

Update section

Short Communication

Cloning and sequence analysis of the glucose-6-phosphate dehydrogenase gene from the cyanobacterium *Synechococcus* PCC 7942

David J. Scanlan, Julie Newman, Mohammed Sebahia, Nicholas H. Mann and Noel G. Carr
Department of Biological Sciences, University of Warwick, Coventry CV4 7AL, UK

Received 23 March 1992; accepted 1 April 1992

Key words: *Synechococcus*, glucose-6-phosphate dehydrogenase, cyanobacterium

Abstract

The glucose-6-phosphate dehydrogenase (EC 1.1.1.49) gene (*zwf*) of the cyanobacterium *Synechococcus* PCC 7942 was cloned on a 2.8 kb *Hind* III fragment. Sequence analysis revealed an ORF of 1572 nucleotides encoding a polypeptide of 524 amino acids which exhibited 41% identity with the glucose-6-phosphate dehydrogenase of *Escherichia coli*.

In cyanobacteria the dissimilation of fixed carbon occurs predominantly via the oxidative pentose phosphate pathway [11]. Glucose-6-phosphate dehydrogenase (G-6-PD, EC 1.1.1.49), apart from being the key enzyme for the entry of fixed carbon into this pathway [see 14], has also been implicated in the supply of reductant to nitrogenase [1]. Because of the central importance of this enzyme in the transition from phototrophic metabolism to heterotrophic metabolism in the dark, the regulation of its activity has been the focus of much attention. To date there are no reports of the characterization, at the nucleotide sequence level, of the G-6-PD gene (*zwf*) from any cyanobacterium or, indeed, from any oxygen-evolving photosynthetic organism. Recently, however, the *zwf* genes from several non-photosyn-

thetic bacterial species [3, 10, 13] have been sequenced and found to exhibit considerable homology both with each other and with G-6-PD genes from eukaryotic organisms. Consequently, we utilized the approach of heterologous hybridization to isolate and characterize a cyanobacterial *zwf* gene.

