

## Molecular Evolution of the Nicotinic Acetylcholine Receptor: An Example of Multigene Family in Excitable Cells

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**Abstract.** An extensive phylogenetic analysis of the nicotinic-acetylcholine-receptor subunit gene family has been performed by cladistic and phenetic methods. The conserved parts of amino acid sequences have been analyzed by CLUSTAL V and PHYLIP software. The structure of the genes was also taken in consideration. The results show that a first gene duplication may have occurred before the appearance of Bilateria. Three subfamilies then appeared: I—the neuronal  $\alpha$ -bungarotoxin binding-site subunits ( $\alpha 7$ ,  $\alpha 8$ ); III—the neuronal nicotinic subunits ( $\alpha 2$ – $\alpha 6$ ,  $\beta 2$ – $\beta 4$ ), which also contain the muscle acetylcholine-binding subunit ( $\alpha 1$ ); and IV—the muscle non- $\alpha$  subunits ( $\beta 1$ ,  $\gamma$ ,  $\delta$ ,  $\epsilon$ ). The Insecta subunits (subfamily II) could be orthologous to family III and IV. Several tissue switches of expression from neuron to muscle and the converse can be inferred from the extant expression of subunits and the reconstructed trees. The diversification of the neuronal nicotinic subfamily begins in the stem lineage of chordates, the last duplications occurring shortly before the onset of the mammalian lineage. Such evolution parallels the increase in complexity of the cholinergic systems.

**Key words:** Nicotinic receptor — Ligand-gated channel — Multigene family — Gene phylogeny

### Introduction

Acetylcholine (ACh) has long been recognized as a neurotransmitter active in Bilateria nervous system and muscle. Two distinct categories of receptors are engaged in the biological effects of ACh: the muscarinic and nicotinic receptors. Muscarinic receptors belong to the superfamily of G-protein-coupled receptors; they consist of single integral proteins with seven transmembrane segments and interact, on their cytoplasmic face, with heterotrimeric G-proteins. Nicotinic receptors (nAChR) belong to the superfamily of ligand-gated ion channels; they are hetero-oligomers composed of five subunits, each with four transmembrane domains (Devillers-Thiéry et al. 1993; Galzi and Changeux 1994). ACh binding causes an ionic channel, most often cationic, to open, resulting in a rapid change in the electrical, and secondarily metabolic, state of the target cell (Greenberg et al. 1986; Bertrand et al. 1993).

The nAChR of striated muscle is the best-characterized member of the ligand gated-ion-channel superfamily (Changeux 1990; Karlin 1993): it is a heteropentamer (with the stoichiometry  $\alpha_2\beta\gamma\delta$ ). According to current models (Bertrand et al. 1993), the ion channel forms along the axis of pseudosymmetry perpendicular to the cell membrane. The subunits share a similar hydrophobic profile with four short hydrophobic domains (MI–MIV) and two long hydrophilic domains. The largest, relatively conserved, hydrophilic domain is located at the amino-terminal side of the subunit polypeptide, and the other, highly variable, joins hydrophobic do-

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Abbreviations:  $\alpha$ -bungarotoxin ( $\alpha$ -Bgt), acetylcholine (ACh), maximum of parsimony (MP), million years ago (MYA), neighbor-joining (NJ), nicotinic acetylcholine receptor (nAChR)

mains MIII and MIV. The amino-terminal hydrophilic domain carries the ACh binding site (Devillers-Thiéry et al. 1993) and faces the synaptic cleft, whereas the other hydrophilic domain is exposed to the cytoplasm.

Molecular cloning and sequencing studies have revealed the existence of a family of genes, expressed in neurons, which code for nAChR subunits **homologous** (see Appendix 1 for definitions of boldface terms) to those of muscle nAChR (Sargent 1993). In the jawed vertebrate nervous systems, several subunits (named  $\alpha 2-\alpha 8$ ) have been identified which share with the muscle-type receptor  $\alpha 1$  subunit the pair of cysteines shown to contribute to the ACh binding site (Wada et al. 1988; Shoepfer et al. 1990; Cockcroft et al. 1992; Karlin 1993). Other homologous chains, lacking the cysteine pair, have been characterized and named non- $\alpha$  or  $\beta 2-\beta 4$ . As for muscle nAChR, the functional neuronal nAChR is an heteropentamer made up by the assembly of  $\alpha$  and  $\beta$  subunits, with a putative stoichiometry in vitro of  $\alpha_2\beta_3$  (Anand et al. 1991; Cooper et al. 1991). The recent evidence that  $\alpha 5$  is coprecipitated with another  $\alpha$  and  $\beta$  subunit in some neuronal nAChRs indicates that more than two different subunits may assemble together to form a receptor molecule (Conroy et al. 1992; Vernallis et al. 1993). In contrast, in reconstituted systems, the  $\alpha 7$  or  $\alpha 8$  subunits can form functional homo-oligomers (Couturier et al. 1990; Revah et al. 1991; Anand et al. 1993). The autoradiographic studies in the brain revealed that  $^3\text{H}$ -nicotine labels receptors formed by subunits  $\alpha 2-\alpha 6$  and  $\beta 2-\beta 4$  but not receptors formed by subunits  $\alpha 7$  and  $\alpha 8$  (Clarke et al. 1985), which are labeled by  $\alpha$ -bungarotoxin ( $\alpha$ -Bgt).

The combinatorial diversity resulting from the assembly of the multiple neuronal subunits results in a wide spectrum of structurally and functionally distinct nAChRs with different pharmacological specificities and ion-channel properties (Role 1992). Such differences have been directly demonstrated in *Xenopus* oocytes and mouse fibroblasts after heterologous expression (Luetje and Patrick 1991; Whiting et al. 1991). Furthermore, multiple functionally distinct types of nAChRs have been detected in different brain areas and subcellular compartments (Mulle et al. 1991).

The nAChR is present in the whole phylum of Bilateria, from nematodes to humans (Gerschenfeld 1973; Darlison et al. 1993; Fleming et al. 1993; Leech and Sattelle 1993). Several nAChR neuronal subunits have been cloned in *Drosophila*, locust, and nematode (Gundelfinger 1992; Fleming et al. 1993). In the insect nervous system, ACh is the major excitatory neurotransmitter, in contrast to vertebrates, where glutamate predominates. At the neuromuscular junction, glutamate is the excitatory transmitter in arthropods, whereas it is ACh in vertebrates. Since some lines of evidence suggest that nAChR is also responsible for neuromuscular transmission in nematodes, molluscs, and annelids (Gerschenfeld 1973; Segerberg and Stretton 1993), it is of interest

to assess whether the original form of nAChR appeared in muscle or in neurons.

The neuronal nAChRs provide a good example of a multigene family differentially expressed in the nervous system. Its evolution deserves comparison with the increase in complexity of the vertebrate nervous system and, in particular, cholinergic systems. Some partial trees have already been constructed, but without comparative methods and without statistical support (Brehm et al. 1991; Cockcroft et al. 1992). Here we provide a molecular phylogenetic study of the whole family of nAChR genes.

## Materials and Methods

The programs used in this work were run on a Sun computer in a UNIX environment. The sequences were loaded from Genbank and EMBL databases (Table 1) with Sequence Analysis Software Package 7.1 from the Genetic Computer Group.

*Alignments of the Sequences.* Alignments were performed using CLUSTAL V software (Higgins and Sharp 1988). This program compares the sequences in pairs according to Wilbur and Lipman (1983) (gap penalty = 3) and builds a preliminary tree by an unweighted pair-group method of arithmetic averages (UPGMA) (Sneath and Sokal 1973). Then the program aligns all sequences in order of decreased similarity according to Feng and Doolittle (1987) (fixed and floated gap penalty = 10). The use of different values of gap cost changed neither topology nor the ratio of branch lengths but did result in a homothetic transformation of the trees. The similarities have been determined by the Dayhoff PAM 250 matrix. The protein sequences were aligned after the following modifications:

Deletions of the signal peptide (corresponding to  $\alpha 1$  aa 1–27), the small nonconserved part in amino-terminal part (corresponding to  $\alpha 1$  R188), the highly variable cytoplasmic region (corresponding to  $\alpha 1$  aa 356–393), and the carboxy-terminal part (corresponding to  $\alpha 1$  aa 452–461). The alignment obtained with 48 sequences shows 394 sites with 357 **informative sites** (Appendix 2).

To determine the branching of the nematode sequence onachr (Fig. 4) which amino-terminal part is not known, a further deletion (corresponding to the 38 amino-terminal aa of  $\alpha 8$  from the Appendix 2 alignment) was performed on 12 sequences. The alignment of the 13 sequences shows 351 sites with 263 informative sites (Appendix 3).

*Sequence Analyses.* Inferences on gene evolution were obtained with the PHYLP 3.5c software of Felsenstein (1993).

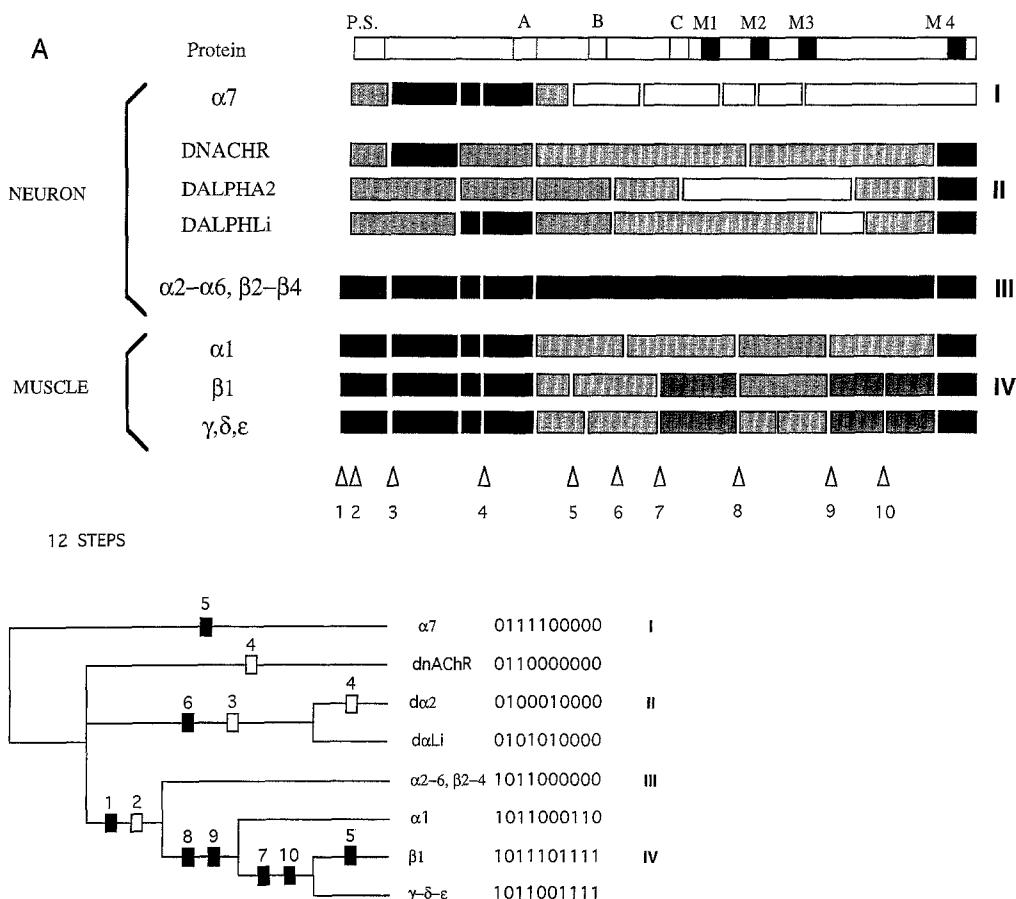
The cladistic method was the maximum of parsimony (MP) (Fitch 1971, program PROTPARS). The mouse 5-HT3 subunit and the rat glycine  $\alpha 3$  subunit were used as outgroups. The use of the rat GABA  $\alpha 1$  subunit instead of the glycine  $\alpha 3$  subunit did not change the results (data not shown). The phenetic method was neighbor-joining (NJ) (Saitou and Nei 1987, program NEIGHBOR). The distance matrix was provided by the Dayhoff PAM matrix (Dayhoff 1979, program PROTDIST). The statistical test used to determine the strength of the trees was bootstrap resampling (Felsenstein 1985) with the SEQBOOT (seed: 5) and CONSENSE programs.

*Analyses of Gene Structure.* The mixed-parsimony algorithm with the Wagner method (Eck and Dayhoff 1966, program MIX) and the

**Table 1.** Genes used in this study. Abbreviations are those used in the text and trees. The first letters represent the species, followed by the name of the subunit<sup>a</sup>

