

## Influence of L-tryptophan and auxins applied to the rhizosphere on the vegetative growth of *Zea mays* L.

MUHAMMAD SARWAR and W.T. FRANKENBERGER, Jr.<sup>1</sup>

Department of Soil and Environmental Sciences, University of California, Riverside, CA 92521, USA.

<sup>1</sup>Corresponding author

Received 28 June 1993. Accepted in revised form 15 November 1993

**Key words:** biologically active substances, indole-3-acetic acid, microbial synthesis of phytohormones

### Abstract

Glasshouse experiments were conducted to evaluate the influence of L-TRP in comparison with indole-3-acetamide (IAM), tryptophol (TOL) and indole-3-acetic acid (IAA) on the growth of *Zea mays* L. var. Early Sunglow. L-TRP ( $25$  to  $2.5 \times 10^{-5}$  mg kg<sup>-1</sup> soil), IAM ( $22$  to  $2.2 \times 10^{-5}$  mg kg<sup>-1</sup> soil), TOL ( $20$  to  $2.0 \times 10^{-5}$  mg kg<sup>-1</sup> soil), and IAA ( $22$  to  $2.2 \times 10^{-5}$  mg kg<sup>-1</sup> soil) were applied as a soil drench to established uniform seedlings. All treatments were applied in a completely randomized design with 10 replicates. IAM had no significant effect on the plant growth parameters. Shoot height, uppermost leaf collar base distance, internodal distance, and shoot dry and fresh weights were significantly improved upon the addition of TOL ( $2.0 \times 10^{-2}$  mg kg<sup>-1</sup> soil), however, the highest concentration ( $20$  mg kg<sup>-1</sup> soil) caused a 14.6% reduction in leaf width. L-TRP ( $2.5 \times 10^{-3}$  mg kg<sup>-1</sup> soil) also had a significant influence on shoot height, uppermost leaf collar base distance, internodal distance and fresh weight of shoot compared with the control. The highest concentration of L-TRP ( $25 =$  mg kg<sup>-1</sup> soil) had a negative effect on leaf width and dry weight of the shoot. The most pronounced response on the corn growth parameters was observed with the application of IAA at lower concentrations ( $2.2 \times 10^{-5}$  to  $2.2 \times 10^{-2}$  mg kg<sup>-1</sup> soil) specifically improving root growth. The highest concentration ( $22$  mg kg<sup>-1</sup> soil) of IAA had a significant negative effect on plant height, leaf width, stem diameter, shoot fresh and dry weight. These findings indicate that L-TRP applied at the appropriate concentrations can have positive effects on corn growth comparable to pure auxins (TOL and IAA).

### Introduction

Plant hormones play a crucial role in controlling plant growth and development. Among the plant hormones known today, auxin (from the Greek term *auxein*, to increase) was the first phytohormone recognized by Darwin (1880). His work on the phototropic response of grass coleoptiles indicated the transfer of a signal from the tip to lower parts of the plant. Later, Went (1926; 1928) was the first to isolate the active substance from an *Avena* coleoptile tip onto agar and

reported that the tropistic response was due to a specific diffusible substance. The isolation of auxin (IAA) from a plant was achieved by Haagen-Smit et al. (1942) and was later confirmed by Berger and Avery (1944). Since then, IAA has been isolated from several other plant sources.

Biosynthesis of auxins is not limited to higher plants. Several microorganisms including fungi (Frankenberger and Poth, 1987; Pokojska and Strzelczyk, 1988), bacteria (Hubbell et al., 1979; Muller et al., 1989), and algae (Florenzano et

al., 1978; Moss, 1965) also have been reported to synthesize auxins. Comparatively much data have been collected on auxin metabolism in higher and lower plants, but little emphasis has been placed on the role of microbially-derived auxins on plant growth.

Soils also have been reported to contain compounds exhibiting strong auxin-like activity (Hammen, 1946; Sheldrake, 1971; Stewart and Anderson, 1942; Whitehead, 1963) and differ in their auxin synthesizing capacity depending on the fertility status and organic matter content (Chandramohan and Mahadevan, 1968; Hammen, 1946). The literature contains ample evidence that the soil microbiota are active in auxin synthesis (Arshad and Frankenberger, 1992; Bric et al., 1991).