Chromosomal DNA from *Synechococcus* PCC 7942 was partially digested with *Sau* 3A and used to construct a library in λ charon 35. A 1.9 kb *Kpn* I-*Xho* I fragment from the plasmid pTC117 which is derived from pTC111 [3] and carries the *Zymomonas mobilis* G-6-PD (*zwf*) gene was used as a hybridization probe to screen the library. One clone from the library hybridized strongly with the 1.9 kb *Kpn* I-*Xho* I fragment and contained a 2.8 kb *Hind* III fragment which also hy-

```

      10          30          50          70          90
ATGACTCCAAACTGCTTGAGAACCCGCTTCGCATTGGACTCCGCCAAGACAAAGTCCCTGAACCCGAAATCCTCGTCATCTTTGGGGCC
M T P K L L E N P L R I G L R Q D K V P E P Q I L V I F G A
      110          130          150          170
ACCGGGGACTTAACCCAGCGCAAACCTGGTGCCTGCCATCTACGAGATGCACCTCGAACGGCGTTTCCGCCAGAAGTACGACGATCGTGGGG
T G D L T Q R K L V P A I Y E M H L E R R L P P E L T I V G
      190          210          230          250
GTGGCTCGGCGGACTGGAGCGATGACTATTTCCGAGAGCACCTTCGCCAAGGGGTTGAACAGTTTGGCGCGGCATTCAAGCAGAGGAA
V A R R D W S D D Y F R E H L R Q G V E Q F G G G I Q A E E
      290          310          330          350
GTTTGGAAACACCTTTGCACAAGGCCTCTTCTTTCGCCCGGGCAACATTGATGACCCCCAGTTTTATCAAACCCTTCGCGATCGCCTTGCG
V W N T F A Q G L F F A P G N I D D P Q F Y Q T L R D R L A
      370          390          410          430          450
AATCTGGATGAGCTGCGGGCACGCGGGCAATCGCACTTTTTACCTCTCGGTGCGACCCCGTTTCTTTGGTGAAGCTGCAAAAACAATC
N L D E L R G T R G N R T F Y L S V A P R F F G E A A K Q L
      470          490          510          530
GGGGCAGCCGGAATGCTTGCCGATCCAGCTAAAACGCGGCTGGTCTGAAAAACCTTTTGGCCGCGATCTCAGCTCCGCTCAGGTGCTG
G A A G M L A D P A K T R L V V E K P F G R D L S S A Q V L
      550          570          590          610          630
AATGCCATCTTGAGAACGTTTGGCGGAAAGCCAGATCTATCGGATTGACCATTACCTCGGCAAGAAGCAGTTCAAACCTCTTAGTT
N A I L Q N V C R E S Q I Y R I D H Y L G K E T V Q N L L V
      650          670          690          710
TTCCGGTTTGCCAAATGCCATTTTTGAGCCGCTCTGGAACCGGCAATACATTGACCATGTCCAATCACGGTGGCTGAAACTGTGGGGTTG
F R F A N A I F E P L W N R Q Y I D H V Q I T V A E T V G L
      730          750          770          790          810
GAAGGGCAGCTGGCTACTACGAAACTGCTGGTGTCTGCGGGATATGGTGCAAAAACCACTTGATGCAGCTCTTCAGCCTGACGGCGATG
E G R A G Y Y E T A G A L R D M V Q N H L M Q L F S L T A M
      830          850          870          890
GAGCCGCAAACTCTCTAGGTGCTGACGGTATCCGTAACGAAAAGGTCAAGGTGGTGAAGCCACACGGTGGCGGATATCGACGATCTC
E P P N S L G A D G I R N E K V K V V Q A T R L A D I D D L
      910          930          950          970
AGTTTGTCTGCGGTGCGGGGCGAGTACAAAGCGGGCTGGATGAATGGCCGCTCTGTGCCCGCCTATCGGGATGAGGAGGGAGCGGATCCC
S L S A V R G Q Y K A G W M N G R S V P A Y R D E E G A D P
      1010          1030          1050          1070
CAGTCGTTTACGCCACCTATGTGCCATGAAATTGCTGGTGCACAACCTGGCGCTGGCAGGGAGTCCCGTTCTATCTACGGACGGGTAAA
Q S F T P T Y V A M K L L V D N W R W Q G V P F Y L R T G K
      1090          1110          1130          1150          1170
CGGATGCCAAAAAGGTGACGGAGATTGCCATTCAAGTTCAAACCGTGGCGCACTTGATGTTCCAGTCAGCCACCCAAAAAGTGAATAGT
R M P K K V T E I A I Q F K T V P H L M F Q S A T Q K V N S
      1190          1210          1230          1250
CCCAACGTCTTAGTGCTGCGGATTGACCCCAATGAAGGCGTGTCTTGGCTTTGAAGTGA AACACCCGGGTTCTCGCAACGGACGCGA
P N V L V L R I Q P N E G V S L R F E V K T P G S S Q R T R
      1270          1290          1310          1330
TCGGTGGATATGGACTTCCGCTACGACACGGCTTTTGGCTCCCCACCCAAGAGGCTATAGCCGCTGCTGGTGGACTGCATGCTCGGC
S V D M D F R Y D T A F G S P T Q E A Y S R L L V D C M L G
      1370          1390          1410          1430
GATCAGACGCTGTTACCCCGCTGATGAGGTTGAAGCGTCTTGGCGGGTGTGACGCGTTACTCGAATCTTGGGATGACCCCGCCCAA
D Q T L F T R A D E V E A S W R V V T P L L E S W D D P R Q
      1450          1470          1490          1510          1530
GCCGCTGGCATTCTTTTTACGAAGCTGGCACTTGGGAGCCGCGCAGAGCGGAGCAGTTGATCAACCGTGATGGTGGCGTTGGCGTGTG
A A G I S F Y E A G T W E P A E A E Q L I N R D G A V G V V
      1550          1570
TCTAGGATCCCTGCAACCCAGCTCAATTCTTCTGGAGATGTTTGATGA
S R I P A T Q L N S S G D V * *

