HYPOTHESIS

Big babies and infant leukemia: a role for insulin-like growth factor-1?

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Several epidemiologic studies have demonstrated that high birthweight is associated with an increased risk of infant leukemia; however, the reason for this relationship is unclear. Biologic data demonstrate that birthweight is correlated positively with circulating levels of insulin-like growth factor-1 (IGF-1). IGF-1 is important in blood formation and regulation and has been shown to stimulate the growth of both myeloid and lymphoid cells in culture. Since infants who develop leukemia are likely to have had at least one transforming event occur *in utero*, we hypothesize that high levels of IGF-1 may both produce a larger baby and contribute to leukemogenesis. *Cancer Causes and Control*, 1996, 7, 553-559

Key words: Birthweight, childhood, IGF-1, infant, leukemia.

Introduction

The majority of epidemiologic studies of childhood leukemia have shown that there is an increased risk of leukemia (approximately twofold) in high birthweight babies, or a deficit of low birthweights among leukemia cases (Table 1).^{1-7,10-11,13,15-16} Moreover, many of the statistically significant associations have been confined to younger children, particularly those diagnosed at less than two years of age.^{6-7,15-16} In earlier studies, some of the positive associations have been found only in males⁵ or have reached statistical significance only in females.^{3,4,7} However, it is difficult to evaluate these gender associations since it is unclear how many cases were infants. In particular, it is important to note that infant leukemia is more common in females, whereas childhood leukemia overall is more common in males.¹⁷ A minority of studies^{8,9,12,14} have failed to demonstrate an association between high birthweight and risk of leukemia. Some earlier studies ascertained cases through death certificates.^{1-4,9} Since it is unknown whether children with higher birthweights experience a better prognosis, this method of case ascertainment could bias the odds ratio toward the null. Overall, we suggest that there is compelling evidence to indicate that high birthweight is a risk factor for infant leukemia.

Increased birthweight also has been associated with other childhood malignancies including Wilms' tumor, neuroblastoma, and astrocytomas.¹⁸ In a recent analysis of over 1,800 children in the National Wilms' Tumor Study, investigators reported that children with Wilms' tumor weighed significantly more at birth than did newborns in the general population.¹⁹ Babies with Beckwith-Wiedemann syndrome (BWS) have generalized macrosomia and associated high birthweight, and have a greatly increased risk of Wilms' tumor over the general population. Patients with BWS recently have been shown to have biallelic (as opposed to the normal monoallelic) expression of insulin-like growth factor II (IGF-2).²⁰ Biallelic expression of IGF-2 also has been demonstrated

