

Brijendra Kumar Kashyap  
Manoj Kumar Solanki *Editors*

# Current Research Trends and Applications in Waste Management

 Springer

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*Editors*

Brijendra Kumar Kashyap  
Department of Biotechnology Engineering  
Institute of Engineering and  
Technology, Bundelkhand University  
Jhansi, Uttar Pradesh, India

Manoj Kumar Solanki  
Department of Life Sciences and Biological  
Sciences  
IES University  
Bhopal, Madhya Pradesh, India

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## About the Editors

**Brijendra Kumar Kashyap** is an assistant professor at the Department of Biotechnology Engineering, Institute of Engineering and Technology, Bundelkhand University (IET, BU), Jhansi, Uttar Pradesh, India. His graduation and post-graduation are from Banaras Hindu University (BHU), Varanasi, India. He had received numerous prestigious awards, including the Young Scientist Award of Society of Bioinformatics and Biological Sciences (SBBS), India, JRF(NET)-CSIR, SRF-CSIR, SRF-ICAR, IIT-Fellowship, ARS(NET)-ICAR, GATE, DBT-Fellowship, etc. He had delivered numerous oral and poster presentations at various national and international conferences and had published more than 24 papers in peer-reviewed journals. He is having more than 15 years of teaching and research experience.

**Manoj Kumar Solanki** is an Associate Professor at IES University in Bhopal, India, and previously served as a scientist at the University of Silesia in Katowice, Poland, from March 2021 to March 2023. He holds a Master's in Microbiology from Barkatullah University (2006) and a PhD in Microbiology from Rani Durgawati University, India (2013). Dr. Solanki's contributions have earned him recognition, including a visiting scientist fellowship from the Guangxi Academy of Agriculture Sciences, China (2013–2015), and a visiting scientist position at the Volcani Center, Agricultural Research Organization, Israel (2016–2020). His diverse research encompasses plant-microbe interactions, soil microbiology, plant disease management, and enzymology, resulting in numerous publications in international journals. Dr. Solanki's primary interest lies in agriculturally significant microorganisms, including bacteria, actinomycetes, fungi, and yeast, and their utilization for soil and crop health management.

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**Part I**

**Introductory Chapters**





# Emerging Frontiers of Microbes as Liquid Waste Recycler

1

Brijendra Kumar Kashyap, Christina Saran, Manoj Kumar Solanki, and Praveen Kumar Divvela

## Abstract

There is a worldwide energy crisis due to massive energy demand and restricted assets. This demand of energy is increasing continuously and exponentially with the increase in population. The nourishment of energy demand should be at least without affecting the environment. The human population is mainly dependent on non-renewable energy resources, especially petroleum products causing deterioration of environment, which will be exhausting in the next few decades. On the other hand, non-exhausting, self-sustainable, environment friendly, and bio-renewable energy sources are underutilized. The non-renewable energy sources are not only getting exhausted after a certain time but also cause carbon emissions to the environment, as one of the agendas in COP 27 (Conference of the parties) held in Egypt. This focuses on sustainable fuel of clean energy projects with zero carbon emission without hampering the climate condition

B. K. Kashyap (✉)

Department of Biotechnology Engineering, Institute of Engineering and Technology, Bundelkhand University, Jhansi, Uttar Pradesh, India

e-mail: [brijendrakashyap@bujhansi.ac.in](mailto:brijendrakashyap@bujhansi.ac.in)

C. Saran

Department of Biotechnology Engineering, Institute of Engineering and Technology, Bundelkhand University, Jhansi, Uttar Pradesh, India

Department of Environmental Microbiology, School of Earth and Environmental Sciences, Babasaheb Bhimrao Ambedkar University, Vidya Vihar, Raebareli Road, Lucknow, Uttar Pradesh, India

M. K. Solanki

Department of Life Sciences and Biological Sciences, IES University, Bhopal, Madhya Pradesh, India

P. K. Divvela

Contec Global Agro Limited, Abuja, Nigeria

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further with the continuously increasing population. Therefore, this chapter focuses on all these issues along with the renewable and sustainable energy sources utilizing organic liquid waste, produced from households, industry, agriculture, dairy, etc., and converting it to energy through a novel technology called microbial Fuel cell (MFC), representing a new form of renewable energy, generating bioelectricity through oxidation of waste. Thus, MFCs have the potency to treat liquid wastewater along with bioenergy generation, and various other applications. A portion of the difficulties and future points of view concerning the energy recuperation from liquid wastewater utilizing MFCs are also discussed.

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**Keywords**

Microbial fuel cell · Liquid waste · Bioremediations · Bioelectricity

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## 1.1 Introduction

The energy requirement is continuously increasing throughout the world. In this regard, fossil fuels have catered a significant portion of the energy demand. Subsequently, this has resulted in depletion of fossil fuel resources as the fossil fuel energy reservoir is in a fixed quantity. Additionally, the combustion of fossil fuels generates lots of greenhouse gas which is an alarming situation for the environment. As a result, looking for low-cost, environmentally friendly alternative energy sources has become a primary concern (Logan 2004).

Addressing waste management and global climate change issues, sustainable development is vital. With exponentially increasing energy demand and finite fossil fuel resources, new alternative sustainable energy solutions are required. In this context, achieving energy demand-treatment of municipal and industrial wastewater is also essential (Gavrilescu and Chisti 2005; Mohan et al. 2007; Li et al. 2014). One of the revived bio-electrochemical concepts and promising technology that is proposed to deal with these aspects is microbial fuel cell (MFC), which principally produce electricity from the anaerobic oxidation of biodegradable organic wastes (Madakka et al. 2020; Pant et al. 2013; Patil et al. 2012). Microorganisms are capable of converting an enormous type of biodegradable natural wastes (organic compound) into CO<sub>2</sub> (carbon dioxide), water, and energy (Potter 1911). MFCs are microbially produced energy and provide a habitat for their growth and metabolic activities (Logan 2004). A general layout of a two-chambered MFC is specified within the anodic compartment, microorganisms can bring forth oxidative conversions, and simultaneous chemical or reductive microbial processes can occur in the cathodic compartment. Electrodes of both compartments are usually separated by a proton or cation exchange membrane and interconnected through an external circuit with an external resistor or load (Rabaey and Verstraete 2005).

## 1.2 What Is Liquid Waste?

Liquid waste or wastewater is a significant problem in the world. Liquid waste may include wastewater, fats, oils, grease, used oil, fluids, solids, gases, sludge and hazardous home liquids. These wastes are hazardous or potentially harmful to human health and the environment. They can be released by commercial items assigned as “liquid industrial waste, for example, cleaning liquids or pesticides as a result of the manufacturing process” (Friedman 1981). According to the environmental protection agency (EPA), liquid waste is any waste material that approves the definition of a liquid and must pass through a 0.45- $\mu\text{m}$  filter at a pressure differential of 75 psi.

### 1.2.1 Sources of Liquid Wastes and Their Pollutants

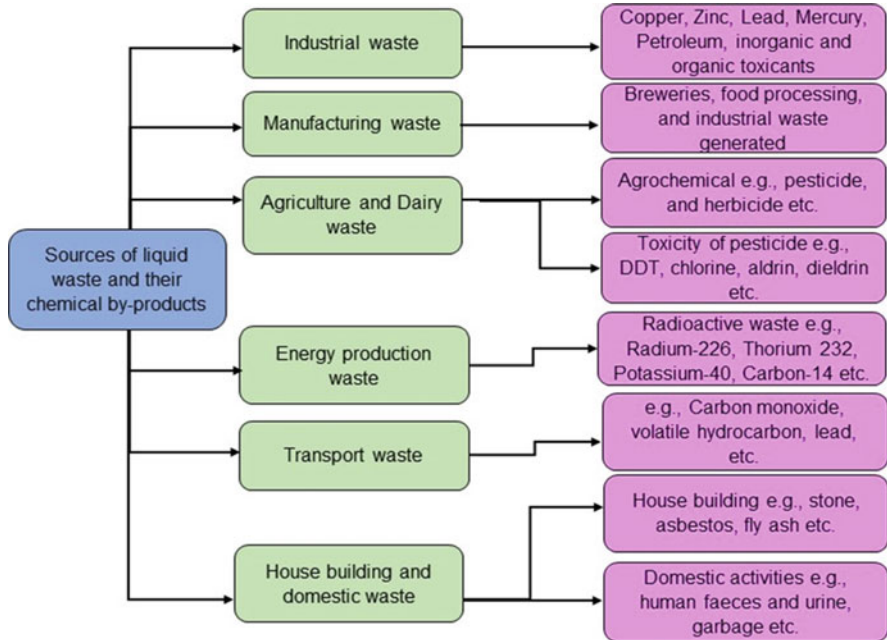
Any product, by-product or type of residue that cannot be used profitably is called waste. A waste outcome is viewed as a pollutant when it harms the environment. Also, waste and pollutants are intricately correlated. In simple words, pollutants are generally waste, but all wastes are not pollutants. Liquid waste may originate from various human activities like- Industrial waste, manufacturing industries, agriculture, dairy, energy production, transport, house building, and domestic activities, as shown in Fig. 1.1.

#### 1.2.1.1 Industrial Waste

In industrial waste, effluents are waste products in liquid forms resulting from various industrial processes. They are released by industries such as petrochemical complexes, fertilizers factories, oil refineries, paper pulp factories, textile, sugar and steel mills, tanneries, distilleries, coal washeries, synthetic material plants for drugs, fibres, rubber plastics etc. (Abbas et al. 2014a, b; Soni et al. 2020; Yadav et al. 2020). The industrial and mills include metals (copper, zinc, lead, mercury etc.), detergents, petroleum, alcohols, acids alkali, phenols, carbamate, cyanide, arsenic, chlorine and many other inorganics and organic toxicants (Devi et al. 2023). All these chemicals discharged from industry are toxic to living beings. They may cause death and sub-lethal effects on the liver, kidneys, reproductive, respiratory and nervous systems (Yadav et al. 2021).

#### 1.2.1.2 Manufacturing Waste

These activities generate a wide variety of waste depending on the nature of raw materials, products, the design and the mode of operation. Generally, manufacturing industries using biological materials (e.g., breweries, food processing, and dairy) generate biodegradable waste of biological substances (Kumar et al. 2020). Microorganisms can frequently use and recycle these biological substances. On the other hand, non-biodegradable raw materials are also used in several sectors, often not biodegradable. They may linger in the environment until it is changed or decomposed by chemical or physical factors (Leow et al. 2018).



**Fig. 1.1** Source of liquid waste with their waste release (*DDT* dichlorodiphenyltrichloroethane)

### 1.2.1.3 Agriculture and Dairy

These activities produced crop residues and manure, which are biodegradable. Most of the pesticides used in agricultural sectors are non-biodegradable. In addition, plastics and copper in feed additives and waste from fossil fuels are recalcitrant to biodegradation. The ammonia gas released from manure and fertilizers to the environment contributes to acid rain. In addition, the discharge of nitrate and phosphate chemicals into surface water and water bodies is not only leading to the formation of massive algae blooms but also contaminating groundwater, making it unfit for drinking (Badgujar and Bhanage 2018).

### Agrochemical

Agrochemicals, such as chemical fertilizer, pesticides and herbicides, contribute to heavy metal and pollution. Pesticides and weedicides are used by human beings to control Crop diseases by pests or to kill weeds and increase crop productivity. These toxic chemicals have created health hazards for livestock, wildlife, fish, other aquatic organisms, birds, mammals and humans. Ecological pesticides and herbicides have created two major serious problems which were not previously anticipated. In the first place, many of them have persisted and accumulated in the environment and have harmed or contaminated numerous animals or plants not intended to be targeted. Secondly, these directly and indirectly affect human health (Rai et al. 2020; Saleh et al. 2020).

## **Pesticides**

The toxicity of pesticides is because of Organo-chlorine pesticides (i.e., Dichlorodiphenyltrichloroethane (DDT), hexachlorocyclohexane, Chlordane, aldrin, Dieldrin, etc.). The reason for this is that sodium, potassium, and magnesium ions decrease adenosine triphosphate activity in the neuromuscular junctions of animals, particularly insects and influence the sensory system in the large zones of axon cockroaches. DDT is also known to affect the efflux of potassium ions from the axons. DDT and other organochlorine pesticides are absorbed from the intestinal tract from the alveoli of the lungs and also through the skin. If the pesticides are in solution, the high concentration of DDT causes brain damage, centrilobular liver necrosis, and liver enlargement in small mammals (Mojiri et al. 2020).

### **1.2.1.4 Energy Production Using Fossil Fuels**

The usage of enormous amounts of water across numerous power plants is essential for energy generation. The majority of power plants in the world generate energy by burning fossil fuels for boiling water. Those results in producing an excess steam inside the plant, the produced steam is used for spinning turbines. Water is also needed for the mining of coal, refining transportation fuels, and extracting petroleum sources. In once-through coal plants, the used water is typically released right back into the source (rivers, lakes, streams, and oceans) which increases the water temperature and causes thermal pollution, which alters life cycle of marine ecosystem (Jin et al. 2019). Thermal pollution of water is extremely harmful for both people and environment. Also, combustion of fossil fuels liberates carbon dioxide and significant amount of carbon monoxide, various oxides of sulphur, nitrogen and water vapours. Carbon dioxide produced during combustion is recycled by photosynthesis, but its increasing concentration in the atmosphere results in global warming. Oxides of sulphur and nitrogen cause acid rain, affecting the natural ecosystem (Tyagi and Lo 2013).

