# **N-methyl-D-aspartate-evoked calcium uptake by kitten visual cortex maintained** *in vitro*

**D. Feldman, J.E. Sherin, W.A. Press, and M.F. Bear** 

Center for Neural Science, Brown University, P.O. Box 1953, Providence, RI 02912, USA

**Summary.** As a functional measure of NMDA receptor effectiveness in kitten striate cortex, the uptake of  $45Ca$ by visual cortical slices was measured after 2 minute bath applications of N-methyl-D-aspartate (NMDA). Significant Ca uptake occured in response to  $12.5-100 \mu M$ NMDA in slices prepared from visual cortex of normal **animals** aged 28-48 days. Basal uptake (in the absence of NMDA) was increased and evoked uptake was decreased in visual cortical slices prepared from age-matched dark-reared animals. Four days of binocular deprivation in otherwise normally reared animals had no effect on basal uptake, but significantly lowered NMDAevoked Ca uptake at agonist concentrations greater than  $25 \mu M$ . These data suggest that even brief manipulations of sensory experience are sufficient to alter visual cortical calcium regulation.

**Key words:** Calcium uptake – NMDA receptor – Striate cortex - Deprivation - Kitten - Synoptic plasticity

## **Introduction**

Neurons in the striate cortex of normal adult cats respond selectively to the visual presentation of oriented bars of light and most are activated by stimulation of either eye (Hubel and Wiesel 1962). Both of these properties depend on the visual environment experienced during a critical period of postnatal development (reviewed by Sherman and Spear 1982; Frégnac and Imbert 1984). For example, prolonged binocular deprivation during the critical period (BD) broadens or eliminates orientation selectivity, and binocular connections **are**  modified after very brief periods of monocular deprivation (MD), strabismus, and reverse suture.

Bienenstock, Cooper and Munro (1982) have proposed a theoretical form of synaptic modification for the visual cortex that appears sufficient to account for

*Offprint requests to:* M.F. Bear (address see above)

these varied experimental results (reviewed by Bear *et at.*  1987). According to this theory, the synaptic efficacy of active geniculocortical inputs increases when the postsynaptic target is concurrently depolarized beyond a "modification threshold",  $\theta_M$ . However, when the level of postsynaptic activity falls below  $\theta_M$ , the strength of active synapses decreases. Importantly, the value of the modification threshold varies as a non-linear function of the average output of the postsynaptic neuron. This feature is important for model stability, and explains why the low level of postsynaptic activity caused by binocular deprivation does not drive the strengths of all cortical synapses to zero. The predictive success and simple basis of the model suggests that some types of visual cortical plasticity may be the result of a molecular mechanism which reflects the model's basic structure.

**Experimental** 

**Brain Research**  9 Springer-Verlag 1990

A growing body of evidence suggests that the activation of NMDA receptors and the entry of calcium postsynaptically may play a role in the mechanism for visual cortical plasticity. The NMDA receptor is a subtype of excitatory amino acid (EAA) receptor which has high affinity for the agonist N-methyl-D-aspartate (NMDA) and is selectively antagonized by 2-amino-5 phosphonovaleric acid, APV (Watkins and Evans 1981). This receptor is associated with a membrane channel that passes  $Ca<sup>2+</sup>$  only when postsynaptic depolarization overcomes a voltage-dependent magnesium ion blockade (Nowak *et al.* 1984). A special role for this receptor **in**  synaptic plasticity is suggested by several lines of evidence. Long-term potentiation (LTP) of synaptic effectiveness, normally resulting from tetanic electrical stimulation of excitatory afferents, cannot be induced when NMDA receptors are blocked in either the CA1 subfield of the hippocampus (Collingridge *et al.* 1983; Harris *et al.* 1984) or the visual cortex of rats (Artola and Singer 1987; Kimura *et aI.* 1988) and kittens (Connors and Bear 1988). Likewise, modification of striate cortex by visual deprivation during the critical period is disrupted by chronic intracortical infusion of APV (Kleinschmidt *et al.* 1987; Gu *et al.* 1989; Bear *et al.* 1990). On the other hand, the application of N-methyl-D-aspartate to hippo-

campal slices *in vitro* can induce a form of synaptic potentiation that can last for 30 min (Collingridge *et aI.*  1983; Kauer *et al.* 1988) or longer (Thibault *et al.* 1989). The idea that elevations in postsynaptic  $[Ca^{2+}]$  trigger the increase in synaptic strength is supported by the finding that intracellular injection of the  $Ca^{2+}$  chelator EGTA blocks the induction LTP in CA1 pyramidal cells (Lynch *et al.* 1983). Further, the intracellular release of  $Ca<sup>2+</sup>$  from the photolabile calcium chelator nitr-5 produces a long-lasting potentiation of synaptic transmission that resembles LTP (Malenka *et al.* 1988). Taken together, these data indicate that the calcium conductance mediated by the NMDA receptor plays a special role in strengthening synaptic relationships in the cortex.

