Classification of GA response, *Rht* genes and culm length in Japanese varieties and landraces of wheat

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Summary

The GA response, *Rht* genes and culm length of 133 Norin varieties, 6 breeding lines and 16 landraces of Japanese wheat were investigated. Out of 133 Norin varieties tested, 103 were GA-insensitive and 30 GA-responsive. The 6 breeding lines were all GA-insensitive. Out of 16 landraces tested, 10 were GA-insensitive and 6 GA-responsive. Among the 10 GA-insensitive landraces, only Daruma had a *Rht1* genotype. The other 9 had a *Rht2* genotype. None of the landraces tested carried both *Rht1* and *Rht2* or *Rht3*. Out of the 103 GA-insensitive Norin varieties, 22 carried only *Rht1*, another 79 carried only *Rht2*, and only Norin 10 and Kokeshikomugi carried both *Rht1* and *Rht2*. No tested variety carried *Rht3*. Some Norin varieties carrying *Rht2* showed tall culms comparable to that of the *rht* tester line Chinese Spring. These results suggest that these varieties had a nullifier or modifier gene(s) or height promoting genes in the background controlling the height-reducing effect of *Rht2*. Conversely, six GA-responsive Norin varieties were as short as Akakomugi which carries the GA-responsive *Rht* genes, *Rht8* and *Rht9*. They also seemed to carry a GA-responsive *Rht* gene or genes, and moreover all but one may carry gene(s) other than the Akakomugi genes. The origin of *Rht1* and *Rht2* of Norin 10 was speculated on the GA-response and *Rht* genotypes of its related varieties and landraces.

Introduction

In bread wheat, *Triticum aestivum* L., two main sources of semi-dwarfing genes have been successfully used for reducing plant height and lodging susceptibility of wheat. Concurrent increases in grain yield have been and are being recorded in various parts of the world. These two sources are the Norin 10 genes *Rht1* and *Rht2* which are characterized by gibberellin (GA) insensitivity and the Akakomugi genes *Rht8* and *Rht9* which are GA sensitivity.

Rht1 is carried on chromosome 4B (Gale & Marshall, 1976; see later – chromosome designation), while *Rht2* is localized at a homeologous position

on the short arm of chromosome 4D (Gale et al., 1975; McVittie et al., 1978). *Rht3*, a stronger GA insensitive gene (Zeven, 1969) carried by Tom Thumb wheats (Morris et al., 1972) is allelic to *Rht1* (Gale & Marshall, 1976). *Rht8* and *Rht9* are carried on chromosomes 2D and 7B, respectively (Law et al., 1981). Although the GA-responsive *Rht* genes are difficult to detect, the *Rht1*, *Rht2* and *Rht3* genes can readily be detected at the seedling stage by their insensitivity to GA (Gale & Law, 1977). Gale et al. (1981) identified and classified these genes in a number of international varieties, and Worland (1986) surveyed European wheats for the presence of GA-insensitive dwarfing genes.

Japanese wheat breeders released 133 Norin va-

rieties between 1929, when Norin 1 was first developed and 1988. Gale et al. (1981) reported that Norin 2 and Norin 10 carried both *Rht1* and *Rht2*, while Aobakomugi carried only *Rht2*. Yamada (1989) identified the GA-insensitive *Rht* genes of 18 Japanese modern varieties and landraces. However, the dwarfing genes of the other Japanese wheats have not been documented yet.

This paper reports the results of a series of experiments in which the GA response, *Rht* genotypes and culm length of 155 Japanese wheats have been studied.

Materials and methods

The 133 Norin varieties, six breeding lines and 16 landraces together with the 15 tester lines were subjected to a seedling GA response test and a field trial to determine final culm length. The tester lines which are shown in Table 1 with their genotypes, were kindly supplied by Dr. M.D. Gale, Cambridge Laboratory, Institute of Plant Science Research, Cambridge.