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Author (year)	Study type (<i>n</i> = leukemia cases)	Definition of high birthweight	Birthweight findings with leukemia (comments)		
MacMahon (1962) ¹	acMahon (1962) ¹ Birth and death certificates: childhood cancer overall and by leukemia (n = 1,323)		Positive association; no age stratification provided		
lversen (1966) ²	Birth and death certificates: leukemia only $(n = 258)$	Not applicable ^a	Deficit of low birthweights in children with leukemia		
Jackson (1968) ³	Twins: leukemia only (<i>n</i> = 50)	Not available	Nearly 70 percent of the leukemic twins were heavier than the non-leukemic co-twin ^{b,c}		
Fasal (1971) ⁴	Case-control: ieukemia only ($n = 802$)	Males (> 9 lb); Females (> 8.5 lb)	Overall odds ratio (OR) = 1.40; males 1.10, females 2.07 ^d		
Wertelecki (1973) ⁵	Sibships: leukemia only $(n = 72)^{e}$	Not applicable ^a	Higher birthweight ranks for cases within sibships ^f		
Hirayama (1980) ⁶	Tumor registry: childhood cancer overall and by leukemia (n = 4,421)	> 4000 g	For cases < 2 years of age at diagnosis, statistically significant higher risk (69%) compared with cases who weighed ≤ 3400 g		
Daling (1984) ⁷	Birth certificates and cancer registry: childhood cancer overall and by leukemia (n = 681)	> 4000 g	For cases < 2 years of age, positive association ^{b,d}		
Shaw (1984) ⁸	Case-control: leukemia only (n = 255)	Not applicable ^a	No association; no age stratification provided		
Eisenberg (1987) ⁹	Birth certificates and death certificates: childhood cancer overall and by leukemia $(n = 1,304)^9$	Not applicable	No association when examining trends for younger age leukemias (including infant deaths, or 1-10 years of age at death)		
Shu (1988) ¹⁰	Case-control: ALL and AML ⁹ (<i>n</i> = 309)	> 3,500 g	For cases < 6 years of age, OR = 1.7, 95% CI = 1.2-2.6); elevated risks observed for both ALL and AML		
Kaye (1991) ¹¹	Birth certificates and cancer registry: ALL (<i>n</i> = 337)	> 3,800 g	For cases < 4 years of age, OR = 2.04 (95% Cl = 1.34-3.12)		
Savitz (1994) ¹²	Case-control: ALL (n = 71)	> 4,000 g	Overall OR = 0.7 (95% CI = 0.2-2.3); no association for cases < 4 years of age ^a		
Cnattingius (1995) ¹³	Case-control: (ALL) (n = 613)	> 4,500 g	For cases < 1 year of age OR = 3.2 (95% Cl = 0.5-19.7); [for ≥ 5 years of age OR = 1.7 (95% Cl = 1.1-2.6)		
Cnattingius (1995) ¹⁴	Case-control: AML (n = 98)	Not available	No association ^b		
Ross (in press) ¹⁵	Case-control: infant AML and ALL (n = 303)	> 4,000 g	OR = 2.28 (95% CI = 1.26-4.13); elevated risks observed for both AML and ALL		
Yeazel (submitted) ¹⁶	Case-control: childhood leukemia overall and by leukemia (n = 1,687)	> 4,000 g	For cases < 2 years of age at diagnosis; ALL, OR = 1.64 (95% CI = 0.97-2.76); AML, OR = 2.46 (95% CI = 1.11-5.47)		

Table 1. Epidemiologic studies examining birthweight and childhood leukemia

^a Comparison of mean birthweights only.
^b Odds ratios (OR) not provided.
^c Comparison of twin birth order and sex of the twin pair revealed statistically significant association only for females.
^d Statistically significant only for females (P < 0.05).

^e Age at diagnosis is unclear. ^f Statistically significant only for males (P < 0.05). ^g ALL = acute lymphoblastic leukemia; AML = acute myeloid leukemia.

in Wilms' tumor tissue of patients with and without BWS.²¹ This indicates that both systemic and local overproduction of growth factors can contribute to development of cancer, as was hypothesized for Wilms' tumor.^{19,22}

In this paper, we suggest that growth factors may play a similar role in the development of infant leukemia. In particular, insulin-like growth factor-1 (IGF-1) plays a major role in hematopoiesis and is a major determinant of birthweight (described below). We hypothesize that IGF-1 contributes to leukemogenesis, and that this manifests in some infants as increased birthweight.

Insulin-like growth factor-1 (IGF-1)

The expression of IGF-1 has been reported in a variety of cells and tissue including liver, bone, heart, kidney, colon, and monocytes.²³ The IGF-1 gene maps to the long arm of chromosome 12; at least five exons have been identified. IGF-1 exerts its biological activity by binding to the IGF-1 receptor, which results in the phosphorylation of several signal transduction molecules.²³ In the circulation, the IGFs are bound to binding proteins, which function as regulators of IGF biologic availability; currently six IGF binding proteins (IGFBP) have been identified.²⁴ The major circulating IGFBP (and thus, most biologically significant) is IGFBP-3; however, in infants, IGFBP-1 and IGFBP-2 also circulate at high levels.