This molecular mechanism is consistent with the theory of Bienenstock *et al.* (1982) assuming that visual cortical NMDA receptors become sufficiently active to increase synaptic strength only when the postsynaptic target is depolarized beyond the modification threshold,  $\theta_M$  (Bear *et al.* 1987; Bear 1988). The theory states that the value of  $\theta_M$  varies as a non-linear function of the average output of the postsynaptic neuron. Does it follow that the effectiveness of NMDA receptor activation in triggering a synaptic modification is also a function of average cortical activity?

As a first step towards addressing this question, we have investigated the effects of visual deprivation on NMDA-stimulated 45Ca uptake by slices of visual cortex maintained *in vitro.* Our results demonstrate that even brief periods of visual deprivation can significantly affect NMDA-stimulated calcium accumulation by visual cortical slices, It remains to be determined, however, if what we observe is a reflection of changes in the synaptic modification threshold.

#### **Methods**

Kittens used in this study are listed in Table 1. Most normally reared animals were purchased immediately post-weaning from Liberty Labs (Liberty Corner, NJ). Visual deprivation by lid suture was carried out under ketamine-xylazine anesthesia 4 days prior to

Table 1. Listed are each animal used in this study, the type of rearing (NR, normal rearing; DR, dark-rearing; BD, binocular deprivation; MD, monocular deprivation), the age when each animal was deprived and sacrificed (in days postnatal), and the calcium accumulated by slices of visual cortex after 2 minute incubations in 0-100  $\mu$ M NMDA and 62.5 mM K. Each value for calcium accumulation is the mean of determinations on 4-8 slices

| Animal          | Rearing    | Age when<br>deprived | Age when<br>sacrificed | Calcium accumulation (nmol per mg protein) |      |      |      |      |      |
|-----------------|------------|----------------------|------------------------|--------------------------------------------|------|------|------|------|------|
|                 |            |                      |                        | NMDA concentration $(\mu M)$ :             |      |      |      |      | K    |
|                 |            |                      |                        | $\bf{0}$                                   | 12.5 | 25   | 50   | 100  |      |
| N1              | NR         |                      | 28                     | 4.33                                       |      | 5.59 |      |      | 5.35 |
| N2              | NR         |                      | 34                     | 3.65                                       | 5.03 | 4.60 | 5.02 | 5.18 | 5.23 |
| N3              | NR         |                      | 34                     | 4.08                                       | 4.40 | 5.08 | 5.33 | 5.92 | 6.40 |
| N <sub>4</sub>  | <b>NR</b>  |                      | 39                     | 3.64                                       | 3.15 | 4.43 | 4.95 | 4.74 | 4.61 |
| N <sub>5</sub>  | NR         |                      | 31                     | 3.40                                       | 4.55 | 5.19 | 5.38 | 5.63 | 5.90 |
| N <sub>6</sub>  | ${\bf NR}$ |                      | 41                     | 2.78                                       | 3.53 | 3.64 | 3.85 | 3.84 | 4.19 |
| N7              | <b>NR</b>  |                      | 46                     | 2.52                                       | 2.58 | 3.22 | 4.64 | 4.87 | 4.25 |
| N8              | <b>NR</b>  |                      | 48                     | 3.09                                       | 3.64 | 4.63 | 5.65 | 5.48 | 4.01 |
| N <sub>9</sub>  | <b>NR</b>  |                      | 86                     | 3.17                                       | 2.69 | 2.83 | 3.75 | 3.22 | 3.54 |
| N10             | <b>NR</b>  |                      | 87                     | 2.39                                       | 3.27 | 2.67 | 3.03 | 3.55 | 3.33 |
| N11             | NR         |                      | 88                     | 3.56                                       | 3.12 | 2.58 | 3.05 | 4.64 | 3.68 |
| N12             | $\rm NR$   |                      | 33                     | 3.22                                       | 3.46 | 4.54 | 3.15 | 5.42 | 4.55 |
| N13             | ${\bf NR}$ |                      | 55                     | 2.72                                       |      | 3.18 |      |      |      |
| N <sub>14</sub> | <b>NR</b>  |                      | 56                     | 2.92                                       |      | 4.05 |      |      |      |
| N <sub>15</sub> | NR         |                      | 57                     | 4.03                                       |      | 4.56 |      |      |      |
| D <sub>1</sub>  | DR         | $\boldsymbol{7}$     | 32                     | 3.59                                       | 3.88 | 4.01 | 3.91 | 4.66 | 4.93 |
| D2              | DR         | $\overline{7}$       | 33                     | 4.33                                       | 4.93 | 5.42 | 5.04 | 6.05 | 5.72 |
| D <sub>3</sub>  | <b>DR</b>  | $\boldsymbol{7}$     | 40                     | 4.49                                       | 4.76 | 4.56 | 5.48 | 5.16 | 5.50 |
| D <sub>4</sub>  | <b>DR</b>  | $\overline{7}$       | 43                     | 4.21                                       | 4.12 | 4.54 | 4.04 | 3.76 | 4.38 |
| D <sub>5</sub>  | <b>DR</b>  | $\tau$               | 32                     | 4.88                                       | 4.84 | 5.15 | 5.55 | 6.72 | 4.92 |
| D <sub>6</sub>  | <b>DR</b>  | 7                    | 46                     | 3.49                                       | 4.18 | 4.80 | 4.74 | 3.89 | 4.57 |
| B1              | <b>BD</b>  | 38                   | 42                     | 2.45                                       | 2.85 | 3.93 | 3.43 | 3.90 | 3.28 |
| B2              | BD         | 41                   | 45                     | 3.00                                       | 3.53 | 2.50 | 2.68 | 3.91 | 4.17 |
| <b>B3</b>       | <b>BD</b>  | 44                   | 48                     | 4.09                                       |      | 5.41 | 5.89 | 5.75 | 5.74 |
| <b>B4</b>       | <b>BD</b>  | 28                   | 32                     | 3.20                                       | 4.16 | 4.22 | 3.96 | 4.36 | 3.72 |
| B <sub>5</sub>  | <b>BD</b>  | 30                   | 34                     | 2.64                                       | 3.09 | 3.57 | 3.14 | 3.41 | 3.21 |
| <b>B6</b>       | <b>BD</b>  | 30                   | 34                     | 2.34                                       | 3.41 | 2.69 | 3.44 | 3.19 | 3.93 |
| M1              | MD         | 37                   | 41                     | 2.48                                       | 4.03 | 3.87 | 5.46 | 5.63 |      |
| M <sub>2</sub>  | MD         | 32                   | 36                     | 3.88                                       | 3.83 | 4.37 | 4.19 | 4.75 |      |
| M <sub>3</sub>  | MD         | 36                   | 40                     | 3.07                                       | 3.55 | 4.78 | 4.29 | 4.70 |      |
| M <sub>4</sub>  | MD         | 28                   | 32                     | 4.54                                       | 6.63 | 4.74 | 4.71 | 6.13 |      |
| M <sub>5</sub>  | MD         | 35                   | 39                     | 4.41                                       | 3.67 | 4.50 | 4.95 | 4.77 |      |

