin Pharmacol 22: 194–197
- Brøsen K, Otton SV, Gram LF (1986a) Imipramine demethylation and hydroxylation: Impact of the sparteine oxidation phenotype. Clin Pharmacol Ther 40: 543–549
- Brøsen K, Klysner R, Gram LF, Otton SV, Bech P, Bertilsson L (1986b) Steady-state concentrations of imipramine and its metabolites in relation to the sparteine/debrisoquine polymorphism. Eur J Clin Pharmacol 30: 679-684
- Brøsen K, Gram LF, Haghfelt T, Bertilsson L (1987) Extensive metabolizers of debrisoquine become poor metabolizers during quinidine treatment. Pharmacol Toxicol 60: 312-314
- Brøsen K, Gram LF (1988) First-pass metabolism of imipramine and desipramine: Impact of the sparteine oxidation phenotype. Clin Pharmacol Ther 43: 400-406
- Clark DWJ, Morgan AKW, Waal-Manning H (1984) Adverse effects from metoprolol are not generally associated with oxidation status. Br J Clin Pharmacol 18: 965–967
- Cooper RG, Evans DAP, Price AH (1987) Studies on the metabolism of perhexiline in man. Eur J Clin Pharmacol 32: 569-576
- Dahl SG (1986) Plasma level monitoring of antipsychotic drugs. Clinical utility. Clin Pharmacokinet 11: 36-61
- Dahl-Puustinen ML, Lidén A, Alm C, Bertilsson L (1989) Disposition of perphenazine is related to the polymorphic debrisoquine hydroxylation in man. Clin Pharmacol Ther (in press)
- Dahlqvist R, Bertilsson L, Birkett DJ, Eichelbaum M, Säwe J, Sjöqvist F (1984) Theophylline metabolism in relation to antipyrine, debrisoquine, and sparteine metabolism. Clin Pharmacol Ther 35: 815-821
- Danhof M, Idle JR, Teunissen MWE, Sloan TP, Breimer DD, Smith RL (1981) Influence of the genetically controlled deficiency in debrisoquine hydroxylation on antipyrine metabolite formation. Pharmacology 22: 349–358
- Dayer P, Balant L, Kupfer A, Striberni R, Leemann T (1985) Effect of oxidative polymorphism (debrisoquine/sparteine type) on hepatic first-pass metabolism of bufuralol. Eur J Clin Pharmacol 28: 317-320
- Dayer P, Desmeules J, Leemann T, Striberni R (1988) Bioactivation of the narcotic drug codeine in human liver is mediated by the polymorphic monooxygenase catalyzing debrisoquine 4hydroxylation. Biochem Biophys Res Commun 152: 411-416
- Dayer P, Gasser R, Gut J, Kronbach T, Robertz GM, Eichelbaum M, Meyer UA (1984) Characterization of a common genetic defect of cytochrome P450 function (debrisoquine-sparteine type of polymorphism). Biochem Biophys Res Commun 125: 374-380
- Eichelbaum M, Spannbrucker N, Steincke B, Dengler HJ (1979 a) Defective *N*-oxidation of sparteine in man: A new pharmacogenetic defect. Eur J Clin Pharmacol 16: 183-187
- Eichelbaum M, Spannbrucker N, Dengler HJ (1979b) Influence of the defective metabolism of sparteine on its pharmacokinetics. Eur J Clin Pharmacol 16: 189-194
- Eichelbaum M, Bertilsson L, Säwe J, Zekorn C (1982) Polymorphic oxidation of sparteine and debrisoquine: Related pharmacogenetic entities. Clin Pharmacol Ther 31: 184–186
- Eichelbaum M, Tomson T, Tybring G, Bertilsson L (1985) Carbamazepine metabolism in man. Induction and pharmacogenetic aspects. Clin Pharmacokinet 10: 80-90
- Eichelbaum M, Mineshita S, Ohnhaus EE, Zekorn C (1986) The influence of enzyme induction on polymorphic sparteine oxidation. Br J Clin Pharmacol 22: 49-53

