

# Chapter 12

## Arbuscular Mycorrhizal Fungi in Organic Versus Conventional Farming



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**Abstract** Arbuscular mycorrhizal fungi (AMF) establish symbiotic relationships with plant roots, enhancing nutrient uptake and promoting plant growth. This chapter discusses various factors that influence AM fungal populations under different farming systems. These factors include soil management routines such as tillage intensity, pesticide usage, fertilizer application, and crop rotation strategies. The aim is to compare the effects of organic farming practices that prioritizes sustainable approaches with those employed in conventional agriculture. The chapter looks at how organic farming methods differ from conventional farming in terms of AM fungal communities, diversity, and functionality. It describes practical strategies for improving mycorrhizal associations through inoculation techniques or by implementing specific agronomic practices that promote a favorable environment for these beneficial microorganisms. Overall, this chapter illuminates the role of AMF in influencing soil health and plant productivity across various agricultural approaches. It emphasizes the importance of taking these symbiotic relationships into account when designing sustainable farming systems that maximize yields while minimizing environmental impacts.

**Keywords** AMF · Monoculture · Crop rotation · Soil management · Beneficial microorganism · Symbiosis

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## 12.1 Introduction

Frank (1885) first used the term “mykorhiza” (mycorrhiza) and noted that nutrients and water were transported to plant roots by endophytic hyphae through mutualistic symbiotic relationships (Frank 1885). Arbuscular mycorrhizal fungi (AMF), as one of the utmost earliest obligate symbioses, are considered the most important soil organisms for agroecosystem sustainability (Martin et al. 2017).

Mycorrhizal symbiosis benefits plants in several ways, including improving their access to immobile nutrients (Basu et al. 2018; Hodge and Fitter 2010; Javot et al. 2007), tolerance to abiotic stress (Aroca et al. 2007; Chourasiya et al. 2021), and protection against pathogens (Morris et al. 2019). Meanwhile, they can play a key role in soil aggregation (Rillig and Mummey 2006), nutrient cycling (Lanfranco et al. 2018), and soil stability (Morris et al. 2019) and have prodigious potential for improving agricultural sustainability (Basu et al. 2018; Bender et al. 2016; Lehmann et al. 2020). As a result of their implications to plant performance and soil health, they play a crucial role in sustaining ecosystem productivity (Castillo et al. 2016; Ma et al. 2021).

More importantly, extensive hyphae networks could be produced by AMF, thereby enhancing water and nutrient uptake by the roots (Smith and Smith 2011). Mycorrhizae can increase the effective root absorption area of the host plants by producing mycelium and forming a mycelial network. Mycorrhizae can also enhance the environmental adaptability and stress resistance of host plants to cope with constraint from an adverse environment. Additionally, AMF are more important in terms of their potential to improve plant growth and health in future climate change scenarios (Wahdan et al. 2021; Johnson et al. 2013). The significance of AMF in improving plant nutrition (especially phosphorus (P)) and improving plant stress tolerance to both biotic and abiotic stress in natural ecosystems or low-input organic agriculture has been widely recognized (Jeffries et al. 2003).

## 12.2 AMF and Agroecosystem

### 12.2.1 *Plant Growth and Productivity*

Although the “high investment and high production” mode of conventional agriculture in the past has significantly increased the yield of crops, with the development of modern agriculture, the “low investment and high production” mode of intensive agriculture is becoming a key strategy of sustainable agricultural development (Shen et al. 2013). To meet the increasing demands of global food production, conserve environmental quality, and respond to more hostile environmental changes, agriculture management is more complex than ever, and simultaneously accomplishing high nutrient utilization efficiency and increasing crop yields have appeared key challenges (Cassman et al. 2003).

AMF provide multiple advantages to plant hosts because they are in direct contact with crop roots (Zhang et al. 2017). AMF are not only important for improving plant resistance to stress and sustaining biogeochemical cycling and ecological functions (Wu et al. 2018) but also for increasing crop yields (Cavagnaro et al. 2015; Rillig et al. 2016). Mounting evidences have suggested that mycorrhizal inoculation can promote the accumulation of grains and aboveground biomass (Ren et al. 2019; Zhao et al. 2015a, b), which being considered to be an effective agronomic measure (Pellegrino and Bedini 2014). The potential mechanism of increased crop yield after inoculation with AMF has received more attention (Bowles et al. 2016; Ren et al. 2019; Zhao et al. 2015). Several studies proved that AMF colonization promotes crop yields and can be a critical factor in making agroecosystems more sustainable (Bender et al. 2016; Thirkell et al. 2017). Recently, much discussion has been devoted to whether AM symbiosis is appropriate for agricultural production (Ryan and Graham 2018; Rillig et al. 2019). AMF symbionts can enhance the water absorption capacity of roots by producing hyphal structures (Zhao et al. 2015a, b) and increase host plant nutrient availability, directly contributing to increased crop yields (Hestrin et al. 2019). Besides, inoculation with AMF can boost the root biomass of host plants and thus produce more root exudates, which may facilitate microbial-driven nitrogen (N) mineralization, and further increase nutrient availability (Mo et al. 2020).

Despite numerous data on the benefits of inoculation of the host plants by AMF, the role of AMF on host plant yield is still under debate. It is known that the respiration rate is increased upon mycorrhization of plants that consumes part of the photosynthetic products (Mortimer et al. 2008). Thus, the enhanced carbon (C) assimilation observed in the mycorrhizal plants is most likely offset by higher respiratory consumption, and the crop yield did not change significantly (Xavier and Germida 2003). Some other studies have revealed that root colonization with AMF reduces crop productivity, crop biomass production, and yield (Berruti et al. 2016; Ryan and Graham 2018; Jayne and Quigley 2014). Divergent influences of AMF on crop productivity might be relevant to experimental conditions, AMF species identity, climate, soil moisture, and soil fertility (Gosling et al. 2016; Jayne and Quigley 2014; Roger et al. 2013; Thirkell et al. 2017). Therefore, further studies are required to elucidate how inoculation with AMF differs in plant production and physiological properties under various conditions.

### ***12.2.2 Quality of Crops***

AMF can enhance plant uptake of nutrients and promote plant health (Gonzalez-Dugo 2010). After colonization of plant roots with fungi, AMF can significantly promote the acquisition of soil mineral nutrients, especially P. The  $^{14}\text{C}$  tracking experiment found that the translocation of carbohydrates from plant to AM symbiotic structures (from root cells to AM mycelium) can promote the uptake of P by the fungi and its further transport to the plant (Bücking and Shachar-Hill 2005).

Additionally, inoculation of date palms with *Albahypha drummondii*, *Pervetustus simplex*, *Septoglomus xanthium*, *Claroideoglomus etunicatum*, *Rhizoglomus irregularis*, and *Funneliformis mosseae* resulted in a higher shoot length and stem diameter as compared with control plant (El Hilali et al. 2022). Inoculation of tomato (*Solanum lycopersicum* L.) with *Rhizophagus intraradices*, *R. irregularis*, *Funneliformis mosseae*, and *Glomus iranicus* resulted in a higher growth index and P content in AMF-treated plants as compared with the control group (Shafiei et al. 2022). Furthermore, the growth rate and consumption index of *Tuta absoluta* larvae feeding AMF tomato leaves were much lower than that on plants without AMF (Shafiei et al. 2022).

Moreover, AMF promote plant water uptake; increase water use efficiency, especially under drought stress conditions; and enhance plant drought resistance (Birhane et al. 2012). In wheat plants, AMF inoculation induces an increase in plant biomass accumulation under long-term growth conditions under drought stress (Al-Karaki et al. 2004).

### 12.2.3 AMF as Biocontrol Agent

AMF play a vital role in boosting the plant resistance to above- and/or belowground pests and diseases (Whipps et al. 2008). AMF-plant symbiosis has been revealed to profoundly alter plant primary and secondary metabolism, including the salicylic acid (SA) and jasmonate (JA) signaling pathways that are critical to plant defenses (Cameron et al. 2013). Plants that have been colonized by AMF prior to pest or pathogen attacks may be systemically primed through defense compound reallocation (Jung et al. 2012), thus allowing rapid and increased expression of defense genes compared with non-AMF plants (Song et al. 2015). AMF can enhance the ability of host plants against fungal, bacterial, viral, and nematode diseases. AMF can significantly inhibit the dispersal of root-knot nematodes and decrease gall numbers and the infection rate and the damage caused by nematodes (Lax et al. 2011). *Glomus mosseae* increased the resistance of different tomato varieties to *Fusarium*, but the extent of resistance was not consistent (Steinkellner et al. 2012). Cucumber, bent grass (*Agrostis stolonifera*), and tomato were inoculated with *Phoma* sp. GS8-2 and *Glomus mosseae*, respectively. *Glomus mosseae* reduced *Rhizoctonia*-caused cucumber root rot and *Fusarium oxysporum*-caused tomato root rot but aggravated bent grass brown spot (Saldajeno et al. 2012). Cosme et al. (2011) explored the effect of inoculation of *Glomus intraradices* on the oviposition of *Lissorhoptus oryzophilus* and found that this insect prefers to lay eggs on mycorrhizal plants. The possible explanation is that AMF colonization improves the root N and P contents; thus, the mycorrhizal plants with better growth were preferred to lay eggs; however, the harm of adults did not increase.

From the agricultural point of view, however, trade-offs that reduce yields may occur between AMF, crops, and pests. In addition to benefiting crop nutrition, AMF colonization also enhances the attraction, quantity characteristics, and quality

measures of plants against herbivores (Hartley and Gange 2009) and therefore improves capabilities of herbivores (Kempel et al. 2010). Moreover, phloem-feeding insects frequently outperform non-AM controls on AMF-colonized plants (Hartley and Gange 2009). AMF management strategies should consider these trade-offs and balance them accordingly.

#### ***12.2.4 Contribution of AMF to Soil Health***

Arbuscular mycorrhiza can directly affect the cycle of inorganic elements such as N, P, K, Ca, and Mg through chemical, physical, and biological activities and play an important role in the biogeochemical cycling of soil nutrients. The impact of arbuscular mycorrhiza on inorganic N is mainly through the hyphae absorbing N from the substrate and transferring it to the host plant and improving the rate of N fixation in N-fixing host plants by alleviating various stresses. The impact of AMF on promoting plant growth is closely associated with the improvement of plant P nutrition by mycorrhizal colonization. In soils with insufficient P supply, the inoculation of AMF can greatly improve the absorption and use of P by the symbiotic partner plants and alleviate the phenomenon of P deficiency in the rhizospheric zone (Recorbet et al. 2013).

AMF improve soil health through external hyphae too, thereby sustaining the constancy of the soil food web and increasing the stability of the soil structure (Finlay 2008), which provides numerous benefits to the host plants, such as defense against pathogens, enhanced salinity tolerance, decreased pH and heavy metals, and biofortification with trace elements in the crops (Ryan and Graham 2018). The huge mycelial network formed by the symbiosis of AMF and crops can extend to deeper and wider soils beyond the rhizosphere trophic zone of plants, thus improving the utilization of elements by plants (Lehmann et al. 2014). Besides facilitating host plant uptake of nutrients and water, AMF expand nutrient entrapment through their mycelial network, increasing soil aggregation through mycelium and glomalin, thereby improving soil nutrient storage and retention (Rillig and Mummey 2006) as well as reducing soil erosion and the risk of nutrient leaching.

#### ***12.2.5 Abiotic Stress Alleviation***

Abiotic stress (e.g., drought, high temperature, salinity, heavy metal stress, and unfavorable soil pH) leads to degradation of soil and poses a serious threat to agricultural production. Therefore, abiotic stress is considered to be one of the main reasons for crop yield reduction worldwide (Singh et al. 2011). AMF can enhance the tolerance of plants to abiotic stress. The extracellular hyphae of AMF can promote the uptake and consumption of nutrients and water by plants, and it is one of the critical factors in facilitating plants to avoid drought damage (Tyagi et al.