sacrifice. Kittens to be dark-reared were taken from a breeding colony operated by the Brown University Animal Care Facility. Mother and kittens were transfered from the colony within 7 days of birth (before the time of natural eye opening) to a standard breeding cage in a darkroom. Care for these animals was provided in accordance with procedures approved by the Brown University Institutional Animal Care and Use Committee.

Prior to sacrifice, each kitten was pre-anesthetized with a small i.m. dose of a ketamine/xylazine mixture (20 and 2.5 mg per kg, respectively) followed by an intravenous or intraperitoneal dose of sodium pentobarbitol (10 mg per kg and upwards as needed to maintain deep surgical anesthesia). Ketamine was used to induce anesthesia because it could be administered easily to kittens in the darkroom. Although there is good evidence that ketamine blocks NMDA receptor channels (Anis *et al.* 1983; Thompson *et al.* 1985; MacDonald *et al.* 1987), we are confident that this did not compromise our experiments. First, the dose was small enough that its anesthetic effects would have worn off prior to the removal of blocks of visual cortex. Second, slices of area 17 were washed in a large volume of artificial cerebrospinal fluid before incubation with NMDA (see below). Finally, pilot studies indicated that anesthesia with sodium pentobarbitol alone did not alter the results of the calcium uptake assay. Because all animals were treated identically, comparisons between groups remain valid.

The anesthetized kitten was placed in a stereotaxic head-holder and a large craniotomy was performed to expose the dorsal surface of the cerebrum. The dura overlying visual cortex was excised and the animal decaptitated. The brain surface was irrigated with cold oxygenated artificial cerebrospinal fluid (ACSF; 120 mM NaC1,  $5 \text{ mM KCl}$ , 1 mM  $MgCl<sub>2</sub>$ , 2 mM  $CaCl<sub>2</sub>$ , 10 mM d-glucose, 20 mM HEPES buffer, pH 7.4) as blocks of visual cortex were gently removed. These blocks, illustrated schematically in Fig. 1A, consisted mostly of area 17. The blocks were cut in the coronal plane into 400 gm slices using a McIlwain tissue chopper and the slices were gently separated in a petri dish containing ACSF at  $4^{\circ}$  C. Slices were allowed to equilibrate for 45 min in a chamber containing continuously oxygenated ACSF at room temperature. Slices within  $\sim$  1.5 mm of the anterior and posterior block incisions were discarded.

After equilibration, cortical slices were transfered into twelve plexiglass cylinders with nylon mesh bottoms which rested in twelve wells containing oxygenated room temperature ACSF (schematically illustrated in Fig. 1B). Each cylinder held 4 slices. ACSF in the wells was continually bubbled with warmed humidified oxygen from small tubes within the cylinders. Slices were exposed to preincubation, incubation, and postincubation media by transfering the cylinders between sets of wells containing the different media (Fig. 1C). Slices in each cylinder were exposed to a unique set of preincubation and incubation solutions, and thus constituted one experimental condition. Most conditions were run in duplicate within each experiment (i.e., 8 slices per condition).

