

Update section

Short communication

Sequence analysis of a cDNA encoding a Group 3 LEA mRNA inducible by ABA or dehydration stress in wheat

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Abstract

A cDNA clone (pMA2005) of a Group 3 LEA (late embryogenesis abundant) protein has been sequenced from wheat. The wheat cDNA clone codes for a protein with ten tandem repeats of an 11 amino acid sequence and has homology to other Group 3 LEAs reported in barley, carrot, cotton and rape (L. Dure *et al.*, *Plant Mol Biol* 12: 475–486, 1989). The deduced amino acid sequence indicates that the wheat protein has a molecular weight of 23 000 and is a basic, hydrophilic protein. Northern analysis with the cDNA clone shows that dehydration of wheat shoot tissue results in increased transcript levels that correlate with increases in endogenous ABA.

Introduction

LEA mRNAs and proteins accumulate in embryonic tissue during seed maturation [4]. LEA mRNAs and proteins can also be induced by the application of abscisic acid (ABA) or by tissue

dehydration. A protective role for these proteins during plant tissue dehydration that could be mediated by ABA has been proposed [2, 5]. LEA proteins deduced from several higher plant cDNA and/or genomic DNA sequences have been compared and fall into three groups based on sequence homologies [5]. In wheat a cDNA clone encoding a Group 1 LEA protein, Em, has been characterized [7]. We report here the sequence analysis of cDNA clone pMA2005 that encodes a Group 3 LEA mRNA from wheat and induction of the mRNA in dehydration-stressed shoot tissue. The deduced protein sequence of the pMA2005 clone contains tandemly repeated 11

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*The nucleotide sequence data reported will appear in the EMBL, GenBank and DDBJ Nucleotide Sequence Databases under the accession number X56882.

amino acid stretches similar to that reported for other Group 3 LEA proteins from barley, carrot, cotton and rape [5].

Materials and methods

A wheat embryonic axes cDNA library, constructed in Lambda Zap (Stratagene, La Jolla, CA) was screened and the cDNA clone pMA2005 selected by differential screening as previously described [8]. Both strands of the

pMA2005 cDNA were sequenced in their entirety using a deletion series (Erase-a Base System, Promega) and dideoxy chain termination (Sequenase Version 2.0 DNA Sequencing Kit, United States Biochemical Corporation) of double-stranded DNA templates. The sequencing data were analyzed using the UWGCG Sequence Analysis software package (version 6.2) [3] with the assistance of the VADMS Center, Washington State University. Poly(A)⁺ RNA was obtained from isolated wheat embryonic axes or from shoot tissue [1]. Northern analysis was con-

- 24 CAGGTCGTGTTCCAAGAAACCAAA

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1  ATG GCC TCC AAC CAG AAC CAG GCC AGC TAC CAC GCC GGC GAG ACC AAG GCC CGC AAC GAG
   Met Ala Ser Asn Gln Asn Gln Ala Ser Tyr His Ala Gly Glu Thr Lys Ala Arg Asn Glu

61  GAG AAG ACC GGG CAG GTG ATG GGC GCG ACC AAG GAC AAG GCG GGG CAG ACC ACG GAG GCC
   Glu Lys Thr Gly Gln Val Met Gly Ala Thr Lys Asp Lys Ala Gly Gln Thr Thr Glu Ala
   [----- 1 -----] [-----]

121 ACG AAG CAG AAG GCC GGC GAG ACC ACG GAG GCC ACG AAG CAG AAG GCC GCC GAG ACC ACG
    Thr Lys Gln Lys Ala Gly Glu Thr Thr Glu Ala Thr Lys Gln Lys Ala Ala Glu Thr Thr
    [----- 2 -----] [----- 3 -----] [-----]

181  GAG GCG GCC AAG CAG AAG GCC AGC GAG ACG GCC GAG GCC ACC AAG CAG AAG GCC GCC GAG
   Glu Ala Ala Lys Gln Lys Ala Ser Glu Thr Ala Glu Ala Thr Lys Gln Lys Ala Ala Glu
   [----- 4 -----] [----- 5 -----]

241  GCC AAG GAC AAG ACG GCG CAG ACG GCG CAG GCG GCC AAG GAG AAG ACG TAC GAG ACG GCG
   Ala Lys Asp Lys Thr Ala Gln Thr Ala Gln Ala Ala Lys Glu Lys Thr Tyr Glu Thr Ala
   [.....] [----- 6 -----] [-----]