Generally, the rhizosphere and rhizoplane microbial community is more active in auxin production than those from non-rhizosphere soil (Brown, 1972; Dvornikov et al., 1970; Kampert et al., 1975; Purushothaman et al., 1974; Strzelczyk and Pokojska-Burdzej, 1984; Strzelczyk et al., 1977). Higher auxin production in the rhizosphere is most likely due to the abundance of substrates and diverse microbial populations (Narayanaswami and Veeraj, 1969). Similarly, Rossi et al. (1984) found a higher amount of auxin-like compounds in corn rhizosphere compared with non-rhizosphere environments, especially during seedling emergence. Arshad and Frankenberger (1991) confirmed these findings by reporting a higher percentage of microorganisms from rhizosphere soil than non-rhizosphere soil capable of producing auxins. However, these microbiota could only affect plant growth if microbially-derived auxins are taken up by plant roots.

L-Tryptophan (L-TRP) is considered as a physiological precursor of auxins in higher plants and microbial production. Purushothaman et al. (1973; 1974) reported synthesis of auxins in TRP-amended soil. Frankenberger and Brunner (1983) confirmed this observation with HPLC-MS and proved that IAA was produced in soil when incubated with L-TRP. Recently, Sarwar et al. (1992) studied auxin biosynthesis in L-TRP-amended soil under the influence of several environmental factors and reported that TRP is a physiological precursor of auxins in soil. Mi-

crobiologically-derived auxins from applied TRP within the rhizosphere can have some major effects on plant growth. Frankenberger and Poth (1987) reported dramatically increased growth of Douglas fir by inoculating with the ectomycorrhizae, *Pisolithus tinctorius*, when soil was amended with L-TRP. Frankenberger et al. (1990) also reported a significantly positive effect of L-TRP comparable with the pure auxins (IAA, indole-3-acetamide, indole-3-lactic acid, etc.) on growth of radish when applied at low concentrations (ng to  $\mu\text{g kg}^{-1}$  soil) at the seedling stage.

Although L-TRP and its various analogs have been tested for their effects on growth of various plants, a thorough investigation dealing with the influence of various levels of TRP and pure auxins (indole-3-acetamide, tryptophol and IAA) when applied to soil on the growth of maize, have not been compared. Thus this study was initiated to compare the effects of microbially-derived auxins from L-TRP and pure auxins with the hypothesis that microbially synthesized auxins in soil amended with L-TRP are subject to plant uptake and evoke a physiological response.

## Materials and methods

A glasshouse study was conducted to evaluate the influence of indole-3-acetamide (IAM), tryptophol (TOL), L-tryptophan (L-TRP), and indole-3-acetic acid (IAA) added to soil on the growth of *Zea mays* L. var Early Sunglow. All compounds were obtained from Sigma (St. Louis, MO). The physicochemical properties of the soil used for the experiment were determined as follows: pH, 8.11; inorganic nitrogen as  $\text{NH}_4\text{-N}$ , 20.3  $\text{mg kg}^{-1}$  and as  $\text{NO}_3\text{-N}$ , 46.8  $\text{mg kg}^{-1}$  soil; orthophosphate-P, 9.1  $\text{mg kg}^{-1}$  soil; organic carbon, 2.7%; moisture content, 14.8%; sandy clay loam texture (47.5% sand and 27.5% clay), a heterotrophic microbial population of  $136 \times 10^5$ ,  $74 \times 10^2$  and  $112 \times 10^3$   $\text{cfu g}^{-1}$  soil enriched on nutrient agar, a minimal salt medium (MSM) plus L-TRP as the sole C source and MSM plus L-TRP as the sole N source, respectively. The soil was selected on the basis of a preliminary study indicating a positive response of soil applied L-TRP on the growth of maize. Surface

(0-15 cm) soil was collected, homogenized and passed through a 2-mm sieve before filling the pots.