## **Radioactive Wastes**

Radioactive isotopes or radionucleotides are forms of an element with unstable atomic nuclei. They decompose with ionizing radiation in the form of Alpha or beta particles or gamma Rays. Many radioisotopes, such as radium- 226, Uranium – 235 or 238, Thorium-232, potassium-40 or carbon- 14, occur naturally. Other radioisotopes, such as Cesium, Cobalt, Iodine, Krypton, Plutonium, and Strontium, are generated industrially as fission products from atomic bomb effects, such as nuclear reactor, or other radiation-related work. Of the over 450 radioactive isotopes that can occur as fission products, only a few are of major ecological concern within the biotic community environment. These radioactive components may become scattered or collected relying on the organic movement of the component and time of radioactivity of the isotopes. However, isotopes may accumulate in human tissue just as plants, and animal radiation exposure from artificial sources are already sufficient to produce serious diseases such as leukaemia and bone tumours, genetic damage, and infant mortality (Petrangeli 2019; Kumaraswamy and Kashyap 2021).

### 1.2.1.5 Transport

This is the major contributor to atmospheric pollution by carbon monoxide, sulphur, nitrogen, volatile hydrocarbon, and lead. It also contaminates air, surface, and soil/underground water with oil and oil products (Sobieraj et al. 2022).

### 1.2.1.6 House Building and Domestic Activities

These activities generate both non-biodegradable (e.g., stone, asbestos, synthetic, fly ash, etc.) and biodegradable (sewage and various waste components) wastes. The chief waste generated by domestic activities is human faeces, urine (a component of sewage), and garbage (consisting of food scraps, plastics, cardboard, tin bottles, etc.). While sewage is biodegradable, it is discharging water bodies. Without proper treatment, it leads to spreading diseases like diarrhoea, hepatitis, etc., reducing oxygen tension or reducing anoxia in water. Separation of these components of garbage would facilitate their biodegradation and recycling. Still, it is not practised mainly due to cost considerations, and the garbage is also dumped into large bits. These activities cause pollution by generating carbon dioxide, carbon monoxide, sulphur, nitrogen, etc. (Noor et al. 2020).

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## 1.3 What Is the Problem Arising from Liquid Waste with Their Static Data?

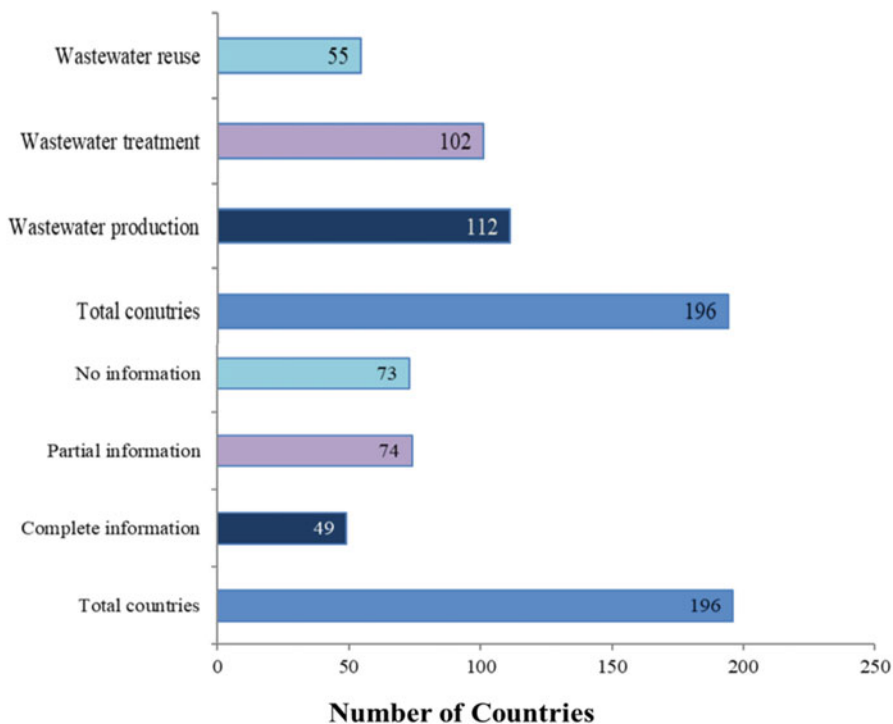
Problems arising from liquid waste are the rise in urban movement and the act of releasing untreated wastewater. The uncontrolled development in urban zones has made arranging and expansion of water usage and made sewage systems troublesome and costly to complete.

It is a typical practice to release untreated sewage into waterways or put it into the farming area, causing significant health and economic risks. The number of families with access to drinking water gracefully has expanded the percentage associated with the urban sewage collection system.

The problem with the current treatment technologies is the lack of sustainability. The conventional centralized system flushes pathogenic bacteria out of the residential area, using large amounts of water, and often combines the domestic wastewater with rainwater, causing the flow of large volumes of the path (Sato et al. 2013).

According to sources of wastewater data: Aquastat, F. A. O. (2019), there is static data from various countries based on wastewater generated, wastewater treated, and wastewater reuse. Out of 196 countries, we could get complete information in 49 countries, partial information in 74, and no information in 73 countries. Also, based on wastewater production (112 countries), wastewater treatment (102 countries), and wastewater reuse (55 countries), various countries are mentioned graphically.

Graphically representation of the above data in the form of complete, partial, and no information in Fig. 1.2a and also no. of countries whose wastewater treatment and reuse are in Fig. 1.2b.

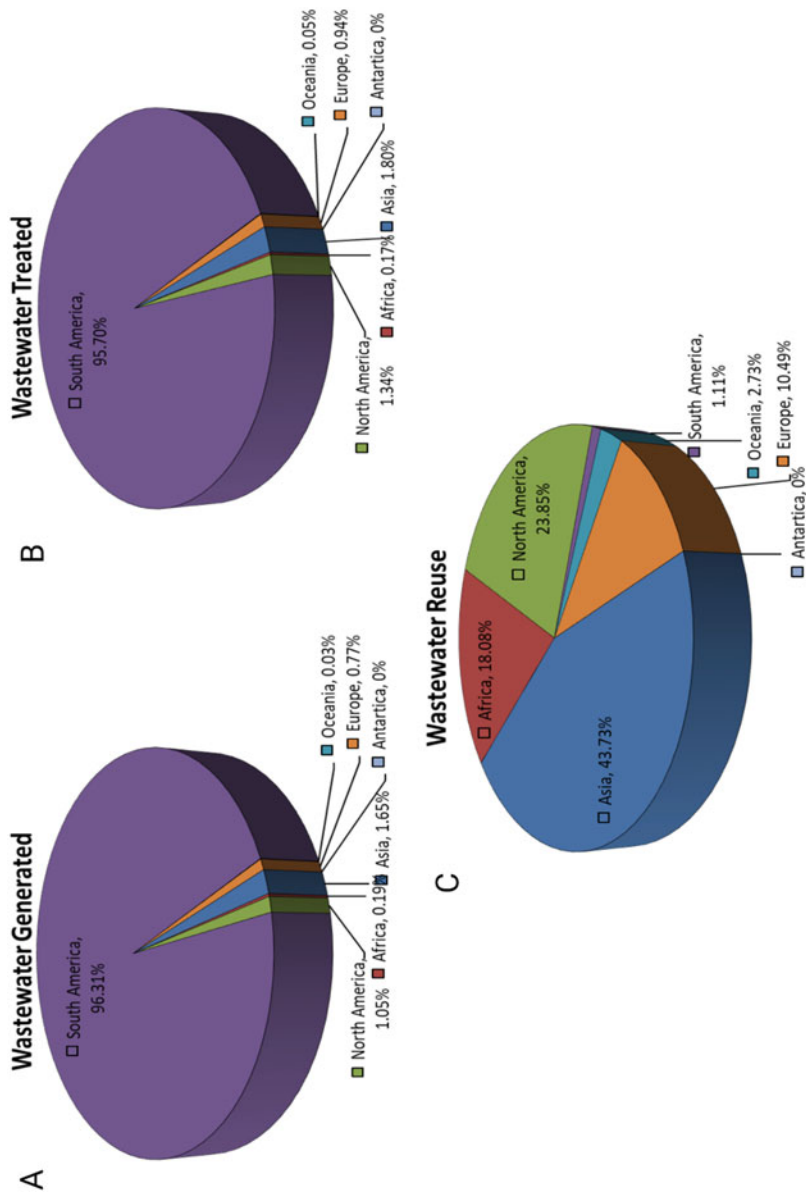


**Fig. 1.2** The availability of complete, partial, or no data on wastewater production, treatment and use and Availability of data regarding each Aspect of wastewater production, treatment, and use at the country level. (Source of wastewater data—Aquastat, F. A. O. (2019))

The graph is a pie-chart by their continents, in which the continent produced more wastewater and treated and reused it. The figure below shows the three different graph-based on wastewater generated, treated, and reused. Figure 1.3a shows the wastewater generated in which continent south America produced 96.31%. It is the maximum generated wastewater and minimum generated by Oceania at 0.03%. Figure 1.3b shows the wastewater treated in the continent south America treated 95.7%, which is the maximum treated and minimum in Oceania at 0.05% in Fig. 1.3c shows the wastewater reuse maximum reuse by continent Asia at 43.73% and the minimum continent\South America 1.11% (all these values are approximate values).

### 1.3.1 Why We Focus on Liquid Waste and How It Is Treated

The focus is on liquid waste, especially wastewater, because, as we know, the availability of sewage and sources is enormous, as well as a by-product of any material it merges with either air, water, or soil. Also, we are using wastewater because most of the industrial and domestic effluents are in a liquid form. Severe



**Fig. 1.3** a, b, and c- show the continent-wise wastewater generated, treated, and reused respectively. (Source of data: Aquastat, F. A. O. (2019))



outcomes occur when these effluent chemicals continuously discharge in the river and freshwater streams. How these chemicals or other by-products are treated or minimized in sewage waste is a question (Hussain et al. 2021).

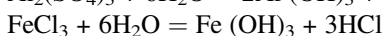
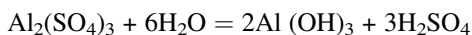
Here, we are focusing on the liquid wastewater or sewage wastewater that will be treated by conventional and advanced methods.

### 1.3.2 Conventional and Advanced Methods for Liquid Wastewater

For the conventional methods, there are some physicochemical methods such as coagulation, flocculation, precipitation, adsorption, ion-exchange, electro-dialysis, and membrane separation that can be applied in wastewater treatment schemes

#### 1.3.2.1 Coagulation and Flocculation

Coagulation and flocculation are significant physicochemical wastewater treatment activities that are used to remove turbidity particles and natural organic materials. Hydrolytic aluminium and iron salts are the most often used coagulants (Kimura et al. 2013). Optimal pH for Al (OH)<sub>3</sub> use is 4.5, and 8 for Fe (OH)<sub>3</sub>



The main disadvantage of these methods is the significant amount of chemical sludge produced. Furthermore, aluminium-based coagulants raise the residual aluminium concentration in purified water. This residual aluminium is connected with a number of issues, including increased turbidity, decreased disinfection efficiency, hydraulic capacity loss, and possible harmful impacts. This method, however, is not generally practical since it necessitates a pH rise post-treatment to prevent corrosion in water distribution networks, which increases the process's cost (Kimura et al. 2013).

Flocculation is the production of bridges between flocs followed by polymer binding of particles into big agglomerates or clumps. Filtration or flotation can then be used to remove the agglomerates. Flocculants may be made from a variety of materials, including polyferricsulfate (PFS) and polyacrylamide (PAM) (Fu and Wang 2011). Despite some turbidity, some flocculants, such as mercaptoacetyl chitosan (MAC) and flocculants based on Konjac graft-poly (acrylamide)-co-sodium xanthate, may efficiently remove heavy metal ions from wastewater. It is impossible to use a universal flocculent due to the differences in particle characteristics (Zinicovscaia and Cepoi 2016). Therefore, flocculent can be divided into several groups:

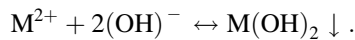
- Non-ionic, with -OH and COOH groups (natural polymers: starch, gums, glues, and alginates).
- Anionic, with -COOH and SO<sub>3</sub>H groups.

- Cationic, with  $-\text{NH}_2$  and  $=\text{NH}$  groups. Because anionic species are less costly than cationic species, they make up the majority of accessible synthetic flocculants.
- Amphoteric, with anionic and cationic groups (proteins).

Inorganic flocculants frequently result in the creation of significant amounts of sludge, whereas natural polymers are biodegradable and more effective.

