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Drought response of a native and introduced Hawaiian grass

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Abstract The alien grass, Pennisetum setaceum, dominates many of the lowland arid regions that once supported native Heteropogon contortus grassland on the island of Hawaii. Response to drought in a glasshouse was compared between these C₄ grasses to test if success as an invader is related to drought tolerance or plasticity for traits that confer drought tolerance. Pennisetum produced 51% more total biomass, allocated 49% more biomass to leaves, and had higher net photosynthetic rates (P_n) on a leaf area basis than *Heteropogon*. Plants of both species under drought produced less total biomass and increased their allocation to roots compared to well-watered plants, but there was no difference between the two species in the magnitude of these responses. The decline in P_n with decreasing leaf water potential (Ψ_1) was greater for *Pennisetum* compared to Heteropogon. Plasticity in the response of P_n to Ψ_{l} , osmotic potentials, and the water potentials at turgor loss in response to drought were not different between the two species. Stomata were more responsive to Δw in Heteropogon than in Pennisetum and for well-watered plants compared to droughted plants. Plasticity for the stomatal response to Δw , however, was not different between the species. There was no evidence that the alien, Pennisetum, had greater plasticity for traits related to drought tolerance compared to the native, Heteropogon. Higher P_n and greater biomass allocation to leaves resulted in greater growth for Pennisetum compared to Heteropogon and may explain the success of Pennisetum as an invader of lowland arid zones on Hawaii.

Key words Plant invasion · Drought acclimation Hawaii · Heteropogon contortus · Pennisetum setaceum

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Introduction

Explanations for invasive ability in plants often stress the importance of genetic (Barrett and Richardson 1986; Gray 1986) and life history (Baker 1974; Mack and Pyke 1983; Groves 1986) characteristics. Ecophysiological characteristics that confer success on an invading plant, by contrast, have received less attention (Bazzaz 1986; Newsome and Noble 1986). Generalizations about the physiological attributes of invasive plants, such as those proposed by Bazzaz (1979, 1986) for early successional species, may be difficult to formulate because of the broad taxonomic diversity of invading plants that collectively display different growth forms, different environmental requirements, and different ecological roles in the communities they invade (Parsons 1972; Baker 1986; Heywood 1989; Mack 1989).

An invading species, by definition, is one that reproduces and spreads in a community in which it has not occurred before (Mack 1985). Baker (1974) argues that successful invading plants will have a high level of phenotypic plasticity allowing exploitation of resources in a new environment that may bear little resemblance to the home range. For example, in arid environments characterized by infrequent or unpredictable precipitation, a plant invader should have physiological plasticity to allow survival when water is limiting and rapid growth when water is abundant. Community dominance by the invader in this case may stem from the invader's greater physiological plasticity compared to natives. Addressing the importance of specific physiological traits as they relate to the invasive ability of plants, therefore, may be inadequate unless one also considers the plasticity of these traits.

Comparisons between ecologically similar native and alien taxa offer a preliminary assessment of those attributes that confer invasive ability. *Pennisetum setaceum* (Forssk.) Chiov, a perennial C_4 grass, was introduced to the Hawaiian islands early in this century and is now the dominant plant on barren lava flows, in coastal dry grasslands, and in coastal, montane and subalpine dry shrublands on the arid (leeward) side of the island of Hawaii (Wagner et al. 1990; D.G. Williams, personal observation). While *Pennisetum* has proliferated on Hawaii, many native plants in the arid coastal zones have undergone a severe reduction in range and abundance (Cuddihy 1989) principally because of development by people and grazing. For example, *Heteropogon contortus* (L.) P. Beauv., a native perennial grass, was once abundant in the arid coastal zones of Hawaii, but is now dominant only on dry lowland cliffs and in scattered remnants in coastal areas (Degener and Degener 1968; Wagner et al. 1990).

In this study we compared drought responses of *Heteropogon* and *Pennisetum* to evaluate their drought tolerance and to test if *Pennisetum*, the alien, has greater plasticity for characters that confer drought tolerance compared to the native, *Heteropogon*.

