# Variations in abscisic acid, indole-3-acetic acid and zeatin riboside concentrations in two Mediterranean shrubs subjected to water stress

M. Lopez-Carbonell<sup>1,\*</sup>, L. Alegre<sup>1</sup>, A. Pastor<sup>1</sup>, E. Prinsen<sup>2</sup> & H. van Onckelen<sup>2</sup>

<sup>1</sup>Department of Plant Biology, Plant Physiology Unit, Faculty of Biology, Barcelona University, Spain; <sup>2</sup>Department of Biology, University of Antwerp (UIA), Wilrijk, Belgium (\* address for correspondence: Departament de Biologia Vegetal, Unitat de Fisiologia Vegetal, Facultat de Biologia, Universitat de Barcelona, Avda, Diagonal, 645. E-08028-Barcelona, Spain. fax: + 34(3) 4112842. e-mail: martal@porthos.bio.ub.es)

Received 1 March 1996; accepted 17 June 1996

Key words: Lavandula stoechas, Rosmarinus officinalis, ABA, IAA, ZR, water stress

# Abstract

Water stress induced an increase in endogenous concentrations of ABA in *Lavandula stoechas* L. plants to 13100 pmol ABA  $g^{-1}$  FW, which may contribute to the maintenance of water relations between the second and the third day of water stress treatment. After the third day, a sharp decrease in ABA levels was observed to 2630 pmol ABA  $g^{-1}$  FW, together with a decrease in water content and water potential and a loss of plant response to water stress. Water deficit did not induce an increase in endogenous ABA concentration, which remained at 514 pmol ABA  $g^{-1}$ FW in Rosmarinus officinalis L., which is more sclerophyllous than L. stoechas. Nevertheless, the relative water content of Rosmarinus officinalis L. after seven days of water stress decreased more than 40% and  $\psi$  reached values of  $-3.2$  MPa. R. officinalis showed lower levels of ABA, but significantly higher levels of IAA and ZR than L. stoechas (4 times and 6 times respectively in well watered-plants). The increase in ABA levels is not a common mechanism in these two Mediterranean shrubs which survive under water stress conditions.

Abbreviations: ABA - abscisic acid;  $d - days$  of water stress treatment; DW - dry weight; FW - fresh weight; IAA - indole-3-acetic acid; RP - Reversed Phase; RWC - relative water content; TW - turgid weight; WC - water content; ZR – zeatin riboside;  $\psi$ , water potential

# 1. Introduction

Lavender (Lavandula stoechas L.) and rosemary (Rosmarinus ofhcinalis L.) are two native Mediterranean evergreen half-shrubs well adapted to survive under water stress conditions by mechanisms of drought tolerance or avoidance. The semi-sclerophyllous to sclerophyllous leaf consistency is a common characteristic of plants growing in Mediterranean climate and is important in avoiding dehydration [7]. Nevertheless, few biochemical studies in xeromorphic leaves have been reported; the high level of secondary metabolites hinders the purification of plant extracts, so more research effort is needed in this field [13]. The knowledge of variations in phytohormone concentrations in Mediterranean shrubs adapted to recurrent drying cycles will provide valuable information about the physiological responses of those plants to water stress.

The functional role of the plant hormone abscisic acid (ABA) in the adaptation of a variety of plants to water stress has been well documented and reviewed [21, 2, 10]. Variations in ABA levels induced by stress do not completely account for the physiological responses of the plant to drought, and an interaction with other chemical signals such as auxins (IAA) and cytokinins requires careful study [ 1 I]. Abiotic stresses alter the levels of plant hormones in general [14] and these variations could act as modulators of plant responses to water stress. Furthermore, each hormone can influence the levels of at least one of the other types of hormone [6] and it is suggested that the variations in endogenous concentrations of different plant growth regulators may modulate plant water balance [3]. Nevertheless, there is scarce and conflicting evidence on the variations of indole-3-acetic acid levels in waterstressed plants [18, 8, 91. A decrease in the cytokinin content of droughted roots and their possible function as root-to-shoot signals have been described [2, 11. However, indole-3-acetic acid and cytokinins do not necessarily decline during the adaptation of plants to water stress [22]. At present, there are still few studies using the same crude plant extract for phytohormone determination in plants subjected to water stress. Nevertheless, such studies may clarify the function of hormones as modulators of plant responses to water stress.