### 1.3.2.2 Precipitation

Chemicals react with heavy metal ions to generate insoluble precipitate, which is then removed from the water by sedimentation or filtering (Fu and Wang 2011). Precipitation is most commonly used to remove metal ions, phosphorus compounds, and radioactive materials. Because of its simplicity, low cost, and automated pH control, hydroxide treatment is the most often utilized precipitation procedure.  $\text{Ca}(\text{OH})_2$  and  $\text{NaOH}$  compounds are used as precipitants. The mechanism of heavy metal removal by chemical precipitation can be presented by the following equation:



The major drawback of hydroxide precipitation is the creation of large amounts of low-density sludge, which causes dewatering and disposal issues. Sulphide precipitation has been shown to be superior than hydroxide precipitation. The main advantages are the high degree of metal removal even at low pH and the possibility of selective metal removal and recovery. Metal sulphide sludge also has greater thickening and dewatering qualities than metal hydroxide sludge. The process's limitations include the generation of hazardous  $\text{H}_2\text{S}$  vapours and sulphide colloidal precipitates (Fu and Wang 2011). Sometimes precipitation is used in combination with coagulation.

### 1.3.2.3 Ion-Exchange

Ion exchange is one of the most often used heavy metal removal procedures in the world. The key benefits of the ion exchange process are metal recovery, greater selectivity, and smaller sludge quantities (Zinicovscaia and Cepoi 2016). The concept is the exchange of ions in a chemically comparable amount between the solid (resin) and liquid (electrolytic solution) phases without any structural change to the resin (Kurniawan et al. 2006). The most common cation exchangers can be divided in the following groups:

- Strong acidic resins with sulfonic acid groups ( $-\text{SO}_3\text{H}$ ),
- Weak acid resins with carboxylic acid groups ( $-\text{COOH}$ ),
- Strong basic anionites containing  $-\text{NH}_2$  groups,
- Weak basic anionites containing amino groups.

Ion exchange techniques are effective for the treatment of wastewater with metal concentrations in the range of <10–100 mg/L, or even higher than 100 mg/L (Kurniawan et al. 2006).

#### **1.3.2.4 Adsorption**

Adsorption is well regarded as an efficient and cost-effective method of pollution removal from wastewater. The concentration of molecules on the surface of a sorbent characterizes the process (Owlad et al. 2009). Adsorption has considerable benefits such as low cost, high availability, profitability, flexibility in design and operation, and process reversibility (Fu and Wang 2011), which is especially relevant from an economic and environmental viewpoint. Because of its huge micropore and mesopore volumes and high surface area, activated carbon is one of the most often utilized sorbents for the removal of organic contaminants from wastewater. Activated carbon is categorized into four categories based on the manufacturing process: powder-activated carbon, granular-activated carbon, activated carbon fibrous, and activated carbon cloth, each of which has a distinct purpose (Owlad et al. 2009). Since activated carbon is a costly sorbent, it cannot be used in complex wastewater treatment systems. As a result, there is a huge potential for the creation of low-cost sorbents made from natural materials or specific waste products from industrial or agricultural activities that are cheap, plentiful, and have extremely low economic expenses (Zinicovscaia and Cepoi 2016). Conventional methods, such as coagulation, precipitation, and adsorption, are used to reduce high concentrations of various organic compounds and metal ions to regulatory required levels. Membrane technology is more efficient when pollutant concentrations are low.

#### **1.3.2.5 Membrane Filtration**

Membrane filtration has received a lot of attention in recent years because it can be used to remove pollutants from various sources. The use of membrane technology in an existing industrial process may reduce costs and overall energy consumption. Existing membrane processes include ultrafiltration (UF), nanofiltration (NF), and reverse osmosis (RO).

##### **Ultrafiltration (UF)**

UF is a procedure for separating heavy metals, macromolecules, and suspended particles from solution employing a permeable membrane with pore sizes ranging from 5 to 20 nm and separating substances with molecular weights ranging from 1000 to 100,000 Da (Fu and Wang 2011). The primary benefits of UF procedures are the lack of chemical usage and the high quality of the end product (pathogen elimination of 90–100%). Regardless, the method is hampered by the expensive expense of the membrane.

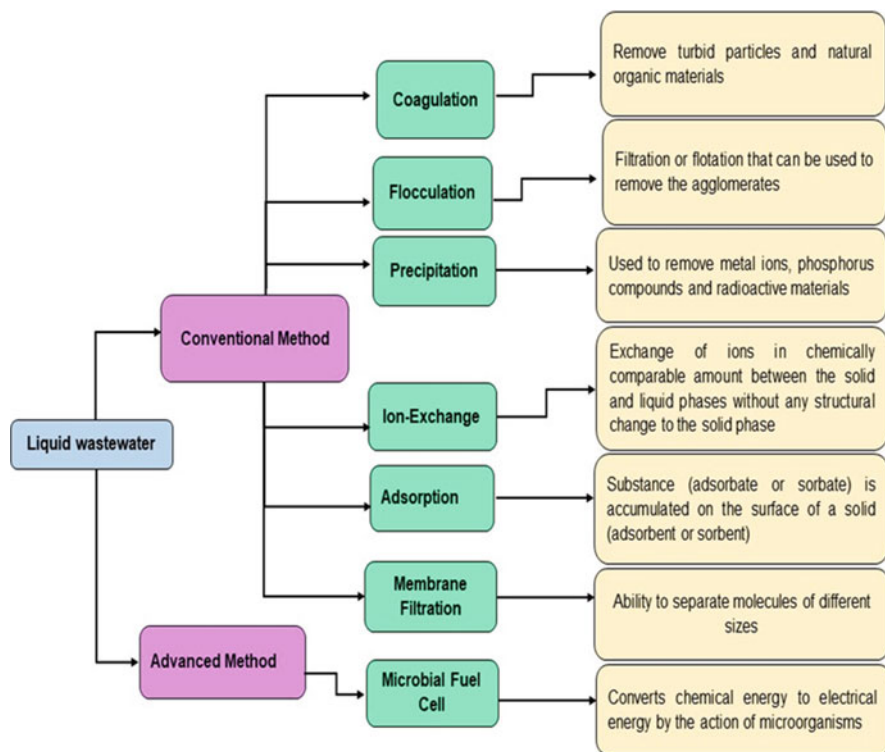
##### **Reverse Osmosis (RO)**

RO is a pressure-driven membrane technology that allows water to flow through while polluting metal ions are trapped. RO is more successful in removing metal ions

from inorganic solutions. Furthermore, the procedure works in a wide pH range of 3–11 and pressure range of 4.5–15 (Fu and Wang 2011). RO also necessitates the employment of high-pressure pumps to drive the water through the semi-permeable membranes, resulting in a reject stream that contains 95–99% of the dissolved salts. The needed pressure is proportional to the concentration of salts in the water. The method’s benefits include minimal cost and excellent efficiency. The primary drawbacks of RO are the high-power consumption caused by the pumping pressures and the costly membrane repair.

**Nanofiltration (NF)**

NF is a technology that is midway between UF and RO and is appropriate for particles with molecular sizes ranging from 0.0001 to 0.001 μm. NF permits monovalent ions to flow through while rejecting a substantial percentage of divalent cations and multivalent ions. The advantages of NF include its high efficiency, low energy consumption, and ease of use. There have been several studies on the removal of heavy metals by NF and RO membranes (Zinicovscaia and Cepoi 2016) (Fig. 1.4).



**Fig. 1.4** Conventional and advanced methods for liquid wastewater remediation

### 1.3.2.6 Advanced Method for Liquid Wastewater

The MFC is used as an advanced method because it has become an innovative renewable energy resource by degrading organic pollutants in wastewater. It is described in Sect. 1.5 of this chapter based on its physical components and its working mechanism.

## 1.4 Role of Microbes

The microorganisms involved in aerobic and anaerobic digestion and their activities are the same as those found in nature. The organic material (biodegradable components) is oxidized to carbon dioxide and water along with the production of biomass and nitrogenous compounds. In wastewater, however, the organic materials are in much higher concentration than in nature. Therefore, the microbial populations and activities are increased accordingly, providing a large surface area for biofilm formation and oxygen exchange in fixed-film processes (Solanki et al. 2020).

### 1.4.1 Aerobic Microbes

Various microorganisms occur in aerobic digestion systems. These are bacteria, protozoa, fungi, viruses, cyanobacteria, and algae.

Bacteria are the most common organisms; their number may be more than  $10^{12}$  cells/mL to  $10^9$  cells/mL).

#### 1.4.1.1 Aerobic Oxidation

Many heterotrophic bacteria are responsible for the Aerobic oxidation of organic molecules. Some important bacteria are, *Sarcina*, *Pseudomonas*, *Streptococcus*, *Salmonella*, *Escherichia*, *Staphylococcus*, *Shigella*, *Aerobacter*, etc. (Liu et al. 2021).

#### 1.4.1.2 Nitrification

Ammonium released from protein/ amino acid degradation is toxic to fish and is undesirable in river waters. Ammonium is converted to nitrate by nitrifying bacteria (*Nitrosomonas* and *Nitrobacter*). Nitrate is much less toxic than ammonia but also causes the eutrophication of river water. The presence of excess nitrate in drinking water may lead to a condition called Blue Baby syndrome in very young ones. The nitrification bacteria are slow to multiply. Therefore, when wastewater contains a high level of ammonia, care must be taken to maintain a high population of bacteria, and organic loading must be carefully regulated (Sadhukhan et al. 2022).

#### 1.4.1.3 Denitrification

The nitrate is ultimately removed from the waste by denitrifying bacteria (e.g., *Alcaligenes*, *Achromobacter*, *Micrococcus*, *Pseudomonas*, etc.). These bacteria

convert nitrate into nitrogen, which is liberated in the atmosphere. Denitrifying bacteria are strictly anaerobic; therefore, denitrification is often achieved by an anaerobic stage following aerobic digestion or by alternating aerobic and anaerobic conditions. Denitrification may also produce various oxides of nitrogen in addition to nitrogen (Dubeux and Sollenberger 2020).

### 1.4.2 Anaerobic Microbes

The anaerobic digestion processes involve a wide variety of organisms, of which bacteria are the most predominant. These microorganisms digest organic molecules, like lipids, carbohydrates, protein, etc., into methane and carbon dioxide (Laurens and Nelson 2020; Verma et al. 2018).

**Sulphate** is used as an electron acceptor by bacteria like *Desulphovibrio* during the oxidation of organic compounds, and they reduce sulphate to sulphur.

**Denitrifying bacteria** oxidized organic substrates and their nitrate as an electron acceptor and liberated nitrogen in the process. At neutral pH, nitrogen is the primary product of this process. But at acidic pH, mainly nitrogen oxides are formed (Zhang et al. 2012).

**Methanogenic bacteria** contain several cofactors not found in other bacteria. Three such Cofactors are involved in reducing carbon dioxide to methane in a stepwise fashion, beginning with methanopterin followed by methanofuran and Coenzyme M (CoM). In the end, the last reaction is catalysed by factor 430 (F430), the prosthetic group of CoM (De Mandal et al. 2020).

The **ATP generation** in methanogens is assumed to involve a proton motive force. According to one model, hydrogen is oxidized by hydrogenase on the surface of the plasma membrane to generate hydrogen ions which drive ATP synthesis. Subsequently, the hydrogen ion is used to reduce carbon dioxide inside the cells. This process also uses up the electrons generated during hydrogen oxidation by hydrogenase (De Mandal et al. 2020).

### 1.4.3 Use of Mixed Microbial Culture

When two distinct microorganisms work together, xenobiotic substances can be entirely degraded. In contrast, neither of them could accomplish this degradation on their own. *Acinetobacter*, for instance, has plasmid-borne genes for the dihydroxylation of one of the rings of 4-chlorobiphenyl, cleavage of the meta ring, and subsequent degradation to yield 4-chlorobenzoate. However, it is unable to degrade this product further. *Pseudomonas putida* strains use the Ortho Pathway to break down the 4-chlorobenzoate ring, producing acetyl-CoA and succinate in the process, but they are unable to use 4-chlorobiphenyl. *Acinetobacter* and *Pseudomonas putida*, all together, decompose the xenobiotic 4-chlorophenyl entirely but are not able to degrade it alone (Marghade et al. 2021).

One bacterium can provide the nutrients needed by another for growth. For instance, *Nocardia cyriacigeorgica* can break down cyclohexene but cannot make biotin. When *Pseudonocardia* species break down cyclohexene, and *Nocardia* cells are lysed, *Pseudomonas* uses these products to grow and release biotin species strains, but it is unable to break down cyclohexene itself. In turn, the biotin encourages *Nocardia*'s growth and cyclohexene's breakdown. Therefore, cyclohexene would be broken down if these two strains were together, but neither one could do it alone (Nawaz et al. 2011; Marghade et al. 2021).

Due to microbial interactions, the biological treatment system, or the microbial population utilized for xenobiotic breakdown, is more stable and typically achieves greater biodegradable rates (Adkins 2019).

