

Historical corner

Dynamics of the history of photosynthesis research

Hiroshi Huzisige¹, * & Bacon Ke², **

¹Okayama City, Japan; ²Walnut Creek, California, U.S.A.

“Qu'on ne dise pas que je n'ai rien fait de nouveau ; la façon de présenter l'information est nouvelle.” Blaise Pascal

Key words: photosynthesis, chlorophyll, reaction center, energy transfer, electron transfer, oxygen evolution, photophosphorylation, carbon dioxide assimilation

Abstract

A personal view of the history of progress in photosynthesis research beginning in the seventeenth century and ending in 1992 is presented in a chart form. The 350-year time span is divided arbitrarily into seven periods by the “development junctures,” which are likened to bamboo joints. The tempo of progress is reflected in the duration of the periods, starting from over 200 years for Period I, which progressively shortens in subsequent periods. This brief introduction highlights some of the events to show the dynamic nature of the progress in photosynthesis research.

1. The development junctures in the history of photosynthesis research

The history of photosynthesis research, as in other fields, has made many periodic, spectacular leaps – what we call “development junctures.” Over the years, the progress of research seems to be gradual, but actually it is not as gradual as it seems. In the course of research, at some point or another, one often faces a “wall,” beyond which nothing can be seen. At that point, transcending all the accepted ideas, some extraordinarily talented scholars begin to develop new experimental materials, procedures, and analytical methods, by which they break through

the wall which had blocked further progress of research at the time. This leads to a “break through” by the efforts of the leading scholars who follow, and great progress is then made possible. As time goes by, another wall appears, and another group of talented scholars lead to another development. Thus, current understanding of photosynthesis has passed through many development junctures (which may be likened to the joints in bamboo).

To some of us, it may seem rather difficult or even impossible to divide the progress of photosynthesis research into several well-defined stages, considering its rapid developments in recent times encompassing many diverse areas. However, in order to fully understand the present stage of photosynthesis research, we need to comprehend what really led to certain discoveries; how much effort was behind a certain idea at the time it was presented; how subsequent research on that topic followed. Furthermore, it is very important to judge the impact of certain erroneous ideas; how they confused subsequent

* Professor emeritus of Okayama University, Okayama, Japan; correspondence address: Yokoi-kami, 507-66, Okayama City, Okayama, 701-11, Japan

** Formerly with the Charles F. Kettering Research Laboratory, Yellow Springs, Ohio, U.S.A.; present address: 5920-A₁ Horsemans Canyon Drive, Walnut Creek, CA 94595, U.S.A.; E-mail: BaconKe@CRL.COM

research for a while; and delayed the progress of the field.

With the above in mind, we present here several outstanding development junctures during the history of photosynthesis research and attempt to view the history of photosynthesis by dividing it, perhaps somewhat arbitrarily, into seven broad periods as the chart that follows this brief introduction shows.

2. Progress of photosynthesis research

2.1. Period I (From the beginning of the recognition of photosynthesis to ca. 1880)

This is the period when each element of photosynthesis, i.e. the nature of the components that are used as well as produced in the process, became established either by observations or by simple experiments. Thus the pioneering works during this period led to the generally accepted outline of photosynthesis. This period was culminated by the publication by the German scientist J. Sachs (1859-1862; I.18 [see ref. list]) of a treatise "Pflanzenphysiologie," that included discussions on the physiological aspects of photosynthesis. His work was followed up by his pupil W. Pfeffer (1874-1892; I.19), who also coined the term "photosynthetic assimilation."

2.2. Period II (ca. 1880 - ca. 1910)

The central figure in the research of this period was the English physiologist F. F. Blackman, who analyzed the relative rates of photosynthesis as affected by the various factors, and the interrelationships between them. He investigated in detail the relationship of the rate of photosynthesis and CO₂ concentration, light intensity, and temperature. As a result, he initiated the concept of a "limiting factor" (1905; II.2), which advocates that photosynthesis is not a simple photochemical reaction, but includes a reaction stage which does not require light. Later, this idea was tested by many researchers, the results were mixed, but in the end this theory was accepted.

Another trend during this period was the research on chlorophyll itself. Although it was recognized early (Dutrochet 1837; I.10) that chlorophyll was an important factor in photosynthesis, it was Willstätter and Stoll who attempted to explain the mechanism of photosynthesis through the various chemical characteristics of chlorophyll. Willstätter and Stoll published "*Untersuchungen über Chlorophyll*" (1913; II.3) and "*Untersuchungen über die Assimilation der Kohlensäure*" (1918; II.8), providing enormous amounts of information (both chemical and optical) on chlorophyll extracted from green leaves. However, in order to explain definitively and substantively the central role chlorophyll plays in photosynthesis, it is necessary to have knowledge on photosynthesis in other areas. This knowledge was lacking in those days. Thus, it was premature to study photosynthesis from the angle of chlorophyll chemistry alone.

Engelmann (1883; II.4) and Winogradsky (1887-1888; II.5) discovered photosynthetic bacteria, which have turned out to be important materials for current research in photosynthesis. During this period, we find an example of a misleading and erroneous hypothesis that was inspired by von Baeyer's research on formaldehyde (1864; II.9); many fruitless experiments were done in order to prove that formaldehyde was the initial carboxylation product formed in photosynthesis. This was a futile attempt and his hypothesis was later rejected by the experiment using carbon isotope ¹⁴C.

2.3. Period III (ca. 1910 - ca. 1938)

Otto Warburg (October 8, 1883 - August 1, 1970) was the center of research activities during this period. Until his research, knowledge concerning the photosynthetic process was fragmented and mostly qualitative. Warburg perfected the manometer to enable the measurement of gas exchange accurately (1919-1920; III.2), and introduced the unicellular green alga, *Chlorella*, as a new experimental material. This alga is not only easy to cultivate but also easy to handle in large quantities. Moreover, unlike higher-plant

leaves used in the past, there was no need to worry about the problem of opening and closing of the stomata or gas diffusion; it became an excellent and popular experimental material for photosynthesis research. Warburg also studied systematically and quantitatively the inhibition of photosynthesis by using various toxic chemicals, thus opening a way to analyze photosynthesis through its inhibition by known chemicals. However, because of his arrogance, he ignored some of the new information obtained by others. Sometimes, by presenting an erroneous theory, such as the unreasonably high quantum yield of photosynthesis, he disturbed the scientific world of his days, and even delayed the progress of the field. Nevertheless, in our opinion, he was a giant in photosynthesis research and made numerous contributions.

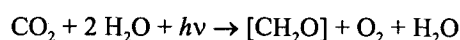
Warburg, using the new experimental material (*Chlorella*) and the new measuring method (manometry), pursued his research vigorously and confirmed the mechanism that photosynthesis consists of (1) a light reaction in which light participates directly, and (2) a dark reaction where light does not participate directly (1919-1920; III.2).

Influenced by his father, Emil Warburg, a scholar in optics, Otto Warburg was familiar with optical instruments, which he introduced effectively into the research of photosynthesis. He set up new measuring equipment and measured, for the first time, the maximum quantum yield of photosynthesis (1922-1923; III.3) and conducted flashing-light experiments (1919-1920; III.2). Later, many scholars worked in the areas pioneered by Warburg and established an important main stream of contemporary photosynthesis research. However, Warburg's extremely high quantum yield of photosynthesis were subsequently challenged by his own student Robert Emerson: the value is now settled as 0.12 in favor of Emerson (IV.5).

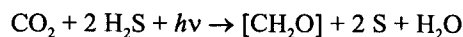
Another main stream of this period was the general formulation of photosynthesis advocated by van Niel (1929; III.9). He made good use of a suggestion by Thunberg (1923; III.10) that photosynthesis is a redox system involving CO₂ re-

duction and H₂O oxidation, and came up with the general equation for photosynthesis. In other words, the main function of photosynthesis is CO₂ reduction by hydrogen donors. The hydrogen donor may be different depending on the organism which performs the photosynthesis, but the process of CO₂ reduction is the same in all photosynthetic organisms:

In the case of plants and cyanobacteria (oxygenic):

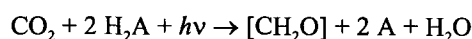


In the case of photosynthetic bacteria (anoxygenic):



or $\text{CO}_2 + 2 \text{H}_2 + h\nu \rightarrow [\text{CH}_2\text{O}] + \text{H}_2\text{O}$

General equation for photosynthesis is then:



Although the concept of redox reactions was also provided by Wurmser (1925-1930; III.8), van Niel's work is clearly responsible for research on photosynthetic bacteria that followed this period (however, see Gest [1993] under III.9). The idea of van Niel was singularly responsible for the modern thinking on the mechanism of photosynthesis. Soon after the establishment of this unitary concept of photosynthesis by van Niel, the very important new concept of the "chlorophyll-containing photosynthetic unit" was advanced by Emerson and Arnold in 1932 (III.4). This concept continued to gain strength (Arnold and Kohn [1934; IV.7]; Gaffron and Wohl [1937; IV.8]; and Wohl [1937-1941; IV.8]).

2.4. Period IV (ca. 1938 - ca. 1954)

During this period many new streams sprang up in the history of photosynthesis research. One of them was the discovery of what we call the "Hill reaction" by Robin Hill (1937-1939; IV.24). Oxygenic photosynthesis takes place in chloroplasts of plant cells. However, in the suspensions

of extracted (non-intact) chloroplasts obtained by grinding plant leaves, the ability to perform normal photosynthesis is lost although Jensen and Bassham later (1966; IV.33) succeeded in preparing active intact chloroplasts which can perform CO₂ fixation. Under the usual circumstances, the decomposition and complete reconstruction, which were very effective means for studying fermentation and respiration, could not be easily applied to photosynthesis research. However, Hill, by adding an oxidizing agent to the chloroplast preparation extracted from green leaves, found the photochemical oxygen-evolution reaction (1937-1939; IV.24) and thus a way into research in the biochemistry of chloroplast components and reactions. Later, many scholars pursued research in this direction by expanding the work on the Hill reaction.

Another major breakthrough was the introduction of the radio-isotope ¹⁴C and much progress was made in the analysis of carbon assimilation in photosynthesis. Sam Ruben and coworkers had pioneered research in this direction (1939-1940; IV.30). They added CO₂ labeled with carbon isotope ¹⁴C (half-life: 20.38 min.) to barley and found that plants have the ability to perform CO₂ fixation without light. After Ruben, Martin Kamen and others utilized the carbon isotope ¹⁴C which has a long half-life (*ca.* 5730 years), remarkable progress was made in the analysis of the path of carbon in photosynthesis. The two major research groups in this field were the Berkeley group (Melvin Calvin, Andy Benson, James Bassham, and others; see IV.31) and the Chicago group (Hans Gaffron, Al Fager, and others; see IV.32).

During this period, the research areas opened up by Warburg became increasingly active. Robert Emerson and his group determined the relationship between the quantum yield of photosynthesis and the wavelength of light used (1943; IV.6); they discovered the "red drop" in photosynthesis: far red light, by itself, was ineffective in photosynthesis. This discovery was followed by Emerson's discovery, in Period V, of the "Enhancement effect" that led to the concept of two-light reactions in photosynthesis.

2.5. Period V (*ca.* 1954 - *ca.* 1968)

From about 1950, photosynthesis research methods became increasingly precise, and collaboration by plant physiologists and physicists with chemists became a commonplace. Biophysics and biochemistry led to an analysis of the mechanism of photosynthesis at the molecular level. This was a golden period for photosynthesis research.

A major achievement concerned the area of biophysics: the use of rapid light-induced difference absorption spectroscopy, *i.e.* through the introduction of improved experimental instruments utilizing the concepts of flash photolysis pioneered by Porter and Norris (Witt and coworkers [1955; V.31]; Kok and coworkers [1959; V.32]) and those dealing with steady-state light (Duysens 1952; IV.10). With these techniques, it became possible to trace small absorption changes of reacting species and to explore primary changes in reaction-center chlorophylls or bacteriochlorophylls (the discovery of P870 by Duysens [1952; IV.10] and Clayton [1963; VI.19]; and that of P700 by Kok and coworkers [1956; V.33]). Essentially, all intermediates (*e.g.*, cytochromes, quinones, etc.) could be fingerprinted by this difference absorption method.

Another major achievement concerned with biophysics was the introduction of the electron-paramagnetic-resonance technique which enables the detection of unpaired electrons produced by light excitation (Commoner and coworkers [1956; V.34]; Calvin and coworkers [1957-1962; V.20]). This technique, like absorption spectrophotometry, has many advantages: it is a non-invasive technique; it is possible to analyze organelles *in vivo*, without destructive effects. It demonstrated an absolute advantage in exploring a majority of intermediates (*e.g.*, reaction-center chlorophylls, semiquinones, iron-sulfur centers, etc.).

On the other hand, classical methods (*e.g.*, manometry) led to the discovery of the Emerson enhancement effect by Emerson and his group (1957; V.7). Hill and Bendall proposed the momentous Z-scheme of photosynthesis. The con-

cept of the two photochemical systems was born, and the two-photosystem concept was immediately confirmed experimentally by several groups (Kok and Hoch [1959-1961; V.14]; Duysens *et al.* [1961; V.15]; and Witt *et al.* [1961; V.16]). Subsequently other explorations of the two photosystems, both by biochemical and biophysical methods, expanded photosynthesis research tremendously. Many important papers were published one after another: discovery of plastocyanin (Kato, 1960; V.48); photosynthetic NADP⁺ reduction (Arnon and coworkers (1951; IV.29); study of the rôle of cytochrome in photosynthesis (Duysens, [1955; V.49]; Chance and Nishimura [1960; V.50]); and the discovery of electron tunneling in bacterial cytochrome oxidation (DeVault and Chance, 1966; V.51).

Another pioneering contribution during this period was that by Al Frenkel (1954; V.44) and by Dan Arnon and coworkers (1954; V.45), who independently discovered photophosphorylation. These findings were pivotal in the later studies on the mechanism of energy conversion in photosynthesis. A few years later, Mitchell proposed the "chemiosmotic hypothesis" (1961; V.46) to explain ATP formation in mitochondria and chloroplasts.

The field of fluorescence of chlorophyll prospered during this period. Following the discovery of quenching of photosystem-II fluorescence by photosystem-I light (Govindjee *et al.*, 1960; V.42), Duysens and Sweers (1963; V.43) proposed that electron acceptor Q, located between the two photosystems, was a quencher in its oxidized state, but not in its reduced state. The non-Q related chlorophyll-*a* fluorescence change was discovered in several laboratories during this period. Furthermore, the temperature dependence of emission spectra down to 4° K, analyzed by Cho and Govindjee (1966-1970; V.30), revealed the validity of Förster's theory (1948; IV.15) on resonance excitation energy transfer in photosynthesis.

Much progress was also made in the analysis of the carbon cycle, and, around 1954 Calvin and Benson established a general principle of

the cycle (V.57). There followed the discovery of a new carbon pathway in C₄ plants by Kortschak and coworkers (1965; V.60) and by Hatch, Slack, and Johnson (1967; V.61), and research in this area made excellent progress.

Although research using electron microscopy began in the early 1940's, continued refinements (Steinmann, [1952; IV.3], Frey-Wyssling [1953; IV.4], and Menke [1962, 1965; V.6]) and development of new techniques such as negative staining (Brenner and Horne, 1959; V.4), and the freeze-etch technique (Moor, Mühlethaler, Waldner and Frey-Wyssling (1961; V.5) provided a detailed and beautiful picture of what we know today about the structure of thylakoids from higher plants, cyanobacteria and photosynthetic bacteria.

2.6. Period VI (*ca.* 1968 - *ca.* 1980)

After going over the various periods, we notice a rather interesting point. As the years pass, the intervals of the development junctures spanning the periods in this history chart that follows become increasingly shorter. This means the progress of learning has become faster and at the same time it suggests that the events marking the development junctures are inevitable. Viewed from the juncture around 1968, it is also clear that photosynthesis research has stepped into the molecular domain.

