

Update section

Short communication

Molecular cloning of the gene encoding developing seed L-asparaginase from *Lupinus angustifolius*

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Abstract

A genomic sequence encoding *Lupinus angustifolius* L-asparaginase has been obtained, and is the first report of this gene from a plant source. The 3.2 kb of DNA sequenced contains a 1136 bp 5' flanking sequence, four exons and three introns. Intron-exon borders were mapped by comparing the genomic sequence with that of a *L. arboreus* cDNA. Primer extension analysis revealed transcription start sites 16 bp and 13 bp 5' of the initiating ATG for *L. angustifolius* and *L. arboreus*, respectively. The 5' flanking region contained sequences associated with seed-specific expression.

In temperate legumes, of which lupin is an example, asparagine is the major compound formed from the assimilation of ammonia and plays a principal role in the transport and metabolism of nitrogen [11]. Asparagine is the major nitrogen compound translocated from the nodules in lupin and accounts for 50 to 70% of nitrogen supplied to lupin seeds [1]. During fruit maturation greater than 80% of asparagine entering the fruit is metabolised. The enzyme L-asparaginase hydrolyses the amide group of asparagine to produce aspartate and ammonia and thus provides a route

for the utilisation of asparagine for the synthesis of amino acids and proteins. Indeed a range of studies have supported the view that asparaginase is a key enzyme involved in asparagine utilisation in lupin and plants in general [2, 18]. In previous work the developing seed L-asparaginase was purified and polyclonal antibodies raised against the protein. These antibodies precipitated asparaginase activity from a partially purified lupin seed extract. Partial protein sequence was obtained from polypeptides identified as asparaginase [12]. A *Lupinus arboreus* developing cDNA

The nucleotide sequence data reported will appear in the EMBL, GenBank and DDBJ Nucleotide Sequence Databases under the accession number X60691.

