Plankton and hydrochemistry of Lake Futalaufquen (Patagonia, Argentina) during the growing season

L. Pizzolon¹, N. Santinelli², M. C. Marinone³ & S. A. Menu-Marque³

 ¹Laboratorio de Ecología Acuática, Universidad Nacional de la Patagonia, 9200 Esquel, Chubut, Argentina
²Facultad de Ciencias Naturales, Universidad Nacional de la Patagonia, 9100 Trelew, Chubut, Argentina
³Departamento de Ciencias Biológicas, Facultad de Ciencias Exactas y Naturales, Universidad de Buenos Aires, Pabellón II, Ciudad Universitaria, 1428 Buenos Aires, Argentina

Received 29 June 1993; in revised form 16 December 1994; accepted 21 February 1995

Key words: Patagonia, oligotrophy, nitrogen limitation, phytoplankton, zooplancton, spatial heterogeneity

Abstract

Plankton communities and hydrochemistry of an oligotrophic lake occupying a glacial valley in Argentinian Patagonia (42 °49'S; 71 °43'W) were studied. Monthly samples at three stations integrated from 0 to 50 m and stratified samples at the site of maximum depth, were taken during the growing season. Transparency was always controlled by glacial silt, and not by phytoplankton. Lake water belongs to the calcium-bicarbonate type, with low conductivity (24 μ S cm⁻¹), and poor buffering capacity. Forty-five phytoplankton taxa were found. Mean phytoplankton density was 49 cells ml⁻¹ and mean biomass 69 μ g l⁻¹. N:P relationships, inorganic nitrogen exhaustion in the photic layer, and correlations between nutrients and phytoplankton density suggests nitrogen as the main limiting factor. Fifteen zooplankton species were found. Mean zooplankton density was 12.2 ind. l⁻¹ and mean biomass 22.9 μ g l⁻¹. Diatoms and Boeckellidae were the dominant planktonic groups. Morphometry and hydrological factors were responsible for horizontal heterogeneity in phytoplankton and chemical variables.

Introduction

This contribution deals with the plankton assemblages of Futalaufquen Lake in relation with hydrological and chemical variables and watershed features. The lake has been sampled sporadically, as part of extensive surveys (Thomasson, 1963; INALI, 1972; José de Paggi & Paggi, 1985; Quirós, 1988b, 1989; Izaguirre *et al.*, 1990). Futalaufquen is a temperate, monomictic, and oligotrophic lake (INALI, 1972) that lies in a pristine area of the Patagonian Andes. A spatial pattern expressed as horizontal heterogeneity *sensu* Margalef (1979) may be hypothesized due to the lake's complex morphology. Present results, obtained during a period of low anthropic impact, could be useful as baseline for the assessment of future perturbations in this watershed.

Table	1.	Morphometrical	parameters	of
Futala	ufq	uen Lake.		

Altitude	518 m a.s.l.
Surface area	44.6 km ²
Mean depth *	101 m
Maximum depth *	168 m
Volume *	4509 km ³
Total watershed area	2920 km ²

* From Quiros et al. (1988a).

Study site

Located on the east slope of the Andes (42 °49'S; 71 °43'W), Futalaufquen Lake, occupies an intermediate position in the lake chain of the Futaleufu River watershed (Fig. 1). Morphometrical parameters are shown in Table 1. The main inflow is Arrayanes River, next in importance are the rivers Desaguadero and Cen-



Fig. 1. Bathymetric map of Futalaufquen Lake (modified from Quirós 1988a) and lake watershed.

tinela. The southwestern arm discharges into Krügger Lake (Fig. 1). Rainfall shows a marked decrease from the Andes (1990 mm y^{-1}) towards the steppe. Intrusive and extrusive igneous rocks, composed by acid and intermediate materials, dominate the watershed

lithology (Viera, pers. com.). The main human impacts are represented by 18 000 tourists each summer, cattle raising and lumbering. About 3000 inhabitants live in the watershed area.

Material and methods

Sampling sites I, II and III (Fig. 1), were monitored monthly from October 1988 to February 1989. Integrated samples for chemical and phytoplankton analyses were taken with a Van Dorn sampler by pooling water from 0, 15, 30 and 45 m. Stratified samples from 0, 50, 100 and 150 m for chemical analyses were taken at the deepest station (II). Main inflows were also sampled for chemical analyses. Transparency and surface temperature were measured with a Secchi disk and a mercury thermometer, respectively. Potentiometric total alkalinity and pH were measured with a VEGA V pH-meter, conductivity (25 °C) with an HORIBA U-27 water-checker, and dissolved oxygen using the Winkler method (A.P.H.A., 1978). Analyses were done within 24 hours after sampling. Unfiltered samples for nutrients and cations were frozen at -20 °C until analysis; samples for silicate-Si were kept at 4 °C. TIN (nitrate-N + nitrite-N) and SRP (soluble reactive phosphorus) were determined after Strickland & Parsons (1972). Silicate-Si was measured with a TECHNI-CON autoanalyzer (Technicon Instr., 1977). Calcium, magnesium, sodium and potassium were analyzed by atomic absorbance spectrophotometry. Hydrological data on daily water levels and the regression model to estimate water discharge were supplied by Agua y Energía Eléctrica (pers. com.).

