## ORIGINAL PAPER

# **Anna Benedetti 9 Giovanni Sebastiani Determination of potentially mineralizable nitrogen in agricultural soil**

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**Abstract** Potentially mineralizable soil N was determined after incubation for 2, 4, 8, 12, 16, 22, and 30 weeks, according to the Stanford and Smith method. A first-order kinetics model was used, and a simulation study was performed using three different statistical methods to estimate potentially mineralizable N and the rate constant  $k$ . The first method was based on the maximum-likelihood approach. The second one relied on nonlinear least square data fitting. The third method was based on linear of logarithmically transformed data. The results of the simulation study suggested that the non-linear least square method was preferable to the others. This method was then applied to real data from 30 different Italian soils. The values obtained for potentially mineralizable N were, on average, 10% of total N (mean standard error = 0.9). The estimated value of k was 0.050 (mean standard error  $= 0.005$ ). Finally, from the values obtained for  $k$  and the results of the simulation, the results indicated that significantly less reliable estimates of potentially mineralizable N were obtained by using data for up to only 22 weeks of incubation.

**Key words** Statistical methods  $\cdot$  Mineralization  $\cdot$ Nitrogen  $\cdot$  First-order kinetics  $\cdot$  Incubation experiment  $\cdot$ Model for N mineralization

## **Introduction**

N mineralization makes a key contribution to the amount of available N in soil. However, no fast, accurate, and seri-

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al methods are available to determine mineralizable N, especially in field conditions (Skjemstad et al. 1988). The numerous references in the literature on this topic distinguish between chemical and biological determinations. The chemical methods are based on the extraction of mineral N using different extractants such as boiling water (Jenkinson 1968; Stanford 1969; Verstraeten et al. 1970), acid, alkaline, and salt solution (e.g.,  $0.01 M$ CaCl<sub>2</sub>), or by determination of the NH $<sub>4</sub><sup>+</sup>$  released during</sub> alkaline permanganate oxidation (Troug 1954; Stanford 1978; Richter et al. 1994). The biological methods make use of soil incubation under controlled conditions of moisture and temperature for a length of time ranging from one to many weeks (Delphin 1986; Catroux et al. 1987; Tabatabai and A1-Khafaji 1988). The results of these methods are difficult to interpret and show their limitations when transferred to open field conditions.

In 1972 decisive progress was made by Stanford and Smith with their method based on leaching, at predetermined times, the N mineralized at  $35^{\circ}$ C, to simulate plant uptake. They described N mineralization by a first-order kinetics model according to the following equation (1):

$$
N_t = N_0(1 - e^{-kt})
$$
\n<sup>(1)</sup>

where  $N_t$  is N mineralized in the time considered,  $N_0$  the potentially mineralizable N in the soil (i.e., the maximum quantity of N that the soil can mineralize), and  $k$  the rate constant expressed week<sup>-1</sup>. Stanford and Smith (1972) estimated  $k$  by using the maximum likelihood method and obtained the value  $0.054 \pm 0.009$  week<sup>-1</sup>. Subsequently, many authors have studied the adequacy of firstorder kinetics to describe the N mineralization rate (Juma et al. 1984; Dendooven et al. 1991) and this is considered to represent a correct description of the mineralization course (Bonde and Roswall 1987). Recent studies have shown that when Stanford and Smith's method is used in a controlled environment the values obtained are too high (Cabrera and Kissel 1988), due to many factors, including the use in the laboratory of disturbed soil samples (Stanford and Epstein 1974; Smith et al. 1977; Smith et al. 1980; Smith et al. 1982; Griffin and Laine 1983;

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Frankenberger and Abkelmagid 1985). In particular, Hadas et al. (1989) pointed out that the mineralization in the field is  $13-26\%$  lower than the calculated values.

The objectives of the present study were (1) to assess by a simulation approach, the goodness of three different statistical methods for estimating  $N_0$  and k in a first-order kinetics model for a special case of correlated errors; (2) to check the adequacy of a first-order kinetics model to describe N mineralization data from 30 Italian soils by applying the best method to the real data, and obtain a mean value for the estimate of the constant rate  $k$  together with an interval of variation; and (3) to study the possibility of achieving reductions in the maximum incubation time because the length of time necessary to carry out the determinations represents the main difficulty in the serial use of this technique (Menasseri et al. 1994).

