

# Identity of inhibitory presynaptic 5-hydroxytryptamine (5-HT) autoreceptors in the rat brain cortex with 5-HT<sub>1B</sub> binding sites

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**Summary.** 1. In rat brain cortex slices preincubated with [<sup>3</sup>H]5-HT, the potencies of 17 5-HT receptor agonists to inhibit the electrically evoked <sup>3</sup>H overflow and the affinities of 13 antagonists (including several  $\beta$ -adrenoceptor blocking agents) to antagonize competitively the inhibitory effect of unlabelled 5-HT on evoked <sup>3</sup>H overflow were determined.

2. The affinities of the compounds for 5-HT<sub>1B</sub> and 5-HT<sub>2</sub> binding sites in rat brain cortex membranes (labelled by [<sup>125</sup>I]cyanopindolol = [<sup>125</sup>I]-CYP in the presence of 30  $\mu$ mol/l isoprenaline and [<sup>3</sup>H]ketanserin, respectively), for 5-HT<sub>1A</sub> binding sites in pig and rat brain cortex membranes (labelled by [<sup>3</sup>H]8-hydroxy-2-(di-n-propylamino)tetralin = [<sup>3</sup>H]8-OH-DPAT) and for 5-HT<sub>1C</sub> binding sites in pig choroid plexus membranes (labelled by [<sup>3</sup>H]mesulergine) were also determined. The affinities of the drugs for the various 5-HT recognition sites ranged over 4–5 log units (the functional experiments revealed the same range of differences between the drugs).

3. There were no significant correlations between the affinities of the drugs at 5-HT<sub>1C</sub> and 5-HT<sub>2</sub> binding sites and their potencies or affinities, determined for the 5-HT autoreceptors. In contrast, significant correlations were found between the potencies or affinities of the drugs for the autoreceptors and their affinities at 5-HT<sub>1A</sub> or 5-HT<sub>1B</sub> binding sites; the best correlations were obtained with the 5-HT<sub>1B</sub> binding site.

4. Some of the drugs investigated were not included in the correlation since their agonistic or antagonistic effects on the autoreceptors were weak and pEC<sub>30</sub> or apparent pA<sub>2</sub> values could not be determined (< 5.5). Among these drugs, 8-OH-DPAT, TVX Q 7821 (2-(4-(4-(2-pyrimidin-yl)-1-piperazinyl)-butyl)-1,2-benzisothiazol-3(2H)one-1,1-dioxide) and spiperone showed a very low affinity for 5-HT<sub>1B</sub> binding sites (pK<sub>D</sub> < 5.3), but a high affinity for 5-HT<sub>1A</sub> binding sites (pK<sub>D</sub> > 7.2).

5. In conclusion, the evidence indicates that the presynaptic 5-HT autoreceptor belongs to the 5-HT<sub>1B</sub> receptor subtype.

**Key words:** Presynaptic 5-HT autoreceptors — 5-HT release — Rat and pig brain cortex — 5-HT binding sites — 5-HT receptor subtypes

## Introduction

The first classification of 5-HT receptors proposed by Gaddum and Picarelli (1957) divided these receptors into two classes: the D receptor was located on the smooth muscle cells of the guinea pig ileum and the M receptor on the nerves of the myenteric plexus. Using radioligand binding studies, Peroutka and Snyder (1979) described 5-HT<sub>1</sub> and 5-HT<sub>2</sub> binding sites in rat brain cortex. The 5-HT<sub>2</sub> sites were identified as receptors and many functional correlates have since been established for these sites (Peroutka et al. 1981; Leysen et al. 1982). It is now known that the D receptor of the guinea pig ileum is identical with the 5-HT<sub>2</sub> receptor (Engel et al. 1985). The question of the functional correlates of the 5-HT<sub>1</sub> recognition site has remained a matter of debate (Peroutka et al. 1981; Middlemiss 1982; Fozard 1983), in part because of the existence of subtypes of the 5-HT<sub>1</sub> binding site.

5-HT<sub>1</sub> recognition sites were subdivided into 5-HT<sub>1A</sub> and 5-HT<sub>1B</sub> subtypes by means of radioligand binding studies performed with [<sup>3</sup>H]5-HT (Pedigo et al. 1981; Deshmukh et al. 1982); the 5-HT<sub>1A</sub> sites had high affinity for spiperone whilst the 5-HT<sub>1B</sub> sites showed low affinity for this compound. Further analysis of the complex competition curves obtained for [<sup>3</sup>H]5-HT binding suggested the existence of a third 5-HT<sub>1</sub> recognition site (Nelson et al. 1983; Engel et al. 1983). Subsequently, Palacios and collaborators by using [<sup>3</sup>H]mesulergine and [<sup>3</sup>H]5-HT described a further subtype, designated 5-HT<sub>1C</sub> concentrated in the choroid plexus of pig and rat (Pazos et al. 1984a, b; Pazos and Palacios 1985). It is now possible to identify three distinct 5-HT<sub>1</sub> binding sites with specific radioligands. 5-HT<sub>1A</sub> sites are labelled by [<sup>3</sup>H]8-OH-DPAT in rat and pig brain membranes (Gozlan et al. 1983; Hoyer et al. 1985b). 5-HT<sub>1B</sub> sites are labelled by [<sup>125</sup>I]CYP (in the presence of 30  $\mu$ M isoprenaline in order to exclude binding to  $\beta$ -adrenoceptors) (Hoyer et al. 1985a, b; Pazos et al. 1985) and 5-HT<sub>1C</sub> receptors are labelled by [<sup>3</sup>]mesulergine in pig choroid plexus (Pazos et al. 1984b; Hoyer et al. 1985b). Furthermore, it is established that [<sup>3</sup>H]5-HT labels all three 5-HT<sub>1</sub> recognition sites in rat brain cortex in equal proportions (Pazos and Palacios 1985; Hoyer et al. 1985b).