```

Fig. 1. Nucleotide sequence and derived amino acid sequence of the 1572 kb ORF encoded on the 2.8 kb *Hind* III fragment from *Synechococcus* PCC 7942.

```

1
a. ....MAVTQTA QACDLV.IFG AKGDLARRKL LPSLYQLEKA GQLNPOTRII GVGRADWDKA AYTQVREAL ETFM.KETID EGLMDLSAR
b. ....MTN TVSTMI.LFG STGDLRQML LPSLYGLDAD GLLADDLRIV CTSRSEYDTD GFRDFAEKAL DRFVSDRLN DDAKAKFLNK
c. MTPKLEENPL RIGLRQDKVP EPQILV.IFG ATGDLTQRKL VPAIYEMHLE RRLPELTIV GVARRODWSDD YFREHLRQGV EQF.GGGIQA EEVWNTFAAG
d. ....MVS EIKTLVTFFG GTGDLAKRKL YPSVFNLYKK GYLQKHFAIV GTARQALNDD EFKQLVRDSI KDFTDQAQA EA....FIEH
      # # ## ** ##** *#* * # * # * # ##* ## ## # # * ## # ##

101
LDFCNLDVND T.....AA FSRLGAMLQD KNRITINYFA MPPSTFGAIC KGLGEAKLNA KPA..RVVME KPLGTSLATS QEINDQVGEY FEECQVYRID
LFYATVDITD P.....TQ FGKLADLCGP VEKGIAIYLS TAPSLFEGAI AGLKQAGLAG PTS..RLALE KPLGQDLASS DHINDAVLKV FSEKQVYRID
LFFAPGNIDD PQFYQLTRDR LANLDELRTG RGNRT.FYLS VAPRFFGEAA KQLGAAGMLA DPAKTRLVVE KPFGRDLSSA QVLNAILQNV CRESQIYRID
FSYRAHDVTD AASYAVLKEA IEAAADKFDI DGNRI.FYMS VAPRFFGTIA KYLKSEGLLA DTGYNRLMIE KPFQTSYDIA AELQMDLENA FDDNQLFRID
#### # * # # # # # # # ##** *#*#* #*#*#* ## #* #*# #*#* *#* * **#* ## #* #*# # #* #*#*#*

201
HYLGKETVLN LLALRFANSL FVNNWDRTI DHVEITVAEE VGIEGRWGYF DKAGQMRDMI QNHLLQILCM IAMSPPSDLS ADSIRDEKVK V.PEVSSPHR
HYLGKETVQN LLTLRFGNAL FEPLWNSKGI DHVQISVAET VGLEGRIGYF DSGSLRDMV QSHILQLVAL VAMEPPAHME ANAVRDEKVK VFRALRPINN
HYLGKETVQN LLVRFANAI FEPLWNRQYI DHVQITVAET VGLEGRAGYF ETAGALRDMV QNHLMQLFSL TAMEPPNSLG ADGIRNEKVK VVQATRLADI
HYLGKEMVQN IAALRFGNPI FDAAWNKDYI KHVQVTLSEV LGVEERAGYF DTAGALLDMI QNHTMQIVGW LAMEKPESFT DKDIRAAKNA AFNALKIYDE
*****#*#* ## **#*#* ##**#*#* #* #*#*#*#*#* #*#*#*#*#* #*#*#*#* #*#*#*#* #*#*#*#* #*#*#*#* #*#*#*#* #*#*#*#*

301
PLQRTRKNRT RAIYCVQKG KVPGYLEEEG ANKSSNTETF VAIRVDIDNW RWAGVPFYLR TGKRLPTKCS EVVVFYKTP E LNLFKESQD LP.QNKLTIR
DTVFTHTVTG QYGAGVSGGK EVAGYIDELG ..QPSDTETF VAIKAHVDNW RWQGVFPYLR TGKRLPARRS EIVVQFKPVP HSIFSSSGGI LQ.PNKLRIV
DOLSLSAVRG QYKAGVMNGR SVPAYRDEEG ADPOSFPTY VAMKLLVDNW RWQGVFPYLR TGKRMPPKVT EIAIQFKTVP HLMFGSATQK VNSPNVLVLR
AEVNKYFVRA QYGAG..DSA DFKPYLEELD VPADSKNNTF IAGELQDLP RWEGVFPYVR SGKRLAAKQT RVDIVFKAGT FNF...GSEQ EAGEAVLSII
# ##* ##* # #*#* #*#*#* #*#*#*#*#* #*#*#*#*#* #*#*#*#*#* #*#*#*#*#* #*#*#*#*#* #*#*#*#*#* #*#*#*#*#*

401
LQPDGVDIQ VLNKVPLGDH K.HNLQITKL DLSYSEFMQ THLADAYERL LLETHRGIGA LFVRRDEVEE AWKWD SITE AWAMNDAPK P..YQAGTWG
LQPDQTIQIS MNVKEPGLDR NGAHMREVWL DLSLTDVFKD RKRRIAYERL HLDLIEGDAT LFVRRDEVEA QWVWIDGIRE GWKANSKPK T..YVSGTWG
IQPNEGVSRL FEVKTPG... SSORTRSVDM DFRYDTAF.G SPTQEAYSRL LVDCMLGDQT LFTRADEVEA SWRVWTPLE SDDPRQAG ISFYEAGTWE
IDPKGAIELK LNAKSYE... DAFNTRTIDL GWTVSDE.DK KNTPEYERM IHDTHMGDGS NFADWNGVSI AWKFVVAISA VYTADKAPLE T..YKSGSMG
##* ##* # #*#* #*#*#* #*#*#* #*#*#* #*#*#* #*#*#* #*#*#* #*#*#* #*#*#* #*#*#* #*#*#* #*#*#* #*#*#* #*#*#*

501
PVASVAMITR DG....RSM EFE.....
PSTAIALAER DG....VTWY D.....
PAEAEQLINR DGAVGVVSR I PATQLNSSGD V
PEASDKLLAA NGDAWVFKG. ....
* # ## #* ##*

