Microbiology and Biotechnology

9 Spfinger-Verlag - 1982

Effect of Hydrolysing and Oxidizing Agents on the Composition and Degradation of Wheat Straw Monosaccharides

J. Miron and D. Ben-Ghedalia*

Institute of Animal Science, Agricultural Research Organization, The Volcani Center, P.O. Box 6, Bet Dagan 50250, Israel

Summary. Wheat straw (WS) was treated with 5% sodium hydroxide, ozone and 5% sulfur dioxide at 70 $^{\circ}$ C for 72 h, and the effect of treatments on monosaccharide composition and in vitro degradability by rumen microorganisms was studied. The major sugars, glucose and xylose, comprising about 90% of the total monosaccharides in the untreated WS were mainly confined to the cell walls. $SO₂$ exerted the greatest solubllizing effect, followed by ozone and NaOH; the respective values for the solubllized cell wall polysaccharides were: 26, 12 and 4.4%. One third of the total phenolics was oxidized by ozone, whereas, SO_2 exerted mostly a solubilizing effect on this fraction, converting 75% of it into soluble phenolics. In the NaOH treated WS 41% of the total phenolics were soluble, as compared to 22% in the untreated. The in vitro digestibility of monosaccharides in the untreated WS were initially high: 50% and 58% for xylose and glucose, respectively and 63% to 80% for the minor sugars. The SO_2 treatment resulted in an overall increase in digestibility of monosaccharides with values lying in the range of 90%. Sodium hydroxide was more efficient than ozone in enhancing the degradability of xylan and total sugars. The digestibility of cell wall sugars was increased from 52.4% to 84.4%, 63.4% and 72.3% by SO_2 , O_3 and NaOH treatments respectively. Based on the present findings, it appears that wheat straw cell wall components are more sensitive to hydrolytic than to oxidative processes aimed at increasing its degradability by rumen microorganisms. SO_2 exerted on WS a multi-effect which was particularly suitable for increasing the digestibility of monosaccharides.

Introduction

The matrix is usually the main target of attack in pretreatmerit studies of lignocellulosics. Its partial dissolution cre-

ates spaces in the cell wall and renders the cellulose fibers to be readily hydrolysed by cellulases. Much interest **has** been focused on the chemical and structural organization of the matrix of the plant cell wall unit, since those factors are major determinants of pretreatment effectiveness. Both hydrolysable and oxidative pretreatments have been used to dissolve matrix components in order to increase substrate degradability by fungal cellulase and by rumen microbes, (Millet et al. 1976; Binder et al. 1980; Ben-Ghedalia et al. 1980; Ben-Ghedalia and Miron, 1981).

Oxidative treatments are directed to mainly affect the lignin moiety, whereas hydrolysing agents are expected to influence the lignin-carbohydrate linkages and the hemicellulose. It is suggested that the response of a given lignocellulosic either to hydrolysis or to oxidative treatments reflects the relative importance of the potentially hydrolysable *versus* non-hydrolysable bonds, as biodegradation obstacles in the substrate.

In this study we examined the effect of sulfur dioxide and sodium hydroxide (as hydrolysing agents) and ozone (as oxidizing agent) on the composition and degradability of wheat-straw cell wall monosaccharides by rumen microorganisms.

Materials and Methods

Plant Material and Treatments. Ground (1 mm) wheat straw was used in this study. For treatment with sodium hydroxide, 100 g of milled straw was mixed with 100 ml of 5% NaOH, and kept at room temperature in plastic containers for 1 week, after which the material was freeze-dried and used for analyses. For ozonation, ground wheat straw was moistened to 40% and ozonized in glass columns $(80 \times 5 \text{ cm})$ until complete decolorization as described by Ben-Ghedalia and Miron (1981). After ozonation the material was freeze dried and used for analyses. For treatment with $SO₂$, ground wheat straw was moistened to 40% and placed in hermetically sealed glass columns (80 \times 5 cm). Gaseous SO₂ was added