Materials and methods

Seeds of Heteropogon contortus were collected in March, 1991 from a population at Puuhonua 'o' Honaunau National Historical Park on the leeward side of Hawaii. The population was deliberately established in 1969, and the community in which it occurs is maintained to provide an example of a native Heteropogon grassland (Wagner et al. 1990; K. Domingo, personal communication). Heteropogon contortus is widespread throughout the tropics and subtropics and is a valuable pasture grass in many arid regions (Hitchcock 1950, Tothill 1968). The race of Heteropogon contortus on Hawaii, however, is cytogenetically and morphologically distinct and may be endemic to the islands (Degener and Degener 1968, Wagner et al. 1990). Seeds of Pennisetum setaceum were collected from greenhouse representatives of populations from 20, 960, and 1995 m elevation on Hawaii that we have used in related studies (Williams 1992; Williams and Black 1993). Seeds were sown in flats in a greenhouse at Washington State University, Pullman, WA, United States (740 m elevation) on 6 August 1991. After germination and growth for 2 weeks, 32 uniformly-sized seedlings with one primary tiller and three to five leaves, from each population (96 plants from Pennisetum and 32 from Heteropogon), were transplanted individually to 5.4 l peat pots filled with a mixture of crushed basalt and weathered granitic sand. Plants were grown at a constant 30° C on benches in a greenhouse under natural lighting, but received supplemental lighting from high pressure metal halide lamps. Average high temperatures in summer in coastal areas on leeward Hawaii are about 33° C (Atlas of Hawaii, 1983; Williams and Black 1993). Photosynthetic photon flux density at mid-day in the glasshouse was $1241 \pm 19 \mu mol pho$ tons m⁻²s⁻¹ on clear days.

All plants were watered daily with 500 ml water. After 4 weeks, plants from each species were randomly assigned to two watering treatments: (1) a well-watered treatment (W), where plants were watered daily with 500 ml water, and (2) a drought treatment (D), where plants were watered only every 6–8 days. A dilute fertilizer solution (Peter's 20-20; 100 ppm N and micronutrients) was used for all watering. The locations of pots on the greenhouse benches were randomized every 2 weeks.

Water relations

After the drought treatment had gone through five cycles, six plants from each species and watering treatment combination were randomly selected for pressure-volume analysis. One young, fully expanded blade (leaf) from each plant was used for analysis following rehydration overnight in a dark cabinet. Pressure-volume methods followed those of Robichaux (1984), where the weight and water potential were measured periodically with a pressure chamber (PMS Instruments Co., Corvallis, Oregon) as the leaves dried on the laboratory bench. Non-linear regression models described by Schulte and Hinckley (1985) were used to estimate osmotic potential at full turgor ($\Psi_{\pi 100}$), osmotic potential at turgor loss ($\Psi_{\pi 0}$), relative water content at turgor loss (RWC₀), bulk modulus of elasticity (*E*), and symplastic water fraction (SWF).

Gas exchange

After six drought cycles, three to five plants from each species and watering treatment combination were randomly selected for measurement of the response of gas exchange to leaf-to-air vapor pressure difference (Δw) with a compensating, null-balance gas exchange system (Bingham et al. 1980) and an infrared gas analyzer (225-MK3, ADC Ltd., Herts, England). Plants were kept well-watered over the 2-day measurement period. Net photosynthesis (P_n), stomatal conductance (g_s), and the ratio of internal to ambient CO₂ concentration (c_i/c_a) were measured on six or seven young, fully expanded leaves placed in the cuvette. Measurements on each plant were made sequentially at Δw ranging from approximately 1.0 to 3.5 kPa at 0.5 kPa intervals. Leaf temperature and photosynthetic photon flux density (PPFD) were maintained at 30° C and 1800 µmol quanta m⁻²s⁻¹, respectively. Carbon dioxide concentration of air in the cuvette was maintained at 350 µl·l⁻¹.

Gas exchange responses to decreasing soil and plant water potentials were evaluated at the end of the seventh drought cycle. Four days into the normal drying cycle for droughted plants, daily watering was terminated for well-watered plants. Gas exchange was measured on plants for both treatments three days later with a portable photosynthesis system (LCA-3, ADC Ltd., Herts, England). Soil water was depleted more rapidly in well-watered plants because of their greater leaf area and measurements were taken when approximately one-third of the plants from each treatment showed visible signs of water stress (e.g., leaf curling and fading color). Carbon dioxide concentration of the air in the chamber during these differential measurements averaged $349.5 \pm 1.4 \ \mu l \ l^{-1}$, while leaf temperature averaged $33.8 \pm 0.1^{\circ}$ C. PPFD was main-tained near 1000 µmol quanta m⁻²s⁻¹, and Δw averaged 3.75 ± 0.05 kPa during measurements. All measurements were made on one day between 0930 and 1630 hours. One fully expanded leaf was placed in the cuvette and gas exchange measurements were recorded two minutes later. Water potential (Ψ_1) of an adjacent leaf from the same plant was measured with a pressure chamber concurrently with the gas exchange measurement. Specific leaf area (SLA) was determined from these samples after assessing leaf area and weighing oven dried samples to the nearest 0.1 mg.