An example of the effect of the development of the techniques of flash-kinetic spectrophotometry and the EPR spectroscopy is the remarkable results produced in this period. This started with the discovery of P680 [1969; VI.28] and X320 (Q_A) [1968; VI.29] by Witt and coworkers, followed by the discovery of P430 (Hiyama and Ke [1971; VI.35]), the discovery of photosystem-I iron-sulfur centers (Malkin and Bearden [1971; VI.36]); and the discovery of electron acceptor A₂ (FeS-X) by McIntosh *et al.* (1975; VI.37). Thus the molecular species involved in the photochemical reactions in photosynthesis became clear.

As mentioned earlier, the discovery of the Emerson enhancement effect led to the idea of

the existence of two photochemical systems in photosynthesis. Combining this idea with the concept of the "photosynthetic unit," another major progress was seen in the substantive analysis of the relationship between the structure and function in photosynthetic systems. The "photosynthetic unit," a rather statistical and vague concept, could now be looked upon as real pigment-protein complexes. The research in this direction includes: the attempt to separate subchloroplast particles representing the two photosystems (Wessels [1962; V.10]; Boardman and Anderson [1964; V.11]; and Vernon and coworkers [1965; V.12]); the electrophoretic separation of the pigment-protein complexes by Ogawa, Obata and Shibata (1966; V.17) and by Thornber and coworkers (1966; V.18); separation and reconstitution of photosystem-I and -II particles (Huzisige and coworkers [1969; VI.8]; Briantais [1969; VI.9]; and later by Ke and Shaw [1972]; and by Lam and Malkin [1982] (see under VI.9).

The pioneering work on the isolation and characterization of the bacterial reaction center by Reed and Clayton [1969; VI.19] ushered in a new period of new and exciting developments.

In the field of analysis of photosynthetic oxygen evolution, which had been lagging in progress, new advances also began to appear. The important achievements were the discovery of a period-of-four change in oxygen evolution, in a series of light flashes, and the advocacy of the linear four-step mechanism for photosynthetic oxygen evolution (Joliot and coworkers [1968-1969; VI.40]; Kok and coworkers [1970; VI.41]). It became clear that four oxidizing equivalents accumulate on a charge accumulator before water is oxidized to O_2 and protons. Later, this accumulator was shown to be in a manganese complex.

The mechanism of energy transformation was analyzed in depth during this period and spectacular results were obtained: manifestation of membrane potential as electrochromic band shifts (Junge and Witt [1968; VI.50]; Jackson and Crofts [1969; V.51]); the role of ubiquinone in cyclic photophosphorylation in photosynthetic bacteria was observed (Horio and coworkers

[1968; VI.49]). Another important development was the discovery of the two-electron gate at the quinone site, first in photosystem II (Bouges-Bocquet [1973; VI.44]; Velthuys and Ames [1974 VI.45]) and then in photosynthetic bacteria (Vermeglio and Clayton [1977; VI.46] and Wraight [1977; VI.47]).

On another front, there was much progress in the direction of genetic analysis of photosynthetic systems: namely, the identification of the nuclear gene for LHCII (Kung, Thornber and Wildman [1972; VI.1]); isolation and sequencing, respectively, of the *psbA* gene for the Q_B (D1) protein by Bedbrook *et al.* (1978; VI.2) and Zurawski *et al.* (1982; VI.3), and sequencing of the *RuBPCase* gene by McIntosh *et al.* (1980; VI.4).

2.7. Period VII (ca. 1980 - present)

In 1985, Fish, Kück and Bogorad found two genes (*psaA* and *psaB*) which encode the high-molecular-weight polypeptides in the P700-containing heterodimer of photosystem I (VII.1). Similar discoveries soon followed for a number of higher plants, eukaryotic algae and cyanobacteria (VII.2). The knowledge thus derived on the amino acid sequences of the *psaA* and *psaB* gene products had a tremendous impact on the studies of structure and function of the P700 complex. The recent avalanche of works on the structure and function of various systems (*e.g.*, see VII.25-28) through site-directed mutagenesis have added truly exciting new areas of research in photosynthesis.

Another trigger for explosive advances was a major breakthrough in the structure of the reaction center by X-ray crystallography, using crystals obtained from the purple non-sulfur bacterium *Rhodospseudomonas viridis*, by Deisenhofer, Michel, Huber and coworkers [1983; VII.13]; and subsequently by Norris, Schiffer and coworkers [1986]; and by Feher and coworkers [1986] for *Rhodobacter sphaeroides* (see under VII.13). Stimulated by this work, active research on similar structural analysis followed with other photosynthetic complexes: Kühlbrandt and Wang

(1991; VII.12) on the electron crystallographic analysis of the light-harvesting Chl *a/b* protein; and Witt and coworkers (1987; VII.14) and Ford and coworkers (1987; VII.15) and more recently by Almog *et al.* (1991; see under VII.15) on the photosystem-I reaction-center complex. Exciting new results on the crystal structure of photosystem-II reaction-center complexes have already begun to appear (VII.43). It is apparent that results from crystal-structure analysis and molecular biology have a huge synergistic impact on our understanding of the structure-function relationship in the photosynthetic apparatus.

Another development is the progress in the analysis of photosystem II: the field was renewed by the isolation of the oxygen-evolving photosystem-II subchloroplast particles by Bertsch, Babcock and Yocum in 1981 [VII.5], followed by the isolation of the photosystem-II core complex (Tang and Ki. Satoh [1985; VII.6]; Ka. Satoh, Ohno and Katoh [1985; VII.7], and finally the isolation of the photosystem-II reaction-center complex D1-D2-cyt-*b559* (Nanba and Satoh [1987; VII.8]), and the demonstration of a similarity as well as differences in structure between photosystem-II and purple photosynthetic bacteria. An extension of this work will greatly advance our understanding of the mechanism of photosynthesis.

The analysis of the charge accumulator in oxygen evolution, mentioned above, also has made progress: EPR signal of the S₂-state (Dismukes and Siderer [1980; VII.23] and others [see under VII.23]) and the S₃-state (Boussac and coworkers [1989; VII.24]); absorbance changes associated with the S-state changes (Brettel, Schlodder, Witt and coworkers [1984; VII.29]; Dekker, van Gorkom and coworkers [1984; VII.30]); involvement of amino acids in the oxygen evolution enzyme (Barry, Babcock *et al.* [1987; VII.25]; Debus and coworkers [VII.26]; Boussac, Rutherford and coworkers [1989; VII.24]).

At the present moment, the following seven areas form the goal of future photosynthesis research:

- (1) Genetic information and its relevance to the biosynthesis, structure, and function of the photosynthetic apparatus;
- (2) Structure and function of the reaction centers;
- (3) Regulation and mechanism of energy transfer;
- (4) Regulation and mechanism of electron transfer;
- (5) Regulation and mechanism of oxygen evolution;
- (6) Mechanism of photophosphorylation; and
- (7) Regulation and mechanism of carbon dioxide assimilation.

Towards these goals, various research workers, trained in diverse areas, are collaborating with each other and proceeding to form a huge network world wide, aiming at the final target of integrating information so that the mechanism of photosynthesis may be understood at the molecular as well as organismic levels.

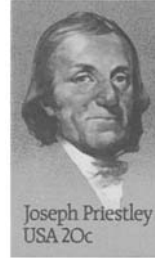
Acknowledgements

The authors thank Dr. Govindjee for his invitation to prepare this article and the history chart for publication in *Photosynthesis Research* and for his generous assistance and counsel in all phases of our preparation. We thank the following colleagues for furnishing the portraits used in the chart: that of Warburg from Dan Arnon; those of Emerson and Kok from Govindjee; and that of Hill from Berger Mayne. The authors are grateful to Dick Malkin for reading the chart and for suggesting entries. We also wish to thank Keiko Ke for assistance in translating the original Japanese text. We thank Elsevier Science Publishers for permission to reproduce the portrait of van Niel which originally appeared on p. xi of Topics in Photosynthesis, Vol. 8, *The Light Reactions*, edited by J. Barber (1987). Abbreviated versions of the history chart appeared previously in H. Huzisige, "Photosynthesis" (in Japanese), University of Tokyo Press (1982) and in B. Ke, "Photosynthesis" (in Chinese), Anhui Educational Publishing House, Hefei (1991). All typeset by BK.

PERIOD I [- ca. 1880]

- Rejection of the Aristotelian dogma that plants derive nourishment from the soil - van Helmont (1648) [1]
- Recognition that plants obtain nourishment from the atmosphere through the leaves - Mariotte (1679) [2]; Hales (1727) [3]
- Observation of the emission of gas bubbles by a submerged illuminated leaf - Bonnet (1754) [4]
- Discovery of the evolution of oxygen by plants - Priestley (1772) [5]
- Discovery of the rôle of light and green color in plant photosynthesis - Ingenhousz (1779) [6]
- "Fixed air" (CO₂) is involved in photosynthesis - Ingenhousz (1798) [7]; Senebier (1782) [8]
- Naming the green-plant pigment "chlorophyll" - Pelletier-Caventou (1818) [9]
- Recognition of the need for chlorophyll in plant photosynthesis - Dutrochet (1837) [10]
- Discovery of chloroplasts in plant cells - von Mohl (1844) [11]
- Proposal that plants convert light energy into chemical energy - Mayer (1845) [12]
- Rejection of the idea that plants are nourished by absorbing humus and water from the soil - De Saussure (1804) [13]; Liebig (1840) [14]
- Development of methods for growing plants in sand with nutrient solutions - Sachs (1853) [15]; Knop (1860-1865) [16]
- Measurement of the "photosynthetic quotient" - Boussingault (1864) [17]

Joseph Priestley



1733-1804



- Conclusion on the basic factors in photosynthesis;
- Recognition of the essential function of chlorophyll;
- Verification that starch is a product formed by photosynthesis;
- Sachs (1859-1862) [18]; Pfeffer (1874-1892) [19]
- Proposed the use of the term "photosynthesis" - Barnes (1893) [20]

PERIOD II [1880-1910]

PERIOD III [ca. 1910 - ca. 1938]

Observation of the chloroplast grana - Meyer (1882) [1]

Isolation of chloroplasts - Granick (1938) [1]

Recognition of the interrelationship between various parameters; Concept of limiting factors - Blackman (1905) [2]

- Quantitative manometry;
- Utilization of new experimental material (*Chlorella*);
- Use of inhibitors;
- Confirmation of light and dark reactions;
- Utilization of photochemical apparatus - Warburg (1911-1920) [2]

First determination of the quantum yield of photosynthesis - Warburg-Negelein (1922-1923) [3]

First flash experiments - Warburg (1919-1920) [2]

Concept of photosynthetic unit - Emerson-Arnold (1932) [4]

Structure of chlorophyll and identity of chlorophyll from different species - Willstätter-Stoll (1906-1926) [3]

First attempt on chlorophyll synthesis - Fischer (1926-1945) [5]

Concept of a chlorophyll-protein complex - "Chloroplastinsymplex" - Stoll (1936) [6]

Parallel measurement of fluorescence and carbon dioxide assimilation - Kautsky *et al.* (1931) [7]

Richard Martin Willstätter



1872-1942

Otto Heinrich Warburg



1883-1970

Cornelis Bernardus van Niel



1897-1985

Discovery of photosynthetic bacteria - Engelmann (1883) [4]; Winogradsky (1887) [5]

Interpretation of the primary photochemistry of photosynthesis as a light-induced oxido-reduction process - Wurmser (1925, 1930) [8]; van Niel (1929) [9]

Recognition that photosynthesis occurs inside the chloroplasts; confirmation of the effectiveness of light absorbed by chlorophyll - Engelmann (1881-1882) [6]

Molisch reaction - Molisch (1896) [7]

PERIOD IV [ca. 1938 - ca. 1954]

Early electron-microscope study of chloroplasts -
Kausche-Ruska (1940) [1];
Granick-Porter (1947) [2]

Electron microscopy of chloroplast structure -
Steinmann (1952) [3];
Frey-Wyssling - Steinmann (1953) [4]

The quantum-yield controversy -
Warburg-Emerson (1939-1951) [5]

Dependence of quantum yield on wavelength; discovery of "red drop" -
Emerson-Lewis (1943) [6]

Chlorophyll-containing photosynthetic unit -
Arnold-Kohn (1934) [7];
Gaffron-Wohl (1936-1941) [8]

Size limit for active chloroplasts -
Thomas-Blaauw-Duysens (1953) [9]

Discovery of P870 -
Duysens (1952) [10]

Evidence that chlorophyll was bound to proteins -
Smith (1938) [11]

Crystallization of chlorophyll lipoprotein -
Takashima (1952) [12]

Crystallization of chlorophylls -
Jacobs *et al.* (1953) [13]

Parallel measurement of fluorescence and photosynthesis -
McAlister-Myers (1940) [14]

Energy transfer between pigment molecules -
Förster (1948) [15];
Dutton *et al.* (1943) [16];
Duysens (1952) [10];
French-Young (1952) [17]

Discovery of delayed light emission -
Strehler-Arnold (1951) [18]

Robert (Robin) Hill



1899-1991

Robert Emerson



1903-1959

Introduced difference spectrophotometry -
Duysens (1952) [10]

Rôle of Mn in photosynthesis -
Pirson (1937) [19]

Hydrogen adaptation in photosynthesis -
Gaffron (1940) [20]

Oxygen in photosynthesis originates in water -
Ruben *et al.* (1941) [21]

Concept of the high-energy phosphate bond -
Lipmann (1941) [22]

First idea of a proton gradient -
Lundegårdh (1946) [23]

Discovery of the Hill reaction -
Hill (1937-1939) [24]

Discovery of cytochrome *f* -
Hill *et al.* (1951-1952) [25]

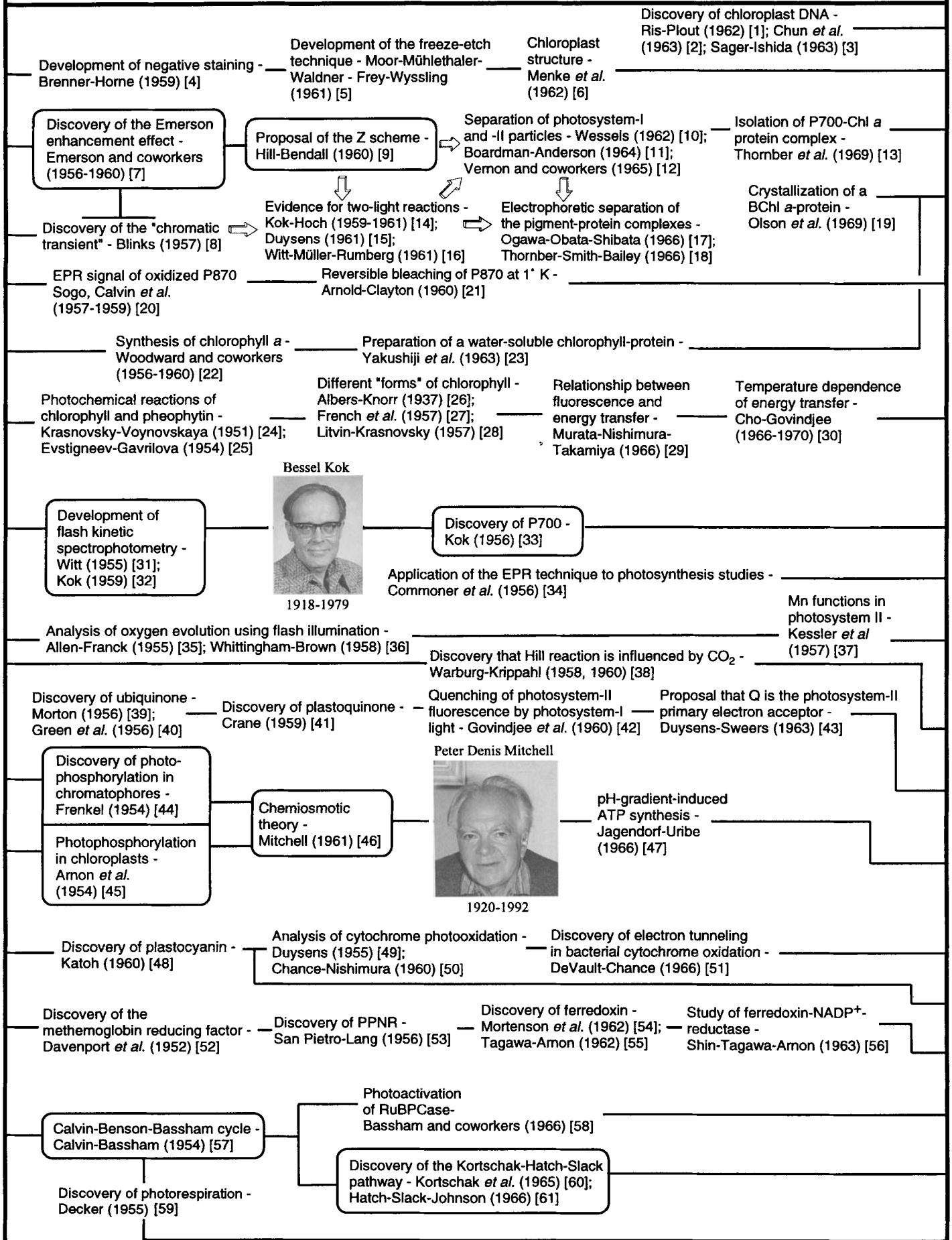
Discovery of the Mehler reaction - Mehler (1951) [26]