L. stoechas and R. officinalis differ in leaf morphology:  $L$ . stoechas is less xeromorphic than  $R$ . officinalis [ 151, and therefore these species could present a different behaviour to water stress. The aim of this work was to study the endogenous concentrations of abscisic acid, indole-3-acetic acid and zeatin riboside in these two shrubs subjected to water stress.

### 2. Materials and methods

### 2.1 Plant material and growth conditions

Two-year-old plants of R. officinalis and L. stoechas obtained from seeds were cultivated in controlled conditions (12 h photoperiod, at a photon flux of 400  $\mu$ mol m<sup>-2</sup> s<sup>-1</sup>, air temperature 25/20 °C day/night and 49% relative humidity). Plants were grown in 17 cm diameter plastic pots filled with a calcic Luvisol (FAO). Plants were watered daily to container capacity: well watered plants. The stress treatment was imposed by lack of watering. Samples were taken early in the morning when  $\psi$  was  $-2.6$  MPa for stressed plants and experiments were continued until stressed plants reached values of  $-3.2$  MPa; this meant witholding water for two, three, and four days in L. stoechas and for three, four, five and six days in R. officinalis. For the studies of water relations and endogenous ABA, IAA and ZR, homogenous apical non-woody shoots ten cm long were selected. The values are means of three samples (collected from separate experiments)  $\pm$ SE.

### 2.2 Water relations

Leaf relative water content (RWC) was calculated as:

$$
RWC = (FW - DW/TW - DW) \times 100,
$$

where  $FW = fresh$  weight in the field;  $DW = Dry$ weight, after drying samples in an oven at 80 $\degree$ C to constant weight;  $TW = \text{target}$  weight, after rehydrating fresh shoots for 24 h.

Water content of shoots (g  $g^{-1}$  DW) was estimated from the fresh and dry weight.

Leaf water potential  $(\psi)$  was obtained from 10 cm long shoots using a pressure chamber (ARIMAD-2) containing a wet filter paper at the bottom of the chamber.

### 2.3 Analysis of endogenous ABA, IAA and ZR

After determination of fresh weight, leaves were immediately frozen in liquid nitrogen and stored at  $-20$  °C until hormone analysis. IAA, ABA and ZR were purified from the same plant extract, as described by Prinsen et al. (1995). After grinding in liquid nitrogen in a mortar and pestle, leaf samples were extracted overnight at  $-20$  °C in 80% MeOH (9 ml  $g^{-1}$  FW), to which DL-cis, trans-[G- $^{3}$ H]-ABA (300 Bq, 2.26 TBq/mmol, Amersham),  $[^{2}H_{6}]$ -ABA (200 ng, for preparation see Milborrow, 1971), (3-[5(n)-  ${}^{3}\text{H}$ ]-indole-acetic acid (300Bq, 788GBq/mmol, Amersham), indole- $[2,4,5,6,7^{-2}H_5]$ -3-acetic acid (200 ng, Aldrich) and  $[3H]$ -trans-zeatin-riboside dialcohol (200 Bq, 1.4TBq mmol<sup>-1</sup>, Amersham) were initially added as tracers for localization  $({}^{3}H)$  and isotope dilution  $(^{2}H)$  purposes. After centrifugation (24,000 g, 15 min,  $4 °C$ ) the extract was purified on a Reversed Phase  $(RP)-C_{18}$  cartridge (500 mg, Varian). The effluent was evaporated in vacuo and the sample was redissolved in 15 ml 0.04 M ammonium acetate buffer pH 6.5. The samples were purified using a DEAE-Sephadex A25 (4 ml) cartridge to which a RP- $C_{18}$  (Varian) cartridge was coupled underneath.