The calcium uptake assay was a modification of the procedure published by Ichida *et al.* (1982). Slices were first warmed to  $35^{\circ}$  C during a 5-min preincubation. The preincubation was also used in some experiments to expose slices to 100 µM D,L-APV. Preincubation was followed by incubation in ACSF solutions containing approximately 2.5  $\mu$ Ci <sup>45</sup>CaCl<sub>2</sub> and either 0, 12.5, 25, 50 or 100  $\mu$ M NMDA. In addition, one condition, lacking NMDA and containing a high concentration of K<sup>+</sup> (62.5 mM KCl, 60 mM NaCl, 1 mM  $MgCl<sub>2</sub>$ , 2 mM CaCl<sub>2</sub>, 10 mM D-glucose, 20 mM HEPES buffer), was used to monitor depolarization-induced calcium uptake. Pilot studies indicated that both NMDA- and  $K^+$ -stimulated calcium uptake by slices were linear beyond 4 min of incubation. In the experiments described here, a 2 min incubation was used. Incubation was terminated by a 10-s wash in a  $4^{\circ}$  C postincubation solution containing La<sup>3+</sup> (120 mM NaCl, 5 mM KCl, 1 mM MgCl<sub>2</sub>, 10 mM LaC13, 10 mM d-glucose, 20 mM HEPES buffer, pH 7.4), followed by a 30-min postincubation in a fresh bath of the same solution (Drapeau and Blaustein 1983; Retz and Coyle 1984). Slices were removed from the postincubation medium by vacuum filtration and sonicated in 1.0 ml deionized distilled water. The  $45Ca$ content of each slice was determined by scintillation counting in



Fig. 1. A Dissection of visual cortical tissue blocks. *Top:* dorsal view of the left hemisphere to illustrate the region of cortex dissected to prepare slices (black region). *Bottom:* medial aspect of a midsagittal view of a cat brain to illustrate the tissue dissected to prepare visual cortical slices (black region). The sagittal incision intersects the splenial sulcus and isolates a block of cortex containing primarily area 17. This block is subsequently cut in the coronal plane into  $400 \mu m$ thick slices. B Schematic illustration of the slice incubation apparatus. Slices rested on the nylon mesh bottoms of plexiglass cylinders that fit snugly into wells containing the various incubation media. Tubing within cylinder walls supplied uninterrupted oxygen during all phases of incubation. Dissolved oxygen within the wells was greater than 85% saturation at  $20^{\circ}$  C. C The slices were exposed to different media simply by transferring the cylinders from well to well

comparison to aliquots of incubation media, which served as standards. Protein content of each slice was determined by the method of Bradford (1976). The calcium accumulation by each slice was expressed in nmoles per mg protein.

Individual slice  $45$ Ca content values were averaged to arrive at a single mean calcium accumulation for each condition in the experiment (conditions run in duplicate were expressed as the single mean of 8 slices). At least 5 separate experiments were performed, and tissue calcium accumulation for each condition was defined as the mean $\pm$  SEM of the individual experimental means for that condition. Calcium content values were compared across NMDA treatments, animal ages, and rearing histories by t-test, regression, and ANOVA, where appropriate. The confidence level for significance in all tests was  $p < 0.05$ .

## **Results**

Evoked increases in calcium accumulation were measured in slices derived from 9 normally reared (NR) animals, aged 28 to 48 days. Slices were subjected to 2 minute incubations in media containing micromolar NMDA and 62.5 mM  $K^+$ , and the results are shown in table 1. NMDA incubation caused a dose-dependent increase in slice calcium accumulation from control levels of  $3.41 \pm 0.19$  nmol/mg to a maximum of  $5.14 \pm 0.23$ nmol/mg with  $100 \mu M$  NMDA. The NMDA-evoked increase, illustrated in Fig. 2A, was significant (1-way





ANOVA,  $p < 0.001$ ). 62.5 mM K<sup>+</sup> treatment also significantly increased calcium accumulation over control levels (correlated nondirectional t-test,  $p < 0.001$ ). To assess more accurately the response to NMDA and  $K^+$ treatment, the results of each experiment were recalculated by subtracting mean basal (unstimulated) calcium accumulation from the mean calcium accumulation of each treated condition. The resulting data represent NMDA- and K+-evoked calcium *uptake* over control levels (Figs. 2B, 3). These data show that NMDA treatment elicited significant calcium uptake (1-way ANOVA,  $p < 0.002$ ).

To determine what part of this uptake occurred through an NMDA receptor-dependent mechanism, slices



Fig. 3. Calcium uptake evoked by 62.5 mM  $K^+$  in slices from age-matched animals that were raised normally (NR), reared in complete darkness (DR) or briefly binocularly deprived (BD). The reduction in  $K^+$ -evoked uptake in slices from DR animals is significant ( $p < 0.05$ )