```

Fig. 2. Alignment of the amino acid sequences of glucose-6-phosphate dehydrogenases from (a) *Escherichia coli*, (b) *Zymomonas mobilis*, (c) *Synechococcus* PCC 7942 and (d) *Leuconostoc mesenteroides* obtained using the PILEUP program [6]. The * symbol indicates residues conserved in all four sequences and the # symbol indicates where a residue found in the *Synechococcus* PCC 7942 protein occurs in at least one of the other proteins.

bridized strongly with the probe. This 2.8 kb *Hind* III fragment was subcloned into pUC19 to yield pNUT1. The nucleotide sequence of the 2.8 kb fragment was determined by the dideoxy chain termination method following a combination of both random and directed subcloning into M13 mp18 and M13 mp10. Analysis of the nu-

cleotide sequence revealed an ORF (524 amino acids) of 1572 nucleotides followed by two stop codons (Fig. 1). Alignment of the amino acid sequence of the predicted polypeptide encoded by this ORF with those of known G-6-PD from prokaryotic sources using the PILEUP program [6] revealed extensive homologies (Fig. 2) strongly

suggesting that this ORF represents the authentic *zwf* gene of *Synechococcus* PCC 7942. The cyanobacterial G-6-PD was estimated to be 60% similar (41% identical) to that of *Escherichia coli* using the GAP alignment program [6]. In addition, the predicted polypeptide includes the canonical sequence (DHYLGKE) of the active site of G-6-PD [9] as recognized in the PROSITE dictionary.