Cytokinins were eluted from the  $RP-C_{18}$  cartridge with 4 ml 80% methanol, evaporated in a speed-vac (VR1 HETO Lab. Eq.), redissolved in 500  $\mu$ l PBS and analyzed by RIA [17] using zeatin-riboside specific polyclonal chicken egg yolk antibodies. Standards and samples were tested in triplicate. The results were expressed as pmol cytokinin-riboside equivalents per gram fresh weight (pmol ZR-equiv.  $g^{-1}$  FW).

ABA and IAA were eluted with 6% formic acid from DEAE-Sephadex A25 and concentrated on a RP

 $C_{18}$  (Varian) cartridge with 5 ml diethyl ether and evaporated in vacuo. A preparative ion supression RP-HPLC (50/49.5/0.5,  $v/v/v$ , methanol/H<sub>2</sub>O/acetic acid; Alltech Econospere C<sub>18</sub>, 100  $\times$  4.6 mm, 3 $\mu$ m; 0.5 ml/min) was coupled to a fluorimetric detector (Schimadzu RF530,  $\overline{\lambda}_{ex}$  285 nm,  $\lambda_{em}$  360 nm). 1 minute fractions were collected during the time interval corresponding to the retention times of the radioactivity of a  $^{14}$ C-IAA (3-indolyl-(1-<sup>14</sup>C)-acetic acid, 100 Bq, 2.18 GBq/mmol, Amersham)-and 3H-ABA (133 Bq) reference respectively. l/10 aliquot of the collected fractions were counted for 14C and 3H of the internal tracer (Packard TRICARB 1500 Liquid Scintillation Analyser).

The ABA samples were methylated with diazomethane [20]; Me-ABA was analyzed by GC-MS separative chemical ionization in selective ion mode (CI-SIM) HP 5890 Series II, coupled to a VG TRIO 2000 quadrupole mass spectrometer (column 25 m, 0.25 mm ID; gas phase He, temperature gradient from 120-240 °C, 15°/min, 250 °C injection temperature, CI gas amonium; retention time of Me-ABA 5.59 min [19]). For the detection of Me-ABA and Me- $^{2}H_{6}$ -ABA, 278 and 284 were used as selective diagnostic ions, respectively [5]. The integrated peak areas corresponding to these diagnostic ions were used for calculations. After methylation, the IAA containing fractions were further derived to IAA-Me-HFB [16]. IAA-Me-HFB was analyzed by GC.MS positive electron impact in selective ion mode  $(EI<sup>+</sup>-SIM)$  HP 5890 Series II, coupled to a VG TRIO 2000 quadrupole mass spectrometer (column 25 m, 0.25 mm ID; gas phase He, temperature gradient from 0-6 min:  $25^{\circ}$ C/min, 6-8 min 50 °C/min; 8-10 min 300 °C, 250 °C injection temperature; retention time IAA-Me-HFB 8.9 min). For the detection of IAA-Me-HFB and  ${}^{2}H_{5}$ -IAA-Me-HFB, 326 and 329 were used as selective diagnostic ions, respectively.

The results were expressed as pmol ABA  $g^{-1}$  FW or pmol IAA  $g^{-1}$  FW and were analyzed using a paired "t" test.

# 3. Results

### 3.1 Water relations

Water status in L. stoechas shoots is shown in Figure 1. After two days of water stress a decline of 30% in RWC and of 45% in water content (WC) was observed.  $\psi$  decreased from  $-0.6$  to  $-2.6$  MPa. There were prac-



Figure 1. Effect of water stress on (a) relative water content (RWC), (b) water content (WC) and (c) water potential  $(\psi)$  of Lavandula stoechas shoots. The vertical bars represent  $\pm$  standard error (n = 3).

tically no differences in plant water relations between two and three days of water stress. After four days of water deficit, RWC and WC declined again: RWC became 60% lower than in well watered plants, WC decreased to 0.94 g  $g^{-1}$  DW and  $\psi$  fell to -3.2 MPa. Thereafter, the experiments were discontinued, since plants appeared completely wilted.