### **Materials and methods**

The research was carried out using 30 soils sampled from the arable layer  $(0-40 \text{ cm})$  of experimental fields of the Institute for Plant Nutrition (ISNP) located in central and northern Italy. These soils are of alluvial, volcanic, and volcanic cyneritic origin and were sampled during the uncultivated period between two crops.

The chemical and physical properties of the soils (Table 1) were determined by Italian Society of Soil Science (1985) methods. Potentially mineralizable N data (Table 2) were measured following the method proposed by Stanford and Smith (1972), partially modified by Benedetti (1983). In this measurement 50 g of soil, air-dried and sieved through a 2-mm screen, was mixed with quartz sand in a 1:1 ratio (sand particle size, 0.2-0.8 mm) and incubated in a Buchner funnel (outer diameter 13 cm), at 60% water-holding capacity and at  $30^{\circ}$ C for 30 weeks. The mineral N in the soil was leached before incubation by adding  $900$  ml CaSO<sub>4</sub> solution and  $100$  ml nutrient solution without N  $(0.002 M CaSO<sub>4</sub> \cdot 2 H<sub>2</sub>O, 0.005 M Ca(H<sub>2</sub>PO<sub>4</sub>),$  $0.0025 M K_2SO_4$ ,  $0.002 M M gSO_4$ ). NO<sub>3</sub>-N and NH<sub>4</sub>+N concentrations were measured periodically (after 2, 4, 8, 12, 16, 22, and 30 weeks) using an Autoanalyzer Techniton II. At the end of the incubation period, exchangeable  $NH_4^+$ -N was extracted, using a 2 NKC1 solution following the Bremner procedure (1965). All analyses were carried out in triplicate.

Statistical analysis

With  $t_1 = 2$ ,  $t_2 = 4$ ,  $t_3 = 8$ ,  $t_4 = 12$ ,  $t_5 = 16$ ,  $t_6 = 22$ ,  $t_7 = 30$ representing the (cumulative) incubation times in weeks, the experimental (cumulative) value  $N_i$  for the N mineralized duringt time  $0-t_i$  was obtained as the sum of the *i* values  $y_1, \ldots, y_i$  for the N mineralized during the first  $i$  consecutive incubations, that is (2):

$$
N_i = \sum_{j=1}^{l} y_j
$$
 (2)

The variables  $y_j$ ,  $j = 1, ..., 7$  are affected by errors, and we assumed the errors  $e_j$  *j* = 1, ..., 7 to be uncorrelated and Gaussiandistributed, with mean zero and variance  $\sigma^2$ . Therefore (3):

$$
N_i = N_0 (l - e^{-\kappa t_i}) + \tilde{e}_i \tag{3}
$$

**Table 1** Some physical and chemical properties of soils tested (total N, organic C, and humus calculated on dry-weight basis; 105 °C). RM, Roma; FR, Frosinone

