ACUTE TOXICITY OF CHROME ELECTROPLATING WASTES TO MICROORGANISMS: ADSORPTION OF **CHROMATE AND** CHROMIUM(VI) ON A MIXTURE OF CLAY AND SAND

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Abstract. Chrome electroplating wastes were collected from two industrial sites and analyzed for color, turbidity, pH, alkalinity, sulfate, chloride, N-ammonia, N-nitrate, N-nitrite, acid hydrolyzable P, dissolved oxygen, biochemical oxygen demand, chemical oxygen demand, chromate and chromium(VI). The effect of these wastes on saprophytic and nitrifying bacteria was studied with varying concentrations of the waste using sucrose substrate as a source of C chain for microorganisms. The use of clay sand mixtures as adsorbents for chromate and chromium(VI) was investigated. Mixtures high in clay content were found to be suitable media for the removal of Cr from the wastes. The clay used was characterized by determination of the following parameters: pH, electrical conductivity, water soluble salts, osmotic pressure, cation exchange capacity, $CaCO₃$ and organic matter.

1. Introduction

It has been observed that the wastes from hard chrome plating are significantly more hazardous than from bright chrome plating (Guillemin and Berode, 1978). The chronic toxicity of Cr(VI) on fathead minow was investigated by Picketing (1980) and Smith and Heath (1979) studied acute toxicity of Cu, chromate, Zn and cyanide to five fresh water fish at different temperatures. Chromium(VI) is also associated with health hazards, and causes nausea and ulceration after long term exposures (McKee and Wolf, 1963; Camp, 1963). Cr can be removed from electroplating wastes by neutralization and electrocoagulation (Grin *et al.,* 1977) and Bornovolokov and Pushkarev (1979) have discussed the use of an electrochemical process for the removal of Cr. Banerjee (1976) described the treatment of effluent from the electroplating industry which can contain unacceptable quantities of KCN and chromates and which can cause corrosion to sewers and disrupt biological treatment. Ajmal *etaI.* (1980) have already reported deterimental effects of pharmaceutical industrial wastes on the microorganisms and it was thought to adopt a similar approach to the study of the hazards of chrome electroplating effluent. It was therefore decided to investigate the toxicity of these wastes to microorganisms and also to develop a cheap method for the removal of chromate and Cr(VI) from the effluents. This paper, therefore, reports toxicity of the electroplating wastes on the two types of bacteria and the removal of chromate and Cr(VI) by passing through columns containing sand: clay mixtures.

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2. Materials and Methods

2.1. LOCATION OF INDUSTRIES

Rainbow Electroplaters (RE) and Chandra Enterprise (CE) industries are situated in the district of Faridabad, Haryana, India. The effluent is dumped into the Gurgaon Canal which flows into the Yamuna River which passes through Indian capital, Delhi.

2.2. COLLECTION OF SAMPLES

The samples of electroplating wastes were collected in 300 mL BOD bottles and stored at 0 °C prior to the determination of physico-chemical characteristics (Standard Methods, 1975; Sawyer, 1960) using a Hach DR-EL/4 model spectrophotometer. A stock solution of sucrose (500 mg L^{-1}) was prepared in pure water (DMW) and used as C chain for microorganisms.

2.3 MICROORGANISMS

The culture of saprophytic microbial population was developed by continuous aeration with a daily increment of sewage (Standard Methods, 1975). The nitrifying bacteria were developed as suggested by Siddiqui *et al.* (1967).

2.4. PREPARATION OF COLUMNS

Soil was collected from the field and sieved through a 200 mesh to obtain pure clay which was then characterized by determination of related physico-chemical properties. The sand was collected from a river in the dry season. Glass columns of diameter 3.5 cm and length 105 cm were used in the adsorption experiments, a plug of glass wool being placed in the bottom of each column and the columns then packed to the height of approximate 80 cm with mixtures of clay and sand in the ratios $0:100$, $25:75$, $50:50$, 75 : 25, and 100 : 0. At first the columns were moistened with water and then 200 mL of each waste were poured in and allowed to stand for 24 hr. After 24 hr the waste was eluted at the rate of 4 drops min^{-1} and analyzed for chromate and Cr(VI).

2.5. PREPARATION OF BOD SETS

Varying volumes of the electroplating wastes were added to the following three systems separately.

- (a) SB + SS in 300 mL BOD bottle
- (b) NB + SS in 300 mL BOD bottle
- (c) $SB + NB + SS$ in 300 mL BOD bottle

where $SB =$ saprophytic bacteria; $NB =$ nitrifying bacteria; and $SS =$ sucrose substrate.

Finally the BOD bottles were filled upto the mark with 500 mg L^{-1} of sucrose substrate and kept for incubation for 15 days at 20 °C. The BODs were monitored at selected times over this period (Ciaccio, 1972).

