

Recovery from Short Term Intense Exercise: Its Relation to Capillary Supply and Blood Lactate Concentration

Per A. Tesch¹ and James E. Wright

Department of Environmental Medicine, Karolinska Institutet, Box 60 400, S-104 01 Stockholm, Sweden
US Army Research Institute of Environmental Medicine, Natick, MA 01760, USA

Summary. Muscle force recovery from short term intense exercise was examined in 16 physically active men. They performed 50 consecutive maximal voluntary knee extensions. Following a 40-s rest period five additional maximal contractions were executed. The decrease in torque during the 50 contractions and the peak torque during the five contractions relative to initial torque were used as indices for fatigue and recovery, respectively. Venous blood samples were collected repeatedly up to 8 min post exercise for subsequent lactate analyses. Muscle biopsies were obtained from m. vastus lateralis and analysed for fiber type composition, fiber area, and capillary density.

Peak torque decreased 67 (range 47–82%) as a result of the repeated contractions. Following recovery, peak torque averaged 70 (47–86%) of the initial value.

Lactate concentration after the 50 contractions was $2.9 \pm 1.3 \text{ mmol} \cdot \text{l}^{-1}$ and the peak post exercise value averaged $8.7 \pm 2.1 \text{ mmol} \cdot \text{l}^{-1}$. Fatigue and recovery respectively were correlated with capillary density ($r = -0.71$ and 0.69) but not with fiber type distribution. A relationship was demonstrated between capillary density and post exercise/peak post exercise blood lactate concentration ($r = 0.64$). Based on the present findings it is suggested that lactate elimination from the exercising muscle is partly dependent upon the capillary supply and subsequently influences the rate of muscle force recovery.

Key words: Fatigue – Mean fiber area – Muscle fiber types – Capillary density – Peak torque

Introduction

Heavy muscular exercise may result in impairment of the potential for further exercise. The reduction in performance capacity, caused by previous exercise, depends upon the mode, intensity and duration of exercise as well as the length of recovery phase. Accumulation of lactate and/or associated shifts in pH have been regarded as the most likely factors responsible for this impairment (cf. Simonson 1971; Karlsson 1979; Tesch 1980). It has been speculated that lactate and an associated increase in H^+ interfere with the formation of cross-bridges in the muscle cell by competing with the Ca^{2+} on the binding sites of troponin (Nakamura and Schwartz 1972; Robertson and Kerrick 1979). Both animal studies (Beatty et al. 1963; Baldwin and Tipton 1972) and recent findings in humans (Tesch et al. 1978; Tesch 1980) have suggested lactate to accumulate at a higher rate in fast twitch (FT or Type II) than in slow twitch (ST or Type I) fibers of exercised muscles. Lactate formed and accumulated in FT fibers may accordingly result in a greater inhibitory effect on the contractile machinery and a greater decrement in muscle force if the muscle is made up by a high percentage of FT fibers than can be expected in a “slow twitch” muscle. Moreover, the over-all metabolic profile of the ST fiber speaks in favor of a higher potential to recover from previous exercise, since one can assume that a ST muscle in comparison to a FT muscle is more efficient in oxidizing lactate due to differences in LDH enzyme and isozyme patterns (Sjödín 1976). Furthermore, a higher rate of lactate release to the blood stream may be expected from ST fibers due to a more developed capillary network (Folkow and Halicka 1968; Andersen and Henriksson 1977).

The following study was conducted to examine the recovery and lactate elimination pattern with regard to the capillary supply of exercising muscle following a short term, intense local performance task

Offprint requests to: P. A. Tesch at the above address

1 Dr. Tesch was on leave from Department of Clinical Physiology, Karolinska Hospital, Stockholm, Sweden

(Thorstensson and Karlsson 1976) which has been shown to induce reduction in maximal voluntary strength.

Subjects, Methods and Materials

Sixteen male subjects volunteered for the study. All were informed of the purpose and the risks associated with the experiments, prior to giving their written consent. The subject group represented a variety in terms of physical activity background including endurance running and advanced strength training programs as well as more general training. Age, height, weight, and maximal oxygen uptake attained on cycle ergometer averaged (\pm SD) 27 ± 4 years, 176 ± 5 cm, 78.5 ± 9.8 kg, and 49 ± 8 ml \cdot kg⁻¹ \cdot min⁻¹.