Gene	Species	Acc. no.	Ref.
b $\alpha$ 1	<i>Bos taurus</i>	X02509	Noda et al. <i>Nature</i> 305:818 (1983)
b $\beta$ 1	<i>Bos taurus</i>	X00962	Tanabe et al. <i>Eur J Biochem</i> 144:11 (1984)
b $\delta$	<i>Bos taurus</i>	X02473	Kubo et al. <i>Eur J Biochem</i> 149:5 (1985)
b $\epsilon$	<i>Bos taurus</i>	X02597	Takai et al. <i>Nature</i> 315:761 (1985)
b $\gamma$	<i>Bos taurus</i>	M28307	Takai et al. <i>Eur J Biochem</i> 143:109 (1984)
c $\alpha$ 2	<i>Gallus domesticus</i>	M07339-44	Nef et al. <i>EMBO J</i> 7:595 (1988)
c $\alpha$ 3	<i>Gallus gallus</i>	M37336	Couturier et al. <i>JBC</i> 265:17560 (1990)
c $\alpha$ 4	<i>Gallus domesticus</i>	X07348-53,99	Nef et al. <i>EMBO J</i> 7:595 (1988)
c $\alpha$ 5	<i>Gallus gallus</i>	J05642	Couturier et al. <i>JBC</i> 265:17560 (1990)
c $\alpha$ 7	<i>Gallus gallus</i>	X68586	Couturier et al. <i>Neuron</i> 5:847 (1990)
c $\alpha$ 8	<i>Gallus gallus</i>	X52296	Schoepfer et al. <i>Neuron</i> 5:35 (1990)
c $\beta$ 2	<i>Gallus domesticus</i>	X53092	Schoepfer et al. <i>Neuron</i> 1:241 (1988)
c $\beta$ 4	<i>Gallus gallus</i>	J05643	Couturier et al. <i>JBC</i> 265:17560 (1990)
c $\delta$	<i>Gallus gallus</i>	K02903	Nef et al. <i>PNAS</i> 81:7975 (1984)
c $\gamma$	<i>Gallus gallus</i>	K02904	Nef et al. <i>PNAS</i> 81:7975 (1984)
d $\alpha$ Li	<i>Drosophila melanogaster</i>	X07194	Bossy et al. <i>EMBO J</i> 7:611 (1988)
d $\alpha$ 2	<i>Drosophila melanogaster</i>	X53583	Sawruk et al. <i>EMBO J</i> 9:2671 (1990)
d $\beta$ 2	<i>Drosophila melanogaster</i>	X55676	Sawruk et al. <i>FEBS Lett</i> 273:177 (1990)
d $\alpha$ chr	<i>Drosophila melanogaster</i>	X04016	Hermans-Borgmeyer et al. <i>EMBO J</i> 5:1503 (1986)
g $\alpha$ 3	<i>Carassius auratus</i>	X54051	Hieber et al. <i>NAR</i> 18:5293 (1990)
g $\beta$ 2	<i>Carassius auratus</i>	X54052	Hieber et al. <i>NAR</i> 18:5307 (1990)
g $\alpha$ 2	<i>Carassius auratus</i>	X14786	Cauley et al. <i>J Cell Biol</i> 108:637 (1989)
g $\alpha$ 3	<i>Carassius auratus</i>	M29529	Cauley et al. <i>J Neurosci</i> 10:670 (1990)
h $\alpha$ 1	<i>Homo sapiens</i>	Y00762	Schoepfer et al. <i>FEBS Lett</i> 226:235 (1988)
h $\alpha$ 3	<i>Homo sapiens</i>	M37981	Mihovilovic et al. <i>J Exp Neurol</i> 111:175 (1991)
h $\alpha$ 5	<i>Homo sapiens</i>	M83712	Chini et al. <i>PNAS</i> 89:1572 (1992)
h $\alpha$ 7	<i>Homo sapiens</i>	X70297	Peng et al. <i>Mol Pharmacol</i> 45:546 (1994)
h $\beta$ 1	<i>Homo sapiens</i>	X14830	Beeson et al. <i>NAR</i> 17:4391 (1989)
h $\beta$ 2	<i>Homo sapiens</i>	X53179	Anand et al. <i>NAR</i> 18:4272 (1990)
h $\beta$ 3	<i>Homo sapiens</i>		Willoughby et al. <i>Neurosci Lett</i> 155:136 (1993)
h $\beta$ 4	<i>Homo sapiens</i>	X68275	Tarroni et al. <i>FEBS Lett</i> 312:66 (1992)
h $\delta$	<i>Homo sapiens</i>	X55019	Luther et al. <i>J Neurosci</i> 9:1082 (1989)
h $\epsilon$	<i>Homo sapiens</i>	X66403	Beeson et al. unpublished
m $\alpha$ er	<i>Mus musculus</i>	M74425	Maricq et al. <i>Science</i> 254:432 (1991)
n $\alpha$ 1	<i>Naja naja</i>	M26388	Neumann et al. <i>PNAS</i> 86:7255 (1989)
o $\alpha$ chr	<i>Onchocerca volvulus</i>	L20465	Ajuh and Egwang unpublished (1993)
r $\alpha$ 2	<i>Rattus norvegicus</i>	L10077	Wada et al. <i>Science</i> 240:330 (1988)
r $\alpha$ 3	<i>Rattus norvegicus</i>	X03440	Boulter et al. <i>Nature</i> 319:368 (1986)
r $\alpha$ 4	<i>Rattus norvegicus</i>	M15681-82	Goldman et al. <i>Cell</i> 48:965 (1987)
r $\alpha$ 5	<i>Rattus norvegicus</i>	J05231	Boulter et al. <i>J Biol Chem</i> 265:4472 (1990)
r $\alpha$ 6	<i>Rattus norvegicus</i>	L08227	Boulter unpublished (1988)
r $\alpha$ 7	<i>Rattus norvegicus</i>	M85273	Seguele et al. <i>J Neurosci</i> 13:596 (1993)
r $\beta$ 2	<i>Rattus norvegicus</i>	—	Deneris et al. <i>Neuron</i> 1:45 (1988)
r $\beta$ 3	<i>Rattus norvegicus</i>	J04636	Deneris et al. <i>J Biol Chem</i> 264:6268 (1989)
r $\beta$ 4	<i>Rattus norvegicus</i>	J05232, M89971, M33951-3, M89989	Boulter et al. <i>J Biol Chem</i> 265:4472 (1990)
r $\delta$	<i>Rattus norvegicus</i>	X74835	Witzemann et al. <i>Eur J Biochem</i> 194:437 (1990)
r $\epsilon$	<i>Rattus norvegicus</i>	X13252	Criado et al. <i>NAR</i> 16:10920 (1988)
r $\gamma$	<i>Rattus norvegicus</i>	X74834	Witzemann et al. <i>Eur J Biochem</i> 194:437 (1990)
r $\text{gly}\alpha$ 3	<i>Rattus norvegicus</i>	M55250	Kuhse et al. <i>J Biol Chem</i> 265:22317 (1990)
s $\alpha$ L1	<i>Schistocerca gregaria</i>	X55439	Marshall et al. <i>EMBO J</i> 9:4391 (1991)
t $\alpha$ 1	<i>Torpedo californica</i>	X00963	Noda et al. <i>Nature</i> 299:793 (1982)
t $\beta$ 1	<i>Torpedo californica</i>	J00964	Noda et al. <i>Nature</i> 301:251 (1983)
t $\delta$	<i>Torpedo californica</i>	J00965	Noda et al. <i>Nature</i> 301:251 (1983)
t $\gamma$	<i>Torpedo californica</i>	J00966	Ballivet et al. <i>PNAS</i> 79:4466 (1982)
x $\alpha$ 1a	<i>Xenopus laevis</i>	X17244	Hartman et al. <i>Nature</i> 343:372 (1990)
x $\alpha$ 1b	<i>Xenopus laevis</i>	X07067	Baldwin et al. <i>J Cell Biol</i> 106:469 (1988)
x $\beta$ 1	<i>Xenopus laevis</i>	U04618	Kullberg et al. <i>Rec Chan</i> (1994) in press
x $\delta$	<i>Xenopus laevis</i>	X07069	Baldwin et al. <i>J Cell Biol</i> 106:469 (1988)
x $\gamma$	<i>Xenopus laevis</i>	X07068	Baldwin et al. <i>J Cell Biol</i> 106:469 (1988)

<sup>a</sup> The six sign codes are the accession numbers of the Genbank-Embl databases. Rat  $\alpha$ 4 is the isoform 4-2



**Fig. 1.** **A** Structure of the subunit genes. Only the exons at least partially coding are represented. The gray level grossly reflects the conservation of the exon through the family (i.e., the presence of an exonic frontier at this place in different subunits, but not the sequence similitude between exons). **A**, **B**, **C**: binding site loops. **M1**, **M2**, **M3**, **M4**: transmembrane segments. The arrowheads mark the informative limits (in a cladistic acceptance). Adapted from Jonas et al. (1990) with

the help of Alain Bessis. **B** Cladogram constructed with the exonic structure of the genes from an MP analysis which gave three equivalent trees. The informative limit of each gene is coded at the right of its name (0: absence; 1: presence). Open box: loss of a limit; filled box: gain of a limit. The dendrogram is arbitrarily rooted. The branch lengths make no sense.

compatibility method (Le Quesne 1969; Estabrook et al. 1976, program CLIQUE) were used to analyze the genomic structure.

**Construction of Figures.** The majority-rule consensus trees were constructed by the program DRAWGRAM. The results of CONSENSE analysis after the bootstrap resamplings are written in ovals on the node considered.

The determination of approximated time divergence between subunits (Fig. 5) is based on the NJ analysis of the Appendix 2 protein alignment. The external branches of the resulting tree showed an approximate molecular clock for each group. We are then able to determine the dates of the last duplications. However, the rates of evolution vary greatly between the subgroups, and the precocious duplications can't be calculated in this way. To determine the date of divergence between two subunits, we averaged the branch lengths and the evolution rate between all the **orthologs**. The estimated rate of evolution is obtained by dividing the branch length by the duration. The dates used are: *Torpedo*/osteichthyans 450 MYA, goldfish/Tetrapoda 405 MYA, *Xenopus*/Amniota 365 MYA, chicken/mammals 310 MYA, mouse/human 110 MYA (Benton 1990). For instance, each external branch inside the  $\delta$ ,  $\epsilon$ ,  $\gamma$  group provides an estimated rate of evolution of between  $3.2 \times 10^{-10}$  and  $7.3 \times 10^{-10}$  substitution per site and per year ( $M = 5.3 \sigma = 1.2$ ), which is not far from a molecular clock.

## Results

### Evolution of the Gene Structure

The number of exons identified in the gene nAChR subunits varies largely in the family although some common features can be recognized (Fig. 1A). Four subfamilies can be identified on the basis of the genomic structure. These are: I—the *neuronal  $\alpha$ -bungarotoxin-binding-site subunit* subfamily; II—the *Arthropoda neuronal subunit* subfamily; III—the *vertebrate neuronal nicotinic subunit* subfamily; and IV—the *muscle subunit* subfamily.

The genes of subfamily IV possess 11 or 12 exons, of which ten are conserved. In subfamily III, the main part of the coding sequence is distributed within a single exon. The structure of  $\alpha_1$  and  $\alpha_7$  genes differs from the two "holotypes" (III or IV). However, it is difficult to determine if the structure of these two genes is mainly **plesiomorph** or contains **autapomorphies**. In order to

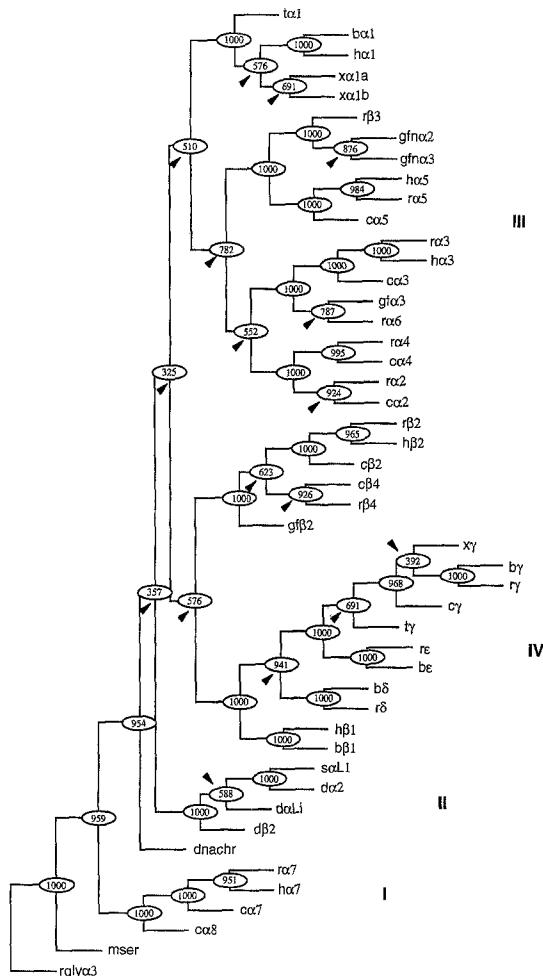
extract information from genomic structure, we made a cladistic analysis of the frontiers between introns and exons. Only the ten informative sites were considered (Fig. 1). A parsimony analysis of two-state character (i.e., presence or absence of the frontier) gave three 12-step cladograms (summarized in Fig. 1B with an arbitrary root corresponding to sequence analyses; see below). A compatibility analysis gave the same results (although automatically rooted at a different point, i.e., between muscle and neuronal genes). One explanation is that the genic structure of type I subunits is mainly made of autapomorphies, whereas that of  $\alpha 1$  is plesiomorphic in subfamily IV. On the basis of gene-structure analysis, this latter subunit would be a sister group of all other subunits of subfamily IV. However, sequence analyses (see below) make  $\alpha 1$  a sister group of subfamily III. A translocation (e.g., between exons 4 and 5) could mask the real onset of the  $\alpha 1$  subunit. Further studies of sequence homology between exons in **paralogs** will help to clarify this issue.

## *Sequences Analysis*

The sequence analysis revealed the existence of the same four subfamilies of nAChR subunits as did analysis of the gene structure (Figs. 2 and 3). We obtained successive divergences of subfamilies I, then II and, at last III and IV. The position of subfamily II as a sister group of subfamily IV is weakly supported by NJ (Fig. 3) analysis and not by MP analysis (Fig. 2). These subunits appeared **polyphyletic** with MP analysis and **monophyletic** with NJ analysis. However, their position was supported by very weak bootstrap score. The  $\beta 2$  and  $\beta 4$  subunits were branched with subfamily IV with a weak bootstrap score. This position may be an artifact resulting from the precocious appearance and the weak divergence of these two subunits.

In subfamily IV, the  $\beta 1$  subunit diverged first followed by the  $\delta$ . A subsequent duplication resulted in the  $\gamma$  and  $\epsilon$  subunits. This latter duplication seems to have occurred shortly before the divergence of *Torpedo* subunits (i.e., before the divergence of the elasmobranch lineage).

In subfamily III, several groupings were present in more than 99% of trials: ( $\beta 2$ ,  $\beta 4$ ), ( $\beta 3$ ,  $\alpha 5$ ), ( $\alpha 2$ ,  $\alpha 4$ ), ( $\alpha 3$ ,  $\alpha 6$ ). A first duplication gave  $\beta 2$  and  $\beta 4$ . The position of  $\beta 2$  and  $\beta 4$  was unstable, jumping between subfamily III and subfamily IV according to the sequences sampled. However, if we consider the gene structure, the neuronal localization, and the pharmacological characteristics of these subunits,  $\beta 2$  and  $\beta 4$  have to be placed in subfamily III. Then we observe the separation of  $\beta 3$  and  $\alpha 5$ . At last, a monophyletic group formed by the two pairs ( $\alpha 4$ ,  $\alpha 2$ ) and ( $\alpha 3$ ,  $\alpha 6$ ) is present in MP and NJ analyses. The position of  $\alpha 1$  does not match the gene-structure analysis. This strange position of  $\alpha 1$  inside the

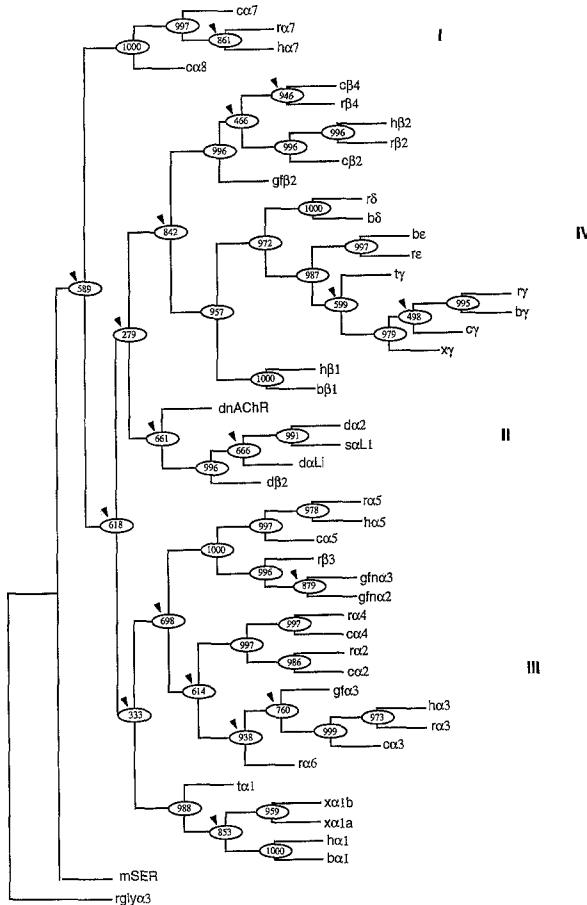


**Fig. 2.** Bootstrap majority-rule consensus tree obtained from 1000 MP replicates (SEQBOOT, PROTPARS and CONSENSE programs) with the alignment shown in Appendix 2. The nodes indicated by an arrowhead are uncertain.

neuronal subgroup is, however, weakly supported by bootstrap scores. The last clear duplications arose in the lineage of teleosteans which possess two homologs of  $\beta 3$  (appeared about 280 MYA), and in Tetrapoda, which have two homologs of the goldfish  $\beta 2$ ,  $\beta 2$ , and  $\beta 4$  sub-units. Moreover, goldfish  $\alpha 3$  is not clearly homolog to tetrapod  $\alpha 3$  (NJ Fig. 3) or  $\alpha 6$  (MP Fig. 2), which could then appear only after the divergence of teleosteans.

### *Evolution of the Stoichiometry*

As the nAChR is an oligomer formed by subunits coded by paralogs, it is reasonable to assume that the primitive receptor resulted from the assembly of just one subunit. Thus, the ability for a subunit to form functional homo-oligomers could reflect a plesiomorphic ("primitive") mode of functioning.  $\alpha$ -Bgt-sensitive homo-oligomers from *Locusta migratoria* (Breer et al. 1985) have been purified and reconstituted in vitro (Hanke and Breer 1986). The  $\alpha$ xL1 subunit of *Schistocerca gregaria* forms



**Fig. 3.** Bootstrap majority-rule consensus tree obtained from 1000 NJ replicates (SEQBOOT, PROTDIST, NEIGHBOR, and CONSENSE programs) with the alignment shown in Appendix 2. The nodes indicated by an arrowhead are uncertain.

functional homo-oligomeric channels (Marshall et al. 1990) blocked by  $\alpha$ -Bgt in vitro.  $\alpha 2$  (also called SAD for *second alpha subunit*), the putative *Drosophila* homolog of locust  $\alpha$ CL1, forms functional receptors alone in *Xenopus* oocytes (Sawruk et al. 1990) though these receptors display an atypical pharmacology. In the same way,  $\alpha 7$  from chicken is able to form homo-oligomers in *Xenopus* oocytes (Couturier et al. 1990; Revah et al. 1991; Anand et al. 1993). In contrast, vertebrate nAChR subunits from subfamilies III and IV, expressed in *Xenopus* oocyte, cannot form functional homo-oligomeric channels.