Fig. 4. LEFT: Effect of APV treatment on calcium uptake evoked by 25  $\mu$ M NMDA. Control slices were incubated in 25  $\mu$ M NMDA alone; APV-treated slices were preincubated in 100 µM D,L-APV for 5 min and then incubated in both 25  $\mu$ M NMDA and 100  $\mu$ M  $D,L$ -APV. Bars represent means ( $\pm$  SEM) of 5 experiments on slices from normally reared animals (N5, N6, N7, N8, N12). APV reduction is significant ( $p < 0.025$ ). RIGHT: Effect of APV treatment on  $K^+$ -stimulated calcium uptake in the same animals

from 5 normal animals were preincubated with  $100 \mu M$  $D,L-APV$  and then incubated with 100  $\mu$ M D,L-APV and  $25 \mu M$  NMDA. The results were compared to APV-free conditions in the same experiments. These experiments showed that APV treatment significantly reduced the calcium uptake evoked by 25  $\mu$ M NMDA. Untreated slices accumulated  $1.24 \pm 0.20$  nmol/mg during incubation with  $25 \mu M$  NMDA, while APV-treated slices accumulated only  $0.34 \pm 0.32$  nmol/mg. This 70% reduction, shown in Fig. 4, was statistically significant (correlated directional t test,  $p < 0.025$ ). In contrast, neither basal calcium content nor  $K^+$ -stimulated calcium uptake were significantly reduced by APV treatment.

Detailed analysis of these experimental results suggested that stimulated calcium uptake declined in slices from animals of increasing age. To examine this possibility further, calcium uptake was measured in slices obtained from 6 additional kittens, aged 55 to 88 days. Only control and  $25 \mu M$  NMDA conditions were run in 86-88 day old animals (Table 1). Slices from these additional animals showed markedly lower NMDA-stimulated increases in calcium content than did those from the 28 to 48 day old animals (2-way ANOVA,  $p < 0.001$ ). In addition, slices from old animals showed significantly reduced  $K<sup>+</sup>$ - stimulated increases in Ca content (noncorrelated t-test,  $p < 0.02$ ). Subsequent analysis showed that 25  $\mu$ M NMDA-stimulated uptake was inversely correlated with increasing animal age ( $R=0.84$ ,  $p < 0.001$ ; Fig. 5). K<sup>+</sup>stimulated uptake also significantly declined with increasing age ( $R = 0.70$ ,  $p < 0.02$ ); however, basal calcium content remained constant  $(R=0.36)$ .

To assess the effects of visual experience on NMDAevoked calcium uptake, slices derived from 6 dark-reared (DR) animals were incubated under conditions identical to those described for normal slices. The DR animals were age-matched to the NR group (mean ages were 37.7 and 37.1 days, respectively). The calcium accumulation of NMDA and  $K^+$ -treated slices from DR animals (Table 1), was not significantly different from that of NR slices. However, basal (unstimulated) Ca accumulation



Fig. 5. Age-related decline in 25  $\mu$ M NMDA-evoked Ca<sup>2+</sup> accumulation. Each point represents  $25~\mu$ M NMDA-evoked calcium uptake over control levels in a single experiment using slices from normal animals. The decline is significant  $(p<0.001)$  and approximates the illustrated line with  $R=0.84$ . K<sup>+</sup>-evoked uptake also declined with age in these animals; however, basal accumulation did not



Fig. 6. Comparison of calcium accumulation by slices derived from monocularly deprived (open boxes) and BD (filled boxes) animals. MD and BD animals were deprived for 4 days. The difference between visual cortical slices from MD and BD kittens is significant  $(p < 0.001)$ 

was significantly elevated in the slices from DR animals (noncorrelated t-test,  $p < 0.01$ ; Fig. 2C). As a result, NMDA-evoked calcium *uptake* was significantly lower in visual cortical slices from DR animals than in slices from NR animals (2-way ANOVA,  $p < 0.001$ ; Fig. 2D). K<sup>+</sup>evoked uptake was also significantly decreased (noncorrelated t-test,  $p < 0.05$ ; Fig. 3).

Slices derived from animals binocularly deprived (BD) by lid suture for 4 days (mean age 39.2 days) were also subjected to the standard incubation conditions (Table 1). NMDA-treated slices from BD animals showed significantly lower calcium accumulation than did slices from NR animals (2-way ANOVA,  $p < 0.001$ ; Fig. 2E). The  $K^+$  treated slices from BD animals showed slightly lower Ca accumulation than did those from NR animals, but this difference did not achieve statistical significance (noncorrelated t-test,  $p < 0.10$ ). Slices from monocularly deprived (MD) animals (mean age 37.6 days) were also incubated in micromolar NMDA (but not in  $62.5 \text{ mM K}^+$ ; see Table 1). Calcium accumulation by these slices was not significantly different from slices prepared from NR animals, but was significantly elevated over slices from BD animals (2-way ANOVA,  $p < 0.001$ ; Fig. 6).

The difference between slices from normal and BD animals was very robust. Not only was the calcium accumulation of NMDA treated slices significantly lower after BD (Fig. 2E), but NMDA-evoked calcium *uptake*  over basal levels was reduced as well (2-way ANOVA,  $p < 0.025$ ). This difference is illustrated in Fig. 2F. K<sup>+</sup>stimulated uptake was slightly but not significantly reduced after BD (Fig. 3). It is noteworthy that the deficit in calcium uptake seen in Fig. 2F was only apparent above 25  $\mu$ M NMDA. Below that, uptake by slices from BD animals was equal to uptake in slices from NR animals.