Photochemical reaction of NADP⁺ -
Vishniac-Ochoa; Tolmach;
Aron *et al.* (1951) [27,28,29]

Tracing the path of carbon in photosynthesis using radio carbon -
Ruben-Kamen-Hassid-DeVault (1939-1940) [30]

The path of carbon in photosynthesis -
Calvin-Benson *et al.* (1945-1954) [31];
Gaffron *et al.* (1945-1951) [32]

PERIOD V [ca. 1954 - ca. 1968]



PERIOD VI [ca. 1968 - ca. 1980]

Nuclear gene for LHCII identified - Kung-Thornber-Wildman (1972) [1] psbA gene for Q_B protein (D1) isolated - Bedbrook *et al.* (1978) [2]; sequenced - Zurawski *et al.* (1982) [3] RuBPCase gene sequenced - McIntosh *et al.* (1980) [4]

Model for the photosynthetic membrane - Miller *et al.*; Staehelin *et al.* (1976-77) [5,6] Lateral asymmetry in thylakoid membrane - Andersson-Anderson (1980) [7]

Separation and reconstitution of photosystem-I and -II particles - Huzisige *et al.* (1969) [8]; Briantais (1969) [9] Separation and analysis of photosystem-II subchloroplast particles - Huzisige (1972) [10]; Satoh-Butler (1978) [11] Isolation of the oxygen-evolving photosystem-II particles from thermophilic cyanobacteria - Stewart-Bendall (1979) [12]; and from red alga - Gantt- Clement-Montral (1983) [13] Photosystem II heterogeneity - Melis-Homann (1975-1976) [14]

Discovery of phycobilisomes - Gantt (1969) [15] Isolation of the light-harvesting Chl *a/b* protein - Thornber *et al.* (1971) [16] X-ray crystallography of a BChl *a*-protein - Fenna-Mathews (1976) [17] Discovery of chlorosomes - Staehelin *et al.* (1978) [18]

Isolation and analysis of the bacterial reaction center - Reed-Clayton (1969) [19]

Subunit structure of the bacterial reaction center - Okamura-Steiner-Feher (1974) [20]

Discovery of BPheo as the intermediary electron acceptor in photosynthetic bacteria - Dutton *et al.* (1975) [21]; Fajer *et al.* (1975) [22]; Parson and coworkers (1975) [23]; Shuvalov-Klimov (1976) [24]; van Grondelle *et al.* (1976) [25]

Split EPR signal of BPheo-Fe²⁺Q_A⁻ - Leigh-Dutton (1972) [26]; Feher *et al.* (1974) [27]

Discovery of P680 and X320 - Döring, Witt and coworkers (1969) [28]; Stiehl-Witt (1968) [29]

Discovery that P680 is a quencher of Chl *a* fluorescence - Butler (1972) [30]; Mauzerall (1972) [31]

Discovery of the photosystem-II primary electron acceptor - pheophytin - Klimov *et al.* (1970) [32]

Split EPR signal of Pheo-Fe²⁺Q_A⁻ - Klimov *et al.* (1980) [33]

Primary electron donor P700 is a "special pair" - Norris-Katz and coworkers (1971-79) [34]

Discovery of P430 (FeS-A/B) - Hiyama-Ke (1971) [35]

Discovery of photosystem-I iron-sulfur centers FeS-A/B - Malkin-Bearden (1971) [36]

Discovery of the electron acceptor FeS-X - McIntosh-Chu-Bolton (1975) [37]

EPR signal II_{VI} attributed to electron donor to P680⁺ - Blankenship-Babcock-Warden-Sauer (1975-1976) [38]

Loss of oxygen-evolution activity correlated with loss of Mn - Cheniae-Martin (1966) [39]

The linear four-step mechanism for photosynthetic oxygen evolution - Joliot *et al.* (1968-1969) [40]; Kok *et al.* (1970) [41]

Relation of thermoluminescence to the "oxygen clock" - Inoue *et al.* (1976) [42]

Tris inactivation of photosystem II and its reactivation - Yamashita-Butler (1968) [43]

Two-electron gate on the acceptor side of photosystem II - Bouges-Bocquet (1973) [44]; Velthuys-Amesz (1974) [45]

Two-electron gate in photosynthetic bacteria - Vermeglio (1977) [46]; Wraight (1977) [47]

CO₂-effect of Warburg is located at the two-electron gate of photosystem II - Govindjee and coworkers (1975, 1976) [48]

Rôle of ubiquinone in cyclic photophosphorylation in photosynthetic bacteria - Horio *et al.* (1968) [49]

Membrane potential and the electrochromic band shift - Junge-Witt (1968) [50]; Jackson-Crofts (1969) [51]

Photooxidation of cytochrome *b559* - Knaff-Arnon (1969) [52]

Isolation of cytochrome *b₆f* complex - Nelson-Neumann (1972) [53]; Hurt-Hauska (1981) [54]

Rieske Fe₂S₂ in photosynthetic bacteria and chloroplasts - Prince *et al.* (1975) [55]; Malkin-Aparicio (1975) [56]

Q-cycle - Mitchell (1976) [57]

Superoxide formation and dismutation - Asada-Kiso (1973) [59]

Crystal structure of plastocyanin - Colman *et al.* (1978) [58] Rôle of thioredoxin (Td) and Fd-Td-reductase - Buchanan *et al.* (1967-1976) [60]

Crystal structure of thioredoxin - Holmgren *et al.* (1975) [61]

RuBPCase subunit structure and biosynthesis - Kawashima-Wildman (1968) [62]

Regulation of carbon metabolism - Miyachi and coworkers (1970) [63]

Carboxylase/oxidase functions of Rubisco - Bowes-Ogren-Hageman (1971) [64]

Regulation mechanism of the Kortschak-Hatch-Slack cycle - Hatch-Slack (1969 review) [65]

Carbon isotope discrimination in photosynthesis - Smith-Epstein (1971) [66]

Discovery of the peroxisomes - Tolbert *et al.* (1968) [67]

PERIOD VII [ca. 1980 -]

psaA/psaB genes for P700-protein (maize) characterized -
Fish-Kück-Bogorad (1985) [1]; and others [2]

Leucine-zipper motif in the psaA/psaB heterodimer -
Webber-Malkin (1990) [3]; Kössel *et al.* (1990) [4]

Isolation of oxygen-evolving
photosystem-II particles from
spinach - Berthold-Babcock-
Yocum (1981) [5]

Isolation of the oxygen-evolving
photosystem-II core complex -
Tang-Satoh (1985) [6];
Satoh-Ohno-Katoh (1985) [7]

Isolation of the
D1-D2-Cyt-b559
complex -
Nanba-Satoh
(1987) [8]

PSI/PSII stoichiometry -
Melis *et al.* (1988) [9];
Graan-Ort-Whitmarsh
(1986-1990) [10]

Reversible phosphorylation of LHC and
regulation of excitation-energy distribution -
Bennett-Arntzen-Steinback-Allen *et al.*
(1977-1981) [11]

Electron crystallographic analysis
of the light-harvesting Chl *a/b*-protein
complex - Kühlbrandt-Wang (1991) [12]

X-ray structural analysis of
the bacterial reaction center -
Deisenhofer-Michel-Huber
(1982) [13]

Crystallization of the photosystem-I
reaction-center complex -
Witt *et al.* (1987-1993) [14];
Ford *et al.* (1987) [15]

Picosecond measurement of charge separation in photosystem II -
Wasielewski and coworkers (1989) [16]

P700 chlorophyll
may be chlorophyll *a'* (?) -
Watanabe
and coworkers
(1985) [17]

Identification of the
electron acceptor A_1 -
Brettel *et al.* (1986) [18];
Mansfield-Evans (1986) [19];
Ikegami and coworkers (1987) [20];
Biggins-Mathis (1988) [21];

Resolution and reconstitution
of PSI protein subunits -
Golbeck-Bryant and
coworkers (1990-1992) [22]

EPR signal of the S_2 -state -
Dismukes-Siderer (1980) [23]

EPR of the S_3 -
state -
Bousaac *et al.*
(1989) [24]

Tyrosine is the
electron donor Z -
Barry-Babcock
(1987) [25]

Tyr-161 (-160) is Z (D)
(electron donor) to P680⁺
Debus *et al.* (1988) [26];
Metz *et al.* (1989) [27];
Vermaas *et al.* (1988) [28]

Absorbance changes associated
with S-state changes -
Brettel-Schlodder-Witt (1984) [29];
Dekker-van Gorkom *et al.* (1984) [30]

Manganese involvement in
photosynthetic oxygen
evolution measured by
EXAFS - Klein, Sauer and
coworkers (1984) [31]

Interpretation
of thermo-
luminescence -
DeVault *et al.* (1989) [32]

Herbicide inhibits photosystem
by displacing Q_B -
Velthuys (1981) [33];
Wraight (1981) [34]

Isolation of *Cyt-bc₁*
complex -
Gabellini *et al.*
(1982) [35]

Isolation of genes of the
Cyt-bc₁ and *Cyt-b_{6/f}* complexes -
Daldal *et al.* (1987) [36];
Hauska *et al.* (1988) [37]

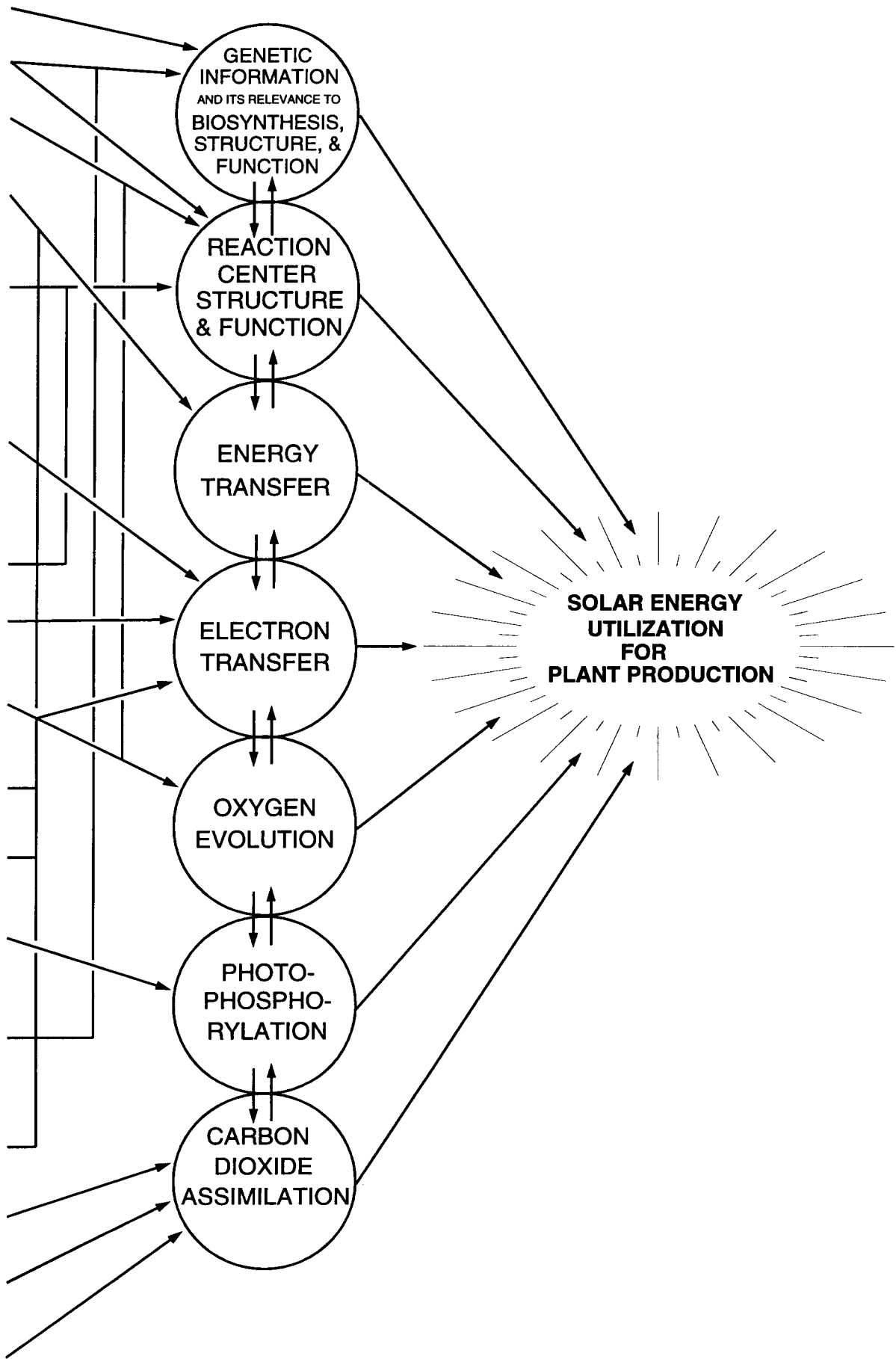
Preparation and characterization
of site-directed mutants of the
Cyt-bc₁ complex -
Daldal and coworkers (1992) [38]

Crystal structure of Fd-NADP⁺-reductase -
Karpus *et al.* (1988) [39]

Discovery of
activase -
Salvucci *et al.*
(1985) [40]

3-dimensional structure of Rubisco
from *Rsp rubrum* - Schneider-
Lindqvist-Brändén-Lorimer (1986) [41]

Carbon isotope discrimination
correlates with water-use efficiency -
Farquhar-Richards (1984) [42]



References

Period I:

- I. 1. **Van Helmont J-B** (1648) *Ortus Medicinae* pp. 108-109
(The Collected Works, edited by his son, FW van Helmont)
Amsterdam (Reprinted, Brussels, 1966)
- I. 2. **Mariotte E** (1679) *Essais de physique: premier essai de la végétation des plantes; seconde essai de physique de la nature de l'air*. Paris
- I. 3. **Hales S** (1727) *Vegetable staticks, or, an account of some statical experiments on the sap in vegetation*. W. Innys, London
- I. 4. **Bonnet C** (1754) *Recherches sur l'usage des feuilles dans les plantes*. Gött. et Leide
- I. 5. **Priestley J** (1772) *Observations on different kinds of air*. Phil Trans Roy Soc London 62:147-264
- I. 6. **Ingenhousz J** (1779) *Experiments upon vegetables, discovering their great power of purifying the common air in the sunshine, and of injuring it in the shade and at night*. Elmsley and Payne, London
- I. 7. **Ingenhousz J** (1798) *Über die Nahrung der Pflanzen und die Düngung des Bodens*. Voigt, Magazin, I (Hft. 2):97-105
- I. 8. **Senebier J** (1782) *Mémoires physicochimiques sur la l'influence de la lumière solaire pour modifier les êtres de trois règnes, surtout ceux du règne végétal*. 3 vols., Chirol, Geneva
- I. 9. **Pelletier J and Caventou JB** (1818) *Notice sur la matière verte des feuilles [chlorophylle]*. Ann Chim Phys IX:194-196
- I. 10. **Dutrochet H** (1837) *De la tendance des végétaux à se diriger vers la lumière*. Compt rend IV:48-50
- I. 11. **Von Mohl H** (1844) *Einige Bemerkungen über den Bau der vegetabilischen Zelle*. Botan Zeitung II:273-277
- I. 12. **Mayer JR** (1845) *Die organische Bewegung in ihrem Zusammenhang mit dem Stoffwechsel*. Heilbronn
- I. 13. **DeSaussure NTh** (1804) *Recherches chimique sur la Vegetation*. Nyon, Paris
- I. 14. **Liebig J** (1840) *Die organische Chemie in ihrer Anwendung auf Agrikultur und Physiologie*. Wieweg, Braunschweig
- I. 15. **Sachs J** (1853) *Über das Wachstum der Pflanzen*. Ziva pp 139-146, 229-236, 293-304, 336-343
- I. 16. **Knop W** (1860) *Über die Ernährung der Pflanzen durch wässerige Lösungen bei Ausschluss des Bodens*. Landw Versuchs-stat Dresden pp 65-99, 270-293
Knop W (1865) *Künstlicher Boden zu Vegetationsversuchen*. Landw Versuchs-stat Dresden pp 341-344
- I. 17. **Boussingault JB** (1864) *De la végétation dans l'obscurité*. Ann sci nat (Paris) I:314-324
- I. 18. **Sachs J** (1859) *Über den Auftreten der Stärke bei der Keimung ölhaltiger Saamen*. Botan Zeitung XVIII:177-183, 185-188
Sachs J (1862) *Übersicht der Ergebnisse der neueren Untersuchungen über das Chlorophyll*. Flora XLV:129-137
Sachs J (1862) *Über den Einfluß des Lichtes auf die Bildung des Amylums in den Chlorophyllkörnern*. Botan Zeitung 20:365-373
- I. 19. **Pfeffer W** (1874) *Die Wirkung farbigen Lichtes auf die Zersetzung der Kohlensäure in Pflanzen*. Würzburg, Botan Inst Arbeit 1:1-76
Pfeffer W (1881) *Pflanzenphysiologie, Ein Handbuch des Stoffwechsels und Kraftwechsels in der Pflanzen*. 2 vols, Leipzig [English edition (1900-1906), translated and edited by

A.J.Ewart: The Physiology of plants. A treatise upon the metabolism and sources of energy in plants. 3 vols, Oxford
Pfeffer W (1892) *Studien zur Energetik der Pflanzen*. Math Phys Abh (Leipzig) 18:149-276

- I. 20. **Barnes CR** (1893) *On the food of green plants*. Botan Gazette 18:403
Barnes CR (1898) *So-called assimilation*. Botan Centrbl LXXVI p.257

Period II:

- II. 1. **Meyer A** (1882) *Über Chlorophyllkörner, Stärkebildner und Farbkörper*. Botan Centrbl 12:314-317
- II. 2. **Blackman FF** (1905) *Optima and limiting factors*. Ann Botan 19:281-295
- II. 3. **Willstätter R and Stoll A** (1913) *Untersuchungen über Chlorophyll*. Justus Springer, Berlin (English translation by F.M. Schertz and A.R. Merz, Science Printing Press, Lancaster, Pennsylvania, 1928)
- II. 4. **Engelmann ThW** (1883) *Bacterium Photometricum: ein Beitrag zur vergleichenden Physiologie des Licht- und Farbensinnes*. Pflüger, Archiv Physiol 30:95-124
- II. 5. **Winogradsky S** (1887) *Über Schwefelbakterien*. Botan Zeitung 45:489 ff.
Winogradsky S (1888) *Zur Morphologie und Physiologie der Schwefelbakterien*. Arthur Felix, Leipzig
- II. 6. **Engelmann ThW** (1881) *Neue Methode zur Untersuchung der Sauerstoffausscheidung pflanzlicher und thierischer Organismen*. Botan Zeitung 39:441-448
Engelmann ThW (1882) *Über Sauerstoffausscheidung von Pflanzencellen im Mikrospectrum*. Botan Zeitung 40:419-426
Gest H ((1988) *Sunbeams, cucumbers and purple bacteria*. Photosynth Res 19:287-308
- II. 7. **Molisch H** (1896) *Eine neue mikrochemische Reaktion auf Chlorophyll*. Ber deut botan Ges 14:16-18
Gest H (1991) *The legacy of Hans Molisch (1856-1937), photosynthesis savant*. Photosynth Res 30:49-57
- II. 8. **Willstätter R and Stoll A** (1918) *Untersuchungen über Assimilation der Kohlensäure*. Springer, Berlin
- II. 9. **Von Baeyer A** (1864) *Über die Wasserentziehung und ihre Bedeutung für das Pflanzenleben und die Gährung*. Ber deut chem Ges 3:63

Period III:

- III. 1. **Granick S** (1938) *Quantitative isolation of chloroplasts from higher plants*. Am J Botany 25:558-561
- III. 2. **Warburg O** (1919-1920) *Über die Geschwindigkeit der photochemischen Kohlensäurezerersetzung in lebenden Zellen (I and II)* Biochem Z 100:230-270; 103:188-217
- III. 3. **Warburg O and Negelein E** (1923) *Über den Einfluß der Wellenlänge auf den Energieumsatz bei der Kohlensäure-assimilation*. Z Physik Chem (Leipzig) 106:191-218
- III. 4. **Emerson R and Arnold W** (1932) *A Separation of the reactions in photosynthesis by means of intermittent light*. J Gen Physiol 15:391-420
Emerson R and Arnold W (1932) *The photochemical reaction in photosynthesis*. J Gen Physiol 16:191-205
- III. 5. **Fischer H and Klarer J** (1926) *Synthesis of porphyrins. VII. Etioporphyrin from cryptopyrrole and hemopyrrole*. Ann Chem 450:181-201

- III. 6. **Stoll A** (1936) *Zusammenhänge zwischen der Chemie des Chlorophylls und seiner Funktion in der Photosynthese*. Naturwissen 24:53-60
- III. 7. **Kautsky H and Hirsch A** (1931) *Chlorophyllfluoreszenz und Kohlensäureassimilation*. Biochem Z 274:423-434
Kautsky H and Zedlith W (1941) *Fluorescence curves of chloroplast grana*. Naturwissen 29:101-102
Kautsky H and Frank U (1943) *Chlorophyllfluoreszenz und Kohlensäureassimilation*. IX. *Apparatur zur Messung rascher Lumineszenzänderung geringer Intensität*. Biochem Z 315: 139-155; X. *Die Chlorophyllfluoreszenz von Ulva lactuca und ihre Abhängigkeit von Temperatur und Lichtintensität*. *ibid* 315: 156-175; XI. *Die Chlorophyllfluoreszenz von Ulva lactuca und ihre Abhängigkeit von Narkotica, Sauerstoff und Kohlendioxyd*. *ibid*. 315:176-206; XII. *Zusammenfassung der bisherigen Ergebnisse und ihre Bedeutung für die Kohlen-säureassimilation*. *ibid*. 315:207-232
- III. 8. **Wurmser R** (1925) *Le rendement énergétique de la photo-synthèse chlorophyllienne*. Annales de physiologie et de physicochimie biologique. 1:47-63
Wurmser R (1930) *Oxydations et réductions*. Les presses universitaires de France, Paris
Wurmser R (1987) *Letter to the editor*. Photosynth Res 13:91-93
- III. 9. **Van Niel CB** (1929) *Photosynthesis in bacteria*. in "Contribution to Marine Biology" pp. 161-169, Stanford University Press, Stanford, California
Van Niel CB (1931) *On the morphology and physiology of the purple and green bacteria*. Arch Mikrobiol 3:1-12
Van Niel CB (1941) *The bacterial photosynthesis and their importance for the general problem of photosynthesis*. Adv Enz 1:263-328
Stanier RY (1961) *Photosynthetic mechanisms in bacteria and plants; a unitary concept*. Bacteriolog Revs 25:1-17
Gest H (1993) *History of concepts of the comparative biochemistry of oxygenic and anoxygenic photosyntheses*. Photosynth Res 35:87-96
- III. 10. **Thunberg T** (1923) *A new way for carbon dioxide to formaldehyde. A contribution to the theory of photosynthesis*. Z physik Chem 106:305-312
Weigert F (1923) *Photochemical comments regarding Thunberg's theory of photosynthesis*. Z physik Chem 106:313-323
- Period IV:
- IV. 1. **Kausche GA and Ruska H** (1940) *Über die Nachweis von Molekülen der Tabakmosaikviren in den Chloroplasten viruskranker Pflanzen*. Naturwissen 28:303-304
- IV. 2. **Granick S and Porter KR** (1947) *Structure of the spinach chloroplasts as interpreted with the electron microscope*. Am J Botany 34:545-550
- IV. 3. **Steinmann E** (1952) *Contribution to the structure of granular chloroplasts*. Experientia, 8:300-301
Steinmann E (1952) *An electron microscope study of the lamellar structure of chloroplasts*. Exptl Cell Res 3:367-372
- IV. 4. **Frey-Wyssling A and Steinmann E** (1953) *Ergebnisse der Feinbau-Analyse der Chloroplasten*. Vierteljahresschr Naturf Ges Zürich 98:20-29
- IV. 5. **Emerson R and Lewis CM** (1939) *Factors influencing the efficiency of photosynthesis*. Am J Botany 26:808-822
Emerson R and Lewis CM (1941) *Carbon dioxide exchange and the measurement of the quantum yield of photosynthesis*. Am J Botany 28:789-804
Warburg O (1948) *Assimilation quotient and photochemical yield*. Am J Botany 35:194-204
Nishimura MS, Whittingham CP and Emerson R (1951) *The maximum efficiency of photosynthesis. A critique of certain manometric methods used for measuring rates of photosynthesis*. Symp Exptl Biol 5:176-210
Rabinowitch E (1951) *Photosynthesis and related processes*. Vol II Pt 1. *Maximum quantum yield of photosynthesis*. pp 1083-1141; (1956) Addendum, Vol II Pt 2 pp 1940-1978 John Wiley-Interscience, New York
Emerson R and Chalmers RV (1955) *Transient changes in cellular gas exchange and the problem of maximum efficiency of photosynthesis*. Plant Physiol 30:504-529
Emerson R (1958) *The quantum yield of photosynthesis*. Annu Rev Plant Physiol 9:1-24
Govindjee R, Rabinowitch E and Govindjee (1968) *Maximum quantum yield and action spectra of photosynthesis and fluorescence*. Biochim Biophys Acta 162:539-544
- IV. 6. **Emerson R and Lewis CM** (1943) *The dependence of the quantum yield of Chlorella photosynthesis on wavelength of light*. Am J Botany 30:165-178
- IV. 7. **Arnold W and Kohn HI** (1934) *The chlorophyll unit in photosynthesis*. J Gen Physiol 18:109-112
- IV. 8. **Gaffron H and Wohl K** (1936) *Zur Theorie der Assimilation*. Naturwissen 24:81-90; 103-107
Wohl K (1937) *Zur Theorie der Assimilation*. I. *Theory of the assimilation unit*. Z Physik Chem Abt B 37:105-121; II. *The assimilation theory of Frank and Herzfeld*. *ibid* 122-147; III. *The dark reaction of assimilation. The Blackman reaction*. *ibid* 169-185; IV. *Mechanism of the assimilation unit*. *ibid* 186-208; V. *General summary*. *ibid* 209-230
Wohl K (1941) *Mechanism of photosynthesis in purple bacteria and green plants*. New Phytologist 40:34-55
- IV. 9. **Thomas JB, Blaauw OH and Duysens LNM** (1953) *On the relationship between size and photochemical activity of fragments of spinach grana*. Biochim Biophys Acta 10:230-239
- IV. 10. **Duysens LNM** (1952) *Transfer of excitation energy in photosynthesis*. Doctoral thesis, State University, Utrecht, the Netherlands
- IV. 11. **Smith EL** (1938) *Solutions of chlorophyll-protein compounds (phylochlorins) extracted from spinach*. Science 88:170-171
Smith EL (1941) *The chlorophyll-protein compound of green leaf*. J Gen Physiol 24:565-585
- IV. 12. **Takashima S** (1952) *Chlorophyll-protein obtained in crystals*. Nature 69:82-183
- IV. 13. **Jacobs EE, Vatter AE and Holt AS** (1953) *Crystalline chlorophyll and bacteriochlorophyll*. J Chem Phys 21:2246-2247
- IV. 14. **McAlister ED and Myers J** (1940) *The time course of photosynthesis and fluorescence observed simultaneously* Smithsonian Inst Pubs Misc Coll 99, #6
- IV. 15. **Förster Th** (1948) *Intermolecular energy transference and fluorescence*. Ann Physik 2:55-75
- IV. 16. **Dutton HJ, Manning WM and Duggar BB** (1943) *Chlorophyll fluorescence and energy transfer in diatom Nitzschia closterium*. J Gen Physiol 47:308-313
- IV. 17. **French CS and Young VK** (1952) *The fluorescence*

