

# Genome of the European Elk Papillomavirus (EEPV)

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## **Abstract**

The genome of the European elk papillomavirus (EEPV) was found to be 8,095 base pairs (bp) long and its genetic organization was similar to that of other papillomaviruses. Ten open reading frames (ORFs), designated E1-E7 and L1-L3, were identified in the genome, all located on one strand. The presence of the L3 ORF is rare among the papillomaviruses and to date has only been identified in the genomes of EEPV, the deer papillomavirus (DPV) and the Cottontail papillomavirus (CRPV). The ORF is well conserved between DPV and EEPV with regard to both length and sequence. Potential promoter regions were identified at the 5'-end of the E6 ORF, at the 3'-end of the E1 ORF and downstream of the L1 ORF. Furthermore, two potential polyadenylation signals were found, one located in the long control region (LCR), downstream of the L1 ORF, and another preceding the L2 ORF. The EEPV genome is closely related to the genome of the DPV, the most highly conserved regions being ORFs E1 (70%), E5 (69%), and L1 (74%).

## **Introduction**

Papillomaviruses belong to the Papovaviridae family and consist of a protein capsid surrounding a circular double-stranded DNA of approximately 8,000 base pairs (bp). They have been isolated and characterized from many different animal species (for references, see 1). The papillomaviruses induce proliferation of either the epithelial layer (papillomas), or the fibroblastic layer (fibromas), or both these layers (fibropapillomas). The majority of the papillomaviruses cause papillomas, but a few also cause fibromas or fibropapillomas.

We have focused our attention on viruses from the two latter groups. Warts induced by the European elk papillomavirus (EEPV) and the deer papillomavirus (DPV) are mainly fibromas, while the reindeer papillomavirus (RPV) and the bovine papillomavirus types 1 and 2 (BPV-1 and 2) give rise to fibropapillomas (2-5). EEPV also causes benign lung fibromas in elks (9). All of these viruses transform cultured murine fibroblast cells and induce sacromas in hamsters (2-4,6-8). Moreover, their genomes are more closely related to one another than to other papillomaviruses. Under stringent hybridization conditions EEPV DNA hybridizes to the genomes of DPV and RPV, but not with the BPV-1 genome that is more distantly related to EEPV (4).

The biological properties of EEPV have been described previously (2) and recently the nucleotide sequence of the early (transforming) region of EEPV has been published (10). We report here the complete nucleotide sequence of the EEPV genome. A molecular analysis of the late region made possible a comparison of regions which are equivalent to previously characterized regions in BPV-1, BPV-2, DPV and human papillomavirus type 6 and 16 (HPV-6 and 16) (11-16).

EEPV was isolated from the epithelial layer of a cutaneous fibropapilloma. The viral DNA was cloned into the plasmid vector pBR322 and subclones were constructed to facilitate sequence analysis as described before (2,10). The nucleotide sequence, obtained by the technique of Maxam and Gilbert (17), was analyzed using the University of Wisconsin Genetic Computer Group (UWGCG) software (18). The strategy for sequencing the late region is shown in Fig. 1 and the complete 8,095 bp nucleotide sequence of EEPV starting with the first nucleotide of the *Hpa*I recognition site is presented in Fig. 2.

The sequence of EEPV DNA revealed that all open reading frames (ORFs) longer than 400 bp are located on one strand (Fig. 3). Ten ORFs were identified, seven in the early region (E1-E7) and three in the late region (L1-L3) (Table 1 and Fig. 3). Three additional potential ORFs were identified, one 309 bp long (ORF $\times$ ) located between the E5 and L2 ORFs, and two located in the long control region

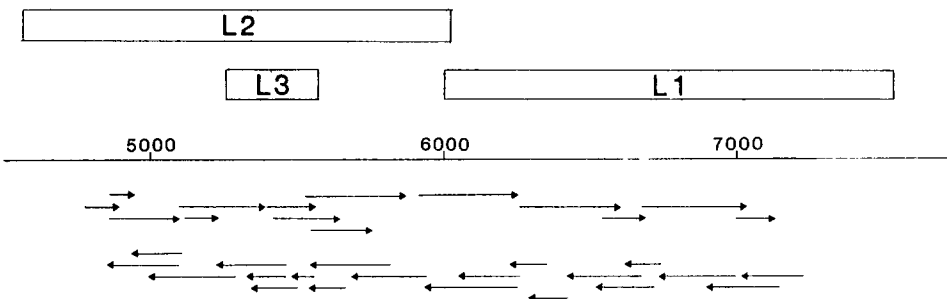


Fig. 1. The strategy for sequencing the late region of EEPV. The arrows show the direction in which the sequence was determined.