The first visit to the laboratory was intended to familiarize subjects with the isokinetic device (Cybex II, Lumex Inc., NY) and the protocol to be used in the experiments. Peak torque, produced at angular velocities corresponding to 30, 180, and 300° \cdot s⁻¹, was recorded during knee extension. After a short rest period a fatigue test consisting of 50 consecutive, maximal voluntary contractions performed at 180° \cdot s⁻¹, was conducted. For further information concerning the experimental set-up, see Thorstensson and Karlsson (1976) and Tesch (1980).

Two days later the actual experiments were carried out. The fatigue test was repeated and following a 40-s rest period five additional consecutive, maximal voluntary contractions were executed. The relative decrease in torque during 50 contractions was used as an index for fatigue. Day to day variation for the fatigue test amounted to 2.6% (coefficient of variation). Peak torque, developed during the following five contractions, relative to initial torque was calculated and expressed as an index for recovery (Fig. 1).

Blood samples for spectrophotometric determination of lactate concentration (Sigma Techn. Bull. 826, 1968) were obtained through an indwelling catheter from an antecubital vein prior to exercise, during the short recovery period, immediately after the additional five contractions, and at each minute up to 8 min post exercise. The concentration immediately post exercise relative to peak post exercise lactate concentration was calculated (Fig. 1).

Muscle biopsies (Bergström 1962) were obtained from m. vastus lateralis of the investigated limb at rest for subsequent histochemical analysis. Tissue specimen were frozen in liquid nitrogen-cooled isopentane and stainings were performed on cross-sections for myofibrillar ATPase (Padykula and Herman 1955), NADH-tetrazolium reductase (Novikoff et al. 1961) and Amylase-PAS (Andersen and Henriksson 1977). Muscle fiber type distribution (%FT, %FT area) and mean muscle fiber area were calculated according to Tesch (1980) while capillary density expressed as cap \cdot mm⁻² and cap \cdot fib⁻¹ were assessed as described elsewhere (Andersen and Henriksson 1977).

Results

Values for peak torque per kilogram body weight at 30, 180, and 300° \cdot s⁻¹, initial peak torque, peak torque decrease during the fatigue test, increase in peak torque during recovery are presented in Table 1 along with information on fiber type distribution, fiber area, and capillary density. The correlation coefficients between peak torque at different angular

Table 1. Mean (\pm SD and range) values for physiological and histochemical variables examined

	Mean (\pm SD)	Range
Physiological variables		
Peak torque at		
30° \cdot s ⁻¹ , Nm \cdot kg ⁻¹ b.w	3.7 (\pm 0.8)	1.7 – 5.1
180° \cdot s ⁻¹ , Nm \cdot kg ⁻¹ b.w	2.3 (\pm 0.4)	1.3 – 3.1
300° \cdot s ⁻¹ , Nm \cdot kg ⁻¹ b.w	1.6 (\pm 0.3)	1.3 – 2.1
Fatigue, % peak torque decrease	67 (\pm 11)	47 – 82
Recovery, % peak torque of initial	70 (\pm 13)	47 – 86
Histochemical variables		
Fiber type distribution		
% FT	46 (\pm 12)	19 – 60
% FT area	51 (\pm 15)	18 – 66
Mean fiber area, μ m ² \cdot 100	66 (\pm 18)	39 – 108
Capillary density		
cap \cdot mm ⁻²	274 (\pm 85)	156 – 489
cap \cdot fib ⁻¹	1.65 (\pm 0.30)	1.12– 2.24

Table 2. Correlation coefficients between knee extension peak torque at different angular velocities, expressed in absolute and relative (to body weight) terms, and mean fiber area and fiber type distribution

	Mean fiber area	% FT area
Peak torque		
30° \cdot s ⁻¹ , Nm	0.47 NS	0.48 NS
30° \cdot s ⁻¹ , Nm \cdot kg ⁻¹	0.46 NS	0.57*
180° \cdot s ⁻¹ , Nm	0.54*	0.61**
180° \cdot s ⁻¹ , Nm \cdot kg ⁻¹	0.55*	0.82***
300° \cdot s ⁻¹ , Nm	0.09 NS	0.57*
300° \cdot s ⁻¹ , Nm \cdot kg ⁻¹	0.41 NS	0.77***

Level of significance is denoted: * $P < 0.05$, ** $P < 0.01$, *** $P < 0.001$, NS = $P > 0.05$

velocities and mean fiber area and fiber type distribution are depicted in Table 2.