#### *Growth of seedlings*

Seeds of *Zea mays* L. cv. Early Sunglow (Burpee, Warminster, PA) were sown directly in plastic pots containing 4.0 kg of sieved field moist soil at the rate of 6 seeds per pot approximately equidistant from each other. Hoagland's mineral solution (500 mL, full-strength) was applied as a one-time application to each pot 10 d prior to treatment with L-TRP and auxins. The controls consisted of treatment with the nutrient solution, but not L-TRP or the auxins. The soil moisture content was maintained at 15% allowing it to dry out before the nutrient solution was applied. Corn seedlings were thinned out to one uniform plant per pot prior to treatment application (10 d after sowing seeds).

#### *Treatment application*

Various concentrations of IAM ( $22$  to  $2.2 \times 10^{-5}$  mg kg<sup>-1</sup>), TOL ( $20$  to  $2.0 \times 10^{-5}$  mg kg<sup>-1</sup>), L-TRP ( $25$  to  $2.5 \times 10^{-5}$  mg kg<sup>-1</sup>) and IAA ( $22$  to  $2.2 \times 10^{-5}$  mg kg<sup>-1</sup>) in solution form were applied to soil 2 d after thinning as a one-time application. All treatments were applied in four different sets, each in a completely randomized design with 10 replicates having one plant per pot.

#### *Growth measurements*

Plant growth was monitored over the entire vegetative growth period and harvested at the appearance of tassels (55 d after sowing). Shoot height was measured up to the top of the uppermost leaf every week throughout the experiment and to the base of the uppermost leaf collar. Stem diameter was measured at the fifth internode with a Vernier caliper and leaf width at the mid-point of the leaf originating from the fifth node. Internodal distance was measured between the fourth and fifth nodes from each plant in each set. After harvesting, shoot and root fresh weights were recorded. Then dry

shoot and root weights were measured following drying at 65°C for 48 h.

#### *Statistical analysis*

The data obtained were subjected to analysis of variance (ANOVA) and all treatment means were compared with the control according to Dunnett's test (Steel and Torrie, 1980).

#### **Results**

This glasshouse experiment was conducted to evaluate the influence of L-TRP in comparison with IAM, TOL and IAA on the growth of *Zea mays* L. Growth parameters measured included shoot height, uppermost leaf collar base distance, leaf width, internodal distance, stem diameter, shoot fresh and dry weight, and root fresh and dry weight.

#### *Shoot height*

There was no significant difference in shoot height upon the application of IAM at  $2.2 \times 10^{-5}$  to  $22$  mg kg<sup>-1</sup> soil (Table 1). TOL promoted shoot height at  $2.0 \times 10^{-2}$  mg kg<sup>-1</sup> soil with a 10.2% increase over the control (Table 2). L-TRP applied at  $2.5 \times 10^{-3}$  mg kg<sup>-1</sup> soil also promoted shoot height by 11.7% while other concentrations were ineffective (Table 3). A dramatic response was observed with the application of IAA, particularly at the highest concentration ( $22$  mg kg<sup>-1</sup> soil). At this concentration, plant height was reduced by 16.4% over the control 5 weeks after treatment. Shoot height was enhanced (7.5% increase) with a lower concentration of IAA ( $2.2 \times 10^{-3}$  mg kg<sup>-1</sup> soil) applied to the rhizosphere (Table 4).

#### *Uppermost leaf collar base distance*

While IAM applied to soil had no significant effect on the uppermost leaf collar base distance, TOL at  $2.0 \times 10^{-2}$  mg kg<sup>-1</sup> soil and L-TRP at  $2.5 \times 10^{-3}$  mg kg<sup>-1</sup> soil promoted this growth parameter by 10.5% and 12.6%, respectively. Overall the most effective treatment was IAA with an increase in the uppermost leaf collar base distance ranging from 15.9 to 22.8% over

Table 1. Growth parameters of Early Sunglow corn as influenced by indole-3-acetamide (IAM) applied to soil (average of 10 replicates)