Qualitative phytoplankton samples were taken with a 25 μ m mesh net by vertical hauls from 50 m to the surface and fixed *in situ* with 5% formaldehyde. Quantitative samples were preserved with acetic lugol. Cell counting was done following the simple aliquot method (Semina, 1975). Between 100 and 200 cells of the most frequent species were counted to achieve a relative precision of 25–30% (Venrick, 1975). Diatoms were processed after Hasle & Syvertsen (1980). Cell biomass was estimated after Trevisan (1978) by measuring 15–20 individuals of each species.

Monthly zooplankton samples were obtained at II. Samples were collected by vertical hauls from 50 m to the surface, using a 75 μ m mesh conical net, and preserved *in situ* with 5% formaldehyde. The number of subsamples for density estimations was determined according to Cassie (1971) (error under 10%). Macrozooplankton was subsampled with a Russell sampler and counted in 5 ml Bogorov chambers. Microzooplankton (rotifers and nauplii) was subsampled with a Hensen-Stempel pipette and counted in 1 ml Sedgwick-Rafter chambers. Thirty to 50 individuals of each category were measured. Crustacean dry weight



Fig. 2. Water discharge through Frey River and Secchi disk transparency of Futalaufquen Lake.



Fig. 3. Spring and summer vertical profiles of nitrate-N at station II in Futalaufquen Lake.

was estimated according to Dumont *et al.* (1975) and Bottrell *et al.* (1976), and that of rotifers according to Ruttner-Kolisko (1977).



Fig. 4. Spring and summer sequence of the main phytoplankton groups at samplings sites I, II and III.

Results

Hydrology and water chemistry

Maximum discharge took place during the thaw and correlated exponentially with transparency (r = -0.91;

p<0.05) (Fig. 2). The euphotic zone estimated from the Secchi disk transparency (Lemoalle, 1981) varied from 30 to 50 m. Surface water temperature rose from 4.4 °C up to 16 °C from October to February.

A theoretical retention time (T_w) of 0.94 y⁻¹ for the whole lake was estimated using historical records (Agua y Energía Eléctrica, 1983). However, the northern branch has 0.48 km³ while the southern branch has a volume of 2.14 km³. The first receives 80% of the lake inflow from Arrayanes River. Therefore, T_w of northern and southern branches differs markedly, being 0.11 and 4.5 y⁻¹ respectively.

Average and standard deviations of chemical variables at I, II and III, are shown in Table 2. Mean conductivity was 24 μ S cm⁻¹; an exceptional value of 56 μ S cm⁻¹ at III, in January, was coincident with high values of Ca, Mg and alkalinity. pH values were always below seven. Cationic relationships (equivalents) were Ca>Mg=Na>K. Silicate-Si showed similar concentrations at all stations, with a maximum of $4.44 \text{ mg } l^{-1}$ (station III, January). Vertical profiles of pH, alkalinity, conductivity, silicate-Si and cations were orthograde or close to that type. Oxygen profiles at II always showed good oxygenation even at the bottom. From October to December, oxygen saturation oscillated around 90%. The minimum saturation (86%) was detected at a depth of 150 m, in November. Total inorganic nitrogen was similar at all sampling sites. All TIN vertical profiles (Fig. 3) were negative clinogrades. SRP vertical profiles were irregular. Molar N:P relationship (TIN/SRP) showed a mean ratio of 0.65 in the euphotic zone, being lowest at III. The rivers Arrayanes, Desaguadero and Centinela presented similar ratios (0.39, 0.58 and 0.59, respectively).

Phytoplankton

Mean phytoplankton density was 50 cells ml⁻¹ (range: 15–155 cells ml⁻¹), with a mean biomass of 69 μ g l⁻¹ (range: 16–261 μ g l⁻¹). Forty-five phytoplankton taxa were found (Table 3), with a mean of 18 taxa per sample. Four taxa contributed 74% of the total biomass, *Synedra nana*, *Peridinium* spp., *Staurastrum tetracerum* and *Oocystis marssonii*.

Bacillariophyceae accounted for the highest relative cell density (41%), followed by Chlorophyceae (35%), Cryptophyceae (10%), Chrysophyceae (7%), and Xanthophyceae (0.5%). The five most frequent species at each station are shown in Table 4. Thirtytwo, 39 and 36 taxa were found at I, II and III, respectively. The spring maximum took place in Octo-