Sample	Location	Texture	pH (1:2.5)	Field capacity $(\%)$	Total N (Kjeldahl) $(\%)$	Organic C (Springer- Klee) $(\%)$	C: N	Humus (organic C $\times$ 1.724) (%)
$\mathbf 1$	Mantova	Silty loam	8.1	24.5	0.08	1.20	15	2.10
$\overline{c}$	Città Castello	Sandy silty clay	7.9	34.5	0.13	1.18	9	2.03
3	Mazzano (RM)	Sandy	7.5	18.0	0.18	2.44	14	4.21
4	Settecamini (RM)	Sandy loam	7.8	17.4	0.05	0.50	13	0.86
5	Celimontana (RM)	Sandy loam	7.2	24.0	0.15	1.91	13	3.29
6	Colleferro (FR)	Sandy silty clay	5.8	40.3	0.04	0.27	7	0.46
$\overline{7}$	Anagni (FR)	Clay loam	5.9	27.6	0.12	1.07	9	1.85
8	Paliano (FR)	Clay	5.8	28.5	0.08	0.09	8	1.35
11	Frosinone	Sandy	7.8	20.0	0.11	0.92	8	1.59
12	Frosinone	Silty loam	6.9	23.9	0.11	1.05	9	1.81
13	Frosinone	Sandy silty loam	7.8	22.2	0.14	1.26	9	2.17
14	Paliano (FR)	Loam	7.2	24.2	0.11	1.18	11	2.03
15	Anagni (FR)	Loam	6.8	20.0	0.07	0.73	10	1.26
16	Anagni (FR)	Loam	6.8	25.9	0.13	1.26	10	2.17
17	Ferentino (FR)	Loam	7.1	22.6	0.19	2.32	12	3.99
18	Paliano (FR)	Clav	6.2	30.9	0.12	1.20	10	2.06
19	Anagni (FR)	Clay	6.5	34.6	0.19	1.76	9	3.03
20	Anagni (FR)	Clay	7.0	41.5	0.09	1.00	11	1.72
21	Paliano (FR)	Clay	5.5	25.7	0.15	1.42	9	2.44
22	Anagni (FR)	Sandy loam	7.6	19.9	0.10	1.16	12	2.00
23	Anagni (FR)	Clay	6.6	33.3	0.10	0.94	9	1.62
24	Paliano (FR)	Clay	6.8	33.9	0.08	0.86	11	1.48
25	Anagni (FR)	Sandy loam	7.6	19.9	0.11	1.20	11	2.06
26	Anagni (FR)	Sandy loam	7.1	20.8	0.10	1.07	11	2.06
27	Anagni (FR)	Sandy loam	6.6	13.4	0.05	0.44	8	0.76
28	Ferentino (FR)	Sandy loam	7.6	17.6	0.11	1.04	9	1.79
29	Ferentino (FR)	Silty loam	6.2	45.9	0.28	3.06	11	5.26
30	Torice (FR)	Silty loam	7.9	22.0	0.11	1.12	10	1.93
31	Paliano (FR)	Silty clay loam	5.7	34.8	0.06	0.57	9	0.198
32	Ferentino (FR)	Loam	6.7	18.3	0.09	0.85	9	1.46

**Table 2** Potentially mineralizable N (mg kg<sup>-1</sup> dry weight); cumulative experimental values

	Weeks							
	0	$\overline{c}$	$\overline{4}$	8	12	16	22	30
1	12.5	7.0	14.5	22.8	26.0	53.0	63.3	70.8
$\overline{\mathbf{c}}$	66.0	19.0	26.0	35.0	35.0	35.0	35.0	35.0
$\overline{\mathbf{3}}$	80.0	41.0	59.0	75.0	95.0	115.0	133.0	155.0
4	8.0	7.4	16.4	20.6	22.8	28.7	35.0	41.0
5	80.0	80.0	135.0	204.0	256.0	304.0	344.0	386.0
6	25.0	0.0	11.0	20.0	20.0	20.0	20.0	20.0
$\overline{7}$	15.0	2.0	13.8	19.6	25.0	35.0	47.6	83.8
8	48.0	14.2	37.6	53.9	67.7	76.1	86.9	98.7
11	17.5	21.3	39.4	44.0	53.6	64.3	76.0	92.0
12	19.2	21.0	29.8	33.3	41.1	50.6	66.7	77.5
13	8.7	13.7	22.1	27.2	37.3	48.1	58.8	77.7
14	13.0	8.0	14.4	21.4	24.7	34.7	42.4	52.3
15	9.8	11.5	18.0	23.4	30.6	39.4	48.4	58.0
16	12.5	15.6	21.1	26.0	33.8	42.3	50.6	63.0
17	42.8	19.9	30.0	34.9	42.0	58.7	71.1	102.2
18	26.6	8.5	14.3	22.8	26.0	36.1	44.3	53.3
19	10.4	18.6	25.3	30.0	37.9	47.3	61.8	95.2
20	15.3	7.8	15.3	23.1	27.1	39.1	46.0	55.2
21	34.6	11.4	17.3	26.6	30.4	39.5	47.0	56.2
22	12.7	10.7	18.3	26.6	33.3	40.3	50.0	62.3
23	14.8	6.4	13.4	22.6	28.9	36.2	46.2	62.4
24	9.9	10.1	15.7	25.0	29.0	39.0	49.3	64.1
25	41.6	6.0	12.4	19.9	26.5	33.3	44.5	56.3
26	10.9	8.3	15.0	22.5	28.8	36.0	44.7	58.6
27	20.2	7.8	14.9	23.5	30.0	36.6	45.9	59.0
28	10.5	6.2	12.2	20.2	26.7	33.5	42.0	58.0
29	90.8	122.3	129.5	172.6	198.0	238.6	302.1	346.8
30	27.9	11.6	17.2	27.7	32.1	42.2	50.5	74.7
31	7.3	5.7	11.4	18.8	23.9	29.4	37.1	47.5
32	9.2	9.2	15.0	22.7	27.5	37.5	44.7	54.1