1	GTAAACAATC	ACCAGATCTT	GCCCGTTTTT	GTGAGCGGGA	AAGCTGGTTA	CAGGTTTATA	60
61	TAAAAGAGCC	CACCCGACAA	GTTTTACAG	ACGGTTT CAG	ATACTTCTAA	TACATGCATG	120
121	TGTGGCGAAT	GCTATGCATA	CCTCACCTGC	ATCTGGTGCA	AGAAGGGCTT	AGATAAGGTA	180
181	GATGCAAAGC	GATGCCATGA	AAAAAAAAAT	AGAATAGCGT	GCAGGAACCG	AAAACATTTG	240
241	GCTGTCTGTA	CATCTTGCCT	GGAAAAATGG	CTGTACCTTG	AAAGGTCCTT	TTTTCTTGGG	300
301	CGACCCATCT	ACCCTGGAGA	CCTGTATGAG	CCCGATCCAT	GGGTCATGTT	CAACGACATT	360
361	AGATGCATGT	ATTGTGGTGG	ATGCCTAAC	CGCGACGAAA	AAGAGAGACA	CAGACTGTTT	420
421	TGTGAAGACT	TCTGGATATT	CAGGCATCAG	GTGCGGGGAC	GTTGTCTACT	CTGCACCAGG	480
481	CATGGTTTAC	GGCCCCCGTA	CAAAGAACA	CCTGCCGCCG	TATGAATCAC	CTCCCCAC	540
541	ACTGCTCCTA	GAGCCAGTTG	CTCCGGTGCA	ACAGACAGGC	ATTCAGGCAC	CGCAGAGGAA	600
601	GCCACCTTCC	CAGAAAGGAC	ACAAAAAAG	ACACAAGAAA	GTTTATTCTG	TGACTGTGCC	660
661	TTGCAATGGA	TGTGACAAAA	ACCTGGAATT	TTGTGCAAGA	ACTTCCAGCG	CCACCATCTT	720
721	AACCGTGC AA	AACCTCTCTG	TGAAAGACCT	AGACTTCTCTG	TGCTCTACTT	CGGAGACCAA	780
781	CCATGGCTGA	AACTGCAGGT	AGCTCGGGGC	AGGGGGGGGG	AGCTTATATC	TGCTTTGAAG	840
841	CCGACTGTAG	CGACTCTGAT	ACAGAGGTTG	ATTCACCTGT	ACAATGCTCT	GATTCAAAGT	900
901	ATGAGGATCT	AGTAGATAAT	GCCAAATCTG	TTCCGGGAAA	CCACTGGGAG	TGTGTTCCAA	960
961	CGCAGGAAAA	AGAGGCGGGA	GAAAGACAGA	TTTTGCCTTT	GAAAAGAAAA	TTCTGTTTGA	1020
1021	GCCCGGGAAC	CTCAGAGGTC	GAGGAGCTTA	GTCCTGGGCT	TGCCGGAACT	AGAATCTCTC	1080
1081	CGCCAAAAGC	AAATCCGGTG	GTTAGGAGAA	GGCTTTTGA	CGCAGGTGGG	AGAGACGCCG	1140
1141	TGGGAACACC	CGGTGATCAT	GAAGTTAATA	GTTCTCTCTG	ACCCAGGAGT	CAGGTACAGT	1200
1201	CGGGAAGTAG	TAGTAGGCTT	TGGGAGGAC	ATCTGGAATC	CATTAAACGAG	CCTGTATAGT	1260
1261	ACGGCAACAT	GGCCGCCGTG	ATGCACAAGT	TGTTCAAGAC	TTTGTACATC	CGGGGTTTTG	1320
1321	GGGAGATAAC	ACGCGTCTTT	CAAAGTGATA	AAACTAACAA	TAAATCAGTG	GTGATAGCAG	1380
1381	CCCATGGCGC	ATCAGAGGTG	CTTTATGCCG	CAAGCTTTGA	AATACCTGAG	AAACCTGCA	1440
1441	GCTACTCTGA	GGCGTCTAGG	AAGGTGTCATG	AGACAGGAAG	CATGCTTTTG	TTCTTAGCTG	1500
1501	TCTTCAATGT	TGGGAAGATG	AGGGAGACTG	TCAGAAAAT	ATTTCCAGT	CTCTTAAACA	1560
1561	CCCGGTGTAG	CCGCTACTFA	TGCAACCCG	CGAAAATTCG	TGGACTATGT	CCTGCTTTAT	1620
1621	TTTGGTTTTAA	GTTGGGGCTC	TCCCCAGCAA	CACAGACGCA	CGGTACGACT	CCGGACTGGA	1680
1681	TTAAGCAGCA	GACCAATGTG	GCCATAAATA	CTGGGGAGGC	TCTTAAATTT	GATTTGGCAA	1740
1741	CAATGGTACA	GTGGGCATAT	GACCACC GGC	TAAACAGAGGA	GTGCAAAATT	GCATATCAAT	1800
1801	ATGCAAAATG	TGCAGGTACA	GACCTAAATG	CGAAAAGCATT	TCTTGCAAGT	ACCAATCAGG	1860
1861	CACGGCTGGT	CAAGGACTGC	TGTACTATGG	TGAAAACATTA	CCTGAGAGCT	GAAGAGCAGT	1920
1921	CATTAACCAT	TTCTGCTTTT	ATTAATAAGGA	GATGCGATAA	TGCAACTGGA	AAAGGCAGTT	1980