		X S	D	х	SD	х	SD
Lake station		I		11		III	
pН		6.69-0.27		6.75-0	.21	6.7	1-0.17
Alkalinity	$(\text{meq } l^{-1})$ 0.26-0.005		5	0.25-0.03		0.29-0.15	
Conductivity	$(\mu S \ cm^{-1})$	23.80-2.90)	23.60-1.82		29.00-15.3	
TIN	$(\mu g l^{-1})$	1.40-1.07	,	1.68-0.96		1.48-1.94	
SRP	$(\mu g l^{-1})$	4.45-2.92		4.76-2.58		7.16-3.32	
SiO3-Si	$(mg l^{-1})$	3.41-0.10		3.43-0.13		3.54-0.52	
Ca	$(mg l^{-1})$	3.52-1.22		3.82-1	.18	6.54	4-4.14
Mg	$(mg l^{-1})$	0.85-0.31		0.94-0.41		0.90-0.44	
Na	$(mg l^{-1})$	1.48-0.08		1.39-0.17		1.48-0.22	
K	$(mg l^{-1})$	0.42-0.06		0.42-0.11		0.39-0.04	
Rivers		Arrayanes		Desagua	adero	Centi	inela
Rivers pH		Arrayanes 6.72-0.37		Desagua 7.32-0	adero	Centi 6.83	inela 3-0.28
Rivers pH Alkalinity	(meq 1 ⁻¹)	Arrayanes 6.72-0.37 0.26-0.03	,	Desagua 7.32-0 0.55-0	adero .33 .01	Centi 6.83 0.40	inela 3-0.28)-0.02
Rivers pH Alkalinity Conductivity	(meq l^{-1}) (μ S cm ⁻¹)	Arrayanes 6.72-0.37 0.26-0.03 21.50-2.69	,	Desagua 7.32-0 0.55-0 55.50-8	adero .33 .01 .50	Centi 6.83 0.40 59.00	nela 3-0.28)-0.02)-21.0
Rivers pH Alkalinity Conductivity TIN	(meq l^{-1}) (μ S cm ⁻¹) (μ g l^{-1})	Arrayanes 6.72-0.37 0.26-0.03 21.50-2.69 0.77-0.61	,	Desagua 7.32-0 0.55-0 55.50-8 2.80-0	adero .33 .01 .50 .56	Centi 6.83 0.40 59.00 1.35	anela 3-0.28 0-0.02 0-21.0 5-1.14
Rivers pH Alkalinity Conductivity TIN SRP	(meq l^{-1}) (μ S cm ⁻¹) (μ g l^{-1}) (μ g l^{-1})	Arrayanes 6.72-0.37 0.26-0.03 21.50-2.69 0.77-0.61 5.83-2.12	,	Desagua 7.32-0 0.55-0 55.50-8 2.80-0 15.35-7	adero .33 .01 .50 .56 .65	Centi 6.83 0.40 59.00 1.35 23.77	inela 3-0.28)-0.02)-21.0 5-1.14 7-21.1
Rivers pH Alkalinity Conductivity TIN SRP SiO ₃ -Si	$(\text{meq } l^{-1})$ $(\mu \text{S cm}^{-1})$ $(\mu \text{g } l^{-1})$ $(\mu \text{g } l^{-1})$ $(\text{mg } l^{-1})$	Arrayanes 6.72-0.37 0.26-0.03 21.50-2.69 0.77-0.61 5.83-2.12 3.15-0.09	, , ,	Desagua 7.32-0 0.55-0 55.50-8 2.80-0 15.35-7 4.40-0	adero .33 .01 .50 .56 .65 .19	Centi 6.83 0.40 59.00 1.35 23.77 5.12	anela 3-0.28 0-0.02 0-21.0 5-1.14 7-21.1 2-0.25
Rivers pH Alkalinity Conductivity TIN SRP SiO ₃ -Si Ca	$(\text{meq } 1^{-1})$ $(\mu \text{S cm}^{-1})$ $(\mu \text{g } 1^{-1})$ $(\mu \text{g } 1^{-1})$ $(\text{mg } 1^{-1})$ $(\text{mg } 1^{-1})$	Arrayanes 6.72-0.37 0.26-0.03 21.50-2.69 0.77-0.61 5.83-2.12 3.15-0.09 3.50-1.49		Desagua 7.32-0 0.55-0 55.50-8 2.80-0 15.35-7 4.40-0 4.80-1	adero .33 .01 .50 .56 .65 .19 .53	Centi 6.83 0.40 59.00 1.35 23.77 5.12 3.91	inela 3-0.28)-0.02)-21.0 5-1.14 7-21.1 2-0.25 I-1.10
Rivers pH Alkalinity Conductivity TIN SRP SiO ₃ -Si Ca Mg	$(meq l^{-1})$ $(\mu S cm^{-1})$ $(\mu g l^{-1})$ $(\mu g l^{-1})$ $(mg l^{-1})$ $(mg l^{-1})$ $(mg l^{-1})$	Arrayanes 6.72-0.37 0.26-0.03 21.50-2.69 0.77-0.61 5.83-2.12 3.15-0.09 3.50-1.49 0.76-0.30		Desagua 7.32-0 0.55-0 55.50-8 2.80-0 15.35-7 4.40-0 4.80-1 1.22-0.	adero 33 01 .50 .56 .65 .19 .53 .09	Centi 6.83 0.40 59.00 1.35 23.77 5.12 3.91 0.98	inela 3-0.28 0-0.02 0-21.0 5-1.14 7-21.1 2-0.25 1-1.10 3-0.28
Rivers pH Alkalinity Conductivity TIN SRP SiO ₃ -Si Ca Mg Na	$(meq l^{-1})$ $(\mu S cm^{-1})$ $(\mu g l^{-1})$ $(\mu g l^{-1})$ $(mg l^{-1})$ $(mg l^{-1})$ $(mg l^{-1})$ $(mg l^{-1})$	Arrayanes 6.72-0.37 0.26-0.03 21.50-2.69 0.77-0.61 5.83-2.12 3.15-0.09 3.50-1.49 0.76-0.30 1.23-0.11		Desagua 7.32-0. 0.55-0. 55.50-8 2.80-0 15.35-7. 4.40-0 4.80-1. 1.22-0. 2.64-0	adero 33 .01 .50 .56 .65 .19 .53 .09 .67	Centi 6.83 0.40 59.00 1.35 23.77 5.12 3.91 0.98 2.83	inela 3-0.28)-0.02)-21.0 5-1.14 7-21.1 2-0.25 1-1.10 3-0.28 3-0.46
Rivers pH Alkalinity Conductivity TIN SRP SiO ₃ -Si Ca Mg Na K	$(meq l^{-1}) (\mu S cm^{-1}) (\mu g l^{-1}) (mg l^{-1}) $	Arrayanes 6.72-0.37 0.26-0.03 21.50-2.69 0.77-0.61 5.83-2.12 3.15-0.09 3.50-1.49 0.76-0.30 1.23-0.11 0.36-0.06		Desagua 7.32-0 0.55-0 55.50-8 2.80-0 15.35-7 4.40-0 4.80-1 1.22-0 2.64-0 0.51-0	adero 33 .01 .50 .56 .65 .19 .53 .09 .67 .13	Centi 6.83 0.40 59.00 1.35 23.77 5.12 3.91 0.98 2.83 0.40	inela 3-0.28 0-0.02 0-21.0 5-1.14 7-21.1 2-0.25 1-1.10 3-0.28 3-0.46 0-0.09