- Evans DAP, Mahgoub A, Sloan TP, Idle JR, Smith RL (1980) A family and population study of genetic polymorphism of debrisoquine oxidation in a white British population. J Med Genet 17: 102-105
- Fonne-Pfister R, Meyer UA (1988) Xenobiotic and endobiotic inhibitors of cytochrome P-450db1 function, the target of the debrisoquine/sparteine type polymorphism. Biochem Pharmacol 37: 3829-3855
- Frielle T, Collins S, Daniel KW, Caron MG, Lefkowitz RJ (1987) Cloning of the cDNA for the human  $\beta_1$ -adrenergic receptor. Proc Natl Acad Sci USA 84: 7920-7924
- Gabris G, Baumann P, Jonzier-Perey M, Bosshart P, Woggon B, Küpfer A (1985) N-Methylation of maprotiline in debrisoquine/mephenytoin-phenotyped depressive patients. Biochem Pharmacol 34: 409-410
- Gleiter CH, Aichele G, Nilsson E, Hengen N, Antonin KH, Bieck PR (1985) Discovery of altered pharmacokinetics of CGP 15 210 G in poor hydroxylators of debrisoquine during early drug development. Br J Clin Pharmacol 20: 81-84
- Gonzalez FJ, Skoda R, Kimura S, Umeno M, Zanger UM, Nebert DW, Gelboin HV, Hardwick JP, Meyer UA (1988) Characterization of the common genetic defect in humans deficient in debrisoquine metabolism. Nature 331: 442-446
- Gram LF (1975) Effects of perphenazine on imipramine metabolism in man. Psychopharmacol Commun 1: 165-175
- Gram LF, Brøsen K (1989) Inhibitors of the microsomal oxidation of psychotropic drugs: Selectivity and clinical significance.
  In: Dahl SG, Gram LF (eds) Clinical pharmacology in psychiatry: From molecular studies to clinical reality. Springer, Berlin Heidelberg New York Tokyo (in press)
- Gram LF, Debruyne D, Caillard V, Boulenger JP, Lacotte J, Moulin M, Zarifian E (1989) Substantial rise in sparteine metabolic ratio during haloperidol treatment. Br J Clin Pharmacol 27: 272-275
- Gram LF, Kragh-Sørensen P, Kristensen CB, Møller M, Pedersen OL, Thayssen P (1984) Plasma level monitoring of antidepressants: Theoretical basis and clinical application. In: Usdin E, Åsberg M, Bertilsson L, Sjöqvist F (eds) Frontiers in biochemical and pharmacological research in depression. Raven Press, New York, pp 399-411
- Gram LF, Overø KF (1972) Drug interaction: Inhibitory of neuroleptics on metabolism of tricyclic antidepressants in man. Br Med J 163: 463-465
- Hirschowitz J, Bennet JA, Semian FP, Garber D (1983) Thioridazine effect on desipramine plasma levels. J Clin Psychopharmacol 3: 376–379
- Inaba T, Jurima M, Mahon WA, Kalow W (1985) In vitro studies of two isozymes of human liver cytochrome P-450-mephenytoin hydroxylase and sparteine monooxygenase. Drug Metab Dispos 13: 443-448
- Inaba T, Otton SV, Kalow W (1980) Deficient metabolism of sparteine and debrisoquine. Clin Pharmacol Ther 27: 547-549
- Iyun AO, Lennard MS, Tucker GT, Woods HF (1986) Metoprolol and debrisoquin metabolism in Nigerians: Lack of evidence for polymorphic oxidation. Clin Pharmacol Ther 40: 387-394
- Jackson PR, Tucker GT, Lennard MS, Woods HF (1986) Polymorphic drug oxidation: pharmacokinetic basis and comparison of experimental indices. Br J Pharmacol 22: 541–550
- Kitchen I, Tremblay J, André, Dring LG, Idle JR, Smith RL, Williams RT (1979) Interindividual and interspecies variation in the metabolism of the hallucinogen 4-methoxyamphetamine. Xenobiotica 7: 397-404
- Knodell RG, Raghvendra KD, Wilkinson GR, Guengerich FP (1988) Oxidative metabolism of hexobarbital in human liver: Relationship to polymorphic S-mephenytoin 4-hydroxylation.
   J Pharmacol Exp Ther 245: 845-849