301  CAG TCG GCC AAG GAG GCG GCC GCC CAG GGC AAG GAC CAG ACC GCC AGC ACC CTC GGC GAG
   Gln Ser Ala Lys Glu Arg Ala Ala Gln Gly Lys Asp Gln Thr Ala Ser Thr Leu Gly Glu
   [----- 7 -----]

361  AAG ACG GAG GCG GCC AAG CAG AAG GCC GCC GAG ACC ACG GAG GCG GCC AGG CAG AAG GCC
   Lys Thr Glu Ala Ala Lys Gln Lys Ala Ala Glu Thr Thr Glu Ala Ala Arg Gln Lys Ala
   [----- 8 -----] [----- 9 -----]

421  GCC GAG GCG ACC GAG GCG GCC AAG CAG AAG GCG TCG GAG ACG GCG CAG TAC ACC AAG GAG
   Ala Glu Ala Thr Glu Ala Ala Lys Gln Lys Ala Ser Glu Thr Ala Gln Tyr Thr Lys Glu
   [----- 10 -----]

481  TCC GCC GTC ACC GGC AAG GAC AAG ACC GGC AGC GTC CTC CAG CAG GCA GGG GAG ACG GTG
   Ser Ala Val Thr Gly Lys Asp Lys Thr Gly Ser Val Leu Gln Gln Ala Gly Glu Thr Val

541  GTG AAC GCC GTG GTG GGC GCC AAG GAC GCC GTG GCC AAC ACG CTG GGC ATG GGC GGG GAC
   Val Asn Ala Val Val Gly Ala Lys Asp Ala Val Ala Asn Thr Leu Gly Met Gly Gly Asp

601  AAC ACC ATC ACC ACC AAG GAC AAC ACC ACC GGC GCC ACC ACC AAG GAC ACC ACC ACC ACC
   Asn Thr Ile Thr Thr Lys Asp Asn Thr Thr Gly Ala Thr Thr Lys Asp Thr Thr Thr

661  ACC AGG AAT CAC TAGACGCATGTTGGCGCTTAATTTGCTTTTGTAGTGTGTTGGTCGTTCCGGGCGCTT
   Thr Arg Asn His

736  CTACATATTTGTATGTTCCACTCTTTTCGTGATTTCAGCTCATTGGTGTAAGTTTGCCTTCGATTGTACTCC

815  TCGTGTCCGGTCTGTATTAGTACGAGTTATGGTCCATATACTTTGGTGTAAGTGGATATCGAGGACACTCGAAGGCGG

894  CAATAAAGTGAATTTC 910

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Fig. 1. Nucleotide sequence of pMA2005. Dideoxy sequencing data of a 934 bp cDNA encoding a Group 3 LEA protein in wheat is shown along with the deduced amino acid sequence. The ten tandem repeats of 11 amino acid tracts are bracketed, underlined and numbered. The truncated version of the repeat is shown underdotted and the unrelated 11 amino acid stretch is located between repeats 7 and 8. A probable polyadenylation signal, AATAAA, is underlined, though a poly(A) tail was not present in the clone.

ducted as previously described [8]. ABA was extracted from shoot tissue and measured by immunoassay using a monoclonal antibody to (+)ABA as previously described [9].

Results and discussion

The cDNA clone pMA2005 was obtained from a cDNA library prepared from ABA-treated embryonic axes of wheat (*Triticum aestivum*) cv. Brevor [8]. The cDNA clone was selected by differentially screening with cDNA prepared from dormant and nondormant axes [8]. In Fig. 1 the nucleotide sequence and deduced amino acid sequence of pMA2005 are shown. The sequenced cDNA has a length of 934 bp, which includes an open reading frame encoding a protein of 224 amino acids plus 5' and 3' flanking regions. The deduced protein sequence of the wheat clone shows homology to the Group 3 LEA proteins. Highest homology was found to the barley clone pHVA1 [6] (91% at the nucleotide level, 95% at the amino acid level). The repeating 11 amino acid tract, typical of Group 3 LEA proteins, is repeated 10 times in the wheat clone (Fig. 1) and 9 times in barley [6]. Both the wheat (Fig. 1) and barley [6] clones contain an identical truncated version of the repeating unit that consists of only 7 of the 11 amino acids. Further, for both the wheat and barley clones the repeating sequences are interrupted by one 11 amino acid stretch that has no homology with the repeating sequence. This unrelated stretch in the wheat clone (Fig. 1) is identical to that of barley [6] except for two amino acid substitutions. Analysis of the entire amino acid sequence shows a hydrophilic, basic polypeptide with a molecular weight of 23 000.