1981	GGTTGAGCAT	TATGAATCTG	TAAAGTTTTC	AAGGCATCGA	GCCCAATGCA	TTTGTAATAG	2040
2041	CCTTGA AAC	ATGGCTGAAA	GGCACC CCAA	AACATAATTG	CATAGCAATT	GTAGGACCCC	2100
2101	CAATAGTGG	GAACTCTCTT	CTGTGCAATA	CCCTCATGTC	GTTTCTGGGA	GGAAAGGTCA	2160
2161	TGACGTTTTG	CAACCACTCC	AGCCACTTCT	GGTTAGCGCC	CCTTACCAGC	TGTAGGGTGC	2220
2221	CCTTGATAGA	TGATGCCACG	CATGCGTGCT	GGAGATACCT	TGACACATAT	CTCAGAAATG	2280
2281	TACTTGACGG	TTATCCAGTT	TGTATTGACA	GAAAGCACAA	ATCCGCTGTG	CAGCTCAAAG	2340
2341	CCCTTCCCTT	TTTGCTAAC	AGTAATATTG	ATGTGCATCG	AGATGAAAAG	TATTTCTATC	2400
2401	TGCAAAAGT	AGTCAAAACC	TTCTATTTCA	AGGAGCCGTG	CCCTGCGCTT	GATACTGGTG	2460
2461	AGCCCTTTT	CTTTTACT	GATGCTGACT	GAAAAAATTT	TTTTGAAAGG	CTATGGGGAG	2520
2521	GATTAGATCT	CAGCGACCAA	GAGGACGAGG	TTGATGAAGA	TGAGTGCAGC	CAGCGATCAT	2580
2581	TTACTTGCAG	CGCAAGAAAC	ACAGATGCAA	TGCATTGAGA	AAGATAGTCG	CCTGTTACAG	2640
2641	GATCATGCAT	GCTATTGGGG	GGCAGTAAGA	AGGGAAAAAC	TGTTATTATA	TGCACCGAGA	2700
2701	ACAAAGGGGT	TAAAAACAAT	TGGGTGTGTG	CCTGTGCCCT	CTTGTCTCTG	TACTGCAGAG	2760
2761	CAAGCGAAGC	AAGCAATATG	CATGCAATTG	ATTGTGGAGG	AATTACTGCA	CAGTCCATGG	2820
2821	GCCAAAGAAC	CATGGTCCCT	TACAGACCTA	AGCTGGGAGA	GATATCAGGC	TGCCCCAAAA	2880
2881	GGGTGTTTTGA	AAAAAGGGCC	CAGAGTGGTG	GAAGTGGAGT	ATGATGGGAA	CTCTTCTAAT	2940
2941	AAGACTTGGT	ATACAGCTTG	GAGTACAGTG	TACGTGCGCG	GAACGGAAAG	GGAGGGCTGG	3000
3001	GAGACTGTCTG	TCTGTGCTGC	AGACGGACAG	GGCATTATT	ATTGCGCCGG	GATGAGCAGT	3060
3061	AAGGTGTACT	TTGAAACCTT	TGAAACTGAT	GCCCGCAGAT	GGAGCAGGAC	GGGGCACTGG	3120
3121	ACTGTGAGGG	ATAACGATGT	GATATATCAT	TCAACCTTTG	GTGCACCCTC	TCACCTTAGA	3180
3181	AACGACAGAG	ACTGCATCGA	AGGATTCTGG	AGCCGACCGC	GGGAGCCGTAG	AGGCTCGAGA	3240
3241	GGGTCCGACA	CACCAAGACAG	AGCCCTGCCT	TACCCTGCTG	CTCGACAATC	CCCCATTTGT	3300
3301	CGCCCCGTCA	GAACTGGCGA	AAACCGGAGT	CGGGCCGTTT	ACCGCCAGGC	TCCCTACAGC	3360
3361	GCACCATCAT	CCCCGGGGAG	TTCCTGGGG	CCCGATTCCC	CCTCCGAGAG	CTCGCCGACG	3420
3421	TGACCCGCTGG	TTTTGTCTACC	AGGACCATCA	GATCCAGCGC	CGCGCTGCC	GGACTCTACA	3480
3481	GACGTAATCG	CAGAGGGTGA	CAAGGAACCT	GAGCGGTTCA	GCATTTCTCT	AAAACCCAGT	3540
3541	GGGCAGCCAT	GTCTGATACT	TAGTGGAAAC	GGAACCAAG	CTAAGTGCTA	TCGTTTCCGG	3600
3601	TGCAAGAGAT	ATTTACAGAG	ACACTATCAG	CACATAACGA	CCACTGGGTG	GACTGTAGGA	3660
3661	GAGCGAGGAT	CTGAAAGGCA	CGGAGATGCC	TGTGTGCTGG	TGACATTCAA	AGACAGTTCC	3720
3721	CAGAGAGGGG	TGTTTTTGA	CGGAGTGCTT	TTGCCACCTG	GAATCGCGC	GCAGGCAGT	3780
3781	ACAATGATTG	CGGACTTTTG	AGAATGTGAC	TCTGGTAGCC	GCAATACTGT	GCAGTTTGCT	3840
3841	CACCTTCATG	TTTTACCTGT	GTAGCACAGA	CTGCTGCATG	CTGTGATGAC	ATACGGTTTTG	3900