i where  $\tilde{e}_i = \sum e_j$ , k is the rate constant expressed per week, and  $t_i$ 

the incubation time in weeks. Then, the variables  $\tilde{e}_i$  i = 1, ..., 7 will also be Gaussian-distributed, with mean zero and covariance matrix  $C$  (Johnson and Wichern 1988) given by (4):

$$
C = \sigma^2 \begin{pmatrix} 1 & 1 & 1 & 1 & 1 & 1 & 1 \\ 1 & 2 & 2 & 2 & 2 & 2 & 2 \\ 1 & 2 & 3 & 3 & 3 & 3 & 3 \\ 1 & 2 & 3 & 4 & 4 & 4 & 4 \\ 1 & 2 & 3 & 4 & 5 & 5 & 5 \\ 1 & 2 & 3 & 4 & 5 & 6 & 6 \\ 1 & 2 & 3 & 4 & 5 & 6 & 7 \end{pmatrix}
$$
 (4)

The joint distribution  $f(N_1, \ldots, N_7)$  for the variables  $N_i$ ,  $i = 1, \ldots, 7$  will then be given by (5):

$$
f(N_1, \ldots, N_7) = \frac{1}{(2\pi)^{7/2} |C|^{1/2}} \exp\left[-\frac{1}{2} \sum_{i,j=1}^7 [N_i - N_0(1 - e^{-kt})] - C_{ij}^{-1} [N_j - N_0(1 - e^{-kt})]\right]
$$
(5)

where  $C_{ii}^{-1}$  is the element  $(i,j)$  of the inverse of the matrix C, and  $\parallel$  indicates the determinant of a matrix. In the maximum likelihood method,  $N_0$  and k are estimated by maximizing the likelihood function f with respect to  $N_0$  and k. This is equivalent to minimizing the function  $F(N_0, k)$  (6):

$$
F(N_0, k) = \sum_{i,j=1}^{7} [N_i - N_0(1 - e^{-kt_i})] C_{ij}^{-1} [N_j - N_0(1 - e^{-kt_j})]
$$
(6)

For a fixed k, the minimum of  $F(N_0, k)$  is found by imposing the  $\partial N_0$ 

derivative 
$$
\frac{\partial^2 V_0}{\partial F(N_0, k)} = 0
$$
. This equation is solved as (7):

$$
N_0 = \frac{\sum_{i,j=1}^{7} [N_i C_{ij}^{-1} (1 - e^{-kt_j}) + (1 - e^{-kt_i}) C_{ij}^{-1} N_j]}{2 \sum_{i,j=1}^{7} [(1 - e^{-kt_i}) C_{ij}^{-1} (1 - e^{-kt_j})]}
$$
(7)

In the last equation,  $N_0$  is a function of k alone, since both  $N_i$  and  $t_i$ ,  $i = 1, \ldots, 7$  are known; thus, by introducing Eq. 7 into Eq. 6, F can be minimized as a function of  $k$  alone. This is done by the iterative Newton method (Wolfe 1978), which was stopped when the percentual difference of the values for  $F$ ,  $N_0$ , and  $k$  corresponding to two consecutive iterations was less than  $10^{-4}$ .

The non-linear least square method (Smith et al. 1980) is based minimizing the sum of the square of the deviations between the experimental data  $N_i$  and the corresponding model prevision  $N_0(1-e^{-kt_i})$ , that is (8):

$$
\tilde{F}(N_0, k) = \sum_{i=1}^{7} [N_i - N_0(1 - e^{-kt_i})]^2
$$
\n(8)

Since the functions F and  $\tilde{F}$  have a similar structure, the same kind of procedure described for F was used to minimize  $\tilde{F}$ .