Table 2. Seasonal mean (X) and standard deviation (SD) values of physical and chemical variables measured in the euphotic zone of sampling sites I, II, and III in Futalaufquen Lake and its main inflows.



Fig. 5. Relationships between phytoplankton density and nutrients in Futalaufquen Lake from October 1988 to February 1989.

	CHLOROPHYCEAE			
BACILLARIOPHYCEAE	Chlamydomonas spp.			
Asterionella formosa Hass.	Coccomyxa lacustris (Chod.) Pasch. 1915			
Ceratoneis arcus Kutzing	Coenocystis subcylindrica Kors 1953			
Cyclotella stelligera Cl. u. Grun.	Cosmarium sp.			
Cymbella affinis Kutzing	Cruciginiella lunaris Lemn			
Cymbella cistula (Hemprich) Grun.	Flakatathrir genevensis (Reverd) Hindak			
Epitemia sorex Kutzing	Fudaring alegans Ehr 1832			
Fragilaria sp.	Decemetractum SD			
Gomphonema olivaceum (Lyngbye) Kutzing	Distrigenhaarium simplex Korshikoff			
Melosira sp.	Norman antique line stimute (C.M. Smith) C.M. Smith			
Navicula spp.	Approcytian anneucum (C.M. Shith) C.M. Shith			
Nitzchia acicularis W. Smith	Docysus marssonii Leiniii. 1878			
Nitzchia frustulum (Kutzing) Grun.	Paulschulzia pseudovolvox (Schulz.) Skuja			
Nitzchia pseudoamphyoxis Husted	Pandorina smithu Chodat			
Rhizosolenia eriensis H.L. Smith	Sphaerocystis schroeteri Chodat			
Synedra nana Meister	Staurastrum tetracerum Ralts			
Synedra ulna (Nitzch.) Ehr.	Staurodesmus triangularis v. subparalellus			
	(G.M. Smith) Teiling			
CHRYSOPHYCEAE	CYANOPHYCEAE			
Dinobryon divergens Imhof.	Anabaena sp.			
Dinobryon sociale Ehrenberg	Aphanocapsa elachista v. conferta West et West			
Malomonas sp.	Coelosphaerium kutzingianum Nageli			
cyst of Chrysophyceae	Oscillatoria sp.			

Table 3. Phytoplankton taxa identified during the growing season in Futalaufquen Lake.

CRYPTOPHYCEAE

Chroomonas sp.

Rhodomonas lacustris Pascher et Ruttner

DYNOPHYCEAE

Peridinium spp.

XANTOPHYECEAE

Gloeochloris sp.

ber except in I (Fig. 4). S. nana and A. formosa codominated at III while Coccomyxa lacustris and O. marssonii codominated at II. From November onwards no relevant differences were observed among sampling sites. The Shannon-Weaver diversity index was minimum in October $(-1.69 \text{ bits ind}^{-1})$ and maximum in November (4.5 bits ind^{-1}). Mean diversity

for all samples was 3.34 bits ind- $^{-1}$, without signifi-

cant differences among sampling sites. Phytoplankton



Fig. 6. Temporal sequence of density and biomass of the main zooplankton groups at station II in Futalaufquen Lake. Rotifer biomass is almost undetectable at this scale.

biomass was well correlated with phytoplankton density (r = 0.63; n = 15). Variance of phytoplankton density was explained (50.4%) by TIN and not by SRP (Fig. 5a and 5b).