#### 1.4.4 Bioremediation

Bioremediation strategies utilize natural frameworks to deal with toxins and are environmentally reliable and a substitute for normal decay. These procedures regularly include bioaccumulation, biosorption, bioaugmentation, and biodegradation (Devi et al. 2022; Kashyap et al. 2019; Solanki et al. 2019). Bioaccumulation is characterized as the ability of the live biomass to assemble the contaminant, which depends on biomass's resilience and take-up limits. The limitation of this procedure is that microbial development is restricted when the toxin focuses are excessively raised for bioaccumulation and such microbial cells need metabolic vitality (Robinson et al. 2001). Biosorption, for the most part, includes the adsorption wonders, any place the pollutants (adsorbate) are adsorbed against regenerative and eco-accommodating adsorbents/biosorbents. The limitation of this strategy is that it cannot be utilized for treating voluminous effluents since the issues are related to removing adsorbed biomass (Kuhad et al. 2004). Bioaugmentation is the strategy for presenting picked species which might be endogenous or exogenous to an intricate domain with contaminations (Joshi et al. 2017). The disadvantage of the bioaugmentation strategy is that the presented bacterial strain might be fruitless to develop or live as they endure some serious hindrances with the ecological toxin (Nzila et al. 2016). Biodegradation is a modest and compelling method of regarding wastewater as it is cheap, eco-accommodating, and naturally appropriate and has less slop-creating properties (Saratale et al. 2011).

#### 1.4.5 Bioremediation by Bacterial Strains

Bioremediation of natural contaminants is founded on microorganisms ordinarily present at the destinations or on microbial inoculants created in the lab and presented at the locales. Certain bacterial, fungal, and algal species are also equipped to collect toxic inorganic contaminants. However, there is no practical strategy for eliminating these microorganisms from the dirt after sequestering the inorganic particles. Therefore, bioremediation of inorganic contaminants is basically founded on appropriate

bacterial species. The biological management processes using a wide range of microorganisms (bacteria, fungi, yeast, and algae) can overcome the limitations because it is cost-effective, produces a reduced amount of sludge, and is eco-friendly to conventional physico-chemical treatment. Different trophic groups of bacteria (i.e., *Pseudomonas*, *Staphylococcus*, *Halomonas*, etc.) have been reported to accomplish a higher extent degradation of many pollutants under the most favourable conditions compared to other microbes. The bacterial method may be able to degradation of the chemical effluents in anaerobic and aerobic conditions or engage a combination of the two (Verma et al. 2021).

Tables 1.1 and 1.2 show bacterial remediation of various chemical and heavy/toxic chemicals usually present in liquid waste, which causes chemical illness and are harmful to the environment.

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## 1.5 Role of Microbial Fuel Cells (MFCs) in Wastewater Treatment

MFC (biofuel device) is a bio-electrochemical device that converts chemical energy into electrical energy by using microorganisms that act as a degradation catalyst of wastewater. The chapter mainly focuses on the use of sewage or liquid waste to produce bioenergy with the help of microorganisms (Obileke et al. 2021).

### 1.5.1 Basic Components of MFCs with Their Factors Affecting Efficiency

A regular/basic MFC comprises an anodic and cathodic chamber isolated by a proton exchange membrane (PEM)/salt bridge. The cathodic chamber usually opens directly to the air, which is shown in Fig. 1.5.

#### 1.5.1.1 Electrode Material

As a conductivity electrode, platinum, platinum black, graphite, carbon paper, graphite felt, and other materials are used. The same electrode material will be used in both chambers. The type of material that would be used in the electrode material will be shown vital effects on efficiency. For better-performing electrode material use will consistently improve the presentation of MFCs on account of various material outcomes in various enactment polarization losses (Saran et al. 2022).

#### 1.5.1.2 pH Buffer and Electrolyte

pH buffer and electrolyte used in the cathode chamber are platinum, platinum black, polyaniline, phosphate, etc. In the event that no buffer is utilized in working MFCs, at that point there will be a self-evident no pH difference between anode as well as cathode chamber. The use of electrolyte is to create a pH discrepancy which expands



**Table 1.1** Microorganisms' tendency to remediate their respective chemicals

Chemical	Microorganisms	References
Sodium (Na <sup>+</sup> )	<i>Rhodobacter sphaeroides</i> , <i>Rhodovulum</i> sp.	Sasaki et al. (2017)
Calcium (Ca <sup>2+</sup> )	<i>Bacillus licheniformis</i> SRB2	Zhao et al. (2019)
	<i>Sporosarcina pasteurii</i> , <i>Bacillus megaterium</i>	Chaparro-Acuña et al. (2018)
Magnesium (Mg <sup>2+</sup> )	<i>Bacillus licheniformis</i> SBR2	Zhao et al. (2019)
Aluminium (Al <sup>3+</sup> )	<i>Vibrio alginolyticus</i>	Purwanti et al. (2019)
Iron (Fe <sup>3+</sup> )	<i>Rhodobacter capsulatus</i> , <i>Pelobacter carbinolicus</i> , <i>Geobacter sulfurreducens</i> , <i>Gallionella capsiferiformas</i> strain ES-2	Gnanaprakasam et al. (2017)
Nitrogen ammonical (NH <sub>4</sub> <sup>+</sup> )	<i>Nitrosomonadales</i> convert NH <sub>4</sub> <sup>+</sup> to NO <sub>2</sub> <sup>-</sup> , <i>Nitrospirales</i> convert NO <sub>2</sub> <sup>-</sup> to NO <sub>3</sub> <sup>-</sup> , <i>Chlorella vulgaris</i> , <i>Bacillus cereus</i> , <i>Pseudomonas putida</i>	Maharjan et al. (2020)
		Gómez-Guzmán et al. (2017)
Carbonate (CO <sub>3</sub> <sup>2-</sup> )	<i>Cyanobacteria</i> , <i>Synechococcus</i> <i>Prochlorococcus</i>	Kamennaya et al. (2012)
Chloride (Cl <sup>-</sup> )	<i>Escherichia coli</i>	Owoseni et al. (2017)
Fluoride (F <sup>-</sup> )	<i>Bacillus flexus</i> PN4	Sakthi Thesai et al. (2020)
	<i>Providencia vermicola</i> KX926492	Mukherjee et al. (2017)
Sulphate (SO <sub>4</sub> <sup>2-</sup> )	<i>Salmonella typhimurium</i> , <i>Clostridium pasteurianum</i>	Gnanaprakasam et al. (2017)
	<i>Desulfovibrio</i> sp., <i>Desulfotomaculum</i> sp.	Piacenza et al. (2018)
Sulphite (SO <sub>3</sub> <sup>2-</sup> )	<i>Chromatium vinosum</i> (as hydrogen sulphite degrade)	Syed et al. (2006)
Nitrate (NO <sub>3</sub> <sup>-</sup> )	<i>Massilia brevitalea</i> , <i>Psychrobacter glacincola</i> , <i>Arthrobacter defluvi</i> , <i>Pseudomonas antarctica</i> , <i>Rhodobacter</i> sp.	Gnanaprakasam et al. (2017)
	<i>Chlorella vulgaris</i> , <i>Bacillus cereus</i> , <i>Pseudomonas putida</i>	Gómez-Guzmán et al. (2017)
Nitrite (NO <sub>2</sub> <sup>-</sup> )	<i>Nitrospora</i> sp., <i>Bradyrhizobium</i> , <i>Nitrospira moscoviensis</i>	Gnanaprakasam et al. (2017)
Phosphate (PO <sub>4</sub> <sup>3-</sup> )	<i>Pseudomonas</i> sp. JPSB12, <i>Enterobacter</i> sp. TPSB20, <i>Flavobacterium</i> sp. TPSB23	Paul and sinha (2015)
	<i>Chlorella vulgaris</i> , <i>Bacillus cereus</i> , <i>Pseudomonas putida</i>	Gómez-Guzmán et al. (2017)
	<i>Accumulibacter</i>	Zou et al. (2014)
Silica (SiO <sub>2</sub> )	<i>Rhodococcus</i> sp. BH4	Lee et al. (2020)
Potassium (K <sup>+</sup> )	<i>Sapindus mukorossi</i>	Jassal et al. (2015)

**Table 1.2** Microorganisms' tendency to remediate their respective chemicals/heavy metals

Chemical	Microorganism	Reference
Arsenic	<i>Pseudomonas chengduensis</i> As11, <i>Bacillus flexus</i> As12	Jebelli et al. (2018)
	<i>Pseudomonas putida</i> strain WB, <i>Geobacter lovleyi</i> , <i>Bacillus selenatarsenatis</i> , <i>Hydrogenophaga</i> sp. strain CL3, <i>Sinorhizobium</i> , <i>Arthrobacter aurescens</i> , <i>Stenotrophomonas</i> sp. strain MM7	Gnanaprakasam et al. (2017)
	<i>Klebsiella pneumonia</i> , <i>Enterobacter</i> sp.	Abbas et al. (2014a, b)
	<i>Corynebacterium glutamicum</i>	Mateos et al. (2006)
Boron	<i>E. coli</i> , <i>Enterococcus faecium</i>	Heim et al. (2015)
	<i>Candida tropicalis</i> , <i>Rhodotorula mucilaginosa</i> , <i>Micrococcus luteus</i> , <i>Bacillus thuringiensis</i> , <i>B. cereus</i> , <i>B. megaterium</i> , <i>B. pumilus</i> , <i>Pseudomonas aeruginosa</i> , <i>Aspergillus versicolor</i>	Laçin et al. (2015)
	<i>Lysinibacillus</i> sp. 21019, <i>B. horneckial</i> DSM23495, <i>Microbacterium</i> sp. CRRI-B	Raja and Omine (2013)
	<i>Variovarox</i> , <i>Shewanella</i> , <i>Mycobacterium</i> , <i>Rhodococcus</i> , <i>B. simplex</i>	Miwa and Fujiwara (2009)
Cadmium	<i>Lactobacillus plantarum</i> CCFM8610	Zhai et al. (2017)
	<i>Pseudomonas</i> sp. M3	Abbas et al. (2014a, b)
	<i>Pseudomonas aeruginosa</i> strain KUCd1	Sinha and Mukherjee (2009)
Lead	<i>Pseudomonas aeruginosa</i> ATCC27853	Babiker et al. (2020)
	<i>Providencia alcalifaciens</i> strain 2EA	Naik et al. (2013)
Cadmium +Lead	<i>Bifidobacterium longum</i> 46, <i>Lactobacillus fermentum</i> ME3, <i>Bifidobacterium lactis</i> Bb12	Halttunen et al. (2007)
Chromium	<i>Lactobacillus plantarum</i> MF042018	Ameen et al. (2020)
	<i>Klebsiella pneumoniae</i> strain MS 1.5, <i>Mangrovibacter yixingensis</i> strain MS2.4	Sanjay et al. (2020)
	<i>Lactobacillus rhamnosus</i> MTCC 1408, <i>L. casei</i> MTCC1423	Mishra et al. (2012)
	<i>Bacillus coagulans</i> , <i>Desulfomacculum reducens</i> , <i>E. coli</i> , <i>Pseudomonad</i> , <i>P. ambigua</i> G-1, <i>P. putida</i> , <i>Enterobacter cloacae</i> , <i>E. coli</i> ATCC33456, <i>Alcaligenes eutrophus</i> AE104, <i>P. fluorescens</i> , <i>B. mycoides</i> , <i>Shewanella oneidensis</i> strain MR-1	Singh (2008)
Copper	<i>Enterococcus faecium</i>	Yilmaz et al. (2010)
	<i>Geobacter metallireducens</i> , <i>Geobacter sulfurreducens</i>	Fang and Achal (2019)
Cyanide	<i>Pseudomonas pseudoalcaligenes</i> CECT5344,	Luque-Almagro et al. (2016)
	<i>Bacillus pumilus</i>	Kandasamy et al. (2015)

(continued)

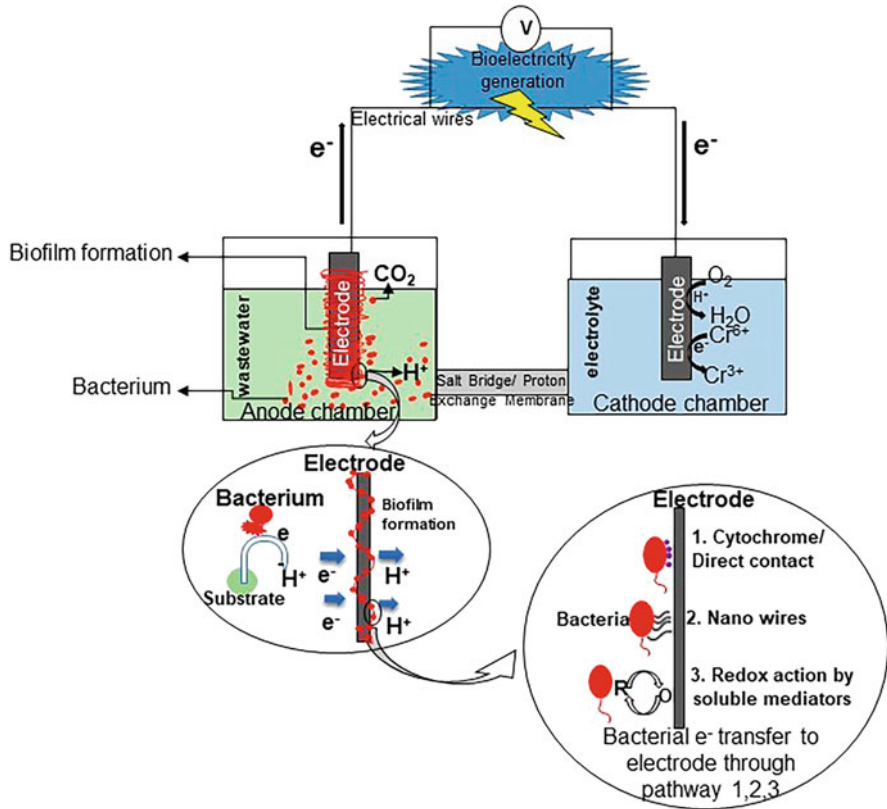
**Table 1.2** (continued)