## *Pharmacological Argument for Monophyly of Neuronal Nicotinic Subfamily*

Although a small number of mutations sometimes suffice to dramatically change the properties of a receptor (e.g., Galzi et al. 1992), the pharmacological properties of the families of ligand-gated ion channels seem to diverge slowly. *Ascaris* muscle (Walker et al. 1992), *Aplysia* (Ono and Salvaterra 1981),  $\alpha$ L1 (insect class 2, Marshall

et al. 1990), dnAChR (insect class 1, Schloss et al. 1988), chicken  $\alpha$ 7 (Couturier et al. 1990; Anand et al. 1993), and vertebrate striated muscle (Lee and Chang 1966; Changeux et al. 1970) receptors are  $\alpha$ -Bgt sensitive. Although  $\alpha$ 1 belongs to subfamily III, the functional  $\alpha$ -Bgt sites of the vertebrate muscle receptor are formed partially by the subunits of subfamily IV. Moreover, if  $\alpha$ 1 is placed as a sister group of all other subunits of subfamily III (as indicated by the gene structure), the loss of  $\alpha$ -Bgt sensitivity in the neuronal nicotinic subfamily is a **synapomorphy**. In addition, *Ascaris* muscle receptor (Walker et al. 1992), *Aplysia* neuronal receptors (Ono and Salvaterra 1981), and the receptor formed by s<sub>o</sub>L1 of *Schistocerca* (Marshall et al. 1990) are sensitive to strychnine, an antagonist of the glycine receptor, and to bicuculline, an antagonist of the GABA<sub>A</sub> receptor.  $\alpha$ 7 is also sensitive to strychnine (Anand et al. 1993). The members of a multigene family can then share pharmacological properties, even after a long divergence (probably more than 1,000 MYA here). Receptors of subfamily III do not seem to be blocked by these antagonists (Clément Léna, personal communication). Overall, the evidence from pharmacological studies further supports the notion of the monophyly of the subfamily III.

## Discussion

The analyses presented in this paper lead to the reconstruction of a global history of nAChR evolution. Although several nodes have not been perfectly resolved, the major relationships between subunits were clarified. Except for  $\alpha 1$  and  $(\beta 2, \beta 4)$  all the analyses performed were congruent.

### *Hypothetically Missing Genes*

Subfamily I diverged before the split insects/vertebrates, and this subfamily could be present in insects. (The cloned insect subunits are orthologous to the subfamily III and IV.)

Neuromuscular transmission via nAChRs is known to occur in nematodes (Gerschenfeld 1973; Walker et al. 1992), annelids, molluscs (Gerschenfeld 1973), and vertebrates but not in insects and crustaceans. The chemical excitation of muscle in the Bilateria nonvertebrates/nonarthropods has to be mediated by subunits which do not belong to the subfamily IV (Fig. 2). Thus, neuromuscular transmission in vertebrates is not **homologous** to that occurring in other phyla.

The  $\epsilon$  subunit seems to be present in the whole Gnathostomata phylum. This is consistent with the reported presence of an  $\epsilon$  subunit in *Xenopus*, yet this subunit has not been cloned in chicken.

The bootstrap confirms that  $\alpha_7$  and  $\alpha_8$  diverged prior

to the separation of Sauropsida and Therapsida. Thus, an  $\alpha 8$  subunit may be present in mammals.

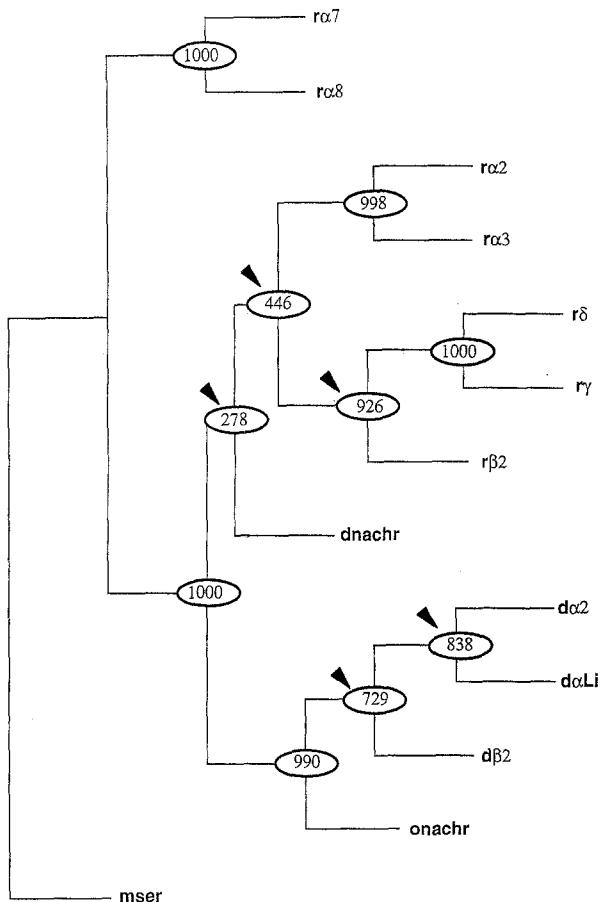
#### *Reconstructed History of the nAChR-Subunit Gene Family*

Based on present and previous results, the history of the nAChR-subunit gene family can be reconstructed as follows (Fig. 5):

We can plausibly assume (still without proof) that in the primitive metazoans (e.g., coelenterates) nAChR was made of a single subunit able to form homo-oligomers. The coelenterates have no true muscle cells but already have multipolar neurons, of ectodermal origin. This first nAChR presumably had a neuronal localization. With the appearance of a third embryonic sheet, the nAChR acquired a novel role in neuromuscular transmission. However, if the nematodes and the molluscs have a muscle nAChR, it is not homologous to the vertebrate subfamily IV. Indeed, this latter plausibly appeared after the differentiation of Deuterostomata. The subunit cloned in *Onchocerca* does not possess the third loop of the ACh binding site (Devillers-Thiéry et al. 1993) and might be a non- $\alpha$  subunit. The NJ analysis of the alignment shown in appendix 3 (Fig. 4) determined this subunit to be an extra group of three *Drosophila* subunits containing two  $\alpha$  and one non- $\alpha$  subunits. The idea of a precocious emergence of the insect subunits is supported by the ability of these subunits to form homo-oligomers in vitro. Assuming that alphaL1 of *Schistocerca* and alpha2 of *Drosophila* are orthologs, the duplication between them and d $\alpha$ L1 is older than the divergence of Orthoptera and Diptera—i.e., older than 300 MYA (Labandeira and Sepkoski 1993). In Deuterostomata, several duplications occurred to give extant subfamily IV, which was complete in vertebrate phylum before the appearance of chondrichthyes (450 MYA) and extant subfamily III, one of the paralogs being expressed in muscle.

Several tissular switches of expression from neuron to muscle or from muscle to neuron can be hypothesized (Fig. 5). Between the divergence of subfamily II and the split subfamily III/subfamily IV (in the chordate lineage), one switch of expression might have given a muscle receptor, possibly homopentameric. After the first duplication between the ancestor of subfamily IV and  $\alpha 1$  (a duplication which is responsible of the heteromeric muscle receptor), a further duplication from  $\alpha 1$  provided a new gene, which expression became neuronal. The evolution of the promoters (and of the transcription regulators) may thus have played a role as important as gene duplication in the diversification of the nAChR family.

The neuronal non- $\alpha$  subunit group is likely to be polyphyletic, whereas the neuronal  $\alpha$  subunits (the “binding subunits”) would form a monophyletic group.  $\alpha 5$ , which lacks some important aromatic amino acids in the third

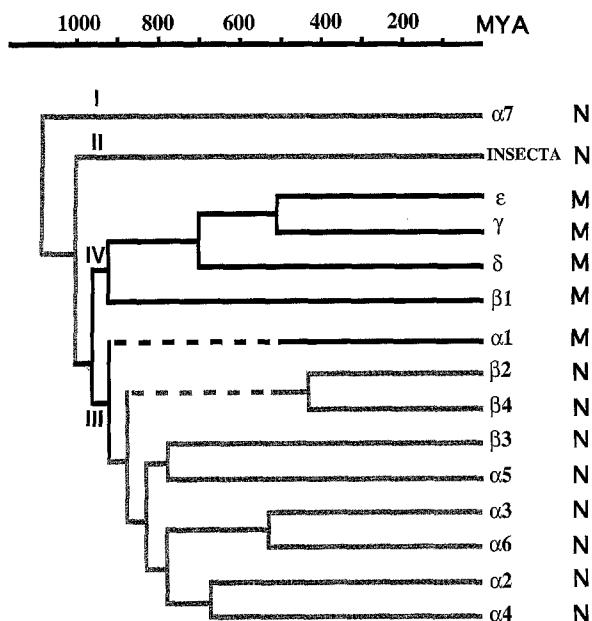


**Fig. 4.** Bootstrap majority-rule consensus tree obtained from 1,000 NJ replicates with the alignment shown in Appendix 3, presenting the possible emergence of the nematode subunit. The nodes indicated by an arrowhead are uncertain.

loop of the ACh binding site, cannot form functional receptors in vitro with any  $\beta$  subunit (Boulter et al. 1990) but is coprecipitated from endogenous material with other  $\alpha$  subunits (Conroy et al. 1992; Vernalis et al. 1993); it thus may represent a new type of “structural” subunit and should therefore be given another name.  $\alpha 5$  and  $\beta 3$  could be called  $\gamma 2$  and  $\gamma 3$ . Then the three types of subfamily III subunits could form monophyletic groups—the tribes  $\alpha$ ,  $\beta$ , and  $\gamma$ .

#### *Growth of the Neuronal Nicotinic Subfamily and Increase in Complexity of the Cholinergic System*

The multiple duplications in subfamily III parallel the progressive increased complexity of the chordate nervous system—in particular, of the cholinergic system. At the beginning of the evolution of this phylum, one subunit was plausibly present in the nervous system, resulting from the duplication of  $\alpha 1$  (in addition to the ancestor of  $\alpha 7$  and  $\alpha 8$ ). The diversity of the group increased during the first 400 MY, until the appearance of Tetrapoda. The whole evolution of the subfamily occurred in



**Fig. 5.** Summary tree, integrating the results of the whole study. The dates of the last divergences have been calculated from the protein alignment of Appendix 2 (cf. Materials and Methods).  $\alpha 7/\alpha 8$ : 380 MYA;  $\epsilon/\gamma$ : 508 MYA;  $\epsilon,\gamma,\delta/\beta 1$ : 711 MYA;  $\epsilon,\gamma,\delta/\beta 1$ : 926 MYA;  $\alpha 3/\alpha 6$ : 529 MYA;  $\alpha 2/\alpha 4$ : 669 MYA;  $\alpha 5/\beta 3$ : 770 MYA. The ages of the precocious divergences have been approximately inferred from the divergence of nematodes (1,000 MYA: Vanfleteren et al. 1994). M: muscle subunit; N: neuronal subunit. The gray branches represent subunits putatively expressed in neurons. The black branches represent subunits possibly expressed in muscle.

the first half of Deuterostomata history (from 600–800 MYA to about 300 MYA). In the first prechordate fossils, we find only two ganglia, the peripheral and the cerebroid ganglia. *Branchiostoma* has only one pseudovesicle in the head. Spinal chord and cholinergic peripheral nervous system were present early in the vertebrate lineage (although the complete autonomous system was reached only in mammals). Lamprey already has five vesicles but the main development of the brain and particularly of the forebrain occurred in Gnathostomata.

In situ hybridization (Deneris et al. 1989; Wada et al. 1989, 1990; Zoli et al. 1995) as well as immunohistochemical (Britto et al. 1992; Hill et al. 1993) studies have shown that, in rat and chicken brain,  $\alpha 4$  and  $\beta 2$  mRNA distribution is diffuse, whereas  $\alpha 2$ ,  $\alpha 3$ ,  $\alpha 5$ ,  $\alpha 6$ ,  $\beta 3$ , and  $\beta 4$  are mainly restricted to a few major cholinergic or cholinoreceptive pathways, which, however, also express  $\alpha 4$  and  $\beta 2$ .  $\beta 2$  has diverged early in the neuronal subfamily history (Figs. 2, 3, and Results).  $\alpha 4$  and  $\beta 2$  could represent a “fossil” expression, which was present in most areas of the ancestral brain. When a duplication occurred, one of the paralogs kept the specific role of the “father gene,” whereas the other paralog had to acquire a new role. This role can be defined by a new domain of expression (like the switches muscle/neuron developed before) or by a modified function. The hypothesis that  $\alpha 4$  maintained its previous role while another paralog ac-

quired a new role is supported by some evidence on the  $\alpha 2$  subunit. In chick brain, the  $\alpha 2$  subunit is restricted to the lateral spiriform nucleus (Daubas et al. 1990) but, in rat brain it is restricted to the interpeduncular nucleus (Wada et al. 1989)—a nonhomolog structure. (The interpeduncular nucleus also exists in the chick brain.) Moreover, there is no homolog of the lateral spiriform nucleus in rat brain, a fact that points to the genesis of this structure after the divergence of the bird lineage. It is attractive to suppose that a gene duplication occurred a short time before the branching of Therapsida and Sauropsida (i.e., before 310 MYA, Benton 1990). This time would have been too short to define the specificity of  $\alpha 2$  (in contrast to  $\alpha 4$ , which maintained its ancient role). Then two independent specificities of expression took place in the two phyla. Accordingly, a transgene with the avian  $\alpha 2$  gene (including the promoter) is expressed throughout the rat brain, mostly in cholinergic structures (motor nuclei and basal telencephalon) (Daubas et al. 1993). (Nevertheless, this distribution corresponds neither to the distribution of endogenous  $\alpha 2$  nor to that of  $\alpha 4$ .)

In the same way, the duplications  $\alpha 3/\alpha 6$  and  $\beta 2/\beta 4$  occurred a little before, or a little after, the split between the teleost and the tetrapod lineages. In the rat brain,  $\alpha 3$  and  $\beta 4$  are mainly expressed in the medial habenula, a cholinergic and cholinoreceptive structure. However, in a teleost fish (*Phoxinus phoxinus*), an immunocytochemical study did not find any cholinergic cell and found only a small number of cholinergic fibers in the habenula (Ekström 1987). If these characteristics are plesiomorph, there could be again a correlation between gene duplications and a further change of function.  $\alpha 4$  and  $\beta 2$  could thus have kept the ancestor role of the neuronal nAChR, whereas other paralogs could have found new functional specificities in the evolving cholinergic systems.

We have shown, on the basis of cladograms and phenograms, that the first duplications in the nAChR occurred before the divergence of nematodes. Several nAChR subfamilies were identified. There is congruence between sequence and gene-structure analyses, and the three subfamilies present in vertebrates correspond closely to the functional subgroups (described from anatomical, pharmacological, and structural considerations). Two phenomena seem to have generated the wealth of the family. First, several switches of expression seem to have occurred from neuron to muscle and the opposite. Second, multiple gene duplications gave the extant number of paralogs. The neuronal nicotinic-subunit subfamily (type III subunits) appeared at the beginning of the chordate phylum and grew until the separation of Sauropsida and Therapsida lineages. This diversification, both quantitative and functional, paralleled the increase in complexity of the cholinergic systems. A link between an increased combinatorial complexity of subunit combinations and a larger plasticity in the functioning of these pathways is plausible.