IAM application (mg kg <sup>-1</sup> )	Shoot height (cm)					Uppermost leaf collar base distance (cm)	Leaf width (mm)	Internodal distance (mm)	Stem diameter (mm)	Shoot		Root	
	2	3	4	5	Weeks following application					fresh wt (g)	dry wt (g)	fresh wt (g)	dry wt (g)
Control	68.6	88.2	101.1	103.7	71.0	5.59	7.91	11.5	87.3	28.1	2.06	1.89	
22	64.6	89.7	104.4	110.5	77.7	5.33	9.00	11.2	89.0	22.5	1.97	1.66	
2.2	68.6	87.4	101.7	105.5	73.2	5.55	8.28	11.1	88.1	26.4	2.21	2.02	
2.2 × 10 <sup>-1</sup>	70.0	87.7	102.0	103.7	69.4	5.54	7.94	11.0	85.7	26.9	2.20	2.02	
2.2 × 10 <sup>-2</sup>	72.4	92.1	105.2	107.2	73.7	5.59	8.29	11.8	92.4	28.4	2.30	1.99	
2.2 × 10 <sup>-3</sup>	72.8	92.9	104.4	106.3	74.6	5.67	8.16	10.6	92.4	29.1	2.46	2.27	
2.2 × 10 <sup>-4</sup>	70.3	88.8	101.2	104.2	66.5	5.54	7.96	11.7	88.2	28.1	2.39	1.86	
2.2 × 10 <sup>-5</sup>	69.8	86.0	101.2	103.5	65.1	5.34	7.88	11.2	86.2	27.8	2.46	2.15	

\*\*\* = Treatment means significantly different from control at  $p \leq .05$  and 0.01, respectively, according to Dunnett's test.

Table 2. Growth parameters of Early Sunglow corn as influenced by tryptophol (TOL) applied to soil (average of 10 replicates)

TOL application (mg kg <sup>-1</sup> )	Plant height (cm)					Uppermost leaf collar base distance (cm)	Leaf width (mm)	Internodal distance (mm)	Stem diameter (mm)	Shoot		Root	
	2	3	4	5	Weeks following application					fresh wt (g)	dry wt (g)	fresh wt (g)	dry wt (g)
Control	63.6	75.3	82.7	84.1	54.1	4.92	5.34	9.6	44.1	21.0	1.43	0.98	
20	57.2	72.3	81.5	83.7	58.3	4.20**	6.27	8.9	40.9	19.8	1.28	0.82	
2.0	61.9	75.9	83.9	85.1	54.9	4.74	5.58	9.5	42.7	20.5	1.12	0.83	
2.0 × 10 <sup>-1</sup>	62.9	76.4	85.4	87.2	55.4	4.83	5.96	9.6	45.0	20.5	1.21	0.88	
2.0 × 10 <sup>-2</sup>	64.3	78.6	88.5*	92.7**	59.8*	4.98	6.80**	10.0	52.2**	24.1**	1.64	1.12	
2.0 × 10 <sup>-3</sup>	72.8	76.0	84.3	88.2	59.5	4.65	6.25	9.3	43.9	20.2	1.26	0.80	
2.0 × 10 <sup>-4</sup>	70.3	76.1	84.6	86.9	59.5	4.52*	6.14	9.8	45.4	20.9	1.35	0.77	
2.0 × 10 <sup>-5</sup>	69.8	72.6	82.0	82.0	56.0	4.67	5.21	9.2	42.0	20.2	1.30	0.94	

\*\*\* = Treatment means significantly different from control at  $p \leq .05$  and 0.01, respectively, according to Dunnett's test.

Table 3. Growth parameters of Early Sunglow corn as influenced by L-tryptophan (L-TRP) applied to soil (average of 10 replicates)

TRP application (mg kg <sup>-1</sup> )	Plant height (cm)					Uppermost leaf collar base distance (cm)	Leaf width (cm)	Internodal distance (cm)	Stem diameter (mm)	Shoot		Root	
	Weeks following application									Shoot fresh wt (g)	Shoot dry wt (g)	Root fresh wt (g)	Root dry wt (g)
	2	3	4	5	5								
Control	63.0	77.6	85.6	88.9	60.4	5.18	5.25	10.3	57.3	15.6	2.47	1.89	
25	61.0	77.4	80.1	81.6	60.4	4.38**	5.72	9.3	46.5	12.5*	1.92	1.39	
2.5	63.6	76.8	83.4	84.0	63.6	4.88	6.35	9.5	55.1	14.8	2.58	2.00	
2.5 × 10 <sup>-1</sup>	65.5	77.9	85.3	88.1	64.6	5.20	6.40	10.4	56.1	17.1	2.49	1.74	
2.5 × 10 <sup>-2</sup>	64.3	77.7	83.7	92.2	64.7	5.42	6.95*	10.4	62.8	17.0	2.98	1.91	
2.5 × 10 <sup>-3</sup>	64.4	78.3	87.8	99.3*	68.0*	5.68	7.31**	11.0	70.2*	18.1	3.20	2.30	
2.5 × 10 <sup>-4</sup>	60.4	77.8	85.4	96.0	61.6	5.60	6.84	10.2	58.1	15.4	2.94	2.09	
2.5 × 10 <sup>-5</sup>	62.2	76.8	84.8	90.8	60.9	5.13	6.43	10.3	57.4	15.5	2.85	1.98	