- Kroemer HK, Mikus G, Kronbach T, Meyer UA, Eichelbaum M (1989) In vitro characterization of the human cytochrome P-450 involved in polymorphic oxidation of propafenone. Clin Pharmacol Ther 45: 28-33
- Kronbach T, Fisher V, Meyer UA (1988) Cyclosporine metabolism in human liver: Identification of a cytochrome P450III gene family as the major cyclosporine metabolizing enzyme explaining interactions of cyclosporine with other drugs. Clin Pharmacol Ther 43: 630-635
- Küpfer A, Preisig R (1984) Pharmacogenetics of mephenytoin: A new drug hydroxylation polymorphism in man. Eur J Clin Pharmacol 26: 753-759
- Küpfer A, Branch RA (1985) Stereoselective mephobarbital hydroxylation cosegregates with mephenytoin hydroxylation. Clin Pharmacol Ther 38: 414-418
- Leemann TP, Dayer P, Meyer UA (1986) Single-dose quinidine treatment inhibits metoprolol oxidation in extensive metabolizers. Eur J Clin Pharmacol 29: 739-741
- Lennard MS, Tucker GT, Silas JH, Freestone S, Ramsay LE, Woods HF (1983) Differential stereoselective metabolism of metoprolol in extensive and poor debrisoquin metabolizers. Clin Pharmacol Ther 34: 732-737
- Lennard MS, Jackson PR, Freestone S, Tucker GT, Ramsay LE, Woods HF (1984) The relationship between debrisoquine oxidation phenotype and the pharmacokinetics and pharmacodynamics of propranolol. Br J Clin Pharmacol 17: 679-685
- Lennard MS, McGourty JC, Silas JH (1988) Lack of relationship between debrisoquine oxidation phenotype and the pharmacokinetics and first dose effect of prazosin. Br J Clin Pharmacol 25: 276-278
- Lewis RV, Lennard MS, Jackson PR, Tucker GT, Ramsay LE, Woods HF (1985) Timolol and atenolol: relationships between oxidation phenotype, pharmacokinetics and pharmacodynamics. Br J Clin Pharmacol 19: 329-333
- Lou YC, Ying L, Bertilsson L, Sjöqvist F (1987) Low frequency of slow debrisoquine hydroxylation in a native Chinese population. Lancet 2: 852-853
- Mahgoub A, Idle JR, Dring LG, Lancaster R, Smith RL (1977) Polymorphic hydroxylation of debrisoquine in man. Lancet 2: 584–586
- Mellström B, Bertilsson L, Lou Y-C, Säwe J, Sjöqvist F (1983) Amitriptyline metabolism: Relationship to polymorphic debrisoquine hydroxylation. Clin Pharmacol Ther 34: 516-520
- Meyer UA, Gut J, Kronbach T, Skoda C, Meier UT, Catin T (1986) The molecular mechanisms of two common polymorphisms of drug oxidation evidence for functional changes in cytochrome P-450 isozymes catalysing bufuralol and mephenytoin oxidation. Xenobiotica 16: 449-464
- Mikus G, Ha HR, Vozeh S, Zekorn F, Follath F, Eichelbaum M (1986) Pharmacokinetics and metabolism of quinidine in extensive and poor metabolizers of sparteine. Eur J Clin Pharmacol 31: 69-72
- Miners JO, Wing LMH, Birkett DJ (1985) Normal metabolism of debrisoquine and theophylline in a slow tolbutamide metaboliser. Austr NZ J Med 15: 348-349
- Nakamura K, Goto F, Ray WA, McAllister CB, Jacqz E, Wilkinson GR, Branch RA (1985) Interethnic differences in genetic polymorphism of debrisoquin and mephenytoin hydroxylation between Japanese and Caucasian populations. Clin Pharmacol Ther 38: 402-408
- Nebert DW, Adesnik M, Coon MJ, Estabrook RW, Gonzalez FJ, Guengerich FP, Gonsalus IC, Johnson EF, Kemper B, Levin W, Phillips IR, Sato R, Waterman MR (1987) The P450 gene super family: Recommended nomenclature. DNA 6: 1-11
- Nordin C, Siwers B, Benitz J, Bertilsson L (1985) Plasma concentrations of nortriptyline and its 10-hydroxymetabolite in de-

pressed patients-relationship to the debrisoquine hydroxylation metabolic ratio. Br J Clin Pharmacol 19: 832-835