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## References

- Anand R, Conroy WG, Schoepfer R, Whiting P, Lindstrom J (1991) Neuronal nicotinic acetylcholine receptors expressed in *Xenopus* oocytes have a pentameric quaternary structure. *J Biol Chem* 266: 11192–11198
- Anand R, Peng X, Lindstrom J (1993) Homomeric and native alpha<sub>7</sub> acetylcholine receptors exhibit remarkably similar but non-identical pharmacological properties, suggesting that the native receptor is a heteromeric protein complex. *FEBS Lett* 327:241–246
- Benton MJ (1990) Phylogeny of the major tetrapod groups: morphological data and divergence dates. *J Mol Evol* 30:409–424
- Bertrand D, Galzi JL, Devillers-Thiéry A, Bertrand D, Changeux JP (1993) Stratification of the channel domain in neurotransmitter receptors. *Curr Opin Cell Biol* 5:688–693
- Boulter J, O’Shea-Greenfield A, Duvoisin R, Connolly JG, Wada E, Jensen A, Gardner PD, Ballivet M, Deneris ES, McKinnon D, Heinemann Patrick J (1990) alpha<sub>3</sub>, alpha<sub>5</sub>, and beta<sub>4</sub>: three members of the rat neuronal nicotinic acetylcholine receptor-related gene family form a gene cluster. *J Biol Chem* 265:4472–4482
- Breer H, Kleene R, Hinz G (1985) Molecular forms and subunit structure of the acetylcholine receptor in the central nervous system of insects. *J Neurosci* 5:3386–3392
- Brehm P, Okamura Y, Mandel G (1991) Ion channel evolution. *Semin Neurosci* 3:355–367
- Britto LRG, Keyser KT, Lindstrom JM, Karten HJ (1992) Immunohistochemical localization of nicotinic acetylcholine receptor subunits in the mesencephalon and diencephalon of the chick (*Gallus gallus*). *J Comp Neurol* 317:325–340
- Changeux JP, Kasai M, Lee CY (1970) The use of snake venom toxin to characterize the cholinergic receptor protein. *Proc Natl Acad Sci USA* 67:1241–1247
- Changeux JP (1990) Functional architecture and dynamics of the nicotinic acetylcholine receptor: an allosteric ligand-gated ion channel. *Fidia Res Found Neurosci Award Lectures* 4:21–168
- Clarke PBS, Schwartz RD, Paul SM, Pert CB, Pert A (1985) Nicotinic binding in rat brain: autoradiographic comparison of [<sup>3</sup>H]acetylcholine, [<sup>3</sup>H]nicotine and [<sup>125</sup>I]alpha-bungarotoxin. *J Neurosci* 5:1307–1315
- Cockcroft VB, Osguthorpe DJ, Barnard EA, Friday AE, Lunt GG (1992) Ligand-gated channels. Homology and diversity. *Mol Neurobiol* 4:129–169
- Conroy G, Vernalis AB, Berg DK (1992) The alpha<sub>5</sub> gene product assembles with multiple acetylcholine receptor subunits to form distinctive receptor subtypes in brain. *Neuron* 9:679–691
- Cooper E, Couturier S, Ballivet M (1991) Pentameric structure and subunit stoichiometry of a neuronal acetylcholine receptor. *Nature* 350:235–238
- Couturier S, Bertrand D, Matter JM, Hernandez MC, Bertrand S, Millar N, Valera S, Barkas T, Ballivet M (1990) A neuronal nicotinic acetylcholine receptor subunit (alpha<sub>7</sub>) is developmentally regulated and forms a homo-oligomeric channel blocked by a-BTX. *Neuron* 5:847–856
- Darlisson MG, Hutton ML, Harvey RJ (1993) Molluscan ligand-gated ion-channel receptors. In: Pichon Y (ed) EXS 63, comparative molecular neurobiology. Birkhäuser, Basel, pp 48–64
- Daubas P, Devillers-Thiéry A, Geoffroy B, Martinez S, Bessis A, Changeux JP (1990) Differential expression of the neuronal acetylcholine receptor alpha<sub>2</sub> subunit gene during chick brain development. *Neuron* 5:49–60
- Daubas P, Salmon AM, Zoli M, Geoffroy B, Devillers-Thiéry A, Bessis A, Médeville F, Changeux JP (1993) Chicken neuronal acetylcholine receptor alpha<sub>2</sub>-subunit gene exhibits neuron-specific expression in the brain and spinal cord of transgenic mice. *Proc Natl Acad Sci USA* 90:2237–2241
- Dayhoff MO (1979) Atlas of protein sequence and structure, vol 5, supplement 3, 1978. National Biomedical Research Foundation, Washington DC
- Deneris ES, Boulter J, Swanson LW, Patrick J, Heinemann S (1989) beta<sub>3</sub>: a new member of nicotinic acetylcholine receptor gene family is expressed in brain. *J Biol Chem* 264:6268–6272
- Devillers-Thiéry A, Galzi JL, Eiselle JL, Bertrand S, Bertrand D, Changeux JP (1993) Functional architecture of the nicotinic acetylcholine receptor: a prototype of ligand-gated ion channels. *J Membr Biol* 136:97–112
- Eck RV, Dayhoff MO (1966) Atlas of protein sequence and structure. National Biomedical Research Foundation, Silver Spring, MD
- Ekström P (1987) Distribution of choline acetyltransferase-immuno-reactive neurons in the brain of a cyprinid teleost (*Phoxinus phoxinus* L.). *J Comp Neurol* 256:494–515
- Estabrook GF, Johnson CS Jr, McMorris FR (1976) A mathematical foundation for the analysis of character compatibility. *Math Biosci* 23:181–187
- Felsenstein J (1985) Confidence limits on phylogenies: an approach using the bootstrap. *Evolution* 39:783–791
- Felsenstein J (1993) PHYLIP (phylogeny inference package) version 3.5c. Distributed by the author. Department of Genetics, University of Washington, Seattle
- Feng DF, Doolittle RF (1987) Progressive sequence alignment as a prerequisite to correct phylogenetic trees. *J Mol Evol* 25:351–360
- Fitch WW (1971) Toward defining the course of evolution: minimum change for a specified tree topology. *Syst Zool* 20:406–416
- Flemming JT, Tornoe C, Riina HA, Coadwell J, Lewis JA, Sattelle DB (1993) Acetylcholine receptor molecules of the nematode *Caeenorhabditis elegans*. In: Pichon Y (ed) EXS 63, comparative molecular neurobiology. Birkhäuser, Basel, pp 65–80
- Galzi JL, Devillers-Thiéry A, Hussy N, Bertrand S, Changeux JP, Bertrand D (1992) Mutations in the channel domain of a neuronal nicotinic receptor convert ion selectivity from cationic to anionic. *Nature* 359:500–505
- Galzi JL, Changeux JP (1994) Ligand-gated ion channel as unconventional allosteric proteins. *Curr Opin Struct Biol* 4:554–565
- Gerschenfeld HM (1973) Chemical transmission in invertebrate central nervous systems and neuromuscular junctions. *Physiol Rev* 53:1–119
- Greenberg ME, Ziff EB, Greene LA (1986) Stimulation of neuronal acetylcholine receptors induces rapid gene transcription. *Science* 234:80–83
- Gundelfinger ED (1992) How complex is the nicotinic receptor system of insects? *TINS* 15:206–211
- Hanke W, Breer H (1986) Channel properties of an insect neuronal acetylcholine receptor protein reconstituted in planar lipid bilayers. *Nature* 321:171–174
- Higgins DG, Sharp PM (1988) CLUSTAL: a package for performing multiple sequence alignments on a microcomputer. *Gene* 73:237–244
- Hill JA, Zoli M, Bourgeois JP, Changeux JP (1993) Immunocytochemical localization of a neuronal nicotinic receptor: The beta<sub>2</sub> subunit. *J Neurosci* 13:1551–1568
- Jonas P, Baumann A, Merz B, Gundelfinger ED (1990) Structure and developmental expression of the Da2 gene encoding a novel nicotinic acetylcholine receptor protein of *Drosophila melanogaster*. *FEBS Lett* 269:264–268
- Karlin A (1993) Structure of nicotinic acetylcholine receptors. *Curr Opin Neurobiol* 3:299–309

- Labandeira CC, Sepkoski JJ Jr (1993) Insect diversity in the fossil record. *Science* 261:310–315
- Lake JA (1990) Origin of the metazoa. *Proc Natl Acad Sci USA* 87: 763–766
- Lee CY, Chang CC (1966) Modes of actions of purified toxins from elapid venoms on neuro-muscular transmission. *Mem Inst Butantan São Paulo* 33:555–572
- Leech CA, Sattelle DB (1993) Acetylcholine receptor/channel of insects. In: Pichon Y (ed) EXS 63, Comparative molecular neurobiology. Birkhäuser, Basel, pp 81–97
- Le Quesne WJ (1969) A method of selection of characters in numerical taxonomy. *Syst Zool* 18:201–205
- Luetje CW, Patrick J (1991) Both  $\alpha$ - and  $\beta$ -subunits contribute to the agonist sensitivity of the neuronal nicotinic acetylcholine receptor. *J Neurosci* 11:837–845
- Marshall J, Buckingham SD, Shingai R, Lunt GG, Goosey MW, Darlington MG, Satelle DB, Barnard EA (1990) Sequence and functional expression of a single  $\alpha$  subunit of an insect nicotinic receptor. *EMBO J* 9:4391–4398
- Mulle C, Vidal C, Benoît P, Changeux JP (1991) Existence of different subtypes of nicotinic acetylcholine receptors in the rat habenulo-interpeduncular system. *J Neurosci* 11:2588–2597
- Ono JK, Salvaterra PM (1981) Snake alpha-toxin effects on cholinergic and noncholinergic responses of *Aplysia californica* neurons. *J Neurosci* 1:259–270
- Revh F, Bertrand D, Galzi JL, Devillers-Thiéry A, Mulle C, Hussy N, Bertrand S, Ballivet M, Changeux JP (1991) Mutations in the channel domain alter desensitization of a neuronal nicotinic receptor. *Nature* 353:846–849
- Role LW (1992) Diversity in primary structure and function of neuronal nicotinic acetylcholine receptor channels. *Curr Opin Neurobiol* 2:254–262
- Saitou N, Nei M (1987) The neighbor-joining method: a new method for reconstructing phylogenetic trees. *Mol Biol Evol* 4:406–425
- Sargent PB (1993) The diversity of neuronal nicotinic acetylcholine receptors. *Annu Rev Neurosci* 16:403–443
- Sawruk E, Schloss P, Betz H, Schmitt B (1990) Heterogeneity of *Drosophila* nicotinic receptors: SAD, a novel developmentally regulated  $\alpha$ -subunit. *EMBO J* 9:2671
- Schloss P, Hermans-Borgmeyer I, Betz H, Gundelfinger ED (1988) Neuronal acetylcholine receptor in *Drosophila*: the ARD protein is a component of a high affinity  $\alpha$ -bungarotoxin binding complex. *EMBO J* 7:2889–2894
- Schoepfer R, Conroy WG, Whiting P, Gore M, Lindstrom J (1990) Brain  $\alpha$ -bungarotoxin binding protein cDNAs and MAbs reveal subtypes of this branch of the ligand-gated ion channel gene superfamily. *Neuron* 5:35–48
- Segerberg MA, Stretton AOW (1993) Actions of cholinergic drugs in the nematode *Ascaris suum*. Complex pharmacology of muscle and motorneurons. *J Gen Physiol* 101:271–296
- Sneath PHA, Sokal RR (1973) Numerical taxonomy. Freeman, San Francisco
- Vanfleteren JR, Van de Peer Y, Blaxter ML, Tweedie SAR, Trotman C, Lu L, Van Hauwaert ML, Moens L (1994) Molecular genealogy of some nematode taxa as based on cytochrome c and globin amino acid sequences. *Mol Phylogen Evol* 3:92–101
- Vernallis AB, Conroy WG, Berg DK (1993) Neurons assemble acetylcholine receptors with as many as three kinds of subunits while maintaining subunit segregation among receptor subtypes. *Neuron* 10:451–464
- Wada K, Ballivet M, Boulter J, Connolly J, Wada E, Deneris ES, Swanson LW, Heinemann S, Patrick J (1988) Functional expression of a new pharmacological subtype of brain nicotinic acetylcholine receptor. *Science* 240:330–334
- Wada E, Wada K, Boulter J, Deneris E, Heinemann S, Patrick J, Swanson LW (1989) Distribution of  $\alpha$ 2,  $\alpha$ 3,  $\alpha$ 4 and  $\beta$ 2 neuronal nicotinic receptor subunit mRNAs in the central nervous system: a hybridization histochemical study in the rat. *J Comp Neurol* 284: 314–335
- Wada E, McKinnon D, Heinemann S, Patrick J, Swanson LW (1990) The distribution of mRNA encoded by a new member of the neuronal nicotinic acetylcholine receptor gene family ( $\alpha$ 5) in the rat central nervous system. *Brain Res* 526:45–53
- Walker RJ, Colquhoun L, Holden-Dye L (1992) Pharmacological profiles of the GABA and acetylcholine receptors from the nematode, *Ascaris suum*. *Acta Biol Hung* 43:59–68
- Whiting P, Schoepfer R, Lindstrom J, Priestley T (1991) Structural and pharmacological characterization of the major brain nicotinic acetylcholine receptor subtype stably expressed in mouse fibroblast. *Mol Pharmacol* 40:463–472
- Wilbur WJ, Lipman DJ (1983) Rapid similarity searches of nucleic acid and protein data banks. *Proc Natl Acad Sci USA* 80:726–730
- Zoli M, Le Novère N, Hill JA, Changeux JP (1995) Developmental regulation of nicotinic receptor subunit mRNAs in the rat central and peripheral nervous system. *J Neurosci* (in press)

## Appendix 1: Glossary of the Cladistic Terms Present in the Text

**Homology:** Two similar characters are homologous when they come from a common ancestor. There is homology between two genes if they arose by duplication. Characters are homologous if (1) they look like each other, (2) they do not coexist in the same organism (the arms and the wings of angels), and (3) if they provide the same phylogenies as other homologous characters.

**Ortholog:** Two orthologs are homologs that have arisen by speciation (the “same” gene in two different species).

**Paralog:** Two paralogs are homologs that have arisen by duplication in the same organism.

**Monophly:** A monophyletic group is formed by one ancestor and all its descendants.

**Polyphyly:** A polyphyletic group is formed by different subgroups, which do not share any common ancestor belonging to the group.

A **plesiomorphy** is an ancestor character. It is not useful in reconstructing phylogenies.

An **autapomorphy** is a derived character, present in one descendant only. It is not useful in reconstructing phylogenies

A **synapomorphy** is a derived character, shared by a monophyletic group of several descendants. The synapomorphies are the only useful characters with which to infer phylogenies

**Informative site:** A character is informative if it exists under at least two states present twice. A character presenting everytime the same state as well as a character different in every compared object are not informative

b $\alpha$ 1 ETRLVAKLFE D--YNSVVVP VEDHRQAVEV TVGLQLIQLI NVDEVNQIVT  
 b $\beta$ 1 EGRLREKLFS --GYDSTVRP AREVGDRVVW SIGLTLAQLI SLNEKDEEMS  
 b $\delta$  EERLIRHLFE EKAYNKELRP AAHKESV-EI SLALTLSNLI SLKEVEETLT  
 b $\epsilon$  ELRLYHHLFD --TYDPGRRP VQEPEDTVTI SLKVTLTNLI SLNEKEETLT  
 b $\gamma$  EERLLGDLMO --GYNPHLRP AEHDSDVVNV SLKLTLTNLI SLNEREALALT  
 c $\alpha$ 2 EDRLFKHLFT G--YNRWSRP VPNTSDVVIV KFGLSIAQLI DVDEKNQMMT  
 c $\alpha$ 3 EHRLYAAFLK N--YNQFVRP VKNASDPVII QFEVSMSQLV KVDEVNQIME  
 c $\alpha$ 4 EERLLKKLFS G--YNKWSRP VANISDVVLV RFGLSIAQLI DVDEKNQMMT  
 c $\alpha$ 5 EDRLFKHLFE D--YQRWVRP VEHLNDTIKI KFGLAISQLV DVDEKNQLMT  
 c $\alpha$ 7 QRKLYKELLK N--YNPLERP VANDSQPLTV YFTLSLMQIM DVDEKNQVLT  
 c $\alpha$ 8 QRRLYRDLLR N--YNRLELRP VMNDSQPIVV ELQLSLLQII DVDEKNQVLI  
 c $\beta$ 2 EERLVEYLL PTRYNKLIIRP ATNGSQLVTI QLMVSLAQLI SVHEREQIMT  
 c $\beta$ 4 EEKLMNHLLS PDRYNKLIIRP AVNSSLVSI ELQVSLAQLI SVNEREQIMT  
 c $\gamma$  EEKLLQDLMT --NYNRHLRP ALRGDQVIDV TLKLTLTNLI SLNEREETLT  
 d $\alpha$ 2 AKRLYDDLLS N--YNRLIRP VSNNTDTVVL KLGRLSQLI DLNLKDQILT  
 d $\alpha$ L1 AKRLYDDLLS N--YNRLIRP VGNNSDRLTV KMGLRLSQLI DVNLKNQIMT  
 d $\beta$ 2 TKRLYDDLLS N--YNRLIRP VVNNTETLTV WGLKLSQLI EVNLKNQVMT  
 dnachr EERLVRDLF R--YNNKLIRP VQNMHQVGV RFGLAFVQLI NVNEKNQVMK  
 g $\alpha$ 3 EDRLFRRLLFR R--YNQFIRP VENVSDPVTV EFEVSIQLV KVDEVNQIME  
 g $\beta$ 2 LRS--DFLLG PERYNKLIIRP AVNKSQQVTI GIKVSLAQLI SVNEREQIMT  
 gfn $\alpha$ 2 EDALLRELFO G--YQRWVRP VQHANHSVVK RFGLKISQLV DVDEKNQLMT  
 gfn $\alpha$ 3 EDTLLRNLLFR G--YQKWVRP ILHANDTITV RFGLKISQLV DVDEKNHLMT  
 h $\alpha$ 1 ETRLVAKLFE D--YSSVVVP VEDHRQVVEV TVGLQLIQLI NVDEVNQIVT  
 h $\alpha$ 3 EHRLFERLFE D--YNEIIIRP VANVSDPVII HFEVSMSQLV KVDEVNQIME  
 h $\alpha$ 5 EDSLLKDLFQ D--YERWVRP VEHLNDKIKI KFGLAISQLV DVDEKNQLMT  
 h $\alpha$ 7 QRKLYKELVK N--YNPLERP VANDSQPLTV YFSLSLIQIM DVDEKNQVLT  
 h $\beta$ 1 EGRLREKLFS --GYDSSVRP AREVGDRVRV SVGLILAQLI SLNEKDEEMS  
 h $\beta$ 2 EERLVEHLLD PSRYNKLIIRP ATNGSELVTI QLMVSLAQLI SVHEREQIMT  
 mserr LLRLSDHLLA --NYKKGVRP VRDWRKPTTV SIDVIMYAIL NVDEKNQVLT  
 r $\alpha$ 2 EDRLFKHLFG G--YNRWARP VPNTSDVVIV RFGLSIAQLI DVDEKNQMMT  
 r $\alpha$ 3 EHRLFQYLFE D--YNEIIIRP VANVSHPVII QFEVSMSQLV KVDEVNQIME  
 r $\alpha$ 4 EERLLKRLFS G--YNKWSRP VGNISDVVLV RFGLSIAQLI DVDEKNQMMT  
 r $\alpha$ 5 EDSLFRDLFE D--YERWVRP VEHLSDKIKI KFGLAISQLV DVDEKNQLMT  
 r $\alpha$ 6 EEQLFHTLFA H--YNRFIRP VENVSDPVTV HFELAITQLA NVDEVNQIME  
 r $\alpha$ 7 QRRLYKELVK N--YNPLERP VANDSQPLTV YFSLSLIQIM DVDEKNQVLT  
 r $\beta$ 2 EERLVEHLLD PSRYNKLIIRP ATNGSELVTI QLMVSLAQLI SVHEREQIMT  
 r $\beta$ 3 EDALLRHLFO G--YQKWVRP VLNSSDIIVK YFGLKISQLV DVDEKNQLMT  
 r $\beta$ 4 EEKLMDDLLN KTRYNNLIRP ATSSSQLISI RLELSLSQLI SVNEREQIMT  
 r $\delta$  EQRLIQHLFE EKGYNKELRP VARKEDIVDV ALSLTLTSNLI SLKEVEETLT  
 r $\epsilon$  ELSLYHHLF D--NYDPECRP VRRPEDTVTI TLKVTLTNLI SLNEKEETLT  
 r $\gamma$  EERLLADLMR --NYDPHRLRP AERDSDVNVN SLKLTLTNLI SLNEREALALT  
 rgly $\alpha$ 3 SDFLDKLMGR TSGYDARIRP -NFKGPPVNV TCNIFINSFG SIAETTMDFR  
 sc $\alpha$ 1 AKRLYDDLLS N--YNRLIRP VSNNTDTVVL KLGRLSQLI DLNLKDQILT  
 t $\alpha$ 1 ETRLVANLLE N--YNKVIIRP VEHHTHFVDI TVGLQLIQLI SVDEVNQIVE  
 t $\gamma$  EGRLIEKLLG --DYDKRIIP AKTLDHIIDV TLKLTLTNLI SLNEKEEALALT  
 x $\alpha$ 1a ESRLLINDLFK S--YNKVVVRP VKAFKDQVVV TVGLQLIQLI NVNEVNQIVT  
 x $\alpha$ 1b ETRLIGDLFA N--YNKVVVRP VETYKDQVVV TVGLQLIQLI NVDEVNQIVS  
 x $\gamma$  EERLLNDLMK --NYNKNLRLP VEKDGDIISV SIKLTLTNLI SLNEKEEALALT  
  