Growth and carbon allocation

All plants were harvested after 14 weeks of growth, i.e., 8 weeks after the initiation of watering treatments. The number of living and dead tillers were counted, and plants were separated into root, culm (culm + sheath), and leaf fractions, and weighed after drying at 65 °C for at least 48 h. The biomass allocated to roots or leaves was calculated as the fraction of root or leaf biomass to total plant biomass.

Statistical analysis

Data were analyzed using the GLM Procedure in the SAS statistical package (Freund et al. 1986). The species by watering treatment interaction was interpreted as a difference in plasticity between the two grasses (population variance for *Pennisetum* was factored out in the analyses). Water relations parameters were analyzed as a factorial experiment (species by watering treatment) in a completely randomized design. Gas exchange responses to Δw or Ψ_1 were compared using regression models with species and Table 1 Means, SEs (in parentheses), and significance of sources of variation for leaf water relations parameters in Pennisetum and Heteropogon subjected to drought and wellwatered treatments in the greenhouse. Four to five plants were measured for each species and treatment combination. Osmotic potential at full turgor ($\Psi_{\pi 100}$), osmotic potential at turgor loss ($\Psi_{\pi 0}$), relative water content at turgor loss (RWC₀), bulk modulus of elasticity (E), and symplastic water fraction (SWF) were estimated from pressure-volume analysis 5 weeks after the start of watering treatments

| ·· = ··· | Ψ_100 | Ψ_0 | RWC | E | SWF |
|----------------------|--------------|--------------|--------------|--------------|--------------|
| | (MPa) | (MPa) | (%) | (MPa) | (%) |
| Heteropogon | | | | | |
| Well-watered | -1.32(0.07) | -1.56(0.12) | 88.23 (2.06) | 25.25 (8.59) | 82.25 (4.76) |
| Drought | -1.51 (0.06) | -1.90 (0.08) | 85.68 (1.78) | 20.50 (2.86) | 65.04 (4.86) |
| Pennisetum | | | | | |
| Well-watered | -1.02(0.04) | -1.11(0.05) | 93.36 (1.13) | 18.95 (3.51) | 69.06 (5.49) |
| Drought | -1.23(0.06) | -1.41(0.10) | 91.17 (2.60) | 23.67 (6.96) | 56.80 (5.58) |
| Sources of variation | · · · · | · · · | · · · · | · · · · | · · · · |
| Species | *** | *** | * | ns | ns |
| Watering treatment | ** | ** | ns | ns | * |
| Species × watering | ns | ns | ns | ns | ns |
| treatment | | | | | |

* P<0.05, ** P<0.01, *** P<0.001

watering treatment defined as class variables and Δw and Ψ_1 defined as continuous variables in SAS. This is equivalent to an analysis of covariance where a significant interaction between Δw or Ψ_1 and a class variable was interpreted as a difference in the regression coefficient (slope) of the response among class variables (Freund et al. 1986). Morphological data were analyzed as a factorial experiment (species by watering treatment). Data were transformed as necessary to comply with assumptions of analysis of variance.

Results

We found no differences among the three Pennisetum populations for their response to drought in this study. Furthermore, transplant and common environment studies (Williams 1992; Williams and Black 1993) and enzyme electrophoresis (D.G. Williams, unpublished observations) have provided data supporting the lack of genetic differentiation among these three populations. Data for the three Pennisetum populations, therefore, were pooled for comparison with Heteropogon.

Water relations

Heteropogon and *Pennisetum* responded to the drought by lowering osmotic potentials 0.2–0.3 MPa (Table 1). Heteropogon had significantly lower $\Psi_{\pi 100}$ and $\Psi_{\pi 0}$ (0.3– 0.5 MPa lower, respectively) than did Pennisetum (Table 1). There was no difference in the amount of reduction of $\Psi_{\pi 100}$ and $\Psi_{\pi 0}$ between the two species (Table 1, ANOVA, $\Psi_{\pi 100}$: F = 0.02, P = 0.88, $\Psi_{\pi 0}$: F = 0.08, P = 0.78). Bulk modulus of elasticity (E) was similar between the two species, and did not change in response to drought (Table 1, ANOVA, F = 0.74, P = 0.40). Relative water content at turgor loss (RWC₀) was lower in Heteropogon than in Pennisetum (Table 1), but was not affected by watering treatment (Table 1, ANOVA, F = 1.56, P = 0.23). There was no significant difference in the symplastic water fraction (SWF) between the two species (Table 1, ANOVA, F = 3.95, P = 0.07). SWF was reduced by the drought treatment but the species by