- Oates NS, Shah RR, Idle JR, Smith RL (1983) Influence of oxidation polymophism on phenformin kinetics and dynamics. Clin Pharmacol Ther 34: 827-834
- Oram M, Wilson K, Burnett D, Al-Dabbagh SG, Idle JR, Smith RL (1982) Metabolic oxidation of methaqualone in extensive and poor metabolizers of debrisoquine. Eur J Clin Pharmacol 23: 147–150
- Otton SV, Inaba T, Kalow W (1984) Competitive inhibition of sparteine oxidation in human liver by  $\beta$ -adrenoceptor antagonists and other cardiovascular drugs. Life Sci 34: 73–80
- Pierce DM, Smith SE, Franklin RA (1987) The pharmacokinetics of indoramin and 6-hydroxyindoramin in poor and extensive hydroxylators of debrisoquine. Eur J Clin Pharmacol 33: 59-65
- Preskorn S (1989) Therapeutic drug monitoring of tricyclic antidepressants: A means of avoiding toxicity. In: Dahl SG, Gram LF (eds) Clinical pharmacology in psychiatry. From molecular studies to clinical reality. Springer, Berlin Heidelberg New York Tokyo, in press
- Roy SD, Hawes EM, McKay G, Korchinski ED, Midha KK (1985) Metabolism of methoxyphenamine in extensive and poor metabolizers of debrisoquin. Clin Pharmacol Ther 38: 128-133
- Schmid B, Bircher J, Preisig R, Küpfer A (1985) Polymorphic dextromethorphan metabolism: Co-segretation of oxidative O-demethylation with debrisoquin hydroxylation. Clin Pharmacol Ther 38: 618-624
- Shaheen O, Patel J, Avant GR, Hamilton M, Wood AJJ (1986) Effect of cirrhosis and debrisoquin phenotype on the disposition and effects of pinacidil. Clin Pharmacol Ther 40: 650-655
- Siddoway LA, Thompson KA, McAllister CB, Wang T, Wilkinson GR, Roden DM, Woosley RL (1987) Polymorphism of propafenone metabolism and disposition in man: clinical and pharmacokinetic consequences. Circulation 75: 785-791
- Siris SG, Cooper TB, Rifbein AE, Brenner R, Liebermann JA (1982) Plasma imipramine concentration in patients receiving concomitant fluphenazine decanoate. Am J Psychiatry 139: 104-106
- Sjöqvist F, Bertilsson L (1986) Slow hydroxylation of tricylic antidepressants – Relationship to polymorphic drug oxidation. In: Kalow W, Goedde HW, Agarwal DP (eds) Ethnic differences in reactions to drugs and xenobiotics. Alan R. Liss, New York, pp 169-188
- Sloan TP, Mahgoub A, Lancaster R, Idle JR, Smith RL (1978) Polymorphism of carbon oxidation of drugs and clinical implications. Br Med J 2: 655-657
- Sloan TP, Lancaster R, Shah RR, Idle JR, Smith RL (1983) Genetically determined oxidation capacity and the disposition of debrisoquine. Br J Clin Pharmacol 15: 443–450
- Speirs CJ, Murray S, Boobis AR, Seddon CE, Davies DS (1986) Quinidine and the identification of drugs whose elimination is

impaired in subjects classified as poor metabolizers of debrisoquine. Br J Clin Pharmacol 22: 739-743

- Steiner E, Iselius L, Alván G, Lindsten J, Sjöqvist F (1985) A family study of genetic and environmental factors determining polymorphic hydroxylation of debrisoquin. Clin Pharmacol Ther 38: 394-401
- Steiner E, Alván G, Garle M, Maguire JH, Lind M, Nilson S-O, Tomson T, McClanahan JS, Sjöqvist F (1987) The debrisoquin hydroxylation phenotype does not predict the metabolism of phenytoin. Clin Pharmacol Ther 42: 326-333
- Steiner E, Dumont E, Spina E, Dahlqvist R (1988a) Inhibition of desipramine 2-hydroxylation by quinidine and quinine. Clin Pharmacol Ther 43: 577-581
- Steiner E, Bertilsson L, Säwe J, Bertling I, Sjöqvist F (1988b) Polymorphic debrisoquine hydroxylation in 757 Swedish subjects. Clin Pharmacol Ther 44: 431-435
- Syvählahti EKG, Lindberg R, Kallio J, de Vocht M (1986) Inhibitory effects of neuroleptics on debrisoquine oxidation in man. Br J Clin Pharmacol 22: 89-92
- Wagner F, Kalusche D, Trenk D, Jänchen E, Roskamm H (1987a) Drug interaction between propafenone and metoprolol. Br J Clin Pharmacol 24: 213–220
- Wagner F, Jänchen E, Trenk D (1987b) Severe complications of antianginal drug therapy in a patient identified as a poor metabolizer of metoprolol, propafenone, diltiazem and sparteine. Klin Wochenschr 65: 1164-1168
- Wang T, Roden DM, Wolfenden HT, Woosley RL, Wood AJJ, Wilkinson GR (1984) Influence of genetic polymorphism on the metabolism and disposition of encainide in man. J Pharmacol Exp Ther 228: 605-611
- Webb NR, Rose TM, Malik N, Marquardt H, Shoyab M, Todaro GJ, Lee DC (1987) Bovine and human cDNA sequences encoding a putative benzodiazepine receptor ligand. DNA 6: 71-79
- Zanger UM, Vilbois F, Hardwick J, Meyer UA (1988) Absence of hepatic cytochrome P450buf I causes genetically deficient debrisoquine oxidation in man. Biochemistry 27: 5447-5454
- Zekorn C, Achtert G, Hausleiter HJ, Moon CH, Eichelbaum M (1985) Pharmacokinetics of N-propylajmaline in relation to polymorphic sparteine oxidation. Klin Wochenschr 63: 1180-1186

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