Figure 2 show water relations in *, <i>officinalis*. The RWC (82.6%) of rosemary well watered plants was lower than that of lavender well watered plants (89%). After three days of water stress a decrease of 25% in RWC and of 45% in WC was seen in rosemary; the  $\psi$  decreased from  $-1.3$  MPa to  $-2.75$  MPa and afterwards a further slight decline in plant water content and  $\psi$  was seen. After six days of water stress, no signs of wilting were observed in rosemary plants.

### 3.2 ABA, IAA and ZR concentrations

The ABA concentration in L. stoechas leaves after three days of water stress (13100  $\pm$  400 pmol ABA  $g^{-1}$  FW) was significantly higher than those of well watered plants (900  $\pm$  700 pmol ABA g<sup>-1</sup> FW). Afterwards, the ABA content decreased to a similar value to that of the well watered plants (Figure 3a).

L. stoechas showed no significant differences in endogenous ZR concentration throughout water stress treatment (Figure 3a). A slight increase was observed after 2 days, followed by a decrease in the ZR content.

The free IAA concentration in  $L$ . stoechas increased over the first two days of water stress, reaching a peak of 189  $\pm$  6 pmol IAA g<sup>-1</sup> FW by the second day; then, it decreased to 63  $\pm$  16 pmol IAA g<sup>-1</sup> FW at three days and rose again on the fourth day of water stress (Figure 3b).

The ABA concentration in  $R$ . officinalis leaves (Figure 4b) from well watered plants was lower than that in leaves of L. stoechas. There were no significant differences in endogenous ABA levels throughout the treatment.

The ZR content of R. officinalis leaves (Figure 4a) increased during the three days of treatment (29  $\pm$  6 nmol ZR equiv.  $g^{-1}$  FW) and then slowly decreased during the remaining experimental period (reaching values of 10.8  $\pm$  2.5 nmol ZR equiv.  $g^{-1}$  FW). The ZR concentration observed in  $R$ . *officinalis* was always higher than that of L. stoechas.

The free IAA concentration in *, <i>officinalis* was significantly higher than IAA concentration in L. stoe*chas*, reaching a peak (1740  $\pm$  260 pmol IAA g<sup>-1</sup> FW) after three days of water stress. Then, it showed a gradual decline during the course of the experiment to  $300 \pm 100$  pmol IAA g<sup>-1</sup> FW by the last day. There were no significant differences between well watered plants and plants subjected to four, five or six days of water stress (Figure 4b).

During the water stress treatment the ratio ZR/ABA decreased from 3.1 in well watered plants to 0.5 after 3



Figure 2. Effect of water stress on (a) relative water content (RWC), (b) water content (WC) and (c) water potential  $(\psi)$  of Rosmarinus *officinalis* shooots. The vertical bars represent  $\pm$  standard error (n = 3).

days. On the fourth day a significant decrease in ABA concentration and a tendency to decrease in ZR levels were observed.



Figure 3. Effect of water stress on endogenous concentration of (a) ZR equiv. and ABA, (b) IAA, in *Lavandula stoechas* leaves.  $(\triangle)$ ZR, ( $\bullet$ ) ABA, ( $\circ$ ) IAA. The vertical bars represent  $\pm$  standard error  $(n = 3)$ .

# 4. Discussion

In both species WC declined more rapidly than RWC indicating that adaptation to water stress was taking place.

The increase observed in the ABA level in L. stoechas after 2 days may explain the lack of differences in WC, RWC and  $\psi$  between 2 and 3 days of water stress treatment. After 3 days, the sharp decline in ABA could be related to a loss of the plant capacity to respond to water stress [8, 9].