Fig. 1 The response of stomatal conductance, net photosynthesis, and the ratio of internal to ambient CO₂ concentration (c_i/c_a) to leaf-to-air vapor pressure difference (VPD) in Heteropogon and Pennisetum subjected to drought (open symbols) and well-watered (closed symbols) treatments. Symbols are individual measurements from plants measured over the range of VPDs. Three to four plants were measured for each species and treatment combination

Fig. 2 The response of stomatal conductance and net photosynthesis to leaf xylem water potential in *Heteropogon* and *Pennisetum* subjected to drought (*open symbols*) and well-watered (*closed symbols*) treatments. Each symbol represents the measurement on one plant 4 days after watering for both treatments



treatment interaction was not significant (Table 1, ANOVA, F = 0.21, P = 0.65).

Gas exchange

Stomatal conductance (g_s) and net photosynthesis (P_n) were higher in Pennisetum than in Heteropogon (ANCO-VA, g_s : F = 142.7, P < 0.0001, P_n : F = 151.8, P < 0.0001). Well-watered plants of both species also exhibited higher g_s and P_n than did plants that had experienced drought (Fig. 1, ANCOVA, g_s : F = 204.6, P < 0.0001, $P_{\rm n}$: F = 328.1, P < 0.0001). Both species responded to increasing leaf-to-air vapor pressure difference (Δw) by reducing g_s and consequently P_n (Fig. 1, ANCOVA, g_s : $F = 113.9, P < 0.0001, P_n: F = 47.0, P < 0.0001$). Stomatal conductance responded more to Δw in *Heteropogon* than in *Pennisetum* (ANCOVA, F = 4.1, P < 0.05) and in well-watered plants compared to droughted plants (Fig. 1; ANCOVA, F = 5.5, P < 0.05). Watering treatments, however, did not differentially affect the two species' stomatal response to Δw as indicated by a lack of a Δw by species by watering treatment interaction (AN-COVA, F = 1.6, P = 0.21). Conversely, there were no differences in the response of P_n to Δw between the two species or watering treatments (ANCOVA, Δw by species: F = 1.2, P = 0.29, Δw by treatment: F = 1.9, P = 0.17). c_i/c_a decreased at higher Δw (Fig. 1, ANCO-VA, F = 25.4, P < 0.0001). The response of c_i/c_a to Δw , however, was not different between the two species (ANCOVA, F = 2.47, P = 0.12) or watering treatments (ANCOVA, F = 0.95, P = 0.33) and the watering treatments did not differentially alter the two species' responses to Δw for both P_n (ANCOVA, F = 1.02, P = 0.32) and c_i/c_a (ANCOVA, F = 3.19, P = 0.08).

The relationship between leaf water potential (Ψ_1) and g_s or P_p varied between the two species and watering treatments. With decreasing Ψ_1 the decline in g_s and P_n was greater for Pennisetum than for Heteropogon (Fig. 2, g_s : ANCOVA, F = 12.8, P < 0.001, P_n : ANCO-VA, F = 5.07, P < 0.05). For *Pennisetum*, photosynthetic rates were near zero at Ψ_1 of -2.5 to -3.5 MPa, while $P_{\rm n}$ in *Heteropogon* reached zero at -3 to -4 MPa. Watering treatments also influenced the response of g_s and $P_{\rm n}$ to decreasing $\Psi_{\rm l}$. Plants exposed to drought had higher g_s and P_n at low Ψ_1 compared to well-watered plants. The magnitude of stomatal adjustment to watering treatment, however, was not different between the two species (ANCOVA, F = 0.01, P = 0.93). The slope of the regression of P_n on g_s , the intrinsic water-use efficiency, was greater for Pennisetum than for Heteropogon (Fig. 3, ANCOVA, F = 7.3, P < 0.01). Watering treatments did not significantly influence the slope of the relationship between g_s and P_n (ANCOVA, F = 0.40, P=0.53) and Heteropogon and Pennisetum did not respond differentially to the watering treatments (Fig. 3, ANCOVA, F = 2.93, P = 0.09). Specific leaf area (SLA) was significantly higher in Heteropogon than in Pennisetum (ANOVA, P < 0.05) and the response of SLA to drought was different between the two species (ANOVA, P < 0.05). Specific leaf area was 276.79 ± 11.11 and 301.66 ± 8.78 cm² g⁻¹ in well-watered and droughted plants of *Heteropogon* and 261.93 ± 8.59 and 233.74 ± 5.08 cm² g⁻¹ in *Pennisetum*.