Offprint requests to: D. Ben-Ghedalia

Monosaccharides	Untreated		Sulfur dioxide		Ozone		5% Sodium hydroxide					
	Compo- sition (g/100) g OM)	Distribution $(\%)$		Compo- sition	Distribution (%)		Compo- sition	Distribution (%)		Compo- sition	Distribution (%)	
		In cell walls	In cell solubles	(g/100) g OM)	In cell walls	In cell solubles	(g/100) g OM)	In cell walls	In cell solubles	(g/100) g OM)	In cell walls	In cell solubles
Glucose	41.6	91.8	8.2	41.4	87.7	12.3	40.6	90.8	9.2	40.9	94.0	6.0
Xvlose	20.2	93.8	6.2	20.8	45.9	54.1	18.4	70.7	29.3	21.2	81.7	18.3
Arabinose	2.84	72.7	27.3	3.10	6.6	93.4	3.01	23.0	77.0	3.44	58.6	41.4
Galactose	0.79	65.3	34.7	0.79	7.7	92.3	0.67	36.0	64.0	0.65	44.8	55.2
Mannose	0.40	65.5	34.5	0.42	33.0	67.0	0.36	58.0	42.0	0.32	59.9	40.1
Rhamnose	0.18	54.6	45.4	0.18	0	100	0.22	14.8	85.2	0.18	47.9	52.1
Uronic acids	4.08	82.1	17.9	4.19	41.3	58.7	4.59	56.1	43.9	4.16	69.7	30.3
Total sugars	70.1	90.5	9.5	70.9	67.7	32.3	67.9	79.1	20.9	70.9	86.5	13.5

Table 1. The composition (g/100 g OM) of monosaccharides and their distribution (%) in cell walls and cell solubles of wheat straw after treatment with sulfur dioxide, ozone and sodium hydroxide

Table 2. The content of cell walls $(CW)^{a}$, CW-glucose polymers^b, CW-non glucose polymers^c, soluble carbohydrates^d, and lignin in wheat straw after treatment with sulfur dioxide, ozone and sodium hydroxide (g 100 g^{-1} OM)

Constituent.		Untreated	Treatment				
			SO ₂	Ozone	5% NaOH		
Cell walls		81.9	55.4	65.2	76.7		
CW-glucose polymers		38.2	36.3	36.9	38.4		
CW-non glucose polymers		25.2	11.7	16.8	22.8		
Soluble carbohydrates		6.7	22.9	14.2	9.5		
Lignin (permanganate)		8.31	7.14	6.44	11.2		
Total phenolics		17.3	16.4	11.2	16.5		
Distribution: In cell walls		78.2	25.9	68.9	58.8		
	In cell solubles	21.8	74.1	31.1	41.2		

 a CW = NDF, neutral detergent fibre = depectinated cell walls

 $\frac{b}{c}$ Total CW glucose

 $\frac{c}{d}$ Total hemicellulose monosaccharides
d. Total monosaccharides soluble in neu

Total monosaccharides soluble in neutral detergent

to the straw at a level of 5 g $100 g^{-1}$ dry matter and then the sealed columns were placed in an oven at 70° C for 72 h, after which the material was aerated, freeze-dried and used for analyses. Each treatment consisted of four replicates.

Analytical Procedures. Analyses were performed on the four individual samples for each treatment, and are referred to as treatment replicates.

Neutral detergent fiber (NDF-fraction), referred in this study as cell-walls, was prepared according to Goering and Van Soest (1970) and served for sugar analyses. The in vitro digestion procedure of Tilley and Terry (1963) was used to measure the digestibility of monosaccharides. The pattern of digestion was followed by running the incubation with rumen liquor for 6 , 12 , 24 , and 48 h and subsequently with acid pepsin for an additional 48 h. Monosaccharides 18% of the uronic acids in that material were found in cell were determined in whole material, NDF and in the in vitro residues, were determined in whole material, NDF and in the in vitro residues, solubles, pointing to the low content of pectic substances after acid hydrolysis, as their alditol acetates. The conditions for solubles in a partial sol hydrolysis and preparation of derivatives were those of Sloneker (1972), and for separation by GLC-those of Bacon and Gordon tion of cell wall sugars with particular effect on the matrix

(1980). Uronic acids in hydrolysates were determined colorimetrically as described by Blumenkrantz and Asboe-Hansen (1973). Total phenolics were estimated according to Morrison (1972).