Chemical	Microorganism	Reference
Manganese	<i>Pseudomonas putida</i> strain MnB1, <i>Pseudomonas</i> sp. strain SK3	Kitjanukit et al. (2017)
Selenium	<i>Lysinibacillus</i> sp., <i>Azospirillum</i> sp., <i>Burkholderia fungorum</i> , <i>Bacillus cereus</i> , <i>Bacillus safensis</i> JG-B5T, <i>Alishewanella</i> sp. WH16-1, <i>Stenotrophomonas maltophilia</i> SeITE02	Sinharoy and Lens (2020)
	<i>Aeromonas</i> sp.VS6, <i>Citrobactor freundii</i> KS8 <i>Pseudomonas fluorescens</i> K27, <i>Enterobacter cloacae</i> SLS1a-1, <i>R. sphaeroids</i> , <i>R.rubrum</i> S1	Piacenza et al. (2018)
	<i>Pseudomonas stutzeri</i> NT-1	Kuroda et al. (2011)
Mercury	<i>Pseudomonas aeruginosa</i> ATCC27853	Babiker et al. (2020)
	<i>Vibrio fluvialis</i>	Saranya et al. (2017)
	<i>Pseudomonas</i> sp. B50A	Giovanella et al. (2016)
	<i>Cupriavidus metallidurans</i> strain MSR3	Rojas et al. (2011)
	<i>Pseudomonas putida</i> spi3	Von canstein et al. (1999)
Zinc	<i>Bacillus megaterium</i> EMCC 1013, <i>Rhizobium rhizogenes</i> EMCC1743, <i>Rhizobium leguminosarum</i> EMCC1130, <i>Azotobacter vinelandii</i> , <i>Nocardiopsis dassenvillei</i>	El-Barbary and El-Badry (2019)
	<i>Bacterium</i> VMSDCM	Mishra et al. (2014)
Antimony	<i>Sinorhizobium</i> sp. GW3	Li et al. (2019)
	<i>Cupriavidus</i> , <i>Moraxella</i> sp. S2	Li et al. (2018)
Tellurium	<i>E. coli</i> , <i>Lactococcus lactis</i> , <i>R. capsulatus</i> , <i>R.rubrum</i> G9, <i>R. capsulatus</i> , <i>P.fluorescens</i> K27, <i>D.gigas</i> , <i>P.aeruginosa</i> ML4262, <i>Stearothermophilus</i> , <i>Mycobacterium tuberculosis</i> , <i>B.beveridgei</i> , <i>B.selenitireduceus</i> , <i>S.barnesii</i> , <i>Shewanella frigidimarins</i> ER-Te-48, <i>Bacillus</i> sp. GT-83,	Piacenza et al. (2018)
	<i>Pseudoalteromonas</i> sp. strain EPR3	Bonificio and Clarke (2014)
	<i>Stenotrophomonas maltophilia</i> TI-1 <i>Ochrobacterium anthropi</i> TI-2, <i>Ochrobactrum anthropi</i> TI-2	Kagami et al. (2012)

the main motivation of the proton spreading from the anode to the cathode chamber, which at last forms an equilibrium (Saran et al. 2022).

### 1.5.1.3 Proton Exchange Membrane (Salt Bridge)

PEM, which uses materials like Nafion, Ultrex, porcelain septum, and others, can alter the internal resistance and concentration polarization loss of MFCs, which in turn affects the power output of the MFCs. Nafion is the most well-liked due to its very selective proton permeability (Obileke et al. 2021).



**Fig. 1.5** Microbial fuel cell with its basic components. ( $CO_2$  carbon dioxide,  $O_2$  oxygen,  $e^-$  electron,  $H^+$  proton/hydrogen ion,  $Cr^{6+}$  and  $Cr^{3+}$  chromium ion)

#### 1.5.1.4 Operating Condition in the Anodic Chamber

Glass, polycarbonate, plexiglass, etc., are used for the chamber. The kind of substrate, concentration, and feed rates are crucial variables in determining how effectively MFCs work. Power density changes with the varied substrates by using a single microbe or a mixed microbial consortium. In batch or continuous flow mode MFCs, the substrate concentration determines the amount of electricity produced (Obileke et al. 2021).

#### 1.5.1.5 Operating Condition in the Cathodic Chamber

The same type of material (glass, polycarbonate, plexiglass, etc.) will be used for both chambers. In the cathode chamber, oxygen is the most commonly used electron acceptor. Power output depends on the concentration level of electron acceptors.

## 1.5.2 Mechanisms of MFCs

As we know, MFC is a bioelectrochemical device that converts chemical energy into electrical energy by the use of microorganisms which utilize the substrate (liquid waste). Also, MFCs simultaneously reuse wastewater and generate electricity. The electricity production from microbes is described as a regular/basic MFC comprising an anode and cathode compartments separated/ distant by a PEM/salt bridge. Microbes in the anode chamber metabolize the organic compounds or substrate, which acts as an electron donor. The metabolism of these organic compounds generates electrons and protons. The electrons first transfer to the anode surface and second migrates via an electrical circuit to the cathode. On the other hand, the flow of protons first migrates to the electrolyte or buffer solution via the PEM/salt bridge. This electron and proton are consumed in the cathode reduction by the electron acceptors and, after that, bioelectricity generation (Chaturvedi and Verma 2016).

## 1.5.3 Types of MFCs

They are broadly classified as a mediator and mediator-less MFC.

### 1.5.3.1 Mediator MFCs

A large portion of the microbial power devices is electrochemically indolent. The mediators strengthen the electrons moving from MFCs to the electrode, such as thionine, methyl viologen, methyl blue, humic acid, or another chemical that enhances the electron transfer. Also, most of the mediators accessible are expensive and toxic.

### 1.5.3.2 Mediator-Less MFCs

This type of MFCs does not require a mediator but electrochemically active bacteria to transfer an electron to the electrode. These electrons are conveyed straightforwardly from the bacterial respiratory catalyst to the electrode. Mediators- less MFCs are a later region of study. Because many aspects determining optimal efficiency, such as bacteria strain, type of PEM, pH, and so on, are poorly understood, Mediators-less MFCs are a later area of study (Kumar et al. 2017).

## 1.5.4 Research Organization on MFCs

### 1.5.4.1 International Status

During the last couple of decades, extensive basic/ fundamental research work has been carried out in many institutes worldwide, a glimpse of which is presented here. The accelerated rate of publication, particularly during the last decade, is quite evident in Fig. 1.5, presented below.

The research in Bio-Energy & Environmental Biotechnology at the Energy and Biotechnology Department of Ecological and Biological Engineering of Oregon State University includes electricity generation using MFCs and Hydrogen production using microbial electrolysis cells (MECs). Currently, the group focuses on reactor design, membrane/cloth selection, electrode development, isolation of exoelectrogens, and system optimization to improve power generation and hydrogen production from various waste biomass. In May 2009, the Department of Earth Sciences at the University of Southern California, Los Angeles, published a paper titled “Electricity production coupled to ammonium in a microbial fuel cell” (He et al. 2009).

MFCs offer great promise for simultaneous wastewater treatment and renewable energy generation. The Penn State group, led by Dr. Bruce Logan, focuses primarily on MFC architecture and factors that will lead to successful scale-up designs. They used air-cathode and aqueous (dissolved oxygen) cathode systems to understand better factors that limit power generation and examine how power density can be increased while using low-cost yet effective materials.

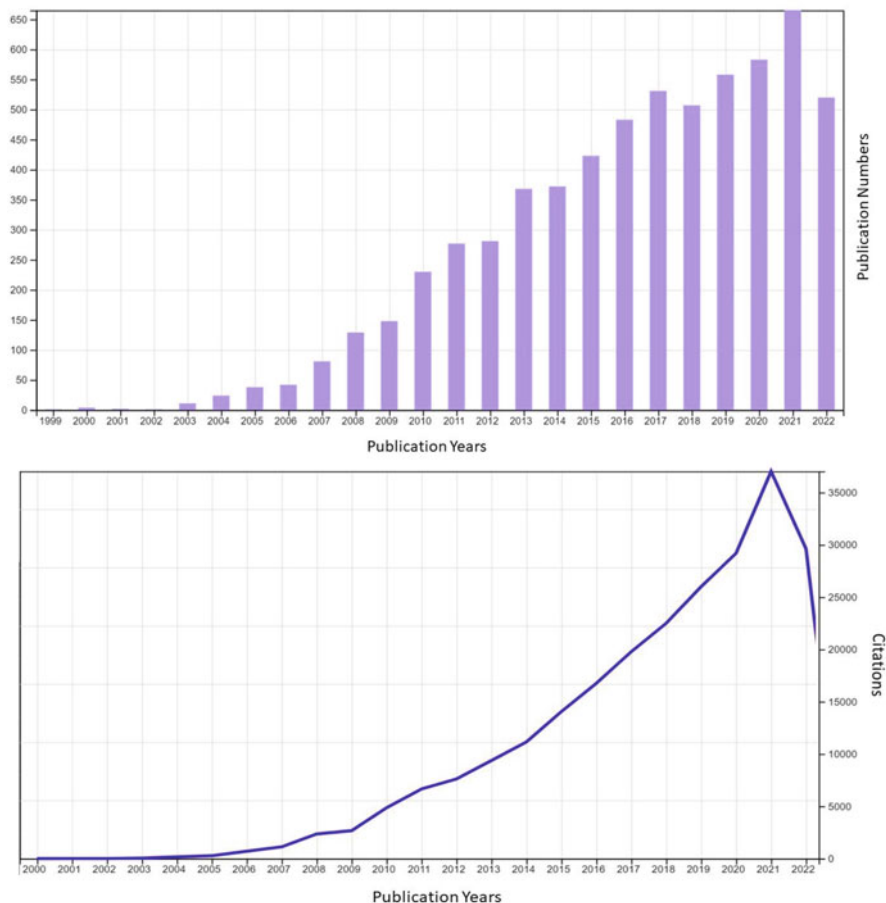
Below is a list of various international institutes working on MFCs.

- Penn State University (USA)—The Logan Group.
- Medical University of South Carolina (MUSC) (USA)—May Lab.
- Gwangju Institute of Science and Technology (Korea) The Energy and Biotechnology Laboratory (EBL).
- Harbin Institute of Technology (HIT) (China) School of Municipal and Environmental Engineering, Advanced Water Management Centre.
- The University of Queensland, St. Lucia, Australia.
- Istituto per l’Ambiente Marino Costiero (IAMC) IST-CNR Section of Messina, Messina, Italy.
- Department of Earth Sciences, University of Southern California, Los Angeles, California.
- Dépt. deGénie Chimique, Ecole Polytechnique de Montréal, Centre-Ville, Montréal, QC, Canada.
- School of Chemical Engineering and Advanced Materials, Merz Court, Newcastle University, Newcastle upon Tyne, UK.
- US Naval Research Laboratory Washington, DC (USA)—The Ringeisen Group.

#### 1.5.4.2 National Status

R&D on biofuel has started more recently (since the year 2000) in India. The rate of publication has accelerated during the last few years, as shown in Fig. 1.6. It is evident that there are only a few institutes which are involved in biofuel cell development, as listed below:

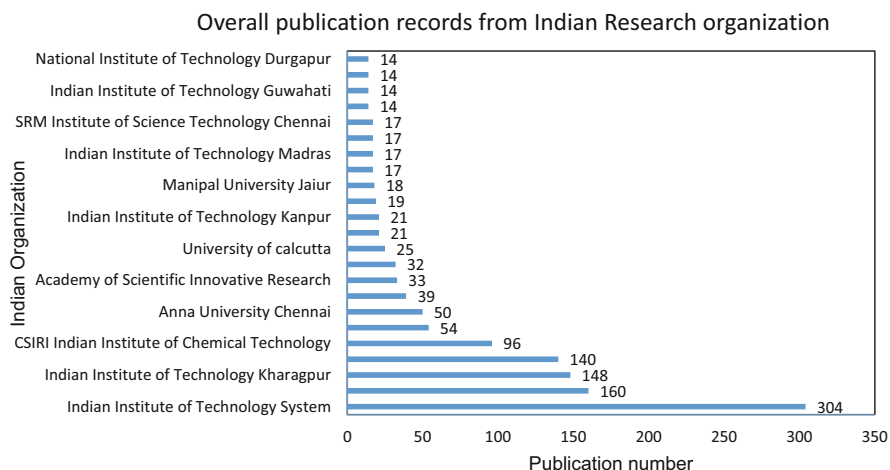
- Indian Institute of Chemical Technology, Bioengineering and Environmental Centre (BEEC), Hyderabad, India.
- Biotechnology Department, IIT Madras, Chennai, India.
- Indian Institute of Technology Delhi, New Delhi.