For the GA response test, seeds were germinated in Petri dishes for 2 days at 20°C then transplanted to a seed trays in a mixture of vermiculite, soil and sand in the ratio of 2:2:1 volume. A proper quantity of compound manure and slaked lime in $323(L) \times 233(W) \times 52(H)$ mm was added. The seedlings were grown at the density of 204 individuals/tray in a growth chamber under controlled light and temperature conditions. After 21 days of cultivation at 20/15°C (day/night) for 16/8 hours and applications of irrigating water containing 10 ppm gibberellic acid (GA_3) every other day, the length of the first leaf sheath (LS) was measured. By this time the growth of the first leaf sheath had ceased. Under these conditions it was possible to distinguish three phenotypes, i.e., fully and partially insensitive to GA and responsive to GA. In the case of the varietal GA response test, control GAuntreated seedlings which received an equal quantity of irrigating water without GA₃ were grown at the same time as the cultivation of the GA-treated seedlings. These GA-treated and GA-untreated

tests were made with 3 replications in which each replication contained 12 seedlings for each variety. The GA response index (GRI) of each variety was calculated using the following numerical expression: (LS of GA-treated seedlings/LS of GA-untreated seedlings) \times 100.

The GA-insensitive Japanese varieties, breeding lines and landraces were hybridized with the *rht*, *Rht1*, *Rht2*, *Rht3* and *Rht1 Rht2* tester lines. The F₁ hybrids were then selfed to produce F₂ populations. A number of 200–800 F₂ seedlings/cross, 6–12 F₁ seedlings/cross were subjected to the GA response test. χ^2 tests were performed to examine the goodness of fit between expected and observed segregations in the F₂ populations from the test crosses. Thereafter the genotypes of the Japanese wheats were identified.

For the measurement of the final culm length, the plants were grown in a field at a spacing of 10 cm within the row and 60 cm between rows in a randomized block design with 2 replications. One replication contained 170 plots, with one plot for each variety. Each plot contained 10 plants of each variety. At the ripening stage, the culm length of 5 plants/plot was measured at the level of the highest culm of the tillers of each plant.

Results and discussion

The GRI value of the GA-responsive tester line Chinese Spring was 240 (Table 1). The GA-treated seedlings of Chinese Spring also showed the distinct phenotype characterized by thin stems with light green leaves and leaf sheaths. The GRI values of the GA-insensitive Rht1 and Rht2 tester lines ranged from 142 to 179, and those of the Rht3 tester lines ranged from 101 to 108 indicating that these lines showed a high level of insensitivity. The GRI values of two Rht1 Rht2 tester lines were 116 and 104. These values were lower than those of the tester lines with single gene Rht1 and Rht2 indicating that these genes additively reduced the GRI value as well as the culm length. Especially in Olesen, the GRI value was comparable to those of the Rht3 tester lines. Fick and Qualset (1973) reported

that Olesen carried a third dwarfing gene in addition to *Rht1* and *Rht2*. The third gene of Olesen may also induce the reduction of the GRI value.

The GRI values of 155 Japanese wheats ranged from 101 in D. W. sel. No. 71 to 302 in Eshima and Haruhikari (Tables 2 and 3). The frequency distribution of the GRI values was discontinuous at 200– 210 (Fig. 1). The wheats having GRI values below 200 did not show the characteristic phenotype such as thin stems with light green leaves and leaf sheaths presented in the GA-responsive wheat plant. Thus the 103 Norin varieties, 6 breeding lines and 10 landraces with GRI values below 200 were classified into the GA-insensitive group (Tables 2 and 3).

The results of the *Rht* gene analysis obtained from the test crosses are shown in Tables 2 and 3. Among the 10 GA-insensitive landraces, only Daruma had a *Rht1* genotype and the other 9 had a *Rht2* genotype. None of the tested landraces carried both *Rht1* and *Rht2* or *Rht3*.

As mentioned above, among the 133 Norin varieties 103 (77%) and 30 (23%) were GA-insensitive and GA-responsive, respectively, indicating that the majority of them were GA-insensitive. Of the 103 GA-insensitive Norin varieties 22 (21%) carried only Rht1, 79 (77%) carried only Rht2, and only 2 (2%), Norin 10 and Kokeshikomugi, carried both Rht1 and Rht2. None of the Norin varieties tested carried Rht3. These results indicated that the majority of the GA-insensitive Norin varieties carried only Rht2.