Growth factors and birthweight

In normal infants, many studies have demonstrated a positive correlation between IGF-1 levels in umbilical cord blood and birthweight^{25-26,28-29,32,34-35,37}or fetal size,²⁷ with only a single contradictory study³¹ (Table 2). Moreover, 40 percent lower IGF-1 levels (compared with normal infants) have been reported in small-for-gestational age newborns, whereas nearly 30 percent higher IGF-1 levels have been found in large-for-gestational age

Table 2. Selected infant cord blood parameters and correlations with birthweight and fetal size (where available)

Author (year)	Sample measured	Outcome measured	IGF-1	IGF-2	IGF-BP1	IGF-BP2	IGF-BP3
Bennett (1983) ²⁵	Cord blood	Birthweight	Positive (0.32)	Positive (0.46 ^a)		_	
Gluckman (1983) ²⁶	Cord blood	Birthweight	Positive (0.37 ^a)	No correlation	_	-	_
Ashton (1985) ²⁷	Cord blood	Fetal weight (15-23 wks gestation)	Positive ^a	No correlation		-	_
Samaan (1987) ²⁸	Cord blood	Birthweight	Positive ^a	_	_	—	_
Lasarre (1991) ²⁹	Cord blood	Birthweight and fetal size	Positive ^a	No correlation		-	_
Salardi (1991) ³⁰	Fetal cord blood	Femoral length (19-37 wks gestation)	Not sensitive enough	_		-	_
Wang (1991) ³¹	Cord blood	Birthweight	Negative correlation ^a	—	Negative correlation		
Delmis (1992) ³²	Cord blood	Birthweight and fetal size	Positive correlation (0.41) ^a		_	-	_
Kubota (1992) ³³	Maternal plasma	Fetal size	Positive correlation (0.61) ^a	Positive correlation (0.36)	_	-	-
Fant (1993) ³⁴	Cord blood	Birthweight	Positive correlation (0.39) ^a	Positive correlation (0.26)	No correlation	Negative correlation (-0.26)	Positive correlation (0.48) ^a
Verhaeghe (1993) ³⁵	Cord blood	Birthweight	Positive correlation (0.48) ⁸	Positive correlation (0.23) ^a	Negative correlation (-0.43) ^a	-	—
Reece (1994) ³⁶	Cord blood	Birthweight	Positive correlation	No correlation	_		No correlation
Simmons (1995) ³⁷	Cord blood	Birthweight	Positive correlation (0.43) ^a			-	

^a Statistically significant (p < 0.05).

newborns.³⁵ In animal studies, IGF-1 is a major determinant of fetal size, as mice lacking the IGF-1 gene are morphologically normal, but are approximately 70 percent smaller than their normal littermates.³⁸

Growth factors, hematopoiesis, and leukemia

Hematopoiesis, as a mesodermal developmental process, is regulated by a number of well-characterized growth factors. These growth factors can be divided into those that regulate early and multilineage bone marrow progenitors [e.g. granulocytic macrophage colony stimulating factors (GM-CSF), stem cell factor (SCF), interleukin-3 (IL3)], and those that function in a lineagerestricted manner (e.g., erythropoietin).39 In addition, there is strong evidence that IGF-1 is important in blood formation including: (i) receptors for IGF-1 are found on cells of hematopoietic origin including monocytes, lymphocytes, erythrocytes, and platelets; (ii) IGF-1 plays a role in regulation of normal B-lymphocyte development; and (iii) IGF-1 stimulates erythropoiesis.40-44 Moreover, in vitro studies show that IGF-1 can stimulate the growth of both myeloid and lymphoid leukemia cells.43