**Fig. 1.6** Histogram shows the year-wise worldwide research publication on microbial fuel cells with their citation analysis

- Indian Institute of Technology Bombay, Mumbai.
- Vellore University.
- Department of Civil Engineering, Indian Institute of Technology, Kharagpur.
- Central Electrochemical Research Institute, Karaikudi, Tamilnadu, India (Ministry of New and Renewable Energy, Government of India 2016).

The last few years have seen considerable research activity in India's biofuel cells, mainly via R&D work sponsored by the MNRE, DST, CSIR, etc. PEM Fuel cell uses an extensive range of materials. Such materials are electrocatalysts, catalyst support, gas diffusion media, microporous materials, hydrophobic materials, hydrophilic materials, different types of carbon, electrolyte, sealants, and conducting coating materials as shown in Fig 1.7 and Table 1.3.



**Fig. 1.7** Total number of research publications on microbial fuel cells from different institutes of India (<https://www.webofscience.com>)

**Table 1.3** Depiction of various work organizations and the various forms of work they execute

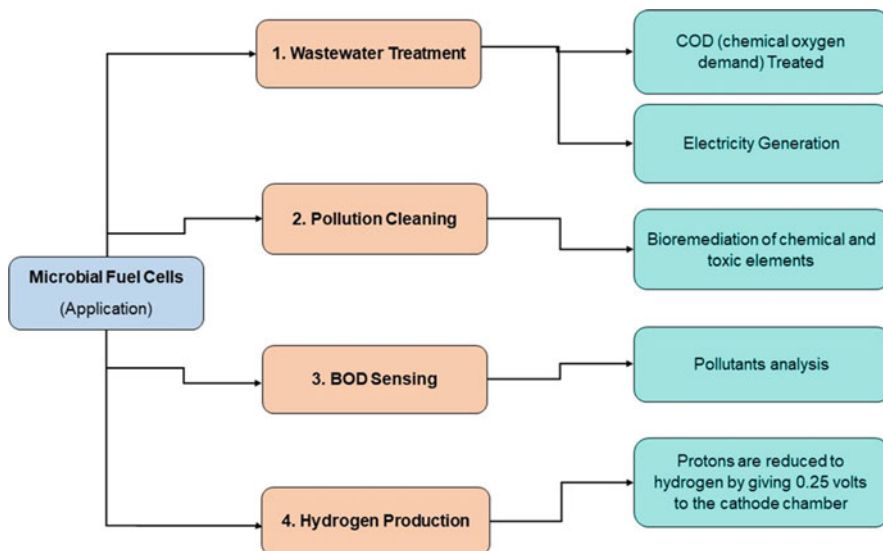
Work organizations	Nature of work
IIT-M, NCCR, IIT-B, IIT-G, IIT-K, IIT-Kh, IIT-R, IIT-H, IISc, BESU, CSIR-CECRI, CSIR-NCL, CSIR-NPL, CFCT-ARCI, CIPET, CSIR-CSMCRI, BITS-Goa, TU, AIIST, PSGIAS, Anna University, UoH, DTU, and many other Universities	<ul style="list-style-type: none"> <li>• Basic Science</li> <li>• Catalysts, Membrane, Bipolar plate</li> <li>• Modelling</li> </ul>
BHEL, CSIR-CECRI, CFCT-ARCI, IIT-B, SSF (closed), ISRO Labs & Def. Labs	<ul style="list-style-type: none"> <li>• Stack and System</li> <li>• Application demonstration</li> </ul>
Tata, M&M, TVS, REVA, NMRL, some CSIR Labs, IITs, BPCL, RIL	<ul style="list-style-type: none"> <li>• System integration using bought-out stacks for demonstration</li> <li>• Demonstration of indigenously</li> </ul>

*IIT* Indian Institute of Technology, *CSIR* Council of Scientific and Industrial Research, *TU* Tamilnadu University, *BITS* Birla Institute of Technology and Science, *DTU* Delhi Technical University

### 1.5.5 Application on Microbial Fuel Cell

Although MFCs have been studied as an alternative energy source, their application is restricted to certain zone only. With further upgrades in configuration, cost-visibility, and execution proficiency are dependent on these close-to-term applications, as shown in Fig. 1.8. It is conceivable to scale up and use MFCs as an environmentally friendly power asset. The clearest utilization of MFCs is the abundance of power. They can be used in the rural area and the urban segment. Even though power generation using energy components has not been very successful on a small scale, large-scale application can be successful. These have a conversion efficiency of fuel to the power of request of 70% or more and are not limited to





**Fig. 1.8** Applications of microbial fuel cells in different areas

the Carnot cycle. Higher energy recovery of 80% to 97% has been accounted for. An ideal approach is to use is to store the electricity in a rechargeable battery.

### 1.5.5.1 Wastewater Treatment

The microbes can generate power while also decomposing effluents. MFCs are straightforwardly under genuine thought as gadgets to deliver electrical force all through the treatment of mechanical, agribusiness, and metropolitan wastewater. When microorganisms oxidize natural compounds in the wastewater, electrons are delivered, yielding a consistent quantity of electrical current. Suppose the power age in this framework can be extended. In that case, MFCs may give another strategy to offset the operating costs of wastewater treatment plants, making advanced wastewater treatment more moderate in both making and industrialized nations. Moreover, MFCs are also mentioned to create less waste when contrasted with the high-sway treatment measure (Li et al. 2014).

### 1.5.5.2 Cleansing Contaminated Lakes and Rivers

MFCs can be used in the bioremediation of water containing characteristic contaminations, for example, toluene and benzene mixes found in gas. The MFCs configuration is changed so the power device floats on the head of contaminated water. The anode is lowered in the water where natural toxic feed the microorganisms, and the cathode float on topor head of the water. Normal pollution is the degeneration of carbon dioxide and water, purifying the contaminated lake or stream. The MFCs can be excused in distant common water waterways, many equivalents to the remote sensor (Chen et al. 2022).

### 1.5.5.3 Biological Oxygen Demand (BOD) Sensing

Another possible use of the MFCs innovation is to use it as a sensor for contamination examination and in situ measure noticing and control. Biological oxygen demand (BOD) is the proportion of split oxygen expected to meet the metabolic necessities of high-impact life structures in water rich in natural issues, for instance, sewage. The related association between the coulombic yield of MFCs and the centralization of adapting characteristic impurities in wastewater makes MFCs possibly usable as BOD sensors. An MFC-type BOD sensor can be saved operational for over 5 years without extra help for more organizational life length than such a BOD sensor detailed in the literature (Do et al. 2020).

### 1.5.5.4 Hydrogen Production

Hydrogen creation by modified MFCs fragmenting away at natural waste may be fascinating for other options. In such a gadget, anaerobic conditions are kept up in the cathode chamber, and an additional voltage of 0.25 volts is to the cathode. Under such conditions, protons are decreased to hydrogen on the cathode. Such adjusted MFCs are named bio-electrochemically helped microbial reactors. (Vishwanathan et al. 2013).

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## 1.6 Challenges of MFCs

MFCs, a promising innovation for power generation by utilizing waste material, experience numerous difficulties that obstruct their commercialization. A fraction of the significant void openings of this innovation are as per the following: -

- The power density obtained by xenobiotics and waste is very low compared to pure carbon sources. This hinders its applicability in waste management and electricity generation for day-to-day purposes. (Chaturvedi and Verma 2016).
- Pure carbon sources cannot be routinely employed for electricity generation because they are expensive compared to waste. (Chaturvedi and Verma 2016).
- The material used for a cathode/ anode and membrane during the scaling up of MFCs is costly and suppresses its commercialization.

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## 1.7 Conclusions and Future Prospects

### 1.7.1 Conclusions

Wastewater is perceived as making a significant commitment to natural contamination. MFCs are an innovation for the creation of power from the metabolism of the microorganism. In this chapter, we interact with considerable liquid waste and its xenobiotic substances, such as a chemical parameter hazardous compound that is extremely dangerous to the environment and toxic to the organism.

MFCs are used for power generation and are transformed into less toxic compounds, which exhibited its other possible use in waste management and pollution control. A large number of microorganisms and a waste assortment of the substrate (including xenobiotics) have been utilized in the creation of power. A significant drawback of this innovation is that power output is very low, and scaling up reductions in power output. This is the principal motivation behind why this innovation has not yet been popularized. Thus, a great deal of work is required so this innovation gets proficient, appropriate, and generally acknowledged.

### 1.7.2 Future Prospects

MFCs are a promising innovation for generating energy using natural substances, particularly from a diverse natural waste source. In any case, there are sure disadvantages, which have impeded making it more material when reasonable applications are concerned. The major drawback of MFCs and possible solutions which can help to enhance the efficiency of MFCs. Drawbacks like low power density can be improved by isolating microorganisms that can transfer electrons to the anode or by generating recombinant strain that shows more excellent electron transfer rates. For electron transfer, many reports have confirmed that a relatively pure culture, a consortium of bacterial cultures, will improve electron transfer. Also, many bacterial cultures produce mediators which efficiently transfer electrodes to the anode. Another drawback is the limited surface area of the electrodes where microorganisms adhere. Studies have been performed on MFC reactors and have resulted in designing more efficient laboratory-scale MFCs. These include the use of air cathode, stacked reactor, and cloth electrode assemblies.

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# Municipal Wastewater Treatment by Microalgae with Simultaneous Resource Recovery: A Biorefinery Approach

# 2

Vishal Singh, Bhola Prasad, and Vishal Mishra

## Abstract

An increase in urbanization and industrialization has led to the increased discharge of wastewater, especially municipal wastewater, causing eutrophication as a large amount of wastewater is discharged into the water bodies without proper treatment. Current municipal wastewater treatment is carried out using the conventional activated sludge process (CAS), where indigenous microbial consortia with external aeration reduce organic matter. But critical issues are associated with the CAS process, including high energy requirements, generation of sludge, and emission of a large amount of carbon dioxide. Therefore, there is a need for alternative strategies in order to deal with these issues. Microalgae-based wastewater treatment process has emerged as a promising alternative technology for treating municipal wastewater. Microalgae offer certain advantages such as sequestration of atmospheric carbon dioxide, effective treatment of wastewater, and resource recovery in the form of microalgal biomass. The current chapter deals with the advancement made during these years for municipal wastewater treatment, including membrane technology, biofilm technology, and photo-sequencing batch reactors. There are also certain disadvantages associated with microalgae-based wastewater, such as scale-up, contamination in raceway ponds, and high energy requirements during the harvesting and dewatering process. In order to recover these costs, a biorefinery approach has been proposed where the microalgal biomass generated during the treatment process is transformed into various products such as biofuel, biochemical, and bioelectricity.

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V. Singh · B. Prasad · V. Mishra (✉)  
School of Biochemical Engineering, IIT (BHU), Varanasi, India  
e-mail: [vishal.bce@itbhu.ac.in](mailto:vishal.bce@itbhu.ac.in)

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**Keywords**

Microalgae · Circular bioeconomy · Wastewater treatment · Bioremediation · Biofuel

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**Abbreviations**

ASP	Activated sludge process
CAS	Conventional activated sludge
CO <sub>2</sub>	Carbon dioxide
COD	Chemical oxygen demand
DIC	Dissolved inorganic carbon
IEA	International Energy Agency
LI	Light intensity
MPBR	Membrane photobioreactor
MR	Mixing rate
N	Nitrogen
NH <sub>4</sub> <sup>+</sup> -N	Ammonium
O <sub>2</sub>	Oxygen
P	Phosphorus
PBR	Photobioreactor
PO <sub>4</sub> <sup>3-</sup> P	Phosphate
RAB	Revolving algal biofilm
TAN	Total ammonia nitrogen
Temp.	Temperature
TKN	Total kjeldahl nitrogen
TN	Total nitrogen
TP	Total phosphorus

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**2.1 Introduction**

Rapid industrialization and urbanization have led to the increased exploitation of natural resources by releasing a large amount of wastewater and greenhouse gases (GHGs). The report of International Energy Agency (IEA) fuel combustion 2019 highlights that 2.2, 4.8, and 9.8 Metric gigatons of CO<sub>2</sub> were emitted by India, the United States, and China alone. The high emission of GHGs triggers climate change and global warming (Arun et al. 2020b). The next disadvantage of industrialization and urbanization is the release of different types of wastewater generated from textile and pharmaceutical industries, agricultural lands, domestic, and municipal wastewater (Zhang et al. 2017; Kadir et al. 2018; Rai et al., 2020; Lellis et al. 2019). The wastewater is rich in various types of nutrients, including both inorganic (macronutrients and micronutrients) and organic nutrients (carbon compounds).