Mean initial peak torque at 180° \cdot s⁻¹ was 178 (102–245) Nm. Peak torque decreased on average of 120 (range 61–201) Nm or 67 (47–82%) following the 50 consecutive contractions. Peak torque following recovery was 70 (47–86%) of the initial value.

Blood lactate concentration at rest, following the 50 contractions and immediately following the final contractions averaged (\pm SD) 1.5 ± 0.3 , 2.9 ± 1.3 , and 5.3 ± 2.3 mmol \cdot l⁻¹. The peak value following exercise was 8.7 ± 2.1 mmol \cdot l⁻¹ (Fig. 1). Blood lactate concentration immediately following the 5th contraction relative to peak lactate concentration after exercise was $62 \pm 21\%$ (Fig. 2).

Peak torque decrease (fatigue) was negatively correlated with recovery ($r = -0.64$, $P < 0.01$). Neither fatigue nor recovery were significantly cor-

EXPERIMENTAL DESIGN

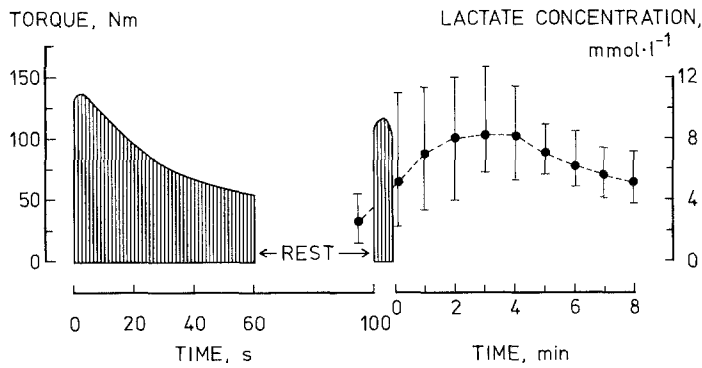


Fig. 1. Schematic description of the experimental design showing force production during exercise and following a 40-s rest period. Values for lactate concentration are mean and range

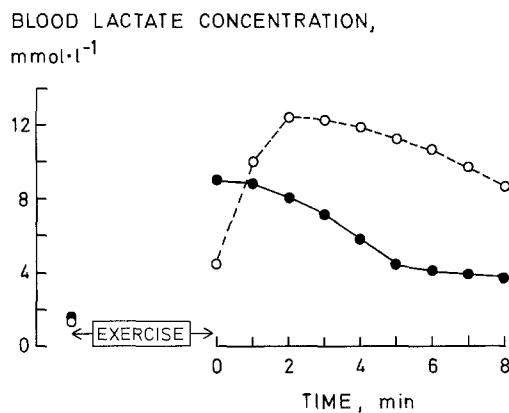


Fig. 2. Examples of post exercise lactate response in two subjects. The lactate concentration immediately post the fifth contraction relative to peak lactate concentration during recovery was calculated. From the picture it is evident that blood lactate response is different in the two individuals exemplified. The subject represented by filled dots (●) had 51% FT area and 364 cap · mm⁻² and the subject represented by open dots (○) had 66% FT area and 156 cap · mm⁻²

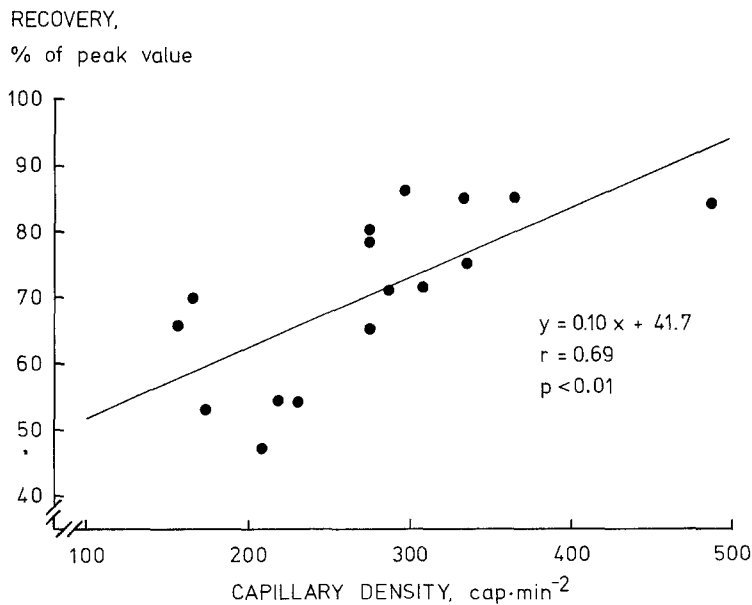


Fig. 3. The relationship of recovery to capillary density

related with fiber type distribution. Fatigue and recovery respectively were, however, correlated with cap · mm⁻² ($r = -0.71$, $P < 0.01$ and $r = 0.69$, $P < 0.01$, Fig. 3). Positive relationships were present between cap · mm⁻² and blood lactate concentration post exercise/peak post exercise ($r = 0.64$, $P < 0.01$,