Leukemia in the infant

As study of the genetic basis of cancer has progressed (e.g., retinoblastoma⁴⁵), it is becoming apparent that tumorigenesis due to mutation in a single gene is probably the exception, not the rule. A more frequent process is a multi-step pathway to tumor development, with accumulation of mutations in a series of genes necessary for true malignant evolution, as has been proposed for colon cancer.46 Evidence exists that leukemogenesis is a multistep phenomenon also, with mutations identified in many oncogenes and tumor suppressor genes, none of which alone seem to be sufficient for malignant transformation. This means that individuals necessarily harbor a significant premalignant clone for some time before clinical diagnosis (although this has not yet been demonstrated in infants). Limpens et al47 recently demonstrated that many normal individuals harbor the translocation, t(14;18), in their B-cells – a translocation clinically associated with follicular lymphoma. Additional evidence indicating the presence of a substantial preleukemic cell-population is the finding of cells containing t(8;21) in marrow of patients in long-term remission of leukemia who do not subsequently relapse.⁴⁸ This implies that while the truly malignant population has been eliminated by therapy, a premalignant clone remains. These findings suggest that a one- or two-hit model is not sufficient for tumorigenesis in many leukemias. Mathematical models have been produced that suggest that leukemogenesis may require the accumulation of at least four mutations or 'hits.'49

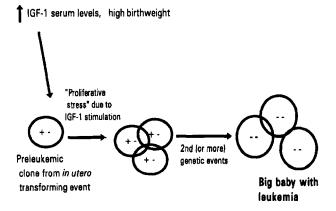
Molecular evidence indicates that some children who

develop leukemia in the first few years of life have had at least one transforming event occur *in utero*; this is clearly true for infants diagnosed in the first few weeks of life.⁵⁰⁻⁵³ In particular, nearly 80 percent of infants with leukemia present with an abnormality involving the *MLL* gene on chromosome band 11q23.¹⁸ Taken together, these data indicate that very young children who develop leukemia likely have a population of preleukemic or leukemic cells while *in utero*.

Hypotheses

There is compelling evidence to indicate that high birthweight is associated with an increased risk of infant leukemia, and with increased circulating levels of IGF-1. Moreover, IGF-1 can stimulate the proliferation of at least a large subset of leukemia cells.40,43-44 One hypothesis to explain the excess of leukemia in high birthweight babies may be that existing preleukemic cells are subjected to 'proliferative stress' by the higher levels of IGF-1 in larger babies (Figure 1). (Alternatively, there could be a decrease in the availability of IGF-1 binding proteins, thus allowing more free IGF-1 in the circulation.) Continued, proliferative stimulation by IGF-1 may predispose to the acquisition of additional genetic abnormalities with evolution to frank leukemia. Thus, IGF-1 could be thought of as a promoter. This hypothesis echoes Greaves' hypothesis relating to leukemia in older children,⁵⁴ which suggests that preleukemic clones acquire additional abnormalities due to proliferative stress applied to lymphocytes by delayed infection in early childhood. Protracted proliferative stimulation of preleukemic clones by hematopoietic growth factors has been implicated in the development of frank leukemia in both preclinical and clinical settings. This phenomenon has been examined most intensively with GM-CSF. Injection of immortalized, but non-leukemic (i.e., preleukemic) myeloid cells

Figure 1. Potential relationships among high birthweight, insulin-like growth factor-1, and infant leukemia.



into GM-CSF transgenic mice, which overexpress GM-CSF 40-fold, leads to death from a leukemic-like illness in two to three months; however, non-transgenic littermates develop no disease.⁵⁵ In the clinic, it has been demonstrated that leukemic myeloblasts from patients with acute myeloid leukemia can be effectively recruited *in vivo* into the S-phase of the cell cycle by the exogenous administration of GM-CSF.⁵⁶ In addition, an increase in the peripheral-blood leukemic-blast count has been observed in some patients receiving GM-CSF for myeloid-lineage leukemias or preleukemic disorders.⁵⁷⁻⁵⁹ IGF-1 may function in a similar role in infants with leukemia.