When they are discharged into the freshwater sources without the proper treatment, causing the problem of eutrophication poses a threat to the natural ecosystem of the freshwater bodies (Bhatia et al. 2020). It was estimated that eutrophication causes a loss of two billion dollars per year as it severely affects fishing and real estate activities (Lavriničs and Juhna 2018).

A large portion of wastewater released every year is constituted by municipal wastewater generated from the urban colonies, institutional setup and small-scale industries (Daverey et al. 2019). The conventional treatment of municipal wastewater is carried out by the activated sludge process (ASP) mediated via the biological approach. In the ASP process, organic matter in the wastewater is degraded via indigenous consortia of microbes and  $O_2$  is supplied to them via an external aeration system. The microbial population in the reactor is maintained via a recycling system that recycles back a portion of sludge into the reactor (Daverey et al. 2019). The main disadvantage of the ASP process is the requirement of a high amount of energy ( $0.3\text{--}0.6\text{ kWh/m}^3$ ), constituting about 26% of the net cost of the treatment process (McCarty et al. 2011; Li et al. 2017). The aeration process alone consumes 47–70% of the total energy required by the treatment process. There have been some advancements in the aeration process. Still, the consumption of a large amount of energy by the ASP process remains a major issue (Gikas 2017). Another critical issue of the ASP process is the disposal of a large amount of activated sludge generated during the process. Removal of per kg of chemical oxygen demand (COD) generates about 0.3–0.5 kg of dry biomass of activated sludge (Liu et al. 2018). The sludge can be utilized in the energy recovery process, but its handling process, which includes thickening, dewatering, and digestion process, consumes about 30% of the total plant energy (Zhou et al. 2013). The third and last critical issue of the ASP process is releasing a large amount of  $CO_2$  during the oxidation process of organic matter by microbes (Singh et al. 2016).

To resolve the issues explained above, microalgae-based treatment of municipal wastewater proved to be a promising technology for the advanced treatment of wastewater with simultaneous recovery of nutrients (Li et al. 2019; Singh and Mishra 2021, 2022). Microalgae are the rapidly growing photoautotrophs that utilize sunlight as energy and  $CO_2$  as a carbon source with the release of  $O_2$  and generate a large amount of biomass (Singh and Mishra 2019). Their  $CO_2$  fixation efficiency is 10 to 50 times higher than terrestrial plants (Langley et al. 2012). In recent years they have been applied to treat municipal wastewater by growing them in open raceway ponds or closed photobioreactors (Daverey et al. 2019). The ample amount of inorganic nutrients such as nitrogen and phosphorus and low toxic elements in municipal wastewater makes it a highly suitable medium for microalgae cultivation (Craggs et al. 2013). Some of the advantages offered by microalgae-based wastewater treatment are given as (1) Overall wastewater treatment is reduced as microalgae can assimilate almost every pollutant with resource recovery; thus, there is no need for additional treatment; (2) the pollutant level in the treated water by microalgae has a deficient level of pollutants satisfying the discharge limit criteria (Whitton et al. 2015); (3) microalgae can efficiently grow in the municipal wastewater with or without the need of external nutrient supplementation (Clarens et al. 2010);

(4) when microalgae are grown in symbiosis with bacteria during the treatment process, they provide  $O_2$  required for oxidation of organic matters by bacteria, thus eliminating the need of external aeration device (Jia and Yuan 2018); (5) microalgal biomass generated the end of the process can be further transformed into biofuels, biogas, fertilizers and feedstock for animals (Raheem et al. 2015; Singh and Mishra 2019). However, various challenges are also associated with microalgae-based wastewater treatment, which include contamination in open raceway ponds, scale-up of closed photobioreactors, the significant cost involved in the harvesting and dewatering process, which incurs about 3–15% of the total cost of the treatment process (Razzak et al. 2017; Fasaei et al. 2018). This cost can be overcome by biorefinery or bio-circular economy approach in which a microalgae-based wastewater treatment process is integrated with the production of energy and other valuable products, as explained in detail in Sect. 2.3 (Bhatia et al. 2020).

Therefore, the current chapter's objective is to provide insights into the recent advancements in the treatment of municipal wastewater by microalgae. It further covers the prospective details of the biorefinery approach for decreasing the treatment process cost.

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## 2.2 Recent Advancements in the Treatment of Municipal Wastewater by Microalgae

Various advancements have been made to treat municipal wastewater by microalgae, including the microalgae-bacterial process, photo-sequencing batch reactor, membrane and biofilm technology, and synchronization of microalgae with yeast and macrophytes explained in the upcoming sections. Table 2.1 represents various microalgal species utilized to treat municipal wastewater with the removal efficiencies of various pollutants and biomass concentrations.

Figure 2.1 represents a schematic diagram for integrating conventional activated sludge process with microalgae technology for the treatment of municipal wastewater and simultaneous production of biomass and transforming it into biofuel, representing a biorefinery concept.

### 2.2.1 Microalgal-Bacterial Process

The microalgal-bacterial process is becoming an alternative method of choice for the treatment of municipal wastewater other than the conventional activated sludge process (CAS), as it demands low energy, low cost, easy operation, and the potential of resource recovery in the form of biomass feedstock (Mata et al. 2010; Quijano et al. 2017; Zhang et al. 2020a). They are a self-sustainable system with mutual synchronization between the microalgae photosynthesis and bacterial respiration processes. Microalgae feed upon the inorganic nutrients such as nitrogen and phosphorus present in the wastewater and assimilate the carbon dioxide generated during bacterial respiration, releasing oxygen. Bacteria then utilize the generated

**Table 2.1** Treatment of municipal wastewater by various microalgal species and removal efficiencies

Microalgae species	Photobioreactor configuration	Experimental condition	Pollutant removal efficiency/removal rate	Biomass concentration/productivity	Biofuel type and concentration	Reference
<i>Nannochloropsis gaditana</i> DEE03	250 mL Erlenmeyer flasks (Batch) and 1 L flat PBR (bioethanol production)	0%–100% wastewater in filtered f/2 medium Temp: $24 \pm 2$ °C pH: 7.8 Air flow rate: 0.5 L/min LI: 80 $\mu\text{mol}/\text{m}^2/\text{s}$	–	$2.33 \pm 0.12$ g/L	89.0 $\pm$ 4.0 mg/g (bioethanol)	Onay (2018)
<i>Pantanalimema rosaneae</i>	Glass reactor	pH: $7.0 \pm 0.2$ LI: 200 $\pm$ 10 $\mu\text{mol}/\text{m}^2/\text{s}$	Influent organics: 92.69%, ammonia: 96.84%, phosphorus: 87.16%	–	–	Ji et al. (2020)
<i>Chlorella sorokiniana</i>	1 L Erlenmeyer flasks	Room temp. Mixing speed: 120 RPM LI: 120 $\mu\text{mol}/\text{m}^2/\text{s}$	Ammonium: 94.29% Phosphate: 83.30%	77.14 mg/L/d	24.91 mg/L/d (lipid)	Ramsundar et al. (2017)
<i>Nannochloropsis oculata</i> and <i>Tetraselmis suecica</i>	250 mL Erlenmeyer flasks	Temp.: $24 \pm 2$ °C Mixing rate: 150 RPM pH: 8.5 LI: 1300 lm	–	1.285 g/L ( <i>N. oculata</i> ) 1.055 g/L ( <i>T. suecica</i> )	–	Reyimu and Özçimen (2017)
<i>Hindakia tetraclotoma</i> ME03	1 L Flat PBR	Air flow rate: 0.5 L/min LI: 150 $\mu\text{mol}/\text{m}^2/\text{s}$	–	$0.78 \pm 0.01$ g/L	$11.2 \pm 0.3$ g/L (bioethanol)	Onay (2019)

(continued)

Table 2.1 (continued)

Microalgae species	Photobioreactor configuration	Experimental condition	Pollutant removal efficiency/removal rate	Biomass concentration/productivity	Biofuel type and concentration	Reference
<i>Chlorella</i> sp.	14 L stirred PBR	pH: 7.4 Temp.: 24 ± 1 °C Room temperature LI: 100 µmol/m <sup>2</sup> /s Mixing rate: 100 RPM	COD: 37.5–45.7%	1.12 g/L	–	Nguyen et al. (2020)
<i>Chlorella</i> sp.	Open raceway pond	pH: 8 Temp.: 24 °C	–	3.6 ± 0.12 g/L	0.925 ± 0.1 g/L	Ashokkumar et al. (2019)
<i>Chlorella zofingiensis</i>	10 L glass column reactor and 240 L outdoor plate bioreactor	8% pig bio-gas slurry in MW Temp.: 25 ± 1 °C LI: 150 µmol/m <sup>2</sup> /s 5% CO <sub>2</sub> mixed air ventilation	TN: 93% TP: 90%	2.5 g/L	–	Zhou et al. (2018)
<i>Chlorella</i> sp.	1 L PBR	pH: 7.33 ± 0.06 LI: 80 µmol/m <sup>2</sup> /s Temp.: 23–27 °C MR: 150 RPM	NH <sub>4</sub> <sup>+</sup> -N: 23.25 ± 1.59 mg/L/d PO <sub>4</sub> <sup>3-</sup> -P: 8.05 ± 0.83 mg/L/d	117.1 ± 2.7 mg/L/d	17.2 ± 0.2 mg/L/d	Cho et al. (2017)
<i>Chlorella pyrenoidosa</i>	1 L cylindrical reactor	Aeration rate: 50 mL/min Temp.: 25 ± 2.0 °C LI: 10,000 lux	NH <sub>4</sub> <sup>+</sup> -N: 91.7%	70 mg/L/d	27.3 mg/L/d (lipid)	Zhou et al. (2020)

<i>Chlorella</i> , diatoms and filamentous cyanobacteria	2 L photo-sequencing batch reactors (PSBR)	Flow rate: 0.7 L/cycle MR: 200 RPM Temp.: 22.2 °C	COD: 87 ± 5% TKN: 98 ± 2% NH <sub>4</sub> <sup>+</sup> -N: 99 ± 3% P accumulation: 9.82 mg/L	–	–	Foladori et al. (2018)
<i>Chlorella pyrenoidosa</i>	1 L PBR	Temp.: 22–28 °C LI: 8000–80,000 lx		0.749 g/L	0.197 g/L	Wang et al. (2019)
<i>Chlorella vulgaris</i>	1.5 L photo-sequencing bioreactors	MR: 50 RPM Temp: 24 ± 2 °C LI: 45 µmol/m <sup>2</sup> /s	COD: 89 ± 4% NH <sub>4</sub> <sup>+</sup> -N: 99 ± 1%	1.1 ± 0.3 g/L	–	Petrini et al. (2020a)
<i>Nostoc ellipsosporium</i>	1 L closed transparent reactors	LI: 2500–6500 lx Aeration rate: 0.05–0.2 vvm Room temp.	N: 87.59% P: 88.31%	2.9 g/L	24.62 wt% (bio-oil yield)	Devi and Parthiban (2020)
<i>Scenedesmus</i> sp.	80 L high rated algal pond	Temp.: 20 °C Paddle speed: 10 RPM Liquid velocity: 0.2 m/s	TN: 60 ± 5% COD: 89 ± 3% P-PO <sub>4</sub> <sup>3-</sup> : 28 ± 7%	12.7 g/m <sup>2</sup> /d	–	Arcila and Buitrón (2017)
<i>Chlorella sorokiniana</i>	50 L Flat panel PBR	LI: 196 µmol/m <sup>2</sup> /s Aeration rate: 0.6 vvm Temp.: 0.6 vvm	Organic matter removal: > 90% DIC: 46–56% PO <sub>4</sub> <sup>3-</sup> -P: 40–60% NH <sub>4</sub> <sup>+</sup> -N: 100%	1 g/L	–	Leite et al. (2019)
<i>Chlorella sorokiniana</i>	2 L integrated sequencing batch reactor system	Temp.: 24 ± 2 °C	COD: 99% TKN: 88% PO <sub>4</sub> <sup>3-</sup> -P: 91% NH <sub>4</sub> <sup>+</sup> -N: 90%	45 mg/d	–	Kotoula et al. (2020)

(continued)



Table 2.1 (continued)