\*\*\* = Treatment means significantly different from control at p ≤ .05 and 0.01, respectively, according to Dunnett's test.

Table 4. Growth parameters of Early Sunglow corn as influenced by indole-3-acetic acid (IAA) applied to soil (average of 10 replicates)

IAA application (mg kg <sup>-1</sup> )	Plant height (cm)					Uppermost leaf collar base distance (cm)	Leaf width (cm)	Internodal distance (cm)	Stem diameter (mm)	Shoot		Root	
	Weeks following application									Shoot fresh wt (g)	Shoot dry wt (g)	Root fresh wt (g)	Root dry wt (g)
	2	3	4	5	5								
Control	58.6	67.8	72.1	73.6	43.9	4.40	4.70	8.9	26.2	7.1	1.59	1.17	
22	39.3**	44.9**	51.1**	61.5**	48.8	2.74**	5.80**	6.2**	13.5**	3.8**	1.12	0.80	
2.2	53.4*	64.2	69.3	71.2	48.5	4.22	5.15	8.3	22.9	6.9	1.64	1.22	
2.2 × 10 <sup>-1</sup>	56.9	67.1	72.8	74.1	48.6	4.56	4.96	8.9	27.6	7.0	2.05	1.68	
2.2 × 10 <sup>-2</sup>	58.3	67.5	71.5	74.6	50.9*	4.67	5.11	9.2	28.8	7.3	2.23*	1.95**	
2.2 × 10 <sup>-3</sup>	60.7	71.7	77.8*	79.1*	51.4*	4.77	5.48	9.1	32.3*	7.4	2.81**	2.49**	
2.2 × 10 <sup>-4</sup>	60.0	69.8	75.7	77.8	53.9**	4.54	5.22	9.1	30.1	7.0	2.56**	2.07**	
2.2 × 10 <sup>-5</sup>	61.0	70.7	74.8	76.9	50.2	4.65	5.18	9.1	29.8	6.8	2.44**	2.00**	

\*\*\* = Treatment means significantly different from control at p ≤ .05 and 0.01, respectively, according to Dunnett's test.

the control with a concentration of  $2.2 \times 10^{-2}$  to  $2.2 \times 10^{-4}$  mg kg<sup>-1</sup> soil (Table 4).

#### *Leaf width*

There was no significant difference in leaf width upon the application of IAM compared with the control. However, upon the addition of TOL (20 mg kg<sup>-1</sup> soil), the leaf width decreased by 14.6%, 15.4% with L-TRP (25 mg kg<sup>-1</sup> soil), and 37.7% with IAA (22 mg kg<sup>-1</sup> soil). All other concentrations of L-TRP and auxins applied had no significant effect on leaf width.

#### *Internodal distance*

The application of IAM had no significant effect on the internodal distance of maize when applied at  $2.2 \times 10^{-5}$  to 22 mg kg<sup>-1</sup> soil. However, the internodal distance was promoted by 27.3% upon the application of TOL at  $2.0 \times 10^{-2}$  mg kg<sup>-1</sup> soil (Table 2). L-TRP also increased the internodal distance from 32.3 to 39.2% when treated with a concentration range of  $2.5 \times 10^{-2}$  to  $2.5 \times 10^{-3}$  mg kg<sup>-1</sup> soil (Table 3). The internodal distance of maize was enhanced upon the application of IAA by 23.4% when applied at 22 mg kg<sup>-1</sup> soil (Table 4).