It has been proposed by various authors (Göthert 1982; Martin and Sanders Bush 1982a; Engel et al. 1983) that 5-HT autoreceptors are of the 5-HT<sub>1</sub> class, however the subtype has not been clearly established. Middlemiss (1984a) has shown that the highly selective 5-HT<sub>1A</sub> agonist, 8-OH-DPAT (Middlemiss and Fozard 1983), does not activate these autoreceptors. It was further shown that pro-

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pranolol and pindolol act as stereoselective antagonists at 5-HT autoreceptors (Middlemiss 1984b, 1985) and it was proposed that these autoreceptors belong to the 5-HT<sub>1B</sub> subtype (Middlemiss 1984b). However, 8-OH-DPAT also shows low affinity for the 5-HT<sub>1C</sub> sites, and neither propranolol nor pindolol distinguishes between 5-HT<sub>1A</sub> and 5-HT<sub>1B</sub> binding (Hoyer et al. 1985a, b). Furthermore function at 5-HT autoreceptors has not yet been correlated with binding data for the various 5-HT<sub>1</sub> receptor subtypes.

In this study we compare the pharmacology of a large series of 5-HT agonists and antagonists from different structural classes at 5-HT autoreceptors with their affinity for 5-HT<sub>1</sub> and 5-HT<sub>2</sub> receptor subtypes in radioligand binding studies. We demonstrate that 5-HT autoreceptors do not belong to the 5-HT<sub>2</sub>, 5-HT<sub>1C</sub> or 5-HT<sub>1A</sub> subtypes and are best classified as 5-HT<sub>1B</sub> receptors.

## Materials and methods

*Preparation of particulate fractions for binding studies.* Male rats (Sandoz own breed, weight about 250 g) were decapitated and the brains rapidly removed and placed on ice. Pig brains were obtained from a local slaughterhouse and placed on ice immediately after death. Frontal cortices of both species and pig choroid plexi were removed and membrane homogenates were prepared as described (Hoyer et al. 1985b; Pazos et al. 1984a, b). The membranes were stored in liquid nitrogen or kept deep frozen at  $-70^{\circ}\text{C}$  until used.

### Radioligand binding experiments

Binding of the tritiated radioligands [<sup>3</sup>H]8-OH-DPAT (5-HT<sub>1A</sub>), [<sup>3</sup>H]ketanserin (5-HT<sub>2</sub>) and [<sup>3</sup>H]mesulergine (5-HT<sub>1C</sub>) was performed as previously described (Hoyer et al. 1985b, Pazos et al. 1984a, b).

[<sup>125</sup>I]CYP binding (5-HT<sub>1B</sub>) was performed in the presence of 30 μmol/l isoprenaline, in order to exclude any interference with β-adrenoceptors (Hoyer et al. 1985a, b). Briefly, 150 μl of membrane suspension (20–30 μg of protein) were added to 50 μl of [<sup>125</sup>I]CYP (100–150 pmol/l) and 50 μl of drug or buffer (Tris HCl 10 mmol/l, NaCl 154 mmol/l, pargyline 10 μmol/l, pH 7.7). Membranes, drugs and the radioligand were dissolved in this buffer. The incubation was carried out in polystyrene microtiter plates (Sterilin 96 holes) in an incubator at 37°C for 90 min. The reaction was stopped by the addition of ice cold Tris HCl 50 mmol/l, pH 7.4 and bound and free ligand were separated by washing and filtering through Gelman A/E glass fiber filters on a cell harvester (Flow Laboratories). After drying under reduced suction, the filters were counted in a WW 1032 gamma counter (Kontron) at 80% counting efficiency.

Competition curves were constructed with 8 to 20 concentrations of competing drugs. As a rule all incubations were carried out in triplicate. Independent experiments were performed at least twice. Protein concentrations were determined according to the method of Bradford (1976).

### Superfusion experiments on cortex slices

Occipitoparietal cortex slices (0.3 mm thick, diameter 3 mm) of male Wistar rats weighing 200–300 g were incubated (60 min) in physiological salt solution (for composition see Engel et al. 1983) containing 0.1 μmol/l [<sup>3</sup>H]5-hydroxy-

tryptamine creatinine sulphate. Subsequently they were superfused with physiological salt solution containing paroxetine or DU 24565 (selective inhibitors of neuronal 5-HT uptake; Petersen et al. 1977; Vaastra et al. 1981; Clasen et al. 1984). Two 2-min periods of electrical field stimulation (20 mA, 2 ms, 3 Hz) were applied to each slice 40 (S<sub>1</sub>) and 90 min (S<sub>2</sub>) after the onset of superfusion. At the end of superfusion, the slices were solubilized, and the radioactivity in the slices and in the superfusate samples was determined by liquid scintillation counting. All further details concerning superfusion, stimulation and experimental protocol were identical to those described by Engel et al. (1983).

### Calculations and statistics

Untransformed data of the saturation and competition *binding* experiments with the various radioligands were analyzed by non-linear regression analysis, based on Feldman's equation (1972), using the computer modeling program SCTFIT as described in detail by DeLean et al. (1980). By *F*-Test analysis the goodness of fit between the models of one or two classes of binding sites was checked and the most appropriate model was determined (Snedecor and Cochran 1973).

Basal <sup>3</sup>H efflux and stimulation-evoked <sup>3</sup>H overflow in *superfusion* experiments were calculated according to Engel et al. (1983). Apparent pA<sub>2</sub> values of 5-HT receptor antagonists were determined as described by Furchgott (1972) for competitive reversible antagonism (see also Göthert et al. 1981).

Results are expressed as arithmetic means (functional studies) or geometric means (radioligand binding studies) ± SEM of *n* independent experiments. Linear regression lines and correlation coefficients were calculated in order to detect and quantify correlations between various drug effects.