Finally, the log-transformed linear least square method, proposed by Stanford and Smith (1972), is based on the equation  $N_t = N_0(1-e^{-kt})$  expressed in logarithmic form (9):

$$
\log (N_0 - N_t) = \log (N_0) - kt/2.303\tag{9}
$$

By fixing a value of  $N_0$  in the left side of this equation, a linear regression between the variables  $log(N_0 - N_i)$  and t can be performed to estimate k. An "optimal" value for  $N_0$  is found by using an iterative procedure which involves repeated regressions based on different choices of  $N_0$ . The best value of  $N_0$  and k is the one that produces a regression with the greatest coefficient of determination  $r^2$ .

### **Results and discussion**

We present here the results from a simulation study used to assess the goodness of using different methods to estimate the parameters  $N_0$  and k of a first-order kinetics model. The results obtained using the best of the three methods on data from 30 soils are also shown.

In the simulation study, we used samples from the statistical distribution of Eq. 5. The value chosen for  $N_0$ was 100, while three different values for  $k$  were used,  $k = 0.03$ , 0.05, 0.07 week<sup>-1</sup>. The value used for  $\sigma^2$  was  $(4 \text{ ppm})^2$ . The three methods were applied to each sample replication by applying each of the three values of  $k$ ; thus estimates of  $N_0$  and k were obtained. The sample average and standard deviation of the estimated quantities were computed for each value of  $k$  and for each of the three methods (Table 3). The number of replications used was 20000, and no significant changes in the averages or standard deviations appeared with only 10000 replications or with different samples. Table 3 also shows

**Table 3** Estimates (means  $\pm$  SD) of  $N_0$  and k obtained by using the maximum likelihood *(ML),* non-linear least square *(NLLS),* and logtransformed linear least square *(LTLS)* methods on simulated data. The values of the root mean square *(RMS)* deviations between the data and the model prediction are also shown



the results obtained using only the time parameter (up to 22 weeks). These results show that the maximum likelihood method performed better than the other two. The average  $N_0$  and k values obtained with the maximum likelihood method were always the closest, or as close as the others, to the true values, and the standard deviations of  $N_0$  and k were always the smallest, or as small as the others. However, the differences between the results of the maximum likelihood and the non-linear least square methods were small and the non-linear least square method consistently gave the best agreement between data and the prediction by the model. This is coherent with the results obtained by Smith et al. (1980), Molina et al. (1980), and Talpaz et al. (1981).

Furthermore, the non-linear least square method has less computation complexity. We therefore prefer this method over the other two for estimating the parameters  $N_0$  and k. We applied it to data from 30 Italian soils.

In Fig. 1, the experimental values of N mineralization for some soils  $(1-8)$  are compared to those of a first-order kinetics model with parameters estimated by the nonlinear least square method. There was generally good agreement between the experimental data and the model for all the soils except soil 7. In this case each of the three methods failed to estimate  $N_0$  and k closely. Though a common pattern in the data suggested the use of a more refined model to allow changes in convexity, we believe that a first-order kinetics model was sufficient to describe these data.

The values of  $N_0$  and k obtained by applying the nonlinear least square method to these data are shown in Table 4. The values obtained for  $N_0$  varied from 21 to 405 ppm, and k ranged from 0.011 to 0.370 week<sup>-1</sup>. When the three soils for which  $k < 0.02$  week<sup>-1</sup> or  $k>0.1$  week <sup>--1</sup> were excluded, the range was reduced to  $0.022 - 0.093$  week<sup>-1</sup>, with an average of 0.050 week<sup>-1</sup> and a mean standard error of 0.005 week<sup>-1</sup>. Further investigations are needed to quantify the variability of the estimates for  $k$  for a single soil. This will allow us to determine whether the variability of the  $k$  values obtained here was a result of sampling variation or not.