Gale et al. (1981) reported that Norin 2 and Norin 10 carried both *Rht1* and *Rht2*, while Aobakomugi carried only *Rht2*. The *Rht* genotypes of Norin 10 and Aobakomugi identified in this study agreed with their results. However, it was found that Norin 2 carried only *Rht2* as shown in Table 3. It is unlikely that the Norin 2 variety tested in this study carried both *Rht1* and *Rht2* because its culm length, 111 cm, was comparable to that of the single gene *Rht2* tester lines or was rather slightly taller.

Among the Norin varieties, the four varieties identified as carriers of *Rht1*; Norin 11, Shirasagikomugi, Mikunikomugi and Gogatsukomugi, and one variety identified as carrying *Rht2*, Norin 52, showed no connections with the recognised sources of the corresponding *Rht* genes in their pedigrees (Fig. 2). Based on pedigree studies, it would be expected that these 4 varieties, Norin 11, Shirasagi-

GRI¹(%) GA reaction² Genotypes Culm length Tester line (cm) Chinese Spring 240 R rht 127 I Rht1 87 SD1 174 I Rht1 86 Highbury 172 Peniamo 62 179 I Rht1 95 Sona 227 166 T Rht1 91 92 I Rht2 SD2 165 I Rht2 76 Maris Hobbit 165 Maris Fundin 142 I Rht2 72 Pitic 62 148 Ι Rht2 101 163 Ι Rht2 80 Sonora Rht3 45 Tom Thumb³ 101 MI MI Rht3 33 D6899 105 Minister Dwarf 108 MI Rht3 41 Norin 10/Brevor 14 116 I Rht1 Rht2 65 104 I Rht1 Rht2 39 Olesen

Table 1. GA response index, GA reactions, Rht genotypes and culm length of tester lines

¹GA response index: (Length of the first leaf sheath (LS) of GA-treated seedlings/LS of GA-untreated seedlings) × 100.

²R: responsive, I: insensitive, MI: markedly insensitive.

³Synonymous to Tom Pouce.



Fig. 1. Frequency distribution of GA response index of Japanese Norin varieties, breeding lines and landraces.

komugi, Mikunikomugi and Gogatsukomugi, should carry *Rht2* and Norin 52 would be predicted to be GA-responsive. The *Rht1* gene of two breeding lines, Saitama 27 and Saitama 29, must have been derived from the landrace Hayakomugi because they were not related to the other possible *Rht1* gene sources in their pedigree, i.e., California/Sojukuakage//Hayakomugi. Both California and Sojukuakage were the GA-responsive varieties whose GRI values were 262 and 233, respectively. These facts supported the above speculation. However since the genotype of Hayakomugi tested in this study was identified as Rht2, it is reasonable to assume that a different Hayakomugi carrying the Rht1 gene had existed.

Worland (1986) reported that, although clearly insensitive, Saitama 27 and its derivatives were less insensitive than would have been expected for materials homozygous for either of the Norin 10 genes. Worland & Petrovic (1988) also reported that the new weak source of GA insensitivity derived from Saitama 27 was controlled by a single gene located on chromosome 4B (see later – chromosome designation) and allelic to *Rht1* and *Rht3*, and it was proposed to temporarily refer to the Saitama 27 allele as *Rht1S* to distinguish it from *Rht1*. However, in this study the GRI value of Saitama 27 was 118 which was lower than those of the *Rht1* tester lines. As mentioned above Saitama 29 has the same pedigree as Saitama 27 and based

Table 2. GA response index, GA reactions, Rht genotypes and culm length of Japanese landraces