2017). Increased drought resistance and better crop performance can be due to the abundance of antioxidative enzymes (superoxide dismutase, catalase, peroxidase) and soluble sugars symbiotically produced by AMF (Huang et al. 2011). AMF also improve plant drought tolerance by improving soil structural stability. It has been reported that AMF hyphae have a rich filamentous hyphae network, which can enhance soil structure and also change soil structure by producing glycoproteins to form aggregates (Singh et al. 2013). Therefore, AMF symbiosis can enhance the drought tolerance of plants by enhancing the stability of soil structure, thereby increasing soil water-holding capacity (Ruiz-Lozano 2003).

The occurrence of AMF greatly enhanced the tolerance of plants to extreme temperature. Under the condition of high temperature stress, root colonization with AMF can enhance the activity of antioxidant enzymes and increase the content of soluble protein and proline in maize (*Zea mays* L.). Plants inoculated with AMF increased the activity of SOD, POD, and CAT by 50%, 40%, and 21%, respectively, indicating that AMF can resist high temperature stress damage by improving the antioxidant capacity of plants, thereby promoting plant growth (Mathur et al. 2018). In addition, low temperature or high temperature induces excessive ROS production under stress, causing oxidative stress response to damage plants. Usually, AMF can effectively promote the production of SOD, POD, CAT, and GR after colonizing host plants and relieve the damage caused by reactive oxygen species. Specifically, they reduce peroxidation of membrane lipids as well as membrane permeability and increase osmotic adjustment substance accumulation in response to temperature stress (Ahanger and Agarwal 2017; Hajiboland et al. 2019).

Under heavy metal stress, AMF change the growth pattern of plants by exploiting and exploring unstressed parts of the environment, alleviating the toxicity of heavy metals through various pathways (Gonzalez-Guerrero et al. 2008). The roots of AMF-inoculated plants can greatly contain heavy metals, which can inhibit the transport of heavy metals from the root system to the aboveground tolerance to adapt to heavy metal stress (Słomka et al. 2011; Zhang et al. 2010). However, mycelia have limited uptake potential of heavy metals, and in highly polluted places, the possibility of mycelium avoiding the hazards of toxic metals is very limited. Therefore, mycorrhizae have developed different strategies to resist heavy metal stress: AMF can secrete glomycin (Ferrol et al. 2009); glomycin participates in heavy metal inactivation by chelating heavy metals in soil. Audet and Charest (2007) revealed that AMF shift from “enhanced uptake” at soil with low concentrations of heavy metals to “metal binding” at soil with high concentrations of heavy metals. The remediation effect of AMF on heavy metals is also reflected in the ability to induce host plants to produce antioxidant enzymes, upregulate the expression of related peroxidase genes, and reduce the detrimental effects of ROS on plants under heavy metal stress (Rozpądek et al. 2014).

Salt stress can negatively affect plant photosynthetic capacity, enzyme activity, protein synthesis, and mineral nutrition and produce osmotic stress and ion stress on plants, thereby affecting the physiological and biochemical balance of plants (Hashem et al. 2016; Iqbal et al. 2015). Salt stress can promote ROS production, cause plant oxidative damage, and interfere with the normal metabolism of plants

(Akyol et al. 2020). AMF can improve the antioxidant capacity of host, reduce  $H_2O_2$  content, inhibit lipid peroxidation, and enhance the salt stress of plants by inducing plants to produce and accumulate osmotic adjustment substances such as glycine betaine, proline, and soluble sugar tolerance (Akyol et al. 2020; Porcel et al. 2015; Zhu 2003). Hidri et al. have found that AMF can improve nutrient (P) acquisition; reduce sodium absorption; enhance water absorption; and improve photosynthesis, chlorophyll content, and availability of antioxidant molecules under saline conditions. Hajiboland et al. (2010) reported that improved salt stress tolerance in tomato plants inoculated with AMF was associated with increased uptake of P, potassium (K), and calcium (Ca) and reduced sodium (Na) toxicity.

### **12.3 Response of Arbuscular Mycorrhizal Fungi to Agricultural Management Practice**

AMF develop symbiosis with most plant species and can assist the host plant in many ways, including better nutrient uptake, drought and salt tolerance, and disease resistance (Smith and Read 2008). These fungi are crucial to the preservation of the soil and the sustenance of plants. The development and survival of fungal propagules, including spores, hyphae, and colonized roots, are necessary for AM fungus to persist in various ecosystems. In the presence of host plants and no soil disturbance, hyphae are considered to be the primary source of inoculum, but spores are thought to be a relatively resistant fungal structure that may be viewed as “long-term” propagules when no viable host plants are present. For agricultural crops, there are management techniques that are widely acknowledged. Numerous agricultural management practices, such as soil tillage, fertilization, and plant protection techniques, have deleterious impact on mycorrhizal symbioses in agroecosystems (Säle et al. 2015; Jansa et al. 2002; Gryndler et al. 2006) (Fig. 12.1). These traditional management techniques can decrease soil biodiversity, especially AMF (Tsiafouli et al. 2015). The technique of organic agriculture, in contrast, can enhance soil quality in terms of boosting the diversity and biomass of soil microbiome (Mäder et al. 2002; Verbruggen et al. 2010) (Fig. 12.1). This section focuses on the discussion of the effects of agricultural management practices on AMF.

#### ***12.3.1 Agrochemical Management Practice***

Agrochemicals are chemical formulations used in industrial agriculture. It refers to chemical fertilizers and biocides, which are substances used for control of hazardous organisms. A biocide can be (1) pesticides including insecticides, herbicides, fungicides, and nematocides; (2) antimicrobials including antibacterials, antifungals, antiviral, antiprotozoals, and antiparasites; and (3) synthetic fertilizers. In addition to

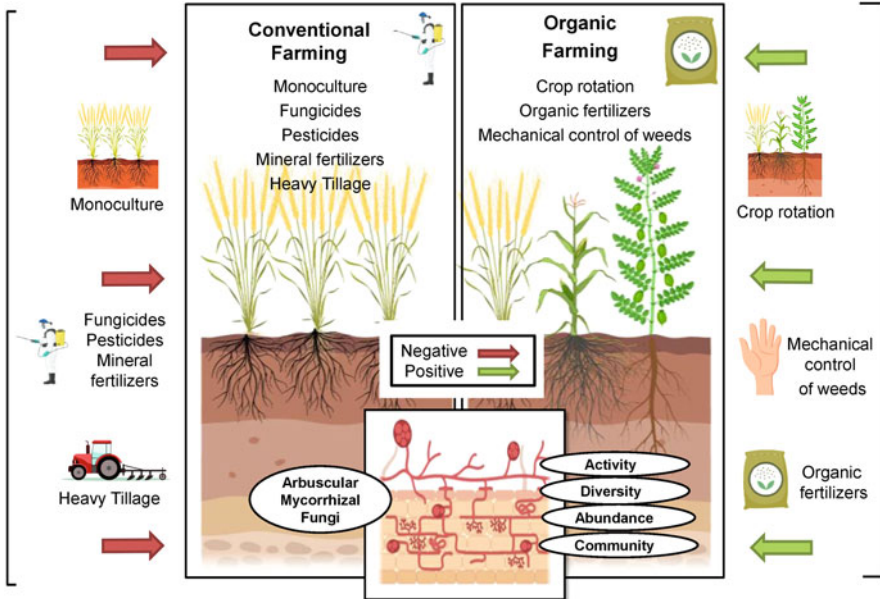


Fig. 12.1 Impact of different agricultural management practices on arbuscular mycorrhizal fungi

potentially contaminating the soil, the application of agrochemicals may have detrimental effects on AMF community (Kurle and Pflieger 1994; Abd-Alla et al. 2000; Lekberg and Koide 2005; Pasaribu et al. 2011; Mathimaran et al. 2007) leading to a decline in AMF spore diversity and abundance (Oehl et al. 2004).

### 12.3.1.1 Pesticides

Insecticides, fungicides, nematicides, and herbicides are the four major categories of pesticides. Pesticides that are applied in agricultural systems could harm both human and environment when used in an unrestrained manner. The quantity of pesticides used and, consequently, their presence in food depend on the farming system which is generally higher in conventional integrated pest management (IPM) than that in organic systems (Atkinson 2009). Therefore, agricultural production has been referred to as “the ghost of conventional agriculture past” (Riedo et al. 2021). Consequently, the application of beneficial soil organisms like AMF contrasts with previous and present conventional agricultural procedures because of the deleterious effects of excessive levels of pesticides on soil microbial communities (Dodd 2000; Montesinos 2003; Jacobsen and Hjelmsø 2014). Such unfavorable outcomes are caused not only by the direct harm of pesticides to soil life, but it also alters the soil ecosystem by favoring weeds and some specific crops through the emergence of pesticide resistance (Baek et al. 2021). When active compounds (pesticides) are administered as a soil drench, seed treatment, and foliar spray or when these



substances exist in runoff from leaf application or drift to the soil, they affect the viability and functionality of AMF structures like spores and hyphae in soil. Additionally, variations in the physiology of the host plant may have indirect impacts on AMF symbiosis.

#### 12.3.1.1.1 Fungicides

It is not surprising that fungicides might have a deleterious impact on AMF. There is a varied sensitivity of AMF to fungicides that explains why negative, neutral, and positive impacts of fungicides have been recorded. Some fungicides target specific families of fungi (e.g., Ascomycota, to which the majority of fungal diseases belong). However, some fungicides, including thiazoles (benomyl and carbendazim), have a substantial adverse impact on Glomeromycota, the phylum of fungi to which AMF belongs. Thus, benomyl causes a reduction in AMF activity in the field (Allison et al. 2007; Chiocchio et al. 2000; Hartnett and Wilson 2002; O'Connor et al. 2009). Additionally, benomyl has a negative effect on nutrient mineralization and decomposition by reducing the diversity and abundance of saprotrophic soil fungus. An alternative fungicide, Topsin M (also known as topsin), was described for use in ecological study including the reduction of AMF (Wilson and Williamson 2008). The dosage used, as well as the fungicide's interactions with the soil matrix (such as adsorption on organic matter or solubility in soil solution), influences how well fungicides work.

#### 12.3.1.1.2 Herbicides

Because the majority of current efforts are focused on aboveground control, herbicide impact on soil biota has got little attention (Kremer 2014). The three herbicides that are most frequently used are paraquat, oxyfluorfen, and glyphosate. The herbicide oxyfluorfen was found to have a negative or indifferent effect on AMF. Recently, genetically modified crops (such as cotton, maize, soybean, and canola) have been released onto the market and include a gene that renders such plants resistant to glyphosate; its use is probably going to grow. Consequently, glyphosate usage as a weed control method was increased particularly in no-till or conservation agriculture (Watrud et al. 2011; Baek et al. 2021). Due to its alleged quick decomposition in soil and minimal toxicity to vertebrates, glyphosate has been viewed as a reasonably advantageous foliar-acting herbicide (Duke and Powles 2008; Weidenhamer and Callaway 2010). Studies examining the effects of glyphosate on AMF reveal a suppression of AM fungal spore germination as well as germ tube growth and a reduction of mycorrhizal count in soil (Zaller et al. 2014) but only at concentrations higher than those advised for usage in the field. When used at acceptable concentrations, glyphosate has no effect on mycorrhiza (Baumgartner et al. 2010; Pasaribu et al. 2011). Although sublethal doses of herbicides may not

cause the plant to die, they can diminish photosynthetic rates to the point where the symbiosis is compromised.