3. Results and Discussion

The physico-chemical characteristics of RE and CE wastes are given in Table I. The wastes were found to be rich in sulfate, chloride, silica, N, P, chromate, and Cr(VI). The wastes were alkaline in nature and had low levels of dissolved oxygen (DO). The low BOD and COD are indicative of the inorganic nature of the wastes. High contents of chromate and Cr(VI) in these wastes led us to study their toxicity to microorganisms, and to develop inexpensive methods for their removal from the electroplating wastes prior to disposal.

Parameters	Rainbow Electro-	Chandra Enterprise (CE)	
	platers (RE)		
1. Color	1125 Pt-Co unit	875 Pt-Co unit	
	(yellowish)	(Pinkish)	
2. Turbidity	500 FTU	200 FTU	
3. pH	11.5	11.0	
4. Alkalinity	2390	2380	
5. Sulfate	200	320	
6. Chloride	680	800	
7. Silica	130	140	
$8. N-NH3$	24	10	
9. N-NO ₂	80	80	
10. $N-NO2$	4.8	ND.	
11. P(reactive)	ND	ND	
12. P(organic)	ND	ND	
13. P(hydrolyzable)	24	90	
14. DO	2.5	2.5	
15. BOD	9	9.5	
16. COD	14	12	
17. Chromate	320	170	
18. $Cr(VI)$	400	200	

TABLE I Physico-chemical characteristics of electroplating wastes

All units except color, turbidity, and pH are expressed in mg L^{-1} .

The results of the investigation of the toxicity of RE and CE wastes to the microorganisms are presented in the Tables II and III. The BOD of sucrose substrate with saprophytic bacteria having no waste (control) showed a rapid stabilization. It is worth reporting that 71, 76, 83, and 98% of the BOD was observed on 5th, 8th, 10th, and 15th day of incubation in the control sets of saprophytic bacteria. Similarly 75 to 99 $\%$ and 77 to 99% were observed in case of nitrifying bacteria and mixture of the two bacteria (saprophytic + nitrifying), respectively. It was observed that 0.5 mL of the RE and CE wastes showed weak toxic effects in repressing the BOD of the system on 5th to 15th days as compared to control. But a pronounced repression of the activity of saprophytic bacteria started with 2 mL of RE waste and increased with an increasing volume of waste (Figure 1). Twelve mL of the waste retarded all microbial activity. The BOD was

 $\%$ inhibition $\frac{1}{1}$ $\frac{3}{1}$ $\frac{3}{8}$ $\frac{6}{4}$ $\frac{5}{1}$ $\frac{3}{1}$ $\frac{6}{1}$ $\frac{5}{1}$ $\frac{1}{2}$
 $\frac{0}{2}$
 $\frac{2}{2}$
 $\frac{0}{2}$
 $\frac{1}{4}$
 $\frac{0}{4}$ 74.7
81.9 -11.1
 $13.3.3.6$ 74.7
81.9 8853737 $RRRRMR$ \mathcal{S}° a sa ta ta ta sa 5th day $mg L^{-1}$ 28881 32725528 39728528 inhibition -4.57 $\frac{31}{31}$
 $\frac{31}{21}$
 $\frac{31}{21}$
 $\frac{31}{21}$
 $\frac{31}{21}$ 13.9
 25.5
 47.2
 79.2 $1, 7, 9, 0, 0, 0, 4, 6, 6, 7, 8, 7$ \sim ន្លួន្គប្តូន្ត \mathcal{S}° 88773732 2222222 .0th day $\rm mg\,L^{-1}$ \$\$\$\$\$1 **FORTESSES** 224488228 inhibition $\frac{1}{6}$ $\frac{1}{2}$ $\frac{1}{2}$ $\frac{1}{2}$ $\frac{0}{2}$ $\frac{0}{2}$ 71.4 -7.5
 18.0
 51.3
 7.5
 7.5
 78.7 \sim \sim° $RF5888$ PREBRAL 2787222 $mg L^{-1}$ 8th day 3233331 $#752225$ 7222728 $\%$ inhibition -3.33328 -1.6
 1.6
 1.5
 3.6
 3.3
 7.3
 8.3 12.9
124845762 **BOD** on given days \sim FSZ\$RI **KERGARAN** maxexaz $mg L^{-1}$ 5th day 8782528 22222222 **HEATSSH** $S = +NB + SS + 0.5$
 $S = +NB + SS + 2$
 $S = +NB + SS + S$
 $S = +S + S$ \bullet $SB + NB + SS + 10$
 $SB + NB + SS + 12$ $SS + NB + SS +$
 $S = + SI + SS +$ $NB + SS + 0.5$
NB + SS + 2
NB + SS + 5
NB + SS + 8 $S1 + S3 + 0.5$
 $S1 + S5 + 2$
 $S1 + S5 + 5$
 $S1 + S5 + 8$
 $S1 + S5 + 8$
 $S1 + S5 + 10$ $NB + SS + 10$
 $NB + SS + 12$ \circ \bullet Experimental $NB + SS +$ $-5S + 8S$ (mL)

 $\%$ inhibition compared to the BOD of control of corresponding day.