## **Discussion**

We have found that NMDA and  $K^+$  evoke uptake of Ca<sup>2+</sup> by slices of kitten visual cortex maintained *in vitro*. The major findings of this study are that this uptake depends importantly upon the postnatal age of the visual cortex and the history of prior visual experience.

The magnitude of NMDA-evoked calcium uptake we measured in P28-48 kitten visual cortex is comparable to values obtained in similar experiments using rat neocortical slices (Ichida *et al.* 1982; Riveros and Orrego 1986; Crowder *et al.* 1987). A variety of pre- and post-synaptic mechanisms could contribute to the measured changes in slice calcium content. Much of the uptake measured under our assay conditions conceivably could occur at presynaptic axon terminals due to impulse activity evoked by NMDA and  $K^+$  application. However, Riveros and Orrego (1986) found in rat neocortical slices that blockade of action potentials (and hence presynaptic  $Ca^{2+}$ entry) with  $1 \mu M$  tetrodotoxin (TTX) failed to reduce calcium uptake stimulated by 150  $\mu$ M N-methyl-D,Laspartate. Postsynaptic calcium uptake is influenced by a number of factors including (1) influx through NMDAreceptor-gated channels (Mayer and Westbrook 1987), (2) influx through voltage-sensitive calcium channels (VSCC's; Miller, 1987), (3) efflux via the  $Ca^{2+}$ - pumping ATPase, and (4) sequestration and release by intracellular calcium buffering systems. The net result of these combined mechanisms is the elevation of intracellular calcium in response to NMDA receptor activation. The APV antagonism of NMDA-evoked calcium uptake we observed confirms that NMDA receptor activation is necessary for calcium uptake. Therefore, we have interpreted the magnitude of NMDA-evoked calcium uptake as a measure of the effectiveness of the NMDA receptor in producing a calcium signal *through these combined mechanisms,* and not through the NMDA-receptor-gated calcium ionophore alone. Because it was not antagonized by APV,  $K^+$ -stimulated uptake is likely to occur via a mechanism that is largely independent of the NMDA receptor.

We found a significant decline in NMDA-evoked calcium uptake in visual cortical slices prepared from animals of increasing age. There are several possible explanations for this result. It is possible that slice protein content increases with age, while the magnitude NMDA-evoked calcium entry does not, causing an apparent decline in calcium uptake per mg protein. We consider this unlikely, however, since basal (unstimulated) calcium content did not decrease significantly with age, as would be expected if protein content were steadily increasing. A second possibility is that slices from older animals were less healthy. Basal (unstimulated) calcium content appears to be a sensitive measure of slice health because it is increased markedly by anoxia and excessive agitation (personal observations). According to this measure, slices from older animals were no less healthy than those from younger animals. A third possibility is that the number of NMDA receptors and/or channels decreases in striate cortex with increasing age. Indeed, recent studies have indicated that

both APV-sensitive glutamate binding (Bode-Greuel and Singer 1989) and MK801 binding to the open NMDA channel (Reynolds and Bear 1989) decline in kitten striate cortex from 5 weeks of age to adulthood. Finally, it is possible that a decrease in the number of VSCC's in striate cortex contributes to the decline in NMDA-stimulated calcium uptake. This possibility is supported by our observation that  $K^+$ -stimulated calcium uptake, which is independent of NMDA receptor activation under our assay conditions, also showed an age-related decline. Furthermore, Bode-Greuel and Singer (1988) have found that 1,4-dihydropyridine binding sites, thought to reflect the density of L-type VSCC's (Miller 1987), decrease in kitten striate cortex during postnatal development.

Taken together, these considerations lead us to suggest that the age-related decrease in NMDA evoked calcium uptake by slices of visual cortex is due largely to a decrease in the number of NMDA- and voltage-gated calcium channels. It is of particular interest that under our assay conditions, calcium uptake evoked by  $25 \mu M$ NMDA became statistically insignificant by 12 weeks of age, considered to be the end of the "critical period" for modification of binocular connections in kitten visual cortex by brief monocular deprivation (Hubel and Wiesel 1970). Thus, these data tend to support the hypothesis that neuronal calcium fluxes play a special role in developmental plasticity in the visual cortex, as suggested by Singer (1987; 1989).

However, experiments using slices prepared from dark-reared animals indicate that there is not always a simple correlation between the occurance of NMDAstimulated calcium uptake and experience-dependent cortical modifications. A period of dark-rearing leaves the striate cortex extremely sensitive to modification by subsequent visual experience (reviewed by Frégnac and Imbert 1984). However, we found a severe deficit in NMDA-evoked calcium uptake, due in part to an unusually high basal calcium content in slices from DR animals. It is possible that DR slices were uniformly less healthy than normal slices, although it is not clear why this would be the case. Alternatively (or in addition), dark-rearing might retard the normal development of NMDA- and voltage-gated calcium channels. This notion is supported by preliminary findings that MK801 binding is substantially lower in the visual cortex of dark-reared as compared with normally reared animals (Reynolds and Bear 1989).