*Drugs used.* [<sup>3</sup>H]5-hydroxytryptamine creatinine sulphate (24.1–26.3 Ci/mmol) and [<sup>3</sup>H]-ketanserin hydrochloride (64 Ci/mmol) NEN, Dreieich, FRG); [<sup>3</sup>H]8-hydroxy-2-(di-n-propylamino)tetralin hydrobromide (105–125 Ci/mmol) ([<sup>3</sup>H]8-OH-DPAT; CEA, Saclay, France); 5-hydroxytryptamine creatinine sulphate (5-HT) and tryptamine hydrochloride (T; Merck, Darmstadt, FRG); 5-methoxytryptamine hydrochloride (5-OCH<sub>3</sub>-T; Fluka, Buchs, Switzerland); 6-hydroxytryptamine (6-HT, Sigma, Munich, FRG); α-methyl-5-hydroxytryptamine creatinine sulphate (α-CH<sub>3</sub>-5-HT; Upjohn, Kalamazoo, MI, USA); 4-hydroxytryptamine creatinine sulphate (4-HT), 5,6-dihydroxytryptamine creatinine sulphate (5,6-DHT) and 5,7-dihydroxytryptamine creatinine sulphate (5,7-DHT) (Regis, Morton Grove, IL, USA); 2-methyl-5-hydroxytryptamine hydrogen maleinate (2-CH<sub>3</sub>-5-HT), β-methyl-5-hydroxytryptamine hydrogen oxalate (β-CH<sub>3</sub>-5-HT), ω-N-methyl-5-hydroxytryptamine oxalate (ω-N-CH<sub>3</sub>-5-HT), N,N-dimethyl-5-hydroxytryptamine binoxalate (NN-(CH<sub>3</sub>)<sub>2</sub>-5-HT), 5-aminotryptamine hydrogenoxalate (5-NH<sub>2</sub>-T), 5-carboxamido-tryptamine hydrogen maleinate (5-CONH<sub>2</sub>-T), unlabelled 8-OH-DPAT, (±)-cyanopindolol (base), (+)-cyanopindolol (base), (–)-cyanopindolol (base), (+)-pindolol (base), (–)-pindolol (base), (±)-4[3-ter-butyl-amino-2-hydroxypropoxy]indol-2-carbonic-acid-isopropylester [(±)-21-009], (–)-[<sup>125</sup>I]-iodo-cyanopindolol (2,175 Ci/mmol) ([<sup>125</sup>I]CYP), [<sup>3</sup>H]mesulergine (85 Ci/mmol) and d-LSD tartrate were synthesized by Sandoz, Basel, Switzerland; metitepin (Hoffmann-La

**Table 1.** Methodological data for in vitro binding assay on various 5-HT-receptors

Receptor Subtype	Ligand	Membranes	$B_{\max}$ fmol/mg protein	$K_D$ nmol/l	Determination of non-specific binding	Reference
5-HT <sub>1A</sub>	[ <sup>3</sup> H]8-OH-DPAT	rat brain cortex <sup>a</sup> pig brain cortex	127 ± 6 73 ± 5	1.9 ± 0.3 2.4 ± 0.4	10 μmol/l 5-HT	Hoyer et al. 1985b
5-HT <sub>1B</sub>	[ <sup>125</sup> I]CYP + 30 μmol/l isoprenaline	rat brain cortex	140 ± 3	0.21 ± 0.01	10 μmol/l 5-HT	Hoyer et al. 1985a
5-HT <sub>1C</sub>	[ <sup>3</sup> H]mesulergine	pig choroid plexus	300 ± 64	1.1 ± 0.1	10 μmol/l 5-HT	Pazos et al. 1984b
5-HT <sub>2</sub>	[ <sup>3</sup> H]ketanserin	rat brain cortex	175 ± 25	0.8 ± 0.2	1 μmol/l mianserin	Pazos et al. 1984a

<sup>a</sup> 5-HT<sub>1A</sub> binding was determined using pig brain cortex membranes. 5-HT<sub>1A</sub> receptors from rat and pig brain cortex are identical (Hoyer et al. 1985b)

Roche, Basel Switzerland); quipazine (Miles, Elkhart, IN, USA); metergoline (Farmitalia, Milan, Italy); DU 24565 (6-nitro,2-(1-piperazinyl)quinoline; Philips-Duphar, Weesp, The Netherlands); (±)-propranolol hydrochloride (ICI, Plankstadt, FRG); 6-chloro-2-(1-piperazinyl)pyrazine hydrochloride (MK 212; Merck, Sharpe and Dohme, Munich, FRG or synthesized by Sandoz, Basel, Switzerland); unlabelled ketanserin and spiroperidol (spiperone) (Janssen, Beerse, Belgium); TVX Q 7821 (2-(4-(4-(2-pyrimidinyl)-1-piperazinyl)butyl)-1,2-benzisothiazol-3(2H)one-1,1-dioxide hydrochloride) (Troponwerke, Köln, FRG); cyproheptadine hydrochloride (Merck, Sharpe and Dohme, Munich, FRG); mianserin hydrochloride (Organon, Oss, Netherlands); isoprenaline hydrochloride (Sigma, Taufkirchen, FRG), paroxetine hydrochloride (Ferrosan, Copenhagen, Denmark).

## Results

### Binding experiments

Table 1 summarizes the values of maximal capacities ( $B_{\max}$ ), the apparent dissociation constants ( $K_D$ ) and the conditions by which non-specific binding in the different assay systems was determined. The competition curves with [<sup>3</sup>H]8-OH-DPAT as well as those with [<sup>125</sup>I]CYP in the presence of isoprenaline were monophasic for 5-HT receptors agonists and antagonists. The same held true for [<sup>3</sup>H]mesulergine binding in pig choroid plexus. The 5-HT<sub>2</sub> binding sites were labelled by [<sup>3</sup>H]ketanserin in rat brain cortex. Some 5-HT receptor agonists, especially 5-HT itself, showed flat or biphasic competition curves with [<sup>3</sup>H]ketanserin. Since the reason for the biphasic shape of the competition curves obtained with 5-HT receptor agonists is unknown only the  $pK_D$  of the main component was taken into account and indicated in Table 2.

### Functional experiments

Unlabelled 5-HT inhibited the electrically evoked <sup>3</sup>H overflow in a concentration-dependent manner, a 60% decrease representing the maximum effect. Qualitatively similar results were obtained with the other indolethylamines investigated (Göthert and Schlicker 1983; Engel et al. 1983) and with LSD; the negative logarithms of the concentrations

producing the half maximum effect ( $pEC_{30}$  values) are listed in Table 2 (in the concentration range investigated the compounds mentioned so far did not affect the basal <sup>3</sup>H-efflux). The evoked <sup>3</sup>H overflow was not affected by TVX Q 7821 (up to 3.2 μmol/l), but was inhibited by unlabelled 8-OH-DPAT 3.2 and 10 μmol/l (by 18 and 38%, respectively). Since, from 1 μmol/l onward, 8-OH-DPAT and TVX Q 7821 enhanced basal <sup>3</sup>H efflux, the effects of both drugs on the evoked <sup>3</sup>H overflow in this concentration range must be regarded with caution.