-1136 ACTAGTGAGTGGCTTGGAAATTCATATAAACATCCAGATCTAAATGCCACCTTTGTATTATTAATAAACTAGTAAATTTGG
-1056 GTGAATATTGATATAGCTCCAAATTTATGTTACTATAAACTCCTGGATATAGTAAAGTTTTTTTCTGAGGGTAAAGTTAT
-976 CTCTATGTTGTCTTCATTGTTGAAAGTACCTAATGGGGTCCCCTATTTTGCATAAATAACATTGTATATAAAATTTGGCA
-896 AAATGGAACCAACAAAACAATAGTAGTGTTCAGCAATATCAATAAAATAGAATCTAAAATTTAACTCTCAATCAATCAG
-816 ATTTAGTAGCCACCCTTATGAAGTCATCAAACCTGAATCTAGTGGGGTTCACATACATAGGGATGCCACAGATGAATGT
-736 CATTAATTTTGTCTTTACCTTATGTGATATAATAATGCATATATATATTTAACATGATATATAATATATTACTATTT
-656 TTATACATATTTTTAATATTCAAACACACTTACAATGAATAATAAAAGATGTACCATTTTATTTGGTATAACTACGAAAAA
-576 TATTCCTCACGAAAGATAAAGTCAGCGTGTTCGTTGATGAAAGGATAAGCCAATGACATGTAAAGTAGATAGAGATTGA
-496 GTTTTAATGTATTCAATACTCGAAGGATAATAACTCACACTTGAAGGAGAGATGTTGGTATTCCAATGTGAATGAAGA
-416 AGTCTCACATTATGCACAAGTGAATGAGTTAAAGAGCATGTAGTATATAATAAGATTTTGGTCAAGATGTAGTGTCAAAC
-336 TTCTTCTATATGTCTTTCACTTAGGTCGTAGAAGAATGTTACACCCCATGAATGAAAATATTCTAACGTTACGAAATATTA
-256 TTTTAAGGAAAACTAATCTTTGAAAATATGTAGCTTATACGAATCAAATATTTTCTTACCCAATAGATCAATACTATA
-176 TCATCTACTAAAATCAATAACCAAAATTAATAATCCCAATCAAAAACCTCACGTGACCAAAAAACCACAAATGAAAAAAGTGA
-96 GTGAAGTATGAGTCACCAAGATCTAAAATTTGAACTGTAATCGCTAAAATTTGCGGTGTCTATTATAAATATACAACACTAGT
-16 ATTCATTCCGTGAGTT
1 TTGAGTTGGAAGAAAAATGGGTGGTGGAGTATAGCTCTGCACGGCGCGCTGGCGACATTCCA
M G G W S I A L H G G A G D I P
65 TTTTCACTGCCACCGGAGCGCGTCCAGCCTCGGGAAGAAGCCTCCGCCACTGCCTCCAAATCGGCTTGAAGCTCTCAA
F S L P P E R R Q P R E E G L R H C L Q I G V E A L K
145 ATCCCAAAAGCCTCCTTTGGACGTTGTAGAAGTGTGtgtagtactatctatctctctctctctctctctctctctctctct
S Q K P P L D V V E L V
225 ttcaatgctgaagctatatatatatatatatatatataataataataataataataataataataatagGTTTCGTGAGTT
V R E L
305 AGAGAATATTCAACATTTCAATGCGGAATAGGATCTGTGTTAACGAATAGTGGAACAGTGGAAATGGAAGCATCAATAA
E N I Q H F N A G I G S V L T N S G T V E M E A S I
385 TGGATGGAAAACTATGAAATGTGGAGCAGTTTCTGGTCTGAGTACAGTTCGAATCCAATTTCACTAGCTCGATTAGTT
M D G K T M K C G A V S G L S T V L N P I S L A R L V
465 ATGGATAAACTCCTCATATATATCTTGCTTTCCAAGGAGCTCAAGATTTTGCTAAACAACAagtaatccttttactctt
M D K T P H I L A F Q G A Q D F A K Q Q
545 tatcatcctttatcttataacatttacatttaactctcactcaactctctctgttactgtatttgacactttacatggt
625 tattttatataatcttttaataatctttgtatataatgctctttacatgagaaaaatgatgaccatattgtatccaattt
705 ttagtagataaacttctctcctcaggactaagaatatacactgtcaatttcaagatagtgagtggtgtgtggttattatg
785 ggtgccatccactagatataatattataatatacactgactaattatagctttcacctaggaatcaatctcaatagctca
865 cttgagtcacaggatcacatgttccgacctaaacttccaagcctatctttttattacctttgttttttaaaaaatattat
945 tttgctcatgtgtcaataagtaattgatttagaacagagaatgagacacgggtttgactttgacaaaacaacaaagtaaa
1025 acatcattggtaaaaaactaaaaacattttgccacctgaactgctttgctggtgttacttttggttccaaactgaacagt
1105 cttgtgcttcatcttttaataatgaaatccctatctttccagGTTGTTGAGACTGTAGATTCAAGTCAATTTTACTGCAGAAA
G V E T V D S S H F I T A E
1185 ATGTTGAAAGGCTAAAGCTGGCAATAGAAGCCAATAGGGTCCAGGtatttttaattttgcatgatgaacaaaaagtagca
N V E R L K L A I E A N R V Q
1265 catttatgttacattgacactcaatctcctcaaatctcatttttttctcctctatgatgtcgattacattagttcttccac
1345 aatgtatagGTTGATTATAGTCAATATAAATATCCCAACCTGCTCAAGATGATGCTGAGAAGGAATACCACCTGCAA
V D Y S Q Y N Y P Q P A Q D D A E K E L P L A
1425 ATGGTGATAGTCAAATGGAAGTGTGGGTGTGTAGCTGTTGATAGCCATGGAAATCTAGCTTCTGCAACACTGCTGGT
N G D S Q I G T V G C V A V D S H G N L A S A T S T G
1505 GGATTAGTTAACAAAATGGTTGGTTCGAATCGGTGACACGCCCTCATAGGTGCCGGAACCTTATGCGAATGAACTTTGTGC
G L V N K M V G R I G D T P L I G A G T Y A N E L C A
1585 AGTTTCTGCAACAGGTAAGGTGAAGCAATAATCTGCGACGGTAGCAAGGGATGTGGTGCACACTCATGGAGTTCAAAG
V S A T G K G E A I I S A T V A R D V A A L M E F K
1665 GCCTTTCTCAAGGAAGCTGCTGATATGTTGTACATGAGCGTACACCAAAAGGCACCTGTTGGTTTGTATTGCTGTGCT
G L S L K E A A D Y V V H E R T P K G T V G A G L I A V S
1745 GCTGCAGGAGAAATTTGCCATGCCTTTAATACAACAGGCATGTTTCAGAGCATCTGCTACTGAGATGGCTATTTCAGAGAT
A A G E I A M P F N T T G M F R A S A T E D G Y S E I
1825 TGCAATTTGCCTACTACCTAAAATAAATGATTGTTTTGAAAGCTGTTCTGTTTTCCATGCTTCCATGTATTCTGACAT
A I W P T T *
1905 TTTTAAATGCTTCATGCATGGTTATATCATATCATAAGGAATAAGAATATTATCGAATTC

Fig. 1. Nucleotide sequence of the gene encoding L-asparaginase from *Lupinus angustifolius*. The 5' flanking sequence is italicised. Coding region from the cap site is in upper case and introns are in lower case.

library was screened with polyclonal antibodies and a positive clone sequenced which, when back-translated, showed homology with se-

quenced asparaginase polypeptides [13]. This paper is the first report of the cloning of a plant gene encoding a L-asparaginase enzyme activity.