### 12.3.1.1.3 Insecticides and Nematicides

The effects of fungicides and herbicides on AMF are far greater than the effects of insecticides and nematicides. Insecticides and nematicides, according to earlier evaluations, had either no adverse effects or moderately positive effects on AMF (Trappe et al. 1984; Hamel and Strullu 2006). However, differential impact of systemic and contact insecticides was documented, just as they were in the case of fungicides (Sarr et al. 2013; Deliopoulos et al. 2008). For example, phoxim, an insecticide/acaricide, was reported to prevent AMF colonization in carrot (*Daucus carota*); however, it was not the case in green onion (*Allium fistulosum*) (Wang et al. 2011a, b). Aldicarb, a nematicide, had no impact on mycorrhizal colonization of potato (*Solanum tuberosum*) (Goulson 2013). Currently, neonicotinoid insecticide class is one of the most popularly used globally due to their broad spectrum of action on many pests of various crops. Numerous civilizations use neonicotinoids extensively. They have drawn a lot of attention because of their extremely detrimental effects on insect diversity, which also have major repercussions for ecosystem services like pollination (Hladik et al. 2018; Malfatti et al. 2023). Neocotinoid effects on AMF have just recently been the subject of studies. For instance, two neonicotinoids, imidacloprid as well as thiamethoxam, were evaluated on the spore germination of two AMF species, *Rhizophagus clarus* and *Glomus albida*. The results showed that both neonicotinoids dramatically decreased spore germination of AMF in a dose-dependent way (Malfatti et al. 2023).

### 12.3.1.2 Fertilizers

Due to the critical symbiotic interaction between hyphae of mycorrhizal fungi and host plants, AMF might be sensitive to alternations in soil nutrients (Hack et al. 2019). Numerous earlier researches have demonstrated that modifying the soil microenvironment with the addition of mineral fertilizer can have (positive, negative, or inconsequential) impacts on the proliferation of AMF. A field study revealed that while P application increased AMF abundance, N application predominantly changed the species composition of AMF (Chen et al. 2014). Nevertheless, according to Xiao et al. (2019), the application of N had an impact on AMF abundance, while the addition of P had an impact on AMF diversity, and the augmentation of N and P had no discernible impact on the AMF community composition. Besides, high soil nutrient levels, such as N and P, can promote AMF sporulation, so adding organic fertilizer helps the soil flora thrive (Qin et al. 2015). Furthermore, soil pH and K have a significant impact on the AMF community structure (Qin et al. 2015).

### ***12.3.2 Soil Tillage***

Rotary soil tillage is regarded as a traditional farming method. Rotary tillage is important because it breaks up and softens the soil while also equally distributing organic materials and nutrients. No-till farming, on the other hand, is a method of crop production in which the soil is not tilled. Crop growth and yield have been observed to be lowered (Huynh et al. 2019), unchanged (Büchi et al. 2017), or increased (da Silva et al. 2022) when compared to rotary tillage cultivation. As previously mentioned, tillage is a factor that influences soil hardness and the alterations in root growth which can either improve or decrease yield (Ren et al. 2018). Additionally, it has been proposed that variations in the species of soil microbes may have an impact on crop yield, which is another element that may change crop growth and yield during plowing (Ma et al. 2020). AMF produce exterior hyphae that provide the host with P that could not be taken up through plant roots (Smith and Read 2008). Intensive soil tillage may have a deleterious impact on AMF biomass and community composition. The disruption of the AMF mycelial network has been proven to be the primary reason why traditional farming generally diminishes the quantity of AMF when compared to no-tillage systems (Gu et al. 2020; Jansa et al. 2006; Castelli et al. 2014). Additionally, substantial research has been done to show how tillage techniques and AMF colonization can change AMF communities in soil and plant roots (Tatewaki et al. 2021; Liu et al. 2022a, b; Lu et al. 2018; Gu et al. 2020; Tatewaki et al. 2021).

### ***12.3.3 Crop Rotation***

Crop rotation is a popular agronomic soil management technique that aims to manage nutrient requirements, maintain healthy soil, reduce pest and disease pressure, and reduce reliance on agricultural chemicals in order to produce high yields that are sustainable (Angus et al. 2015). Crop rotation slows the growth of weeds and plant diseases (Krupinsky et al. 2002; Liebman and Dyck 1993). It also hastens the buildup of soil organic C and soil N (Havlin et al. 1990; Van Eerd et al. 2014). Additionally, a system with a greater variety of rotating crops is linked to a greater diversity and richness of soil microbial life (Venter et al. 2016). The benefits of crop rotation systems are influenced by the plant species chosen and the order in which they are planted (Yang and Kay 2001; West and Post 2002). Despite its significance, it can also impact soil microbial communities, particularly AMF, which are imperative for plant growth. The existence of nonhost crops in the rotation has been proven to have an impact on root colonization and spore abundance (Arihara and Karasawa 2000). Extremely little AMF variety has reportedly been found in monoculture (Burrows and Pflieger 2002; Oehl et al. 2003). Crop rotations appear to be a simple and low-cost farming technique associated with increased AMF sporulation in many upland farming systems. AMF sporulation increased in a maize-common

bean crop rotation system compared to a maize monoculture, and AMF sporulation correlates with the crop previously planted on the site (Oruru and Njeru 2016). Similarly, crop rotation with mycorrhizal plants and AMF inoculation both improve maize growth performance in limed, acid sulfate soil (Higo et al. 2010). Crop rotation also aids plant nutrient absorption. Improved N uptake by maize plants in a maize-common bean crop rotation system is one example (Oruru and Njeru 2016). Similarly, Ratnayake et al. (2017) revealed that rotations of crops can boost nutrient availability in soil and stocks of C compared with monocultures.

## **12.4 Comparing Arbuscular Mycorrhizal Fungi in Organic and Conventional Farming**

### ***12.4.1 Benefits and Limitations of Organic and Conventional Farming***

Organic and conventional farming both have advantages and disadvantages. Although conventional farming is frequently associated with higher yields and lower costs, it is harmful to the environment, reduces biodiversity, and poses potential health risks. In addition, traditional farming controls synthetic inputs (such as pesticides, chemical fertilizers, and amendments) consistently, disregarding the naturally occurring spatial-longitudinal heterogeneity of soil and crop conditions within fields. As a result, organic farming is emerging as a viable option for productive and sustainable agriculture although it is characterized by high cost and relatively unstable yield (Jaacks et al. 2022). Organic farming relies on the use of organic fertilizers, biofertilizers, and organominerals, which helps to avoid the negative environmental effects and toxicities associated with chemicalized farming. In addition, organic agriculture increases the agroecosystem's resistance to the negative effects of climate change by enhancing the sequestration and conservation of C in soil systems (Gamage et al. 2023).

### ***12.4.2 Influence of Organic and Conventional Farming on Arbuscular Mycorrhizal Diversity and Community Composition***

Agricultural management strategies and differences in land-use types can have a significant impact on AMF populations (Martinez and Johnson 2010; Zhao et al. 2015a, b). Understanding the effects of organic and conventional agricultural practices on the diversity and community composition of mycorrhizal symbiosis is essential for agricultural production and sustainable soil utilization. Several studies have been undertaken in agricultural contexts for comparing the effect of organic and

**Table 12.1** Recent studies comparing the effect of organic and conventional management approaches on arbuscular mycorrhizal fungi

Studied plants	Major findings	Applied techniques	References
Maize root	The colonization intensity, operational taxonomic unit, and taxonomic diversity were all greater on conventional farms compared with that on organic farms	Sequencing of V4–V5 hyper-variable regions of the fungal 18S rRNA gene	Chen et al. (2022)
Wheat	Conventional agriculture had a substantial impact on the AMF population but led to a decline in AMF diversity	Sequencing of 18S rRNA gene amplicons	Wahdan et al. (2021)
Cereal crops	Organic farming had a positive effect on the AMF diversity, which influenced plant productivity	Sequencing of the LSU region of ribosomal DNA	Manoharan et al. (2017)
Apple	AMF abundance was substantially greater in organically managed than in conventionally managed orchards. Also, organic orchards harbored significantly different AMF communities	454 pyrosequencing of SSU rRNA gene amplicons	van Geel et al. (2015)
Maize and potato	A decline was observed in the diversity of mycorrhizal taxa in conventionally maintained fields	T-RFLP of 25S rRNA gene fragments	Verbruggen et al. (2012)
Maize and potato	The average number of AMF taxa was greater in areas treated organically. In addition, AMF abundance significantly increased after transition to organic agriculture	Terminal restriction fragment-length polymorphism (T-RFLP) of PCR-amplified large subunit rRNA gene fragments	Verbruggen et al. (2010)
Onion	Farming systems did not influence AMF diversity	Sequencing of the partial 18S-ITS1–5.8S-ITS2 rDNA region	Galván et al. (2009)
Potatoes, winter wheat, and beetroots	Mycorrhizal spore abundance and species diversity were considerably greater in organic systems than that in conventional ones	Morphological identification	Oehl et al. (2004)

conventional management approaches on AMF diversity (Table 12.1). Almost all studies indicated that organic farming is capable of sustaining greater diversity of AMF than conventional farming and showed the necessity of an improved diversity for long-term crop productivity (Manoharan et al. 2017).

## 12.5 Digging Deeper: Factors to Consider When Applying Arbuscular Mycorrhizal Fungi in Organic Farming

The environmentally benign, innovative, and sustainable method of organic farming increases agricultural output and the quality of life for many farmers. The role of AMF in helping soil fertility, sustainability, and crop production improvement is one of the most researched and well-known topics in agronomy. When incorporating AMF into organic farming practices, several factors need to be taken into consideration.

Firstly, arbuscular mycorrhizal fungal communities can be influenced by the identity of the host plant. Furthermore, plant traits, such as root exudates and morphological characteristics, play a critical role in determining the specificity of AM symbiosis (Ramana et al. 2023). Hence, various studies have shown that the presence of suitable AMF species is required in a given soil system. For instance, Gao et al. (2020) reported a high specificity between roots of cotton plant and the mycorrhizal fungus, *Rhizophagus irregularis* CD1. They also reported an increase in growth and yield, boll number per plant, as well as the maturity of the fiber cotton when inoculated with *Rhizophagus irregularis* CD1 as compared with non-inoculated plant. After mycorrhizal inoculation, both the expression of particular P transporter family genes and the P concentration in cotton biomass were dramatically increased.

Secondly, the presence of other soil microorganisms should also be considered. Some microorganisms, such as certain bacteria and fungi, may cooperate with AMF to facilitate the host plant growth (Feng et al. 2023). Therefore, it is essential to maintain a balanced microbial community to support the symbiotic relationship between AMF and plants. For instance, a recent study revealed that some isolates of AMF (e.g., *Gigaspora* sp.) and bacteria (e.g., *Sphingomonas*) assisted their host plants thrive in the native soil (Feng et al. 2023). The combined use of *Bacillus subtilis* and AM fungi resulted in superior yield-related indexes and biofortification in wheat grains (Yadav et al. 2020).

Thirdly, combining AMF with suitable sustainable agronomic practices can maximize the expected benefits from plant-arbuscular mycorrhizal symbiosis. For instance, the use of biofertilizers to agricultural soils has a positive impact on crop productivity and soil fertility. Combining the exogenous AMF *Rhizoglyphus irregularis* (DAOM) with phospho-compost resulted in considerably increased shoot and root dry weights as well as yield in lettuce plants compared to control plants (Anli et al. 2022). In addition, soil characteristics, particularly soil total organic C, N, P, and GRSP content, were considerably enhanced in soils treated with biofertilizers and *Rhizoglyphus irregularis* (DAOM) after 2 months of field testing compared to the control (Anli et al. 2022).

Finally, regular monitoring and evaluation of AMF colonization and effectiveness are essential. This can be accomplished by sampling and analysing roots, and assessing plant growth and nutrient acquisition. To maximize the benefits provided by AMF, management practices may need to be adjusted based on the results.

## 12.6 Arbuscular Mycorrhizal Fungi and Climate-Smart Organic Agriculture

Climate change is one of the most significant concerns influencing the way of life of people worldwide. Climate change and some agricultural management practices, such as tilling and excessive chemical use, have all led to land degradation and crop output decline. AMF are capable of mitigating future environmental disturbances. Field investigations and greenhouse tests under simulated climate warming indicate that temperature increases may have a positive effect on AM symbiosis (Hu et al. 2015). Furthermore, it has been suggested that the stimulation of AMF by elevated atmospheric carbon dioxide (CO<sub>2</sub>) is a key mechanism for promoting soil C sequestration by boosting C inputs to soil and by shielding organic C from degradation via aggregation (Cheng et al. 2012). In a unique field experiment that simulates future climate, Wahdan et al. (2021) found that organic agricultural practices significantly increased total richness of AMF under future climate conditions compared to all other treatments, indicating that organic farming not only mitigates the effects of climate change but also increases richness of AMF under future climate conditions (Wahdan et al. 2021).