#### *Stem diameter*

The addition of IAM, TOL and L-TRP had no significant effect on the stem diameter of maize. However, the application of IAA at 22 mg kg<sup>-1</sup> soil decreased the stem diameter by 30.3% (Table 4). All other concentrations of IAA tested had no significant effect on stem diameter.

#### *Shoot fresh weight*

The addition of IAM had no significant effect on the shoot fresh weight of maize, while TOL at  $2.0 \times 10^{-2}$  significantly enhanced this growth parameter by 18.4% (Table 2). L-TRP at  $2.5 \times 10^{-3}$  mg kg<sup>-1</sup> soil promoted the shoot fresh weight by 22.5% (Table 3). IAA showed both inhibitory and stimulatory effects, depending on the concentration applied. At the higher concentration of 22 mg IAA kg<sup>-1</sup> soil, shoot fresh weight decreased by 48.5%, while at  $2.2 \times 10^{-3}$

mg kg<sup>-1</sup> this growth parameter was enhanced by 23.3% (Table 4).

#### *Shoot dry weight*

The application of IAM had no significant effect on the shoot dry weight of maize. However, TOL had a positive influence at  $2.0 \times 10^{-2}$  mg kg<sup>-1</sup> soil with a 14.8% enhancement (Table 2). The addition of L-TRP and IAA decreased the shoot dry weight at the highest concentrations applied (25 and 22 mg kg<sup>-1</sup>, respectively) by 19.9% (L-TRP) and 46.5% (IAA) (Tables 3 and 4).

#### *Root fresh weight*

The application of IAM, TOL and L-TRP had no significant effect on the root fresh weight, however IAA stimulated the root weight at concentrations ranging from  $2.2 \times 10^{-5}$  to  $2.2 \times 10^{-2}$  mg kg<sup>-1</sup> soil with a 40.3 to 76.7% increase (Table 4).

#### *Root dry weight*

The addition of IAM, TOL and L-TRP had no significant effect on the root dry weight of maize, however IAA stimulated this growth parameter by 66.6 to 112.8% with concentrations ranging from  $2.2 \times 10^{-5}$  to  $2.2 \times 10^{-2}$  mg kg<sup>-1</sup> soil (Table 4).

### **Discussion**

Previous studies have shown that auxins produced by microorganisms in the vicinity of the root tip can have a significant effect on plant growth (Pilet, 1977). Attempts were made to prevent nutrient and water stress, hence the response to the treatments of L-TRP and auxins (TOL and IAA) were most likely a hormonal response.

The increase in plant height in response to a dilute concentration of L-TRP, TOL and IAA represents a typical growth response produced by auxins which are inhibitory at high concentrations and stimulatory at lower levels. Similar results have been demonstrated by Robertson et

al. (1976) who reported that high concentrations of IAA strongly inhibited seed germination of lettuce. Likewise, reduced seedling growth of oats by high auxin concentrations was observed by Hwang and Perse (1940). Frankenberger et al. (1990) reported a positive effect of L-TRP on the growth parameter of radish (*Raphanus sativus*) when applied at low concentrations at the seedling stage and was comparable with the effect of auxins (IAM and indole-3-lactic acid).

Many parameters of plant growth such as uppermost leaf collar base distance, internodal length, and shoot fresh weight were all significantly improved by lower concentrations of L-TRP and the auxins (TOL and IAA) applied. Our study also showed a strong root growth stimulation by IAA at the lower levels. IAA was found to be strongly inhibitory to many of the growth parameters such as shoot height, leaf width, internodal distance, stem diameter and shoot and root fresh weights, when applied at the highest concentration (22 mg kg<sup>-1</sup> soil). Similar findings were reported by Opatrna et al. (1988) who demonstrated root growth inhibition of *Chenopodium rubrum* by IAA at a rate of 1 mM, while lower concentrations stimulated the root growth of pea seedlings (Zelena et al., 1988). In this study, L-TRP also inhibited leaf width and shoot dry weight at the highest concentration (25 mg kg<sup>-1</sup> soil). The inhibitory effects of L-TRP may be a result of a slow and continuous release of IAA in soil due to microbial action.