Zooplankton

Fifteen zooplankton species were recorded: 3 cladocerans, 4 copepods and 8 rotifers (Table 6). Boeckella michaelseni was always dominant both in density and biomass (Fig. 6). Mesocyclops araucanus and Parabotreas sarsi were the only predator species, representing on average a very low proportion of the whole community. Cladocerans were uniformly scarce (Fig. 6). The most important species were Daphnia middendorffiana (sensu Paggi, 1973) in November and December, and Bosmina chilensis during the rest of the season. Conochilus unicornis was the dominant rotifer



Fig. 7. Relationships between phytoplankton and zooplankton total biomass in Futalaufquen Lake during the growing season.

in all sampling dates, probably owing to its colonial habit. Mean density was 12.2 ind 1^{-1} , with a biomass of 22.9 μ g 1^{-1} . The community composition showed a great constancy, with smooth fluctuations in density and biomass (Fig. 6). Total species number varied between 8 and 13, and crustaceans between 5 and 7. Zooplankton minima were recorded in November and December, while maxima corresponded to early spring and midsummer. The summer density peak was mainly determined by rotifers. Mean macrozooplankton size (762 μ m) tracked the changes in *Daphnia* density. Phytoplankton biomass explained 52% of zooplankton biomass variance (r=0.72; p<0.05) (Fig. 7).

Discussion

Futalaufquen Lake waters are calcium-bicarbonate type, with low conductivity and poor buffering capacity. This is explained by the watershed geochemistry, dominated by igneous rocks, a feature shared with other lakes and rivers of the Patagonian Andes (Campos, 1984; Pedrozo *et al.*, 1993). Cationic relationships and absolute values were the same found during the summer of 1984 (Quiros, 1989). Maxima of alkalinity, conductivity, and Ca found in January at III, may be due to rain and land washout four days before sampling. Concentrations of SRP, Ca and K were higher at III. Variances were also particularly high at III (Table 2), indicating that it was the most unstable station. This finding confirms the riverine characteristics of the northern branch, also evidenced by the short

Station I	%	Station II	%	Station III	%
R. eriensis	11.9	C. lacustris	32.6	S. nana	30.8
D. divergens	11.9	O. marssonii	9.6	A. formosa	8.4
R. lacustris	10.5	R. eriensis	8.2	S. schroeteri	7.9
Chlamydomonas sp.	8.6	R. lacustris	6.7	R. eriensis	7.5
S. schroeteri	8.1	A. formosa	5.1	R. lacustris	5.8

Table 4. Relative frequency (%) of the five most frequent phytoplankton species at sampling sites I, II and III in Futalaufquen Lake.

Table 5. Zooplankton species identified during the growing season in Futalaufquen Lake.

CLADOCERA

Bosmina chilensis Daday, 1902

Bosmina longirostris (O.F. Muller, 1785)

Daphnia middendorffiana Fischer, 1851

COPEPODA

Calanoida

Boeckella michaelseni (Mrazek, 1901)

Parabroteas sarsi (Daday, 1901)

Cyclopoida

Mesocyclops araucanus Löffler, 1961

Tropocyclops prasinus v. meridionalis (Kiefer, 1931)

ROTIFERA

Bdelloidea

Collotheca mutabilis (Hudson, 1885)

Collotheca pelagica (Rousselet, 1893)

Conochilus unicornis (Rousselet, 1892)

Euchlanis dilatata Ehrenberg, 1832

Keratella thomassoni Thomasson, 1957

Polyarthra vulgaris Carlin, 1943

Synchaeta pectinata Ehrenberg, 1832

retention time (40 days). The chemical composition of the main inflow (Table 2) was determinant of lake water chemistry.

We have found several evidences of N-limitation. The spring phytoplankton maximum seems to decay due to nitrate-N exhaustion in photic layers, as shown by TIN vertical profiles (Fig. 3). This is a common feature shared by many lakes on the western slope of the Andes (Campos, 1984). On the other hand, the N:P molar ratios of all lake arms and the main inflows, were always below 1 after October, being 1.4 the maximum value observed. These values are far from the colimitation range of 11-26.5 (Thayer, 1974; Fricker, 1980; Ram & Plotkin, 1983), considering N:P as (NO₃ + NO₂ + NH₄)-N/SRP. Ammonia-N was undetectable in a preliminary survey of the Lake. The maximum N:P ratio was 1.4, being always lower than unit after October. Nostoc sp. colonies (2-4 cm diameter), a biological indicator of N-limitation (Margalef, 1983), were recorded at the bottom of coastal waters. Regression-correlation analysis (Figs 5a, 5b) also highlights the importance of nitrogen instead of phosphorus in explaining phytoplankton density variance. All approaches used suggest nitrogen as the main limiting factor in Futalaufquen Lake. This feature could also be shared by upstream lakes of the watershed, as suggest the low N:P ratio of the Arrayanes River (0.39). However, Quirós (1989) using a different methodology, found in a single survey at the end of summer a TN:TP ratio of 242, indicating P-limitation. Phytoplankton communities evolve in a non-equilibrium dynamic (Harris, 1986; Sommer, 1989b) and therefore, could be limited by more than one factor at the same time (Dodds et al., 1989), or by different factors along the year (Pizzolon, unpublished data; Pedrozo et al., 1993; Diaz et al., 1994). Despite the worldwide extend of phosphorus limitation in temperate lakes proposed by Schindler (1978), recent data on Patagonia

Table 6. Mean seasonal density (ind l^{-1}) and biomass ($\mu g \ l^{-1}$) for the main zooplankton groups at Station II in Futalaufquen Lake. Calanoida and Cyclopoida include adults and copepodites.