Metitepin, metergoline and quipazine caused a parallel shift to the right of the concentration-response curve of unlabelled 5-HT for its inhibitory effect on evoked <sup>3</sup>H overflow (Engel et al., 1983). The same held true for a series of  $\beta$ -adrenoceptor antagonists including (±)-cyanopindolol and its enantiomers (Schlicker et al., 1985), (±)-propranolol, (±)-21009 and (-)-pindolol as well as for the piperazine derivative MK 212 (data not shown).

From the shifts at the level of the  $EC_{30}$  value of 5-HT, apparent  $pA_2$  values of the antagonists were calculated (Table 2). (+)-Pindolol, spiroperidol and cyproheptadine, each up to 3.2 μmol/l, did not affect the concentration-response curve of 5-HT. The inhibitory effects of other indolethylamines and of LSD were also antagonized by metitepin (Langer and Moret 1982; Göthert and Schlicker 1983). By contrast, metitepin or (±)-cyanopindolol, each 1 μmol/l, did not affect the inhibitory effect of 8-OH-DPAT 10 μmol/l (data not shown).

### Correlations between binding affinities and functional effects

The  $pEC_{30}$ , apparent  $pA_2$  and  $pK_D$ -values of the 5-HT receptor agonists or antagonists for 5-HT autoreceptor-mediated effects and for the affinities at the various 5-HT binding sites ranged over 4–5 log units (Fig. 1). No significant correlation was found between the affinities of the drugs (expressed as  $pK_D$  values) at 5-HT<sub>1C</sub> or 5-HT<sub>2</sub> binding sites and the  $pEC_{30}$  values of the agonists for the inhibition of evoked <sup>3</sup>H-overflow (Figs. 1C and D) or the apparent  $pA_2$  values of antagonists for the antagonism of the inhibition produced by unlabelled 5-HT (Figs. 1G and H; see Table 3 for statistical parameters). However, there were significant correlations between  $pEC_{30}$  or apparent  $pA_2$ -values and the affinities of the drugs at 5-HT<sub>1B</sub> or 5-HT<sub>1A</sub> binding sites (Figs. 1A, B, E, F). The correlation coefficients ( $r$ ) for the

**Table 2.** pEC<sub>30</sub> and apparent pA<sub>2</sub> values of 5-HT receptor agonists and antagonists at 5-HT-autoreceptors and pK<sub>D</sub> values of these drugs at 5-HT<sub>1A</sub>, 5-HT<sub>1B</sub>, 5-HT<sub>1C</sub> and 5-HT<sub>2</sub>-binding sites

Compounds <sup>a</sup>	pEC <sub>30</sub> (agonists) <sup>b</sup> or apparent pA <sub>2</sub> (antag.) <sup>c</sup> : 5-HT autoreceptors	pK <sub>D</sub> <sup>d</sup> : 5-HT binding sites			
		5-HT <sub>1B</sub>	5-HT <sub>1A</sub>	5-HT <sub>1C</sub>	5-HT <sub>2</sub>
<b>Agonists</b>					
1 5-HT	6.76	7.63 ± 0.12 (10)	8.51 ± 0.08 (6)	7.48 ± 0.07 (3)	5.53 ± 0.37 (5)
2 5-OCH <sub>3</sub> -T	6.45	6.40 ± 0.23 (3)	8.04 ± 0.15 (4)	7.36 ± 0.31 (4)	5.59 ± 0.32 (4)
3 2-CH <sub>3</sub> -5-HT	4.21	4.44 ± 0.16 (3)	5.60 ± 0.04 (3)	5.84 ± 0.19 (2)	4.99 ± 0.37 (3)
4 (±)α-CH <sub>3</sub> -5-HT	4.62	6.00 ± 0.25 (4)	7.07 ± 0.17 (5)	7.23 ± 0.21 (4)	6.90 ± 0.18 (4)
5 (±)β-CH <sub>3</sub> -5-HT	6.46	5.51 ± 0.20 (3)	7.05 ± 0.08 (3)	6.31 ± 0.16 (2)	5.60 ± 0.32 (2)
6 N,N(CH <sub>3</sub> ) <sub>2</sub> -5-HT	5.38	6.04 ± 0.30 (4)	7.60 ± 0.28 (4)	7.15 ± 0.09 (4)	6.42 ± 0.12 (2)
7 5-CONH <sub>2</sub> -T	7.65	8.29 ± 0.12 (3)	9.67 ± 0.08 (3)	6.23 ± 0.15 (3)	4.83 ± 0.03 (2)
8 4-HT	6.21	5.98 ± 0.22 (4)	7.02 ± 0.04 (2)	7.39 ± 0.30 (3)	6.14 ± 0.15 (2)
9 5,6-DHT	5.47	5.21 ± 0.13 (3)	6.02 ± 0.08 (4)	6.15 ± 0.05 (2)	5.01 ± 0.04 (2)
10 5,7-DHT	4.27	3.66 ± 0.34 (3)	4.90 ± 0.04 (3)	4.29 ± 0.16 (3)	< 4 (3)
11 5-NH <sub>2</sub> -T	5.96	6.17 ± 0.13 (4)	6.15 ± 0.17 (4)	7.15 ± 0.17 (2)	5.49 ± 0.24 (2)
12 6-HT	5.91	5.23 ± 0.31 (4)	5.80 ± 0.04 (2)	5.26 ± 0.13 (3)	4.94 ± 0.29 (2)
13 ω-N-CH <sub>3</sub> -5-HT	7.16	7.35 ± 0.27 (2)	8.31 ± 0.14 (3)	6.55 ± 0.21 (2)	6.74 ± 0.31 (3)
14 T	5.31	4.99 ± 0.34 (3)	6.77 ± 0.05 (2)	7.33 ± 0.02 (2)	—
15 8-OH-DPAT	< 5.5	4.22 ± 0.10 (4)	8.73 ± 0.11 (7)	5.24 ± 0.19 (4)	5.04 ± 0.09 (4)
16 LSD	5.50	6.82 ± 0.15 (3)	8.59 ± 0.06 (6)	7.93 ± 0.06 (5)	8.62 ± 0.06 (6)
17 TVXQ 7821	< 5.5	3.87 ± 0.27 (3)	7.73 ± 0.10 (3)	4.53 ± 0.27 (3)	5.07 ± 0.11 (3)
<b>Antagonists (in brackets: autoreceptor studies, concentrations investigated, μmol/l)</b>					
18 metitepin (1.0)	6.99	7.28 ± 0.03 (5)	7.10 ± 0.13 (5)	7.56 ± 0.07 (4)	8.76 ± 0.09 (7)
19 quipazine (10.0)	5.29	6.51 ± 0.14 (6)	5.49 ± 0.05 (7)	6.73 ± 0.09 (3)	6.20 ± 0.03 (5)
20 metergoline (1.0)	6.29	7.39 ± 0.09 (8)	8.10 ± 0.10 (5)	9.19 ± 0.10 (3)	9.03 ± 0.14 (6)
21 MK 212 (320)	4.04	5.03 ± 0.13 (4)	5.32 ± 0.03 (5)	6.16 ± 0.15 (2)	4.76 ± 0.14 (5)
22 (±)cyanopindolol (0.032)	8.29	8.28 ± 0.11 (6)	8.27 ± 0.12 (4)	4.44 ± 0.5 (3)	4.53 ± 0.13 (4)
23 (+)cyanopindolol (1.0)	6.83	7.25 ± 0.13 (4)	7.27 ± 0.16 (3)	—	4.60 ± 0.13 (3)
24 (-)cyanopindolol (0.032)	8.30	8.74 ± 0.05 (3)	8.64 ± 0.11 (3)	—	4.58 ± 0.04 (3)
25 (+)pindolol (3.2)	< 5.5	5.29 ± 0.08 (4)	5.88 ± 0.05 (5)	4.18 ± 0.3 (3)	4.42 ± 0.08 (7)
26 (-)pindolol (1.0)	6.57	7.19 ± 0.03 (10)	7.66 ± 0.04 (6)	4.31 ± 0.07 (5)	4.53 ± 0.20 (3)
27 (±)propranolol (0.32)	6.76	7.07 ± 0.10 (4)	6.48 ± 0.06 (3)	—	6.12 ± 0.22 (4)
28 (±)21-009 (0.075)	7.49	8.53 ± 0.04 (7)	7.76 ± 0.14 (4)	5.05 ± 0.05 (3)	4.58 ± 0.21 (4)
29 spiperone (3.2)	< 5.5	5.27 ± 0.06 (3)	7.18 ± 0.26 (4)	5.94 ± 0.02 (3)	8.76 ± 0.03 (3)
30 cyproheptadine (3.2)	< 5.5	5.32 ± 0.22 (3)	6.45 ± 0.07 (3)	7.86 ± 0.32 (4)	8.46 ± 0.14 (8)