Results

The composition of monosaccharides in wheat straw and their distribution in cell walls and in cell solubles is shown in Table 1. The major sugars, glucose and xylose, comprising about 90% of the total monosaccharides in the untreated WS, were mainly confined to the cell walls. Only

Table 3. The composition of the cell walls (NDF fraction) after treatment of wheat straw with sulfur dioxide, ozone and sodium hydroxide (g $100 g^{-1}$ sugars)

Monosaccharides Untreated		Treatment			
		SO ₂	Ozone	5% NaOH	
Glucose	60.2	75.7	68.8	62.8	
Xylose	29.9	19.9	24.2	28.3	
Arabinose	3.26	0.41	1.30	3.31	
Galactose	0.81	0.12	0.45	0.48	
Mannose	0.41	0.28	0.39	0.31	
Rhamnose	0.16	0.	0.06	0.06	
Uronic acids	5.28	3.59	4.80	4.74	
Total	100	100	100	100	

components. SO_2 exerted the greatest effect, followed by ozone and NaOH. The phenomena are summarized and shown in Table 2. The solubilized cell wall material was partly recovered in the soluble sugar fractions. It is assumed that phenolic substances and labile lignins account for the remainder. The content of permanganate-lignin suggested to represent the core and functional lignin was reduced, mainly by the ozone but to a lesser extent also by the $SO₂$ treatment.

Table 3 presents the monosaccharide profiles of the cell walls in treated and untreated WS. The solubilization of xylans has caused a reduction in cell wall xylose and increased the proportion of glucose particularly in the $SO₂$ and O_3 treated WS.

The in vitro digestiblity of monosaccharides in whole WS is shown in Table 4. The digestibilities of both major and minor sugars in the untreated WS, were initially high: 50 and 58% for xylose and glucose, respectively and 63% to 80% for the minor sugars. The $SO₂$ treatment resulted in an overall increase in digestibility of monosaccharides with values lying in the range of 90%. It is assumed that these results are close to the maximal response which is potentially attainable in chemically-treated natural lignocellulosics. Sodium hydroxide was more efficient than ozone in enhancing the in vitro digestibility of glucose, xylose and total sugars. The fact that this difference was not expressed in the organic matter digestibility of the NaOH and O_3 treated WS, implies that soluble products of lignin degradation contributed notably to the digestible fraction of the ozonated WS.

Table 5 presents the degradability of cell wall monosaccharides. Generally, the values were lower for the cell-wall than for the whole plant material. The effect of $SO₂$ was not confined to the solubilization of cell wall sugars (Ta-

Table 4. In vitro digestibility^a (%) of monosaccharides of whole wheat straw after treatment with sulfur dioxide, ozone and sodium hydroxide

Monosaccharides	Untreated	SO ₂	Ozone	5% NaOH	
Glucose	58.1 ± 0.80	88.2 ± 0.96	70.4 ± 0.55	75.1 ± 0.45	
Xylose	49.8 ± 1.63	90.8 ± 0.68	58.3 ± 0.86	74.0 ± 0.70	
Arabinose	71.9 ± 1.01	96.4 ± 0.38	85.8 ± 0.31	83.2 ± 0.24	
Galactose	77.4 ± 0.48	90.0 ± 1.62	74.0 ± 1.65	77.6 ± 0.92	
Mannose	79.7 ± 2.40	87.2 ± 1.58	73.1 ± 3.28	64.4 ± 2.33	
Rhamnose	64.5 ± 7.0	84.4 ± 3.17	73.4 ± 4.25	52.7 ± 3.36	
Uronic acids	63.4 ± 1.32	90.5 ± 0.12	78.5 ± 1.06	69.8 ± 1.02	
Total sugars	56.9 ± 1.03	89.5 ± 0.78	68.5 ± 0.21	74.6 ± 0.49	
I VOMD ^b	46.2 ± 0.83	81.7 ± 0.36	68.4 ± 0.70	67.5 ± 0.58	