Landrace	GRI ¹ (%)	GA reaction ²	Genotype	Culm length
				(cm)
Akadaruma	150	Ι	Rht2 ³	91
Akadaruma 7	113	Ι	Rht2	88
Akakawaaka	252	R		162
Akakomugi	243	R	Rht8 Rht94	98
Daruma	190	Ι	Rht1	103
Daruma 2	223	R	-	109
Eshima ⁵	302	R	_	134
Hayakomugi	162	I	Rht2 ^{3,6}	90
Hiroshima Shipree	149	I	$Rht2^3$	102
Sapporoharukomugi	258	R	_	160
Shirochabo	144	I	Rht2 ³	105
Shirodaruma	150	I	Rht2 ³	78
Shirodaruma Sai 1	117	I	Rht2	80
Shisen 1	227	R	· _	109
Shou Fultz	136	Ι	Rht2	101
Yushoki 347	156	I	$Rht2^3$	80

¹GA response index: (Length of the first leaf sheath (LS) of GA-treated seedlings/LS of GA-untreated seedlings) \times 100.

²R: responsive, I: insensitive.

³Cited from Yamada (1989).

⁴Cited from Law et al. (1981).

⁵Synonymous to Ejima.

⁶ This may not be correct as many Southern European wheats carry a weak GA insensitive allele from Saitama 27 (Worland & Petrovic, 1988). This allele located at the same position as *Rht1/Rht3* on 4B (see later – chromosome designation) appears to originate from Hayakomugi. Therefore this should not carry *Rht2*.

-: Genes not identified.

Norin No.	Variety and breeding line	GRI ¹ (%)	GA reaction ²	Genotypes	Culm length (cm)
1	Norin 1	167	I	Rht2	97
2	Norin 2	159	Ι	Rht2	111
3	Norin 3	276	R	_	153
4	Norin 4	159	I	Rht2	90
5	Norin 5	149	Ι	Rht2	81
6	Norin 6	157	Ι	Rht2	90
7	Norin 7	168	Ι	Rht2	87
8	Norin 8	250	R		143
9	Norin 9	145	Ι	Rht2	80
10	Norin 10	115	Ι	Rht1 Rht2 ³	72
11	Norin 11	172	I ·	Rht14	92
12	Norin 12	145	Ι	Rht2	92
13	Norin 13	150	Ī	Rht2	89
14	Norin 14	158	Ī	Rht1	100
15	Norin 15	253	R	_	111
16	Norin 16	146	T	Rht2	84
10	Norin 17	140	I	Rht2	90
18	Norin 18	130	T	Rht2	84
10	Norin 10	133	T	Rht?	07 07
19	Norin 20	228	P	-	103
20	Norin 21	157	I		88
21	Norin 22	137	P		100
22	Norin 22	223	R D	_	109
25	Norin 24	233	K I	- Dh+7	107
24	Norin 24 Norin 25	177	T	R/112 DL+7	80
25	Norin 25 Narin 26	100	. I . T	N#12 DL+1	04
20	Norm 20 Navia 27	101	I D	Nm	74 110
27	Norin 27	237	K T	- DL/2	112
28	Norin 28	141	I D	Rni2	135
29	Norin 29	2/8	K	- nt-0	125
30	Norin 30	151	I D	KN12	19
31	Norin 31	221	ĸ		120
32	Norin 32	161	· 1	Knti DL 2	95
33	Norin 33	158		Rht2	120
34	Norin 34	233	ĸ	-	103
35	Norin 35	224	ĸ	-	128
36	Norin 36	225	R	-	107
37	Norin 37	226	R	-	109
38	Norin 38	266	R	-	113
39	Norin 39	265	R	-	123
40	Norin 40	242	R		123
41	Norin 41	216	R	-	102
42	Norin 42	238	R	-	88
43	Norin 43	246	R	-	93
44	Norin 44	155	I	Rht2	91
45	Norin 45	241	R	-	88
46	Norin 46	254	R	-	98
47	Norin 47	266	R		98
48	Norin 48	162	Ι	Rht2	77
49	Norin 49	173	Ι	Rht1	94
50	Norin 50	139	I	Rht2	84