Fig. 4) and blood lactate concentration and recovery ($r = 0.55$, $P < 0.05$).

Multiple regression analyses revealed that 95% ($R = 0.98$) of the variance in recovery from fatigue could be explained by fiber type distribution, capillary density, fatigue, and peak torque.

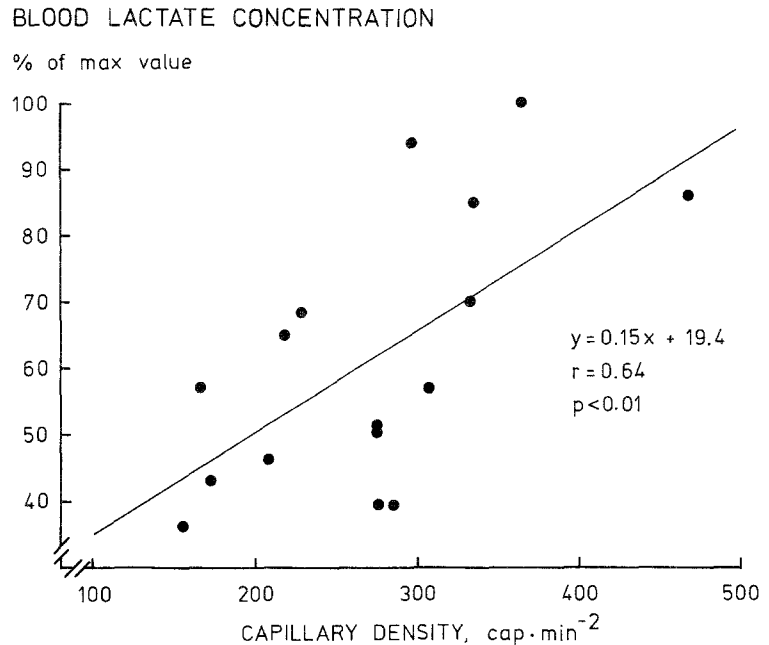


Fig. 4. The relationship of post exercise/peak post exercise blood lactate concentration to capillary density

Discussion

In contrast to previous studies (Thorstensson and Karlsson 1976; Nilsson et al. 1977; Komi and Tesch 1979; Tesch 1980) using the same experimental set-up no significant relation was present between force decline and the relative distribution of FT fibers. Hence, it is obvious that the fiber type – fatigue relationship can be altered.

Animal studies (Kugelberg and Edström 1968; Kugelberg and Lindgren 1979) have demonstrated that fatigue properties of single motor units were related to the oxidative capacity of the innervated muscle fibers. Recently, Ivy et al. (1982), using a protocol similar to our own, were able to demonstrate a negative relationship ($r = -0.63$) between the respiratory capacity of the exercising muscle and rate of fatigue development whereas lower correlations were established between fiber type composition and muscle fatigue. It was speculated that muscle fatigability could be altered as a result of changes in the muscle respiratory capacity, induced by physical training, without affecting fiber type composition. The finding here of a relationship between fatigue index and capillary density could be interpreted in the same way. That is, the capillary supply reflects the over-all metabolic profile of the exercising muscle and parallels both mitochondrial content (Brodal et al. 1977) and the activity of enzymes regulating oxidation (Hudlická 1982).

In the present study voluntary muscle strength was found to decrease by an average of 67% and after a brief recovery period contractile function was still depressed by 30%. To a certain extent recovery in

muscle contractility was influenced by the magnitude of the preceding reduction in muscle strength. Individuals who were most susceptible to fatigue also exhibited the smallest recovery in muscle force.