<sup>a</sup> For explanation of abbreviations see Methods (Drugs used)

<sup>b</sup> Derived from concentration-response curves for the inhibitory effects of 5-HT receptor agonists on the electrically evoked <sup>3</sup>H overflow from rat brain cortex slices preincubated with [<sup>3</sup>H]5-HT. Neuronal 5-HT uptake was blocked throughout superfusion by paroxetine 3.2 μmol/l (indolethylamines, LSD) or DU 24565 1 μmol/l (8-OH-DPAT, TVX Q 7821). The pEC<sub>30</sub> values of indolethylamines were taken from Göthert and Schlicker (1983) and Engel et al. (1983)

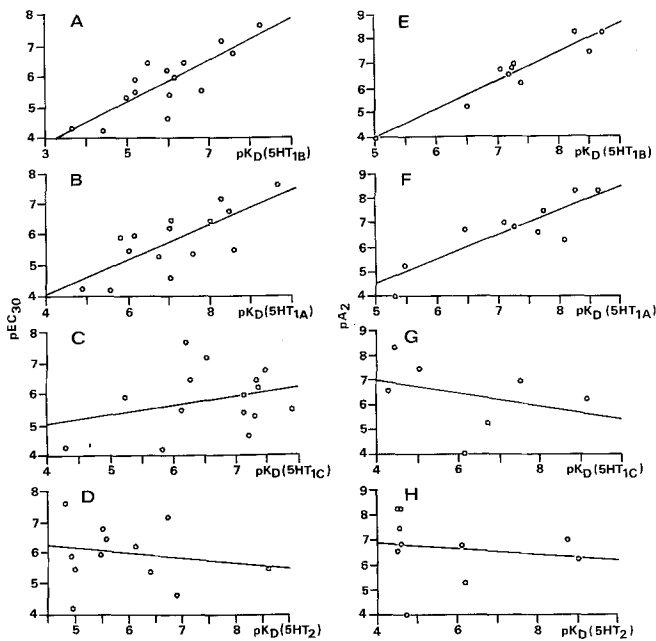
<sup>c</sup> Apparent pA<sub>2</sub> values of the compounds for their antagonism against unlabelled 5-HT were determined as described in the text. Neuronal 5-HT uptake was blocked throughout superfusion by DU 24565 1 μmol/l. The apparent pA<sub>2</sub> values of metitepin, quipazine, metergoline and cyproheptadine were taken from Engel et al. (1983). Absolute values of <sup>3</sup>H efflux in the control experiments (5-HT and 5-HT receptor antagonists absent): basal efflux determined immediately before S<sub>2</sub>, 0.15 ± 0.03 nCi/5 min (corresponding to a fractional rate of <sup>3</sup>H efflux of 0.0052 ± 0.0002/min; related to tissue tritium); <sup>3</sup>H overflow evoked by S<sub>2</sub>, 0.17 ± 0.04 nCi (corresponding to 2.80 ± 0.30% of tissue tritium)

<sup>d</sup> Geometric means ± SEM of (*n*) independent experiments. For *n* = 2 the maximal deviations from the mean value are indicated

correlations with 5-HT<sub>1B</sub> binding sites (Figs. 1A and E, Table 3) were higher than those for the correlations with 5-HT<sub>1A</sub> binding sites (Figs. 1B and F, Table 3). In the binding studies, the best discrimination between 5-HT<sub>1A</sub> and 5-HT<sub>1B</sub> binding sites could be obtained with unlabelled 8-OH-DPAT and TVX Q 7821, which selectively displayed a high affinity at the 5-HT<sub>1A</sub> binding sites (pK<sub>D</sub> 5-HT<sub>1A</sub> = 8.73 and 7.73 respectively; Table 2). Our findings that the electrically evoked <sup>3</sup>H overflow was only marginally inhibited by 8-OH-DPAT (3.2 μmol/l) and not affected at all by TVX Q 7821 were not compatible with similarities between the sites mediating the functional effects and the

5-HT<sub>1A</sub> binding sites, but rather with similarities between the functional sites and the 5-HT<sub>1B</sub> binding sites (pK<sub>D</sub> of 4.22 and 3.87, respectively).