Table 3. GA response index, GA reactions, Rht genotypes and culm length of Japanese Norin varieties and breeding lines

Table 3.	Continued

Norin No.	Variety and	GRI ¹ (%)	GA reaction ²	Genotypes	Culm length
	breeding line			o the type to	(cm)
51	Norin 51	231	R		100
52	Norin 52	158	l T	Rht2*	8/
53	Norin 53	163		Rhtl	96
54	Norin 54	151	I ,	Rht2	84
55	Norin 55	157	1	Rht2	99
56	Norin 56	159	1	Rht2	94
57	Norin 57	153	1	Rht2	89
58	Norin 58	139	1	Rht2	110
59	Norin 59	177	I	Rht2	90
60	Norin 60	167	I	Rht2	96
61	Norin 61	188	I	Rht2 ⁵	92
62	Norin 62	248	R	-	117
63	Norin 63	163	I	Rht2	95
64	Norin 64	142	I	Rht2	85
65	Norin 65	164	I	Rht2	102
66	Norin 66	163	I	Rht2	104
67	Norin 67	152	Ι	Rht2	97
68	Norin 68	142	I	Rht2	86
69	Norin 69	170	I	Rht1	101
70	Norin 70	160	Ι	Rht2	104
71	Norin 71	162	I	Rht2	103
72	Norin 72	157	I	Rht2	92
73	Norin 73	158	I	Rht2	89
74	Norin 74	169	I	Rht2	97
75	Norin 75	292	R		131
76	Yuvakekomugi	192	I	Rht?	101
77	Susonokomugi	166	Ī	Rht?	101
78	Mutsubenkei	177	Ĩ	Rht2	124
79	Ivokomugi	182	T	Rht?	03
80	Hatamasari	263	P	1(112	95 100
81	Aobakomugi	163	I	- PL+3 ³	07
87	Nanbukomugi	160	I	N/112 DL+25	97
83	Akateukikomugi	107	I	Rht2	91
84	Nukishaha	162	I T		8J 00
0 4 95	Likorikomugi	170	I T	R <i>nu</i> 2 (DL+1)6	00
85	Muskakamugi	172	I T	$(Rni2)^{2}$	94
00 07	Ebicultomusi	130	I	Rhi2 DL42	99
07	EOISUKOIIIUgi	173	L T	Rhi2	90
80	Hitsumikomugi	172	1 T	Rht2	104
89	Kokesnikomugi	135	I	Rht1 Rht2	77
90	Okukomugi	174	1	Rht2	115
91	Sakyukomugi	164	1	Rht2	96
92	Yutakakomugi	193	I	Rht2	92
93	Danchikomugi	177	I	Rht2	93
94 of	Furutsumasari	276	R	-	112
95 07	Shirasagikomugi	188	I	Rht1 ⁴	92
96	Junreikomugi	184	I	Rht1	89
9 7	Kitakamikomugi	169	I	Rht2°	101
98	Fujimikomugi	183	I	Rht1	94
99	Hayatokomugi	162	I	Rht2	95
100	Mikunikomugi	179	I	Rht1⁴	93