A major motivation in performing these experiments was to determine if a brief period of binocular deprivation changed the effectiveness of NMDA receptor activation in producing a postsynaptic calcium signal. NMDA receptor activation can, under some circumstances, increase synaptic effectiveness in the visual cortex (Kleinschmidt *et al.* 1987; Artola and Singer 1987; Kimura *et al.* 1989). Presumably this effect is mediated by postsynaptic calcium entry, as shown for LTP in the CA1 region of the hippocampus (Lynch *et al.* 1983; Malenka *et al.* 1988). Theoretical considerations had led Bear *et aI.* (1987) to propose that that the level of postsynaptic activation at which sufficient  $Ca^{2+}$  enters through

NMDA channels to increase synaptic strength (the modification threshold,  $\theta_M$ ) should vary depending on the history of prior cell activity (see Introduction). Specifically, a brief period of binocular deprivation should reduce the value of  $\theta_M$ . If this regulation were manifested by changes in NMDA receptor effectiveness, then a brief period of BD should produce enhanced calcium uptake in response to NMDA.

Indeed, 4 days of binocular deprivation did produce significant changes in NMDA- and  $K^+$ -evoked calcium uptake. However, instead of exhibiting increased uptake, BD cortex showed decreased uptake from normal. This reduction in Ca uptake was a specific consequence of binocular deprivation as monocular lid closure had no effect on NMDA-stimulated calcium uptake in visual cortex. Furthermore, the reduced Ca uptake in slices from BD animals is not the same as that observed in slices from DR animals, because BD did not raise basal uptake from normal levels; the stimulated uptake was selectively reduced.

A number of mechanisms could contribute to the reduction in NMDA-evoked calcium uptake in striate cortex after brief binocular deprivation. For example, BD may reduce the number of NMDA receptors and/or channels, reflecting the loss of synaptic effectiveness thought to underlie the physiological consequences of BD (Freeman *et al.* 1983). In addition, because 1 mM  $Mg<sup>2+</sup>$  was present in our incubation media, the response to NMDA presumably depended upon the postsynaptic membrane potential. Hence, reduced calcium uptake could reflect decreased excitability in visual cortex of BD animals. It is also possible that BD causes a reduction in the number of voltage-sensitive calcium channels. This hypothesis is supported by our observation that  $K^+$ stimulated calcium uptake also appeared to be reduced in the cortex of BD kittens (although this change did not achieve statistical significance). Alternatively, since much of the 45Ca we measure in our slices is likely to be associated with intracellular stores, it is possible that BD produces a decrease in the effectiveness of intracellular calcium buffering mechanisms.

Regardless of the mechanisms involved, the data clearly show that a brief period of binocular deprivation is sufficient to cause significant changes in visual cortical calcium regulation. The relationship between these changes and the modification threshold  $(\theta_M)$  proposed by Bienenstock *et al.* (1982) remains unclear. The use of more refined methods is now warranted to investigate the mechanisms by which NMDA stimulated calcium uptake is changed, and to assess the impact of these changes on cortical function.

*Acknowledgements.* This work was supported by NIH grant NS 06 929, ONR contract N 00 014-81-K0136, and a Sloan Foundation fellowship. We thank K. Retz for helpful suggestions on the assay, H. Colman for technical assistance, and R. Neve and I. Reynolds for valuable criticism of the manuscript. We also thank L. Cooper for valuable discussions of all aspects of the study.

#### **References**

Anis NA, Berry SC, Burton NR, Lodge D (1983) The dissociative anesthetics, ketamine and phencyclidine, selectively reduce ex-

citation of central neurones by N-methyl-D-aspartate. Br J Pharmacol 79 : 565-575