	Calanoida	Cyclopoida	nauplii	Cladocera	Rotifera
Density	4.22	0.48	3.60	0.30	3.62
(range)	1.68-7.17	0.03-0.88	1.37-6.66	0.07-0.57	0.72-9.27
Biomass	16.02	2.72	1.13	2.92	0.16
(range)	7.0-27.82	0.24-5.87	0.41-2.05	0.55-7.45	0.04-0.42

(Pedrozo *et al.*, 1993; Soto *et al.*, 1994; Diaz & Pedrozo, 1994) and the present results, suggest the existence of a greater number of N-limited lakes than expected, in temperate South America.

Silicate-Si concentration was high, although lower than in Chilean and New Zealand lakes (Campos, 1984; Duthie & Stout, 1986). Silicate-Si vertical profiles were always orthograde and its concentration did not change over time. Correlation with diatoms was not significant. Compared with published data (Werner, 1977; Tilman *et al.*, 1982; Sommer, 1989b; Diaz, 1994), the values recorded in Futalaufquen Lake do not seem to be limiting for diatoms growth.

The most surprising feature of phytoplankton succession was the simultaneous dominance in October of the large diatom Synedra nana (3127 μ m³) at III, and of the small Chlorophyceae Coccomyxa lacustris $(20.7 \ \mu m^3)$ at II. This event does not agree with the general theory of phytoplankton succession, which states a sequence that begins in spring with a bloom of small cells that grow fast in turbulent waters (Sommer, 1989a). Margalef (1983) reported Synedra acus as a frequent species in potamoseston, while Duthie & Stout (1986) also found the suspension of heavy cells without floating mechanisms in a New Zealand lake exposed to strong winds. The northern branch is characterized by winds frequently over 18 m s⁻¹, and a retention time typical of rivers. Both are synergic factors of turbulence, which can explain the dominance of S. nana at III. We cannot explain the dominance of the chlorophyceans C. lacustris and O. marssonii at II in October instead of the small diatoms expected in this time of the year (Tilman et al., 1982; Harris, 1986; Sommer, 1989a; Diaz et al., 1994).

Decreased water transparency after the spring maximum was explained by glacial silt inflow, as shown by the highly significant correlation between water discharge and transparency (Fig. 2), and not by phytoplankton growth. Phytoplankton biovolume has proved to be an element of lake eutrophication assessment at least as valuable as chlorophyll *a* (Rott, 1984). Maximum (0.26 mg l^{-1}) and mean phytoplankton biomass (0.069 mg l^{-1}) allow us to consider this lake as oligotrophic (Fricker, 1980; Rott, 1984; Margalef, 1983).

The zooplankters present in the lake are widely distributed in Andean Patagonia, where calanoid copepods are dominant, cladocerans scarce and diversity low (Soto & Zúñiga, 1991; Marinone & Menu-Marque, unpublished data). Crustacean density and biomass in February (8.6 ind 1^{-1} or 22.8 μ g 1^{-1}) were in good agreement with records of March 1984 (13.9 ind l^{-1} or 27.5 μ g l^{-1}) by Marinone & Menu-Marque (unpublished data). Biomass values were comparable to those published for oligotrophic, deep, stratified lakes of Europe (Herzig, 1979). Mean macrozooplankton biomass (21.7 μ g l⁻¹) is among the lowest levels recorded in the world's literature (Ivanova, 1987). Spring and summer zooplankton maxima have been recorded in Chilean Araucanian lakes (Campos, 1984; Campos et al., 1987), when the community experiences strong changes (Zúñiga & Domínguez, 1978). Other coincidences with these lakes are the dominance of centropagid copepods over cladocerans, which indicates oligotrophy (Campos et al., 1987), and the parallel oscillations of phyto and zooplankton which would suggest that there is no top-down control of phytoplankton (Campos, 1984). The presence of a large cladoceran such as Daphnia denotes low predation pressure by larval or planktivorous fish, while the reduction in Daphnia size in January could be related to the fish recruitment season. Chilean lakes seem to have higher zooplankton densities and higher proportions of cladocerans and cyclopoids (Domnguez & Zúñiga, 1979) than lakes east of the Andes (Marinone & Menu-Marque, unpublished). These features are probably related to the lower elevation and higher temperature of Chilean lakes, that render them slightly

more productive than Argentine lakes (Quirós & Drago, 1985). Sporadic records on phyto and zooplankton (Thomasson, 1963; INALI, 1972; José de Paggi & Paggi, 1985) indicate that the dominant species remain the same, thus failing to show water quality changes during the last decades.