Table 3. Continued

Norin No.	Variety and breeding line	GRI ¹ (%)	GA reaction ²	Genotypes	Culm length (cm)
101	Shimofusakomugi	152	I	Rht2	88
102	Miyaginokomugi	151	Ι	Rht2	91
103	Nichirinkomugi	168	Ι	Rht2	89
104	Haruhikari	302	R	-	130
105	Ushiokomugi	173	I	Rht2	90
106	Omasekomugi	183	Ι	Rht2	84
107	Hiyokukomugi	196	I	Rht1	81
108	Mukakomugi	158	I	Rht2	112
109	Zenkojikomugi	181	I	Rht2	86
110	Kobushikomugi	188	I	Rht2	86
111	Haruminori	257	R	-	115
112	Sakigakekomugi	187	I	Rht2	83
113	Hachimankomugi	180	Ι	Rht2	85
114	Horoshirikomugi	176	I	$Rht2^5$	96
115	Takunekomugi	153	I	Rht2	88
116	Hanagasakomugi	171	Ι	Rht2	92
117	Shiroganekomugi	172	Ι	Rht1 ⁵	72
118	Gogatsukomugi	192	Ι	Rht1 ⁴	80
119	Toyohokomugi	178	Ι	Rht2	88
120	Setokomugi	199	Ι	Rht1 ⁵	85
121	Chikushikomugi	193	Ι	Rht1 ⁵	84
122	Shirowasekomugi	171	Ι	Rht1	73
123	Asakazekomugi	176	Ι	Rht1 ⁵	78
124	Fukuhokomugi	188	I	Rht1 ⁵	83
125	Minaminokomugi	177	Ι	Rht1	84
126	Chihokukomugi	176	Ι	Rht2 ⁵	86
127	Wakamatsukomugi	177	Ι	Rht2	84
128	Fukuwasekomugi	264	R	_	80
129	Nishikazekomugi	173	Ι	Rht1 ⁵	79
130	Haruyutaka	127	Ι	Rht1	103
131	Shiranekomugi	134	I	Rht2	86
132	Airakomugi	126	Ι	Rht2	. 85
133	Kovukikomugi	130	I	$(Rht2)^6$	92
	Chugoku 131	144	Ι	Rht1	79
	D.W. sel. No. 71	101	MI	Rht1 Rht2	65
	Kanto 100	180	Ι	Rht1	80
	Saitama 27	118	Ι	Rht1	98
	Saitama 29	144	Ι	Rht1	87
	Sanin 1	129	Ι	Rht2	95

¹GA response index: (Length of the first leaf sheath (LS) of GA-treated seedlings/LS of GA-untreated seedlings) \times 100. ²R: responsive, I: insensitive, MI: markedly insensitive.

³After Gale et al. (1981).

⁴Aberrant genotypes based on their pedigrees.

⁵Cited from Yamada (1989).

⁶Estimated from their pedigrees.

-: Genes not identified.





















Fig. 2. Pedigrees of Norin varieties.

The upper side of each cross combination is the female parent and the lower side is the male parent. Synonym: Nakanaga = Chunaga, Eshima = Ejima, Eshimashinriki = Ejimashinriki, Shinriki = Shinrikimugi = Shinrikikomugi.

on pedigree studies the Rht1 gene of 10 varieties, Norin 26, Norin 32, Norin 49, Norin 69, Junreikomugi, Fujimikomugi, Hiyokukomugi, Setokomugi, Chikushikomugi and Minaminokomugi, must have been derived from Saitama 29 (Fig. 2). The GRI value of Saitama 29 was 144. Out of the 10 descendants of Saitama 29 mentioned above, indeed the GRI values of the 6 varieties, Norin 26, Junreikomugi, Fujimikomugi, Hiyokukomugi, Setokomugi and Chikushikomugi, were above 180, and especially the GRI value of Setokomugi was 199 which was the highest value among the GRI values recorded in the GA-insensitive wheats. However, the GRI values of the other 4 descendants were not as high but comparable to those of the *Rht1* tester lines. Thus the results in this study did not necessarily provide consistent evidence that

the weak GA-insensitive allele occurred at the *Rht1* locus in the Japanese wheats tested.

According to Inazuka (1971), Norin 10 was developed from the cross between Turkey Red and Fultz Daruma. Fultz Daruma is a breeding line which was developed from the cross between Shirodaruma and Glassy Fultz. Shirodaruma and Glassy Fultz are considered to have been derivatives of Daruma and Fultz (GRI = 224), respectively (Fig. 2). The Rht genes of Norin 10 must have been derived from Fultz Daruma because the female parent Turkey Red was GA-responsive (GRI = 271) and a tall variety with a culm length of 139 cm. Although the existing Fultz Daruma was not GA-insensitive (GRI = 279) and moreover a tall variety with a culm length of 123 cm, Fultz Daruma which had been used as the parent of Norin 10 appeared to carry both Rht1 and Rht2