## 12.7 Future Directions in Sustainable Forestry: AMF Response to Management Practices in Forests

AMF play a major role in the establishment and succession of plant communities, driving the restoration process of degraded land (Manaut et al. 2015; Karthikeyan and Krishnakumar 2012). AMF is one of the pivotal factors determining species diversity, community structure, and productivity in terrestrial ecosystems, and they regulate the resource allocation among adjacent plants through the underground mycelial network, thereby affecting the coexistence and competition among plants (Smith et al. 2010). Zobel and Öpik (2014) stated that the inherent relationships between AMF and the plant community can be explained by “driver” and “passenger” hypotheses, which illustrated the driving effect of AMF on the plant. Generally, AMF will govern the plant community structure of a certain successional stage, and the specific plant community of this successional stage will affect the colonization of the AMF communities, and the varied AMF communities will further affect the plant community in the next successional stage (Renker et al. 2004). In addition, plant-soil feedback is also a manifestation of AMF-driven plant community succession. Positive feedback can promote community development in the early stage, while negative feedback can facilitate plant species replacement and drive community succession (Kikvidze et al. 2010).

The influence of AMF on plant fitness can be attributed to improved seedling establishment, plant growth, a higher tolerance to belowground plant pathogens, better water acquisition, and improved soil structure and stability in forest

ecosystems (Newsham et al. 1995). Through the removal of nutrients, disturbance of soil or forest floor, and altered microclimates, intensive forest management practices can negatively affect AMF biodiversity (Sharmah and Jha 2014). It has been suggested that intensive forest clearcutting may affect the distribution and composition of AMF communities (Moora et al. 2014; Mummey et al. 2010), favoring some AMF taxa and disfavoring others (Gottshall et al. 2017). Intensive management practices can significantly decrease AMF biomass in soil and glomalin-related soil protein (GRSP) content, of which understory removal, mineral fertilization, and soil tillage are major practices with profound negative effects on AMF (Qin et al. 2017). AMF can produce glomalin, which plays the role of “super glue” in the formation of stable soil structure, which can significantly increase the water percolating capacity and stability of soil (Burrows 2014).

In addition, the abundance of AMF taxa may be correlated with the level of management, and AMF diversity varies depending on management techniques as seen by the high AMF biodiversity in non-managed fields and low diversity in intensively managed fields (Sharmah and Jha 2014). In Moso bamboo forests, AMF abundance and community composition are altered by long-term intensive management, leading to a decrease in soil aggregation (Qin et al. 2017). Moreover, when AMF communities are strongly altered, Moso bamboo forests have been intensively managed for long-term experience soil erosion and organic C decomposition. Because of intensive management practices, AMF communities differ from those of old stands, but the ecosystem can still provide a “symbiotic service” needed to restore old-growth understory plant communities (Uibopuu et al. 2009).

In abandoned slash-and-burn forests, which have been disturbed for 2 or 3 years, AMF communities were similar to those in mature forests (de León et al. 2018). Studies indicate that AMF communities and forest regeneration may be threatened by clearcutting tropical rain forests (Stürmer and Siqueira 2011), whereas slash-and-burn management may have relatively minor effects (Rillig 2004; Stürmer and Siqueira 2011). According to Aguilar-Fernández et al. (2009), slash-and-burn maintained similar levels of species richness in forest and pasture plots. AMF communities that develop after forest conversion to pasture seem to be defined more by the persistent effects derived from conversion and management which can be manifested by variations in composition of plant species, soil quality, and microclimates than by the slash-and-burn process (Sharmah and Jha 2014).

The composition of AMF communities was significantly affected by land use, with communities in the clearcut region being the most distinct (de León et al. 2018). Under various land-use regimes, changes in AMF communities may be caused by both biotic and abiotic interactions. Host plants assist AMF establishment via different types of propagules that arrive at a location of interaction (Fonseca et al. 2017), although abiotic factors filtering environment are more significant components in community development than AMF competition (Bouffaud et al. 2016). According to Oehl et al. (2010), soil type and land-use intensity are the primary factors determining the composition and richness of AMF, and different agricultural practices including monoculture and narrow cereal mono-crop rotations can significantly reduce the biodiversity and richness of AMF in comparison to grasslands.



In terms of plant community structures and production, the biological and functional variety of AMF is crucial to forest ecosystems (Jiang et al. 2017). AMF populations in differently managed forests had distinct effects on the development of various forest plant species; in general, the old growth inoculum had a greater beneficial impact than the inoculum from the young forest. The response of plant communities to the severity of forest management may be mediated by mycorrhizal interactions since land use influences the composition of AM fungus populations (Uibopuu et al. 2012). Thinning is a crucial silvicultural strategy that has an impact on a variety of forest features, including overstory, understory, soil properties (Trentini et al. 2017; Zhou et al. 2016), and microbial communities (Dang et al. 2018). In comparison with the late successional species, the early successional species considerably increased the rate of AMF colonization (Zangaro et al. 2003). The findings of Lu et al. suggested that the expansion of Chinese fir may have reached a turning point in the 32-year period as the number of shared OTUs continuously climbed from 9-year, 17-year, and 23-year to 32-year while decreasing in 45-year (Lu et al. 2019). The colonization, biomass, and diversity of AMF considerably increased with the emergence of *Cunninghamia lanceolata*, and the biomass contribution of AMF to glomalin-related soil protein (GRSP) showed a strong association with soil organic C (Liu et al. 2022a, b). Zhou et al. found that the topsoil C storage was significantly enhanced by in-forest planting of *Sarcardra glabra*, while the diversity and composition of the related AMF community were not significantly affected (Zhou et al. 2022). In-forest planting of *Sarcardra glabra* was beneficial for the forest and ecologically safe for the *C. lanceolata* AMF (Zhou et al. 2022). Wang et al. found that the rate of AMF colonization was substantially and negatively linked with soil P, suggesting that low P soil would be a better choice for using AMF in afforestation (Wang et al. 2019).

## 12.8 Conclusions

AMF play a vital role in ecosystem function, soil structure maintenance through C sequestration, and water retention. As a result, whether farmers use organic or conventional methods has an impact not only on agricultural productivity but also on overall environmental sustainability. This chapter investigated the impact of organic vs. conventional farming practices on AMF using existing research and empirical evidence.

For starters, organic farming has been found to be more conducive to promoting AMF diversity and abundance due to its emphasis on sustainable and eco-friendly practices. The absence of synthetic fertilizers and pesticides promotes a more balanced and natural ecosystem, which promotes the sporulation and activity of AMF. As a result, organic farming systems have the potential to significantly improve soil fertility, nutrient cycling, and plant health. Conventional farming, on the other hand, has been shown to have negative effects on AMF populations because of its heavy reliance on agrochemical inputs. Synthetic pesticides and

fertilizers can disrupt the symbiotic association between AMF and plant host, resulting in a reduction in AMF diversity and activity. This, in turn, can have deleterious impacts on soil health and nutrient availability.

More research is needed in the future to gain a better understanding of the mechanisms that govern interactions between farming practices and AMF communities. This knowledge will be extremely useful in directing agricultural policies and practices toward more environmentally sound and sustainable approaches. Efforts should also be made to educate farmers, policymakers, and consumers about the significance of AMF and the potential benefits of organic farming in promoting their growth.

## References

- Abd-Alla MH, Omar SA, Karanxa S (2000) The impact of pesticides on arbuscular mycorrhizal and nitrogen-fixing symbioses in legumes. *Appl Soil Ecol* 14:191–200. [https://doi.org/10.1016/S0929-1393\(00\)00056-1](https://doi.org/10.1016/S0929-1393(00)00056-1)
- Aguilar-Fernández M, Jaramillo VJ, Varela-Fregoso L et al (2009) Short-term consequences of slash-and-burn practices on the arbuscular mycorrhizal fungi of a tropical dry forest. *Mycorrhiza* 19:179–186. <https://doi.org/10.1007/s00572-009-0229-2>
- Ahanger MA, Agarwal R (2017) Potassium up-regulates antioxidant metabolism and alleviates growth inhibition under water and osmotic stress in wheat (*Triticum aestivum* L). *Protoplasma* 254:1471–1486. <https://doi.org/10.1007/s00709-016-1037-0>
- Akyol TY, Yilmaz O, Uzilday B et al (2020) Plant response to salinity: an analysis of ROS formation, signaling, and antioxidant defense. *Turk J Bot* 44:1–13
- Al-Karaki G, McMichael B, Zak J (2004) Field response of wheat to arbuscular mycorrhizal fungi and drought stress. *Mycorrhiza* 14:263–269. <https://doi.org/10.1007/s00572-003-0265-2>
- Allison VJ, Rajaniemi TK, Goldberg DE, Zak DR (2007) Quantifying direct and indirect effects of fungicide on an old-field plant community: an experimental null-community approach. *Plant Ecol* 190:53–69. <https://doi.org/10.1007/s11258-006-9190-8>
- Angus JF, Kirkegaard JA, Hunt JR et al (2015) Break crops and rotations for wheat. *Crop Pasture Sci* 66(6):523–552
- Anli M, Boutasknit A, Ait-El-Mokhtar M et al (2022) Improving lettuce yield and quality of an agricultural soil using a combination of arbuscular mycorrhizal fungus and phosphate-green wastes compost. *Gesunde Pflanzen* 74:205–217. <https://doi.org/10.1007/s10343-021-00603-0>
- Arihara J, Karasawa T (2000) Effect of previous crops on arbuscular mycorrhizal formation and growth of succeeding maize. *Soil Sci Plant Nutr* 46(1):43–51. <https://doi.org/10.1080/00380768.2000.10408761>
- Aroca R, Porcel R, Ruiz-Lozano JM (2007) How does arbuscular mycorrhizal symbiosis regulate root hydraulic properties and plasma membrane aquaporins in *Phaseolus vulgaris* under drought, cold or salinity stresses? *New Phytol* 173:808–816. <https://doi.org/10.1111/j.1469-8137.2006.01961.x>
- Atkinson D (2009) Soil microbial resources and agricultural policies. In: Azcón-Aguilar C, Barea JM, Gianinazzi S, Gianinazzi-Pearson V (eds) *Mycorrhizas-functional processes and ecological impact*. Springer, Berlin/Heidelberg, pp 1–16
- Audet P, Charest C (2007) Dynamics of arbuscular mycorrhizal symbiosis in heavy metal phytoremediation: meta-analytical and conceptual perspectives. *Environ Pollut* 147:609–614. <https://doi.org/10.1016/j.envpol.2006.10.006>
- Baek Y, Bobadilla LK, Giacomini DA et al (2021) Evolution of glyphosate-resistant weeds. *Rev Environ Contam Toxicol* 255:93–128. [https://doi.org/10.1007/398\\_2020\\_55](https://doi.org/10.1007/398_2020_55)