Fig. 2. The complete 8095 nucleotide sequence of EEPV, starting with the first nucleotide in the *Hpa*I recognition sequence. The sequence data for the early (transforming) region is from Ahola et al. (10).

3901	CTTTTGTTC	TGGGGCTCAC	ATTTGGACTA	CAGCTGATGC	TACTTGTCTT	TCTGCTGTTT	3960
3961	TFCTTCTCG	TATGGTGGGA	CCAGTTTGGC	TGCCGTTGTG	AAAACATGCA	GTTGTAATA	4020
4021	GTGTATATTG	AGGTGTAGAT	ATTCATTTGA	TCCTGTACAT	ACATTTTCTC	TATTTTTTTA	4080
4081	AAAAATGCTG	GTTGATAAAC	ATACATAGTT	CACAAAACAGG	TCATTTCCATA	ACGTCAACAA	4140
4141	TACTTTTTAT	AGCCGTGGAG	TCTCTTATCT	CTGTCCCTTT	GTTTCTCTCT	TTTGAACGTG	4200
4201	TATCCGAGTC	AGCAAGTGCC	ATTTTTGTGT	CCATTTTCTT	CAAGTGCCAA	TCTCCTAACT	4260
4261	GCATCCCCAA	GAGCTGGTGT	AACGTGAGAA	GAACAGTCGA	GAATCTAGCC	TTTGAAGAC	4320
4321	TTTCAACCC	CAAAACAGACC	ATGTGCATAC	CTCCTGTAAC	AGCAAAGCTG	CCACGACATT	4380
4381	GAAGCCTGCA	TGAAATATTG	TTTACTATTG	TTTTTGCTGC	TATTTGCTGGG	GCAGTGGAA	4440
4441	CCAATGTGGG	TATTGTACT	AATTGTCTGG	TGTCCATTG	TACTTTTATT	TGAGTTGCAT	4500
4501	TTTGTGGAAC	ACTTCCACATG	ACAGGCTGCA	CTGCAGCGCC	CTCATCTCAT	CCCTAAATTT	4560
4561	AATAAACCTT	CCCTTATTTA	ACCCCTACCA	TGGCGCCTCG	GCGAGTAAAG	CGTGCAATG	4620
4621	TCTATGACCT	GTATCGCACA	TGCAAGCAGG	CAGGCACCTG	TCCTCCGGAT	GTGATACCTA	4680
4681	AGGTGGAAAG	GAAAGCAGTA	GCAGACAAGA	TATTGCAGTA	TGGAAGCATTG	GCCTTTTATT	4740
4741	TAGGCGCCCT	AGGCATTTGA	ACAGGTTCTG	GAAAGCCAGG	AACAGGAGTC	GATATTCCAC	4800
4801	TCAGAGGTGG	GGGCTCTACC	ACTTCACTAT	CAAGCAAACC	TTTTGTGGG	GGGATACCTT	4860
4861	TAGAAACCTT	AGAAGGGATA	GGGGCATTCC	GGCCTGGCAT	AGTGGAAAGT	GCAGGGCCTG	4920
4921	CTTTAGAAGG	CATTCCTCC	GACGCACCAG	CAGTTGTCC	GCAGGAGCCA	TCTGGAGTGG	4980
4981	ATGAGGGGTT	AAAGTGGGCTA	GATATTTCCA	GGGAATTAAG	CAGGAAACAA	ATPCTCAGCT	5040
5041	TTCTCCACCC	TGAGGGTCCG	GATGATATTG	CAGTACTTGA	GCTAAGGCCA	CGTCAACATG	5100
5101	ATCAGGCACA	TTTGTGTCT	ACAAGCACAC	ACCCAAATCC	ACTGTTTCAG	GGTCTGTGAC	5160
5161	AGCAGGCACG	AAATATTGCA	GAAACATCTG	GTGCAGAAAA	CGTTTTTGTG	GGTGGAAAGT	5220
5221	GCATTTGGAAG	CAATGCAGGA	GAGGACATTG	AACTGACACT	GTTTGTCTGAA	CCAAGGACAA	5280
5281	GTACACCTGA	GGTGCCTATT	AAACGTTCTC	GGGGCATTTC	CAATTGGTTC	AGCAGGCGCT	5340
5341	ACTATACACA	GGTACCTGTC	GAAAGCCAG	ACGAGATTGC	TGCTGCAGGC	TCGTATGCT	5400
5401	TTGAGAATCC	TGTATACGAT	TCAAAGCGGT	TCAAACCTGC	GCAGCAGCCG	GCATATTACT	5460
5461	TACAGGATGA	AGTCTCTGTC	ACTGGGCGGG	ACGCTGCAAG	ATTTGCTGGC	GGACCCTCGT	5520
5521	GCAAGGATTG	GTGGAGTCTG	ATCACACGAC	CCACTAGTCT	TGGAAACAGT	AGTCCGGTGA	5580
5581	GGGTAGGCC	TCTTTATCAT	TTACGATCCT	CTTTCAGCAC	TATCCATAGT	CCTGAGACAA	5640
5641	TAGAGCTAAT	ACCCACAGTA	CTTGAGGATG	ATACTGAGGT	GCTTACAGTT	GTCTTCGAGA	5700
5701	GAGACTAGCT	TTTTGATGAT	GTGGATTTGG	ACAGTATAGC	AAAGTGACAGT	CCATTACTAC	5760
5761	CTGAGCGGCA	TCACCTTGCT	TTTGGAGCAA	GGCGGTCTCA	CATTCCAATT	GTGGCACGAC	5820