The hypothesis on lake horizontal heterogeneity was supported by chemical and phytoplankton community results. Phytoplankton periodicity in these lakes could be influenced by the rainfall regime, as suggested by Thomasson (1963). Not only vertical mixing is important to explain phytoplankton succession, but also horizontal transport from the watershed. Pulses of nutrients can reach the lake through Arrayanes River, generating a spatial pattern expressed as horizontal heterogeneity (Margalef, 1979). A gradient of perturbations, sensu Margalef (1975), decreasing from the northern branch to the southern one can be hypothesized and partially evidenced in the present study. The northern branch is the most directly exposed to chemical and biological perturbations induced by Arrayanes River, while the southern branch is the most protected. On the other hand, during hydrologically stable periods the differences among stations would be minimum.

Acknowledgments

We would like to thank Ing. Miriam Solis and the Centro Nacional Patagónico, Mr Roberto Cerdá and the Administración de Parques Nacionales, Lic. Viviana Sastre, Dra Mónica Díaz, Dr Oscar Romero, Lic. Alicia Moretto, and Mr Esteban Nuñez. This study was the first part of the project 'Limnology of the Futaleufú River watershed (Chubut, Argentina)', supported by the grants PIA N 0610/87 from CONICET, and PI N 056/88 from the Consejo de Investigaciones de la Universidad Nacional de la Patagonia. We greatly appreciate the critical revision of the manuscript by Prof Dr Heinz Löffler.

References

- American Public Health Association, 1980. Standard methods for the examination of water and wastewater. 15th ed. A.P.H.A., N.Y., 1134 pp.
- Agua y Energía Eléctrica, División Recursos Hídricos, 1983. Estadística hidrológica hasta 1983. I: Fluviometría. Red Hidrometeorológica. Ministerio de Obras y Servicios Publicos de la Nación. Buenos Aires, 230 pp.

- Bottrell, H. H., A. Duncan, Z. M. Gliwicz, E. Grigierek, A. Herzig, A. Hillbricht-Ilkowska, H. Kurasawa, P. Larsson & T. Weglenska, 1976. A review of some problems in zooplankton production studies. Norw. J. Zool. 24: 419–456.
- Campos, H., 1984. Limnological studies of Araucanian lakes. Verh. int. Ver. Limnol. 22: 1319–1327.
- Campos, H., W. Steffen, O. Parra, P. Domínguez & G. Agüero, 1987. Estudios limnológicos en el lago Caburgua (Chile). Gayana (Botánica) 44: 61–84.
- Cassie, R. M., 1971. Sampling and statistics. In Edmondson, W. T. & G. G. Winberg (eds), A manual on methods for the assessment of secondary productivity in fresh waters. Blackwell Scientific Publications, Oxford, IBP Handbook 17: 174–209.
- Dfaz, M. & F. L. Pedrozo, 1994. Nutrient limitation in Andean-Patagonian lakes at latitude 40–41 °S. Submitted to Arch. Hydrobiol, 35 pp.
- Dodds, W. K, K. R. Johnson & J. C. Priscu, 1989. Simultáneous nitrogen and phosphorus deficiency in natural phytoplankton assemblages: theory, empirical evidence, and implications for lake management. Lake Reserv. Mgmt 5: 21-26.
- Domínguez, P. & L. Zúñiga, 1979. Perspectiva temporal de la entomostraca fauna limnética del Lago Ranco (Valdivia, Chile). Anales Mus. Hist. Nat. Valparaíso 12: 53-58.
- Dumont, H. J., I. Van de Velde & S. Dumont, 1975. The dry weight estimate of biomass in a selection of Cladocera, Copepoda and Rotifera from the plankton, periphyton and benthos of continental waters. Oecologia 19: 75–97.
- Duthie, H. C. & V. M. Stout, 1986. Phytoplankton periodicity of the Waitaki Lakes, New Zealand. Hydrobiologia 138 (Dev. Hydrobiol. 33): 221–236.
- Fricker, H., 1980. OECD eutrophication programme. Regional Project. Alpine Lakes. Swiss Federal Institute Water Resources and Water Pollution Control & OECD. Dübendorf, 234 pp.
- Harris, G. P., 1986. Phytoplankton ecology. Structure, function and fluctuation. Chapmann & Hall, N.Y., 384 pp.
- Hasle, F. & E. E. Syvertsen, 1980. The diatom genus *Ceratulina*: Morphology and taxonomy. Bacillaria 3: 79–113.
- Herzig, A., 1979. The zooplankton of the open lake. In Löffler, H. (ed.), Neusiedlersee: The limnology of a shallow lake in Central Europe. Monogr. Biol. 37: 281–335.
- INALI, 1972. Patagonian Lakes. In: Instituto Nacional de Limnología. Report on IBP/PF projects. CONICET, Santa Fe, Argentina, 56 pp.
- Ivanova, M. B., 1987. Relationships between zooplankton development and environmental conditions in different types of lakes in the zone of temperate climate. Int. Revue ges. Hydrobiol. 72: 669–684.
- Izaguirre, I., P. Del Giorgio, I. O'Farrel & G. Tell, 1990. Clasificación de 20 cuerpos de agua andino-patagónicos (Argentina) en base a la estructura del fitoplancton estival. Cryptogamie, Algol. 11: 31–46.
- José de Paggi, S. J. & J. C. Paggi, 1985. Zooplancton de los cuerpos de agua preexistentes en el área del Embalse Amutui Quimei (Cuenca del Río Futaleufú). Neotropica 31: 119–131.
- Lemoalle, J., 1981. Photosynthetic production and phytoplankton in the euphotic zone of some African and temperate lakes. Revue Hydrobiol. trop. 14: 31–37.
- Margalef, R., 1975. External factors and ecosystem stability. Schweiz. Z. Hydrol. 37: 102–117.
- Margalef, R., 1979. The organization of space. Oikos 33: 152-159.
- Margalef, R., 1983. Limnología. Omega, Barcelona, 1010 pp.
- Paggi, J. C., 1973. Contribución al conocimiento de la fauna de cladóceros dulceacuícolas argentinos. Physis 32: 105-114.

