Short communication

# Effect of temperature shock on the dynamics of abscisic acid and wheat germ agglutinin accumulation in wheat cell culture

F.M. Shakirova, M.V. Bezrukova & I.F. Shayakhmetov

Department of Biochemistry and Cytochemistry, Ufa Scientific Centre, Russian Academy of Sciences, pr. Octyabrya 69, Ufa, 450054, Russia

Received 2 June 1995; accepted 20 October 1995

Key words: abscisic acid, temperature shock, wheat germ agglutinin, wheat callus cells

#### Abstract

Abscisic acid (ABA) and lectin content was immunoassayed in wheat cell cultures affected by temperature stress. The elevated temperature (40 °C) resulted in a 7-fold increase in the level of ABA and a 10-fold increase in that of lectin. The increase in the lectin content in cells was preceded by ABA accumulation. It is suggested that this ABA increase induces the synthesis of lectin, which in addition to stress proteins, play an important role in controlling mechanisms of plant adaptation to unfavourable environments.

Abbreviations: ABA – abscisic acid; WGA – wheat germ agglutinin

# 1. Introduction

Wheat germ agglutinin (WGA), a typical representative of cereal lectins, is involved in the development of plant resistance to various stresses [2]. Abscisic acid (ABA) is known to participate in the regulation of lectin synthesis. ABA accumulation preceeds lectin accumulation in the case of fungal pathogenesis [5], drought, osmotic shock [1] and salinity [8]. However, there are no data on whether quantitative changes in ABA induced by temperature shock affect lectin levels. Consequently, we studied the effect of high temperature on the dynamics of ABA and WGA contents in wheat callus tissue.

## 2. Materials and methods

# 2.1 Plant material

Callus cells were formed from immature embryos of wheat (*Triticum durum* L.) cv. Bezenchukskaya 139, obtained from Chishminsky Crop Production, Bashko-

rtostan, Russia after 10 days of subcultivation on Murashige and Skoog nutrient medium [7].

# 2.2 Treatment of wheat callus cells

For temperature stress treatment, tubes with calli were placed in thermostatically-controlled conditions at 40 °C whilst control tubes were kept at 24 °C. After 4, 9, 14 and 18 h of treatment, samples of callus tissue were fixed in liquid nitrogen.

## 2.3 Quantitative estimation of ABA and WGA content

Callus tissue was homogenized in 80% ethanol and kept at  $4^{\circ}$ C for 16 h to extract ABA (meanwhile lectin remained insoluble [6]). After centrifugation at 10,000 g for 10 min. the ethanol extract was evaporated to an aqueous residue from which ABA was partitioned into diethyl ether. The procedure of purification and ABA immunoassay was carried out according to [10]. After removal of ethanol, proteins were extracted from the pellet with 0.05 N HCl for 1 h at room temperature. The extract was centrifuged at 15,000 g for 10

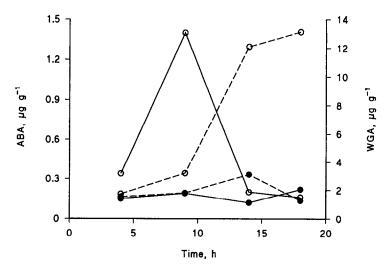


Figure 1. Effect of heat shock on the dynamics of ABA and lectin content in wheat callus cells. Solid lines – ABA, broken lines – WGA. ● 24 °C (control), ○ 40 °C. The ratios of standard deviation to average were less than 12%.

min; the supernatant was neutralized with 1 M sodium phosphate buffer (pH 7.2) and aliquots of the resulting crude plant extract were tested for WGA content by ELISA using specific rabbit antibodies to WGA and anti-rabbit peroxidase-labelled antibodies as described previously [5].

#### 3. Results and discussion

High temperature resulted in a sharp ABA accumulation in cells (Figure 1). Thus, 4 h after callus was exposed to 40 °C the ABA content doubled and after 9 h increased by 7 times, in comparison with controls. Subsequently, ABA content decreased in comparison to the controls. These large ABA changes indicated that cells cultivated at 40 °C suffered from severe stress which caused major changes in their metabolic activity. These results are consistent with the literature reporting increases in ABA level in response to different unfavourable factors in the environment [4, 10]. It has been proposed that ABA acts as a common mediator in defence responses of plants to stresses [4].

In parallel with ABA determinations, we analysed lectin content in the same callus tissues. Figure 1 shows that heat shock resulted in an increase in WGA level nearly 10 times greater than in control cells after 18 h of exposure. The progressive lectin accumulation which followed the sharp increase in ABA level may be the result of a rise in *de novo* lectin synthesis induced by stress. This agrees with the results of Spadoro-Tank and

Etzler [9] who showed induction of the synthesis of a lectin-related protein in *Dolichos biflorus* cell suspension culture affected by temperature shock.

Under stress conditions ABA induces the synthesis of stress proteins [4]. This process is observed within a background of suppression of some major proteins present in cells under normal conditions [4]. However, together with stress proteins there are some proteins contained by normal cells [3] which are increased during plant adaptation to stresses. Wheat lectin is probably an example of such proteins, the quantitative increase in lectin level being one of the universal nonspecific mechanisms of plant protection against the influence of unfavourable environmental conditions.

#### References

- Cammue BPA, Broekaert WF, Kellens JTC, Raikhel NV and Peumans WJ (1989) Stress-induced accumulation of wheat germ agglutinin and abscisic acid in roots of wheat seedlings. Plant Physiol 91: 1432–1435
- Chrispeels MJ and Raikhel NV (1991) Lectins, lectin genes and their role in plant defense. Plant Cell 3: 1-9
- Ericson MC and Alfinito ShCH (1983) Protein produced during salt stress in tobacco cell culture. Plant Physiol 72: 31
- Hong B, Barg R and Ho TD (1991) Developmental and organspecific expression of an ABA and stress-induced protein in barley. Plant Mol Biol 18: 663-674
- Khairullin RM, Shakirova FM, Maksimov IV and Bezrukova MV (1993) Change in the content of lectin, abscisic and indolylacetic acids in the wheat plants infected with Septoria nodorum Berk. Fiziol Biokhim Kult Rast 25: 138-144
- Lutsik MD, Panasjuk EN and Lytsik AD (1981) Lectins. L'vov: Vysshaya Shkola

- Murasige T and Skoog F (1962) A revised medium for rapid growth and bioassays with tobacco tissue cultures. Physiol Plant 15: 473-497
- Shakirova FM, Bezrukova MV and Khairullin RM (1993) The increase in lectin level in wheat shoots under the action of salt stress. Izv Russ Acad Sci 1: 142–145
- Spadoro-Tank JP and Etzler ME (1988) Heat shock enhances the synthesis of a lectin-related protein in *Dolichos biflorus* cell suspension cultures. Plant Physiol 88: 1131–1135
- Yamaleev AM, Yarullina LG, Shakirova FM, Karavaiko NN, Kudojarova GR, Mustafina AR and Moshkov IE (1989) Effect of fungicide baytan on indolylacetic and abscisic acids levels in wheat plants infected with root rot. Soviet Plant Physiol 36: 399-403