# **Adaptation of the thylakoid membranes of pea chloroplasts to light intensities. I. Study on the distribution of chlorophyllprotein complexes**

#### TA-YAN LEONG and JAN M. ANDERSON

CSIRO, Division of Plant Industry, GPO Box 1600, Canberra ACT 2601, Australia

*(Received." 17August 1983; in revised form: 8 December 1983)* 

Key words: chl a/chl b ratios, chlorophyll-proteins, light adaptation, photosynthesis, thylakoid membranes

Abstract. The effect of light intensity (16 h white light and 8 h dark) during growth of pea plants at 20 °C on the chlorophyll composition and on the relative distribution of chlorophyll amongst the various chlorophyll-protein of pea thylakoids was studied. The chl a/chl b ratios increased from 2.1 to 3.2 as light intensity during growth varied from 10 to 840 $\mu$ Em<sup>-2</sup> s<sup>-1</sup>. This function can be described by two straight lines intersecting at a transition point of approximately  $200 \mu E m^{-2} s^{-1}$ . Similar discontinuities in the responses were observed in the changes in the relative distribution of chlorophyll amongst the various chlorophyll-protein complexes. This demonstrates that the chl a/chl b ratio of the various thylakoids is a good indicator of changes in the relative distribution of chlorophyll. As the chl a/chl b ratio decreased, the amount of chlorophyll associated with photosystem I complexes decreased, that with photosystem II core reaction centre complex was halved, and that with the main chl a/b-proteins of the light-harvesting complex was markedly increased.

# **Introduction**

It is well established that higher plants adapt to different light intensities during growth in such a way that plants grown under high light intensities have a greater efficiency of photosynthetic quantum conversion than plants grown under low light intensities. This photoadaptation is accompanied by specific changes in composition, structure and function of the chloroplasts [10, 11, 12, 18]. Part of the functional change is associated with measurable changes in the chemical composition of the thylakoid membranes including varying chl a/chl b ratios, varying xanthophylls/ $\beta$ -carotene ratios, changing amounts of electron carriers such as plastoquinone, cytochromes f,  $b_6$  and b-559, and changes in ribulose-l,5-bisphosphate carboxylase concentrations. High light grown plants have higher chl a/chl b ratios, lower xanthophyll/ /3-carrotene ratios, and higher levels of electron carriers per unit chlorophyll [11, 12, 17, 22, 27]. Changes in the pigment composition are well documented but the changes in the relative amounts of chlorophyll-protein complexes in the thylakoid membranes during photoadaptation are less well

Abbreviations:  $ch = chlorophyll$ ,  $PS = photosystem$ ,  $SDS = sodium dodecyl sulplate$ , Tricine = N-tris (hydroxymethyl) methylglycine.

understood [13] mainly because, prior to 1978, only two main chlorophyllproteins were resolved by gel electrophoresis and the amounts of free chlorophyll were relatively high [27].

Recent developments in mild SDS polyacrylamide gel electrophoresis procedures have led to high resolution of cholorophyll-protein complexes with the liberation of very little free chlorophyll and carotenoid [1, 2, 14, 24]. However, studies on changes in the distribution of chlorophyll in the various chlorophyll-protein complexes during photoadaptation have been limited to two studies with plants adapted to only two light intensities, namely, high and low light intensities [19, 20, 23]. These studies showed that the chloroplasts from low-light grown plants have more light-harvesting chl a/b-proteins and less photosystem I-chlorophyll a-protein complexes than chloroplasts from high-light grown plants [13, 19, 20]. However, it is not clear whether these changes are linear with respect to light intensity during growth since only two light intensities were investigated.

In this study, pea seedlings were grown at different light intensities. Analyses of the chlorophyll-protein complexes in the isolated thylakoid membranes show that the changes in the chlorophyll composition, as reflected by the chl a/chl b ratio and relative amounts of chl-proteins, with respect to light intensity can be represented by two straight lines with a transition point at about 200  $\mu$ E m<sup>-2</sup> s<sup>-1</sup>.