based on the photograph of Fultz Daruma and Norin 10 taken by Matsumoto (1968). The photograph shows that the culm of Fultz Daruma is clearly shorter than that of Norin 10. As shown in Table 2, Daruma carried only Rhtl and Shirodaruma carried only Rht2. Therefore one of the Norin 10 genes must have been derived from Glassy Fultz. The variety called Glassy Fultz was not tested in this study. However, Shou Fultz identified as an Rht2 carrier (Table 2) seems to be identical with Glassy Fultz because 'Shou' in Japanese corresponds to 'glassy' in English. If this assumption is valid, the following speculation can be made about the sources of the genes of Norin 10: The Rht1 gene of Norin 10 may not have been derived from the existing Shirodaruma but from the existing Daruma, the Rht2 gene of Norin 10 might have been derived from Glassy Fultz, and the Rht2 gene of Glassy Fultz might have arisen by an outcrossing or a spontaneous mutation. The latter phenomenon could actually occur. For example, the Rht10 gene of the dwarf mutant Aibian 1 (Wang et al., 1982) is located on the short arm of chromosome 4D (Sasakuma & Izumi, 1983), at the same locus as Rht2 (Börner & Mettin, 1988).

The culm lengths of most of the wheats carrying GA-insensitive Rht gene or genes were less than 105 cm (Tables 1, 2 and 3, Fig. 3). However, there were 7 Norin varieties, Norin 2, Norin 24, Norin 33, Norin 58, Mutsubenkei, Okukomugi and Mukakomugi, whose culm lengths exceeded 105 cm though all of them carried the Rht2 gene. Norin 33 and Mutsubenkei especially had tall culms comparable to that of the *rht* tester line Chinese Spring. These varieties may have a nullifier or modifier gene(s) or height promoting genes in the background (Gale & Youssefian, 1985) controlling the height-reducing effect of Rht2. On the other hand, there were 6 GA-responsive Norin varieties, Norin 42, Norin 43, Norin 45, Norin 46, Norin 47 and Fukuwasekomugi, that were as short as Akakomugi which carries the GA-responsive Rht genes, Rht8 and Rht9 (Worland et al., 1984). They therefore appear to carry GA-responsive dwarfing gene or genes. As all of them except for Norin 43 are not related to Akakomugi they may carry previously unidentified height reducing genes. In its pedigree



Fig. 3. Frequency distribution of culm length of every *Rht* genotype group containing *Rht* tester lines, Japanese Norin varieties, breeding lines and landraces.

Norin 43 is related to Akakomugi 3 which is probably a derivative of Akakomugi. Therefore Norin 43 may carry the same genes as Akakomugi.

The results of this study show that the Rht1 and Rht2 genes have been successfully used by Japanese wheat breeders unconsciously selecting for GA insensitive gneotypes. These two genes will certainly also be useful in the future. The increased knowledge of the GA insensitive genotype is obviously useful for the selection of parents, especially when wheat breeders are trying to incorporate some Rht gene or genes into their breeding materials or avoid major segregation for culm length. Gale & Gregory (1977) reported that in the case of the incorporation of the Norin 10 genes the breeding method combining the GA seedling response test with the single seed descent technique could remove most of the unwanted tall genotypes as F_2 seedlings by completely omitting the F_2 field sowing. Recent studies have indicated that the Norin 10 genes exhibit pleiotropic effects on other plant characters including grain quality, and that the phenotypic expression of those genes is affected by their interactions with the components of agricultural environments as well as by other major genes (Gale & Youssefian, 1985; Allan, 1989). Yamada (1989) reported that the geographical distribution of the Rht genotypes in the Japanese modern varieties was clearly localized suggesting that *Rht1* and *Rht2* genotypes have a different adaptability to agro-ecological environments. Such accumulated information on these genes will certainly be useful for the breeders in devising their strategies to develop high yielding wheat varieties with improved quality.

Chromosome designation

It was approved at the Seventh International Wheat Genetics Symposium that the chromosome designations of 4A and 4B were interchanged (Gale & Snape, 1988).

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