Using an identical fatigue protocol, pronounced muscle lactate accumulation occurred and concentrations remained the same or decreased only slightly 30 s post exercise (Tesch 1980). In that study, however, a rapid restitution of muscle force took place. Also, blood lactate concentration after exercise was lower in subjects with muscles which decreased most in muscle force, rich in FT fibers, and producing most lactate. It was speculated that individual variations in capillary supply of the exercising muscle had impact on elimination of lactate from the muscle. The fact that recovery from short term intense exercise was shown to be correlated with the density of the capillary bed is thus supportive.

The relationship between capillary supply and recovery might be explained in that capillary supply simply is paralleling the oxidative capacity of the muscle. Support for this assumption was found in that the magnitude of recovery, using the present experimental procedures, was correlated with citrate synthase activity, a marker for oxidative capacity (Tesch et al. 1983). Thus the possible role of oxidative metabolism in recovery processes (Nassar-Gentina et al. 1978) was implied and since fatigue index was not negatively related to the activity of this enzyme a causal relationship does not seem implausible. It is also possible that capillary density is a true reflection of the open capillary surface area and thus the blood transport system. Jorfeldt et al. (1978)

found the rate of lactate release from exercising muscle to blood to be maximal at muscle lactate concentrations of approximately $5 \text{ mmol} \cdot \text{kg}^{-1} \text{ w.w}$ in spite of a further increased blood flow. Thus membrane transport or uptake and elimination by the vascular system seemed to be impeded above that lactate level. The authors did not indicate the magnitude of the individual range in this response. It is therefore impossible to speculate whether the levelling off observed was influenced by peripheral factors such as capillary supply.

From the present findings we cannot elucidate which factor exerted the greatest influence on recovery. However, it can be concluded that muscle force recovers at a faster rate than lactate is eliminated.

Dynamic muscle strength correlated with the relative area occupied by FT fibers rather than with muscle fiber size irrespective of contraction speed (angular velocity). Mean fiber area of a muscle has been proposed to be a reliable measure of its total cross-sectional area (Häggmark et al. 1978); probably also mirroring the volume of the examined muscle. Our data can therefore be interpreted to imply that fiber type composition and selective FT fiber hypertrophy are both important factors for muscle force production at slow velocities as well as during high speed contractions (Coyle et al. 1979; Gregor et al. 1979; Tihanyi et al. 1982); this permits a refinement of the general view that muscle mass per se is the predominant factor influencing force production (Ikai and Fukunaga 1968). This pattern was recently confirmed in a sample of more than 100 physically active subjects (Tesch, unpublished observations).

In summary, both fatigue and recovery of knee extensor muscles were found to correlate with the capillary density of that muscle. Since lactate release to the blood stream was correlated with capillary supply, it is suggested that recovery processes are influenced by the rate of lactate disappearance from the muscle.

Acknowledgements. The views and opinions contained in this report are those of the authors and are not to be construed as official or as reflecting the views of the Department of the US Army. This study was partly supported by grants from the Swedish Medical Research Council Project No. 5020 and the Research Council of the Swedish Sports Federation.