# **Materials and methods**

Pea *(Pisum sativum* L.) seedlings were grown in vermiculite in growth cabinets at  $20 \pm 0.5$  °C. The light source was a bank of white fluorescence tubes supplemented with tungsten incandescent lamps (the spectral property of the light source was as published [15]), operated on a 16 hour light/8 hour dark regime. Light intensities, measured with a Li-Cor Quantum Photometer (model LI-185A), were regulated by adjusting the distance between the seedlings and the light source as well as by using neutral density screens.

Chloroplats were isolated as previously reported [15]. Thylakoids were washed in glass-distilled water, followed by two washes and resuspension in 50 mM Tricine (pH 8.0)  $(1-4 \text{ mg chl/ml})$  prior to storage in liquid nitrogen. Total chl and chl a/chl b ratios were determined in 80% acetone [8].

Thylakoid membranes (50 $\mu$ g chl) were solubilized without prior lipid extraction in a solution (100  $\mu$ ) containing 50 mM Tris (pH 8.0), 10% (v/v) glycerol and SDS to give an SDS/chl ratio of 7.5 (w/w). Samples equivalent to  $17.5 \mu$ g chl were immediately applied to SDS polyacrylamide tube gels (4% stacking and 8% separation gels, 6 mm in diameter), as previously described [2]. The gels had been pre-electrophoresed at 3 mA per tube for 15 min, electrophoresis was for 45 min at  $4^{\circ}C$  [2]. Immediately after electrophoresis, these 'green' gels were scanned at 675 and 650 nm on a Varian 635 spectrophotometer fitted with a gel-scanning attachment. The relative distribution of chlorophyll on the gels was estimated as previously described



Figure 1. Effect of light intensity during growth on the chl a/chl b ratio in pea thylakoids isolated from plants grown at different light intensities. Circles represent data from one lot of plants, and squares represent data from another lot grown a few weeks apart.

[1] and were averages of three experiments each with duplicates. Standard deviations were within  $\pm$  7%.

#### **Results**

#### *Photoadaptation of chl a/chl b ratio*

The chl a/chl b ratios of pea thylakoids adapted to different light intensities ranged from 3.2 to 2.1 (Figure 1). The plot of light intensity during growth against chl a/chl b ratios shows clearly that the chl a/chl b ratios increase sharply in a linear manner at low light intensity in the region below  $200 \mu E$  m<sup>-2</sup> s<sup>-1</sup>, but increase gradually and linearly at higher light intensities between 200 to  $840 \mu E \text{ m}^{-2} \text{s}^{-1}$ . While it is well known that plants grown under low intensity light and shade plants have lower chl a/chl b ratios than plants grown under high intensity light and sun plants [10, 11, 12, 17, 22], this experiment with pea thylakoids is the first evidence showing that there is a bilinear response of chl a/chl b ratios to light intensity with an unexpected transition point at about  $200 \mu E m^{-2} s^{-1}$ .

# *Distribution of chlorophyll between the chlorophyll-protein complexes*

The pea thylakoids from plants adapted to different light intensities were solubilized at  $4^{\circ}$ C with an SDS/chl weight ratio of 7.5/1 and subjected to discontinuous SDS polyacrylamide gel electrophoresis at 4°C. Eight chlcontaining bands were resolved (Figure 2) which were identified by their previously characterized spectral properties [1]. They are (in order of increasing mobility): CP1a<sup>1</sup>, CP1a<sup>2</sup>, CP1, LHCP<sup>1</sup>, LHCP<sup>2</sup>, CPa, LHCP<sup>3</sup> and free chlorophyll. Three of these chl-protein bands are associated with PS I: CP1 is the  $\beta$ -carotene-P700-Chl a-protein and CPla<sup>1</sup> and CPla<sup>2</sup> are partially dissociated PS I complexes which include CP1, the antenna chl a/b-proteins of PSI (LHC-I) and other polypeptides. The other four chlorophyll bands are associated with PS II: CPa is the presumed reaction centre complex of PS II and LHCP<sup>1</sup>, LHCP<sup>2</sup> and LHCP<sup>3</sup> are three chl a/b-proteins with chl a/chl b