This assumption is further supported by two 5-HT receptor antagonists, spiperone and cyproheptadine, which discriminate also quite clearly between the 5-HT<sub>1B</sub> and the 5-HT<sub>1A</sub> site. Both spiperone and cyproheptadine were inactive at the autoreceptor (up to a concentration of 3.2 μmol/l) and had low affinities for 5-HT<sub>1B</sub> sites (pK<sub>D</sub> values: 5.27 and 5.32, respectively), whereas their affinities for 5-HT<sub>1A</sub> sites were higher by about 1–2 orders of magnitude (7.18 and 6.45, respectively).



**Fig. 1A–H.** Correlation of  $pEC_{30}$  or apparent  $pA_2$  values from superfusion experiments with  $pK_D$  values from binding studies. The affinity values and the potencies for the calculation of the linear regression are taken from Table 2. The correlation lines were calculated with  $pEC_{30}$  values for 5-HT receptor agonists (*left side*: A, B, C, D) and with apparent  $pA_2$  values for 5-HT receptor antagonists (*right side*: E, F, G, H) and with  $pK_D$  values derived from specific binding experiments. The correlation coefficients and the statistical significance are given in Table 3

**Table 3.** Parameter estimation ( $y = mx + b$ ) from linear regression analysis in Fig. 1

Parameter	5-HT <sub>1B</sub>	5-HT <sub>1A</sub>	5-HT <sub>1C</sub>	5-HT <sub>2</sub>
5-HT receptor agonists ( $pEC_{30}$ versus $pK_D$ )				
<i>m</i>	0.67	0.56	0.28	-0.16
<i>b</i>	1.81	1.82	3.93	6.92
<i>r</i>	0.82	0.74	0.28	0.18
<i>p</i>	0.001	0.0018	0.32	0.57
5-HT receptor antagonists (apparent $pA_2$ versus $pK_D$ )				
<i>m</i>	1.15	0.98	-0.26	-0.11
<i>b</i>	-1.73	-0.41	8.03	7.33
<i>r</i>	0.95	0.86	0.33	0.15
<i>p</i>	0.001	0.0013	0.48	0.67

*m* = slope; *b* = intercept on the *y*-axis; *r* = correlation coefficient; *p* = significance level

## Discussion

The aim of this study was to characterize the pharmacology of 5-HT autoreceptors of the rat brain cortex and to determine to which 5-HT receptor subtype autoreceptors belong. This was done by combining functional and radioligand binding studies.

In rat brain cortex membranes, [<sup>125</sup>I]CYP (in the presence of 30 μmol/l isoprenaline) (Hoyer et al. 1985a, b; Pazos et al. 1985) and [<sup>3</sup>H]ketanserin (Leysen et al. 1982; Pazos et al. 1984a, b) label 5-HT<sub>1B</sub> and 5-HT<sub>2</sub> binding sites, respectively. In pig cortex and choroid plexus membranes,

[<sup>3</sup>H]-8-OH-DPAT (Hoyer et al. 1985b) and [<sup>3</sup>H]mesulergine (Pazos et al. 1984b) are suitable to identify 5-HT<sub>1A</sub> and 5-HT<sub>1C</sub> binding sites respectively. Furthermore, it has been shown that all three subtypes of 5-HT<sub>1</sub> binding sites occur in the rat brain cortex (Hoyer et al. 1985b).

Under the conditions of the present investigation, the electrically evoked <sup>3</sup>H overflow from the rat brain cortex slices reflects quasi-physiological serotonin release (Göthert and Weinheimer 1979; Classen et al. 1984), the latter is under the modulatory control of inhibitory presynaptic autoreceptors (for reviews, see Héry and Ternaux 1981; Starke 1981; Göthert 1982, 1985).

Tryptamine and its 5-hydroxy, 5-methyl, 5-methoxy, 5-amino, and 5-carboxamido derivatives as well as LSD, act as agonists at the presynaptic 5-HT autoreceptors (Göthert 1982, Göthert and Schlicker 1983, Engel et al. 1983). 8-OH-DPAT 1 μmol/l did not affect the evoked serotonin release in the present study an observation which confirms that of Middlemiss (1984a). Hamon et al. (1984), however, found a decrease in evoked 5-HT release by 34% (basal tritium efflux not affected), which could be antagonized by metergoline. In our study, 8-OH-DPAT inhibited the evoked serotonin release only at concentrations exceeding 1 μmol/l, at which basal <sup>3</sup>H efflux was markedly enhanced. The inhibitory effect of 8-OH-DPAT 10 μmol/l was not attenuated by metitepin or (±)-cyanopindolol, excluding that 5-HT autoreceptors are involved. TVXQ 7821, a putative anxiolytic, which displays affinity for 5-HT<sub>1A</sub> binding sites in the nanomolar range (Dompert et al. 1985), did not evoke <sup>3</sup>H overflow at 3.2 μmol/l.

The 5-HT receptor antagonists, metitepin, quipazine and metergoline have been shown to produce parallel shifts to the right of the concentration-response curve of unlabelled 5-HT for its inhibitory effect on 5-HT release from the rat brain cortex and hypothalamus (Göthert 1980, Schlicker and Göthert 1981, Martin and Sanders-Bush 1982b, Middlemiss 1984a). Data suggesting the possibility of antagonism by MK 212 at the 5-HT autoreceptor was first described by Baumann and Waldmeier (1981), and the parallel shift by MK 212 of the concentration-response curve of 5-HT, found in the present study, supports this suggestion.

Propranolol and pindolol were characterized as stereoselective antagonists at the 5-HT autoreceptors of the rat brain cortex by Middlemiss (1984b, 1985) and Richards (1985); in addition, the β-adrenoceptor antagonists (±)-cyanopindolol and its enantiomers (present study, Schlicker et al. 1985) and (±)-21009 (present study) block the autoreceptors.