Numerous studies have been directed at analysing the regulation of activity of G-6-PD activity in cyanobacteria particularly in connection with the light-dependent control of activity. The metabolites implicated in regulation include NADPH [1, 12], ATP [8] and thioredoxin [5], the latter having a role in regulating G-6-PD activity in chloroplasts [2]. Thioredoxin can function as a protein disulphide reductase and has been shown to reduce the disulphides in certain proteins, such as the activation of chloroplast fructose-1,6-bisphosphatase [4]. In this context it is worth noting that the amino acid sequence reported here for the *Synechococcus* PCC 7942 enzyme contains only two cysteine residues and consequently both residues would be involved in such regulation. Neither of the two cysteines present in the cyanobacterial polypeptide are conserved residues in the other prokaryotic enzymes and one of the cysteines (Cys-188), which replaces a conserved phenylalanine in the other prokaryotic enzymes, lies immediately prior to the active site region defined by the sequence DHYLGKE.

Acknowledgements

We thank Dr T. Conway for the kind gift of plasmid pTC117. Computer-assisted analysis of the nucleotide sequence information was carried out via the Seqnet facility of the SERC Daresbury Laboratory, U.K.

References

1. Apte SK, Rowell P, Stewart WDP: Electron donation to ferredoxin in heterocysts of the N_2 -fixing alga *Anabaena cylindrica*. Proc R Soc B200: 1–25 (1978).
2. Ashton AR, Brennan T, Anderson LE: Thioredoxin-like activity of thylakoid membranes. Plant Physiol 66: 605–608 (1980).
3. Barnell WO, Yi KC, Conway T: Sequence and genetic organization of a *Zymomonas mobilis* gene cluster that encodes several enzymes of glucose metabolism. J Bact 172: 7227–7240 (1990).
4. Clancey CJ, Gilbert HF: Thiol/disulfide exchange in the thioredoxin-catalyzed reductive activation of spinach chloroplast fructose-1,6-bisphosphatase. J Biol Chem 262: 13545–13549 (1987).
5. Cossar JD, Rowell P, Stewart WDP: Thioredoxin as a modulator of glucose-6-phosphate in a N_2 -fixing cyanobacterium. J Gen Microbiol 130: 991–998 (1984).
6. Devereux J, Haeberli P, Smithies O: A comprehensive set of sequence analysis programs for the VAX. Nucl Acids Res 12: 387–395 (1984).
7. Eichorn M, Corbus B: Metabolic role of glucose-6-phosphate dehydrogenase in photoautotrophic organisms. Biochem Physiol Pfl 183: 449–475 (1988).
8. Grossman A, McGowan RE: Regulation of the glucose-6-phosphate dehydrogenase in blue-green algae. Plant Physiol 55: 658–662 (1975).
9. Jeffery J, Wood I, McLeod A, Jeffery R, Jornvall H: Glucose-6-phosphate dehydrogenase. Characterization of a reactive lysine residue in the *Pichia jadinii* enzyme reveals a limited structural variation in a functionally significant segment. Biochem Biophys Res Commun 160: 1290–1295 (1989).
10. Lee WT, Flynn TG, Lyons C, Levy HR: Cloning of the gene and amino acid sequence for glucose-6-phosphate dehydrogenase from *Leuconostoc mesenteroides*. J Biol Chem 266: 13028–13034 (1991).
11. Pelroy RA, Bassham JA: Photosynthetic and dark carbon metabolism in unicellular blue-green algae. Arch Microbiol 86: 25–38 (1972).
12. Pelroy RA, Kirk MR, Bassham JA: Photosystem II regulation of macromolecule synthesis in the blue-green alga *Aphanocapsa* 6714. J Bact 128: 623–632 (1976).
13. Rowley DL, Wolf Jr RE: Molecular characterization of the *Escherichia coli zwf* gene encoding glucose-6-phosphate dehydrogenase. J Bact 173: 968–977 (1991).
14. Smith AJ: Modes of cyanobacterial carbon metabolism. In: Carr NG, Whitton BA (eds) The Biology of the Cyanobacteria, pp. 47–85. Blackwell Scientific Publications, Oxford (1982).