- Artola A, Singer W (1987) Long-term potentiation and NMDA receptors in rat visual cortex. Nature 330:649-652
- Bear MF, Cooper LN, Ebner FF (1987) A physiological basis for a theory of synapse modification. Science 237 : 42-48
- Bear MF (1988) Involvement of excitatory amino acid receptors in the experience-dependent development of visual cortex. In: Cavalheiro EA, Lehman J, Turski L (eds) Frontiers in excitatory amino acid research. Liss AR, New York, pp 393-401
- Bear MF, Kleinschmidt A, Gu Q, Singer W (1989) Disruption of experience-dependent synaptic modifications in striate cortex by infusion of an NMDA receptor antagonist. J Neurosci (in press)
- Bienenstock EL, Cooper LN, Munro PW (1982) Theory for the development of neuron selectivity: orientation specificity and binocular interaction in visual cortex. J Neurosci 2:32-48
- Bode-Greuel KM, Singer W (1988) Developmental changes of the distribution of binding sites for organic  $Ca^{2+}$ -channel blockers in cat visual cortex. Exp Brain Res 70:266-275
- Bode-Greuel KM, Singer W (1989) The development of N-methyl-D-aspartate receptors in cat visual cortex. Dev Brain Res 46 : 197-204
- Bradford MM (1976) A rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein dye binding. Anal Biochem 72:248-254
- Collingridge GL, Kehl SJ, McLennan H (1983) Excitatory amino acids in synaptic transmission in the Schaffer collateralcommisural pathway of the rat hippocampus. J Physiol 334:33-46
- Conners BW, Bear MF (1988) Pharmacological modulation of long term potentiation in slices of visual cortex. Neuroscience Abstr 14:298.8
- Crowder J, Croucher M, Bradford H, Collins J (1987) Excitatory amino acid receptors and depolarization-induced  $Ca^{2+}$  influx into hippocampal slices. J Neurochem 48:1917- 1924
- Drapeau P, Blaustein MP (1983) Initial release of 3H dopamine from rat striatal synaptosomes: correlation with calcium entry. J Neurosci 3:703-713
- Freeman RD, Mallach R, Hartley S (1983) Responsivity of normal kitten striate cortex deteriorates after brief binocular deprivation. J Neurophysiol 45:1074-1084
- Frégnac Y, Imbert M (1984) Development of neuronal selectivity in the primary visual cortex of the cat. Physiol Rev 64: 325-434
- Gu Q, Bear MF, Singer W (1989) Blockade of NMDA receptors prevents ocularity changes in kitten striate cortex after reversed monocular deprivation. Dev Brain Res (in press)
- Harris EW, Ganong AH, Cotman CW (1984) Long-term potentiation involves activation of N-methyl-D-aspartate receptors. Brain Res 323 : 132-137
- Hubel DH, Wiesel TN (1962) Receptive fields, binocular interactions and functional architecture in the cat's visual cortex. J Physiol 160:106-154
- Hubel DH, Wiesel TN (1970) The period of susceptibility to the physiological effects of unilateral eye closure in kittens. J Physiol 206: 419-436
- Ichida S, Tokunaga H, Moriyama M, Oda Y, Tanaka S, Kita T (1982) Effects of neurotransmitter candidates on  $45Ca$  uptake by cortical slices of rat brain: stimulatory effect of L-glutamic acid. Brain Res 248 : 305-311
- Kauer JA, Malenka RC, Nicoll RA (1988) NMDA application potentiates synaptic transmission in the hippocampus. Nature 334:250-252
- Kimura F, Tsumoto T, Nishigori A, Shirokawa T (1988) Long-term synaptic potentiation and NMDA receptors in the rat pup visual cortex. Neuroscience Abstr 14:81.10
- Kimura F, Nishigori A, Shirokawa T, Tsumoto T (1989) Long-term potentiation and N-methyl-D-aspartate receptors in the visual cortex of young rats. J Physiol (in press)
- Kleinschmidt A, Bear MF, Singer W (1987) Blockade of "NMDA" receptors disrupts experience-dependent plasticity of kitten striate cortex. Science 238 : 355-358
- Lynch G, Larson J, Kelso S, Barrioneuvo S, Schlotter F (1983) Intracelluiar injections of EGTA block induction of hippocampal long-term potentiation. Nature 305 : 719-721
- MacDonald JF, Miljkovic Z, Pennefather P (1987) Use-dependent block of excitatory amino acid currents in cultured neurons by ketamine. J Neurophysiol 58:251
- Malenka RC, Kauer JA, Zucker RS, Nicoll RA (1988) Postsynaptic calcium is sufficient for potentiation of hippocampal synaptic transmission. Science 242: 81-84
- Mayer ML, Westbrook GL (1987) The physiology of excitatory amino acids in the vertabrate central nervous system. Prog Neurobiol 28 : 197-276
- Miller RJ (1987) Multiple calcium channels and neuronal function. Science 235 : 46-52
- Nowak L, Bregostovski, Ascher P, Herbert A, Prochiantz A (1984) Magnesium gates glutamate-activated channels in mouse central neurones. Nature 307:462-465
- Retz KC, Coyle JT (1984) The differential effects of excitatory amino acids on uptake of  ${}^{45}CaCl<sub>2</sub>$  by slices from mouse striatum. Neuropharmacology 23 : 89-94
- Reynolds IJ, Bear MF (1989) NMDA receptor development in the visual cortex of cats. Soc Neurosci Abs (in press)
- Riveros N, Orrego F (1986) N-methylaspartate-activated calcium channels in rat brain cortex slices: effect of calcium channel blockers and of inhibitory and depressant substances. Neuroscience 17 : 541-546
- Sherman S, Spear PD (1982) Organization of visual pathways in normal and visually deprived cats. Physiol Rev 62:738-855
- Singer W (1987) Activity-dependent self-organization of synaptic connections as a substrate of learning. In: Changeaux JP, Konishi M (eds) The neural and molecular basis of learning. John Wiley and Sons, New York, pp 301-336
- Singer W (1989) Ontogenetic self-organization and learning. In: McGaugh JL, Weinberger NM, Lynch G (eds) Brain organization and memory: cells, systems and circuits. Oxford University Press, New York (in press)
- Thibault O, Joly M, Muller D, Schottler F, Dudek S, Lynch G (1989) Long-lasting physiological effects of bath applied Nmethyl- D-aspartate. Brain Res 476:170-173
- Thompson AM, West DC, Lodge D (1985) An N-methylaspartate receptor-mediated synapse in rat cerebral cortex: a site of action of ketamine ? Nature 313 : 479-481
- Watkins JC, Evans RH (1981) Excitatory amino acid transmitters. Ann Rev Parmacol Toxicol 21 : 165-204

Received July 13, 1989 / Accepted October 16, 1989