 b $\alpha$ 1 TNVRLKQWV DYNLKWNPDY YGGVKKIHIP SEKIWRPDLV LYNNADGDFR  
 b $\beta$ 1 TKVYLDLWEW DYLRLSWDPEE HEGIDSLRIS AESVWLDPDV LLNNNDGNFD  
 b $\delta$  TNVWIEQGWT DSRLQWDAED FGNISVLRP ADMVWLPEIV LENNNNDGSFQ  
 b $\epsilon$  TSVWIGIDWQ DYRLNYSKGD FGGVETLVRP SELVWLPEIV LENNIDGQFG  
 b $\gamma$  TNVWIEMQWC DYRLRWDPRD YGGLWVLRP STMWRPDIY LENNVGVFE  
 c $\alpha$ 2 TNVWLKQEWY DYKLRWNPDY FDNYTSIRVP SEMIWIPDIV LYNNADGEFA  
 c $\alpha$ 3 TNLWLKHIWN DYKLRWNPDY YGGAEFIRVP SGQIWKPDIV LYNNAVGDFQ  
 c $\alpha$ 4 TNVWVKQEWY DYKLRWDQPE YENVTSIRVP SELIWRPDIY LYNNADGDFR  
 c $\alpha$ 5 TNVWLKQEWY HVKLRWNPDY YAGITSIRVP SDSIWIPDIV LYDNADGRFE  
 c $\alpha$ 7 TNIWLQMYWT DYHQLQNVSE YPGVKNVRFP DGLIWKPDIL LYNSADERFD  
 c $\alpha$ 8 TNAWLQMYWV DYIYLSDQYE YPGVQNLVRP SDQIWVPDIL LYNSADERFD  
 c $\beta$ 2 TNVWLQEWY DYRLTWKPED FDNMKKVRLP SKHIWLDPDV LYNNADGMYE  
 c $\beta$ 4 TNVWLQEWY DYRLAWKPSD YEGINMLRIP AKHIWLDPDV LYNNADGTYE  
 c $\gamma$  TNVWIEMQWS DYRLRWDPDK YDDIQQLRVP SAMVWLDPDV LENNIDGTFE  
 d $\alpha$ 2 TNVWLHEHWQ DHKFKWDPSE YGGVTELYVP SEHIWLDPDV LYNNADGEYV  
 d $\alpha$ L1 TNVWVEQEWN DYKLNKWPDD YGGVDTLHVP SEHIWHPDV LYNNADGNYE  
 d $\beta$ 2 TNLWVKQRWF DYKLRWDPEE YGGVEQLYVP SEHIWVPDV LYNNWDGNYE  
 dnachr SNVWLRLVWY DYQLQWDEAD YGGIGVLRP PDKVWKPDIV LFNNADGNYE  
 g $\alpha$ 3 TNLWLRLHIWN DYKLNKWLPAE FDGIEFIRVP SNKIWRPDIY LYNNAVGDFL

gf $\beta$ 2	TNVWL/TQEWL	DYRLVWDPNE	YEGLKKLRIP	SQHIWLPDIV	LYNNADGVYE
gfn $\alpha$ 2	TNVWLWQEWL	DYKLWRNPEN	YGGITSIRVP	SESIWLPDIV	LYENADGRFE
gfn $\alpha$ 3	TNVWLWQEWT	DYKLWRNPED	YGGITSIRVP	SETIWLPDIV	LYENADGRFE
h $\alpha$ 1	TNVRLKQQWV	DYNLKWNPDD	YGGVKKIHIP	SEKIWRPDVV	LYNNADGDFA
h $\alpha$ 3	TNLWLKQIWN	DYKLKWNPSD	YGGAEFMRVP	AQKIWKPDIV	LYNNAVGDFQ
h $\alpha$ 5	TNVWLKQEWI	DVKLRWNPDD	YGGIKVIRVP	SDSSWTPDIV	LFDNADGRFE
h $\alpha$ 7	TNIWLQMSWT	DHYLQWNVSE	YPGVKTVRFP	DGQIWKPDLIL	LYNSADERFD
h $\beta$ 1	TKVYLDLEWT	DYRLSWDPAE	HEGIDSLRIT	AESVWLPDVV	LLNNNDGNFD
h $\beta$ 2	TNVWLQEWE	DYRLTWKPEE	FDNMKVRLP	SKHIWLPDVV	LYNNADGMYE
mser	TYIWYRQYWT	DEFLOQWTPED	FDNVTKLSIP	TDWIWVPDLIL	INEFVDVG-K
r $\alpha$ 2	TNVWLKQEWN	DYNVRWDPAE	FGNVTSLRVP	SEMIWLPDIV	LYNNADGEFA
r $\alpha$ 3	TNLWLKQIWN	DYKLKWKPDS	YQGVEFMRVP	AEKIWKPDIV	LYNNADGDFQ
r $\alpha$ 4	TNVWVKQEWH	DYKLRWDPGD	YENVTSIRIP	SELIWRPDIV	LYNNADGDFA
r $\alpha$ 5	TNVWLKQEWI	DVKLRWNPDD	YGGIKIIRVP	SDSLWIPDIV	LFDNADGRFE
r $\alpha$ 6	TNLWLRHVW	DYRLCWDPTE	YDGIELTRVP	ADNIWKPDIV	LYNNAVGDFQ
r $\alpha$ 7	TNIWLQMSWT	DHYLQWNMSE	YPGVKNVRFP	DGQIWKPDLIL	LYNSADERFD
r $\beta$ 2	TNVWLQEWE	DYRLTWKPKH	FDMNKVRLP	SKHIWLPDVV	LYNNADGMYE
r $\beta$ 3	TNVWLQEWL	DQKLRWNPEE	YGGINSIKVP	SESLWLPDIV	LFENADGRFE
r $\beta$ 4	TSIWLQEWL	DYRLAWNSSC	YEGVNILRIP	AKRVWLPDIV	LYNNADGTYE
r $\delta$	TNVWIDHAWI	DSRLQWNANE	FGNITVIRLP	SDMVWLPEIV	LENNNDGSFO
r $\varepsilon$	TSVWIGIEWQ	DYRLNFSKDD	FAGVEILRVP	SEHVWLPEIV	LENNIDGQFG
r $\gamma$	TNVWIEMQWC	DYRLRWDPKD	YEGLWILRVP	STMVWQPDIV	LGNNVDGVFE
rgly $\alpha$ 3	VNIFLRQKWN	DPRLAYSEYP	DDSDLDDPSM	LDSIWKPDLF	FANEKGANFH
scl1	TNVWLEHEWQ	DHKFRWDPAE	YGGVTELYVP	SEHIWLPDIV	LYNNADGEYV
t $\alpha$ 1	TNVRLRQQWI	DVRLRWNPAD	YGGIKKIRLP	SDDVWLPDLV	LYNNADGDFA
t $\gamma$	TNVWIEIQWN	DYRLSWNTSE	YEGIDLVRIP	SELLWLPDVV	LENNVDGQFE
x $\alpha$ 1a	TNVRLKQQWE	DVHLKWDPED	YGGIKKVRIP	SSDIWRPDIV	LYNNADGDFA
x $\alpha$ 1b	TNIRLKQQWR	DVNWKWDPAK	YGGVKKIRIP	SSDVWSPDLV	LYNNADGDFA
x $\gamma$	TNVWVEMQWK	DYRLSWDPND	YHGISMRRIP	STSVWLPDVG	LENNVDGTFD
b $\alpha$ 1	IVKFTKVLLD	YTGHITWTTP	AIFKSYCEII	VTHFPFDEQN	CSMKGWTWTY
b $\beta$ 1	VALDINVVVS	SDGSMRWQPP	GIYRSSCSIQ	VTYFPFDWQN	CTMVFSYSY
b $\delta$	ISYSCNVLIY	PSGSVYWLPP	AIFRSSCPIS	VTYFPFDWQN	CSLFSSLKY
b $\varepsilon$	VAYEANVLVS	EGGYLSWLPP	AIYRSTCAVE	VTYFPFDWQN	CSLVFRSQTY
b $\gamma$	VALYCNVLVS	PDGCVYWLPP	AIFRSSCPVS	VTFPFWDWQN	CSLIFQSQTY
c $\alpha$ 2	VTHMTKAHLF	SNGKVKWVPP	AIYKSSCSID	VTYFPFDQQN	CKMKFGSWTY
c $\alpha$ 3	VDDKTKALLK	YTGDVTWIPP	AIFKSSCKID	VTYFPFDYQN	CTMKFGWSY
c $\alpha$ 4	VTHLTKAHLF	YDGRIKWMPP	AIYKSSCSID	VTFPFWDQQN	CKMKFGSWTY
c $\alpha$ 5	GT-STKTVVK	YDGTIATWTPP	VNYKSSCTID	VTFPFWDLQN	CSMKFGSWTY
c $\alpha$ 7	ATFHTNVLVN	SSGHCQYLPP	GIFKSSCYID	VRWFPPFDVQK	CNLKFGSWTY
c $\alpha$ 8	ATFHTNVLVN	YSGSCQYIP	GILKSTCYID	VRWFPPFDVQK	CDLKFGSWTH
c $\beta$ 2	VSFYSNAVIS	YDGSIFWLPP	AIYKSACKIE	VKHFPPFDQQN	CTMKFRSWTY
c $\beta$ 4	VSLYTNAIVQ	NNGSIRWLPP	AIYKSACKIE	VKHFPPFDQQN	CTLKFRSWTY
c $\gamma$	ITLYTNVLVY	PDGSIYWLPP	AIYRSSCSII	VTYFPFDWQN	CTMVFQSQTY
d $\alpha$ 2	VTTMTKAILH	YTGKVVWTPP	AIFKSSCEID	VRYFPFDQQT	CFMKFGSWTY
d $\alpha$ Li	VTIMTKAILH	HTGKVVWKP	AIYKSFCID	VEYFPFDEQT	CFMKFGSWTY
d $\beta$ 2	VTLMTKATLK	YTGEVFWEPP	AIYKSSCEMN	VEYFPYDEQI	CFMKFGSWTY
dnachr	VRYKSNVLIY	PTGEVLWVPP	AIYQSSCTID	VTYFPFDQQT	CIMKFGSWTF
gfo $\alpha$ 3	VEDKTKALLK	YDGTITWVPP	AIFKSSCPMD	ITYFPFDYQN	CSMKFGSWTY
gf $\beta$ 2	VSFYCNAVVS	NTGDIWLP	AIYKSACAIE	VRNFPFDQQN	CTLKFRSWTY
gfn $\alpha$ 2	GSLMTKAIVR	YNGMITWTPP	ASYKSACTMD	VTFPFDRQN	CSMKFGSWTY
gfn $\alpha$ 3	GSLMTKAIVR	FNGTIMWTPP	ASYKSSCTMD	VTFPFDRQN	CSMKFGSWTY
h $\alpha$ 1	IVKFTKVLLQ	YTGHITWTPP	AIFKSYCEII	VTHFPFDEQN	CSMKGWTWTY
h $\alpha$ 3	VDDKTKALLK	YTGEVTWIPP	AIFKSSCKID	VTYFPFDYQN	CTMKFGWSY
h $\alpha$ 5	GT-STKTVIR	YNGTVTWTPP	ANYKSSCTID	VTFPFDLQN	CSMKGFWTY
h $\alpha$ 7	ATFHTNVLVN	SSGHCQYLPP	GIFKSSCYID	VRWFPPFDVQH	CKLKFGWSY
h $\beta$ 1	VALDISVVVS	SDGSVRWQPP	GIYRSSCSIQ	VTYFPFDWQN	CTMVFSYSY
h $\beta$ 2	VSFYSNAVVS	YDGSIFWLPP	AIYKSACKIE	VKHFPPFDQQN	CTMKFRSWTY
mser	SPNIPYVYVH	HRGEVQNYKP	LQLVTACSLD	IYNFPFDVQN	CSLTFTSWLH
r $\alpha$ 2	VTHMTKAHLF	FTGTVHWVPP	AIYKSSCSID	VTFPFWDQQN	CKMKFGSWTY
r $\alpha$ 3	VDDKTKALLK	YTGEVTWIPP	AIFKSSCKID	VTYFPFDYQN	CTMKFGWSY
r $\alpha$ 4	VTHLTKAHLF	YDGRVQWTPP	AIYKSSCSID	VTFPFWDQQN	CTMKFGSWTY
r $\alpha$ 5	GA-STKTVVR	YNGTVTWTQP	ANYKSSCTID	VTFPFDLQN	CSMKFGSWTY
r $\alpha$ 6	VEGKTKALLK	YDGVIWTTPP	AIFKSSCPMD	ITFFPFDHQN	CSLKFGSWTY
r $\alpha$ 7	ATFHTNVLVN	ASGHQCQLPP	GIFKSSCYID	VRWFPPFDVQQ	CKLKFGWSY
r $\beta$ 2	VSFYSNAVVS	YDGSIFWLPP	AIYKSACKIE	VKHFPPFDQQN	CTMKFRSWTY
r $\beta$ 3	GSLMTKAIVK	SSGTVSWTPP	ASYKSSCTMD	VTFPFDRQN	CSMKFGSWTY
r $\beta$ 4	VSVYTNVIVR	SNGSIQWLPP	AIYKSACKIE	VKHFPPFDQQN	CTLKFRSWTY
r $\delta$	ISYACNVLVS	DSGHVTWLPP	AIFRSSCPIS	VTYFPFDWQN	CSLKFSSSLKY