Growth and biomass allocation

Total biomass at harvest was lower in droughted plants of *Heteropogon* and *Pennisetum*, but *Pennisetum* pro-



Fig. 3 The relationship between stomatal conductance and net photosynthesis in *Heteropogon* and *Pennisetum* subjected to drought (*open symbols*) and well-watered (*closed symbols*) treatments. Data are the same as for Fig 2



Fig. 4 The distribution of biomass among roots, culms, and leaves at harvest in *Heteropogon* and *Pennisetum* subjected to drought (D) and well-watered (W) treatments. For each treatment for *Pennisetum* 48 plants were harvested and 16 plants were harvested for each treatment for *Heteropogon*

Table 2 Means, SEs (in parentheses), and significance of sources of variation for morphological characters in *Pennisetum* and *Heteropogon* subjected to droughted and well-watered treatments in the glasshouse at harvest

| | Total number of tillers | Number of live tillers | Total biomass (g) | Root fraction (g g ⁻¹) | Leaf fraction (g g ⁻¹) |
|--------------------------------------|-------------------------------|------------------------------|-------------------------|--|--|
| Heteropogon | | | | | |
| Well-watered | 69.22 (7.09) | 69.22 (7.09) | 18.37 | 0.26 | 0.31 |
| Droughted | 18.75 | 16.58 | 5.42 | (0.01) (0.02) | (0.01) (0.01) |
| Pennisetum | (1.05) | (2.10) | (0.55) | (0.02) | (0.01) |
| Well-watered | 107.97 (2.93) | 107.97 (2.93) | 27.24 (0.79) | 0.28 (0.01) | 0.44 (0.01) |
| Droughted | 32.49 | 28.45 | 8.78 | 0.30 | 0.44 (0.01) |
| Sources of variation | (111) | (1.00) | (0) | (0,01) | (0.01) |
| Species Watering treat- | *** | ** | *** | ns | *** |
| ment | *** | *** | *** | * | ns |
| Species × watering treat- ment | ns | ns | ns | ns | ns |

* P<0.05, ** P<0.01, *** P<0.001

duced more biomass than *Heteropogon* regardless of watering treatment (48 and 62% more in the well-watered and drought treatments, respectively) (Fig. 4, Table 2). The effect of watering treatment on biomass was similar for the two species (species by watering treatment interaction, ANOVA, F = 0.34, P = 0.56). Greater biomass in *Pennisetum* was accompanied by greater biomass allocation to leaves compared to that in *Heteropogon*, but biomass allocation to roots was not different between the two species (Table 2, ANOVA, F=0.31, P=0.58). Heteropogon and Pennisetum responded to drought by increasing biomass allocation to roots (23% and 7% increase, respectively), but there was no difference in the magnitude of response between the two species (Table 2, ANOVA, F = 0.12, P = 0.73). Both the total number of tillers and the number of living. tillers at harvest for both species were significantly greater in plants from the well-watered treatment compared to plants from the drought treatment (Table 2). Pennisetum produced 55 and 68% more total tillers than Heteropogon in the well-watered and drought treatments, respectively (Table 2), but the species by treatment interaction was not significant (ANOVA, F = 0.97, P = 0.33).

Discussion

High photosynthetic capacity and high allocation to leaves may be common attributes of aggressive alien grasses in arid environments. Maximum photosynthetic rates were higher and biomass allocation to leaves was greater in *Pennisetum* compared to *Heteropogon*, regardless of the watering treatment. These characteristics allowed *Pennisetum* to accumulate more biomass than *Heteropogon* even under drought in our glasshouse study. Similarly, Baruch et al. (1985) found that invasive C_4 grasses in the tropical savannas of Venezuela have higher photosynthetic rates, allocate more biomass to leaves and have higher growth rates than the native grass, *Trachypogon plumosus*. Even the greater productivity of the alien C_3 grass Ammophila arenaria compared to the native C_3 Elymus mollis of dunes along the Pacific coast of North America apparently relates to the alien's greater allocation of biomass to leaves (Pavlik 1983).

Accumulating more biomass is positively correlated with competitive ability and fitness in plants (Harper 1977). Greater vegetative growth when water is plentiful as well as under recurring drought may allow Pennisetum to maintain dominance in the arid zones on Hawaii. In addition, high photosynthetic rates and high allocation to leaves may be advantageous in these environments where disturbance by fire and grazing is frequent (Cuddihy 1989) and the availability of water is infrequent and seasonal (Atlas of Hawaii 1983; Williams and Black 1993). Although our experiment was conducted under lower light levels than those that *Heteropogon* and Pennisetum experience in the field (1250 µmol photons m⁻²s⁻¹ compared to about 2200 µmol photons m⁻ $2s^{-1}$ for field sites on Hawaii), the relative photosynthetic and growth rate differences between the two grasses under natural conditions on Hawaii is likely similar.