Northern analysis with clone pMA2005 has shown that gene expression is high in embryos from mature wheat seed and that this message can be enhanced by incubation of embryos in ABA [8]. To test whether the message accumulates in shoot tissue when ABA levels are elevated by water stress, we measured transcript levels in dehydrated shoot tissue. As shown in Fig. 2A, dehydration of wheat shoots also induces high

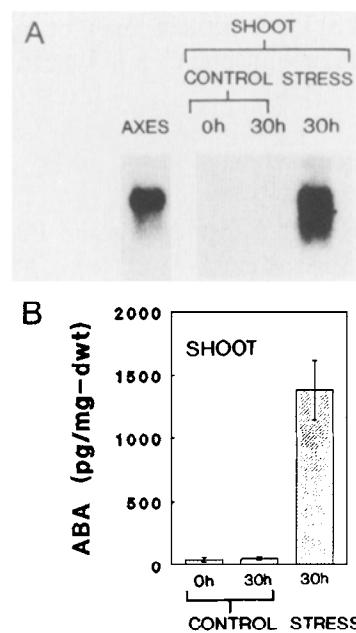


Fig. 2. Northern analysis of the wheat Group 3 LEA transcript and ABA measurement in wheat. A. Two-day old wheat seedlings were slowly dehydrated by placement in open petri dishes over 25% glycerol (v/v) in a desiccator and incubated at 22 °C for 30 hours. Control seedlings were placed on moistened blotter paper and incubated under the same conditions. Poly(A)⁺ RNA (0.5 μg) obtained from isolated mature wheat embryonic axes (non-stressed) or from the shoot tissue was separated by formaldehyde-agarose gel electrophoresis and transferred to a nylon membrane. Northern analysis conducted with the pMA2005 cDNA revealed transcripts of approximately 1 kb. B. ABA levels in control tissue at 0 and 30 hours and in shoot tissue dehydrated for 30 hours. ABA was measured by immunoassay [9].

levels of pMA2005 mRNA compared to hydrated controls. Induction of the wheat clone transcript correlates with endogenous increases in ABA in dehydrated shoot tissue (Fig. 2B). An antibody has been produced against a fusion protein developed from the cDNA sequence of pMA2005. Upon dehydration a protein that is strongly cross-reactive with the antibody accumulates in wheat shoots. The protein is not present in unstressed shoots (J.L. Ried and M. Walker-Simmons, unpublished data). While the function of the pMA2005 protein product is not yet certain, the induction of gene expression for this highly hydrophilic protein in dehydrated tissue suggests that

this Group 3 LEA protein may be part of a drought tolerance mechanism in wheat.

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References

1. Badley JE, Bishop GA, St. John T, Frelinger JA: A simple, rapid method for the purification of poly (A)⁺ RNA. *Biotechniques* 6: 114–116 (1988).
2. Close TJ, Kortt AA, Chandler PM: A cDNA-based comparison of dehydration-induced proteins (dehydrins) in barley and corn. *Plant Mol Biol* 13: 95–108 (1989).
3. Devereux J, Haerberli P: A comprehensive set of sequence analysis programs for the VAX. *Nucl Acids Res* 12: 387–395 (1984).
4. Dure L III, Greenway SC, Galau GA: Developmental biochemistry of cottonseed embryogenesis and germination. XIV. Changing mRNA populations as shown by *in vitro* and *in vivo* protein synthesis. *Biochemistry* 20: 4162–4168 (1981).
5. Dure L III, Crouch M, Harada J, Ho THD, Mundy J, Quatrano R, Thomas T, Sung ZR: Common amino acid sequence domains among the LEA proteins of higher plants. *Plant Mol Biol* 12: 475–486 (1989).
6. Hong BM, Uknes SJ, Ho THD: Cloning and characterization of a cDNA encoding a mRNA rapidly induced by ABA in barley aleurone layers. *Plant Mol Biol* 11: 495–506 (1988).
7. Litts JC, Colwell GW, Chakerian RL, Quatrano RS: The nucleotide sequence of a cDNA clone encoding the wheat Em protein. *Nucl Acids Res* 15: 3607–3618 (1987).
8. Morris CF, Anderberg RJ, Goldmark PJ, Walker-Simmon MK: Molecular cloning and expression of abscisic acid-responsive genes in embryos of dormant wheat seeds. *Plant Physiol*, 95: 814–821 (1991).
9. Walker-Simmons M, Kudrna DA, Warner RL: Reduced accumulation of ABA during water stress in a molybdenum cofactor mutant of barley. *Plant Physiol* 90: 728–733 (1988).