The growth parameters of maize were not influenced by IAM. Even at the highest application (22 mg kg<sup>-1</sup>) there was no inhibitory effect. Since IAM is not found in plant tissues (Kosuge et al., 1983), the enzymes necessary to metabolize this auxin may be lacking, thus there was no hormonal response (Leopold, 1980).

This work indicates that L-TRP and auxins (TOL and IAA) can have an ecological effect in modifying plant growth and development. The mechanism of action of L-TRP and auxins on plant growth may be attributed to the direct uptake of these compounds by plant roots, a change in the balance of rhizosphere microflora discouraging root pathogens or by microbial conversion into metabolites resulting in a beneficial rhizosphere for plant growth.

## References

- Arshad M and Frankenberger W T Jr 1991 Microbial production of plant hormones. *Plant and Soil* 133, 1–8.
- Arshad M and Frankenberger W T Jr 1992 Microbial production of plant growth regulators. *In* *Soil Microbial Ecology*. Ed. B Metting, pp 307–347. Marcel Dekker, Inc., New York.
- Berger J and Avery G S Jr 1944 Isolation of an auxin precursor and an auxin (indole acetic acid) from maize. *Am. J. Bot.* 31, 199–203.
- Bric J M, Silverstone S E and Bostock R M 1991 Rapid in situ assay for indoleacetic acid production by bacterial immobilized on a nitrocellulose membrane. *Appl. Environ. Microbiol.* 57, 535–538.
- Brown M E 1972 Plant growth substances produced by microorganisms of soil and rhizosphere. *J. Appl. Bacteriol.* 35, 443–451.
- Chandramohan D and Mahadevan A 1968 Indoleacetic acid metabolism in soils. *Curr. Sci.* 37, 112–113.
- Darwin C 1880 *The Power of Movements in Plants*. London.
- Dvornikov T P, Skryabin G K, and Suvorov N N 1970 Enzymatic transformation of tryptamine by fungi. *Microbiol.* 39, 32–35.
- Florenzano G, Balloni W and Materassi R 1978 Algal organic matter and plant growth. *Zentral Blatt.* 133, 379–384.
- Frankenberger W T Jr and Brunner W 1983 Method of detection of auxin, indole-3-acetic acid in soils by high performance liquid chromatography. *Soil Sici. Soc. Am. J.* 47, 237–241.
- Frankenberger W T Jr and Poth M 1987 Biosynthesis of indole-3-acetic acid by the pine ectomycorrhizal fungus, *Pisolithus tinctorius*. *Appl. Environ. Microbiol.* 53, 2908–2913.
- Frankenberger W T Jr, Chang A C and Arshad M 1990 Response of *Raphanus sativus* to the auxin precursor, L-tryptophan applied to soil. *Plant and Soil* 129, 235–241.
- Haagen-Smit A J, Leach W D and Bergren W R 1942 The estimation, isolation and identification of auxins in plant materials. *Amer. J. Bot.* 29, 500–506.
- Hamence J H 1946 The determination of auxins in soils including a note on synthetic substances. *Analyst* 71, 111–116.
- Hubbell D H, Tien T M, Gaskins M H, and Lee J 1979 Physiological interactions in the *Azospirillum*-grass root association. *In* *Associative N<sub>2</sub>-Fixation*. Eds. P B Vose and A P Ruschel. CRC Press, Inc., Boca Raton, FL.
- Hwang Y and Pearse H L 1940 The response of seeds and seedlings to treatment with indolyacetic acid. *Ann. Bot.* 4, 31–37.
- Kampert M, Strzelczyk E and Pokojska A 1975 Production of auxins by bacteria isolated from the roots of pine seedlings (*Pinus silvestris* L.). *Acta Microbiol. Pol.* 7, 135–143.
- Kosuge T, Comai L and Glass N L 1983 Virulence determinants in plant pathogen interactions. *In* *Plant Molecular Biology*. Ed. R B Goldbert. pp 167–177. Alan R. Liss, Inc., New York.
- Leopold A C 1980 Hormonal regulating systems in plants. *In*