References

- Andersen P, Henriksson J (1977) Capillary supply of the quadriceps femoris muscle of man: adaptive response to exercise. *J Physiol* 270: 677–690
- Baldwin KM, Tipton CM (1972) Work and metabolic patterns of fast and slow twitch skeletal muscle contracting in situ. *Pflügers Arch* 334: 345–356
- Beatty CH, Peterson RD, Bocek RM (1963) Metabolism of red and white muscle fiber groups. *Am J Physiol* 204: 939–942
- Bergström J (1962) Muscle electrolytes in man. *Scand J Clin Lab Invest [Suppl]* 68
- Brodal P, Ingjer F, Hermansen L (1977) Capillary supply of skeletal muscle fibers in untrained and endurance-trained men. *Am J Physiol* 232: H705–H712
- Coyle EF, Costill DL, Lesmes GR (1979) Leg extension power and muscle fiber composition. *Med Sci Sports* 11: 12–15
- Folkow B, Halicka HD (1968) A comparison between “red” and “white” muscle with respect to blood supply, capillary surface area and oxygen uptake during rest and exercise. *Microvasc Res* 1: 1–14
- Gregor RJ, Edgerton VR, Perrine JJ, Campion DS, Debus C (1979) Torque-velocity relationships and muscle fiber composition in elite female athletes. *J Appl Physiol* 47: 388–392
- Häggmark T, Jansson E, Svane B (1978) Cross-sectional area of the thigh muscle in man measured by computed tomography. *Scand J Clin Lab Invest* 38: 355–360
- Hudlická O (1982) Growth of capillaries in skeletal and cardiac muscle. *Circ Res* 50: 451–461
- Ikai M, Fukunaga T (1968) Calculation of muscle strength per unit cross-sectional area of human muscle by means of ultrasonic measurement. *Int Z angew Physiol Einschl Arbeitsphysiol* 26: 26–32
- Ivy JL, Sherman WM, Miller JM, Maxwell BD, Costill DL (1982) Relationship between muscle $\dot{Q}O_2$ and fatigue during repeated isokinetic contractions. *J Appl Physiol: Respirat Environ Exercise Physiol* 53: 470–474
- Jorfeldt L, Juhlin-Dannfelt A, Karlsson J (1978) Lactate release in relation to tissue lactate in human skeletal muscle during exercise. *J Appl Physiol: Respirat Environ Exercise Physiol* 44: 350–352
- Karlsson J (1979) Localized muscular fatigue: Role of muscle metabolism and substrate depletion. In: Hutton RS, Miller DI (eds). *Exercise Sport Sci Rev* 7. Franklin Institute Press, Philadelphia, pp 1–42
- Komi PV, Tesch P (1979) EMG frequency spectrum, muscle structure, and fatigue during dynamic contractions in man. *Eur J Appl Physiol* 42: 41–50
- Kugelberg E, Edström L (1968) Differential histochemical effects of muscle contractions on phosphorylase and glycogen in various types of fibres: relation to fatigue. *J Neurol Neurosurg Psychiatry* 31: 415–423
- Kugelberg E, Lindegren B (1979) Transmission and contraction fatigue of rat motor units in relation to succinate dehydrogenase activity of motor unit fibres. *J Physiol* 288: 285–300
- Nakamura Y, Schwartz A (1972) The influence of hydrogen ion concentration on calcium binding and release by skeletal muscle sarcoplasmic reticulum. *J Gen Physiol* 59: 22–32
- Nassar-Gentina V, Passonneau JV, Vergara JL, Rapoport SI (1978) Metabolic correlates of fatigue and of recovery from fatigue in single frog muscle fibers. *J Gen Physiol* 72: 593–606
- Nilsson J, Tesch P, Thorstensson Å (1977) Fatigue and EMG of repeated fast voluntary contractions in man. *Acta Physiol Scand* 101: 194–198
- Novikoff AB, Shin WY, Drucker J (1961) Mitochondrial localization of oxidative enzymes: staining results with two tetrazolium salts. *J Biophys Biochem Cytol* 9: 47–61
- Padykula HA, Herman E (1955) The specificity of the histochemical method for adenosine triphosphatase. *J Histochem Cytochem* 3: 170–195
- Robertson SP, Kerrick WGL (1979) The effects of pH on Ca^{2+} -activated force in frog skeletal muscle fibers. *Pflügers Arch* 380: 41–45
- P. A. Tesch and J. E. Wright: Fatigue, Recovery and Lactate

- Sigma Technical Bulletin 826 (1968) Sigma Chemical Co, St Louis MO
- Simonson E (1971) Physiology of work capacity and fatigue. Charles C Thomas, Springfield Ill
- Sjödín B (1976) Lactate dehydrogenase in human skeletal muscle. *Acta Physiol Scand* [Suppl] 436
- Tesch P (1980) Muscle fatigue in man with special reference to lactate accumulation during short term intense exercise. *Acta Physiol Scand* [Suppl] 480: 1-90
- Tesch P, Sjödín B, Karlsson J (1978) Relationship between lactate accumulation, LDH activity, LDH isozyme and fibre type distribution in human skeletal muscle. *Acta Physiol Scand* 103: 40-46

- Tesch PA, Wright JE, Daniels WL, Sjödín B (1983) Physical performances and muscle metabolic characteristics. In: Knuttgen HG, Vogel JA, Poortmans J (eds). *Int. Series on Sport Sciences. 13. Biochemistry of Exercise. Human Kinetics Publishers. Champaign. IL.* pp 258-263
- Thorstensson A, Karlsson J (1976) Fatiguability and fibre composition of human skeletal muscle. *Acta Physiol Scand* 98: 318-322
- Tihanyi J, Apor P, Fekete GY (1982) Force-velocity-power characteristics and fiber composition in human knee extensor muscles. *Eur J Appl Physiol* 48: 331-343

Accepted May 29, 1983