Figure 2. Unstained SDS-polyacrylamide gels after electrophoresis of chloroplast thylakoids from peas grown under different light intensities. Light intensities during growth are (from left to right) 840, 610, 440, 215, 165, 93 and 42  $\mu$ E m<sup>-2</sup> s<sup>-1</sup>. The designation of the chlorophyll-proteins resolved is referred to in the text.



Figure 3. Relative distribution of chlorophyll in the chlorophyll-protein complexes as a function of light intensities during growth.

ratios of 1.3-1.1 belonging to the main light-harvesting complex of PS II  $(LHC-II)$   $[1-3]$ .

Inspection of the green gels (Figure 2), suggests that the amount of chlorophyll associated with  $CPIa<sup>1</sup>$  and  $LHCP<sup>1</sup>$  increases as light intensity during growth decreases. The reverse is true for the amount of chlorophyll associated with CP1. This was confirmed when the gels were scanned at 650 and 675 nm to determine the average relative amounts of chlorophyll associated with each of the chl-protein bands. The relative distribution of chlorophyll in the chl-protein complexes with respect to light intensities during growth (Figure

3) shows clearly that as light intensity during growth increases the relative percentage of chl associated with  $CPIa<sup>1</sup>$  decreases sharply until the light intensity is above  $200~\mu\text{E m}^{-2}$  s<sup>-1</sup>. At intensities above approximately  $200~\mu$ Em<sup>-2</sup> s<sup>-1</sup>, the relative amount of CP1a<sup>1</sup> remains quite constant. This bilinear type of response towards light intensity with a transition point at about 200  $\mu$ E m<sup>-2</sup> s<sup>-1</sup> is observed in the relative distribution of chlorophyll in all of the chl-protein complexes (Figure 3). As light intensity during growth increases, the relative amount of chlorophyll associated with  $\text{CPIa}^1$ , LHCP<sup>1</sup>, LHCP<sup>2</sup> and LHCP<sup>3</sup> decreases while that associated with CP1a<sup>2</sup>, CP1 and CPa increases in a bilinear manner.

In higher plants, most of the PS II complexes and their associated chl a/b-proteins (LHC-II) are located in the appressed membranes of grana partitions, whereas PS I complex is mainly located in stroma-exposed thylakoids  $[4, 7]$ . Hence the domains of PS II and PS I chlorophyll are largely separated. However, there is still the possibility of some sharing of light energy between PSI and PS II in non-appressed membranes, and it is reported that part of the LHCP pool may be mobile under phosphorylated conditions [cf. 9]. Nevertheless, it is of interest to compare the amounts of chlorophyll associated with the chlorophyll-proteins of PS II and PS I. Since there is some free chlorophyll in the SDS polyacrylamide gels and the value is not constant, it should be noted that the comparisons between the PS I and PS II chlorophyll contents are relative only.

The relative amount of chlorophyll associated with PS I (sum of  $\text{CP1a}^1$ ,  $CP1a<sup>2</sup>$  and CP1) (Table 1, Figure 4) is not markedly affected by light intensity during growth above the  $200~\mu$ Em<sup>-2</sup> s<sup>-1</sup> transition point. Below this point PS I is influenced by light intensity changes. On the other hand, there is a decrease in the relative amount of chl associated with PS II (sum of CPa, LHCP<sup>1</sup>, LHCP<sup>2</sup>, and LHCP<sup>3</sup>) at higher light intensity during growth; thus PS II is more sensitive to light intensity changes. This is also reflected in PS II/PS I chlorophyll content ratio. However, it is clear that in all cases, the bilinear response is observed.