- spectra of red algae and the transfer of energy from phycoerythrin to phycocyanin and chlorophyll.* J Gen Physiol 35:873-890
- IV. 18. **Strehler BL and Arnold W** (1951) *Light production by green plants.* J Gen Physiol 34:809-820
Arnold WA (1991) *Experiments.* Photosynth Res 27:73-82
- IV. 19. **Pirson A** (1937) *A study of the nutrition and metabolism of Fontinalis and Chlorella.* Z Bot 31:193-267
- IV. 20. **Gaffron H** (1940) *Carbon dioxide reduction with molecular hydrogen in green algae.* Am J Botany 27:273-283
- IV. 21. **Ruben S, Randall M, Kamen MD and Hyde JL** (1941) *Heavy oxygen (¹⁸O) as tracer in the study of photosynthesis.* J Am Chem Soc 63:877-878
- IV. 22. **Lipmann F** (1941) *Metabolic generation and utilization of phosphate bond energy.* Adv Enz 1:99-162
Lipmann F (1971) *Wondering of a biochemist.* Wiley-Interscience, New York
- IV. 23. **Lundegårdh H** (1946) *Transport of water and salts through plant tissues.* Nature 157:575-578
Lundegårdh H (1947) *Mechanism of absorption, transport, accumulation, and secretion of ions.* Annu Rev Biochem 16:503-528
- IV. 24. **Hill R** (1937) *Oxygen evolved by isolated chloroplasts.* Nature 139:181-182
Hill R (1939) *Oxygen evolution by isolated chloroplasts.* Proc Roy Soc ser B 127:192-210
- IV. 25. **Hill R and Scarisbrick R** (1951) *The hematin compounds of leaves.* New Phytologist 50:98-111
Davenport HE and Hill R (1952) *The preparation and some properties of cytochrome f.* Proc Roy Soc ser B 139:327-345
- IV. 26. **Mehler A** (1951) *Studies on reactions of illuminated chloroplasts. I. Mechanism of the reduction of O₂ and other Hill reagents.* Arch Biochem Biophys. 33:65-77; II. *Stimulation and inhibition of the reaction with molecular oxygen.* ibid 34:339-351
- IV. 27. **Vishniac W and Ochoa S** (1951) *Photochemical reduction of pyridine nucleotides by spinach grana and coupled carbon dioxide fixation.* Nature 167:768-769
- IV. 28. **Tolmarch LJ** (1951) *Effects of triphosphopyridine nucleotide upon oxygen evolution and carbon dioxide fixation by illuminated chloroplasts.* Nature 167:946-948
- IV. 29. **Arnon DI** (1951) *Extracellular photosynthetic reactions.* Nature 167:1008-1010
- IV. 30. **Ruben S, Kamen MD, Hassid WZ and DeVault D** (1939) *Photosynthesis with radio-carbon.* Science 90:570-571
Ruben S, Kamen MD and Hassid WZ (1940) *Photosynthesis with radioactive carbon. II. Chemical properties of the intermediates.* J Am Chem Soc 62:3443-3450
Kamen MD (1989) *Onward into a fabulous half-century.* Photosynth Res 21:137-144
- IV. 31. **Calvin M and Benson AA** (1948) *Path of carbon in photosynthesis.* Science 107:476-480
- IV. 32. **Gaffron H, Fager EW and Rosenberg JL** (1951) *Intermediates in photosynthesis: Formation and transformation of phosphoglyceric acid.* Symp Exptl Biol 5:262-283
Gaffron H and Fager EW (1951) *The kinetics and chemistry of photosynthesis.* Annu Rev Plant Physiol 2:87-114
- IV. 33. **Jensen RG and Bassham JA** (1966) *Photosynthesis by isolated chloroplasts.* Proc Nat Acad Sci USA 56:1095-1101
- Period V:
- V. 1. **Ris H and Plout W** (1962) *Ultrastructure of DNA containing areas in chloroplasts of Chlamydomonas.* J Cell Biol 13:383-391
- V. 2. **Chun EHL, Vaughan, Jr MH and Rich A** (1963) *The isolation and characterization of DNA associated with chloroplast preparations.* J Mol Biol 7:130-141
- V. 3. **Sager R and Ishida MR** (1963) *Chloroplast DNA in Chlamydomonas.* Proc Nat Acad Sci USA 50:725-730
- V. 4. **Brenner S and Horne RW** (1959) *A negative staining method for high resolution electron microscopy of virus.* Biochim Biophys Acta 34:103-110
- V. 5. **Moor H, Mühlethaler K, Waldner H and Frey-Wyssling A** (1961) *A new freezing ultramicrotome.* J Biophys Biochem Cytol 10:1-13
- V. 6. **Menke W** (1962) *Structure and chemistry of plastids.* Annu Rev Plant Physiol 13:27-44
Menke W (1965) *Feinbau und Entwicklung der Plastiden.* Ber dtsch Bot Ges 77:340-354
Menke W (1972) *40 Jahre Versuche zur Aufklärung der molekularen Struktur der Chloroplasten.* Jahrbuch der Max-Planck-Gesellschaft zur Förderung der Wissenschaften E.V. pp. 132-155
Menke W (1990) *Retrospective of a botanist.* Photosynth Res 25:77-82
- V. 7. **Emerson R, Chalmers RV and Cederstrand C** (1957) *Some factors influencing the long-wave limit of photosynthesis.* Proc Nat Acad Sci USA 43:133-143
Emerson R and Rabinowitch E (1960) *Red drop and rôle of auxiliary pigments in photosynthesis.* Plant Physiol 35:477-485
Govindjee and Rabinowitch E (1960) *Two forms of chlorophyll a in vivo with distinct photochemical functions.* Science 132:355-356
Govindjee R, Rabinowitch E and Govindjee (1968) *Maximum quantum yield and action spectra of photosynthesis and fluorescence.* Biochim Biophys Acta 162:539-544
- V. 8. **Blinks LR** (1957) *Chromatic transients in photosynthesis of red algae.* Research in Photosynthesis (H Gaffron et al, eds) pp. 444-449. Interscience Publishers
- V. 9. **Hill R and Bendall F** (1960) *Function of the two cytochrome components in chloroplasts. A working hypothesis.* Nature 186: 136-137
- V. 10. **Wessels JSC** (1962) *Separation of the two photochemical systems of photosynthesis by digitonin fragmentation of spinach chloroplasts.* Biochim Biophys Acta 65:561-564
- V. 11. **Boardman NK and Anderson JM** (1964) *Isolation from spinach chloroplasts containing different proportions of chlorophyll a and chlorophyll b and their possible rôle in the light reactions of photosynthesis.* Nature 203:166-167
- V. 12. **Vernon LP and Shaw ER** (1965) *Photochemical activities of spinach chloroplasts following treatment with the detergent Triton X-100.* Plant Physiol 40:1269-1277
Vernon LP, Shaw ER and Ke B (1966) *A photochemically active particle derived from chloroplasts by the action of the detergent Triton X-100.* J Biol Chem 241:4101-4109
- V. 13. **Thornber JP** (1969) *Comparison of a Chl a-protein complex from a blue-green alga with chlorophyll-protein complexes obtained from green bacteria and higher plants.*

- Biochim Biophys Acta 172:230-241
Dietrich WE and Thornber JP (1971) *The P700 chlorophyll a-protein of a blue-green alga*. Biochim Biophys Acta 245:482-493
Shiozawa JA, Alberte RS and Thornber JP (1974) *The P700-chlorophyll a protein. Isolation and some characteristics of the complex in higher plants*. Arch Biochem Biophys 165:388-397
- V. 14. **Kok B and Hoch G** (1961) Spectral changes in photosynthesis. in "Light and life" (McElroy WD and Glass B, eds) pp. 397-416. Johns Hopkins Press, Baltimore
- V. 15. **Duysens LNM, Ames J and Kamp BM** (1961) *Two photochemical systems in photosynthesis*. Biochim Biophys Acta 190:188-190
Duysens LNM (1988) *The discovery of the two photosynthetic systems: a personal account*. Photosynth Res 21:61-79
- V. 16. **Witt HT, Müller A and Rumberg B** (1961) *Experimental evidence for the mechanism of photosynthesis*. Nature 191:194-195
Witt HT, Müller A and Rumberg B (1961) *Oxidized cytochrome and chlorophyll in photosynthesis*. Nature 192:967-969
- V. 17. **Ogawa T, Obata F and Shibata K** (1966) *Two pigment proteins in spinach chloroplasts*. Biochim Biophys Acta 112:223-234
- V. 18. **Thornber JP, Smith CA and Bailey JL** (1966) Partial characterization of two chlorophyll-protein complexes isolated from spinach-beet chloroplasts. Biochem J 100:14-15
- V. 19. **Olson JM, Filmer D, Radloff R, Romano C and Sybesma C** (1963) *The protein-chlorophyll-770 complex from green bacteria*, in "Bacterial photosynthesis" (Gest H, San Pietro A and Vernon LP, eds) pp. 423-431. Antioch Press, Yellow Springs, OH
- V. 20. **Sogo PB, Pon NG and Calvin M** (1957) *Photo spin resonance in chlorophyll-containing plant material*. Proc Nat Acad Sci USA 43:287-293
Sogo PG, Jost M and Calvin M (1959) *Evidence for free radical production in living cells exposed to ionizing radiation*. Radiation Res 1:511-518
- V. 21. **Arnold W and Clayton RK** (1960) *The first step in photosynthesis: Evidence for its electronic nature*. Proc Nat Acad Sci USA 46:769-776
- V. 22. **Woodward RB, Ayer WA, Beaton JM, Bickelhaupt F, Bonnett R, Buchschacher P, Closs GL, Dutler H, Hannah J, Hauck FP, Itô L, Angemann A, LeGoff E, Leimgruber W, Lwowski W, Sauer J, Valenta Z and Volz H** (1960) *The total synthesis of chlorophyll*. J Am Chem Soc 82:3800-3802
Woodward RB (1960) *Total synthesis of chlorophylls*. Angew Chem 72:651-662
- V. 23. **Yakushiji E, Uchino K, Sigimura Y, Shiratori I and Takamiya F** (1963) *Isolation of water-soluble chlorophyll protein from the leaves of Chenopodium album*. Biochim Biophys Acta 75:293-298
- V. 24. **Krasnovsky AA and Voynovskaya KK** (1951) *Reversible photochemical reduction and oxidation of bacteriochlorophyll and bacteriopheophytin*. Doklady Akad Nauk SSSR 81:879-882
Krasnovsky AA (1992) *Excited chlorophyll and related problems*. Photosynth Res 33:173-193
- V. 25. **Evstigneev VB and Gavrilova VA** (1954) *Photoreduction of pheophytins a and b*. Doklady Akad Nauk SSSR 96:1201-1204
- V. 26. **Albers VM and Knorr HV** (1937) *Forms of chlorophyll a in living plants*. Plant Physiol 12:833-843
- V. 27. **French CS and Huang HS** (1957) *The shape of the red absorption band of chlorophyll in live cells*. Carnegie Institution Washington Year Book No 56, pp. 266-268
French CS (1958) *Various forms of chlorophyll a in plants*. Brokhaveen Symp Biol 11:65-73
French CS and Elliott RF (1958) *The absorption spectra of chlorophylls in various algae*. Carnegie Institution Washington Year Book No 57, pp. 278-286
French CS (1957) *Derivative spectrophotometry*. Symp Instrumentation and control, Instr soc Am, Berkely, CA, USA
French CS (1960) *The chlorophylls in vivo and in vitro*, in "Encycl Plant Physiol" (Ruhland W, ed) 5:257-297 Springer
Brown JS and French CS (1961) *The long wavelength forms of Chl a*. Biophys J 1:539-550
Smith JHC and French CS (1963) *The major and accessory pigments in photosynthesis*. Annu Rev Plant Physiol 14:181-224
- V. 28. **Litvin FF and Krasnovsky AA** (1957) *An investigation of the intermediate stages of formation of chlorophyll in etiolated leaves based on measurement of the fluorescence spectra*. Doklady Akademii Nauk SSSR 117:106-109
- V. 29. **Murata N, Nishimura M and Takamiya A** (1966) *Fluorescence of chlorophyll in photosynthetic systems. I. Analysis of weak light effect in isolated chloroplasts*. Biochim Biophys Acta 112:213-222; *II. Induction of fluorescence in isolated chloroplasts*, *ibid* 120:23-33; *III. Emission and action spectrum of fluorescence. Three emission bands of Chl a and the energy transfer between pigment systems*. *ibid* 126:234-243
- V. 30. **Cho F, Spencer J and Govindjee** (1966) *Emission spectra of Chlorella at very low temperatures (-269° to -196°)* Biochim Biophys Acta 126:174-176
Cho F and Govindjee (1970) *Low-temperature (4-77° K) spectroscopy of Chlorella: temperature dependence of energy transfer efficiency*. Biochim Biophys Acta 216:139-150
Cho F and Govindjee (1970) *Low-temperature (4-77° K) spectroscopy of Anacystis: temperature dependence of energy transfer efficiency*. Biochim Biophys Acta 216:151-161
- V. 31. **Witt HT** (1955) *Kurzzeitige Absorptionsänderungen beim Primärprozess der Photosynthese*. Naturwissen 42:72-73
Witt HT (1955) *The primary processes of photosynthesis*. Z Elektrochem 59:981-986
Rüppel H and Witt HT (1969) *Measurement of fast reactions by single and repetitive excitation with pulses of electromagnetic radiation*. Methods Enzym 16:316-380 (Kustin K, ed) Acad Press, New York
Witt HT (1991) *Functional mechanism of water splitting photosynthesis*. Photosynth Res 29:55-77
- V. 32. **Kok B** (1959) *Light-induced absorption changes in photosynthetic organisms. II. A split-beam difference spectrophotometer*. Plant Physiol 34:184-192
- V. 33. **Kok B** (1956) *On the reversible absorption change at 705 m μ in photosynthetic organisms*. Biochim Biophys Acta 22:399-401
Kok B (1957) *Light-induced absorption changes in photosynthetic organisms*. Acta Botan Neerl 6:316-336
Kok B (1961) *Partial purification and determination of the oxidation reduction potential of the photosynthetic chlorophyll*

- complex absorbing at 700 m μ . *Biochim Biophys Acta* 48:527-533
- V. 34. **Commoner B, Heise JJ and Townsend J** (1956) *Light-induced paramagnetism in chloroplasts*. *Proc Nat Acad Sci USA* 42:710-718
- V. 35. **Allen FL and Franck J** (1955) *Photosynthetic oxygen evolution by flashes of light*. *Arch Biochem Biophys* 58:124-143
- V. 36. **Whittingham CP and Brown AH** (1958) *Oxygen evolution from algae illuminated by short and long flashes*. *J Exptl Botany* 9:311-319
- V. 37. **Kessler E, Arthur W and Brugger JE** (1957) *Influence of manganese on delayed light emission, fluorescence, photo-reduction, and photosynthesis in algae*. *Arch Biochem Biophys* 71:326-335
- V. 38. **Warburg O and Krippahl G** (1958) *Hill Reaktionen*. *Z Naturforsch* 13b:509-514
Warburg O and Krippahl (1960) *Notwendigkeit der Kohlen-säure für die Chinoxin und Ferricyanid-Reaktionen in grünen Grana*. *Z Naturforsch* 15b:367-369
Blubaugh DJ and Govindjee (1988) *The molecular mechanism of the bicarbonate effect at the plastoquinone reductase site of photosynthesis*. *Photosynth Res* 19:85-128
- V. 39. **Morton RA** (1958) *Ubiquinone*. *Nature* 182:1764-1767
Morton RA (1965) *Quinones as biological catalysts*. *Endeavour* 24:81-86
- V. 40. **Green DE, Hatefi Y and Fechner WF** (1959) *On the rôle of coenzyme Q in electron transport*. *Biochem Biophys Res Commun* 1:45-48
- V. 41. **Crane FL** (1959) *Isolation of two quinones with coenzyme Q activity from alfalfa*. *Plant Physiol* 34:546-551
- V. 42. **Govindjee, Ichimura S, Cederstrand C and Rabinowitch E** (1960) *Effect of combining far-red light with shorter wave light on the excitation of fluorescence in Chlorella*. *Arch Biochem Biophys* 89:322-323
- V. 43. **Duysens LNM and Sweers HE** (1963) *Mechanism of two photochemical reactions in algae as studied by means of fluorescence, in "Studies on Microalgae and Photosynthetic Bacteria"* pp. 353-372. Special issue of *Plant Cell Physiology*, Soc Plant Physiologists, University of Tokyo Press, Tokyo
- V. 44. **Frenkel AW** (1954) *Light-induced phosphorylation by cell-free preparations of photosynthetic bacteria*. *J Am Chem Soc* 76:5568-5569
Frenkel AW (1993) *Recollections*. *Photosynth Res*, 35:103-116
- V. 45. **Arnon DI, Allen MB and Whatley FR** (1954) *Photosynthesis by isolated chloroplasts*. *Nature* 174:394-396
Arnon DI, Whatley FR and Allen MB (1957) *Triphosphopyridine nucleotide as a catalyst of photosynthetic phosphorylation*. *Nature* 180:182-185
Arnon DI (1984) *The discovery of photosynthetic phosphorylation*. *TIBS* 9:258-262
- V. 46. **Mitchell P** (1961) *Coupling of phosphorylation to electron and hydrogen transfer by a chemiosmotic type of mechanism*. *Nature* 191:144-148
Mitchell P (1961) *Chemiosmotic coupling in oxidative and photosynthetic phosphorylation*. Glynn Res, Bodmin, Cornwall, England.
- V. 47. **Jagendorf AT and Uribe E** (1966) *ATP formation caused by acid-base transition of spinach chloroplasts*. *Proc Nat Acad Sci USA* 55:170-177
- V. 48. **Katoh S** (1960) *A new copper protein from Chlorella Ellipsoidea*. *Nature*, 186:533-534
- V. 49. **Duysens LNM** (1955) *Rôle of cytochrome and pyridine-nucleotide in algal photosynthesis*. *Science* 121:210-213
- V. 50. **Chance B and Nishimura M** (1960) *On the mechanism of chlorophyll-cytochrome interaction: The temperature insensitivity of light-induced cytochrome oxidation in Chromatium*. *Proc Nat Acad Sci USA* 46:19-24
- V. 51. **DeVault D and Chance B** (1966) *Studies of photosynthesis using a pulsed laser: Temperature dependency of cytochrome oxidation rate in Chromatium vinosum. Evidence for tunneling*. *Biophys J* 6:825-847
DeVault D (1989) *Tunneling enters biology*. *Photosynth Res* 22:5-10
- V. 52. **Davenport HE, Hill R and Whatley FR** (1952) *A natural factor catalyzing reduction of methemoglobin by isolated chloroplasts*. *Proc Roy Soc Ser. B* 139:346-358
- V. 53. **San Pietro A and Lang HM** (1956) *Accumulation of reduced pyridine nucleotide by illuminated grana*. *Science* 124: 118
San Pietro A and Lang HM (1958) *Photosynthetic pyridine nucleotide reductase. I. Purification and properties of the enzyme from spinach*. *J Biol Chem* 231:211-229
- V. 54. **Mortenson LE, Valentine RC and Carnahan JE** (1962) *An electron transport factor from Clostridium pasteurianum*. *Biochem Biophys Res Commun* 7:448-452
- V. 55. **Tagawa K and Arnon DI** (1962) *Ferredoxins as electron carriers in photosynthesis and in the biological production and consumption of hydrogen gas*. *Nature* 195:537-543
Tagawa K and Arnon DI (1968) *Oxidation-reduction potentials and stoichiometry of electron transfer in ferredoxin*. *Biochim Biophys Acta* 153:602-613
- V. 56. **Shin M, Tagawa K and Arnon DI** (1963) *Crystallization of ferredoxin-TPN reductase and its role in the photosynthetic apparatus of chloroplasts*. *Biochem Z* 338:84-96
- V. 57. **Calvin M and Bassham JA** (1962) *The path of carbon in photosynthesis*. WA Benjamin Inc, NY
Calvin M (1989) *Forty years of photosynthesis and related activities*. *Photosynth Res* 21:3-36
- V. 58. **Pedersen TA, Kirk M and Bassham JA** (1966) *Light-dark transients in levels of intermediate compounds during photosynthesis in air-adapted Chlorella*. *Physiol Plant* 19:219-231
- V. 59. **Decker JP** (1955) *A rapid postillumination deceleration of respiration in green leaves*. *Plant Physiol* 30:82-84
- V. 60. **Kortschak HP, Hartt CE and Burr GO** (1965) *Carbon dioxide fixation in sugarcane leaves*. *Plant Physiol* 40:209-213
- V. 61. **Hatch MD and Slack CR** (1966) *Photosynthesis by sugarcane leaves, A new carboxylation reaction and the pathway of sugar formation*. *Biochem J* 101:103-111
Hatch MD, Slack CR and Johnson HS (1967) *Further studies on a new pathway of photosynthetic carbon dioxide fixation in sugar-cane and its occurrence in other plant species*. *Biochem J* 102:417-422
Hatch MD (1992) *I can't believe my luck*. *Photosynth Res* 33:1-14
- Period VI:
- VI. 1. **Kung SD, Thornber JP and Wildman SG** (1972) *Nuclear DNA codes for the photosystem II chlorophyll-protein of chloroplast membranes*. *FEBS Lett* 24:185-188

