Update section

Sequence

Nucleotide sequence of an osmotin cDNA from the *Nicotiana tabacum* cv. White Burley generated by the polymerase chain reaction

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Salt stress-induced changes in gene expression have been observed in a number of plant species, in both whole plants and cultured cells. In tobacco and tomato salt adaptation is associated with the accumulation of a 26 kDa protein, which has been termed osmotin in tobacco [1, 2, 3]. Highly homologous cDNA clones encoding osmotin or an osmotin-like protein have been isolated and sequenced from libraries constructed from the roots of salt-adapted tomato (*Lycopersicon esculentum* VFNT) plants, and from saltadapted suspension cultures of tobacco (*Nicotiana tabacum* Wisconsin-38) [3, 4].

The present communication describes the nucleotide sequence of a PCR-amplified osmotin cDNA from salt-adapted suspension cultures of the tobacco cultivar White Burley, using primers derived from published sequence from the Wisconsin-38 cultivar [4]. The cDNA was cloned in both orientations into the phagemid vector pTZ (Pharmacia) and the sequence determined by the dideoxynucleotide chain termination method [5], each position being determined on average three times. The coding region is 738 bp, 6 bp longer

than that of Wisconsin-38, and the predicted molecular mass of the encoded preprotein is 26697 Da. Assuming a signal processing site identical to the Wisconsin-38 osmotin [6], the molecular mass of the mature protein would be 24283 Da.

The main differences between the White Burley sequence, shown in Fig. 1, and the Wisconsin-38 sequence [4] can be summarised as follows: (a) nucleotide substitutions occur at 11 positions, 4 of them resulting in no change in amino acid. The remaining substitutions result in 5 changes in amino acid sequence, 3 of them being conservative changes; (b) there are 7 nucleotide insertions and 1 deletion, resulting in 2 additional amino acids; (c) there is a region of considerable variation from nucleotide positions 446–465 where 2 nucleotide substitutions followed by an insertion and a compensatory deletion result in changes in 7 consecutive amino acids.

Although the physiological function of osmotin is unknown, we are currently investigating its role in salt adaptation by introducing the gene into a different cellular environment.

The nucleotide sequence data reported will appear in the EMBL, GenBank and DDBJ Nucleotide Sequence Databases under the accession number X61679.

G NLRSSFVFFLLALVTYT ATGGGCAACTTGAGATCTTCTTTTGTTTTCTTCCTCCTTGCCTTGGTGACTTATACTTAT 10 20 30 40 50 60 40 R CP T V W I G G G R R L D R G Q T W V I N A P R G ATAGGCGGTGGCCGGCGTCTCCGATCGAGGCCAAACTTGGGTGATCAATGCGCCACGAGGT 130 140 150 160 170 180 T N M A R V W G R T N C N F N A A G K G ACTAATATGGCACGTGTATGGGGCCGTACTAATTGTAACTTCAATGCTGCTGGTAGGGGGT 190 200 210 220 230 240 от б DCG G V L C с т 0 G w G ACGTGCCAAACCGGTGACTGTGGTGGAGTCCTACAGTGCACCGGGTGGGGTAAACCACCA 250 260 270 280 290 300 N T L A E Y A L D Q F S G L D F W D I S AACACCTTGGCTGAATACGCTTTGGACCAATTCAGTGGTTAGATTTCTGGGACATTTCT 310 320 330 340 350 $\begin{array}{c|c} C & H & I \\ \hline \\ TGCCATGCAATCCATTGTACGGCTAATATACGCGAATGTCCCCGCGAACTTAGGGTTCCCC \\ 430 & 440 & 450 & 460 & 470 & 480 \end{array}$ G G C N N P C T T F G G Q Q Y C C T Q G GGAGGATGTAATAACCCTTGTACTACATTGGAGGAGCAACAATATTGTTGCACACAAGA 500 510 520 530 540 Q D D РТ S TFT C ΡG G s T TACCCACAAGATGATCCTACTAGCACTTTTACTTGCCCTGGTGGTAGTACAAATTATAGG 610 620 630 640 650 660 V I F C P N G Q A H P N F P L E M P G S GTTATCTTTTGTCCTAATGGTCAAGGTCACCCAAATTTTCCCTTGGAAATGCCTGGAAGT 670 680 690 700 710 720 V A Е к GATGAAGTGGCTAAGTAG. 730

Fig. 1. Nucleotide sequence and derived amino acid sequence of an osmotin cDNA clone from tobacco (*Nicotiana tabacum* cv. White Burley). The arrow indicates the likely cleavage site for the removal of the signal sequence. Variations with the Wisconsin-38 sequence are indicated: nucleotide substitutions are underlined once, insertions are underlined twice and the deletion point is indicated by $\hat{}$. Amino acid substitutions are boxed and insertions are circled.

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