 $\frac{a}{b}$ Means with their SE

In vitro organic matter digestibility

Table 5. In vitro digestibility^a (%) of cell wall (NDF) monosaccharides of wheat straw after treatment with sulfur dioxide, ozone and sodium hydroxide

Monosaccharides	Untreated	SO ₂	Ozone	5% NaOH
Glucose	55.8 ± 2.08	86.5 ± 1.09	70.1 ± 0.33	74.1 ± 0.47
Xylose	46.5 ± 1.72	80.1 ± 1.49	45.6 ± 1.50	70.5 ± 0.87
Arabinose	61.4 ± 1.39	86.3 ± 2.77	56.0 ± 1.62	78.3 ± 0.41
Uronic acids	55.4 ± 1.57	77.2 ± 0.36	61.0 ± 1.97	59.7 ± 2.92
Total sugars ^b	52.4 ± 1.09	84.4 ± 1.15	63.4 ± 0.24	72.3 ± 0.57

 $\frac{a}{b}$ Means with their SE

Including the minor sugars

XYLOSE HARIDE IONOSA با
ح \tilde{c} GEST o F > Z 100 80 60 40 20 0 GLUCOSE **I** I I I I I I I I I I ARABINOSE 100 80 60 40 20 0 0 612 24 TOTAL SUGARS **II 1 I** 1 URONIC ACIDS **I** <u>I</u> I <u>I I I I I I</u> **1,8 0 12 24 /,8** ORGANIC MATTER II **I I** 0612 24 48 INCUBATION TIME *(h)*

Fig. 1. The pattern of in vitro digestion of monosaccharides in wheat straw treated with: \blacktriangle , sulfur dioxide; \triangle , ozone; and \triangle , sodium hydroxide; o, represents untreated material

ble 1) but was expressed also in an increase in the degradability of the residual glucose, xylose and total sugars by 55, 72, and 61%, respectively. The digestibility of total cell wall sugars was increased from 52.4% to 84.4, 63.4 and 72.3% by the SO_2 , O_3 and NaOH treatments, respectively.

Figures 1 and 2 show the in vitro degradation pattern of monosaccharides and organic matter in whole plant material and in cell walls. Due to the solubilization of cell wall components by SO_2 and ozone, the behaviour of the digestion curves in the cell walls and whole plant material is different. In the cell walls of the $SO₂$ and $O₃$ treated WS, there was a lag time of between 6 to 12 h before the degradation of glucose, xylose and arabinose had started. However, since those cell wall sugars in the $SO₂$ treated material were potentially highly digestible, they reached the highest values of digestibility at 48 h of incubation.

Discussion

Hydrolysis and/or oxidation are the main routes by which biological systems decompose natural lignocellulosics. Most of the readily hydrolysable bonds are found within or associated to the matrix polysaccharides. Including: the labile glycosidic bonds between xylan and arabinofuranosyl residues (Bailey 1973), acetyl-ester bonds (Bacon and Gordon

Fig. 2. The pattern of in vitro digestion of cell wall monosaccharides of wheat straw treated with: \triangle , sulfur dioxide; \Box , ozone; and Δ , sodium hydroxide; \circ , represents untreated material