- Recent Developments in Plant Sciences. Ed. S P Sen. pp 33–41. Today and Tomorrow Publ., New Delhi, India.
- Moss B 1965 Apical dominance in *Fucus vesiculosus*. New Phytol. 64, 387–392.
- Muller M, Deigele L and Ziegler H 1989 Hormonal interaction in the rhizosphere of maize (*Zea mays* L.) and their effect on plant development. Z. Pflanzenernähr. Bodenk. 152, 247–254.
- Narayanaswami R and Veerajau F 1969 IAA synthesis in paddy soil as influenced by ammonium sulfate fertilization. Curr. Sci. 38, 517–518.
- Opatrna J, Josefusova Z and Pavlova L 1988 The role of IAA and ABA in the root growth regulation. In Proc. of Symposium, Sept. 28–Oct. 2, 1987. Liblice, Czechoslovakia. Eds. M. Kutacek, R S Bandurski and J. Krekule. pp 339–340. Academia, Prague.
- Pilet P E (Ed.) 1977 Plant growth regulation. In Proc. of 9th Internatl. Conf. on Plant Growth Substances. May 30–Sept. 4, 1976. Lausanne, Switzerland. Springer-Verlag Pub., New York.
- Pokojska A and Strzelczyk E 1988 Effect of organic acids on production of auxin-like substances by ectomycorrhizal fungi. Symbiosis 6, 211–224.
- Purushothaman D K, Balaraman K and Oblisami G 1973 Indoleacetic acid metabolism in soil as influenced by pesticide application. Curr. Sci. 42, 356–366.
- Purushothaman D, Marimuthu T, Venkataramanan C V and Kesavan R 1974 Role of actinomycetes in the biosynthesis of indoleacetic acid in soil. Curr. Sci. 43, 413–414.
- Rigaud J 1970 Biosynthesis of indole-3-acetic acid in connection with the metabolism of tryptophol and indole-3-acetaldehyde in rhizobium. Physiol. Plant. 23, 171–178.
- Robertson J, Hillman J R and Berrie A M M 1976 The involvement of indole acetic acid in the thermodormancy of lettuce fruits, *Lactuca sativa* cv. Grand Rapids. Planta 131, 309–313.
- Rossi W, Grappelli A and Pietrosanti W 1984 Phytohormones in soil after atrazine application. Folia Microbiol. 29, 325–329.
- Sarwar M, Arshad M, Martens D A and Frankenberger W T Jr 1992 Tryptophan-dependent biosynthesis of auxins in soil. Plant and Soil 147, 207–215.
- Sheldrake A R 1971 The occurrence and significance of auxins in the substrate of bryophytes. New Phytol. 70, 519–526.
- Steel R G D and Torrie J H 1980 Principles and Procedures of Statistics: A Biometrical Approach. 2nd Ed. pp 49–31. McGraw-Hill, New York.
- Stewart W S and Anderson M S 1942 Auxins in some American soils. Bot. Gaz. 103, 570–575.
- Strzelczyk E and Pokojska-Burdzej A 1984 Production of auxins and gibberellin-like substances by mycorrhizal fungi, bacteria and actinomycetes isolated from soil and the mycorrhizosphere of pine (*Pinus silvestris* L.). Plant and Soil 81, 185–194.
- Strzelczyk E, Sitek J M and Kowalski S 1977 Synthesis of auxins from tryptophan and tryptophan-precursors by fungi isolated from mycorrhizae of pine (*Pinus silvestris* L.). Acta Microbiol. Pol. 26, 255–264.
- Went F W 1926 On growth-accelerating substances in the coleoptile of *Avena sativa*. Proc. Kon. Akad. Wetensch. Amsterdam 30, 10–19.
- Went F W 1928 Wuchsstoff und Wachstum. Rec. Trav. Bot. Neerl. 25, 1–116.
- Whitehead D C 1963 Some aspects of the influence of organic matter on soil fertility. Soil Fertil. 26, 217–223.
- Zelena E, Kutacek M and Cermak V 1988 Fate of root applied indolylacetic acid and its influence on the growth of intact plants. In Physiology and Biochemistry of Auxins in Plants. Eds. M Kutacek, R S Bandurski and J. Krekule. pp 371–376. Proceedings of the Symposium, Sept. 28–Oct. 2, 1987. Liblice, Czechoslovakia. SPB Academic Publ.

Section editor: A C Borstlap