- Basu S, Rabara RC, Negi S (2018) AMF: the future prospect for sustainable agriculture. *Physiol Mol Plant Pathol* 102:36–45. <https://doi.org/10.1016/j.pmpp.2017.11.007>
- Baumgartner K, Fujiyoshi P, Smith R et al (2010) Weed flora and dormant-season cover crops have no effects on arbuscular mycorrhizae of grapevine. *Weed Res* 50(5):456–466. <https://doi.org/10.1111/j.1365-3180.2010.00793.x>
- Bender SF, Wagg C, van der Heijden MG (2016) An underground revolution: biodiversity and soil ecological engineering for agricultural sustainability. *Trends Ecol Evol* 31:440–452. <https://doi.org/10.1016/j.tree.2016.02.016>
- Berruti A, Lumini E, Balestrini R et al (2016) Arbuscular mycorrhizal fungi as natural biofertilizers: let's benefit from past successes. *Front Microbiol* 6:1559. <https://doi.org/10.3389/fmicb.2015.01559>
- Birhane E, Sterck FJ, Fetene M et al (2012) Arbuscular mycorrhizal fungi enhance photosynthesis, water use efficiency, and growth of frankincense seedlings under pulsed water availability conditions. *Oecologia* 169:895–904. <https://doi.org/10.1007/s00442-012-2258-3>
- Bouffaud M-L, Creamer RE, Stone D et al (2016) Indicator species and co-occurrence in communities of arbuscular mycorrhizal fungi at the European scale. *Soil Biol Biochem* 103:464–470. <https://doi.org/10.1016/j.soilbio.2016.09.022>
- Bowles TM, Barrios-Masias FH, Carlisle EA et al (2016) Effects of arbuscular mycorrhizae on tomato yield, nutrient uptake, water relations, and soil carbon dynamics under deficit irrigation in field conditions. *Sci Total Environ* 566:1223–1234. <https://doi.org/10.1016/j.scitotenv.2016.05.178>
- Büchi L, Wendling M, Amossé C et al (2017) Long and short term changes in crop yield and soil properties induced by the reduction of soil tillage in a long term experiment in Switzerland. *Soil Tillage Res* 174:120–129. <https://doi.org/10.1016/j.still.2017.07.002>
- Bücking H, Shachar-Hill Y (2005) Phosphate uptake, transport and transfer by the arbuscular mycorrhizal fungus *Glomus intraradices* is stimulated by increased carbohydrate availability. *New Phytol* 165:899–912. <https://doi.org/10.1111/j.1469-8137.2004.01274.x>
- Burrows R (2014) Glomalin production and infectivity of arbuscular-mycorrhizal fungi in response to grassland plant diversity. *Am J Plant Sci* 5(1):103–111. <https://doi.org/10.4236/ajps.2014.51013>
- Burrows RL, Pflieger FL (2002) Arbuscular mycorrhizal fungi respond to increasing plant diversity. *Can J Bot* 80(2):120–130. <https://doi.org/10.1139/b01-138>
- Cameron DD, Neal AL, van Wees SC et al (2013) Mycorrhiza-induced resistance: more than the sum of its parts? *Trends Plant Sci* 18:539–545. <https://doi.org/10.1016/j.tplants.2013.06.004>
- Cassman KG, Dobermann A, Walters DT et al (2003) Meeting cereal demand while protecting natural resources and improving environmental quality. *Annu Rev Environ Resour* 28:315–358. <https://doi.org/10.1146/annurev.energy.28.040202.122858>
- Castelli M, Urcoviche RC, Gimenes RMT et al (2014) Arbuscular mycorrhizal fungi diversity in maize under different soil managements and seed treatment with fungicide. *J Food Agric Environ* 12:486–491
- Castillo C, Borie F, Oehl F et al (2016) Arbuscular mycorrhizal fungi biodiversity: prospecting in Southern-Central zone of Chile. A review. *J Soil Sci Plant Nutr* 16:400–422
- Cavagnaro TR, Bender SF, Ashgari HR et al (2015) The role of arbuscular mycorrhizas in reducing soil nutrient loss. *Trends Plant Sci* 20:283–290. <https://doi.org/10.1016/j.tplants.2015.03.004>
- Chen YL, Zhang X, Ye JS et al (2014) Six-year fertilization modifies the biodiversity of arbuscular mycorrhizal fungi in a temperate steppe in Inner Mongolia. *Soil Biol Biochem* 69:371–381. <https://doi.org/10.1016/j.soilbio.2013.11.020>
- Chen J, Li J, Yang Y, Wang Y, Zhang Y, Wang P (2022) Effects of conventional and organic agriculture on soil arbuscular mycorrhizal fungal community in low-quality farmland. *Front Microbiol* 13:914627. <https://doi.org/10.3389/fmicb.2022.914627>
- Cheng L, Booker FL, Tu C et al (2012) Arbuscular mycorrhizal fungi increase organic carbon decomposition under elevated CO<sub>2</sub>. *Science* 337:1084–1087. <https://doi.org/10.1126/science.1224304>

- Chiocchio V, Venedikian N, Martínez AE et al (2000) Effect of the fungicide benomyl on spore germination and hyphal length of the arbuscular mycorrhizal fungus *Glomus mosseae*. *Int Microbiol* 3:173–175
- Chourasiya D, Gupta MM, Sahni S et al (2021) Unraveling the AM fungal community for understanding its ecosystem resilience to changed climate in agroecosystems. *Symbiosis* 84(3):295–310. <https://doi.org/10.1007/s13199-021-00761-9>
- Cosme M, Stout MJ, Wurst S (2011) Effect of arbuscular mycorrhizal fungi (*Glomus intraradices*) on the oviposition of rice water weevil (*Lissorhoptrus oryzophilus*). *Mycorrhiza* 21:651–658. <https://doi.org/10.1007/s00572-011-0399-6>
- da Silva GF, Calonego JC, Luperini BCO et al (2022) Soil–plant relationships in soybean cultivated under conventional tillage and long-term no-tillage. *Agronomy* 11(19):2657. <https://doi.org/10.3390/plants11192657>
- Dang P, Gao Y, Liu J et al (2018) Effects of thinning intensity on understory vegetation and soil microbial communities of a mature Chinese pine plantation in the Loess Plateau. *Sci Total Environ* 630:171–180. <https://doi.org/10.1016/j.scitotenv.2018.02.197>
- de León DG, Neuenkamp L, Moora M et al (2018) Arbuscular mycorrhizal fungal communities in tropical rain forest are resilient to slash-and-burn agriculture. *J Trop Ecol* 34:186–199. <https://doi.org/10.1017/S0266467418000184>
- Deliopoulos T, Haydock PJJ, Jones PW (2008) Interaction between arbuscular mycorrhizal fungi and the nematocidal aldicarb on hatch and development of the potato cyst nematode, *Globodera pallida*, and yield of potatoes. *Nematology* 10:783–799. <https://doi.org/10.1163/156854108786161427>
- Dodd JC (2000) The role of arbuscular mycorrhizal fungi in agro- and natural ecosystems. *Outlook Agric* 29(1):63–70. <https://doi.org/10.5367/000000000101293059>
- Duke SO, Powles SB (2008) Glyphosate: a once-in-a century herbicide. *Pest Manag Sci* 64:319–325. <https://doi.org/10.1002/ps.1518>
- El Hilali R, Symanczik S, El Kinany S et al (2022) Cultivation, identification, and application of arbuscular mycorrhizal fungi associated with date palm plants in Drâa-Tafilalet oasis. *Rhizosphere* 22:100521. <https://doi.org/10.1016/j.rhisph.2022.100521>
- Feng Z, Liu X, Qin Y et al (2023) Cooperation of arbuscular mycorrhizal fungi and bacteria to facilitate the host plant growth dependent on soil pH. *Front Microbiol* 14:1116943. <https://doi.org/10.3389/fmicb.2023.1116943>
- Ferrol N, González-Guerrero M, Valderas A et al (2009) Survival strategies of arbuscular mycorrhizal fungi in Cu-polluted environments. *Phytochem Rev* 8:551–559
- Finlay RD (2008) Ecological aspects of mycorrhizal symbiosis: with special emphasis on the functional diversity of interactions involving the extraradical mycelium. *J Exp Bot* 59:1115–1126. <https://doi.org/10.1093/jxb/ern059>
- Fonseca MB, Dias T, Carolino MM et al (2017) Belowground microbes mitigate plant-plant competition. *Plant Sci* 262:175–181. <https://doi.org/10.1016/j.plantsci.2017.06.006>
- Frank B (1885) Über die auf Wurzelsymbiose beruhende Ernährung gewisser Bäume durch unterirdische Pilze. <https://doi.org/10.1111/j.1438-8677.1885.tb04240.x>
- Galván GA, Parádi I, Burger K et al (2009) Molecular diversity of arbuscular mycorrhizal fungi in onion roots from organic and conventional farming systems in the Netherlands. *Mycorrhiza* 19:317–328. <https://doi.org/10.1007/s00572-009-0237-2>
- Gamage A, Gangahagedara R, Gamage J, Jayasinghe N, Kodikara N, Suraweera P, Merah O (2023) Role of organic farming for achieving sustainability in agriculture. *Farming System 1* (1): 100005, <https://doi.org/10.1016/j.farsys.2023.100005>
- Gao X, Guo H, Zhang Q et al (2020) Arbuscular mycorrhizal fungi (AMF) enhanced the growth, yield, fiber quality and phosphorus regulation in upland cotton (*Gossypium hirsutum* L.). *Sci Rep* 10(1):2084. <https://doi.org/10.1038/s41598-020-59180-3>
- Gonzalez-Dugo V (2010) The influence of arbuscular mycorrhizal colonization on soil–root hydraulic conductance in *Agrostis stolonifera* L. under two water regimes. *Mycorrhiza* 20:365–373. <https://doi.org/10.1007/s00572-009-0294-6>

- Gonzalez-Guerrero M, Melville LH, Ferrol N et al (2008) Ultrastructural localization of heavy metals in the extraradical mycelium and spores of the arbuscular mycorrhizal fungus *Glomus intraradices*. *Can J Microbiol* 54:103–110. <https://doi.org/10.1139/W07-119>
- Gosling P, Jones J, Bending GD (2016) Evidence for functional redundancy in arbuscular mycorrhizal fungi and implications for agroecosystem management. *Mycorrhiza* 26:77–83. <https://doi.org/10.1007/s00572-015-0651-6>
- Gottshall CB, Cooper M, Emery SM (2017) Activity, diversity and function of arbuscular mycorrhizae vary with changes in agricultural management intensity. *Agric Ecosyst Environ* 241:142–149. <https://doi.org/10.1016/j.agee.2017.03.011>
- Goulson D (2013) An overview of the environmental risks posed by neonicotinoid insecticides. *J Appl Ecol* 50:977–987. <https://doi.org/10.1111/1365-2664.12111>
- Gryndler M, Larsen J, Hřšelová H et al (2006) Organic and mineral fertilization, respectively, increase and decrease the development of external mycelium of arbuscular mycorrhizal fungi in a long-term field experiment. *Mycorrhiza* 16:159–166. <https://doi.org/10.1007/s00572-005-0027-4>
- Gu S, Wu S, Guan Y et al (2020) Arbuscular mycorrhizal fungal community was affected by tillage practices rather than residue management in black soil of northeast China. *Soil Tillage Res* 198: 104552. <https://doi.org/10.1016/j.still.2019.104552>
- Hack CM, Porta M, Schäufele R et al (2019) Arbuscular mycorrhiza mediated effects on growth, mineral nutrition and biological nitrogen fixation of *Melilotus alba* Med. in a subtropical grassland soil. *Appl Soil Ecol* 134:38–44. <https://doi.org/10.1016/j.apsoil.2018.10.008>
- Hajiboland R, Aliasgharzadeh N, Laiegh SF et al (2010) Colonization with arbuscular mycorrhizal fungi improves salinity tolerance of tomato (*Solanum lycopersicum* L.) plants. *Plant Soil* 331: 313–327. <https://doi.org/10.1007/s11104-009-0255-z>
- Hajiboland R, Joudmand A, Aliasgharzad N, Tolra R, & Poschenrieder C (2019). Arbuscular mycorrhizal fungi alleviate low-temperature stress and increase freezing resistance as a substitute for acclimation treatment in barley. *Crop & Pasture Science*, 70:218–233. <https://doi.org/10.1071/cp18385>
- Hamel C, Strullu D (2006) Arbuscular mycorrhizal fungi in field crop production: potential and new direction. *Can J Plant Sci* 86:941–950. <https://doi.org/10.4141/P05-099>
- Hartley SE, Gange AC (2009) Impacts of plant symbiotic fungi on insect herbivores: mutualism in a multitrophic context. *Annu Rev Entomol* 54:323–342. <https://doi.org/10.1146/annurev.ento.54.110807.090614>
- Hartnett DC, Wilson GWT (2002) The role of mycorrhizas in plant community structure and dynamics: lessons from grasslands. In: Smith SE, Smith FA (eds) *Diversity and integration in mycorrhizas*. *Developments in plant and soil sciences*, vol 94. Springer, Dordrecht. [https://doi.org/10.1007/978-94-017-1284-2\\_31](https://doi.org/10.1007/978-94-017-1284-2_31)
- Hashem A, Abd-Allah EF, Alqarawi AA et al (2016) The interaction between arbuscular mycorrhizal fungi and endophytic bacteria enhances plant growth of *Acacia gerrardii* under salt stress. *Front Microbiol* 7:1089. <https://doi.org/10.3389/fmicb.2016.01089>
- Havlin JL, Kissel DE, Maddux LD et al (1990) Crop rotation and tillage effects on soil organic carbon and nitrogen. *Soil Sci Soc Am J* 54(2):448–452. <https://doi.org/10.2136/sssaj1990.03615995005400020026x>
- Hestrin R, Hammer EC, Mueller CW et al (2019) Synergies between mycorrhizal fungi and soil microbial communities increase plant nitrogen acquisition. *Commun Biol* 2:233. <https://doi.org/10.1038/s42003-019-0481-8>
- Higo M, Isobe K, Kang DJ et al (2010) Inoculation with arbuscular mycorrhizal fungi or crop rotation with mycorrhizal plants improves the growth of maize in limed acid sulfate soil. *Plant Prod Sci* 13(1):74–79. <https://doi.org/10.1626/pp.s.13.74>
- Hladik ML, Main AR, Goulson D (2018) Environmental risks and challenges associated with neonicotinoid insecticides. *Environ Sci Technol* 52:3329–3335. <https://doi.org/10.1021/acs.est.7b06388>