There were no significant correlations between the affinities of the drugs at 5-HT<sub>1C</sub> and 5-HT<sub>2</sub> binding sites and their potencies or affinities determined for the 5-HT autoreceptor-mediated effect, thus excluding the possibility that these binding sites and receptors have properties in common. In contrast, significant correlations were found between the potencies or affinities of the drugs determined for the 5-HT autoreceptors and their affinities at 5-HT<sub>1A</sub> and 5-HT<sub>1B</sub> binding sites. However, this is not surprising since the pharmacological profile of 5-HT<sub>1A</sub> and 5-HT<sub>1B</sub> recognition sites for tryptamine derivatives is rather similar. Nevertheless the correlations were best with 5-HT<sub>1B</sub> sites, both when agonists and when antagonists were investigated. An even more clear cut distinction between both recognition sites becomes evident when selective agonists which are not tryptamine

derivatives, (like 8-OH-DPAT and TVX Q 7821) and 5-HT antagonists (like spiroperidol and cyproheptadine) are taken into account. These drugs had no measurable affinity to the autoreceptors and very low affinity to the 5-HT<sub>1B</sub> sites, but displayed moderate to high affinity to the 5-HT<sub>1A</sub> sites with pK<sub>D</sub> values greater than 6.4.

In conclusion the results of the present investigation indicate that the presynaptic 5-HT autoreceptors of the rat brain cortex belong to the 5-HT<sub>1B</sub> receptor subtype.

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## References

- Baumann PA, Waldmeier PC (1981) Further evidence for negative feedback control of serotonin release in the central nervous system. *Naunyn-Schmiedeberg's Arch Pharmacol* 317:36–43
- Bradford MM (1976) A rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein-dye binding. *Anal Biochem* 72:248–254
- Classen K, Göthert M, Schlicker E (1984) Effects of DU 24565 (6-nitroquipazine) on serotonergic and noradrenergic neurones of the rat brain and comparison with the effects of quipazine. *Naunyn-Schmiedeberg's Arch Pharmacol* 326:198–202
- De Lean A, Stadel JM, Lefkowitz RJ (1980) A ternary complex model explains the agonist-specific binding properties of the adenylate cyclase-coupled  $\beta$ -adrenergic receptor. *J Biol Chem* 255:7108–7117
- Deshmukh PP, Nelson DL, Yamamura HJ (1982) Localization of 5-HT<sub>1</sub> receptor subtypes in rat brain by autoradiography. *Fed Proc* 41:1338
- Dompert WU, Glaser T, Traber J (1985) <sup>3</sup>H-TVX Q 7821: identification of 5-HT<sub>1</sub> binding sites as target for a novel putative anxiolytic. *Naunyn-Schmiedeberg's Arch Pharmacol* 328:467–470
- Engel G, Göthert M, Müller-Schweinitzer E, Schlicker E, Sistonen L, Stadler PA (1983) Evidence for common pharmacological properties of [<sup>3</sup>H]5-hydroxytryptamine binding sites, presynaptic 5-hydroxytryptamine autoreceptors in CNS and inhibitory presynaptic 5-hydroxytryptamine receptors on sympathetic nerves. *Naunyn-Schmiedeberg's Arch Pharmacol* 324:116–124
- Engel G, Hoyer D, Kalkman HO, Wick M (1985) Pharmacological similarity between the 5-HT<sub>D</sub> receptor on the guinea pig ileum and the 5-HT<sub>2</sub> binding site. *Brit J Pharmacol* 84:106P
- Feldman HA (1972) Mathematical theory of complex ligand-binding systems at equilibrium: some methods of parameter fitting. *Anal Biochem* 48:317–338
- Fozard JR (1983) Functional correlates to 5-HT<sub>1</sub> recognition sites. *TIPS* 288–289
- Furchgott RF (1972) The classification of adrenoceptors (adrenergic receptors). An evaluation from the standpoint of receptor theory. In: Blaschko H, Muscholl E (eds) *Handbook of experimental pharmacology. Catecholamines*, vol XXXIII. Springer, Berlin Heidelberg New York, pp 283–333
- Gaddum JH, Picarelli ZP (1957) Two kinds of tryptamine receptor. *Br J Pharmacol Chemother* 12:323–328
- Göthert M (1980) Serotonin-receptor-mediated modulation of Ca<sup>2+</sup>-dependent 5-hydroxytryptamine release from neurones of the rat brain cortex. *Naunyn-Schmiedeberg's Arch Pharmacol* 314:223–230
- Göthert M (1982) Modulation of serotonin release in the brain via presynaptic receptors. *Tr Pharmacol Sci* 3:437–440
- Göthert M (1985) Effects of 5-hydroxytryptamine at presynaptic and neural sites. In: Bevan JA et al. (eds) *Vascular neuroeffector mechanisms*. Elsevier, Amsterdam, pp 315–320
- Göthert M, Schlicker E (1983) Autoreceptor-mediated inhibition of <sup>3</sup>H-5-hydroxytryptamine release from rat brain cortex slices by analogues of 5-hydroxytryptamine. *Life Sci* 32:1183–1191
- Göthert M, Weinheimer G (1979) Extracellular 5-hydroxytryptamine inhibits 5-hydroxytryptamine release from rat brain cortex slices. *Naunyn-Schmiedeberg's Arch Pharmacol* 310:93–96
- Göthert M, Huth H, Schlicker E (1981) Characterization of the receptor subtype involved in alpha-adrenoceptor-mediated modulation of serotonin release from rat brain cortex slices. *Naunyn-Schmiedeberg's Arch Pharmacol* 317:199–203
- Gozlan H, El Mestikawy S, Pichat L, Glowinski J, Hamon M (1983) Identification of presynaptic serotonin autoreceptors using a new ligand: <sup>3</sup>H-PAT. *Nature* 305:140–142
- Hamon M, Bourgoin S, Gozlan H, Hall MD, Goetz C, Artaud F, Horn AS (1984) Biochemical evidence for the 5-HT agonist properties of PAT [8-hydroxy-2-(di-n-propylamino)tetralin] in the rat brain. *Eur J Pharmacol* 100:263–276
- Héry F, Ternaux JP (1981) Regulation of release processes in central serotonergic neurons. *J Physiol (Paris)* 77:287–301
- Hoyer D, Engel G, Kalkman HO (1985a) Characterization of the 5-HT<sub>1B</sub> recognition site in rat brain: binding studies with (–)[<sup>125</sup>I]iodocyanopindolol. *Eur J Pharmacol* (in press)
- Hoyer D, Engel G, Kalkman HO (1985b) Molecular pharmacology of 5-HT<sub>1</sub> and 5-HT<sub>2</sub> recognition sites in rat and pig brain membranes. Radioligand binding studies with [<sup>3</sup>H]5-HT, [<sup>3</sup>H]8-OH-DPAT, (–)[<sup>125</sup>I]iodocyanopindolol, [<sup>3</sup>H]mesulergine and [<sup>3</sup>H]ketanserin. *Eur J Pharmacol* (in press)
- Langer SZ, Moret C (1982) Citalopram antagonizes the stimulation by lysergic acid diethylamide of presynaptic inhibitory serotonin autoreceptors in the rat hypothalamus. *J Pharmacol Exp Ther* 222:220–226
- Leysen JE, Niemegeers CJE, Van Nueten JM, Laduron PM (1982) [<sup>3</sup>H]Ketanserin (R41468), a selective <sup>3</sup>H-ligand for serotonin<sub>2</sub> receptor binding sites. Binding properties, brain distribution and functional role. *Mol Pharmacol* 21:301–314
- Martin LL, Sanders-Bush E (1982a) Comparison of the pharmacological characteristics of 5-HT<sub>1</sub> and 5-HT<sub>2</sub> binding sites with those of serotonin autoreceptors which modulate serotonin release. *Naunyn-Schmiedeberg's Arch Pharmacol* 321:165–170
- Martin LL, Sanders-Bush E (1982b) The serotonin autoreceptor: antagonism by quipazine. *Neuropharmacology* 21:445–450
- Middlemiss DN (1982) Multiple 5-hydroxytryptamine receptors in the central nervous system of the rat. In: Belleruche J, de (ed) *Presynaptic receptors: mechanisms and functions*. Ellis Horwood, Chichester, pp 46–74
- Middlemiss DN (1984a) 8-Hydroxy-2-(di-n-propylamino)tetralin is devoid of activity at the 5-hydroxytryptamine autoreceptor in rat brain. Implications for the proposed link between the autoreceptor and the [<sup>3</sup>H]5-HT recognition site. *Naunyn-Schmiedeberg's Arch Pharmacol* 327:18–22
- Middlemiss DN (1984b) Stereoselective blockade at [<sup>3</sup>H]5-HT binding sites at the 5-HT autoreceptor by propranolol. *Eur J Pharmacol* 101:289–293
- Middlemiss DN (1985)  $\beta$ -Adrenoceptor antagonists and blockade of the 5-HT autoreceptor in rat frontal cortex. *Br J Pharmacol* 84:189P
- Middlemiss DN, Fozard JR (1983) 8-Hydroxy-2-(di-n-propylamino)-tetralin discriminates between subtypes of the 5-HT<sub>1</sub>-recognition site. *Eur J Pharmacol* 90:151–153
- Nelson DL, Schnellmann R, Smit M (1983) <sup>3</sup>H-Serotonin binding sites: Pharmacological and species differences. In: Segawa T, Yamamura HI, Kurijama K (eds) *Molecular pharmacology of neurotransmitter receptors*. Raven Press, New York, pp 103–114
- Pazos A, Palacios JM (1985) Quantitative autoradiographic mapping of serotonin receptors in the rat brain. 1. Serotonin-1 receptors. *Brain Res* 346:205–230
- Pazos A, Engel G, Palacios JM (1985)  $\beta$ -adrenoceptor blocking agents recognize a subpopulation of serotonin receptors in brain. *Brain Res* 343:403–408