The ratio of chlorophyll content associated with light-harvesting chlproteins of PS II [LHC-II] to that of the PS II core reaction centre complex, i.e. the LHCP<sup>1+2+3</sup>/CPa ratio varies from 3.5 at high light to 8.6 at low light intensity (Table I, Figure 5), showing again the bilinear response and a transition point at about  $200~\mu$ E m<sup>-2</sup> s<sup>-1</sup>. This indicates again that the lightharvesting chl-proteins of PS II are most sensitive to high light intensities than other chl-protein complexes. The marked increase in the LHCP<sup>1+2+3</sup>/CPa ratio from 3.5 to 8.6 with decreasing light intensities occurs because of an increase in LHCP<sup>1+2+3</sup> and a concomitant decrease in CPa. These changes result in significantly greater amounts of chl b in PS II as the light intensity is decreased. Similar increases in the total amount of chlorophyll associated with the main chl a/b-proteins,  $LHCP^{1+2+3}$ , as well as a concomitant two-fold decrease in CPa were observed in pea thylakoids isolated from plants grown

Table 1. Comparison of the relative distribution of chlorophyll in the chl-proteins associated with PS I and PS II in thylakoids from peas adapted to light intensities

Intensities during growth $(\mu E \text{ m}^{-2} \text{ s}^{-1})$	chl a chl b	$PSI^*$ $chl(\%)$	PS II* chl $(\%)$	PS II* PS I	$LHCP^{1+2+3}$ CPa
840	3.21	26.0	55.0	2.12	3.55
735	3.07	26.1	54.1	2.07	3.60
610	3.01	27.2	53.2	1.96	4.16
440	2.93	26.1	58.4	2.24	4.26
215	2.70	25.5	60.6	2.38	4.46
165	2.73	25.0	60.5	2.42	4.31
93	2.55	23.4	63.0	2.69	5.24
42	2.32	22.6	65.8	2.91	5.39
10	2.13	21.5	64.4	3.00	8.61

 $PS I^* = \text{CPIa}^1 + \text{CPIa}^2 + \text{CPI}$ ;  $PS II^* = \text{CPa} + \text{LHCP}^1 + \text{LHCP}^2 + \text{LHCP}^3$ 



Figure 4. Relative distribution of chlorophyll content in PS I and PS II as a function of light intensity during growth.

under different light regimes which varied not only in intensity but also in quality [15].

### *Relationship between chl a/b ratio and chlorophyll-protein complexes*

Since all chlorophyll and carotenoid molecules are non-covalently bound in stoichiometric amounts to a few specific intrinsic proteins which are organized as light-harvesting assemblies of PSI and PS II [5, 27], the changes in chl a/chl b ratios as a result of photoadaptation should also be reflected in the amounts of chlorophyll associated with the main chl-proteins. Although exact measurements of the amount of chlorophyll associated with the two photosystems is not yet feasible by these methods, approximate values may be obtained as stated by summing  $CPIa<sup>1</sup>$ ,  $CPIa<sup>2</sup>$  and  $CPI$  for PS I, and summing CPa, LHCP<sup>1</sup>, LHCP<sup>2</sup>, and LHCP<sup>3</sup> for PS II. When these relative



Figure 5. Effect of light intensity during growth on the  $(LHCP<sup>1</sup> + LHCP<sup>2</sup> +$  $LHCP<sup>3</sup>$  )/Cpa ratio.



Figure 6. Relationship between chl a/chl b ratios and the relative chlorophyll content of PS II and PS I.

chlorophyll contents are plotted against chl a/chl b ratio (Figure 6) it becomes clear that the relationship between PS II chlorophyll content and chl a/chl b ratio is linear except for points scattered at the extreme ends, and, that between PSI chlorophyll content and chl a/chl b ratio is also linear (Figure 6).