- Hodge A, Fitter AH (2010) Substantial nitrogen acquisition by arbuscular mycorrhizal fungi from organic material has implications for N cycling. *Proc Natl Acad Sci* 107:13754–13759. <https://doi.org/10.1073/pnas.1005874107>
- Hu Y, Wu S, Sun Y et al (2015) Arbuscular mycorrhizal symbiosis can mitigate the negative effects of night warming on physiological traits of *Medicago truncatula* L. *Mycorrhiza* 25:131–142. <https://doi.org/10.1007/s00572-014-0595-2>
- Huang Z, Zou Z, He C et al (2011) Physiological and photosynthetic responses of melon (*Cucumis melo* L.) seedlings to three *Glomus* species under water deficit. *Plant Soil* 339:391–399. <https://doi.org/10.1007/s11104-010-0591-z>
- Huynh HT, Hufnagel J, Wurbs A et al (2019) Influences of soil tillage, irrigation and crop rotation on maize biomass yield in a 9-year field study in Müncheberg, Germany. *Field Crops Res* 241: 107565. <https://doi.org/10.1016/j.fcr.2019.107565>
- Iqbal N, Umar S, Khan NA (2015) Nitrogen availability regulates proline and ethylene production and alleviates salinity stress in mustard (*Brassica juncea*). *J Plant Physiol* 178:84–91. <https://doi.org/10.1016/j.jplph.2015.02.006>
- Jaacks LM, Serupally R, Dabholkar S et al (2022) Articles Impact of large-scale, the government legislated and funded organic farming training on pesticide use in Andhra Pradesh, India: a cross-sectional study. *Lancet Planet Health* 6:e310–e319. [https://doi.org/10.1016/S2542-5196\(22\)00062-6](https://doi.org/10.1016/S2542-5196(22)00062-6)
- Jacobsen CS, Hjelmsø MH (2014) Agricultural soils, pesticides and microbial diversity. *Curr Opin Biotechnol* 27:15–20. <https://doi.org/10.1016/j.copbio.2013.09.003>
- Jansa J, Mozafar A, Anken T et al (2002) Diversity and structure of AMF communities as affected by tillage in a temperate soil. *Mycorrhiza* 12:225–234. <https://doi.org/10.1007/s00572-002-0163-z>
- Jansa J, Wiemken A, Frossard E (2006) The effects of agricultural practices on arbuscular mycorrhizal fungi. *Geol Soc Lond, Spec Publ* 266(1):89–115. <https://doi.org/10.1144/GSL.SP.2006.266.01.08>
- Javot H, Penmetsa RV, Terzaghi N et al (2007) A *Medicago truncatula* phosphate transporter indispensable for the arbuscular mycorrhizal symbiosis. *Proc Natl Acad Sci* 104:1720–1725. <https://doi.org/10.1073/pnas.0608136104>
- Jayne B, Quigley M (2014) Influence of arbuscular mycorrhiza on growth and reproductive response of plants under water deficit: a meta-analysis. *Mycorrhiza* 24:109–119. <https://doi.org/10.1007/s00572-013-0515-x>
- Jeffries P, Gianinazzi S, Perotto S et al (2003) The contribution of arbuscular mycorrhizal fungi in sustainable maintenance of plant health and soil fertility. *Biol Fertil Soils* 37:1–16. <https://doi.org/10.1007/s00374-002-0546-5>
- Jiang J, Moore JA, Priyadarshi A et al (2017) Plant-mycorrhizal interactions mediate plant community coexistence by altering resource demand. *Ecology* 98:187–197. <https://doi.org/10.1002/ecy.1630>
- Johnson NC, Angelard C, Sanders IR et al (2013) Predicting community and ecosystem outcomes of mycorrhizal responses to global change. *Ecol Lett* 16:140–153. <https://doi.org/10.1111/ele.12085>
- Jung SC, Martinez-Medina A, Lopez-Raez JA et al (2012) Mycorrhiza-induced resistance and priming of plant defenses. *J Chem Ecol* 38:651–664. <https://doi.org/10.1007/s10886-012-0134-6>
- Karthikeyan A, Krishnakumar N (2012) Reforestation of bauxite mine spoils with *Eucalyptus tereticornis* Sm. seedlings inoculated with arbuscular mycorrhizal fungi. *Ann For Res* 55: 207–216. <https://doi.org/10.15287/afr.2012.61>
- Kempel A, Schmidt AK, Brandl R et al (2010) Support from the underground: induced plant resistance depends on arbuscular mycorrhizal fungi. *Funct Ecol* 24:293–300. <https://doi.org/10.1111/j.1365-2435.2009.01647.x>

- Kikvidze Z, Armas C, Fukuda K et al (2010) The role of arbuscular mycorrhizae in primary succession: differences and similarities across habitats. *Web Ecol* 10:50–57. <https://doi.org/10.5194/we-10-50-2010>
- Kremer RJ (2014) Environmental implications of herbicide resistance: soil biology and ecology. *Weed Sci* 62:415–426. JSTOR. <http://www.jstor.org/stable/43700669>
- Krupinsky JM, Bailey KL, McMullen MP et al (2002) Managing plant disease risk in diversified cropping systems. *Agron J* 94(2):198–209. <https://doi.org/10.2134/agronj2002.1980>
- Kurle JE, Pfeleger FL (1994) The effects of cultural practices and pesticides on VAM fungi. *Mycorrhizae and plant health*. APS Press, Minnesota, pp 101–131
- Lanfranco L, Fiorilli V, Gutjahr C (2018) Partner communication and role of nutrients in the arbuscular mycorrhizal symbiosis. *New Phytol* 220:1031–1046. <https://doi.org/10.1111/nph.15230>
- Lax P, Becerra AG, Soteras F et al (2011) Effect of the arbuscular mycorrhizal fungus *Glomus intraradices* on the false root-knot nematode *Nacobbus aberrans* in tomato plants. *Biol Fertil Soils* 47:591–597. <https://doi.org/10.1007/s00374-010-0514-4>
- Lehmann A, Veresoglou SD, Leifheit EF et al (2014) Arbuscular mycorrhizal influence on zinc nutrition in crop plants—a meta-analysis. *Soil Biol Biochem* 69:123–131. <https://doi.org/10.1016/j.soilbio.2013.11.001>
- Lehmann J, Bossio DA, Kögel-Knabner I et al (2020) The concept and future prospects of soil health. *Nat Rev Earth Environ* 1:544–553. <https://doi.org/10.1038/s43017-020-0080-8>
- Lekberg Y, Koide R (2005) Arbuscular mycorrhizal fungi, rhizobia, available soil P and nodulation of groundnut (*Arachis hypogaea*) in Zimbabwe. *Agric Ecosyst Environ* 110(3–4):143–148. <https://doi.org/10.1016/j.agee.2005.03.011>
- Liebman M, Dyck E (1993) Crop rotation and intercropping strategies for weed management. *Ecol Appl* 3(1):92–122. <https://doi.org/10.2307/1941795>
- Liu W, Ma K, Wang X et al (2022a) Effects of no-tillage and biologically-based organic fertilizer on soil arbuscular mycorrhizal fungal communities in winter wheat field. *Appl Soil Ecol* 178:104564. <https://doi.org/10.1016/j.apsoil.2022.104564>
- Liu Z, Han Y, Lai F et al (2022b) Contribution of arbuscular mycorrhizal fungal communities to soil carbon accumulation during the development of *Cunninghamia lanceolata* plantations. *Forests* 13:2099. <https://doi.org/10.3390/f13122099>
- Lu X, Lu X, Liao Y (2018) Effect of tillage treatment on the diversity of soil arbuscular mycorrhizal fungal and soil aggregate-associated carbon content. *Front Microbiol* 9:2986. <https://doi.org/10.3389/fmicb.2018.02986>
- Lu N, Xu X, Wang P et al (2019) Succession in arbuscular mycorrhizal fungi can be attributed to a chronosequence of *Cunninghamia lanceolata*. *Sci Rep* 9:18057. <https://doi.org/10.1038/s41598-019-54452-z>
- Ma G, Kang J, Wang J et al (2020) Bacterial community structure and predicted function in wheat soil from the North China plain are closely linked with soil and plant characteristics after seven years of irrigation and nitrogen application. *Front Microbiol* 11:506. <https://doi.org/10.3389/fmicb.2020.00506>
- Ma Y, Zhang H, Wang D et al (2021) Differential responses of arbuscular mycorrhizal fungal communities to long-term fertilization in the wheat rhizosphere and root endosphere. *Appl Environ Microbiol* 87:e00349–e00321. <https://doi.org/10.1128/aem.00349-21>
- Mäder P, Fliessbach A, Dubois D et al (2002) Soil fertility and biodiversity in organic farming. *Science* 296(5573):1694–1697. <https://doi.org/10.1126/science.1071148>
- Malfatti ALR, Filho LCJO, Carniel LSC et al (2023) Risk assessment tests of neonicotinoids on spore germination of arbuscular mycorrhizal fungi *Gigaspora albida* and *Rhizophagus clarus*. *J Soils Sedim* 23:1295–1303. <https://doi.org/10.1007/s11368-022-03419-1>
- Manaut N, Sanguin H, Ouahmane L et al (2015) Potentialities of ecological engineering strategy based on native arbuscular mycorrhizal community for improving afforestation programs with carob trees in degraded environments. *Ecol Eng* 79:113–119. <https://doi.org/10.1016/j.ecoleng.2015.03.007>