5821	CAGGTGTCCA	AACTGGTACA	GTGATTTGTA	CACGTCAGAT	GGCTGAAAA	TCGTGTTACG	5880
5881	TGTCGGACAA	TGGAGGACAG	GAGTCCACAG	AGACGCCAC	TGTGGTAATC	AATGGCAACA	5940
5941	TTAATGTGTC	CATGGAATAT	TTTAGGCATT	ACTATTTGCA	CCCTAGCCTT	CTAGGTCGCA	6000
6001	AACGAAAACG	TCTATTCCGT	TAATGTTTTA	CAGATGGCGT	TCTGGCAGCC	TAGTCAAAGG	6060
6061	CTATACCTGC	CTCCACACCC	TGTGACAAAG	GTGCTGTGCT	CTGAGCAATA	TATTAGCGT	6120
6121	AAGGACGTAT	TTTATCACGG	GGAGACGGAG	CGCATGTCTA	CTGTAGGGCA	TCCATATTAT	6180
6181	GAAATTAAC	AATCAGGGTC	TGGGAAAACC	ATTCCAAAGG	TTTCACTTAA	TCAATTCGT	6240
6241	GTTTTTCGGA	TCTTACTGCC	GGATCCCAAC	CAGTTTGCTC	TTCAGATAAA	TAGCATGTAT	6300
6301	GACCCAAAGT	AGGAAAAGCT	AGTCTGGGCT	GTTGTGGGGG	GTTGCTGTCT	AGGACGACAA	6360
6361	CCTTTAGGTG	GCTCTGTTTC	AGGACATTCG	TATCAGAACA	CTCTGATTGA	TGCGGAGAA	6420
6421	GTTAGTAAAA	AGGTAATATC	ACAGGGCACA	GATGACAGGA	AGCAGGGAGG	CATGGACGTC	6480
6481	AAGCAACAGC	AAATTCCTACT	GCTAGGATGC	ACCCACAGTA	TTGGTGAGTA	TTGGACAAC	6540
6541	GCTAGGCCCT	GCCTTACAGA	TAGGCCAGAG	ACTGGCTCCT	GCCCCCTAT	AGAACTAAAA	6600
6601	AACAAACCTA	TAGAAGATGG	TGATATGATG	GATATTGGCT	TTGGTGCAGC	TAATTTCAA	6660
6661	GAGTTAAATG	CCACAAAAGT	AGATPCTCCCT	TTAGATATTG	AAAAGATAT	TTGTTGTAT	6720
6721	CCTGATTTAT	TAAAGATGAC	TGAAGAAGCG	GCTGGCAACA	GATAGTTTTT	TTTTGTCTCG	6780
6781	AAAGAACAAG	TTTTATGTTG	CCACATCTGG	TCGCGTGGGG	GTACCGACAA	AGAAATGCCT	6840
6841	CCAGAGGCAT	ACTTCTGAA	GCCAAAGGTT	GGGGACAAA	CACAGAAAAT	GCCTAGTATT	6900
6901	CTTTTGGGAG	TGCCAAGTGG	CAGTTTAGTT	TCTACAGATG	GACAATTGTT	TAATAGACCT	6960
6961	TACTGGCTGT	TTCCGTGACA	GGGCATGAAT	AATGGCATAT	GCTGGCTTAA	TCAACTGTTT	7020
7021	GTTACTGTTG	GTGACAATA	AAGAGGAACC	ACATTAACCA	TTCAGGTGCC	TACATCCGGG	7080
7081	TCCCCACTCA	CTGAATATGA	CACGAGCAA	TTTAATGTTT	TTCAAAGGCA	TTTGAAGAA	7140
7141	TATAAGCTTG	CCTTTGTATT	TCAGCTTTGC	TCTGTCACTC	TAAGTCCAGA	AACCGTCTCA	7200
7201	CATCTCCAGG	GGTTAATGCC	TTCCATCTCTG	GAACACTGGG	ATATTAACT	GCAGCCTGCT	7260
7261	ACGTCTCGTA	TTCTTGAGGA	TACTTACAGA	TATCTTGAAT	CACCTGCTAC	TGCTATGTCA	7320
7321	GATAATGTAA	CCCTATGGG	ACCTGAAGAT	CCCTATGCTG	GTTTAAAGTT	TTGGGAGGTT	7380
7381	AACTAAAAAG	AAAGGTTGTC	TCTTGTACTT	GATCAATTTT	CTCTGGGACG	CGGTTTCTT	7440
7441	GCGCAGCAAG	GATTAGGGTG	CAGTACTAGA	AAGAGGGTTG	CACTGGTCCC	TAAGGTACC	7500
7501	GAAAAAAGGA	TTGTTAGGAA	AAGAAGAAAG	GGGAATTAAG	GGCATGAAAT	CTTAAAAACT	7560
7561	GCTGTGTTTG	CTAAATAAAT	GCAATTTTTT	TTATGTGTCA	AGAGTTTATG	TGCTATGTCC	7620
7621	TGCTGTTCAG	TCCAACCTGC	ACCCACCCCG	GTGCTGGCAT	CTGATTAGAC	GCAGTGTCA	7680
7681	CAGCTTTATG	AAAAGCAGAC	ACTTGGCTAG	ACACACAGGC	CGCTGGCGCC	CTCATCGAAT	7740
7741	TGGCGCACCG	CTGGCGTTGG	GATCAAAAT	TCCTCTACCG	GCTGGGGTTG	TTAAAGCGCC	7800
7801	CTTCTGTAC	CGTTCGCGT	AGGCCCTCTT	CTCTCCCTTC	AGCGCTACCG	CTCCCGGTTG	7860