Pennisetum and *Heteropogon* had contrasting patterns of stomatal response to drought. While Pennisetum had a stronger stomatal response to decreased leaf water potential than Heteropogon, Heteropogon exhibited greater stomatal response to Δw than did *Pennise*tum. These contrasting responses may reflect different, but not mutually exclusive processes that limit water loss and protect tissues from rapid dehydration during drought. The capacity to limit transpiration by closing stomata under conditions of high evaporative demand apparently is not related to invader status for Pennisetum and Heteropogon. Stomatal conductance declined 20–35% in Heteropogon but only 15% in Pennisetum over a range of Δw . Furthermore, prior exposure to soil drought reduced the stomatal response to Δw for both grasses. Reductions in P_n with increasing Δw were apparently due to stomatal limitations and not reductions in photosynthetic capacity since c_i/c_a declined concomitantly with g_s . Stomatal response to Δw may minimize water loss and tissue dehydration under extreme evaporative demand (Hall et al. 1976) and variation among species in stomatal response to Δw has been found in other studies (Jarvis 1980; Losch and Tenhunen 1981; Turner et al. 1984). Our results show that the native, Heteropogon, has a greater capacity to respond to leafto-air vapor pressure difference than does the alien, Pennisetum. Other comparisons between introduced and native C₄ grasses (Baruch et al. 1985), however, have shown no such differences.

The capacity for stomatal adjustment was not an important characteristic distinguishing *Heteropogon* and *Pennisetum*. Both species showed stomatal adjustment to water deficits; the magnitude of stomatal adjustment, however, did not vary between the two species. Maintenance of photosynthesis at low Ψ_1 is an important characteristic of drought tolerant plants (Ludlow et al. 1983; 1985) and *Heteropogon* maintained higher photosynthetic rates at lower Ψ_1 than did *Pennisetum*. However, stomatal closure in response to decreasing Ψ_1 may protect leaf tissues from damage by limiting water loss (Schulze and Hall 1982) and *Pennisetum* exhibited more rapid stomatal closure in response to decreasing Ψ_1 than did *Heteropogon*.

Stomatal adjustment, or the change in the response of stomata to decreases in $\Psi_{\rm b}$, is likely a consequence of tissue osmotic adjustment to maintain turgor at low Ψ_1 (Turner and Jones 1980). Pennisetum and Heteropogon showed some ability to adjust osmotically, however, the degree of this adjustment was small compared to that for other C₄ grasses (Wilson et al. 1980; Ludlow et al. 1985). Solute accumulation in the guard cells and/or maintenance of turgor in the bulk leaf tissue, or both, could result in stomatal adjustment to water deficits (Pierce and Raschke 1980; Ludlow et al. 1985). The small degree of osmotic adjustment observed in this experiment may have been a consequence of the artificial conditions under which plants were grown, which may have limited full drought acclimation (Ludlow et al. 1985).

Heteropogon and Pennisetum maintained positive carbon gain at Ψ_1 values well below our estimate of the point of turgor loss in the bulk tissue. Overnight tissue rehydration, which is often performed prior to pressurevolume analysis, has been shown to increase estimates of osmotic potential (Evans et al. 1990) and may have influenced our measurements of $\Psi_{\pi 100}$ and $\Psi_{\pi 0}$. Alternatively, guard cells may maintain turgor at lower Ψ_1 than the bulk tissue or stomatal closure may be responsive to signals other than leaf water potential, such as symplast volume (Meinzer et al. 1990; Saliendra and Meinzer 1991) or root water potential (Gollen et al. 1985, Turner 1986, Chaves 1990). In addition, gas exchange was measured 2 weeks (and two drought cycles) after analysis of pressure-volume relationships; plants may have further acclimated to growing conditions which could have led to discrepancies between Ψ_{π} and gas exchange responses to Ψ_1 .

The alien, *Pennisetum*, showed no greater capacity for physiological acclimation or plasticity in response to drought than the native, *Heteropogon*. Only a few studies have compared physiological plasticity of alien and native plants. Caldwell et al. (1981) found that the alien tussok grass *Agropyron desertorum* had greater flexibility of carbon allocation following simulated grazing than did the native *A. spicatum*, which partially accounted for the alien's greater grazing tolerance. Black et al. (1994), working with the same two *Agropyron* species found that the alien *A. desertorum* also possesses greater flexibility for root growth and activity when exploiting soil nutrient patches. In contrast, *Pennisetum* and *Heteropogon* from Hawaii shared a similar capacity to adjust physiologically to drought; the more vigorous growth of *Pennisetum* across both watering treatments could not be accounted for by greater physiological plasticity. Alternatively, the higher photosynthetic rates and proportionately greater allocation of biomass to leaves in *Pennisetum* more adequately explains its greater growth compared to *Heteropogon*. Successful invasions by tropical grasses in aridlands, therefore, may be related to their capacity to persist through drought, but rapidly exploit water when it is available.