- Pedrozo, F., S. Chillrud, P. Temporetti & M. Diaz, 1993. Chemical composition and nutrient limitation in rivers and lakes of northern Patagonian Andes (39.5–42 ° S; 71 ° W) (Rep. Argentina). Verh. int. Ver. Limnol. 25: 207–214.
- Quirós, R., 1988a. Mapas batimétricos y parámetros morfométricos de Lagos Patagónicos del Neuquén, de Río Negro y del Chubut (Argentina). Informe Técnico INIDEP, Dep. Aguas Continentales 5: 1–8.
- Quirós, R., 1988b. Relationships between air temperature, depth, nutrients and chlorophyll in 103 Argentinian lakes. Verh. int. Ver. Limnol. 23: 647-658.
- Quirós, R., 1989. Relaciones entre niveles de pigmentos fotosintéticos y diversos factores ambientales en ambientes acuáticos de la República Argentina. PhD Thesis, Facultad de Ciencias Exactas y Naturales, Universidad de Buenos Aires, 258 pp.
- Ram, N. M. & S. Plotkin, 1983. Assessing aquatic productivity in the Housatonic River using the algal assay: bottle test. Wat. Res. 17: 1095-1106.
- Rott, E., 1984. Phytoplankton as a biological parameter for the trophic characterization of lakes. Verh. int. Ver. Limnol. 22: 1078– 1085.
- Ruttner-Kolisko, A., 1977. Suggestions for biomass calculation of planktonic rotifers. Ergebn. Limnol. 8: 71–76.
- Schindler, D. W., 1978. Factors regulating phytoplankton production and standing crop in the World's freshwaters. Limnol. Oceanogr. 23: 478–486.
- Semina, H. J., 1978. The size of cells. In: A. Sournia (ed.), Phytoplankton manual. UNESCO, Paris: 233-237.
- Sommer, U., 1989a. Preface. In U. Sommer (ed.), Plankton Ecology. Succession in plankton communities. Springer: 1–8.

- Sommer, U., 1989b. The role of competition for resources in phytoplankton succession. In U. Sommer (ed.), Plankton Ecology. Succession in plankton communities. Springer: 57–106.
- Soto, D. & L. Zúñiga, 1991. Zooplankton assemblages of Chilean temperate lakes: a comparison with North American counterparts. Rev. Chilena de Hist. Nat. 64: 569–81.
- Soto, D., H. Campos, W. Steffen, G. Agüero, O. Parra & L. Zúñiga. 1994. The Torres del Paine lake district (Chilean Patagonia): A case of potentially N-limited lakes and ponds. Arch. Hydrobiol. (Supp.) 99: 181–197.
- Strickland, J. D. H. & T. R. Parsons, 1968. A practical handbook of seawater analyses. Bull. Fish. Res. Bd Can.: 167.
- Thayer, G. W., 1974. Identity and regulation of nutrients limiting phytoplankton production in the shallow estuaries near Beaufort, N.C. Oecologica 14: 75–92.
- Thomasson, K., 1963. Araucanian Lakes. Acta Phytogeogr. Suecica 47: 1–139.
- Tilman, D., S. Kilham & P. Kilham, 1982. Phytoplankton community ecology: The role of limiting nutrients. Annu. Rev. Ecol. Syst. 13: 349–372.
- Trevisan, R., 1978. Nota sull'uso dei volumi algali per la stima della biomasa. Riv. Idrobiol. 17: 345–357.
- Venrick, E. L., 1975. Estimating cell numbers. In A. Sournia (ed.), Phytoplankton manual, UNESCO, Paris: 176–180.
- Werner, D., 1977. The Biology of Diatoms. Blackwell Scientific Publications, Oxford: 309–310.
- Zúñiga, L. R. & P. T. Domínguez, 1978. Entomostracos planctónicos del Lago Riñihue (Valdivia, Chile): distribución temporal de la taxocenosis. Anal. Mus. Hist. Nat. Valparaíso 11: 89–95.