In addition to growth stimulation, IGF-1 also can function to protect hematopoietic cells from apoptosis.⁶⁰ Apoptosis, or programmed cell death, plays a central role in the normal mechanisms for the physiologic elimination of lymphocytes during lymphopoiesis, as well as in the elimination of lymphocytes that have incurred DNA damage from mutagens or irradiation.⁶¹⁻⁶² Induction of apoptosis is also an important way in which drugs used to treat leukemia (such as steroids and DNA topoisomerase II inhibitors) kill cells.⁶³ Therefore, if IGF-1 is involved in the pathogenesis of infant leukemia, it also may contribute to the poor results achieved with chemotherapy treatment of these children.⁶⁴⁻⁶⁵

An alternative hypothesis for the excess leukemia in high birthweight babies is that the presence of preleukemic cells secreting IGF-1 leads to increased birthweight in the infant, although there are less data to support this theory. In some leukemia cases studied, IGF-1 has been shown to function as an autocrine growth factor, being produced by the leukemia cells and, in turn, stimulating the growth of those same cells.⁴⁰ The additional production of IGF-1 *in utero* produced by an autocrine loop, together with endogenous IGF-1, could have the effect of both increasing birthweight and stimulating proliferation of the preleukemic cells, increasing the opportunity for the acquisition of the additional genetic mutations for the evolution into frank leukemia.

Future investigations

Approximately 11 percent of babies weigh more than 4,000 grams at birth.⁶⁶ Using a relative risk estimate of 2.0 from the more recent epidemiologic studies,¹⁵⁻¹⁶the attributable risk of infant leukemia due to high birthweight could approach 10 percent. Technology is now available to begin to understand the biology behind the epidemiologic data. If high levels of IGF-1 produce a big baby and contribute to leukemia, we would expect to find both IGF-1 and IGF-1 receptor mRNA in the leukemic cells of the infant (as evidence of an autocrine loop), in addition to high serum levels of IGF-1. Epidemiologic and molecular biologic investigations should be able to address these important questions.

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References

- 1. MacMahon B, Newill VA. Birth characteristics of children dying of malignant neoplasms. JNCI 1962; 28: 231-44.
- Iversen T. Leukaemia in infancy and childhood. A material of 570 Danish cases. Acta Paediatr Scand [Suppl] 1966; 167: 25-50.
- 3. Jackson EW, Norris FD, Klauber MR. Childhood leukemia in California-born twins. *Cancer* 1968; 23: 913-9.
- 4. Fasal E, Jackson EW, Klauber MR. Birth characteristics and leukemia in childhood. JNCI 1971; 47: 501-9.
- 5. Wertelecki W, Mantel N. Increased birthweight in leukemia. *Pediatr Res* 1973; 7: 132-8.
- Hirayma T. Descriptive and analytical epidemiology of childhood malignancy in Japan. In: Kobayashi N, ed. Recent Advances in Management of Children With Cancer. Tokyo, Japan: The Children's Cancer Association of Japan, 1980: 27-43.
- Daling JR, Starzyk P, Olshan AF, Weiss NS. Birthweight and the incidence of childhood cancer. JNCI 1984; 72: 1039-41.
- 8. Shaw G, Lavey R, Jackson R, Austin D. Association of childhood leukemia with maternal age, birth origin, and paternal occupation. Am J Epidemiol 1984; 119: 788-95.
- 9. Eisenberg DE, Sorahan T. Birthweight and childhood cancer deaths. *JNCI* 1987; **78**: 1095-100.
- Shu XO, Gao YT, Brinton LA, et al. A population-based case-control study of childhood leukemia in Shanghai. Cancer 1988; 62: 635-44.
- 11. Kaye SA, Robison LL, Smithson WA, Gunderson P, King FL, Neglia JP. Maternal reproductive history and birth characteristics in childhood acute lymphoblastic leukemia. *Cancer* 1991; 68: 1351-5.
- Savitz DA, Ananth CV. Birth characteristics of childhood cancer cases, controls, and their siblings. *Ped Hematol Oncol* 1994; 11: 587-99.
- Cnattingius S, Zack MM, Ekbom A, et al. Prenatal and neonatal risk factors for childhood lymphatic leukemia. JNCI 1995; 87: 908-14.
- Cnattingius S, Zack MM, Ekbom A, et al. Prenatal and neonatal risk factors for childhood myeloid leukemia. Cancer Epidemiol Biom Prev 1995; 4: 441-5.
- Ross JA, Potter JD, Shu XO, Reaman GH, Lampkin B, Robison LL. Evaluating the relationships among maternal reproductive history, birth weight, and risk of infant leukemia. A report from the Childrens Cancer Group. Ann Epidemiol (in press).
- Yeazel MW, Ross JA, Buckley JD, Woods WG, Ruccione K, Robison LL. High birthweight and risk of specific childhood cancers: a report from the Children's Cancer Group. (Submitted).
- Gurney JG, Severson RK, Davis S, Robison LL. Incidence of cancer in children in the United States. Sex-, race-, and 1-year age-specific rates by histologic type. *Cancer* 1995; 75: 2186-95.
- Ross JA, Davies SM, Potter JD, Robison LL. The epidemiology of childhood leukemia with a focus on infants. *Epidemiologic Rev* 1994; 16: 243-72.