- VI. 2. **Bedbrook JR, Link G, Coen DM, Bogorad L and Rich A** (1978) *Maize plastid gene expressed during photoregulated development*. Proc Nat Acad Sci USA 75:3060-3064
- VI. 3. **Zurawski G, Bohnert HJ, Whitfield PR and Bottomley W** (1982) *Nucleotide sequence of the gene for the M_r 32,000 thylakoid membrane protein from *Spinacia oleracea* and *Nicotiana debnevi* predicts a totally conserved translational product of M_r 38,950*. Proc Nat Acad Sci USA 79:7699-7703
- VI. 4. **McIntosh L, Poulson C and Bogorad L** (1980) *Chloroplast gene sequence for the large subunit of ribulose biphosphate-carboxylase of maize*. Nature 288:556-559
- VI. 5. **Miller KR and Staehelin LA** (1976) *Analysis of the thylakoid outer surface: Coupling factor is limited to unstacked membrane regions*. J Cell Biol 68:30-47
- VI. 6. **Staehelin LA, Armond PA and Miller KR** (1977) *Chloroplast membrane organization at the supramolecular level and its functional implications*. Brookhaven Symp Biol 28:278-315
- VI. 7. **Andersson B and Anderson JM** (1980) *Lateral heterogeneity in the distribution of chlorophyll-protein complexes of the thylakoid membranes of spinach chloroplasts*. Biochim Biophys Acta 593:427-440
Anderson JM and Andersson B (1982) *The architecture of photosynthetic membranes: The lateral and transverse organization*. Trends Biochem Sci 7:288-292
- VI. 8. **Huzisige H, Usiyama H, Kikuti T and Azi T** (1969) *Purification and properties of the photoactive particles corresponding to photosystem II*. Plant Cell Physiol 10:441-455
- VI. 9. **Briantais JM** (1969) *Reestablishment of a link between two particles isolated by Triton X-100*. Progr Photosyn Res, Proc Intern Congr (1969) 1:174-178. Verlag C. Lichtenstern, Munich
Ke B and Shaw ER (1972) *Reconstitution of photosystems I and II using fragments fractionated from spinach chloroplasts by Triton treatment*. Biochim Biophys Acta 275:192-198
Lam E and Malkin R (1982) *Reconstitution of the chloroplast noncyclic electron transport pathway from water to NADP with three integral protein complexes*. Proc Nat Acad Sci USA 79:5494-5498
- VI. 10. **Huzisige H and Yamamoto Y** (1972) *Analysis of photosystem II using particle II preparation. I. Experimental evidence supporting the idea of involvement of two light reactions in photosystem II of green plant photosynthesis*. Plant Cell Physiol 13:477-491; (1973) *II. Action spectra for Hill activities and fluorescence properties of variously-treated particle II preparations*. *ibid* 14:953-963
Huzisige H and Takimoto N (1974) *Analysis of photosystem II using particle II preparation. III. Roles of cytochrome b559 with different redox potentials and plastocyanin in the photosynthetic electron transport system*. Plant Cell Physiol 15:1099-1113
- VI. 11. **Satoh Ki and Butler WL** (1978) *Low temperature spectral properties of subchloroplast fractions purified from spinach*. Plant Physiol 61:373-379
- VI. 12. **Stewart AC and Bendall D** (1979) *Preparation of an active, oxygen-evolving photosystem-II particle from a blue-green alga*. FEBS Lett 107:308-312
Ke B, Inoue H, Babcock GT, Fang ZX and Dolan E (1982) *Optical and EPR characterization of oxygen-evolving system-II subchloroplast fragments isolated from the thermophilic blue-green alga *Phormidium laminosum**. Biochim Biophys Acta 682:297-306
- VI. 13. **Clement-Metral JD and Gantt E** (1983) *Active oxygen-evolving photosystem-II phycobilisome particle from *Porphyridium cruentum**. Advances in Photosynthesis Research, 6th Intern Congr Photosynthesis vol 1: pp 453-456
- VI. 14. **Melis A and Homann PH** (1975) *Kinetic analysis of the fluorescence induction in 3-(3,4-dichlorophenyl)-1,1-dimethylurea poisoned chloroplasts*. Photochem Photobiol 21:431-437
Melis A and Homann PH (1976) *Heterogeneity of the photochemical centers in system II of chloroplasts*. Photochem Photobiol 23:343-350
- VI. 15. **Gantt E** (1969) *Properties and ultrastructure of phycoerythrin from *Porphyridium cruentum**. Plant Physiol. 44: 1629-1638
- VI. 16. **Thorner JP** (1971) *Chlorophyll proteins: Light-harvesting and reaction center components of plants*. Annu Rev Plant Physiol 26:127-158
- VI. 17. **Fenna RE and Mathews BW** (1976) *Structure of a bacteriochlorophyll a-protein from *Prosthecochloris aestuarii**. Brookhaven Symp Biol 28:170-182
Mathews BW and Fenna RE (1980) *Structure of a green bacteriochlorophyll protein*. Accts Chem Res 13:309-317
Tronrud DE, Schmid MF and Mathews BW (1986) *Structure and amino acid sequence of a bacteriochlorophyll a protein from *Prosthecochloris aestuarii* at 1.9 Å resolution*. J Mol Biol 188:443-454
- VI. 18. **Staehelin LA, Golecki JR, Fuller RC and Drews G** (1978) *Visualization of the supramolecular architecture of chlorosome (*Chlorobium* type vesicles) in freeze-fractured cells of *Chloroflexus aurantiacus**. Arch Microbiol 119: 269-277
Sprague SG, Staehelin LA, DiBartolomeis MJ and Fuller RC (1980) *Isolation and development of chlorosomes in the green bacterium *Chloroflexus aurantiacus**. J Bacteriol 147: 1021-1031
Feick RG and Fuller RC (1984) *Topography of the photosynthetic apparatus of *Chloroflexus aurantiacus**. Biochemistry 23:3693-3700
- VI. 19. **Reed DW and Clayton RK** (1969) *Isolation of a reaction center fraction from *Rhodospseudomonas sphaeroides**. Biochem Biophys Res Commun 30:471-475
Clayton RK (1963) *Toward the isolation of a photochemical reaction center in *Rhodospseudomonas sphaeroides**. Biochim Biophys Acta 75:212-223
Clayton RK (1988) *Memories of many lives*. Photosynth Res 19:205-224
Gingras G and Jolchine G (1969) *Isolation of a P870-enriched particle from *Rhodospirillum rubrum**. Progr Photosynth Res (Tübingen) 1:209-216
Feher G (1971) *Some physical and chemical properties of a bacterial reaction-center particle and its primary photochemical reactants*. Photochem Photobiol 14:373-387
- VI. 20. **Okamura MY, Steiner LA and Feher G** (1974) *Characterization of reaction centers from photosynthetic bacteria. I. Subunit structure of the protein mediating the primary photochemistry in *Rhodospseudomonas sphaeroides* R-26*. Biochemistry 13:1394-1403
Steiner LA, Okamura MY, Lopes LA, Moskowitz E and Feher G (1971) *Characterization of reaction centers from*

- photosynthetic bacteria. II. Amino acid composition of the reaction center protein and its subunits in Rhodospseudomonas sphaeroides R-26.* Biochemistry 14:1403-1410
- VI. 21. **Dutton PL, Kaufmann KJ, Chance B and Rentzepis PM** (1975) *Picosecond kinetics of the 1250 nm band of the Rps. sphaeroides reaction center: The nature of the primary photochemical intermediate state.* FEBS Lett 60 275-280
- VI. 22. **Fajer J, Brune DC, Davis MS, Forman A and Spaulding LD** (1975) *Primary charge separation in bacterial photosynthesis: oxidized bacteriochlorophylls and reduced pheophytin.* Proc Nat Acad Sci USA 72:4956-4960
- VI. 23. **Rockley MG, Windsor MW, Cogdell RJ and Parson WW** (1975) *Picosecond detection of an intermediate in the photochemical reaction of bacterial photosynthesis.* Proc Nat Acad Sci USA 72:2251-2255
- VI. 24. **Shuvalov VA and Klimov VV** (1976) *The primary photoreaction in the complex cytochrome-P890-P760 (bacteriopheophytin-760) of Chromatium minutissimum at low redox potentials.* Biochim Biophys Acta 440:587-599
- VI. 25. **Van Grondelle R, Romijn JC and Holmes NG** (1976) *Photoreduction of the long-wavelength bacteriopheophytin in reaction centers and chromatophores of the photosynthetic bacterium Chromatium vinosum.* FEBS Lett 72:187-192
- VI. 26. **Leigh JS and Dutton PL** (1972) *The primary electron acceptor in photosynthesis.* Biochem Biophys Res Commun 46:414-421
- VI. 27. **Feher G, Isaacson RA, McElroy JD, Ackerson LC and Okamura MY** (1974) *On the question of the primary acceptor in bacterial photosynthesis: Manganese substitution for iron in reaction centers of Rhodospseudomonas sphaeroides R-26.* Biochim Biophys Acta 368:135-139
- VI. 28. **Döring G, Renger G, Vater J and Witt HT** (1969) *Properties of the photoactive chlorophyll a_H in photosynthesis.* Z Naturforsch 24b:1139-1143
- VI. 29. **Stiehl HH and Witt HT** (1968) *Die kurzzeitigen ultravioletten Differenzspektren bei der Photosynthese.* Z Naturforsch 23b:220-224
- Stiehl HH and Witt HT** (1969) *Quantitative treatment of the function of plastoquinone in photosynthesis.* Z Naturforsch 24b:1588-1598
- VI. 30. **Butler WL** (1972) *On the primary nature of fluorescence yield changes associated with photosynthesis.* Proc Nat Acad Sci USA 69:3420-3422
- VI. 31. **Mauzerall D** (1972) *Light-induced fluorescence changes in Chlorella, and the primary photoreactions for the production of oxygen.* Proc Nat Acad Sci USA 69:1358-1362
- Sonneveld A, Rademaker H and Duysens LNM** (1979) *Chlorophyll fluorescence as a monitor of nanosecond reduction of the photooxidized P680⁺ of photosystem II.* Biochim Biophys Acta 548:536-551
- VI. 32. **Klimov VV, Klevanik AV, Shuvalov VA, and Krasnovsky AA** (1977) *Reduction of pheophytin in the primary light reaction of photosystem II.* FEBS Lett 82:183-186
- VI. 33. **Klimov VV, Dolan E, Shaw ER and Ke B** (1980) *Interaction between the intermediary electron acceptor (pheophytin) and a possible plastoquinone-iron complex in photosystem-II reaction centers.* Proc Nat Acad Sci USA 77:7227-7231
- VI. 34. **Norris JR, Uphaus RA, Crespi HL and Katz JJ** (1971) *Electron spin resonance of chlorophyll and the origin of signal I in photosynthesis.* Proc Nat Acad Sci USA 68:625-628
- Shipman LL, Cotton TM, Norris JR and Katz JJ** (1976) *New proposal for structure of special pair chlorophyll.* Proc Nat Acad Sci USA 73:1791-1794
- Katz JJ** (1990) *Green thoughts in a green shade.* Photosynth Res 26:143-160
- VI. 35. **Hiyama T and Ke B** (1971) *A new photosynthetic pigment, "P430": Its possible rôle as the primary electron acceptor of photosystem I.* Proc Nat Acad Sci USA 68:1010-1013
- Ke B** (1973) *The primary electron acceptor of photosystem I.* Biochim Biophys Acta 301:1-33
- VI. 36. **Malkin R and Bearden AJ** (1971) *Primary reactions of photoreduction of a bound chloroplast ferredoxin at low temperature as detected by EPR spectroscopy.* Proc Nat Acad Sci USA 68:16-19
- Ke B, Hansen RE and Beinert H** (1973) *Oxidation-reduction potentials of bound iron-sulfur proteins of photosystem I.* Proc Nat Acad Sci USA 70:2491-2495
- Evans MCW, Reeves SG and Cammack R** (1974) *Determination of the oxidation-reduction potential of the bound iron-sulfur proteins of the primary electron acceptor complex of photosystem I in spinach chloroplasts.* FEBS Lett 49:111-114
- VI. 37. **McIntosh AR, Chu M and Bolton JR** (1975) *Flash photolysis electron spin resonance studies of the electron acceptor species at low temperature in photosystem I spinach chloroplast particles.* Biochim Biophys Acta 276:308-314
- Evans MCW, Sihra CK, Bolton JR and Cammack R** (1975) *Primary electron acceptor complex of photosystem I in spinach chloroplasts.* Nature 256:668-670
- VI. 38. **Babcock GT and Sauer K** (1975) *A rapid light-induced transient in electron paramagnetic resonance signal II activated upon inhibition of photosynthetic oxygen evolution.* Biochim Biophys Acta 376:315-328
- Blankenship RE, Babcock GT, Warden JT and Sauer K** (1975) *Observation of a new EPR transient in chloroplasts that may reflect the electron donor to photosystem II at room temperature.* FEBS Lett 51:287-293
- Warden JT, Blankenship and Sauer K** (1976) *A flash photolysis ESR study of photosystem II signal II_v, the physiological donor to P680⁺.* Biochim Biophys Acta 423:462-478
- VI. 39. **Cheniae GM and Martin IF** (1966) *Studies on the function of manganese in photosynthesis.* In Energy Conversion by the Photosynthetic Apparatus, Brookhaven Symp Biol #19:406-417
- VI. 40. **Joliot P and Joliot A** (1968) *A polarographic method for detection of oxygen production and reduction of Hill reagent by isolated chloroplasts.* Biochim Biophys Acta 153:625-634
- Joliot P, Barbieri G and Chabaud R** (1969) *Un nouveau modèle des centres photochimiques du système II.* Photochem Photobiol 10:309-329
- VI. 41. **Kok B, Forbush B and McGloin M** (1970) *Cooperation of charges in photosynthetic O₂ evolution. I. A linear four-step mechanism.* Photochem Photobiol 11:457-475
- Joliot P and Kok B** (1975) *Oxygen evolution in photosynthesis,* in "Bioenergetics of Photosynthesis," pp. 387-412 (Govindjee, ed) Acad Press, NY
- VI. 42. **Inoue Y, Ichikawa T and Shibata K** (1976) *Development of thermoluminescence bands during greening of wheat leaves*