 $R.$  officinalis is more sclerophyllous than  $L.$  stoechas and transpiration rates are always lower than those of L. stoechas. Although  $\psi$  reached  $-3.2$  MPa, no signs of stress were observed in these  $R$ . *officinalis* plants;



Figure 4. Effect of water stress on endogenous concentration of (a) ZR equiv., (b) ABA and IAA, in Rosmarinus officinalis leaves. ( $\triangle$ ) ZR, ( $\bullet$ ) ABA, ( $\circ$ ) IAA. The vertical bars represent  $\pm$  standard

error  $(n - 3)$ .

after an initial decrease in water content and  $\psi$ , practically no differences in these parameters from 3 to 6 d were observed. Leaf xeromorphy enables plants to reduce water loss and maintain RWC.

In L. stoechas the increase in ABA content appeared to be correlated with drought resistance in plants and a sharp decline in ABA levels observed at severe water stress conditions could be related with the loss of capacity of plant to withdraw stress. In well watered plants the bulk leaf ABA concentration was about 16 times higher than that of IAA. After 2 days the endogenous IAA rose approximately three fold with respect to those of well watered plants. Similar results have been reported for aralia plants subjected to stress [9]. It was hypothesized [14] that, under stress conditions which impair plant growth, the cytokinin activity decreases, while the ABA acitivity increases. Nevertheless, no significant differences in zeatin riboside levels were recognized in our study. Our results agree with those of Zholkevich and Pustovoitova (1993), who found that IAA and cytokinins do not necessarily decline during the adaptation to water stress.

 $R.$  officinalis showed no significant changes in ABA content, which could be related to the lack of differences observed in water relations between 3 and 6 d. In comparison to L. stoechas, the endogenous ABA concentration in  $R$ . *officinalis* well watered plants was 3 times lower. ABA endogenous concentration of water stressed  $R$ . *officinalis* plants was found to be up to 30 times lower than those observed in L. stoechas water stressed plants. Levitt (1980) stated that drought avoidance is largely morphological-anatomical in nature, thus it could explain the lack of the common response to water stress such as an increase in the endogenous ABA concentration. Nevertheless, the levels of IAA and ZR were significantly higher than those observed in  $L$ . stoechas. In well watered plants of  $R$ . officinalis endogenous ZR concentration was 6 fold higher than in L. stoechas and it remained high throughout the water stress treatment. Similar behaviour was observed for endogenous concentrations of IAA. Hormonal interactions may affect hormonal levels in plant tissues. High levels of cytokinins increase the levels of IAA and reduce the levels of ABA [6]. The ability of one hormone to affect the level of another is probably of great significance in the regulation of plant growth and plant responses to water stress, and merits more critical assessment. The complexity of these interactions introduces a new dimension to the mechanism of hormone action. The high concentrations of ZR and IAA observed in R. officinalis could be related with the delay in leaf senescence. Meanwhile, higher levels of ABA observed in L. stoechas contribute to the acceleration of leaf senescence [4].

The increase in endogenous ABA concentration in response to water stress may not be a general mechanism in these two autochthonous Mediterranean shrubs adapted to water stress avoidance. Anatomical and ultrastructural characteristics, and also hormonal interactions, should be taken into account.

# Acknowledgements

Part of this work has been supported by the Research Project of CICYT PB92-815, Spain. H.V.O. is a Research Director and E.P. is a Research Associate of the Belgian National Fund for Scientific Research. We are grateful to Dr. R. Rycroft for the correction of the English manuscript.