Genes encoding L-asparaginase have, however, been cloned from bacterial [6] and yeast [10] sources. The gene from *Lupinus arboreus* is known to be expressed in the developing seed [12] and northern hybridisation experiments have shown maximum mRNA levels 30 days after anthesis [13]. Other seed-specific and embryo-specific genes have been cloned including B-conglycinin [3], rice lectin [21] and pea convicilin [14]. These are all seed storage proteins, whereas L-asparaginase is an enzyme activity implicated in the amino acid metabolism of the developing seed. It has been suggested [15] that embryo development relies on the seed coat for the supply of amino acids.

DNA was prepared from nuclei obtained from the radicles of germinating seeds of *Lupinus angustifolius* [L] var. Uniharvest [5], the DNA partially digested with *Sau* 3aI and fragments of mean size 15 kb to 20 kb ligated into the lambda vector EMBL3 and packaged [8]. This library was screened by plaque hybridisation using a *Lupinus arboreus* developing seed L-asparaginase cDNA probe [13]. Four positive clones were identified from a screen of 250000 and one of these, pPMB100, was used in the sequencing experiments. DNA was sequenced by the dideoxy chain termination method [17] and the sequence assembled using the computer programs of Stockwell [20]. Intron-exon borders were mapped by comparison with the *L. arboreus* cDNA sequence [13]. A 39 bp oligonucleotide, complementary to the RNA, was synthesized and this primer was annealed to and extended against RNA templates [9] from *L. arboreus* and *L. angustifolius* developing seed, and from *L. angustifolius* leaf, nodule and root. The primer corresponds to a position between 86 and 125 bp from the initiating ATG, and was chosen as there was absolute homology between the *L. angustifolius* genomic sequence and that of the *L. arboreus* cDNA clone [13] at this position. *L. arboreus* RNA gave a product 142 bp long, consistent with a 5' untranslated region of 13 bp and *L. angustifolius* RNA gave a major product at 145 bp, consistent with a 5' untranslated region of 16 bp (data not shown). An S1 nuclease protection [14]

experiment confirmed 5' untranslated regions of 13 bp and 16 bp, respectively for *L. arboreus* and *L. angustifolius* transcripts. Figure 1 shows the DNA sequence of a 3.1 kb *Eco* RI fragment obtained from pPMB 100. There is 94% homology between the *L. arboreus* cDNA sequence and the *L. angustifolius* genomic sequence. The gene consists of a 1136 bp 5' flanking sequence, exon 1 of 180 bp, intron 1 of 112 bp, exon 2 of 234 bp, intron 2 of 615 bp, exon 3 of 87 bp, intron 3 of 126 bp and exon 4 of 492 bp (to the TAA stop codon). Intron-exon boundaries followed the general exon/GT..intron..AG/exon rule. Two consensus poly(A) addition signals were observed in the 3' untranslated region.

Peptide sequence data obtained from purified *L. arboreus* asparaginase protein [12] was aligned against the genomic data. The first peptide bridged the gap between the 5' terminus of the cDNA and the presumed initiating methionine residue of the full-length coding region. The other seven peptides were aligned with other regions of the gene, except for exon 3. Several elements previously reported to be associated with seed- or embryo-specific expression are present in the 5' flanking region of the L-asparaginase gene:

- A 6 bp repeat, A(AGC)CCCA, implicated in embryo-specific expression has been reported [3]; this 'ACCCCA' motif is present at position -292;
- Sequences called 'RY repeats' have been implicated in the regulation of seed lectin, legumin, vicilin and seed albumin genes [4], and such an element - AATGCATA - is present at -700;
- An (ATC)AACACA(AC)(ATC) consensus sequence is present in the 5' region of many seed protein genes [7], and this motif is present at -632.

These three motifs are present in the L-asparaginase 5' flanking region at positions considerably more distant to the cap site than in previous reports and, furthermore, they occur at only one position in the sequence.

As well as containing sequences associated with seed- and/or embryo-specific expression an AAAGAT motif associated with legume nodule/root expression [19] was seen at -610, -563 and -365. The significance of this observation is

not known. A number of other repeats of unknown function were also observed. Of particular interest is the sequence CTAAAATT which is repeated three times at -49, -77 and -166. The significance of this motif remains to be elucidated and is the subject of ongoing investigation, particularly as it is the only significant feature of the sequence identified (aside from the TATA and CAAT motifs) in the first 200 nucleotides 5' from the cap site.

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