# **Discussion**

Although it is well known that the chl a/chl b ratios of higher plants and some green algae increase with higher light intensity during growth [12, 19, 22, 25], it was not clear whether this rate of photoadaptation is linear. Since plants adapted to high light intensity have 'sun' type chloroplasts and those adapted to low light intensity have 'shade' type chloroplasts [11, 12, 19], it is possible that photoadaptation of a given species might be different at high and low light intensities. By studying peas grown at various light intensities, we have now shown for the first time that the photoadaptation of the photosynthetic units of PS II and PSI as reflected by the chl a/chl b ratio is not simply linear with respect to light intensity, but bilinear with a transition point (Figure 1). That is, the rate of photoadaptation of the chl a/chl b ratio at higher light intensities is different from that at lower light intensities. This is also reflected in the relative distribution of chl-protein complexes in the thylakoid membranes (Figures 3, 4 and 5). The light intensity at which this transition point occurs varies from 150 to  $250 \,\mu\text{E m}^{-2} \text{ s}^{-1}$ , reflecting perhaps variations in the measurement of the various parameters.

It is well known that chl a and chl b are present in almost equal amounts in the main LHCP (LHC-II) of PS II (chl a/chl b ratio of  $1.1-1.3$ ) [27]. However, there is also some chl b present in PS I, since recent evidence indicates that PS I has specific, minor chl a/b-proteins with chl a/chl b ratios greater than 1.3, and with apoproteins distinct from those of the main chl a/b-proteins of PS II light-harvesting complexes [6, 26, 29]. Most of the chl b present in thylakoid membranes is associated with LHC-II. In spinach thylakoids, LHC-I accounts for only 8% of the total chlorophyll content of thylakoids, and 14% of the total chlorophyll b [21]. Therefore it is possible from the chl a/chl b ratio to roughly predict the abundance of chlorophyll associated with PS II or PS I. Thus, a high chl  $a$ /chl b ratio would mean less LHC-II and thus lower PS II chlorophyll content and higher PS I chlorophyll content and *vice versa.* This is indeed observed in the present study (Figure 6 and Table 1); higher chl a/chl b ratios indicate higher PS I chlorophyll content and lower PS II chlorophyll content.

As light intensity during growth decreases, relatively more chl becomes associated with PS II (Figure 4) to ensure higher efficiency of lightharvesting in PS II. Furthermore, the chlorophyll composition of PS II changes since the ratio of  $L H C P^{1+2+3}/C P a$  increases dramatically when light intensity is decreased (Figure 5). There is strong evidence that LHC-II is essential for thylakoid stacking [4]. The immediate consequence of increased LHC-II should be more stacked thylakoid membranes, since PS II is localized mainly in stacked membranes [4]. This is indeed the case: a higher degree of stacking is found in low light intensity-adapted thylakoids than those adapted to high light intensity [17, 20].

The chl a/chl b ratios of pea plants grown at high light intensity (3.2 at  $840 \mu \text{E m}^{-2} \text{ s}^{-1}$ ) are comparable to 2.8 of normally grown spinach, a sun plant [3, 4]. On the other hand, the chl a/chl b ratio of pea plants grown at low light intensity (2.1 at  $10 \mu \text{E m}^{-2} \text{ s}^{-1}$ ) is comparable also to 2.2 of *Alocasia,* an extreme shade plant when grown in its natural habitat [3, 4]. Thus, it seems at least with regard to chl a/chl b ratios that pea plants have adapted efficiently to the light intensities used in the present study. However, while the ratio of PS II chlorophyll content to PS I chlorophyll content of 2.1 for peas grown at high light intensity of  $840~\mu E m^{-2} s^{-1}$  is comparable to the value of 2.0 of normally grown spinach [3, 4], the value of 3.0 for peas grown at low light intensity of  $10 \mu E \text{ m}^{-2} \text{ s}^{-1}$  is different from the value of 5.0 reported for the shade plant, *Alocasia* in natural habitat [3, 4].

Although it is shown that pea plants adapt very efficiently to light intensity during growth by changing their chl a/chl b ratios, therby markedly changing the composition of the chl-proteins associated with PS I and PS II, these changes are limited and do not appear to reach the ultimate limits observed in shade plants where the light quality is also markedly different.