# References

- 1. Bano A, Dorffling K, Bettin D and Hahn H (1993) Abscis acid and cytokinins as possible root-to-shoot signals in xylem sap of rice plants in drying soil. Australian Journal of Plant Physiology 20: 109-115
- 2. Davies WJ and Jones HG (1991) Abscisic acid. Oxford: Bio Scientific Publishers
- 3. Davies WJ and Zhang J (1991) Root signals and the regulation of growth and development of plants in drying soil. Annual Review of Plant Physiology and Plant Molecular Biology 42: 55-76
- 4. Goldthwaite JJ (1987) Hormones in plant senescence. In: Davies PJ (ed) Plant Hormones and Their Role in Plant Growth and Development, pp 553-573. Martinus Nijhoff Publishers
- 5. Gray RT, Mallaby R, Ryback G and Willians VP (1974) Mas spectra of methyl abscisate and isotopically labelled analogues. J. C. S. Perkin II: 919-924
- 6. Letham DS, Higgins TJV, Goodwin PB and Jacobsen JV ( 1978) Phytohormones in retrospect. In: Letham DS, Goodwin PB and Higgins TJV (eds) Phytohormones and Related Compounds: A Comprehensive Treatise, pp l-27. North-Holland: Elsevier
- 7. Levitt L (1980) Water, radiation, salt and other stresses. In: Kowslowsky TT (ed) Response of Plants to Environmental Stresses, pp 93-128. New York: Academic Press
- 8. López-Carbonell M, Alegre L and Van Onckelen H (1994a) Effects of water stress on cellular ultrastructure and on concentrations of endogenous abscisic acid and indole-3-acetic acid in Farsia japonica leaves. Plant Growth Regulation 14: 29-35
- 9. López-Carbonell M, Alegre L and Van Onckelen H (1994) Changes in cell ultrastructure and endogenous abscisic acid and indole-3-acetic acid concentrations in Fatsia japonica leaves under polyethylene glycol-induced water stress. Plant Growth Regulation 15: 165-174
- 10. Mansfield TA and McAinsh MR ( 1995) Hormones as regulators of water balance. In: Davies J (ed) Plant Hormones. 2nd Edition, pp 598-616. Kluwer Academic Publisher
- 11. Masia A, Pitacco A, Braggio Land Giulivo C (1994) Hormonal responses to partial drying of the root system of Helianfhus annuus. Journal of Experimental Botany 45: 69-76.
- 12. Milborrow BW (1971) Abscisic acid. In: Goodwin TW (ed) Aspects of Terpenoid Chemistry and Biochemistry, pp 137- 15 1. London: Academic Press
- 13. Mooney HA (1985) The impact of environmental stress on plant performance in mediterranean-climate ecosystems: Differing levels of analysis. In: Tenhunen JD, Catarino FM, Lange OL and Oechel WC (eds) Plant Response to Stress. Nato. Asi. Series. Serie G. Ecological Science, pp 661-668. London, Paris, Tokyo: Springer-Verlag
- 14. Morgan PW (1990) Effects of abiotic stresses on plant hormone systems. In: Alscher RC and Cumming JR (eds) Stress Responses in Plants: Adaptation and Acclimation Mechanisms, pp 113-146. New York: Wiley-Liss, Inc
- 15. Oshran G (1989) Plant Pheno-Morphological Studies in Mediterranean Type Ecosystems. Dordrecht, Kluwer Academic Publisher, pp 57-83.
- 16. Pilet PE and Saugy M (1985) Effect of applied and endogenous indole-3-acetic acid on maize root growth. Planta 164: 254-258
- 17. Prinsen E, Redig P, Stmad M, Galis I, Van Dongen W. and Van Gnckelen H (1995) Quantifying Phytohormones in Transformed Plants. In: Gartland KMA and Duvey MR (eds) Methods in Molecular Biology, vol. 44, pp 245-262. Totowa, NJ: Agrobacterium Protocols. Humana Press Inc
- 18. Rimbaut JM and Pilet PE (1994) Water stress and indole-3 acetic acid content of maize roots. Planta 193: 502-507
- 19. Rivier L, Milon H and Pilet PE (1977) Gas Chromatographymass spectrometric determinations of abscisic acid levels in the cap and the apex of maize roots. Planta 134: 23-27
- 20. Schlenk H and Gellerman JL (1960) Esterification of fatty acids with diazomethane on a small scale. Anal. Chem. 32: 1412-1414
- 21. Zeevaart JAD and Creelman RA ( 1988) Metabolism and physiology of abscisic acid. Annual Review of Plant Physiology and Plant Molecular Biology 39: 439-473
- 22. Zholkevich VN and Pustovoitova TN (1993) Growth and phytohormone content in Cucumis sativus L. leaves under water deficiency. Russian Journal of Plant Physiology 40: 585- 589