rε	VAYDCNVLVY	EGGSVSWLPP	AIYRSTCAVE	VTYFPFDWQN	CSLIFRSQTY
rγ	VALYCNVLVS	PDGCIYWLPP	AIFRSSCSIS	VTYFPFDWQN	CSLVFQSQTY
rglyc3	EVTTDNKLLR	INGNVLYSIR	LTLTLCSCPMD	LKNFPMDVQT	CIMQLESFGY
sα1	VTTMTKAVLH	HTGKVWWTTP	AIFKSSCEID	VRYFPFDQQT	CFMKFGSWTY
tα1	IVHMTKLLLLD	YTGKIMWTPP	AIFKSYCEII	VTHFPFDQQN	CTMKLGWWTY
tγ	VAYYANVLVY	NDGSMYWLPP	AIYRSTCPIA	VTYFPFDWQN	CSLVFRSQTY
xα1a	IVQETKVLLD	YTGKIIWLPP	AIFKSYCEMI	VTYFPFDLQN	CSMKGTVWTY
xα1b	ISKDTKILLE	YTGKITWTPP	AIFKSYCEII	VTYFPFDQQN	CSMKGTVWTY
xγ	IALYTNTLVS	SDGSMYWLPP	AIYRSSCPVV	VTYFPFDWQN	CSIVFQSQTY
bα1	DGSVVVINPE	SDPDLSNFME	SGEWVIKESR	GWKHWFYAC	CPST--PYLD
bβ1	DSSEVSLQTG	LSIHEGTFFIE	NGQWEIHKP	SRLIQPSVDP	RGGGEGRREE
bδ	TTKEITLSLK	QAIDPEGFTE	NGEWEIVHRP	ARVNVDPS-V	PLDSPNR-QD
bε	NAEEVEVFVA	VDIRTEAYTE	NGEWAIDFCP	G-VIRRHDGD	SAGGPGE-TD
bγ	STNEINLQLS	QEIDPEAFTE	NGEWAIRHRP	AKMLLDEAA-	PAEEAGH-QK
cα2	DKAKIDLENM	EHVDLKDYWE	SGEWAIINAI	GRYNNSKKYDC	CTEI--Y-PD
cα3	DKAKIDLVLI	GSMNLKDYWE	SGEWAIKAP	GYKHDIKYNC	CEEI--Y-TD
cα4	DKAKIDLVSM	HSVDQLDYWE	SGEWEI NAV	GNYNSKKYEC	CTEI--Y-PD
cα5	DGSQVDIILE	DYVDKRDFFD	NGEWEIVTAT	GSKGNRTDGC	CW---Y-PF
cα7	GGWSLDLQMQ	-EADISGYIS	NGEWDLVGIP	GKRTERFYEC	CKEP--Y-PD
cα8	SGWLIDLQML	-EADISNYIS	NGEWDLVGVP	GKRNELYYEC	CKEP--Y-PD
cβ2	DRTEIDLVLK	SEASLDDFTP	SGEWDIVALP	GRRNENPDD-	--ST--Y-VD
cβ4	DHTEIDMVLK	TSASMDDFTP	SGEWDIVALP	GRRTENPLD-	--PN--Y-VD
cγ	SANEINLLLT	VEIDPEAFTE	NGEWAIKHRP	ARKIINSGRF	TPDDIQY-QQ
dα2	DGDQIDLKHI	SGIDLREYYP	SVEWDILGVP	AERHEKYYPC	CAEP--Y-PD
dαLi	DGYMVDLRHL	K-IDLQDYYI	SVEWDIMRVP	AVRNEKFYSC	CEEP--Y-LD
dβ2	NGAQVDLKH	D-IDLTEFYL	SVEWDILEVP	ATKNEEYYPD	TLEP--F-SD
dnachr	NGDQVSLA-L	YNVDSLSDYWK	SGTWIDIEVP	AY-LN VYEGD	SNHP--TETD
gfa3	DKAKIDLVLI	GSVNLKDFWE	SGEWEIIDAP	GYKHDIKYNC	CEEI--Y-PD
gfβ2	DRTEIDLVLT	SDASRDDYTP	SGEWDIVSLP	GRKNEDPND-	--LT--Y-LD
gfnα2	DGNMVKLVL	NQVDRSDFD	NGEWEILSAT	GVKGSRQDSH	LS----Y-PY
gfnα3	DGTMVDLTL	DAVDRKDFFD	NGEWEILNAT	GQRGSRRDG	YS----Y-PY
hα1	DGSVVAINPE	SDPDLSNFME	SGEWVIKESR	GWKHSVTYSC	CPDT--PYLD
hα3	DKAKIDLVLI	GSMNLKDYWE	SGEWAIKAP	GYKHDIKYNC	CEEI--Y-PD
hα5	DGSQVDIILE	DQVDKRDFD	NGEWEIVSAT	GSKGNRTDSC	CW---Y-PY
hα7	GGWSLDLQMQ	-EADISGYIP	NGEWDLVGIP	GKRTERFYEC	CKEP--Y-PD
hβ1	DSSEVLTQTG	LGHEGTFFIE	NGQWENIHKP	SRLIQPPGDP	RGGREGQRQE
hβ2	DRTEIDLVLK	SEASLDDFTP	SGEWDIVALP	GRRNENPDD-	--ST--Y-VD
mser	TIQDINITLW	RRSDKSIFIN	QGEWELLEV-	--FPQFKEF	SIDISNSYAE
rα2	DKAKIDLEQM	ERVDLKDYWE	SGEWAIINAT	GTYNNSKKYDC	CAEI--Y-PD
rα3	DKAKIDLVLI	GSMNLKDYWE	SGEWAIKAP	GYKHEIKYNC	CEEI--Y-QD
rα4	DKAKIDLVS	HSDQLDFWE	SGEWIVDAV	GTYNTRKYEC	CAEI--Y-PD
rα5	DGSQVDIILE	DQVRTDFFD	NGEWEIMSAM	GSKGNRTDSC	CW---Y-PY
rα6	DKAEIDLII	GSVDMNDFWE	NSEWEIVDAS	GYKHDIKYNC	CEEI--Y-TD
rα7	GGWSLDLQMQ	-EADISSYIP	NGEWDLMGIP	GKRNEKFYEC	CKEP--Y-PR
rβ2	DRTEIDLVLK	SDASLDDFTP	SGEWDIAILP	GRRNENPDD-	--ST--Y-VD
rβ3	DGTMVDLILI	NEVDRKDFD	NGEWEILNAK	GMKGNRREGF	YS----Y-PF
rβ4	DHTEIDMVLK	SPAIDMDDFTP	SGEWDIVALP	GRRTVNPQD-	--PS--Y-VD
rδ	TAKEIRLSK	QEIDPEGFTE	NGEWEIVHRA	AKVNVDPS-V	PMDSTNH-QD
rε	NAEEVELIFA	VDIRTAATFE	NGEWAIDYCP	G-MIRHYEGG	STEDPGE-TD
rγ	STSEINLQLS	QEIDPEAFTE	NGEWAIRHRP	AKMLLDPVT-	PAEEAGH-QK
rglyc3	TMNDLIFEWQ	DEAPVQQFL	KEEKDLRYCT	KHYNTGKFTC	-----
sα1	DGDQIDLKHI	N-IDLREYYP	SVEWDILGVP	AERHEKYYPC	CAEP--Y-PD
tα1	DGTVKSISPE	SDPDLSTFME	SGEWVMKDYR	GWKHWVYYTC	CPDT--PYLD
tγ	NAHEVNLQLS	AEDPEDFTE	NGEWTIRHRP	AKKNYNW-QL	TKDDTDF-QE
xα1a	DGTLVVINPE	NDPDLSNFME	SGEWYMKDYR	CWKHWVYYDC	CPET--PYLD
xα1b	DGSLLVINPE	RDPDLSNFMA	SGEWMMKDYR	CWKHWVYYTC	CPDK--PYLD
xγ	SANEIELLLT	VDIRTAATFE	NGEWAIKHMP	AKRIINH-RL	PRDDVNY-QQ
bα1	ITYHFVMQRL	PLYFIVNVII	PCLLFSFLTG	LVFYLPTDSG	-EKMTLSISV
bβ1	VTFYLIIRRK	PLFYLVNVIA	PCILITLLAI	FVFYLPDPAG	-EKMGSLISA
bδ	VTFYLIIRRK	PLFYVINILV	PCVLISFMIN	LVFYL PADCG	-EKTSMAISV
bε	VIYSLIIRRK	PLFYVINII	PCVLISGLVL	LAYFLPAQAG	GQKCTVAINV
cα2	ITFYFVIRRL	PLFYTINLII	PCLLISCLTV	LVFYLPSDCG	-EKITLCISV
cα3	ITYSLYIRRL	PLFYTINMII	PCLLISFLTV	LVFYLPSDCG	-EKVTL CISV
cα4	ITYSFIRRL	PLFYTINLII	PCLLISCLTV	LVFYLPSSEC	-EKITLCISV
cα5	VTYSFIIRRL	PLFYTLFLII	PCIGLSFLTV	LVFYLPSNEA	-EKISLCTSV
cα7	ITFTVTMRRR	TLYYGLNLLI	PCVLISALAL	LVFLLPADSG	-EKISLGITV

c $\alpha$ 8	VTYTITMRRR	TLYYGLNLLI	PCVLISGLAL	LVFLLPADSG	-EKISLGITV
c $\beta$ 2	ITYDFIIRRK	PLFYTIINLII	PCILITSLAI	LVFYLPSDCG	-EKMTLCISV
c $\beta$ 4	VTYDFIIRRK	PLFYTIINLII	PCVLITSLAI	LVFYLPSDCG	-EKMTLCISV
c $\gamma$	VIFYLIQRK	PLFYIINIIV	PCVLISMAV	LVYFLPAKAG	GQKCTVSINV
d $\alpha$ 2	IFFNITLRRK	TLFYTVNLII	PCVGISYLSV	LVFYLPSDCG	-EKIALCISI
d $\alpha$ Li	IVFNLTLLRK	TLFYTVNLII	PCVGISFLSV	LVFYLPSDCG	-EKISLCISI
d $\beta$ 2	ITFKLTMRRK	TLFYTVNLIV	PCVALTFLTV	LVFYLPSDCG	-EKVTLCCISI
dnachr	ITYFIIRRK	TLFYTVNLIL	PTVLISFLCV	LVFYLPAEAG	-EKVTLGISI
g $\alpha$ 3	ITYSFYIRRL	PLFYTIINLII	PCLLISFLTI	LVFYLPSDCG	-EKVTLCCISV
g $\beta$ 2	ITYDFVIKRK	PLFYTIINLII	PCVLITSLAI	LVFYLPSDCG	-EKVTLCCMSV
gfno2	ITYSFILKRL	PLFYTLFLII	PCLGLSFLTV	LVFYLPSDEG	-EKVSLSTSV
gfno3	VTYSFILKRL	PLFYTLFLII	PCLGLSFLTV	LVFYLPSDEG	-EKLLLSTSV
ha1	ITYHFVMQRL	PLYFIVNVII	PCLLFSFLTG	LVFYLPTDSCG	-EKMTLSISV
ha3	ITYSLYIRRL	PLFYTIINLII	PCLLISFLTV	LVFYLPSDCG	-EKVTLCCISV
ha5	VTYSFVIKRK	PLFYTLFLII	PCIGLSFLTV	LVFYLPSNEG	-EKICLCTSV
ha7	VTFTVTMRRR	TLYYGLNLLI	PCVLISALAL	LVFLLPADSG	-EKISLGITV
h $\beta$ 1	VIFYLIIRRK	PLFYLVNVIA	PCILITLLAI	FVFYLPPDAG	-EKMGSLIFA
h $\beta$ 2	ITYDFIIRRK	PLFYTIINLII	PCVLITSLAI	LVFYLPSDCG	-EKMTLCISV
mser	MKFYVIIRR	PLFYAVSLLL	PSIFLMVVDI	VGFCFLPPDSCG	-ERVSFKITL
r $\alpha$ 2	VTYYFVIRRL	PLFYTIINLII	PCLLISCLTV	LVFYLPSSEC	-EKITLCISV
r $\alpha$ 3	ITYSLYIRRL	PLFYTIINLII	PCLLISFLTV	LVFYLPSDCG	-EKVTLCCISV
r $\alpha$ 4	ITYAFIIRR	PLFYTIINLII	PCLLISCLTV	LVFYLPSSEC	-EKVTLCCISV
r $\alpha$ 5	ITYSFVIKRK	PLFYTLFLII	PCIGLSFLTV	VVFYLPSNEG	-EKISLCISV
r $\alpha$ 6	ITYSFYIRRL	PMFYTIINLII	PCLFISFLTV	LVFYLPSDCG	-EKVTLCCISV
r $\alpha$ 7	CTYTVTMRR	TLYYGLNLLI	PCVLISALAL	LVFLLPADSG	-EKISLGITV
r $\beta$ 2	ITYDFIIRRK	PLFYTIINLII	PCVLITSLAI	LVFYLPSDCG	-EKMTLCISV
r $\beta$ 3	VTYSFVLRRL	PLFYTLFLII	PCLGLSFLTV	LVFYLPSDEG	-EKLSLSTSV
r $\beta$ 4	VTYDFIIRR	ALFYTIINLII	PCVLITSLAI	LVFYLPSDCG	-EKMTLCISV
r $\delta$	VTFYLIIRR	PLFYIINIIV	PCVLISFMIN	LVFYLPGDCG	-EKTSVAISV
re	VIYTLIIRR	PLFYVINIIV	PCVLISGLVL	LAYFLPAQAG	GQKCTVSINV
r $\gamma$	VVFLYLIQRK	PLFYVINIIV	PCVLISSSVAI	LIYFLPAKAG	GQKCTVATNV
rgly $\alpha$ 3	IEVRFHRLQ	MGYYLIQMYI	PSLLLIVLW	VSFWINMDAA	PARVALGITT
s $\alpha$ 1	IFFNITLRRK	TLFYTVNLIV	PCVGISYLSV	LVFYLPAEAG	-EKIALCISI
t $\alpha$ 1	ITYHFIMQRI	PLYFVNVII	PCLLFSFLTG	LVFYLPTDSCG	-EKMTLSISV
t $\gamma$	IIFFLIQRK	PLFYIINIIA	PCVLISSLVV	LVYFLPAQAG	GQKCTLISIV
x $\alpha$ 1a	ITYHFLQRL	PLYFIVNVII	PCLLFSFLTG	LVFYLPTDSCG	-EKITLVSIV
x $\alpha$ 1b	ITYHFVQLRL	PLYFIVNVII	PCLLFSFLTG	LVFYLPTDSCG	-EKMTLSISV
x $\gamma$	IVFYLIQRK	PLFYIINIIV	PCVLISFVSI	LVYFLPAKAG	GQKCTVSINI
b $\alpha$ 1	LLSLTVFLLV	IVELIPSTSS	AVPLIGKYML	FTMVFVIASI	IITVIVINTH
b $\beta$ 1	LLTLTVFLLL	LADKVPETSL	SPVIIKYL	FTMVLVTFSV	ILSVVVLNLH
b $\delta$	LLAQSVFLLL	ISKRLPATSM	AIPLIGKFL	FGMVLVTMVV	VICVIVLNH
b $\epsilon$	LLAQTVFLFL	IAQKTPETSL	SPVLLGRYLI	FVMVVATLIV	MNCVIVLNVS
b $\gamma$	LLAQTVFLFL	VAKKVPETSQ	AVPLISKYLT	FLLVVITLIV	VNAVVVLNVS
c $\alpha$ 2	LLSLTVFLL	ITEIIPSTSL	VIPLIGEYLL	FTMIFVTLSI	IITVFLNVH
c $\alpha$ 3	LLSLTVFLLV	ITETIPSTSL	VIPLIGEYLL	FTMIFVTLSI	VITVFVFLNVH
c $\alpha$ 4	LLSLTVFLL	ITEIIPSTSL	VIPLIGEYLL	FTMIFVTLSI	IITVFLNVH
c $\alpha$ 5	LVSLTVFLLV	IEIIIPSSSK	VIPLIGEYLV	FTMIFVTLSI	VITVFVFLNVH
c $\alpha$ 7	LLSLTVFMLL	VAEIMPATSD	SVPLIAQYFA	STMIIVGLSV	VVTVIVLQYH
c $\alpha$ 8	LLSLTVFMLL	VAEIMPATSD	SVPLIAQYFA	SIMVIVGLSV	VVTVFLQFH
c $\beta$ 2	LLALTIVFLL	ISKIVPPTSL	DVPLVGKYLM	FTMVLVTFSI	VTSVCVFLNVH
c $\beta$ 4	LLALTIVFLL	ISKIVPPTSL	DVPLIGKYLM	FTMVLVTFSI	VTSVCVFLNVH
c $\gamma$	LLAQTVFLFL	IAQKVPETSQ	AVPLIGKYLT	FLMVVTVVIV	VNAVIVLNVS
d $\alpha$ 2	LLSQTMFLL	ISEIIPSTSL	ALPLLGKYLL	FTMILLVGLSV	VITIIIILNH
d $\alpha$ Li	LLSLTVFFLL	LAEEIPPTSL	TVPLLGKYLL	FTMMLVTLFSV	VVTIAVLNVN
d $\beta$ 2	LVSLTVFFLL	LAEEIPPTSL	AVPLLGKYLL	FTMILVSLSV	WTTCVFLNIH
dnachr	LLSLVVFL	VSKILPPTSL	VLPLIAKYLL	FTFIMNTVSI	LVTVIIINWN
g $\alpha$ 3	LLSLTVFLLV	ITETIPSTSL	VIPLIGEYLL	FTMIFVTLSI	VITVFVFLNVH
g $\beta$ 2	LLALTIVFLL	ISKIVPPTSL	AVPLIGKYLM	FTMVLVTFSI	VTSVCVFLNVH
gfno2	LVSLTVFLLV	IEIIIPSSSK	VIPLIGEYLL	FIMIFVTLSI	IVTIFVINH
gfno3	LVSLTVFLLV	IEIIIPSSSK	VIPLIGEYLL	FIMIFVTFSI	IVTFLFVINH
h $\alpha$ 1	LLSLTVFLLV	IVELIPSTSS	AVPLIGKYML	FTMVFVIASI	IITVIVINTH
h $\alpha$ 3	LLSLTVFLLV	ITETIPSTSL	VIPLIGEYLL	FTMIFVTLSI	VITVFVFLNVH
h $\alpha$ 5	LVSLTVFLLV	IEIIIPSSSK	VIPLIGEYLV	FTMIFVTLSI	MVTVFAINIH
h $\alpha$ 7	LLSLTVFMLL	VAEIMPATSD	SVPLIAQYFA	STMIIVGLSV	VVTVIVLQYH
h $\beta$ 1	LLALTIVFLL	LADKVPETSL	SPVIIKYL	FTMVLVTFSV	ILSVVVLNLH
h $\beta$ 2	LLALTIVFLL	ISKIVPPTSL	DVPLVGKYLM	FTMVLVTFSI	VTSVCVFLNVH
mser	LLGYSVFLII	VSDTLPAT-I	GTPLIGVYFV	VCMALLVISL	AETIFIVRLV
r $\alpha$ 2	LLSLTVFLL	ITEIIPSTSL	VIPLIGEYLL	FTMIFVTLSI	VITVFVFLNVH