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TABLE III

BOD studies showing toxicity of CE waste on microorganisms

 $\%$ inhibition 12.2
 21.4
 31.4
 47.4
 78.8 $\frac{133}{230.7}$ 6.9 **19.5** $-0.271.54$
 $-0.277.54$ 15.9
 77.5 $\overline{5}$ \mathbf{L} \mathbf{I} $\frac{1}{6}$ 88558888 85885733 $R522222$ Sth day mgL^{-1} **ARAQUE ESSES** $\frac{2}{6}$ inhibition -5.3
 -5.3
 -5.3
 -5.3 -1.7
 -2.5
 -2.5
 -3.4
 -0.6
 -4.5 76.1 $\frac{28}{1}$ $\frac{28}{1}$ $\frac{33}{1}$ $\frac{35}{1}$ $\frac{37}{1}$ $\frac{36}{1}$ $\frac{37}{1}$ $\frac{36}{1}$ $\frac{37}{1}$ \sim EXPREZE 222222222 ******* oth day $mg L^{-1}$ $= 5.492728$ 22222222 %inhibition -141
 784
 194
 313 44.31 55.9 -0.5
 -0.7
 -0.3
 -0.7
 -0.3
 -0.3
 -0.3
 -0.0 45.4 9.0
 8.3 32.4 34.5 $\frac{3}{1}$ RRRG582 \sim FFF8SS# **FRREERSHE** $mg\,L^{-1}$ 8th day 4772728728 7422222222222 ******* $\overline{5}$ $\%$ inhibition -257
 -177
 -177
 -179
 -59
 -89 -1.4777282 11.7
 23.1
 34.1 51.2 69.8 83.0 **BOD** on given days $\mathbf{I} = \mathbf{I}$ $RF88572$ \sim **ARGESSENT** RRESSERT $mg L^{-1}$ 5th day 3582382 277272288 33588582 $S\overline{B} + NB + SS + 0.5$
 $S\overline{B} + NB + SS + 2$
 $S\overline{B} + NB + SS + 5$
 $S\overline{B} + SS + SS + 8$ \circ $SS + 20$ $SB + NB + SS + I2$ $+$ SS + 25 $SS + NB + SS +$
 $+ SS + BS +$ 0.5 0.5 $NB + SSS + 2$
 $NB + SSS + 5$
 $NB + SSS + 8$
 $NB + SSS + 12$ $NB + SS + 20$
 $NB + SS + 25$ $SB + SS + 12$
 $SB + SS + 20$ \bullet \sim 0 ∞ \circ Experimental $S = S + S$ $SB + SS +$ $SS + SS +$ $NB + SS +$ $+$ SS $S = 1 + S$ $NB + SS +$ $SB + NB +$ conditions $SB + NB$ $\frac{1}{4}$ \overline{a} $_{\rm SB}$

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 $\%$ inhibition compared to the BOD of control of corresponding day.

Fig. 1. BOD as a function of days of incubation.

repressed upto 14, 20, 20, and 21 $\%$ on 5th, 8th, 10th, and 15th day in the sets containing 10 mL of RE waste. In general the percent inhibition decreased with the increase in the time of incubation, whereas the inhibition increased with the increase in the volume of waste. The inhibition in the sets of saprophytic bacteria ranged from 5.9 to 79.8 $\%$ on the 5th day and 13.1 to 79.5% on the 15th day of incubation. Similarly the same volumes of the waste were added for nitrifying bacteria but in this case inhibition in bacterial activity was only observed when 15 mL of waste was used. The same volumes of the wastes were added to the mixture of saprophytic and nitrifying bacteria and it was found that only when 15 mL of the waste had been added the microbial activity was retarded.

The studies conducted with the CE waste under the identical conditions showed similar results to those with RE waste but the volume of waste which inhibited the microbial activity of saprophytic, nitrifying and a mixture of both were 25, 30, and 30 mL, respectively. The reason for this high volume required for the inhibition may be attributed to the low concentration of chromate and Cr(VI) in the CE waste (Table I).