Fig. 2. (continued)

7861	GCATGGTAAG	TAGGCCGGTCA	TTGTCTGAAGA	GAAGTGGTAA	GCAAGTCCGA	ACAAGAAAAA	7920
7921	TGCTTGGCGC	AACGCTGACG	GTAGTCGCTA	CCGTCGCGCG	TGCTCGCTTT	TCTAAGAAAT	7980
7981	GCTCAAACGG	TCTCTTGCTA	GCTCTGCTCC	TATTGGCTGT	GCTGAAATTA	CTCACGCCGC	8040
8041	TTTGCCGTGA	CCGTGAACGG	TTTTGAATCC	TACTTTTTTCT	CAGGGAATGA	TTGTT	8095

Fig. 2. (continued)

(LCR), 263 and 371 bp long respectively (LCR1 and LCR2). The L1 and L2 ORFs in EEPV have their counterparts in all papillomaviruses studied so far, while the L3 ORF has only been identified in the genomes of DPV and the Cottontail rabbit papillomavirus (CRPV) (14,19). The L3 ORF is 321 bp long in both EEPV and DPV, and parts of these two ORFs show a high degree of homology (Fig. 4), whereas no significant homology was found when the L3 ORF of EEPV was compared to that of CRPV. The L1 and L2 ORFs have been shown to code for viral structural proteins (20), but no functional activity has yet been assigned to the L3 ORF.

When compared with other papillomavirus genomes, EEPV shows a higher degree of homology to DPV than to any other papillomaviruses. The ORFs L1 (74%), E1 (70%), and E5 (69%) are best conserved (Table 2a). The extent of homology between the L1 ORF of EEPV and either DPV, BPV-1, BPV-2, HPV-6, or HPV-16 was determined and is shown in Table 2b. The homologies of other ORFs between EEPV, DPV, and BPV-1 are listed in Table 2a. A noteworthy feature of the L1 ORFs in the papillomaviruses sequenced to date is the presence of a conserved postulated splice acceptor site preceding the putative initiator ATG in this ORF

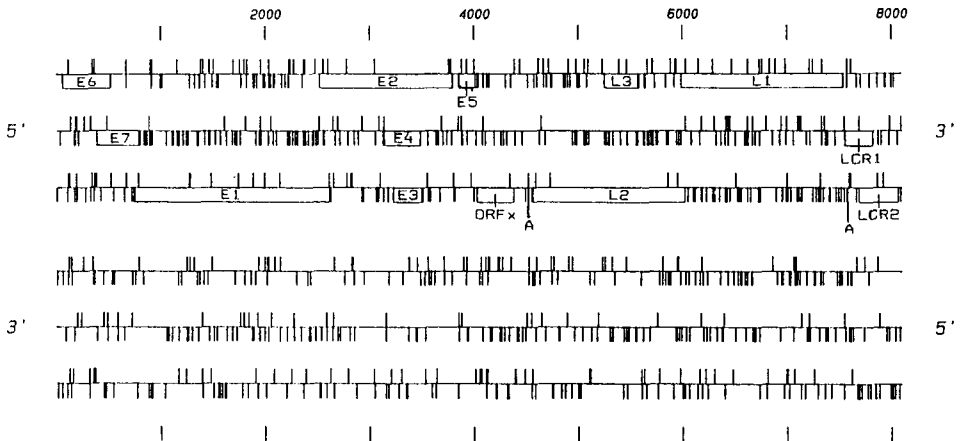


Fig. 3. Genomic organization of EEPV, including all ATG initiation codons (vertical bars above the lines) and termination codons (vertical bars beneath the lines). The ORFs in the genome are boxed. Potential polyadenylation signals are indicated with A. The sequence data for the early (transforming) region is from Ahola et al. (10).



*Table 2a.* Degree of homology at the nucleotide (NT) and amino acid (AA) sequence levels between different ORFs in EEPV, DPV and BPV-1, measured from the first ATG where applicable.

ORF	DPV		BPV-1	
	NT (%)	AA (%)	NT (%)	AA (%)
E1	70	71	57	54
E2	65	56	55	44
E3	—	—	53	28
E4	67	58	52	37
E5	69	72	60	53
E6	58	54	46	43
E7	54	46	46	40
L1	74	73	66	72
L2	60	64	43	37
L3	38	28	—	—

The sequence data was obtained from references 10-12, 14.