- Leisenring WM, Breslow NE, Evans IE, Beckwith JB, Coppes MJ, Grundy P. Increased birthweights of National Wilms' Tumor Study patients suggest a growth factor excess. *Cancer Res* 1994; 54: 4680-3.
- Weksberg R, Shen DR, Fei YL, Song QL, Squire J. Disruption of insulin-like growth factor 2 imprinting in Beckwith-Wiedemann Syndrome. Nat Genet 1993; 5: 143-50.
- Ogawa O, Eccles MR, Szeto J, et al. Relaxation of insulinlike growth factor II gene imprinting implicated in Wilms' tumour. Nature 1993; 362: 749-51.
- Olshan AF. Wilms' tumor, overgrowth, and fetal growth factors. A hypothesis. Cancer Genet Cytogen 1986; 21: 303-7.
- 23. Insulin-like growth factors. In: Pimental E, ed. Handbook of Growth Factors. Vol II. Peptide Growth Factors. Boca Raton, FL (USA): CRC Press, Inc., 1994: 55-95.
- Baxter RC. Insulin-like growth factor binding proteins in the human circulation: a review. Horm Res 1994; 42: 140-4.
- Bennett A, Wilson DM, Liu F, Nagashima R, Rosenfeld RG, Hintz RL. Levels of insulin-like growth factors I and II in human cord blood. J Clin Endocinol Metab 1983; 57: 609-12.
- Gluckman PD, Johnson-Barrett JJ, Butler JH, Edgar BW, Gunn TR. Studies of insulin-like growth factor-I and -II by specific radioligand assays in umbilical cord blood. *Clin Endocrin* 1983; 19: 405-13.
- 27. Ashton IK, Zapf J, Einschenk I, MacKenzie IZ. Insulin-like growth factors (IGF) 1 and 2 in human foetal plasma and relationship to gestational age and foetal size during midpregnancy. *Acta Endocrinologica* 1985; 110: 558-63.
- Samaan NA, Schultz PN, Johnston DA, Creasy RW, Gonik B. Growth hormone, somatomedin C, and nonsuppressible insulin-like activity levels compared in premature, small, average birth weight, and large infants. *Am J Obstet Gynecol* 1987; B;157: 1524-8.
- 29. Lasarre C, Hardouin S, Daffos F, Forestier F, Frankenne F, Binoux M. Serum insulin-like growth factors and insulinlike growth factor binding proteins in the human fetus. Relationships with growth in normal subjects with intrauterine growth retardation. *Ped Res* 1991; 29: 219-25.
- Salardi S, Orsini LF, Cacciari F, Righetti F, Donati S, Mandini M. Growth hormone, insulin-like growth factor I, insulin and C-peptide during human fetal life: in-utero study. *Clin Endocrinol* 1991; 34: 187-90.
- Wang HS, Lim J, English J, Irvine L, Chard T. The concentration of insulin-like growth factor-I and insulin-like growth factor-binding protein-1 in human umbilical cord serum at delivery: relation to fetal weight. J Endrocrinol 1991; 129: 459-64.
- 32. Delmis J, Drazancic A, Ivanisevic M, Suchanek E. Glucose, insulin, HGH, and IGF-1 levels in maternal serum, amniotic fluid and umbilical venous serum: a comparison between late normal pregnancy and pregnancies complicated with diabetes and fetal growth retardation. *J Perinat Med* 1992; 20: 47-56.
- Kubota T, Kamada S, Taguchi M, Aso T. Determination of insulin-like growth factor-2 in feto-maternal circulation during human pregnancy. Acta Endocrinologica 1992; 127: 359-65.
- Fant M, Salafia C, Baxter RC, et al. Circulating levels of IGFs and IGF binding proteins in human cord serum. Relationships to intrauterine growth. *Regulatory Peptides* 1993; 48: 29-39.
- 35. Verhaeghe J, Van Bree R, Van Herck EV, Laureys J, Bouillon R, Van Assch FA. C-peptide, insulin-like growth factors I and II, and insulin-like growth factor binding protein-1 in