1980), phenolic esters (Higuchi et al. 1967; Hartley 1973), and lignin-hemicellulose complexes (Morrison 1974; Vered et al. 1981). Notwithstanding, it is widely accepted that most of the chemical bonds linking together the phenolic units in the lignin macromolecule are non-hydrolysable or fairly resistent to mild hydrolysis (Sarkanen 1981). Lignin, on the other hand, is sensitive to oxidation and is being degraded in nature by oxidative processes (Crawford and Crawford 1980; Eggeling and Sahm 1981). It is suggested therefore, that the quantitative ratio of hemicellulose to core lignin in the matrix of a given lignocellulosic, determines whether the substrate is more sensitive to hydrolysis or oxidative treatments. Core lignin is defined in this study, as the non hydrolysable cell wall lignin determined according to the permanganate method (Goering and Van Soest 1970). In Gramineae straws, the ratio of matrix polysaccharides to core lignin, is in the range of 3 to 4 (see Table 2 and Theander and Aman 1980) whereas, in dicotiledonous sources of lignocellulosics (as for example in cotton stalks or grape branches) the ratio is about 1 (Ben-Ghedalia, unpublished). This means that in terms of quantity and space, three quarters of the matrix in the Gramineae straws is composed of and occupied by polysaccharides, mainly branched xylans, representing in terms of sensitivity to hydrolysis, all the above mentioned. This is probably the reason why mild hydrolysing pretreatments are effective when applied

to Gramineae straws (this study and numerous others), while proving completely inefficient when applied to dicotiledonous lignocellulosics (Miron and Ben-Ghedalia 1981; Sneddon et al. 1981).

In this study, SO_2 was the most effective treatment in terms of cell wail dissolution (Table 2) which consequently increased the in vitro digestibility of WS monosaccharides to about 90% (Table 4). It is suggested that the main effect of $SO₂$ was mediated via the hydrolysis and dissolution of matrix polysaccharides (Table 1); this effect being complemented by solubillzation of cell wall phenolics (Table 2). $SO₂$ is able to hydrolyse labile ether bonds within the lignin molecule (Sarkanen 1981) and to sulfonate the "new born" hydroxyls as well as others attached to the aliphatic "tail" of the phenolic units (Forss et al. 1966). And since the lignin molecules of WS are much smaller than those found in woody materials (Ben-Ghedalia, unpublished), they became easily soluble (as sulfonated lignins) under the mild conditions of this study. Thus, it is assumed that the effect of $SO₂$ on the lignin of WS was a combined action of hydrolysis and sulfonation.

The primary effect of O_3 was expressed in partial oxidation of the phenolic fraction (Table 2) and the consequent release of the lignin associated carbohydrates. Secondary hydrolysis of matrix polysaccharides by the organic acids produced during ozonation (Ben-Ghedalia et al. 1982) cannot be ruled out. However, the comparison of in vitro digestibilities of sugars in the O_3 and NaOH treated straw (Tables 4 and 5, Figs. 1 and 2) points to the somewhat better effect of the NaOH treatment, particularly with respect to xylan degradability. This might lead to the conclusion that hydrolytic and swelling effects, as probably exerted by NaOH (Chesson 1981; Bacon et al. 1981) are of major importance when referred to pretreating WS, exceeding even the partial dissolution of cell wall material. $SO₂$ at 70 °C exerts a multi-effect which is particularly suitable for tackling the chemical and structural problemacy of the WS cell wall, in order to enhance its degradability by biological systems.

Acknowledgements. This research was supported by a grant from the United States - Israel (Binational) Agricultural Research $\&$ Development Fund (BARD).

References

- Bacon JSD, Chesson A, Gordon AH (1981) Deacetylation and enhancement of digestibility. Agric Environ 6:115-126
- Bacon JSD, Gordon AH (1980) The effects of various deacetylation procedures on the nylon bag digestibility of barley straw and grass cell walls recovered from sheep faeces. J Agric Sci Camb 94:361-367
- Bailey RW (1973) Structural carbohydrates. In: Butler GW, Bailey W (eds) Chemistry and Biochemistry of Herbage. Academic Press, New York, pp 157-209
- Ben-Ghedalia D, Miron J (1981) Effect of sodium hydroxide, ozone and sulphur dioxide on the composition and in vitro digestibility of wheat straw. J Sci Food Agric 32: 224-228