r $\alpha$ 3	LLSLTVFLLV	ITETIPSTSL	VIPLIGEYLL	FTMIFVTLSI	VITVFVLNVH
r $\alpha$ 4	LLSLTVFLLL	ITEIIIPSTSL	VIPLIGEYLL	FTMIFVTLSI	VITVFVLNVH
r $\alpha$ 5	LVSLTVFLLV	IEEIIPSSSK	VIPLIGEYLV	FTMIFVTLSI	MVTVFAINIH
r $\alpha$ 6	LLSLTVFLLV	ITETIPSTSL	VIPLVGLEYLL	FTMIFVTLSI	VVTVFVLNIH
r $\alpha$ 7	LLSLTVFMLL	VAEIMPATSD	SVPLIAQYLP	STMIIVGLSV	VVTIVLRYH
r $\beta$ 2	LLALTVFLLL	ISKIVPPPTSL	DVPLVGKYLM	FTMVLVTFSI	VTSVCVLNVH
r $\beta$ 3	LVSLTVFLLV	IEEIIPSSSK	VIPLIGEYLL	FIMIFVTLSI	IVTVFVINHV
r $\beta$ 4	LLALTTFLLL	ISKIVPPPTSL	DIPLIGKYLL	FTMVLVTFSI	VTTVCVLNVH
r $\delta$	LLAQSVFLLL	ISKRKPATSM	AIPLVGKFLL	FGMVLVTMVV	VICVIVLNH
r $\epsilon$	LLAQTVFLFL	IAQKIPETSL	SVPLLGRYLI	FVMVVATLIV	MNCVIVLNVS
r $\gamma$	LLAQTVFLFL	VAKVPETSQ	AVPLISKYLT	FLMVVTILIV	VNSVVVLNVS
rgly $\alpha$ 3	VLTMTTQSSG	SRAASPKVSY	-VKAIDIWMA	VCLLFV-FSA	LLEYAAVNFI
s $\alpha$ 1	LLSQTMFFLL	ISEIIIPSTSL	ALPLLGKYLL	FTMVLVGLSV	VITIMVLNVH
t $\alpha$ 1	LLSLTVFLLV	IVELIPSTSS	AVPLIGKYM	FTMIFVISSI	IITVVVINTH
t $\gamma$	LRAQTVFLFL	IAQKVPETSL	SVFMVFSMLIV	MNCVIVLNVS	
x $\alpha$ 1a	LLSLVVFLLV	IVELIPSTSS	AVPLIGKYL	FVMFVSIASI	VITVIVINTH
x $\alpha$ 1b	LLSLTVFLLV	IVELIPSTSS	AVPLIGKYM	FTMVFVIASI	IITVIVINTH
x $\gamma$	LRAQTVFLFL	VAQKIPETST	SVPLIVKYLT	FLMVVTITIV	ANAVIVLNIS
b $\alpha$ 1	HRSPST-HVM	PEWVRKVFD	TIPNIMFFST	MKRP--PEVK	SAIEGIKYIA
b $\beta$ 1	HRSPHT-HQM	PLWVRQIFIH	KLPLYL---G	LKRPLPPELR	EVVSSISYIA
b $\delta$	FRTPST-HVL	SEPVKKLFLE	TLPEIL---H	MSRPLFSELK	PAVDGANFIV
b $\epsilon$	LRTPTT-HAM	SPRLRYVVLLE	LLPQLL---G	SGAPAAPEIR	CCVDAVNFI
b $\gamma$	LRSPHT-HSM	ARGVRKVFLR	LLPQLL---R	MHVRAAPAIQ	ACVEACNLIA
c $\alpha$ 2	HRSPST-HTM	PHWRSFFLG	FIPRWLFFF	MKRLPSPSIL	RALEGVQYIA
c $\alpha$ 3	YRTPKT-HTM	PVWVRTIFLN	LLPRIMF---	MTRPLSPEMR	DAIESVKYIA
c $\alpha$ 4	HRSPRT-HTM	PDWVRRVFLD	IVPRLLF---	MKRPMSPALK	LAEGVHYIA
c $\alpha$ 5	HRSSSTHNAM	APWVRKIFLH	KLPKLLC---	MRS---	LE AALDSIRYIT
c $\alpha$ 7	HHDPDG-GKM	PKWTRVILLN	WCAWFL---	MKRPGDPLA	KILEEVRYIA
c $\alpha$ 8	HHDPAQ-GKM	PRWVRVILLN	WCAWFL---	MKKPTIPVIV	KILEEVQFIA
c $\beta$ 2	HRSPPTT-HTM	PPWVRTLFLR	KLPALLF---	MKQ---	PGLE EAEGVRFIA
c $\beta$ 4	HRSPST-HTM	PPWVKLVFLE	RLPAYLF---	MKR---	PEVQ EAIDGVSFIA
c $\gamma$	LRTPN-T-HSM	SQRVRQVWLH	LLPRYL---	G MHMPASPEIR	ACVEACNHIA
d $\alpha$ 2	YRKPST-HKM	RPIWRSFFIK	RLPKLL---	LMR---	VELE KAIHNVMFIQ
d $\alpha$ Li	FRSPVT-HRM	APWVQRLFIQ	ILPKLLC---	IER---	PEME KTIEGSRFIA
d $\beta$ 2	FRSPST-HNM	SRLVRKLFHL	FMPKLM---	MRR---	TEVL QALRAVRFIA
dnachr	FRGPRT-HRM	PMYIRSIFLH	YLP AFLF---	MKR---	PEAS KATEAVEFIA
g $\alpha$ 3	YRTPMT-HTM	PSWVRTVFLR	ALPRVML---	MRRPVSPEIK	QAIESVKYIA
g $\beta$ 2	HRSPST-HYM	PEWVKCVFLH	KLPAFLL---	MRR---	PDVD EAIDGVRFIA
gfna2	HRSSATYHPM	SPWVRSLSFLQ	RLPHLLC---	MRGNLINLLE	QATNSVRYIS
gfna3	HRSSATYHPM	APWVKSLFLQ	RLPRLLC---	MRGHWIALLE	KATHSVHYIS
h $\alpha$ 1	HRSPST-HVM	PNWVRKVFD	TIPNIMFFST	MKRP--PEVK	SAIEGIKYIA
h $\alpha$ 3	YRTPTT-HTM	PSWVKTVFLN	LLPRVMF---	MTRPLSPEIK	EAIQSVKYIA
h $\alpha$ 5	HRSSSTHNAM	APLVRKIFLH	TLPKLLS---	MRS---	LE AALDSIRYIT
h $\alpha$ 7	HHDPDG-GKM	PKWTRVILLN	WCAWFL---	MKRPGDPLA	KILEEVRYIA
h $\beta$ 1	HRSPHT-HQM	PLWVRQIFIH	KLPLYL---	R	LKRPLPPELR EVVSSISYIA
h $\beta$ 2	HRSPTT-HTM	APWVKVVFLE	KLPALLF---	MQQ---	PGLR EAEGVRFIA
mser	HKQ-DLQRPV	PDWLRHLVLD	RIAWILC---	LPREASLAVR	GLLQELSSIR
r $\alpha$ 2	HRSPST-HNM	PNWVRVALLG	RVPRWLM---	MNRPLSPQIQ	KALEGVHYIA
r $\alpha$ 3	YRTPTT-HTM	PTWKAVFLN	LLPRVMF---	MTRPLSPEIK	EAIQSVKYIA
r $\alpha$ 4	HRSPRT-HTM	PAWVRRVFLD	IVPRLLF---	MKRP-SPALT	RAEGVQYIA
r $\alpha$ 5	HRSSSTHNAM	APWVRKIFLH	KLPKLLC---	MRS---	LE AALDCIRYIT
r $\alpha$ 6	YRTPAT-HTM	PKWVKTMFQ	VFP S ILM---	MRRPHPPDVE	DVIDSVQFIA
r $\alpha$ 7	HHDPDG-GKM	PKWTRIILLN	WCAWFL---	MKRPGDPLA	KILEEVRYMP
r $\beta$ 2	HRSPTT-HTM	APWVKVVFLE	KLPTLLF---	LQQ---	PGLR EAEGVRFIA
r $\beta$ 3	HRSSSTYHPM	APWVKRLFLQ	RLPRWLC---	MKDPLVAFLE	KASESIRYIS
r $\beta$ 4	HRSPST-HTM	ASWVKECFHL	KLPTFLF---	MKR---	PDLQ EAEGVSFIA
r $\delta$	FRTPST-HVL	SEGVKKFFLE	TLPKLL---	H	MSRPLFNEMK PAVDGANFIV
r $\epsilon$	LRTPTT-HAT	SPRLRQILLE	LLPRL---	G LSPPAPEVR	CCVDAVNFI
r $\gamma$	LRSPHT-HSM	ARGVRKVFLR	LLPQLL---	R MHVHASPAIQ	ACVDACNLMA
rgly $\alpha$ 3	SRQHKELRKR	KNKTEAFALE	KFYRFLSFTA	YG--MGPCLO	AK-DGVVPKG
s $\alpha$ 1	YRKPST-HKM	APWVRKVFLR	RLPKLL---	LMR---	VELE KAIHNVLFIQ
t $\alpha$ 1	HRSPST-HTM	PQWVRKIFID	TIPNVMFFST	MKRA--PDVK	SAIEGIKYIA
t $\gamma$	LRTPN-T-HSL	SEKIKHLFLG	FPLKYL---	G MQLEFAPEIK	SCVEACNFIA
x $\alpha$ 1a	HRSPST-HIM	PQWLKKIFIE	TIPRVMFFST	MKRP--PDVK	SAIEGAKYVA
x $\alpha$ 1b	HRSPST-HTM	PPWVRKIFIE	TIPNIMFFST	MKRP--PDVK	SAIEGIKYIA
x $\gamma$	LRTPN-T-HSM	SSTVRELCLR	TVPRLL---	R MHLRAAPEIR	TCVEACCHIA

b $\alpha$ 1 LLSLTVFLLV IVELIPSTSS AVPLIGKYML FTMVFVIASI IITVIVINTH  
 b $\beta$ 1 LLTLCVFLLL LADKVPETSL SVPIIIKYLM FTMVLVTFSV ILSVVVNLH  
 b $\delta$  LLAQSVFLLL ISKRLPATSM AIPLIGKFLL FGMVLVTMVV VICVIVLNH  
 b $\epsilon$  LLAQTVFLFL IAQKTPETSL SVPLLGRYLI FVMVVATLIV MNCVIVLNVS  
 b $\gamma$  LLAQTVFLFL VAKKVPETSQ AVPLISKYLT FLLVVTILIV VNAVVLNVS  
 c $\alpha$ 2 LLSLTVFLLL ITEIIPSTSL VIPLIGEYLL FTMIFVTLSI IITVFVLNVH  
 c $\alpha$ 3 LLSLTVFLLV ITETIPSTSL VIPLIGEYLL FTMIFVTLSI VITVFVLNVH  
 c $\alpha$ 4 LLSLTVFLLL ITEIIPSTSL VIPLIGEYLL FTMIFVTLSI IITVFVLNVH  
 c $\alpha$ 5 LVSLTVFLLV IEEIIPSSSK VIPLIGEYLV FTMIFVTLSI VITVFAINIH  
 c $\alpha$ 7 LLSLTVFMLL VAEIMPATSD SVPLIAQYFA STMIIIVGLSV VVTVIVLQYH  
 c $\alpha$ 8 LLSLTVFMLL VAEIMPATSD SVPLIAQYFA SIMVIVGLSV VVTVLVLQFH  
 c $\beta$ 2 LLALTVFLLL ISKIVPPPTSL DVPLVGKYLM FTMVLVTFSI VTSVCVLNVH  
 c $\beta$ 4 LLALTVFLLL ISKIVPPPTSL DVPLIGKYLM FTMVLVTFSI VTSVCVLNVH  
 c $\gamma$  LLAQTVFLFL IAQKVPETSL AVPLIGKYLT FLMVVTVVIV VNAVVLNVS  
 d $\alpha$ 2 LLSQTMFLL ISEIIPSTSL ALPLLGKYLL FTMLLVGLSV VITIIILNIH  
 d $\alpha$ L LLSLTFFFLL LAEIIIPPTSL TVPLLGKYLL FTMLLVTLSV VVTIAVLNVN  
 d $\beta$ 2 LVSLTVFFFLL LAEIIIPPTSL AVPLLGKYLL FTMLLVLSV WTTVCVLNIH  
 dnachr LLSLTVVFLLL VSKILPPPTSL VLPLIAKYLL FTFIMNTVSI LVTVIIINWN  
 g $\alpha$ 3 LLSLTVFLLV ITETIPSTSL VIPLIGEYLL FTMIFVTLSI VITVFVLNVH  
 g $\beta$ 2 LLALTVFLLL ISKIVPPPTSL AVPLIGKYLM FTMVLVTFSI VTSVCVLNVH  
 gfn $\alpha$ 2 LVSLTVFLLV IEEIIPSSSK VIPLIGEYLL FIMIFVTLSI IVTIFVINVH  
 gfn $\alpha$ 3 LVSLTVFLLV IEEIIPSSSK VIPLIGEYLL FIMIFVTFSI IVTLFVINVH  
 h $\alpha$ 1 LLSLTVFLLV IVELIPSTSS AVPLIGKYML FTMVFVIASI IITVIVINTH  
 h $\alpha$ 3 LLSLTVFLLV ITETIPSTSL VIPLIGEYLL FTMIFVTLSI VITVFVLNVH  
 h $\alpha$ 5 LVSLTVFLLV IEEIIPSSSK VIPLIGEYLV FTMIFVTLSI MVTVFAINIH  
 h $\alpha$ 7 LLSLTVFMLL VAEIMPATSD SVPLIAQYFA STMIIIVGLSV VVTVIVLQYH  
 h $\beta$ 1 LLTLCVFLLL LADKVPETSL SVPIIIKYLM FTMVLVTFSV ILSVVVNLH  
 h $\beta$ 2 LLALTVFLLL ISKIVPPPTSL DVPLVGKYLM FTMVLVTFSI VTSVCVLNVH  
 mser LLGYSVFLII VSDTLPAT-I GTPLIGVYFV VCMALLVSL AETIFIVRLV  
 r $\alpha$ 2 LLSLTVFLLL ITEIIPSTSL VIPLIGEYLL FTMIFVTLSI VITVFVLNVH  
 r $\alpha$ 3 LLSLTVFLLV ITETIPSTSL VIPLIGEYLL FTMIFVTLSI VITVFVLNVH  
 r $\alpha$ 4 LLSLTVFLLL ITEIIPSTSL VIPLIGEYLL FTMIFVTLSI VITVFVLNVH  
 r $\alpha$ 5 LVSLTVFLLV IEEIIPSSSK VIPLIGEYLV FTMIFVTLSI MVTVFAINIH  
 r $\alpha$ 6 LLSLTVFLLV ITETIPSTSL VIPLVGEYLL FTMIFVTLSI VVTVFVLNIH  
 r $\alpha$ 7 LLSLTVFMLL VAEIMPATSD SVPLIAQYLP STMIIIVGLSV VVTVIVLRYH  
 r $\beta$ 2 LLALTVFLLL ISKIVPPPTSL DVPLVGKYLM FTMVLVTFSI VTSVCVLNVH  
 r $\beta$ 3 LVSLTVFLLV IEEIIPSSSK VIPLIGEYLL FIMIFVTLSI IVTVFVINVH  
 r $\beta$ 4 LLALTFFLLL ISKIVPPPTSL DIPLIGKYLL FTMVLVTFSI VTTVCVLNVH  
 r $\delta$  LLAQSVFLLL ISKRLPATSM AIPLVGKFLL FGMVLVTMVV VICVIVLNH  
 re LLAQTVFLFL IAQKIPETSL SVPLLGRYLI FVMVVATLIV MNCVIVLNVS  
 r $\gamma$  LLAQTVFLFL VAKKVPETSQ AVPLISKYLT FLMVVVTILIV VNSVVVLNVS  
 rgly $\alpha$ 3 VLTMTTQSSG SRASLPKVSY -VKAIDIWMA VCLLFV-FSA LLEYAAVN  
 sc $\alpha$ 1 LLSQTMFFLL ISEIIPSTSL ALPLLGKYLL FTMLLVGLSV VITIMVLNVH  
 tc $\alpha$ 1 LLSLTVFLLV IVELIPSTSS AVPLIGKYML FTMIFVISSI IITVVVINTH  
 t $\gamma$  LLAQTIFLFL IAQKVPETSL NVPLIGKYLI FVMFVSMILIV MNCVIVLNVS  
 x $\alpha$ 1a LLSLTVVFLLV IVELIPSTSS AVPLIGKYML FTMVFVIASI VITVIVINTH  
 x $\alpha$ 1b LLSLTVFLLV IVELIPSTSS AVPLIGKYML FTMVFVIASI IITVIVINTH  
 x $\gamma$  LLAQTVFLFL VAQKIPETST SVPLIVKYLT FLMVVVTITIV ANAVIVLNIS