umbilical cord serum: correlations with birth weight. Am J Obstet Gynecol 1993; 169: 89-97.

- 36. Reece EA, Wiznitzer A, Le E, Homko CJ, Behram H, Spencer EM. The relation between human fetal growth and fetal blood levels of insulin-like growth factors I and II, their binding proteins, and receptors. *Obstet Gynecol* 1994; 84: 88-95.
- 37. Simmons D. Interrelation between umbilical cord serum sex hormones, sex hormone-binding globulin, insulin-like growth factor 1, and insulin in neonates from normal pregnancies and pregnancies complicated by diabetes. J Clin Endocrinol Metab 1995; 80: 2217-21.
- Baker J, Liu JP, Robertson EJ, Efstratiadis A. Role of insulin-like growth factors in embryonic and postnatal growth. *Cell* 1993; 75: 73-82.
- Perentesis JP, Kersey JH. Biologic therapy in leukemia. In: Henderson ES, Lister TA, Greaves MF, eds. *Leukemia, 6th Ed.* London, UK: WB Saunders, 1996.
- Baier TG, Jenne EW, Blum W, Shonberg D, Hartmann KK. Influence of antibodies against IGF-1, insulin or their receptors on proliferation of human acute lymphoblastic leukemia cell lines. *Leuk Res* 1993; 16: 807-14.
- Sanders M, Sorba S, Daniak N. Insulin-like growth factors stimulate erythropoiesis in serum-substituted umbilical cord blood cultures. *Exper Hematol* 1993; 21: 25-30.
- Jardieu P, Clark R, Mortensen D, Dorshkind K. In vivo administration of insulin-like growth factor-1 stimulates primary B lymphopoiesis and enhances lymphocyte recovery after bone marrow transplantation. J Immunol 1994; 152: 4320-7.
- 43. Estrov Z, Meir R, Barak Y, Zaizov R, Zadik Z. Human growth hormone and IGF-1 enhances the proliferation of leukemic blasts. J Clin Oncol 1991; 9: 394-9.
- 44. Shimon I, Shpilberg O. The insulin-like growth factor system in regulation of normal and malignant hematopoiesis. *Leuk Res* 1995; 19: 233-40.
- Knudson AG, Meadows AT, Nichols WW, Hill R. Chromosomal deletion and retinoblastoma. N Engl J Med 1976; 295: 1120-3.
- Volgelstein V, Fearon ER, Hamilton SR, et al. Genetic alterations during colorectal-tumor development. N Engl J Med 1988; 319: 525-32.
- Limpens J, Stad R, Vos C, et al. Lymphoma-associated translocation t(14;18) in blood B cells of normal individuals. Blood 1995; 85: 2528-36.
- Nucifora G, Larson RH, Rowley JD. Persistence of the 8;21 translocation in patients with acute myeloid leukemia type M2 in long term remission. *Blood* 1993; 82: 712-5.
- Morris JA. A mutational theory of leukemogenesis. J Clin Pathol 1989; 42: 337-40.
- Wasserman R, Galili N, Ito Y, Reichard BA, Shane S, Rovera G. Predominance of fetal type DJH joining in young children with B-precursor lymphoblastic leukemia as evidence for an in utero transforming event. J Exper Med 1992; 176: 1577-81.
- 51. Steenbergen EJ, Verhagen OJ, Van Leeuwen EF, et al. B-precursor acute lymphoblastic leukemia third complementarily determining regions predominantly represent an unbiased recombination repertoire: leukemic transformation frequently occurs in fetal life. Euro J Immunol 1994; 24: 900-8.
- 52. Ford AM, Ridge SA, Cabrera ME, et al. In utero rearrangements in the trithorax-related oncogene in infant leukemias. *Nature* 1993; 363: 358-60.