- Manoharan L, Rosenstock NP, Williams A et al (2017) Agricultural management practices influence AMF diversity and community composition with cascading effects on plant productivity. *Appl Soil Ecol* 115:53–59. <https://doi.org/10.1016/j.apsoil.2017.03.012>
- Martin FM, Uroz S, Barker DG (2017) Ancestral alliances: plant mutualistic symbioses with fungi and bacteria. *Science* 356:eaad4501. <https://doi.org/10.1126/science.aad4501>
- Martinez TN, Johnson NC (2010) Agricultural management influences propagule densities and functioning of arbuscular mycorrhizas in low-and high-input agroecosystems in arid environments. *Appl Soil Ecol* 46:300–306. <https://doi.org/10.1016/j.apsoil.2010.07.001>
- Mathimaran N, Ruh R, Jama B et al (2007) Impact of agricultural management on arbuscular mycorrhizal fungal communities in Kenyan ferralsols. *Agric Ecosyst Environ* 119(1–2):22–32. <https://doi.org/10.1016/j.agee.2006.06.004>
- Mathur S, Sharma MP, Jajoo A (2018) Improved photosynthetic efficacy of maize (*Zea mays*) plants with arbuscular mycorrhizal fungi (AMF) under high temperature stress. *J Photochem Photobiol B* 180:149–154. <https://doi.org/10.1016/j.jphotobiol.2018.02.002>
- Mo F, Han J, Wen XX et al (2020) Quantifying regional effects of plastic mulch on soil nitrogen pools, cycles, and fluxes in rain-fed agroecosystems of the Loess Plateau. *Land Degrad Dev* 31:1675–1687. <https://doi.org/10.1002/ldr.3548>
- Montesinos E (2003) Plant-associated microorganisms: a view from the scope of microbiology. *Int Microbiol* 6:221–223. <https://doi.org/10.1007/s10123-003-0141-0>
- Moora M, Davison J, Öpik M et al (2014) Anthropogenic land use shapes the composition and phylogenetic structure of soil arbuscular mycorrhizal fungal communities. *FEMS Microbiol Ecol* 90:609–621. <https://doi.org/10.1111/1574-6941.12420>
- Morris EK, Morris D, Vogt S et al (2019) Visualizing the dynamics of soil aggregation as affected by arbuscular mycorrhizal fungi. *ISME J* 13:1639–1646. <https://doi.org/10.1038/s41396-019-0369-0>
- Mortimer P, Pérez-Fernández M, Valentine A (2008) The role of arbuscular mycorrhizal colonization in the carbon and nutrient economy of the tripartite symbiosis with nodulated *Phaseolus vulgaris*. *Soil Biol Biochem* 40:1019–1027. <https://doi.org/10.1016/j.soilbio.2007.11.014>
- Mummey DL, Clarke JT, Cole CA et al (2010) Spatial analysis reveals differences in soil microbial community interactions between adjacent coniferous forest and clear cut ecosystems. *Soil Biol Biochem* 42:1138–1147. <https://doi.org/10.1016/j.soilbio.2010.03.020>
- Newsham K, Fitter A, Watkinson A (1995) Multi-functionality and biodiversity in arbuscular mycorrhizas. *Trends Ecol Evol* 10(10):407–411. [https://doi.org/10.1016/S0169-5347\(00\)89157-0](https://doi.org/10.1016/S0169-5347(00)89157-0)
- O'Connor P, Manjarrez M, Smith SE (2009) The fate and efficacy of benomyl applied to field soils to suppress activity of arbuscular mycorrhizal fungi. *Can J Microbiol* 55(7):901–904. <https://doi.org/10.1139/w09-035>
- Oehl F, Sieverding E, Ineichen K et al (2003) Impact of land use intensity on the species diversity of arbuscular mycorrhizal fungi in agroecosystems of Central Europe. *Appl Environ Microbiol* 69(5):2816–2824. <https://doi.org/10.1128/aem.69.5.2816-2824.2003>
- Oehl F, Sieverding E, Mäder P et al (2004) Impact of long-term conventional and organic farming on the diversity of arbuscular mycorrhizal fungi. *Oecologia* 138:574–583. <https://doi.org/10.1007/s00442-003-1458-2>
- Oehl F, Laczko E, Bogenrieder A et al (2010) Soil type and land use intensity determine the composition of arbuscular mycorrhizal fungal communities. *Soil Biol Biochem* 42:724–738. <https://doi.org/10.1016/j.soilbio.2010.01.006>
- Oruru MB, Njeru EM (2016) Upscaling arbuscular mycorrhizal symbiosis and related agroecosystems services in smallholder farming systems. *Biomed Res Int* 2016:4376240. <https://doi.org/10.1155/2016/4376240>
- Pasaribu A, Mohamad RB, Awang Y et al (2011) Growth and development of symbiotic Arbuscular mycorrhizal fungi, *Glomus mossea* (Nicol. and Gerd.), in alachlor and glyphosate treated soils. *Afr J Biotechnol* 10(55):11520–11526. <https://doi.org/10.5897/AJB11.1200>



- Pellegrino E, Bedini S (2014) Enhancing ecosystem services in sustainable agriculture: biofertilization and biofortification of chickpea (*Cicer arietinum* L.) by arbuscular mycorrhizal fungi. *Soil Biol Biochem* 68:429–439. <https://doi.org/10.1016/j.soilbio.2013.09.030>
- Porcel R, Redondo-Gómez S, Mateos-Naranjo E et al (2015) Arbuscular mycorrhizal symbiosis ameliorates the optimum quantum yield of photosystem II and reduces non-photochemical quenching in rice plants subjected to salt stress. *J Plant Physiol* 185:75–83. <https://doi.org/10.1016/j.jplph.2015.07.006>
- Qin H, Lu K, Strong PJ et al (2015) Long-term fertilizer application effects on the soil, root arbuscular mycorrhizal fungi and community composition in rotation agriculture. *Appl Soil Ecol* 89:35–43. <https://doi.org/10.1016/j.apsoil.2015.01.008>
- Qin H, Chen J, Wu Q et al (2017) Intensive management decreases soil aggregation and changes the abundance and community compositions of arbuscular mycorrhizal fungi in Moso bamboo (*Phyllostachys pubescens*) forests. *For Ecol Manag* 400:246–255
- Ramana JV, Tylisanakis JM, Ridgway HJ et al (2023) Root diameter, host specificity and arbuscular mycorrhizal fungal community composition among native and exotic plant species. *New Phytol* 239:301–310. <https://doi.org/10.1111/nph.18911>
- Ratnayake RR, Perera B, Rajapaksha R et al (2017) Soil carbon sequestration and nutrient status of tropical rice based cropping systems: Rice-Rice, Rice-Soya, Rice-Onion and Rice-Tobacco in Sri Lanka. *Catena* 150:17–23. <https://doi.org/10.1016/j.catena.2016.11.006>
- Recorbet G, Abdallah C, Renaut J et al (2013) Protein actors sustaining arbuscular mycorrhizal symbiosis: underground artists break the silence. *New Phytol* 199:26–40. <https://doi.org/10.1111/nph.12287>
- Ren B, Li X, Dong S, Liu P et al (2018) Soil physical properties and maize root growth under different tillage systems in the North China Plain. *Crop J* 6:669–676. <https://doi.org/10.1016/j.cj.2018.05.009>
- Ren A-T, Zhu Y, Chen Y-L et al (2019) Arbuscular mycorrhizal fungus alters root-sourced signal (abscisic acid) for better drought acclimation in *Zea mays* L. seedlings. *Environ Exp Bot* 167: 103824. <https://doi.org/10.1016/j.envexpbot.2019.103824>
- Renker C, Zobel M, Öpik M et al (2004) Structure, dynamics, and restoration of plant communities: do Arbuscular Mycorrhizae matter? In: Temperton VK, Hobbs RJ, Nuttle T et al (eds) *Assembly rules and restoration ecology: bridging the gap between theory and practice*. Island Press, pp 189–229
- Riedo J, Wettstein FE, Rösch A et al (2021) Widespread occurrence of pesticides in organically managed agricultural soils—the ghost of a conventional agriculture past? *Environ Sci Technol* 55:2919–2928. <https://doi.org/10.1021/acs.est.0c06405>
- Rillig MC (2004) Arbuscular mycorrhizae and terrestrial ecosystem processes. *Ecol Lett* 7:740–754. <https://doi.org/10.1111/j.1461-0248.2004.00620.x>
- Rillig MC, Mummey DL (2006) Mycorrhizas and soil structure. *New Phytol* 171:41–53. <https://doi.org/10.1111/j.1469-8137.2006.01750.x>
- Rillig MC, Sosa-Hernández MA, Roy J et al (2016) Towards an integrated mycorrhizal technology: harnessing mycorrhiza for sustainable intensification in agriculture. *Front Plant Sci* 7:1625. <https://doi.org/10.3389/fpls.2016.01625>
- Rillig MC, Aguilar-Trigueros CA, Camenzind T et al (2019) Why farmers should manage the arbuscular mycorrhizal symbiosis. *New Phytol* 222:1171–1175. <https://doi.org/10.1111/nph.15602>
- Roger A, Colard A, Angelard C et al (2013) Relatedness among arbuscular mycorrhizal fungi drives plant growth and intraspecific fungal coexistence. *ISME J* 7:2137–2146. <https://doi.org/10.1038/ismej.2013.112>
- Rozpądek P, Weźowicz K, Stojakowska A et al (2014) Mycorrhizal fungi modulate phytochemical production and antioxidant activity of *Cichorium intybus* L. (Asteraceae) under metal toxicity. *Chemosphere* 112:217–224. <https://doi.org/10.1016/j.chemosphere.2014.04.023>

- Ruiz-Lozano JM (2003) Arbuscular mycorrhizal symbiosis and alleviation of osmotic stress. New perspectives for molecular studies. *Mycorrhiza* 13(6):309–317. <https://doi.org/10.1007/s00572-003-0237-6>
- Ryan MH, Graham JH (2018) Little evidence that farmers should consider abundance or diversity of arbuscular mycorrhizal fungi when managing crops. *New Phytol* 220:1092–1107. <https://doi.org/10.1111/nph.15308>
- Saldajeno MGB, Ito M, Hyakumachi M (2012) Interaction between the plant growth-promoting fungus *Phoma* sp. GS8-2 and the arbuscular mycorrhizal fungus *Glomus mosseae*: impact on biocontrol of soil-borne diseases, microbial population, and plant growth. *Australas Plant Pathol* 41:271–281. <https://doi.org/10.1007/s13313-011-0101-7>
- Säle V, Aguilera P, Laczko E et al (2015) Impact of conservation tillage and organic farming on the diversity of arbuscular mycorrhizal fungi. *Soil Biol Biochem* 84:38–52. <https://doi.org/10.1016/j.soilbio.2015.02.005>
- Sarr B, Ndiaye F, Ndiaye M et al (2013) Effet de deux types d'insecticides sur la mycorrhization arbusculaire et le développement de deux variétés de pomme de terre (*Solanum tuberosum*). *Int J Biol Chem Sci* 7:1902–1909. <https://doi.org/10.4314/ijbcs.v7i5.10>
- Shafiei F, Shahidi-Noghabi S, Sedaghati E (2022) The impact of arbuscular mycorrhizal fungi on tomato plant resistance against *Tuta absoluta* (Meyrick) in greenhouse conditions. *J Asia Pac Entomol* 25(3):101971. <https://doi.org/10.1016/j.aspen.2022.101971>
- Sharmah D, Jha D (2014) Diversity of arbuscular mycorrhizal fungi in disturbed and undisturbed forests of Karbi Anglong Hills of Assam, India. *Agric Res* 3:229–238. <https://doi.org/10.1007/s40003-014-0110-1>
- Shen J, Li C, Mi G, Li L et al (2013) Maximizing root/rhizosphere efficiency to improve crop productivity and nutrient use efficiency in intensive agriculture of China. *J Exp Bot* 64:1181–1192. <https://doi.org/10.1093/jxb/ers342>
- Singh LP, Gill SS, Tuteja N (2011) Unraveling the role of fungal symbionts in plant abiotic stress tolerance. *Plant Signal Behav* 6:175–191. <https://doi.org/10.4161/psb.6.2.14146>
- Singh PK, Singh M, Tripathi BN (2013) Glomalin: an arbuscular mycorrhizal fungal soil protein. *Protoplasma* 250:663–669. <https://doi.org/10.1007/s00709-012-0453-z>
- Słomka A, Kuta E, Szarek-Lukaszewska G et al (2011) Violets of the section *Melanium*, their colonization by arbuscular mycorrhizal fungi and their occurrence on heavy metal heaps. *J Plant Physiol* 168:1191–1199. <https://doi.org/10.1016/j.jplph.2011.01.033>
- Smith SE, Read DJ (2008) Arbuscular mycorrhizae. In: Smith SE, Read DJ (eds) *Mycorrhizal symbiosis*, 3rd edn. Academic, London, pp 13–187
- Smith SE, Smith FA (2011) Roles of arbuscular mycorrhizas in plant nutrition and growth: new paradigms from cellular to ecosystem scales. *Annu Rev Plant Biol* 62:227–250. <https://doi.org/10.1146/annurev-arplant-042110-103846>
- Smith SE, Facelli E, Pope S et al (2010) Plant performance in stressful environments: interpreting new and established knowledge of the roles of arbuscular mycorrhizas. *Plant Soil* 326:3–20. <https://doi.org/10.1007/s11104-009-9981-5>
- Song Y, Chen D, Lu K et al (2015) Enhanced tomato disease resistance primed by arbuscular mycorrhizal fungus. *Front Plant Sci* 6:786. <https://doi.org/10.3389/fpls.2015.00786>
- Steinkellner S, Hage-Ahmed K, García-Garrido JM et al (2012) A comparison of wild-type, old and modern tomato cultivars in the interaction with the arbuscular mycorrhizal fungus *Glomus mosseae* and the tomato pathogen *Fusarium oxysporum* f. sp. *lycopersici*. *Mycorrhiza* 22:189–194. <https://doi.org/10.1007/s00572-011-0393-z>
- Stürmer SL, Siqueira JO (2011) Species richness and spore abundance of arbuscular mycorrhizal fungi across distinct land uses in Western Brazilian Amazon. *Mycorrhiza* 21:255–267. <https://doi.org/10.1007/s00572-010-0330-6>
- Tatewaki Y, Higo M, Isobe K (2021) Community structure of arbuscular mycorrhizal fungi in the roots of maize under different of preceding winter cropping and tillage systems in south Kanto region of Japan. *Soil Microorg* 75:23–31. (In Japanese)