Using the  $N_{(30)}$  and  $N_0$  values in Table 4, we observed that in about  $25\%$  of the samples examined  $N_0$  50% higher than the experimental data  $(N_{(30)})$ . However, another  $25\%$  of the samples presented the same value for  $N_0$  and  $N_{(30)}$ . In the remaining samples,  $N_0$  ranged from 30% to 10% higher than  $N_{(30)}$ . For the highest k values

**Table 4** Values of potentially mineralizable N  $(N_0)$  and rate constant  $(k)$  estimated by applying the non-linear least square method to the data from the incubated soils. The values of the root mean square *(RMS)* deviations between the data and the prediction are also shown

<b>RMS</b>
$(week-1)$ (ppm)
5.1
0.8
9.0
2.5
9.2
2.9
nd
3.1
6.6
6.4
4.0
2.1
2.7
4.1
7.4
2.0
7.3
2.0
2.5
2.5
1.8
2.7
1.2
2.1
1.8
1.7
32.0
4.2
1.2
1.9



Table 5 Values of NH $^{+}_{4}$ -N determined after extraction in 2 M KCl at the end of the incubation period (30 weeks)

Soil	$NH4+ -N$	Soil	$NH4+ -N$
no.	(ppm)	no.	(ppm)
$\mathbf{1}$	13.6	18	3.5
	tr	19	4.7
$\frac{2}{3}$	8.8	20	3.2
	2.0	21	2.7
$\frac{4}{5}$	9.5	22	8.5
6	2.0	23	10.9
7	2.6	24	2.8
8	2.0	25	7.3
11	6.0	26	7.3
12	3.4	27	5.1
13	5.9	28	3.9
14	5.6	29	2.2
15	3.2	30	1.7
16	5.1	31	8.3
17	8.0	32	1.6

we obtained the maximum of variability between  $N_0$  and  $N_{(30)}$ . These results suggest that statistical analysis is required for a correct evaluation of  $N_0$  (Stanford and Smith 1972). In addition, Table 4 shows that the ratio between total N and  $N_0$  was characterized, in more than  $65%$  of the samples, by values ranging from 7 to 13. Only three samples were largely outside this range (37, 23, and 19). This result is important because this ratio might be suitable as a rapid index of potentially mineralizable N obtained by measuring only the total N content.

These results were independent of the organic matter content of the soils. In fact, we did not find the highest values of  $N_0$  in soils with the highest humus contents.

Finally, the possibility of reducing the incubation time was examined, by simulation as described above, using the data up to 22 weeks. The results are shown in Table 3. Significantly less reliable estimates of  $N_0$  were obtained using 22 weeks of data. In fact, in some cases, the standard deviation of  $N_0$  was twice the value obtained using 30 weeks of data. Furthermore, with a "low" value of  $k$ , i.e.  $k = 0.03$  week<sup>-1</sup>, even using 30 weeks of data, the standard deviation of  $N_0$  was about three times higher than those corresponding to  $k = 0.05$  or 0.07 week<sup>-1</sup>. In this case, a maximum incubation time greater than 30 weeks should be used. This is in agreement with the work of Mary and Remy (1979) and Menasseri et al. (1994).

The NH $_4^+$ -N values determined in a 30-day incubation period with 2 M KC1 are reported in Table 5. Since all values were very low  $(< 14$  ppm), we concluded that all the available N had been extracted during the experiment.

#### **Conclusions**

The results reported in this paper show that a first-order kinetics model can be successfully used to describe N mineralization in agricultural soils in equilibrium conditions, i.e. when there is no influence from farming activities, because in this second case the first-order kinetics model failed (Alianiello and Benedetti 1994). According to Dendooven et al. (1991), this failure can be explained by the fact that the effects of growing plants, organic matter amendments, or N-fertilizer dressing on N-mineralization capacity were not taken into account and cannot be described by a first-order kinetics model.

The results from the simulation study suggest that the non-linear least square method is preferable to the maximum likelihood and the log-transformed linear least square methods for estimating the parameters  $N_0$  and k of the first-order model.

The value of  $k$  estimated by using data from 30 Italian soils was  $0.050$  (mean standard error = 0.005). The present results confirm that it is not possible to obtain reliable values of  $N_0$  if the maximum incubation time is shorter than 30 weeks, and in some cases a maximum incubation time of longer than 30 weeks is necessary. Finally, the  $N_0$  parameter is a measure of potentially mineralizable N and can give useful information on biological activity in soil (Brookes 1992). However, further work is necessary to analyse the  $N_0$  parameter with a forecasting mathematical model structured on the pedoclimatic environment, in order to obtain practical information on rationalization of N fertilization (Stanford and Epstein 1974; Cabrera and Kissel 1988; Hadas et al. 1989).

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