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References

- Atlas of Hawaii 2nd edn. (1983) Department of Geography, University of Hawaii, University of Hawaii Press, Honolulu
- Baker HG (1974) The evolution of weeds. Annu Rev Ecol Syst 5:1-24
- Baker HG (1986) Patterns of plant invasion in North America. in Mooney HA, Drake JA (eds) Ecology of biological invasions of North Americal and Hawaii (Ecological Studies 58). Springer, New York, pp 44–57
- Barrett SCH, Richardson BJ (1986) Genetic attributes of invading species. In Groves RH, Burdon JJ (eds) Ecology of biological invasions. Academy of Science, Canberra, pp 21–33
- Baruch Z, Ludlow MM, Davis R (1985) Photosynthetic responses of native and introduced grasses from Venezuelan savannas. Oecologia 67:388-393
- Bazzaz FA (1979) The physiological ecology of plant succession. Annu Rev Ecol Syst 10:351-371
- Bazzaz FA (1986) Life history of colonizIng plants: some demographic, genetic, and physiological features. In Mooney HA, Drake JA (eds) Ecology of biological Invasions of North America and Hawaii (Ecological Studies 58). Springer, New York, pp 96–110
- Bingham GE, Coyne PI, Kennedy RB, Jackson WL (1980) Design and fabrication of a minicuvette system for measuring leaf photosynthesis and stomatal conductance under controlled conditions. Lawrence Livermore Laboratory, University of California, Lawrence, California
- Black RA, Richards JH, Manwaring JL (1994) Nutrient uptake from enriched soil microsites for three Great Basin perennials. Ecology 75:110–122
- Caldwell MM, Richards JH, Johnson DA, Nowak RS, Dzurec RS (1981) Coping with herbivory: photosynthetic capacity and resource allocation in two semiarid *Agropyron* bunchgrasses. Oecologia 50:14-24
- Chaves MM (1990) Effects of water deficits on carbon assimilation. J Exp Bot 42:1-16
- Cuddihy LW (1989) Vegetation zones of the Hawaiian islands. In Stone CP, Stone DB (eds) Conservation biology in Hawaii. University of Hawaii Press, Honolulu, pp 27-35
- Cuddihy LW, Stone CP (1990) Alteration of native Hawaiian vegetation. University of Hawaii Press, Honolulu
- Degener O, Degener I (1968) Flora Hawaiiensis, Fam. 47, Heteropogon contortus. Published privately, Honolulu, Hawaii, USA