- Thirkell TJ, Charters MD, Elliott AJ et al (2017) Are mycorrhizal fungi our sustainable saviours? Considerations for achieving food security. *J Ecol* 105:921–929. <https://doi.org/10.1111/1365-2745.12788>
- Trappe JM, Molina R, Castellano M (1984) Reactions of mycorrhizal fungi and mycorrhiza formation to pesticides. *Annu Rev Phytopathol* 22:331–359. <https://doi.org/10.1146/annurev.py.22.090184.001555>
- Trentini CP, Campanello PI, Villagra M et al (2017) Thinning of loblolly pine plantations in subtropical Argentina: impact on microclimate and understorey vegetation. *For Ecol Manag* 384:236–247. <https://doi.org/10.1016/j.foreco.2016.10.040>
- Tsiafouli MA, Thébault E, Sgardelis SP et al (2015) Intensive agriculture reduces soil biodiversity across Europe. *Glob Chang Biol* 21(2):973–985. <https://doi.org/10.1111/gcb.12752>
- Tyagi J, Sultan E, Mishra A et al (2017) The impact of AMF symbiosis in alleviating drought tolerance in field crops. In: Varma A, Prasad R, Tuteja N (eds) *Mycorrhiza—nutrient uptake, biocontrol, ecorestoration*. Springer, Cham. [https://doi.org/10.1007/978-3-319-68867-1\\_11](https://doi.org/10.1007/978-3-319-68867-1_11)
- Uibopuu A, Moora M, Saks U et al (2009) Differential effect of arbuscular mycorrhizal fungal communities from ecosystems along management gradient on the growth of forest understorey plant species. *Soil Biol Biochem* 41:2141–2146. <https://doi.org/10.1016/j.soilbio.2009.07.026>
- Uibopuu A, Moora M, Õpik M et al (2012) Temperate forest understorey species performance is altered by local arbuscular mycorrhizal fungal communities from stands of different successional stages. *Plant Soil* 356:331–339. <https://doi.org/10.1007/s11104-011-1116-0>
- Van Eerd LL, Congreves KA, Hayes A et al (2014) Long-term tillage and crop rotation effects on soil quality, organic carbon, and total nitrogen. *Can J Soil Sci* 94(3):303–315. <https://doi.org/10.4141/cjss2013-093>
- van Geel M, Ceustermans A, van Hemelrijck W et al (2015) Decrease in diversity and changes in community composition of arbuscular mycorrhizal fungi in roots of apple trees with increasing orchard management intensity across a regional scale. *Mol Ecol* 24:941–952. <https://doi.org/10.1111/mec.13079>
- Venter ZS, Jacobs K, Hawkins et al (2016) The impact of crop rotation on soil microbial diversity: a meta-analysis. *Pedobiologia* 59(4):215–223. <https://doi.org/10.1016/j.pedobi.2016.04.001>
- Verbruggen E, Van Der Heijden MGA, Weedon JT, Kowalchuk GA, Røling WFM (2012). Community assembly, species richness and nestedness of arbuscular mycorrhizal fungi in agricultural soils. *Molecular Ecology*, 21: 2341–2353. <https://doi.org/10.1111/j.1365-294X.2012.05534.x>
- Verbruggen E, Røling WFM, Gamper HA et al (2010) Positive effects of organic farming on below-ground mutualists: large-scale comparison of mycorrhizal fungal communities in agricultural soils. *New Phytol* 186:968–979. <https://doi.org/10.1111/j.1469-8137.2010.03230.x>
- Wahdan SFM, Reitz T, Heintz-Buschart A et al (2021) Organic agricultural practice enhances arbuscular mycorrhizal symbiosis in correspondence to soil warming and altered precipitation patterns. *Environ Microbiol* 23:6163–6176. <https://doi.org/10.1111/1462-2920.15492>
- Wang FY, Shi ZY, Tong RJ et al (2011a) Dynamics of phoxim residues in green onion and soil as influenced by arbuscular mycorrhizal fungi. *J Hazard Mater* 185:112–116. <https://doi.org/10.1016/j.jhazmat.2010.09.004>
- Wang FY, Tong RJ, Shi ZY et al (2011b) Inoculations with arbuscular mycorrhizal fungi increase vegetable yields and decrease phoxim concentrations in carrot and green onion and their soils. *PLoS One* 6:16949. <https://doi.org/10.1371/journal.pone.0016949>
- Wang J, Wang GG, Zhang B et al (2019) Arbuscular mycorrhizal fungi associated with tree species in a planted forest of Eastern China. *Forests* 10(5):424. <https://doi.org/10.3390/f10050424>
- Watrud LS, King G, Londo JP et al (2011) Changes in constructed Brassica communities treated with glyphosate drift. *Ecol Appl* 21:525–538. <https://doi.org/10.1890/09-2366.1>
- Weidenhamer JD, Callaway RM (2010) Direct and indirect effects of invasive plants on soil chemistry and ecosystem function. *J Chem Ecol* 36(1):59–69. <https://doi.org/10.1007/s10886-009-9735-0>

- West TO, Post WM (2002) Soil organic carbon sequestration rates by tillage and crop rotation: a global data analysis. *Soil Sci Soc Am J* 66(6):1930–1946. <https://doi.org/10.2136/sssaj2002.1930>
- Whipps J, Hand P, Pink D, Bending GD (2008) Phyllosphere microbiology with special reference to diversity and plant genotype. *J Appl Microbiol* 105:1744–1755. <https://doi.org/10.1111/j.1365-2672.2008.03906.x>
- Wilson GWT, Williamson MM (2008) Topsin-M: the new benomyl for mycorrhizal-suppression experiments. *Mycologia* 100:548–554. <https://doi.org/10.3852/08-024r>
- Wu S, Hu Y, Zhang X, Sun Y et al (2018) Chromium detoxification in arbuscular mycorrhizal symbiosis mediated by sulfur uptake and metabolism. *Environ Exp Bot* 147:43–52. <https://doi.org/10.1016/j.envexpbot.2017.11.010>
- Xavier LJ, Germida JJ (2003) Selective interactions between arbuscular mycorrhizal fungi and *Rhizobium leguminosarum* bv. *viciae* enhance pea yield and nutrition. *Biol Fertil Soils* 37:261–267. <https://doi.org/10.1007/s00374-003-0605-6>
- Xiao D, Che R, Liu X et al (2019) Arbuscular mycorrhizal fungi abundance was sensitive to nitrogen addition but diversity was to phosphorus addition in karst ecosystems. *Biol Fertil Soils* 55:457–469. <https://doi.org/10.1007/s00374-019-01362-x>
- Yadav R, Ror P, Rathore P et al (2020) Bacteria from native soil in combination with arbuscular mycorrhizal fungi augment wheat yield and biofortification. *Plant Physiol Biochem* 150:222–233. <https://doi.org/10.1016/j.plaphy.2020.02.039>
- Yang XM, Kay BD (2001) Rotation and tillage effects on soil organic carbon sequestration in a typic Hapludalf in Southern Ontario. *Soil Tillage Res* 59(3–4):107–114. [https://doi.org/10.1016/S0167-1987\(01\)00162-3](https://doi.org/10.1016/S0167-1987(01)00162-3)
- Zaller JG, Heigl F, Ruess L, Grabmaier A (2014) Glyphosate herbicide affects belowground interactions between earthworms and symbiotic mycorrhizal fungi in a model ecosystem. *Sci Rep* 4:5634. <https://doi.org/10.1038/srep05634>
- Zangaro W, Nisizaki S, Domingos J, Nakano E (2003) Mycorrhizal response and successional status in 80 woody species from south Brazil. *J Trop Ecol* 19:315–324. <https://doi.org/10.1017/S0266467403003341>
- Zhang H-H, Tang M, Chen H, Zheng C-L, Niu Z-C (2010) Effect of inoculation with AM fungi on lead uptake, translocation and stress alleviation of *Zea mays* L. seedlings planting in soil with increasing lead concentrations. *Eur J Soil Biol* 46:306–311. <https://doi.org/10.1016/j.ejsobi.2010.05.006>
- Zhang R, Vivanco JM, Shen Q (2017) The unseen rhizosphere root–soil–microbe interactions for crop production. *Curr Opin Microbiol* 37:8–14. <https://doi.org/10.1016/j.mib.2017.03.008>
- Zhao C, Fu S, Mathew RP, Lawrence KS, Feng Y (2015a) Soil microbial community structure and activity in a 100-year-old fertilization and crop rotation experiment. *J Plant Ecol* 8:623–632. <https://doi.org/10.1093/jpe/rtv007>
- Zhao R, Guo W, Bi N, Guo J, Wang L, Zhao J, Zhang J (2015b) Arbuscular mycorrhizal fungi affect the growth, nutrient uptake and water status of maize (*Zea mays* L.) grown in two types of coal mine spoils under drought stress. *Appl Soil Ecol* 88:41–49. <https://doi.org/10.1016/j.apsoil.2014.11.016>
- Zhou L, Cai L, He Z, Wang R, Wu P, Ma X (2016) Thinning increases understory diversity and biomass, and improves soil properties without decreasing growth of Chinese fir in southern China. *Environ Sci Pollut Res* 23:24135–24150. <https://doi.org/10.1007/s11356-016-7624-y>
- Zhou H, Ouyang T, Liu L, Xia S, Jia Q (2022) In-Forest planting of high-value herb *Sarcandra glabra* enhances soil carbon storage without affecting the diversity of the arbuscular mycorrhizal fungal community and composition of *Cunninghamia lanceolata*. *Microorganisms* 10(9):1844. <https://doi.org/10.3390/microorganisms10091844>
- Zhu J-K (2003) Regulation of ion homeostasis under salt stress. *Curr Opin Plant Biol* 6:441–445. [https://doi.org/10.1016/s1369-5266\(03\)00085-2](https://doi.org/10.1016/s1369-5266(03)00085-2)
- Zobel M, Öpik M (2014) Plant and arbuscular mycorrhizal fungal (AMF) communities—which drives which? *J Veg Sci* 25:1133–1140. <https://doi.org/10.1111/jvs.12191>