Ben-Ghedalia D, Shefet G, Miron J (1980) Effect of ozone and am-

monium hydroxide treatments on the composition and in vitro digestibility of cotton straw. J Sci Food Agric 31:1337-1342

- Ben-Ghedalia D, Shefet G, Miron J, Dror Y (1982) Effect of ozone and sodium hydroxide treatments on some chemical characteristics of cotton straw. J Sci Food Agric (in press)
- Binder A, Pelloni L, Fiechter A (1980) Delignification of straw with ozone to enhance biodegradability. Eur J Appl Microbiol Biotechnol 11:1-5
- Blumenkrantz N, Asboe-Hansen G (1973) New method for quantitative determination of uronic acids. Anal Biochem 54: 484- 489
- Chesson A (1981) Effects of sodium hydroxide on cereal straws in relation to the enhanced degradation of structural polysaccharides by rumen microorganisms. J Sci Food Agric 32:745-758
- Crawford D, Crawford R (1980) Microbial degradation of lignin. Enzyme Microbiol Technol 2:11-22
- Eggeling L, Sahm H (1981) Degradation of Lignin-related aromatic compounds by nocardia spec DSM 1069 and specificity of demethylation. In: Schaal, Pulverer (eds) Actinomycetes, Suppl 11. Gustav Fischer, Stuttgart New York, p 361-365
- Forss K, Fremer K, Stenlund B (1966) Spruce lignin and its reactions in sulfite cooking I. The structure of lignin II. The reactions in sulfite cooking. Paper and Timber 48:I, 565-574; II, 669-671
- Goering HK, Van Soest PJ (1970) Forage fiber analysis. USDA Agricultural Handbook No. 379
- Hartley RD (1973) Carbohydrate esters of ferulic acid as components of ceil-walls of Lolium multiflorum. Phytochemistry 12:661-665
- Higuchi T, Ito Y, Shimada M, Kawamura I (1967) Chemical properties of milled wood lignin of grasses. Phytochemistry 6: 1551-1556
- MiLlet MA, Baker AJ, Satter CD (1976) Physical and chemical pretreatment for enhancing cellulose saccharification. Biotechnol & Bioeng Symp 6:125-153
- Morrison IM (1972) A semi-micro method for determination of lignin and its use in predicting the digestibility of forage crops. J Sci Food Agric 23:455-463
- Morrsion IM (1974) Structural investigations on the lignin-carbohydrate complexes of Lolium perenne. Biochem J 139:197- 204
- Miron J, Ben-Ghedalia D (1981) Effect of chemical treatments on the degradability of cotton straw by rumen microorganisms and by fungal cellulase. Biotechnol Bioeng 23:1427-1437
- Sarkanen KV (1981) Principles and practical approaches to chemical and hydrothermal delignification. In: Domsch KH, Ferranti MP, Theander O (eds) Improved Utilization of Lignocellulosic Materials for Animal Feed. OECD/COST workshop. Braunschweig, pp 19-35
- Sloneker JH (1972) Gas liquid chromatography of alditol acetates. In: Whistler RL, BeMiller JN (eds) Methods in Carbohydrate Chemistry. Academic Press, New York London, p 20-24
- Sneddon DN, Thomas VM, Roffler RE, Murray G (1981) Laboratory investigations of hydroxide-treated sunflower or alfalfa-grass silage. J Anim Sci 53:1623-1628
- Theander O, Aman P (1980) Chemical composition of some forages and various residues from feeding value determinations. I Sci Food Agric 31:31-37
- Tilley JM, Terry RA (1963) A two-stage technique for the in vitro digestion of forage crops. J Brit Grassland Soc 18:104-111
- Vered Y, Milstein O, Flowers HM, Gressel J (1981) Biodegradation of wheat straw lignocarbohydrate complexes (LCC) i. Dynamics of liberation of hot aqueous LCC's from wheat straw and partial characterization of the products. Eur J Appl Microbiol Biotechnol 12:183-188