- Mahmoud HH, Ridge SA, Behm, FG, et al. Intrauterine monoclonal origin of neonatal concordant acute lymphoblastic leukemia in monozygotic twins. Med Ped Oncol 1995; 24: 77-81.
- 54. Greaves MJ, Chan LC. Is spontaneous mutation the major 'cause' of childhood acute lymphoblastic leukemia? Br J Hematol 1986; 64: 1-13.
- Metcalf D, Rasko JEJ. Leukemic transformation of immortalized FDC-P1 engrafted in GM-CSF transgenic mice. *Blood* 1993; 7: 878-86.
- 56. Cannistra SA, Groshak P, Griffin JD. Granulocyte-macrophage colony-stimulating factor enhances the cytotoxic effects of cytosine arabinoside in acute myeloblastic leukemia and in the myeloid blast crisis phase of chronic myelogenous leukemia. *Leukemia* 1989; 3: 328-34.
- 57. Buchner T, Hiddemann W, Koenigsmann M, et al. Recombinant human granulocyte-macrophage colony stimulating factor after chemotherapy in patients with acute myeloid leukemia at higher age after relapse. *Blood* 1991; **78**: 1190-7.
- Ganser A, Volkers B, Greher J, et al. Recombinant human granulocyte-macrophage colony-stimulating factor in patients with myelodysplastic syndromes – a phase I/II trial. Blood 1989; 73: 31-7.
- 59. Herrmann F, Lindemann A, Klein H, Lubbert M, Shulz G, Mertelsmann R. Effect of recombinant human granulocytemacrophage colony-stimulating factor in patients with myelodysplastic syndrome with excess blasts. *Leukemia* 1989; **31**: 335-8.

- 60. Williams GT, Smith CA. Molecular regulation of apoptosis. genetic controls on cell death. *Cell* 1993; 74: 777-9.
- 61. Fisher DE. Apoptosis in cancer therapy: crossing the threshold. *Cell* 1994; 78: 539-42.
- 62. Pui CH, Frankel LS, Carroll AJ, et al. Clinical characteristics and treatment outcome of childhood acute lymphoblastic leukemia with the t(4;11)(q21;q23). A collaborative study of 40 cases. *Blood* 1991; 77: 440-7.
- Neiman PE, Thomas SJ, Loring G. Induction of apoptosis during normal and neoplastic B-cell development in the bursa of Fabricus. Proc Natl Acad Sci USA 1991; 88: 5857-61.
- Muta K, Krantz SB. Apoptosis of human erythroid colonyforming cells is decreased by stem cell factor and insulin-like growth factor I as well as erythropoietin. J Cell Physiol 1993; 156: 264-71.
- 65. Reaman G, Zeltzer P, Bleyer WA, et al. Acute lymphoblastic leukemia in infants less than one year of age. A cumulative experience of the Childrens Cancer Study Group. Blood 1985; 3: 1513-21.
- 66. Department of Health and Human Services. Compilations of Data on Natality, Mortality, Marriage, Divorce, and Induced Terminations of Pregnancy. No 1. Hyattsville, MD (USA): DHHS, May 1989; DHHS Publication No. (PHS) 89-1951, Vital and Health Statistics Supplements to the Monthly Vital Statistics Report. Series 24.