It is clear from the percent inhibition data (Tables II and III) that the nitrifying bacteria can mildly tolerate effects of wastes as compared to the saprophytic bacteria. In the sets of saprophytic bacteria with 10 mL of RE waste on 5th, 8th, 10th, and 15th day the percent inhibition observed was 79.8, 73.6, 75.8, and 79.5 $\frac{9}{6}$ as compared to the 76.3, 71.4, 72.1, and 74.4 $\%$ of corresponding days in the sets of nitrifying bacteria. A similar trend was found in the sets of CE waste.

The physico-chemical analysis of the clay is presented in Table IV. The cation exchange capacity of the clay ranged between 192.4 and 195.5 meq per 100 g of clay determined as per Ganguly (1952). The clay used for the studies had pH 7.9 to 8.0, electrical conductivity 3.50 \times 10⁻⁴ to 4.0 \times 10⁻⁴ mhos cm⁻¹, osmotic pressure 0.79 to 0.82, calcium carbonate 8.14 to 10.7% and organic matter 0.84 to 0.86%. The soil used was low in organic matter and also had a low cation exchange capacity (Alexander, 1961). The eluted wastes (RE and CE) from different columns were analysed and compared with the initial chromate and Cr(VI) concentrations (Table V). The first column (clay: sand, $0:100$) showed slight adsorption of chromate and Cr(VI). However, the adsorption increased with an increasing proportion of clay. The fifth column filled with pure clay adsorbed nearly 95.6 to 96 $\%$ chromate and 99 to 99.2 $\%$ Cr(VI) (Table V). The eluant so obtained was free from color, chromate and Cr(VI). So the clay appears to provide a good and cheap means for the removal of chromate and Cr(VI) from electroplating waste before discharge to the sewer or onto the land.

Chromate and $Cr(VI)$ before adsorption	Adsorbed chromate and $Cr(VI)$ (clay: sand)					
	0:100	25:75	50:50	75:25	100:0	
RE waste (i)						
Chromate (320 mg L^{-1})	6.4(2)	96(30)	214 (67)	265 (83)	307 (96)	
$Cr(VI)$ (400 mg L ⁻¹)	7.6(1.9)	140 (35)	304 (76)	356 (89)	396 (99)	
(ii) CE waste						
Chromate (170 mg L^{-1})	3.06(1.8)	52.7 (31)	113 (67)	141 (83)	161 (95)	
$Cr(VI)$ (200 mg L ⁻¹)	3.00(1.5)	69.4 (35)	152 (76)	178 (89)	98 (99)	

TABLE V Adsorption of chromate and Cr(VI) on clay : sand in five different ratios for 24 hr

Values in parenthesis shows percent adsorption.

4. Conclusion

These studies clearly indicated that the electroplating waste which contains chromate and Cr(VI) contents is inimical to the saprophytic and nitrifying bacteria. RE waste was much toxic as it contained higher concentration of chromate and Cr(VI) in comparison to CE waste. Clay proved to be a good absorpent for the chromate and Cr(VI) due to high cation exchange capacity and strong binding force. This needs further research as it may be proved to be cheapest treatment of electroplating wastes in the forth coming future.

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References

Ajmal, M., Ahmad, A., Hasan, M. Z., and Nomani, A. A.: 1980, *Water, Air, and Soil Pollut.* 13, 447.

Alexander, M.: 1961, *Introduction to Soil Microbiology,* John Wiley and Sons, Inc., New York.

Banerjee, R. K.: 1976, *Ind. Chem. Manuf* 14, 43.

Bornovolokov, P. P. and Pushkarev, V. V.: 1979, *Stroit Mater.* 5, 19.

Camp, T. R.: 1963, *Water and its Impurities,* Reinhold, New York.

Ciaccio, L. L.: 1972, *Water and Water Pollution Hand Book*, Marcel Dekker, Inc., New York, Vol. 3.

Ganguly, A. K.: 1952, *Y. Phys. Colloid Chem.* 55, 1417.

Grin, N. V., Solovev, V. I., and Glushko, L. S.: 1977, *Gig Sanit* 6, 97.

Guillemin, M, P. and Berode, M.: 1978, *Ann. Occup. Hyg.* 21, 105.

McKee, J. E. and Wolf, H. W.: 1963, *Water Quality Criteria,* Publication 3A, California State Water Resources Control Board, Saerameuto, Calif.

Pickering, Q. H.: 1980, *Archives of Environmental Contamination and Toxicology* 9, 405.

Sawyer, C. N.: 1960, *Chemistry for Sanitary Engineers,* McGraw Hill Book Company, Inc., New York.

Siddiqui, R. H., Specce, R. E., Engelbrecht, R. S., and Schmidt, J. W.: 1967, *J.W.P.C.F.* 39, 579.

Smith, M. J. and Heath, A. G.: 1979, *Bull. Environ. Contam. Toxicol.* 22, 113.

Standard Methods for Examination of Water and Waste Water: 1975, 14th edn., Am APHA, AWWA, and WPFC.