- Evans RD, Black RA, Link SO (1990) Rehydration-induced changes in pressure-volume relationships of *Artemisia tridentata*. Nutt. ssp. *tridentata*. Plant Cell Environ 13:455–461
- Freund RJ, Littell RC, Spector PC (1986) SAS system for linear models. SAS Institute, Cary, North Carolina
- Gollen T, Turner NC, Schulze E-D (1985) The responses of stomata and leaf gas exchange to vapour pressure deficits and soil water content III. In the sclerophyllous woody species *Nerium oleander*. Oecologia 65:356–362
- Gray AJ (1986) Do invading species have definable genetic characteristics? Phil Trans R Soc London B314:655-674
- Groves RH (1986) Plant invasions of Australia: an overview. In Groves RH, Burdon JJ (eds) Ecology of biological invasions. Cambridge University Press, Cambridge, pp 137-149
- Hall AE, Schulze E-D, Lange OL (1976) Current perspectives of steady state stomatal responses to environment. In: Lange OL, Kappen L, Schulze E-D, (eds) Water and plant life. (Ecological Studies 19). Springer, Berlin Heidelberg New York pp 169–188
- Harper, JL (1977) The population biology of plants. Academic Press, New York
- Heywood VH (1989) Patterns, extents and modes of invasions by terrestrial plants. In: Drake JA, Mooney HA, Castri F di, Groves RH, Kruger FJ, Rejmanek M, Williamson M (eds) Biological invasions: a global perspective. (SCOPE series 37). John Wiley and Sons, New York, pp 31-60
- Hitchcock, AS (1950) Manual of the grasses of the United States (Miscellaneous Publication 200). United States Department of Agriculture, Washington, DC, USA
- Jarvis, PG (1980) Stomatal response to water stress in conifers. In: Turner NC, Kramer PJ (eds) Adaptation of plants to water and high temperature stress. John Wiley and Sons, New York, pp 105–122
- Losch R, Tenhunen JD (1981) Stomatal responses to humidityphenomenon and mechanism In: Jarvis PG, Mansfield TA (eds) Stomatal physiology. Cambridge University Press, Cambridge, pp 137-161
- Ludlow MM, Chu ACP, Clements RJ, Kerslake RG (1983) Adaptation of species of *Centrosema* to water stress. Aust J Plant Physiol 10:119–130
- Ludlow MM, Fisher MJ, Wilson JR (1985) Stomatal adjustment to water deficits in three tropical grasses and a tropical legume grown in controlled conditions and in the field. Aust J Plant Physiol 12:131–149
- Mack RN (1985) Invading plants: their potential contribution to population biology. In White J (ed) Studies on plant demography: a festschrift for John L. Harper. Academic Press, London, pp 127–142
- Mack RN (1989) Temperate grasslands vulnerable to plant invasions: characteristics and consequences. In Drake JA, Mooney HA, Castri F di, Groves RH, Kruger FJ, Rejmanek M, Williamson M (eds) biological invasions: a global perspective. (SCOPE series 37), John Wiley and Sons, New York, pp 155–180
- Mack RN, Pyke DA (1983) The demography of *Bromus tectorum* L.: variation in time and space. J Ecol 71:69–93
- Meinzer FC, Grantz DA, Goldstein G, Saliendra NZ (1990) Leaf water relations and maintenance of gas exchange in coffee cultivars grown in drying soil. Plant Physiol 94:1781- 1787
- Newsome AE, Noble IR (1986) Ecological and physiological characters of invading species. In Groves RH, Burdon JJ (eds) Ecology of biological invasions. Cambridge University Press, Cambridge, pp 1–20
- Parsons JJ (1972) Spread of African pasture grasses to the American tropics. J Range Manage 25:12-17
- Pavlik M (1983) Nutrient and productivity relations of the dune grasses Ammophila arenaria and Elymus mollis II. Growth and patterns of dry matter and nitrogen allocation as influenced by nitrogen supply. Oecologia 57:233–238
- Pierce M, Raschke K (1980) Correlation between loss of turgor and accumulation of abscisic acid in detached leaves. Planta 148:174-182

- Robichaux RH (1984) Variation in the tissue water relations of two sympatric Hawaiian *Dubautia* species and their natural hybrid. Oecologia 65:75–81
- Saliendra NZ, Meinzer FC (1991) Symplast volume, turgor, stomatal conductance and growth in relation to osmotic and elastic adjustment in droughted sugarcane. J Exp Bot 42:1251–1259
- Schulte PJ, Hinckley TM (1985) A comparison of pressure-volume curve data analysis techniques. J Exp Bot 36:1590–1602
- Schulze E-D, Hall AE (1982) Stomatal responses, water loss and CO₂ assimilation rates of plants in contrasting environments. In: Lange OL, Nobel PS, Osmond CB, Zeigler H (eds) Physiological plant ecology II. Water relations and carbon assimilation. (Encyclopedia of Plant Physiology 12B). Springer, Berlin, pp181-230
- Tothill JC (1968) Variation and apomixis in *Heteropogon contor*tus, Gramineae. Bol Soc Argent Bot 12:188-201
- Turner NC (1986) Adaptation to water deficits: a changing perspective. Aust J Plant Phys 13:175–190
- Turner NC, Jones MM (1980) Turgor maintenance by osmotic adjustment: a review and evaluation. In Turner NC, Kramer PJ (eds) Adaptation of plants to water and high temperature stress. John Wiley and Sons, New York, pp 89–103

- Turner NC, Schulze E-D, Gollan T (1984) The responses of stomata and leaf gas exchange to vapour pressure deficits and soil water content. I. Species comparisons at high soil water content. Oecologia 63:338–342
- Wagner WL, Herbst DR, Sohmer SH (1990) Manual of the flowering plants of Hawaii University of Hawaii Press, Honolulu
- Williams DG (1992) Physiological ecology of the invasive grass Pennisetum setaceum on Hawaii. Ph.D. Dissertation, Washington State University, Pullman, Washington, USA
- Williams DG, Black RA (1993) Phenotypic variation in contrasting temperature environments: growth and photosynthesis in *Pennisetum setaceum* from different altitudes on Hawaii. Funct Ecol 7:623–633
- Wilson JR, Ludlow MM, Fisher MJ, and Schulze E-D (1980) Adaptation to water stress of the leaf water relations of four tropical forage species. Aust J Plant Phys 7:207–220