A possible explanation for the unexpected change in the linear rate of photoadaptation by pea plants is that they are sun plants which tend, therefore, to behave in a 'normal' way in the light intensity ranges similar to those encountered in their 'natural' habitat, i.e. light intensities above the transition point (approximately  $200~\mu\text{E m}^{-2}$  s<sup>-1</sup>). However, below approximately  $200 \mu \text{E m}^{-2}$  s<sup>-1</sup>, pea plants appear to be less able to cope with the lower light intensities that are more characteristic for shade plants, and so they have to incorporate more drastic changes in the relative distribution of chlorophyll in the various chlorophyll-protein complexes. Hence, the bilinear nature of the response curves.

In view of this unexpected regulatory effect of light intensities on the distribution of chlorophyll-protein complexes, we have also examined the effects of light intensities on the other photosynthetic components of the thylakoids. The results of that investigation are described in the following paper [16].

#### **Acknowledgements**

We thank Mr. I. Dawson and Dr. I. Wardlaw of the Phytotron for providing facilities to grow the peas used in this study.

### **References**

- 1. Anderson JM, Waldron JC and Thorne SW (1978) Chlorophyll-protein complexes of spinach and barley thylakoids. Spectral characterization of six complexes resolved by an improved electrophoretic procedure. FEBS Lett 92:227-233
- 2. Anderson JM (1980) P-700 content and polypeptide profile of chlorophyll-protein complexes of spinach and barly thylakoids. Biochim Biophys Acta 591:113-126
- Anderson JM (1980) Chlorophyll-protein complex of higher plant thylakoids: Distribution, stoichiometry and organization in the photosynthesis unit FEBS Lett 117:327-331
- 4. Anderson JM (1982) The role of chlorophyll-protein complexes in the function and structure of chloroplast thylakoids. Mol Cell Biochem 46:161-172
- 5. Anderson JM, Barrett J and Thorne SW (1981) Chlorophyll-protein complexes of photosynthetic eukaryotes and prokaryotes: properties and functional organization. In Akoyunoglou G, ed. Photosynthesis III. Structure and molecular organization of the photosynthetic apparatus, pp 301-315. Philadelphia. Balaban International Science Services
- 6. Anderson JM, Brown JS, Lam E and Malkin R (1983) Chlorophyll b: an integral component of photosystem I of higher plant chloroplasts. Photochem Photobiol: 38:205-210
- 7. Andersson B and Anderson JM (1980) Lateral heterogeneity in the distribution of chlorophyll-protein complexes of the thylakoid membranes of spinach chloroplasts. Biochim Biophys Acta 593:427-440
- 8. Arnon DI (1949) Copper enzymes in isolated chloroplasts. Polyphenoloxidase in Beta Vulgaris. Plant Physiot 24:1-15
- 9. Bennett J (1983) Regulation of photosynthesis by reversible phosphorylation of the light harvesting chlorophyll a/b-protein. Biochem J  $212:1-13$
- 10. Bj6rkman O (1973) Comparitive studies on photosynthesis in higher plants. In Giese A, ed. Current topics in photobiology, photochemistry and photophysiology, Vol 8: pp 1-63. New York: Academic Press
- 11. Bj6rkmann O (1981) Responses to different quantum flux densities. In Lange OK, Nobel PS, Osmond CB and Ziegler H eds. Encyclopaedia of Plant Physiology Vol 12A: Physiological Plant Ecology, pp 57-107. Berlin: Springer-Verlag
- 12. Boardman NK (1977) Comparative photosynthesis of sun and shade plants. Annu Rev Plant Physiol 28:355-377