- Pazos A, Hoyer D, Palacios JM (1984a) Mesulergine, a selective serotonin-2 ligand in the rat cortex, does not label these receptors in porcine and human cortex: evidence for species differences in brain serotonin-2 receptors. *Eur J Pharmacol* 106:531–538
- Pazos A, Hoyer D, Palacios JM (1984b) The binding of serotonergic ligands to the porcine choroid plexus: characterization of a new type of serotonin recognition site. *Eur J Pharmacol* 106:539–546
- Pedigo NW, Yamamura HJ, Nelson DL (1981) Discrimination of multiple [<sup>3</sup>H]-5-hydroxytryptamine binding sites by neuroleptic spiperone in rat brain. *J Neurochem* 36:220–226
- Peroutka SJ, Snyder SH (1979) Multiple serotonin receptors: Differential binding of [<sup>3</sup>H]-5-hydroxytryptamine, [<sup>3</sup>H]lysergic acid diethylamide and [<sup>3</sup>H]-spiroperidol. *Mol Pharmacol* 16:687–699
- Peroutka SJ, Lebovitz RM, Snyder SH (1981) Two distinct central serotonin receptors with different physiological functions. *Science* 212:827–829
- Petersen EN, Olsson SO, Squires RF (1977) Effects of 5-HT uptake inhibitors on the pressor response to 5-HT in the pithed rat. The significance of the 5-HT blocking property. *Eur J Pharmacol* 43:209–215
- Richards MH (1985) Efflux of <sup>3</sup>H-5-hydroxytryptamine from rat hypothalamic slices by continuous electrical stimulation: Frequency-dependent responses to serotonergic antagonists and 5-hydroxytryptamine. *Naunyn-Schmiedeberg's Arch Pharmacol* 329:359–366
- Schlicker E, Göthert M (1981) Antagonistic properties of quipazine at presynaptic serotonin receptors and  $\alpha$ -adrenoceptors in rat brain cortex slices. *Naunyn-Schmiedeberg's Arch Pharmacol* 317:204–208
- Schlicker E, Göthert M, Hillenbrand K (1985) Cyanopindolol is a highly potent and selective antagonist at the presynaptic serotonin autoreceptor in the rat brain cortex. *Naunyn-Schmiedeberg's Arch Pharmacol* 331:398–401
- Snedecor GW, Cochran WG (1973) In: Snedecor GW, Cochran WG (eds) *Statistical methods. Curvelinar regression*, 6th edn. The Iowa State University Press, Ames, pp 447–471
- Starke K (1981) Presynaptic receptors. *Annu Rev Pharmacol Toxicol* 21:7–30
- Vaastra WJ, Deiman-Van Aalst WMA, Eigeman L (1981) DU 24565, a quipazine derivative, a potent selective serotonin uptake inhibitor. *Eur J Pharmacol* 70:195–202

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