*Table 2b.* The degree of homology between L1 ORFs in EEPV, DPV, BPV-1, BPV-2, HPV-6 and HPV-16, measured from the first ATG in each ORF.

L1	NT (%)	AA (%)
DPV	74	73
BPV-1	66	72
BPV-2	66	71
HPV-6	52	46
HPV-16	56	46

The sequence data was obtained from reference 11-16.

(Table 3). This splice acceptor site has indeed been shown to be used in the case of BPV-1 by S1-analysis (22).

The G + C content of the EEPV genome is 47.7%. However, the bases are not evenly distributed in the genome. For example, the E3 and E4 ORFs (bp 3143-3497) have a G + C content of 62%. In the middle of the genome, including the E5 ORF, there is a T-rich area with a T-content of 35% (bp 3865-4566). A similar T-rich area has been observed in DPV DNA (14).

In the genome of BPV-1, promoter regions have been identified near positions 89 ( $P_{89}$ ), 2443 ( $P_{2443}$ ), and 3080 ( $P_{3080}$ ) (for references see 21). Recently, three additional promoter regions were identified in BPV-1 at positions 7185 ( $P_{7185}$ ), 7940 ( $P_{7949}$ ), and within the region 7214-7256, designated the major late promoter ( $P_L$ ) (21,22). Potential promoter regions were identified in the EEPV genome at positions 87, corresponding to  $P_{89}$  in BPV-1 (Fig. 5) and at 2392 corresponding to  $P_{2443}$  in BPV-1. A ten base pair repeated sequence (bp 7591-7600 and bp 7606-7615) was identified in EEPV. This has 8 of 10 nucleotides (nt) in common with the





flanking area of P<sub>7185</sub> (bp 7180–7189) in the BPV-1 genome. Moreover, a similar repeated sequence is present in DPV (bp 7811–7826 and 7836–7851). Upstream of the P<sub>L</sub> region in BPV-1 there is a sequence homologous to the simian virus 40 (SV40) late promoter element GGTACCTAACC that has been shown to be important for efficient utilization of the SV40 major late transcriptional start site (21, 23, 24). It is interesting that this sequence homology is present in EEPV at approximately the corresponding location as in BPV-1, whereas no such sequence is found in this part of the genome in DPV. No area similar to the BPV-1 promoter region P<sub>3080</sub> has been identified in the genome of EEPV.

Two potential polyadenylation (pA) signals (AATAAA) are present in the EEPV genome (Fig. 3). One is located immediately upstream of the L2 ORF (bp 4561) (putative early pA signal) and the other is located downstream of the L1 ORF (bp 7575) (putative late pA signal). Potential pA signals have been found at the corresponding locations in the genomes of DPV and BPV-1 (11, 14).

The repeated motif ACCGN<sub>4</sub>CGGT that has recently been shown to be part of two E2-binding regions in BPV-1 (25, 26), is present in five copies in the LCR of EEPV (Table 4).

Taxonomically, elk and deer belong to the Cervidae family, and they show a similar host response to papillomavirus infections, i.e., mainly fibromas develop. Besides the biological similarities between EEPV and DPV, it is interesting that the molecular analysis indicates a higher degree of evolutionary relationship between them than with the bovine fibropapillomaviruses BPV-1 and BPV-2. On the basis of the sequence homology and the characteristic host response to infection, EEPV and DPV could be classed as a subgroup of fibromaviruses to distinguish them from the fibropapillomaviruses.

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*Table 4.* Putative E2 binding sites in EEPV.