- 13. Brown JS, Alberte RS and Thornber JP (1974) Comparative studies on the occurence and spectral composition of chlorophyll-protein complexes in a wide variety of plant material. In Avron M, ed. Proc 3rd Int Congr on Photosynthesis, pp 1951-1962. Amsterdam: Elsevier
- 14. Henriques F and Park RB (1978) Characterization of three new chlorophyllprotein complexes. Biochem Biophys Res Comm 81:1113-1118
- 15. Leong T-Y and Anderson JM (1983) Changes in composition and function of thylakoid membranes as a result of photosynthetic adaptation of chloroplasts from pea plants grown under different light conditions. Biochim Biophys Acta 723:391-399
- 16. Leong T-Y and Anderson JM (1984) Adaptation of the thylakoid membranes of pea chloroplasts to light intensities II. Study on electron transport capacities, coupling factor  $(CF<sub>1</sub>)$  activity and rates of photosynthesis. Photosynthesis Research. This issue, 105-116
- 17. Lichtenthaler HK (1981) Adaptation of leaves and chloroplasts to high quanta fluence rates. In Akoyunoglou G, ed. Photosynthesis VI, Photosynthesis and productivity, photosynthesis and environment, pp 273-287. Philadelphia, Pa.: Balaban International Science Services
- 18. Lichtenthaler HK, Buschmann, C, D611 M, Fietz H-J, Bach T, Kozel U, Meier D and Rahmsdorf U (1981) Photosynthetic activity, chloroplast ultrastructure, and leaf characteristics of high-light and low-light plants and of sun and shade leaves. Photosynthesis Res 2:115-141
- 19. Lichtenthaler HK, Kuhn G, Prenzel U, Buschmann C and Meier D (1982) Adaptation of chloroplast-ultrastructure and of chlorophyll-protein levels to high-light and low-light growth conditions. Z Naturforsch 37C:464-475
- 20. Lictenthaler HK, Kuhn G, Prenzel U and Meier D (1982) Chlorophyll-protein levels and degree of thylakoid stacking in radish chloroplasts from high-light, low-light and bentazon treated plants. Physiol Plant 56:183-188
- 21. Melis A and Anderson JM (1983) Structural and functional organization of the photosvstems in spinach chloroplasts: antenna size, relative electron transport capacity and chlorophyll composition. Biochim Biophys Acta 724:473-484
- 22. Melis A and Harvey GW (1981) Regulation of photosystem stoichiometry, chlorophyll a and chlorophyll b content and relation to chloroplast ultrastructure. Biochim Biophys Acta 637:138-145
- 23. Nurmi A and Vapaavuori E (1982) Chlorophyll-protein complexes in *Salix* sp. 'Aquatica gigantea' under strong and weak light. I. Spectral characterization of the chlorophyll protein complexes. Plant and Cell Physiol 23:785-790
- 24. Remy R and Hoarau J (1978) New form of chlorophyll-protein complexes from thylakoids of different photosynthesizing organisms. In Akoyunoglou G and Argyroudi-Akoyunoglou JH eds. Chloroplast Development, pp 235-240. Amsterdam: Elsevier
- 25. Senger H and Fleischhacker PH (1978) Adaptation of the photosynthetic apparatus of *Scenedesmus obliquus* to strong and weak light conditions. I. Differences in pigments, photosynthetic capacity, quantum yield and dark reactions. Physiol Plant  $43.35 - 42$
- 26. Ish-Shalom D and Ohad I (1983) Organization of chlorophyll-protein complexes

114

of photosystem I in *Chlamydomonas reinhardii.* Biochim Biophys Acta 722: 498-507

- 27. Thornber JP, Markwell JP and Reinman S (1979) Plant chlorophyll-protein complexes: Recent advances. Photochem Photobiol 29:1205-1216
- 28. Wild A (1979) Physiology of photosynthesis in higher plants. The adaptation of photosynthesis to light intensity and light quality in higher plants. Ber Deutsch Bot Ges 92:341-364
- 29. Wollman FA and Bennoun P (1982) A new chlorophyll-protein complex related to photosystem 1 in *Chlamydomonas reinhardii.* Biochim Biophys Acta 680:352-360