**Appendix 2:** Continued

c $\alpha$ 8	IIDVDEKNQV	LITNAWLQMY	WVDIYLSWDQ	YEYPGVQNLR	FPSDQIWVPD
d $\alpha$ 2	LIDLNLKDQI	LTINNVLEHE	WQDHFKWKDP	SEYGGVTELY	VPSEHIWLPD
d $\alpha$ Li	LIDVNLKNQI	MTTNVWVEQE	WNDYKLKWNP	DDYGGVDTLH	VPSEHIWHPD
d $\beta$ 2	LIEVNLKNQV	MTTNLWVKQR	WFDYKLRWDP	EYGGVEQLY	VPSEHIWVPD
dnachr	LINVNEKNQV	MKSNNWLRLV	WYDYQLQWDE	ADYGGIVLRL	LPPDKVWKPD
mser	IILNVDEKNQV	LTTYIWYRQY	WTDEFQWTP	EDFDNVTKLS	IPTDSIWVPD
onachr	IIDVHEIDQI	MTCSVWLKVQ	WIDKKLSWNP	EIYGGVSVL	VPYEMVWVPD
r $\alpha$ 2	LIDVDEKNQV	MTTNVWLNQK	WNDYNVRWDP	AEGFNVTSLR	VPSEMIWIPD
r $\alpha$ 3	LVKDEVNQI	METNLWLKQI	WNDYKLKWKP	SDYQGVEFMR	VPAEKIWKP
r $\alpha$ 7	IIMDVDEKNQV	LTNTNIWLQMS	WTDHYLQWNM	SEYPGVKNVR	FPDGQIWKP
r $\beta$ 2	LISVHEREQI	MTTNVWLTQE	WEDYRLTWKP	QHFDNMKVRL	LPSKHIWLPD
r $\delta$	LISLKEVEET	LTINNVWIDHA	WIDSRLQWNA	NEFGNITVLR	LPSDMVWLPE
r $\gamma$	LISLNEREAA	LTINNVWIEMQ	WCDYRLRWDP	KDYEGLWILR	VPSTMVWQPD
c $\alpha$ 8	ILLYNSADER	FDATFHTNVL	VNYSGSCQYI	PPGILKSTCY	IDVRWFPFDV
d $\alpha$ 2	IVLYNNADGE	YVVTMTKAI	LHYTGKVVWT	PPAIFKSSCE	IDVRYFPFDQ
d $\alpha$ Li	IVLYNNADGN	YEVTIMTKAI	LHHTGKVVWK	PPAIYKSFCE	IDVEYFPFD
d $\beta$ 2	IVLYNNWDGN	YEVTLMKTAK	LKYTGEVFWE	PPAIYKSSCE	MNVEYFPYDE
dnachr	IVLFNNADGN	YEVRYKSNVL	IYPTGEVLWV	PPAIYQSSCT	IDVTYFPFDQ
mser	ILINEFVDVG	-KSPNIPYVY	VHHRGEVQNY	KPLQLVTACS	LDIYNFPFDV
onachr	IVLYNTVDSN	YINITISTKAT	LRYDGQVTWD	SPAIFKTLQ	IDVRWFPFD
r $\alpha$ 2	IVLYNNADGE	FAVTHMTKAH	LFFGTGTVHWV	PPAIYKSSCS	IDVTFFPFQ
r $\alpha$ 3	IVLYNNADGD	FQVDDKTKAL	LKYTGEVWTI	PPAIFKSSCK	IDVTYFPFDY
r $\alpha$ 7	ILLYNSADER	FDATFHTNVL	VNASGHCQYL	PPGIFKSSCY	IDVRWFPFDV
r $\beta$ 2	VVLYNNADGM	YEVSFYNSNAV	VSYDGSIFWL	PPAIYKSACK	IEVKHFPFDQ
r $\delta$	IVLENNNDGS	FQISYACNVL	VSDSGHVTWL	PPAIFRSSCP	ISVTYFPFDW
r $\gamma$	IVLGNNVDGV	FEVALYCNVL	VSPDGCYIWL	PPAIFRSSCS	ISVTYFPFDW
c $\alpha$ 8	QKCDLKFGSW	THSGWLIDLQ	MLEA-DISNY	ISNGEWDLVG	VPGKRNELYY
d $\alpha$ 2	QTCFMKFGSW	TYDGDQIDLK	HISGIDLREY	YPSVEWDILG	VPAERHEKYY
d $\alpha$ Li	QTCFMKFGSW	TYDGYMVDLR	HLK-IDLQDY	YISVEWDIMR	VPAVRNEKFY
d $\beta$ 2	QICFMKFGSW	TYNGAQVDLK	HL-DIDLTEF	YLSVEWDILE	VPATKNEEYY
dnachr	QTCIMKFGSW	TFNGDQVSLA	LYN-VDLSDY	WKGSTWDIIE	VPAYLNV--Y
mser	QNCSLTFTSW	LHTIQDINIT	LWRRSDKSIF	INQGEWELLE	VFPQFKEFSI
onachr	QNCHFKFGSW	TYTEDLLNLE	LLDSIDLSDY	YPSVEWDIMS	RIAKRRTKNY
r $\alpha$ 2	QNCKMKFGSW	TYDKAKIDLE	QMERVDLKDY	WESGEWAIIN	ATGTYNSKKY
r $\alpha$ 3	QNCTMKFGSW	SYDKAKIDLV	LIGSMNLKDY	WESGEWAIIK	APGYKHEIKY
r $\alpha$ 7	QQCKLKFGSW	SYGGWSLDLQ	MQEA-DISSY	IPNGEWDLMG	IPGKRNEKFY
r $\beta$ 2	QNCTMKFRSW	TYDRTEIDLV	LKSDASLDDF	TPSGEWDIIA	LPGRRNENPD
r $\delta$	QNCSLKFSLL	KYTAKEIRLS	LKQEIDPEGF	TENGGEWEIVH	RAAKVNVDP-
r $\gamma$	QNCSLVFQSQ	TYSTSEINLQ	LSQEIDPEAF	TENGGEWAIRH	RPAKMLLDP-
c $\alpha$ 8	E--CCKEPYP	DVTYTITMRR	RTLYYGLNLL	IPCVLISGLA	LLVFLLPADS
d $\alpha$ 2	--PCCAEPYP	DIFFNITLRR	KTLFYTVNLI	IPCVGISYLS	VLVFYL PADS
d $\alpha$ Li	--SCCEEPLY	DIVFNLTLRR	KTLFYTVNLI	IPCVGISFLS	VLVFYLPSDS
d $\beta$ 2	--PDTLEPFS	DITFKLTMRR	KTLFYTVNLI	VPCVALTFLT	VLVFYLPSDS
dnachr	EGDSNHPTET	DITFYIIIIRR	KTLFYTVNLI	LPTVLISFLC	VLVFYLPAEA
mser	DIS---NSYA	EMKFYVIIRR	RPLFYAVSLL	LPSIFLMVVD	IVGFCLPPDS
onachr	LTSFSDEAFI	DIIFYLELRR	KPLFYTVNVL	FPCVGISFLT	IVAFYLPFRS
r $\alpha$ 2	--DCCAEIYP	DVTYYFVIRR	LPLFYTIINLI	IPCLLISCLT	VLVFYLPSSEC
r $\alpha$ 3	--NCCEEIYQ	DITYSLYIIRR	LPLFYTIINLI	IPCLLISFLT	VLVFYLPSDC
r $\alpha$ 7	E--CCKEPYP	RCTYTVTMRR	TTLYYGLNLL	IPCVLISALA	LLVFLLPADS
r $\beta$ 2	--DS---TYV	DITYDFIIRR	KPLFYTIINLI	IPCVLITS LA	ILVFYLPSDC
r $\delta$	SVPMDSTNHQ	DVTFYLIIRR	KPLFYIINIL	VPCVLISFMI	NLVFYLPGDC
r $\gamma$	VTPAEEAGHQ	KVVFYLLIQR	KPLFYVINII	VPCVLISSVA	ILIIYFLPAKA
c $\alpha$ 8	G-EKISLGIT	VLLSLTVFML	LVAEIMPATS	DSVPLIAQYF	ASIMVIVGLS
d $\alpha$ 2	G-EKIALCIS	ILSQTMMFL	LISEIIPSTS	LALPLLGKYL	LFTMMLVGLS
d $\alpha$ Li	G-EKISLCIS	ILSLSLTVF	LLAEIIPSTS	LTVPLLGKYL	LFTMMLVTL
d $\beta$ 2	G-EKVTLCIS	ILVSLTVF	LLAEIIPSTS	LAVPLLGKYL	LFTMILVSL
dnachr	G-EKVTLCIS	ILSLSLTVF	LLAEIIPSTS	LAVPLLGKYL	LFTMILVSL
mser	G-ERVSFKIT	LLLGYSVFL	IVSDTLPAT-	IGTPLIGVYF	VVCMALV
onachr	G-EKVTLCIL	ILVALTVF	LLKDIIIPATS	IALPLFGKYL	LFTMIMVSL
r $\alpha$ 2	G-EKITLCIS	VLLSLTVF	LITEIIPSTS	LVIPLIGEYL	LFTMIFVTL
r $\alpha$ 3	G-EKVTLCIS	VLLSLTVF	VITETIPSTS	LVIPLIGEYL	LFTMIFVTL
r $\alpha$ 7	G-EKISLGIT	VLLSLTVF	LVAEIMPATS	DSVPLIAQYF	PSTMIIIVGLS
r $\beta$ 2	G-EKMTLCIS	VLLALTVF	LISKIVPPTS	LDVPLVGKYL	MFTMVLVTF
r $\delta$	G-EKTSVAIS	VLLAQSVF	LISKRLPATS	MAIPLVGKFL	LFGMVLVTF
r $\gamma$	GGQKCTVATN	VLLAQTVF	LVAKKVPETS	QAVPLISKYL	TFLMVVTILI

Appendix 3: Alignment of 13 nAChR Subunits Made by the CLUSTAL V Software—Total Sites: 351; Informative Sites: 263

$\alpha_8$	VVVTVLVLQF	HHDPQAGKM	PRWVRVILLN	WCAWFLRMKK	PTIPVIVKIL	
$\alpha_2$	VVITIIILNI	HYRKPSHTKM	RPIWRSFFIK	RLPKLL-LMR	V---ELEKAI	
$\alpha_{Li}$	VVVTIAVLNV	NFRSPVTHRM	APWVQRLFIQ	ILPKLLECER	P---EMEKTI	
$\beta_2$	VWTTVCVLNI	HFRSPSTHNM	SRLVRKLFHL	FMPKLMRMRR	T---EVLOAL	
$\alpha_{nachr}$	ILVTVIIINW	NFRGPRTHRM	PMYIRSIFLH	YLPAFLFMKR	---PEASKAT	
$\alpha_{ser}$	LAETIFIVRL	VHKQDLQRPV	PDWLRLVLD	RIAWILCLPR	EASLAVRGLL	
$\alpha_{nachr}$	VLVTVISLNL	HFRSPSTHRM	PIWKWLFLR	ILPKILFMRR	---HVIKAF	
$\alpha_2$	IVITVFVLNV	HHRSPSTHNM	PNWVRVALLG	RVPRLMMNR	PLSPQIQKAL	
$\alpha_3$	IVITVFVLNV	HYRTPTTHTM	PTWVKAVFLN	LLPRVMFMTR	PLSPEIKEAI	
$\alpha_7$	VVVTIVLRLY	HHHDPDGGKM	PKWTRIILLN	WCAWFLRMKR	PGDPDLAKIL	
$\beta_2$	IVTSVCVLNV	HHRSPPTTHTM	APWVKVVFLE	KLPTLLFLQQ	---PGLREAV	
$\delta$	VVICVIVLNI	HFRTPSTHVL	SEGVKKFFLE	TLPKLLLHMSR	PLFNEMKPAV	
$\gamma$	VVNSVVVLNV	SLRSPHTHSM	ARGVRKVFLR	LLPQLLRMHV	HASPAIQACV	
$\alpha_8$	EEVQFIAMRF	RKQDEGEEIC	SEWKFAAAVI	DRLCLVAFTL	FAIIC TFTIL	M
$\alpha_2$	HNVMFIQHHM	QRQDEFNAED	QDWGFVAMVM	DRLFLWLWMI	ASLVGT-FVI	L
$\alpha_{Li}$	EGSRFIAQHV	KNKDKFESVE	EDWKYVAMVL	DRMFLWIFAI	ACVVGTLAII	L
$\beta_2$	RAVRFIAQHI	KDADKDNEIV	EDWKFVSMVL	DRFFLWLFTL	SCVFGTLAII	C
$\alpha_{nachr}$	EAVEFIAEHL	RNEDLYIQT	EDWKYVAMVI	DRLQLYIFFI	VTTAGTVGIL	M
$\alpha_{ser}$	QELSSIRHFL	EKRDEMREVA	RDWLRVGYVL	DRLLFRIYLL	AVLAYSITLV	T
$\alpha_{nachr}$	ENVCFIAQLL	KKKDREAMID	EDWKFVARVL	DRLFLLLFSI	ACFLGTILIL	F
$\alpha_2$	EGVHYIADRL	RSEDADSSVK	EDWKYVAMVV	DRIFLWLFI	VCFLGTIGLF	L
$\alpha_3$	QSVKYIAENM	KAQNVAKEIQ	DDWKYVAMVI	DRIFLWVFIL	VCILGTAGLF	L
$\alpha_7$	EEVRYMPTAY	RCQDESEVIC	SEWKFAACVV	DRLCLMAFSV	FTIIC TIGIL	M
$\beta_2$	DGVRFIADHM	RSEDDDQSVR	EDWKYVAMVI	DRLFLWIFVF	VCVFGTVGMF	L
$\delta$	DGANFIVNHM	RDQNSYNEEK	DNWNQVARTV	DRCLCLFVVTP	VMVVGTAWIF	L
$\gamma$	DACNLMLARAR	HQQSHFDSDGN	EEWLLVGRVL	DRVCFMLMS	LFICGTAGIF	L

**Appendix 3:** Continued