Position	ACCGNNNNCGGT
7777	ACCGCTGCCGGT
7809	ACCGTTCCCGGT
7847	ACCGTCCCGGT
7950	ACCGTCCCGGT
8050	ACCGTGAACGGT

## References

1. Pfister H. Biology and biochemistry of papillomaviruses, *Rev Physiol Biochem Pharmacol*, 99:111-181, 1984.
2. Stenlund A., Moreno-Lopez J., Ahola H., & Pettersson U. European elk papillomavirus; characterization of the genome, induction of tumors in animals, and transformation in vitro, *J Virol* 48:370-376, 1983.
3. Groff D.E., Sundberg J.P., & Lancaster W.D. Extrachromosomal deer fibromavirus DNA in deer fibromas and virus-transformed mouse cells, *Virology* 131:546-550, 1983.
4. Moreno-Lopez J., Ahola H., Eriksson A., Bergman P., & Pettersson U. Reindeer papillomavirus transforming properties correlate with a highly conserved E5 region, *J Virol* 61:3394-3400, 1987.
5. Olson C., Gordon D.E., Robl M.G., & Lee K.P. Oncogenicity of bovine papillomavirus, *Arch Environ Health* 19:827-837, 1969.
6. Koller L.D., & Olson C. Attempted transmission of warts from man, cattle, and horses and of deer fibroma, to selected hosts. *J Invest Dermatol* 58:366-368, 1972.
7. Dvoretzky I., Shober R., Chattopadhyay S.K., & Lowy D.R. A quantitative in vitro focus assay for bovine papilloma virus. *Virology* 103:369-375, 1980.
8. Friedman J.C., Levy J.P., Lasneret J., Thomas M., Boiron M., & Bernard J. Induction de fibromes sous-cutanes chez le hamster dore par inoculation d'extraits acellulaires de papillomes bovins. *C R Acad Sci, Paris*, 257:2328-2331, 1963.
9. Moreno-Lopez J., Mörner T., Pettersson U. Papillomavirus DNA associated with pulmonary fibromatosis in European elk, *J Virol* 57:1173-1176, 1986.
10. Ahola H., Bergman P., Ström A.C., Moreno-Lopez J., & Pettersson U. Organization and expression of the transforming region from the European elk papillomavirus (EEPV), *Gene* 50:195-205, 1986.
11. Chen E.Y., Howley P.M., Levinson A.D. & Seeburg P.H. The primary structure and genetic organization of the bovine papillomavirus type 1 genome, *Nature* 299:529-534, 1982.
12. Ahola H., Stenlund A., Moreno-Lopez J., & Pettersson U. Sequences of bovine papillomavirus type 1 DNA—functional and evolutionary implications, *Nucl Acid Res* 11:2639-2650, 1983.
13. Potter H.L.Jr., & Meinke W.J. Nucleotide sequence of bovine papillomavirus type 2 late region, *J Gen Virol* 66:187-193, 1985.
14. Groff D.E., & Lancaster W.D. Molecular cloning and nucleotide sequence of deer papillomavirus, *J Virol* 56:85-91, 1985.
15. Schwartz A., Durst M., Demankowski C., Lattermann O., Zech R., Wolfspurger E., Suhai S., & zur Hausen H. DNA sequence and genome organization of genital human papillomavirus type 6b, *EMBO J* 2:2341-2348, 1983.
16. Seedorf K., Krämmer G., Durst M., Suhai S., & Röwekamp W.G. Human papillomavirus type 16 DNA sequence, *Virology* 145:181-185, 1985.
17. Maxam A., & Gilbert W. Sequencing end-labeled DNA with base-specific chemical cleavage, *Methods enzymol* 65:499-560, 1980.
18. Devereux J., Haeblerli P., & Smithies O. A comprehensive set of sequence analysis programs for the VAX, *Nucl Acids Res* 12:387-395, 1984.
19. Giri I., Danos O., Yaniv M. Genomic structure of the cottontail rabbit (Shope) papillomavirus. *Proc Natl. Acad Sci USA* 82:1580-1584, 1985.
20. Pilacinski W.P., Glassman D.L., Krzyzek A., Sadowski P.L., & Robbins A.K. Cloning and expression in *Escherichia Coli* of the bovine papillomavirus L1 and L2 open reading frames. *Biotechnology* 1:356-360, 1984.
21. Baker C.C., & Howley P.M. Differential promoter utilization by the bovine papillomavirus in transformed cells and productively infected wart tissues, *EMBO J* 6:1027-1035, 1987.
22. Stenlund A., Bream G.L., & Botchan M.R. A promoter with an internal regulatory domain is part of the origin of replication in BPV-1. *Science* 236:1666-1671, 1987.

23. Brady J., Radonovich M., Vodkin M., Natarajan V., Thoren M., Das G., Janik J., & Salzman P. Site-specific base substitution and deletion mutations that enhance or suppress transcription of the SV40 major late RNA. *Cell* 31:625-633, 1982.
24. Brady J., Radonovich M., Thoren M., Das G., & Salzman N.P. Simian virus 40 major late promoter: an upstream DNA sequence required for efficient in vitro transcription. *Mol Cell Biol* 4:133-141, 1984.
25. Spalholz B., Lambert P.F., Yee C.L., & Howley P.M. Bovine papillomavirus transcriptional regulation: localization of the E2-responsive elements of the long control region. *J Virol* 61:2128-2137, 1987.
26. Moskaluk C., & Bastia D. The E2 "gene" of bovine papillomavirus encodes an enhancer-binding protein. *Proc Natl Acad Sci USA* 84:1215-1218, 1987.
27. Danos O., Katiwala M., & Yaniv M. Human papillomavirus 1a complete DNA sequence: a novel type of genome organization among papovaviridae. *EMBO J* 1:231-236, 1982.
28. Zachow K.R., Ostrow R.S., & Faras A.J. Nucleotide sequence and genome organization of human papillomavirus type 5. *Virology* 158:251-254, 1987.
29. Fuchs P.G., Iimer T., Weninger J., & Pfister H. Epidermodysplasia verruciformis-associated human papillomavirus 8: genomic sequence and comparative analysis. *J Virol* 58:626-634, 1986.
30. Dartmann K., Schwartz E., Gissman L., & zur Hausen H. The nucleotide sequence and genome organization of human papillomavirus type 11. *Virology* 151:124-130, 1986.
31. Cole S.T. & Danos O. Nucleotide sequence and comparative analysis of the human papillomavirus type 18 genome. *J Mol Biol* 193:599-608, 1986.
32. Cole S.T., & Streeck R.E. Genome organization and nucleotide sequence of human papillomavirus type 33, which is associated with cervical cancer. *J Virol* 58:991-995, 1986.