- under continuous and intermittent illumination. *Photochem Photobiol* 23:125-130
- Demeter S and Govindjee** (1989) *Thermoluminescence from plants*. *Physiol plant* 75:121-130
- VI. 43. **Yamashita T and Butler WL** (1968) *Photoreduction and photophosphorylation with Tris-washed chloroplasts*. *Plant Physiol* 43:1978-1986
- Yamashita T, Tsuji J and Tomita G** (1971) *Reactivation of the Hill reaction of Tris-washed chloroplasts*. *Plant Cell Physiol* 12:117-126
- VI. 44. **Bouges-Bocquet B** (1973) *Electron transfer between the two photosystems in spinach chloroplasts*. *Biochim Biophys Acta* 314:250-256
- VI. 45. **Velthuys BR and Amez J** (1974) *Charge accumulation at the reducing side of photosystem II of photosynthesis*. *Biochim Biophys Acta* 333:85-94
- VI. 46. **Vermeglio A** (1977) *Secondary electron transfer in reaction centers of Rhodospseudomonas sphaeroides. Out-of-phase periodicity of two for the formation of ubisemiquinone and fully reduced ubiquinone*. *Biochim Biophys Acta* 459:516-524
- Vermeglio A and Clayton RK** (1977) *Kinetics of electron transfer between the primary and the secondary electron acceptors in reaction centers from Rhodospseudomonas sphaeroides*. *Biochim Biophys Acta* 461:159-165
- VI. 47. **Wraight CA** (1977) *Electron acceptors of photosynthetic bacterial reaction centers. Direct observation of oscillatory behaviour suggesting two closely equivalent ubiquinones*. *Biochim Biophys Acta* 459:525-531
- Wraight CA** (1979) *The rôle of quinones in bacterial photosynthesis*. *Photochem Photobiol* 30:767-776
- VI. 48. **Wydrzynski T and Govindjee** (1975) *A new site of bicarbonate effect in photosystem II of photosynthesis: evidence from chlorophyll fluorescence transients in spinach chloroplasts*. *Biochim Biophys Acta* 387:403-408
- Govindjee, Pulles MPJ, Govindjee R, Van Gorkom HJ and Duysens LNM** (1976) *Inhibition of the reoxidation of the secondary electron acceptor of photosystem II by bicarbonate depletion*. *Biochim Biophys Acta* 449:602-605
- VI. 49. **Okayama S, Yamamoto N, Nishikawa K and Horio T** (1968) *Rôles of ubiquinone-10 and rhodoquinone in photosynthetic formation of adenosine triphosphate by chromatophores from Rhodospirillum rubrum*. *J Biol Chem* 243:2995-2999
- VI. 50. **Junge W and Witt HT** (1968) *On the ion-transport system of photosynthesis – Investigation on a molecular level – Z Naturforsch* 23b:244-254
- VI. 51. **Jackson and Crofts A** (1969) *High-energy state in chromatophores from Rhodospseudomonas sphaeroides*. *FEBS Lett* 4:185-189
- VI. 52. **Knaff DB and Arnon DI** (1969) *Light-induced oxidation of a chloroplast b-type cytochrome at -189°C*. *Proc Nat Acad Sci USA* 63:956-962
- VI. 53. **Nelson N and Neumann J** (1972) *Isolation of a cytochrome b₆f particle from chloroplasts*. *J Biol Chem* 247:1817-1824
- Ke B, Sugahara K and Shaw ER** (1975) *Further purification of "Triton Subchloroplast Fraction I" (TSF-I) particles. Isolation of a cytochrome-free high-P700 particle and a complex containing cytochrome f and b₆, plastocyanin and iron-sulfur protein(s)*. *Biochim Biophys Acta* 408:12-25
- VI. 54. **Hurt E and Hauska G** (1981) *A cytochrome b₆f complex of five polypeptides with plastoquinol-plastocyanin-oxidoreductase activity from spinach chloroplasts*. *Eur J Biochem* 117:591-599
- VI. 55. **Prince RC, Lindsay JG and Dutton PL** (1975) *The Rieske iron-sulfur center in mitochondrial and photosynthetic systems: E_m/pH relationships*. *FEBS Lett* 51:108-111
- VI. 56. **Malkin R and Aparicio PJ** (1975) *Identification of a g=1.90 high-potential iron-sulfur protein in chloroplasts*. *Biochem Biophys Res Commun* 63:1157-1160
- VI. 57. **Mitchell P** (1976) *Vectorial chemistry of the molecular mechanics of chemiosmotic coupling. Power transmissions by protonicity*. *Biochem Soc Trans* 4:399-430
- Mitchell P** (1976) *Possible molecular mechanism of the protonmotive function of cytochrome systems*. *J Theor Biol* 62:327-367
- VI. 58. **Colman PM, Freeman HC, Guss GM, Murata M, Norris VA, Ramshaw JAM and Venkatappa MP** (1978) *X-ray crystal structure analysis of plastocyanin at 2.7 Å resolution*. *Nature* 272:319-324
- Guss JM and Freeman HC** (1983) *Structure of oxidized poplar plastocyanin at 1.6 Å resolution*. *J Mol Biol* 169:521-563
- Collyer CA, Guss JM, Sugimura Y, Yoshizaki F and Freeman HC** (1990) *Crystal structure of plastocyanin from a green alga, Entermorpha prolifera*. *J Mol Biol* 211:267-632
- VI. 59. **Asada K and Kiso K** (1973) *Initiation of aerobic oxidation of sulfite by illuminated spinach chloroplasts*. *Eur J Biochem* 33: 253-257
- VI. 60. **Buchanan BB, Kalberer PP and Arnon DI** (1967) *Ferredoxin-activated fructose diphosphatase in isolated chloroplasts*. *Biochem Biophys Res Commun* 29:74-79
- Schürmann P, Wolosiuk RA, Breazeale VD and Buchanan BB** (1976) *Two proteins function in the regulation of photosynthetic CO₂ assimilation in chloroplasts*. *Nature* 263:257-258
- Buchanan BB** (1980) *Thioredoxin*. *Annu Rev Plant Physiol* 31:341-374
- VI. 61. **Holmgren A, Soderberg B-O, Eklund H and Brändén C-I** (1975) *Three-dimensional structure of E. coli thioredoxin-S2 to 2.8 Å resolution*. *Proc Nat Acad Sci USA* 72:2305-23097
- VI. 62. **Kawashima N and Wildman SG** (1970) *Fraction I protein*. *Annu Rev Plant Physiol* 12:325-358
- Kawashima N and Wildman SG** (1972) *Studies on fraction I protein. IV. Mode of inheritance of primary structure in relation to whether chloroplast or nuclear DNA contain the codes for chloroplast protein*. *Biochim Biophys Acta* 262:42-49
- VI. 63. **Ogasawara N and Miyachi S** (1970) *Regulation of CO₂ fixation in Chlorella by light of varied wavelength and intensities*. *Plant Cell Physiol* 11:1-14
- Hogetsu D and Miyachi S** (1970) *Effect of oxygen on the light enhanced dark CO₂ fixation in Chlorella cells*. *Plant Physiol* 45:178-182
- Miyachi S and Hogetsu D** (1970) *Effects of preillumination with light of different wavelengths on subsequent dark CO₂ fixation in Chlorella cells*. *Can J Bot* 48:1203-1207
- VI. 64. **Bowes G, Ogren WL and Hageman RH** (1971) *Phosphoglycolate production catalysed by ribulose diphosphate carboxylase*. *Biochem Biophys Res Commun* 45:716-722

- Laing WA, Ogren WL and Hageman RH** (1975) *Bicarbonate stabilization of ribulose 1,5-diphosphate carboxylase*. *Biochemistry* 14:2269-2275
- VI. 65. **Hatch MD and Slack CR** (1970) *Photosynthetic CO₂-fixation pathways*. *Annu Rev Plant Physiol* 21:141-162
- VI. 66. **Smith B and Epstein S** (1971) *Two categories of ¹³C/¹²C ratio for higher plants*. *Plant Physiol* 47:380-384
- VI. 67. **Tolbert NE, Oeser A, Kisaki T, Hageman RH and Yamasaki RK** (1968) *Peroxisomes from spinach leaves containing enzymes related to glycolate metabolism*. *J Biol Chem* 243:5179-5184
- Tolbert NE** (1981) *Metabolic pathways in peroxisomes and glyoxysomes*. *Annu Rev Biochem* 50:133-157
- Period VII:
- VII. 1. **Fish LE, Kück U and Bogorad L** (1985) *Two partially homogeneous adjacent light-inducible maize chloroplast genes encoding polypeptides of the P700 chlorophyll a protein complex of photosystem I*. *J Biol Chem* 260:1413-1421
- VII. 2. **Lehmbeck J, Rasmussen OF, Bookjans GB, Jepsen BR, Stummann BM and Henningsen KW** (1986) *Sequence of two genes in pea chloroplast DNA coding for 84 and 82 kD polypeptides of the photosystem-I complex*. *Plant Mol Biol* 7:3-10
- Ohyama K, Fukuzawa H, Kohchi T, Shirai H, Sano T, Sano S, Umesono K, Shiki Y, Takeuchi M, Chang Z, Aota S, Inokuchi H and Ozeki H** (1986) *Chloroplast gene organization deduced from complete sequence of liverwort *Marchantia polymorpha* chloroplast DNA*. *Nature* 322:572-574
- Shinozaki K, Ohme M, Tanaka M, Wakasugi T, Hayashida N, Matsubayashi T, Zaita N, Chunwongse J, Obokata J, Yamaguchi-Shinozaki K, Ohto C, Torazawa K, Meng BY, Sugita M, Deno H, Kamogashira T, Yamada K, Kusuda J, Takaiwa F, Kato R, Tohdoh N, Shimada H and Sugiura M** (1986) *The complete nucleotide sequence of the tobacco chloroplast genome: its gene organization and expression*. *EMBO J* 5:2043-2049
- Kirsch W, Seyer and P Herrmann RG** (1987) *Nucleotide sequence of the clustered genes for two P₇₀₀ chlorophyll a apoproteins of the photosystem-I reaction center and the ribosomal protein S14 of the spinach plastid chromosome*. *Curr Genet* 10:843-855
- Kück U, Choquet Y, Schneider M, Dron M and Bennoun P** (1987) *Structural and transcription analysis of two homologous genes for the P700 chlorophyll a-apoproteins in *Chlamydomonas reinhardtii*: evidence for in vivo transsplicing*. *EMBO J* 6:2185-2195
- Cantrell A and Bryant DA** (1987) *Molecular cloning and nucleotide sequence of the *psaA* and *psaB* genes of the cyanobacterium *Synechococcus* sp. PCC 7002*. *Plant Mol Biol* 9:453-468
- Cushman JC, Hallick RB and Price CA** (1988) *The two genes for the P₇₀₀ chlorophyll a apoproteins on the *Euglena gracilis* chloroplast genome contain multiple introns*. *Curr genet* 13:159-171
- Hiratsuka J, Shimada H, Whittier R, Ishibashi T, Sakamoto M, Mori M, Kondo C, Honji Y, Sun C-R, Meng BY, Li YQ, Kanno A, Nishizawa Y, Hirai A, Shinozaki K and Sugiura M** (1989) *The complete sequence of the rice (*Oryza sativa*) chloroplast genome: intermolecular recombination between distinct tRNA genes accounts for a major plastid DNA inversion during the evolution of cereals*. *Mol Gen Genet* 217:185-194
- VII. 3. **Webber AN and Malkin R** (1990) *Photosystem-I reaction-center proteins contain leucine zipper motifs. A proposed role in dimer formation*. *FEBS Lett* 264:1-4
- VII. 4. **Kössel H, Döry I, Igloi G and Maier R** (1990) *A leucine-zipper motif in photosystem I*. *Plant Mol Biol* 15:497-499
- Smart LB, Warren PV, Golbeck JH and McIntosh L** (1993) *Mutational analysis of the structure and biogenesis of the photosystem-I reaction center in the cyanobacterium *Synechocystis* sp. PCC 6803*. *Proc Nat Acad USA* 90:1132-1136.
- VII. 5. **Berthold DA, Babcock GT and Yocum CF** (1981) *A highly resolved oxygen-evolving photosystem II preparation from spinach thylakoid membranes*. *FEBS Lett* 134:231-234
- Yamamoto Y, Ueda T, Shinkai H and Nishimura M** (1982) *Preparation of O₂-evolving photosystem-II subchloroplasts from spinach*. *Biochim Biophys Acta* 679:347-350
- Kuwabara T and Murata N** (1982) *Inactivation of photosynthetic oxygen evolution and concomitant release of three polypeptides in the photosystem II particles of spinach chloroplasts*. *Plant Cell Physiol* 23:533-539
- VII. 6. **Tang XS and Satoh Ki** (1985) *The oxygen-evolving photosystem-II core complex*. *FEBS Lett* 179:60-64
- VII. 7. **Satoh Ka, Ohno T and Katoh S** (1985) *An oxygen-evolving complex with a simple subunit structure – “a water-plastoquinone-oxidoreductase” – from the thermophilic cyanobacterium, *Synechococcus* sp.* *FEBS Lett* 180:326-330
- Ghanotakis DF, Demetriou DM and Yocum CF** (1987) *Isolation and characterization of an oxygen-evolving photosystem-II reaction center preparation and a 28 kDa Chl-a binding protein*. *Biochim Biophys Acta* 891:15-21
- VII. 8. **Nanba O and Satoh Ki** (1987) *Isolation of a photosystem-II reaction center containing D1 and D2 polypeptides and cytochrome b559*. *Proc Nat Acad Sci USA*. 84:109-112
- Ghanotakis DF, dePaula JC, Demetriou DM, Bowlby NR, Petersen J, Babcock GT and Yocum CF** (1989) *Isolation and characterization of the 47 kDa protein and the D1-D2-cytochrome b-559 complex*. *Biochim Biophys Acta* 974:44-53
- Gounaris K, Chapman DJ and Barber J** (1989) *Isolation and characterization of a D1/D2/cytochrome b559 complex from *Synechococcus* 6803*. *Biochim Biophys Acta* 973:296-301
- VII. 9. **Melis A, Guenther GE, Morrissey PJ and Ghirardi ML** (1988) *Photosystem II heterogeneity in chloroplasts*. In “Applications of Chlorophyll Fluorescence,” (Lichtenthaler HK, ed.) pp. 33-43. Kluwer Acad Publ, Dordrecht
- VII. 10. **Graan T and Ort DR** (1986) *Detection of oxygen-evolving photosystem II centers inactive in plastoquinone reduction*. *Biochim Biophys Acta* 852:320-330
- Ort DR and Whitmarsh J** (1990) *Inactive photosystem II centers: a resolution of discrepancies in photosystem II quantitation*. *Photosynth Res* 23:101-104
- VII. 11. **Bennett J** (1977) *Phosphorylation of chloroplast membrane polypeptides*. *Nature* 269:344-346
- Bennett J** (1979) *Chloroplast phosphoproteins: Phosphorylation of polypeptides of the light-harvesting chlorophyll protein*

- complex. *Eur J Biochem* 99:133-137
- Bennett J** (1980) *Chloroplast phosphoproteins: Evidence for a thylakoid-bound phosphoprotein phosphatase*. *Eur J Biochem* 104:85-89
- Bennett J, Steinback K and Arntzen CJ** (1980) *Chloroplast phosphoproteins: Regulation of excitation energy transfer by phosphorylation of thylakoid-membrane polypeptides*. *Proc Nat Acad Sci USA* 77:5253-5257
- Allen JF, Bennett J, Steinback K and Arntzen CJ** (1981) *Chloroplast protein phosphorylation couples plastoquinone redox state to distribution of excitation energy between photosystems*. *Nature* 291:25-29
- VII. 12. **Kühlbrandt W and Wang DN** (1991) *Three dimensional structure of plant light-harvesting complex determined by electron crystallography*. *Nature* 350:130-134
- VII. 13. **Deisenhofer J, Epp O, Mikki K, Huber R and Michel H** (1984) *X-ray structural analysis of a membrane protein complex. Electron density map at 3 Å resolution and a model of the chromophores of the photosynthetic reaction center from Rhodospseudomonas viridis*. *J Mol Biol* 180:385-398
- Deisenhofer J and Michel H** (1991) *High resolution structures of photosynthetic reaction centers*. *Annu Rev Biophys Chem* 20:247-266
- Chang CH, Tiede D, Tang J, Smith U, Norris J and Schiffer M** (1986) *Structure of Rhodospseudomonas sphaeroides reaction center*. *FEBS Lett* 205:82-86
- Norris J and Schiffer M** (1990) *Photosynthetic reaction centers in bacteria*. *Chem Eng News* July 30, pp. 22-37
- Allen JP, Feher G, Yeates TO, Komiyama H and Rees DC** (1987) *Structure of the reaction center from Rhodospseudomonas sphaeroides R-26: The cofactors*. *Proc Nat Acad Sci USA* 84:5730-5734
- Allen JP, Feher G, Yeates TO, Rees DC, Deisenhofer J, Michel H and Huber R** (1986) *Structure homology of reaction centers from Rhodospseudomonas sphaeroides and Rhodospseudomonas viridis as determined by x-ray diffraction*. *Proc Nat Acad Sci USA* 83:8589-8593
- Feher G, Allen JP, Okamura MY and Rees DC** (1989) *Structure and function of bacterial photosynthetic reaction centers*. *Nature* 339:111-116
- VII. 14. **Witt I, Witt HT, Gerken S, Sängler W, Dekker JP and Rögner M** (1987) *Crystallization of reaction center I of photosynthesis. Low concentration crystallization of photoactive protein complexes from the cyanobacterium Synechococcus sp.* *FEBS Lett* 221:260-264
- Witt HT, Krauß N, Hinrichs W, Witt I, Fromme P and Saenger W** (1992) *Three-dimensional structure of Synechococcus sp. photosystem I reaction center with a 6 Å resolution*. *Proc. 9th Intern Congr Photosynth I:521-528*
- Krauß N, Hinrichs W, Witt I, Fromme P, Pritzkow W, Dauter Z, Betzel C, Wilson S, Witt HT and Saenger W** (1993) *Three-dimensional structure of system I of photosynthesis at 6 Å resolution*. *Nature* 361:326-331
- VII. 15. **Ford RC, Picot D and Garavito RM** (1987) *Crystallization of the photosystem I reaction center*. *EMBO J* 6:1581-1586
- Almog O, Shoham G, Michaeli D and Nechustai R** (1991) *Monomeric and trimeric forms of photosystem I reaction center of Mastigocladus laminosus: Crystallization and preliminary characterization*. *Proc Nat Acad Sci USA* 88:5312-5316
- VII. 16. **Wasielewski MR, Johnson DG, Seibert M and Govindjee** (1989) *Determination of the primary charge separation rate in isolated photosystem II reaction centers with 500 femtosecond time resolution*. *Proc Natl Acad Sci USA* 86:524-528
- Wasielewski MR, Johnson DG, Govindjee, Preston C and Seibert M** (1989) *Determination of the primary charge separation rate in photosystem II reaction centers at 15 K*. *Photosynth Res* 22:89-100
- VII. 17. **Watanabe T, Nakazato M, Mazeki H, Hongu A, Konno M, Saitoh S and Honda K** (1985) *Chlorophyll a epimer and pheophytin a in green leaves*. *Biochim Biophys Acta* 807:110-117
- Watanabe T, Kobayashi M, Hongu A, Nakazato M, Hiyama T and Murata N** (1985) *Evidence that chlorophyll a' dimer constitutes the photochemical reaction center 1 (P700) in photosynthetic apparatus*. *FEBS Lett* 191:252-256
- Watanabe T, Kobayashi M, Maeda H, Oba T, Yoshida S, van den Meent EJ and Ames J** (1992) *Function of the C13'-epimer chlorophylls in type I photosystem reaction centers*. *Proc. 9th Intern Congr Photosynth III:3-10*
- VII. 18. **Brettel K, Sétif P and Mathis P** (1986) *Flash-induced absorption changes in photosystem I at low temperature: evidence that the electron acceptor A₁ is vitamin K₁*. *FEBS Lett* 203:220-224
- Brettel K** (1988) *Electron transfer from A₁⁻ to an iron-sulfur center with t_{1/2}=200 ns at room temperature in photosystem I. Characterization by flash absorption spectroscopy*. *FEBS Lett* 239:93-98
- VII. 19. **Mansfield RW and Evans MCW** (1986) *UV optical difference spectrum associated with the reduction of electron acceptor A₁ in photosystem I of higher plants*. *FEBS Lett* 203:225-229
- VII. 20. **Ikegami I, Sétif P and Mathis P** (1987) *Absorption studies of photosystem I photochemistry in the absence of vitamin K₁*. *Biochim Biophys Acta* 894:414-422
- Itoh S, Iwaki M and Ikegami I** (1987) *Extraction of vitamin K₁ from photosystem I particles by treatment with diethyl ether and its effects on the A₁⁻ EPR signal and system I photochemistry*. *Biochim Biophys Acta* 893:508-516
- VII. 21. **Biggins J and Mathis P** (1988) *Functional rôle of vitamin K₁ in photosystem I of the cyanobacterium Synechocystis 6803*. *Biochemistry* 27:1494-1500
- VII. 22. **Golbeck JH and Bryant DA** (1991) *Photosystem I*, in "Current Topics in Bioenergetics," vol. 16, pp. 83-177. *Acad Press, New York*
- VII. 23. **Dismukes GC and Siderer Y** (1980) *Intermediates of a polynuclear manganese center involved in photosynthetic oxygen evolution*. *FEBS Lett* 121:78-80
- Dismukes GC and Siderer Y** (1980) *Intermediates of a polynuclear manganese center involved in photosynthetic oxidation of water*. *Proc Nat Acad Sci USA* 78:274-278
- Hansson Ö and Andréasson L-E** (1982) *EPR detectable magnetically interacting manganese ions in the photosynthetic oxygen-evolving system after continuous illumination*. *Biochim Biophys Acta* 679:261-268
- Brudvig GW, Casey JL and Sauer K** (1983) *Properties of the S₂ state associated with O₂ evolution*. *Biochim Biophys Acta* 723:366-371
- VII. 24. **Boussac A, Zimmermann JL and Rutherford AW** (1989) *EPR signals for modified charge accumulation states of*

- the oxygen-evolving enzyme in Ca^{2+} -deficient photosystem II. *Biochemistry* 28:8984-8989
- VII. 25. **Barry BA and Babcock GT** (1987) Tyrosine radicals are involved in the photosynthetic oxygen evolving system. *Proc Nat Acad Sci USA* 84:7099-7103
- Babcock GT, Barry BA, Debus RJ, Hoganson CW, Atamian M, McIntosh L, Sithole I and Yocum CF** (1989) Water oxidation in photosystem 2. From radical chemistry to multielectron chemistry. *Biochemistry* 28:9557-9565
- VII. 26. **Debus RJ, Barry BA, Sithole I, Babcock GT and McIntosh L** (1988) Directed mutagenesis indicates that the donor to P680^{*} in photosystem II is tyrosine-161 of the D1 polypeptide. *Biochemistry* 27:9071-9074
- Debus RJ, Barry BA, Babcock GT and McIntosh L** (1988) Site-directed mutagenesis identifies a tyrosine radical involved in the photosynthetic oxygen-evolving system. *Proc Nat Acad Sci USA* 85:427-430
- VII. 27. **Metz JG, Nixon PJ, Rögner M, Brudvig GW and Diner BA** (1989) Directed alteration of the D1 polypeptide of photosystem II: Evidence that tyrosine-161 is the redox component, Z, connecting the oxygen-evolving complex to the primary electron donor, P680. *Biochemistry* 28:6960-6969
- VII. 28. **Vermaas WFJ, Rutherford AW and Hansson Ö** (1988) Site-directed mutagenesis in photosystem II of the cyanobacterium *Synechocystis* sp PCC 6803: Donor D is a tyrosine residue in the D2 protein. *Proc Nat Acad Sci USA* 85:8477-8481
- VII. 29. **Brettel K, Schlodder E and Witt HT** (1984) Nanosecond reduction kinetics of photooxidized chlorophyll a_n (P680) in single flashes as a probe for the electron pathway. H^+ -release and charge accumulation in the O_2 -evolving complex. *Biochim Biophys Acta* 766:403-415
- Witt HT, Schlodder E, Brettel K and Saygin Ö** (1986) Reaction sequences from light absorption to the cleavage of water in photosynthesis. *Ber Bunsenges Physik Chem* 90:176-179
- VII. 30. **Dekker JP, van Gorkom HJ, Wensink J and Ouwehand L** (1984) Absorbance difference spectra of the successive redox states of the oxygen-evolving apparatus of photosynthesis. *Biochim Biophys Acta* 767:1-9
- Dekker JP, Plijter JJ, Ouwehand L and Van Gorkom HJ** (1984) Kinetics of manganese redox titrations in the oxygen-evolving apparatus of photosynthesis. *Biochim Biophys Acta* 767:176-179
- Lavergne J** (1991) Improved UV-visible spectra of the S-transitions in the photosynthetic oxygen-evolving system. *Biochim Biophys Acta* 1060:175-188
- VII. 31. **Goodin DB, Yachandra VK, Britt RD, Sauer K and Klein MP** (1984) The state of manganese in the photosynthetic apparatus determined by X-ray absorption spectroscopy. *J de Physique, Colloque C8*, supplement No. 12, pp. 1121-1128
- Sauer K, Guiles RD, McDermott AE, Cole J, Yachandra VK, Zimmermann J-L, Klein MP, Dexheimer SL and Britt RD** (1988) Spectroscopic studies of manganese in photosynthetic oxygen evolution. *Chim Scripta* 28A:87-91
- VII. 32. **DeVault D, Govindjee and Arnold W** (1983) Energetics of photosynthetic glow peaks. *Proc Nat Acad Sci USA* 80:983-987
- VII. 33. **Velthuys BR** (1981) Electron dependent competition between plastoquinone and inhibitors for binding to photosystem II. *FEBS Lett* 126:277-281
- VII. 34. **Wraight C** (1981) Oxidation reduction physical chemistry of the acceptor quinone complex in bacterial photosynthetic reaction centers: evidence for a new model of herbicide activity. *Isr J Chem* 21:248-354
- VII. 35. **Gabellini N, Boywer JR, Hurt E, Melandri BA and Hauska H** (1982) A cytochrome bc_1 complex with ubiquinol-cytochrome c_2 oxidoreductase activity from *Rhodospseudomonas sphaeroides* GA. *Eur J Biochem* 126:105-111
- VII. 36. **Daldal F, Davidson E and Cheng S** (1987) Isolation of the structural genes for the Rieske Fe-S protein, cytochrome b and cytochrome c_1 - all components of the ubiquinol-cytochrome c_2 oxidoreductase complex of *Rhodospseudomonas capsulata*. *J Mol Biol* 195:1-12
- VII. 37. **Hauska G, Nitschke W and Herrmann RG** (1988) Amino acid identities in the three redox center-carrying polypeptides of the cytochrome $bc_1/b_6/f$ complexes. *J Bioenerg Biomembr* 20:211-228
- VII. 38. **Davidson E, Ohnishi T, Atta-Asafo-Adjei E and Daldal F** (1992) Potential ligands to the [2Fe-2S] Rieske cluster of the cytochrome bc_1 complex of *Rb. capsulatus* probed by site-directed mutagenesis. *Biochemistry* 31:3342-3351
- Davidson E, Ohnishi T, Tokito M and Daldal F** (1992) *Rhodobacter-capsulatus* mutants lacking the Rieske FeS protein form a stable cytochrome bc_1 subcomplex with an intact quinone-occupancy Q_o site model. *Biochemistry* 31:3351-3358
- Gray KA, Davidson E and Daldal F** (1992) Mutagenesis of methionine-183 drastically affects the physicochemical properties of cytochrome c_1 of the bc_1 complex of *Rhodobacter capsulatus*. *Biochemistry* 31:11864-11873
- VII. 39. **Karplus PA, Daniels MJ and Herriott JR** (1991) Atomic structure of ferredoxin-NADP⁺ reductase: prototype for a structurally novel flavoenzyme family. *Science* 251:60-66
- VII. 40. **Salvucci ME, Portis, Jr AR and Ogren WL** (1985) A soluble chloroplast protein catalyzes ribulosebiphosphate carboxylase/oxygenase in vivo. *Photosynth Res* 7:193-201
- VII. 41. **Schneider G, Lindqvist Y, Brändén C-I and Lorimer G** (1986) Three-dimensional structure of ribulose-1,5-bisphosphate carboxylase/oxygenase from *Rhodospirillum rubrum* at 2.9 Å resolution. *EMBO J* 5:3411-3415
- Schneider G, Lindqvist Y and Lindqvist T** (1990) Crystallographic refinement and structure of ribulose-1,5-bisphosphate carboxylase from *Rhodospirillum rubrum* at 1.7 Å resolution. *J Mol Biol* 211:989-1008
- Knight S, Andersson T and Brändén C-I** (1990) Crystallographic analysis of ribulose-1,5-bisphosphate carboxylase from spinach at 2.4 Å resolution. Subunit interactions and active site. *J Mol Biol* 215:113-160
- VII. 42. **Farquhar GD and Richards RA** (1984) Isotopic composition of plant carbon correlates with water-use efficiency of wheat genotypes. *Aust J Plant Physiol* 11:539-552
- VII. 43. **Adir N, Okamura MY and Feher G** (1992) Crystallization of the photosystem-II reaction center. *Proc. 9th Intern Congr photosynth II*:195-198 □