Microalgae species	Photobioreactor configuration	Experimental condition	Pollutant removal efficiency/removal rate	Biomass concentration/productivity	Biofuel type and concentration	Reference
<i>Chlorella vulgaris</i>	Cylindrical glass reactors	Temp.: 25 °C LI: 2000 lx MR: 300 RPM	COD: 93% PO <sub>4</sub> <sup>3-</sup> -P: 91% NH <sub>4</sub> <sup>+</sup> -N: 90%	1.96 g/L	–	Amini et al. (2020)
<i>Scenedesmus obliquus</i>	1 L Erlenmeyer flasks	Temp: 25 ± 2 °C LI: 100 µmol/m <sup>2</sup> /s	TP: 95.72% TN: 80.30% NH <sub>4</sub> <sup>+</sup> -N: 87.25% COD: 85.43%	0.891 ± 0.012 g/L	0.477 ± 0.073 g/L	Qu et al. (2020a)
<i>Scenedesmus</i> sp.	5 L batch polyethylene terephthalate (PET) bioreactors	Air flow rate: 0.5 VVM pH: 7	Nitrate: 96% TAN: 100% PO <sub>4</sub> <sup>3-</sup> -P: 3%	0.98 ± 0.10 g/L	–	Walls et al. (2019)
<i>Tetraselmis</i> sp. NKG2400013	Flat-shaped glass flasks	Temp.: 25 °C LI: 130 µmol/m <sup>2</sup> /s	N: 98 ± 0% P: 82 ± 2%	157 ± 5 mg/L/d	5.5 ± 1.8 mg/L/d	Aketo et al. (2020)
<i>P. kessleri</i> NKG021201		Air flow rate: 0.8 L/L/min	N: 98 ± 0% P: 20 ± 3%	101 ± 1 mg/L/d	39 ± 1 mg/L/d	
<i>C. Saccharophilum</i> NKH13			N: 99 ± 0% P: 39%	127 ± 9 mg/L/d	35 ± 10 mg/L/d	
<i>Chlorella sorokiniana</i>	Conical flasks	Temp.: 25 °C LI: 4000 lux	N: 100% P: 39.3%	25.0 ± 0.1 mg/L/d	–	Chen et al. (2020)
<i>Chlorella vulgaris</i>	1 L MPBR	LI: 101.5 to 112.3 µmol/m <sup>2</sup> /s Air flow rate: 0.5 L/min pH: 6.8–7.6	–	1.84 g/L 1.72 g/L	25.76 mg/L/d 29.57 mg/L/d	Gao et al. (2019)
<i>Scenedesmus obliquus</i>		Temp.: 25–28 °C				
<i>Chlorella pyrenoidosa</i>	1 L Erlenmeyer flask	Temp.: 20.65 °C, pH: 7.72 LI: 2500 lux	NH <sub>4</sub> <sup>+</sup> -N: 98.72% PO <sub>4</sub> <sup>3-</sup> -P: 76.29%	5.36 g/L	–	Singh and Mishra (2020)

<i>Chlorella pyrenoidosa</i>	PBR	LI: 86.5 $\mu\text{mol}/\text{m}^2/\text{s}$ pH: 7.56 Temp.: 30 °C	N: 96.7% P: 98%	1.001	–	(Gao et al. 2021)
Microalgae consortia	Raceway Pond	LI: 428.52 $\mu\text{mol}/\text{m}^2/\text{s}$ pH: 8 Temp.: 16.5 °C	N: 99% P: 90.16%	0.601	–	Lage et al. (2021)

Temp. temperature, LI light intensity, MR mixing rate,  $\text{NH}_4^+$ -N ammonium,  $\text{PO}_4^{3-}$ -P phosphate; N nitrogen, P phosphorus, COD chemical oxygen demand, TAN Total Ammonia Nitrogen, TN Total Nitrogen, TP Total Phosphorus, DIC dissolved inorganic carbon, TKN Total Kjeldahl Nitrogen, PBR photobioreactor



sludge process was developed to eliminate these limitations that utilized engineered microalgal-bacterial granules. The process successfully achieved high REs of 96.84%, 92.69%, and 87.16% for ammonia, organic components, and phosphorous, respectively, within 6 h of operation. No external aeration was supplied to the process (Ji et al. 2020). They also concluded that a mutually symbiotic relationship occurred between the microalgae and bacteria, which was essential in obtaining the above results and self-sustaining the system for a longer time (Ji et al. 2020).

Another limitation in applying the microalgal-bacterial process was the design process of PBR, as the kinetics and parameters used for the ASP may not be applicable for the PBR (Brindley et al. 2010; Qu et al. 2020b). The reason for this can be the difference in the PBR's growth and decay rate of the microalgal-bacterial process (Decostere et al. 2016). Therefore, a method based on the respirometry approach was used by Petrini et al. (2020b) to determine the kinetics of the microalgal-bacterial consortium treating municipal wastewater (Petrini et al. 2020b). Respirometry is a cheap and fast method in which the process's DO (dissolved oxygen) concentration is continuously measured via an automated system. After that, the DO curve is plotted from which the net Oxygen Uptake Rate (OUR, considered negative) of the consortium and net Oxygen Production Rate (OPR, considered positive) of the microalgae are calculated by the slope of the curve. At last, the gOPR (gross oxygen production rate) is calculated by the difference between OPR and OUR (Tang et al. 2014; Ippoliti et al. 2016). Based upon the calculation of Petrin et al. (2020), gOPR was found to be  $9.8 \pm 0.2 \text{ mg O}_2 \text{ g TSS}^{-1} \text{ h}^{-1}$  and this  $\text{O}_2$  was applied for the degradation of COD at the maximum rate of  $19.3 \text{ TSS}^{-1} \text{ h}^{-1}$  (Petrini et al. 2020b).

### 2.2.2 PSBR (Photo-Sequencing Batch Reactor)

The application of the microalgal-bacterial consortium for wastewater treatment has been further extended in photo-sequencing batch reactors (PSBR). An ASP comprising of sequencing batch reactor (SBR) has been applied for the treatment of municipal and agro-industrial wastewater at low and medium scales (Sirianuntapiboon et al. 2005; Wang et al. 2011). SBR offers advantages such as high RE, flexible operation, and an effective control system (Dionisi et al. 2001). Microalgae have been introduced in the SBR process to form a synergistic microalgal-bacterial system to improve its potential for resource recovery. Such an SBR system is called PSBR (Liu et al. 2017). Foladori et al. (2018) cultivated a microalgal-bacteria consortium in PBR to treat municipal wastewater and also evaluated DO, pH, and ORP profiles. No external aeration was supplied to the reactor, and RE of  $87 \pm 5\%$  for COD and  $98 \pm 2\%$  for total kjeldahl nitrogen (TKN) was obtained (Foladori et al. 2018). However, it should also be noted that an appropriate amount of microalgae inoculum should be supplied to the reactor to maintain the system's excellent performance, as the introduction of microalgae impacts the original microbial flora (Ye et al. 2018). When the microalgae concentration is above  $4.60 \text{ mg Chl/L}$ , it will inhibit the growth of certain bacteria phylum,

including Bacterioidetes and Actinobacteria, and hamper the stable operation of PSBR (Ye et al. 2018).

### 2.2.3 Supplementation of External Nutrient Source

It has been reported that low-nutrient concentration in municipal wastewater limits its application for microalgae cultivation (Chu et al. 1996). Leite et al. (2019) also reported that municipal wastewater they received from the centralized Brazilian system was highly diluted and not fit for microalgae cultivation both technically and economically (Leite et al. 2019). One of the methods applied to increase the nutrient concentration was the supplementation of artificial nutrient media, which will increase the overall production cost (Lv et al. 2010; Phukan et al. 2011; Itoiz et al. 2012; Lam and Lee 2013; Miriam et al. 2017). Biogas slurry can prove to be an alternative nutrient supplementation source instead of artificial nutrient media. It contains a high amount of nutrients, thus reducing nutrient limitation in municipal wastewater (Wang and Lan 2011). Zhou et al. (2018) cultivated *Chlorella zofingiensis* in the municipal wastewater where pig biogas slurry was supplied as the sole supplementation source of nutrients (Zhou et al. 2018). Their study reported that keeping the concentration of pig biogas slurry up to 8% in municipal wastewater produced significant results. REs of up to 93% for total nitrogen (TN) and 90% for TP were obtained with a 2.5 g/L concentration of biomass and increased lipid productivity by 8% compared to the BG11 medium (Zhou et al. 2018). The problem of nutrient limitation can also be solved by mixing municipal wastewater with another source of wastewater that may have a high-nutrient concentration, such as livestock effluent (Leite et al. 2019). Leite et al. (2019) carried out the pilot-scale cultivation of *Chlorella sorokiniana* in the flat panel PBR by mixing municipal wastewater with piggery wastewater. Biomass concentration reached up to 1 g/L with 46–56% REs for DIC, 40–60% for orthophosphate, and 100% for ammonia (Leite et al. 2019).

Utilization of the tail gas of the power plant to meet the demand for inorganic carbon sources during the cultivation of microalgae in wastewater has gained much importance during these years (Packer 2009; Ho et al. 2010; Sydney et al. 2010; Yoo et al. 2010; Lam et al. 2012). The use of tail gas increases biomass and lipid productivity and is also helpful in successfully sequestering CO<sub>2</sub> from the environment (Tu et al. 2019). During the cultivation of *C. pyrenoidosa* in the wastewater, tail gas was supplied from the power plant, which increased dry biomass weight and lipid productivity by 84.92% and 74.44%, respectively. Their study also suggests that pretreatment of tail gas by desulfurization and denitrification is also needed in order toxic material (Tu et al. 2019).

### 2.2.4 Membrane Photobioreactor

In the membrane photobioreactor (MPBR), a membrane made up of microfilters is equipped in the PBRs (Gao et al. 2014). Membrane act as a solid-liquid barrier during the cultivation of microalgae in semi-continuous or continuous mode. The filtration module eliminates the problem of a washout as microalgal cells can be retained for a longer duration of time with the continuous and ample supply of wastewater (Honda et al. 2012; Singh and Thomas 2012; Gao et al. 2014; Sun et al. 2018). As hydraulic retention time (HRT) is increased in the MPBR, wastewater containing low-nutrient concentration can also be used to cultivate microalgae (Gao et al. 2016, 2018; Sheng et al. 2017). They also offer other advantages, such as high sludge concentration, high RE, and small footprint (Sun et al. 2018). Several studies have reported that the biomass productivity of microalgae in MPBR is higher than in conventional PBR (Honda et al. 2012; Gao et al. 2014, 2018). Gao et al. (2019) cultivated two green microalgae strains, *Chlorella vulgaris* and *Scenedesmus obliquus*, in MPBR using municipal wastewater having a low-nutrient concentration in the continuous mode (Gao et al. 2019). The result indicated that even though the low-nutrient medium was used for cultivation, the lipid content was increased by 29.8% and 36.9% in *C. vulgaris* and *S. obliquus*, respectively, thus proving MPBR a valuable tool for cultivating microalgae in a low-nutrient medium (Gao et al. 2019). The application of MPBR was further extended to treat wastewater by microalgae-bacteria consortia (Amini et al. 2020). *Chlorella vulgaris* and bacterial inoculum from activated sludge were cultivated in MPBR in semi-continuous mode. RE of 93%,  $88 \pm 1\%$ , and  $84 \pm 1\%$  for COD,  $\text{N-NH}_4^+$ , and  $\text{P-PO}_4^{3-}$ , respectively, were obtained. Also, the biomass concentration reached up to 1.96 g/L. Thus, the above results indicated that MPBR is useful in both semi-continuous and continuous modes (Amini et al. 2020).

### 2.2.5 Biofilm Technology

One of the significant problems that hinder the scale-up of the microalgae cultivation system is a less efficient harvesting system, as microalgal cells have low separability in the suspended cultures (Zhu et al. 2017a, b). To tackle this, biofilm technology has been developed in which the microalgal cells are grown on the carrier surface and can be easily separated from the effluent (Wang et al. 2017, 2018a, b). After that, cells are mechanically separated from the carrier surface (Wang et al. 2018a, b). Biofilm technology performs the wastewater treatment process more efficiently and economically as they possess a high mass transfer rate and high penetration efficiency of light (Mantzorou and Ververidis 2019). Carriers supporting microalgal cell growth play an essential role in biofilm technology. Various biofilm technology that has been applied both at lab and pilot scale includes rotating biofilm reactors (Christenson and Sims 2012), algal turf scrubber (Wang et al. 2018a, b), and vertical biofilm reactors (Podola et al. 2017). Zhang et al. (2018) modified the traditional raceway pond by introducing vertical algal biofilm and accessed its efficiency for

wastewater treatment and biomass production (Zhang et al. 2018). Their results showed that this modified raceway pond could efficiently remove COD, TN, and TP at 7.52, 6.76, and 0.11 g/m<sup>2</sup>/day removal rates. Moreover, lipid productivity reached 7.47–10.10 tonnes/hectare/year (Zhang et al. 2018). In another study, revolving algal biofilm (RAB) reactors were used to treat wastewater generated after sludge sedimentation at pilot scale mode. RE of 80% and 87% were obtained for TP and TKN, respectively, while 100% RE was obtained for NH<sub>4</sub><sup>+</sup>-N and PO<sub>4</sub><sup>3-</sup>-P (Zhao et al. 2018).

But the reported carriers used for the biofilm technology are expensive in nature. Therefore, the study has shifted towards cheap carriers such as natural materials that include loofah sponge (Zhang et al. 2019), filter papers (Aljerf 2018), jute (Cao et al. 2013), linen (Kesaano and Sims 2014), etc. One of the added advantages of these materials is that they have micropores and various functional groups on their surface that function as adsorbent surfaces and are involved in the nutrient removal process with the microalgal cells (Riahi et al. 2017). Zhang et al. (2020b) designed a PBR in which pine sawdust was used as a biofilm carrier and assessed its efficiency for treating both synthetic and real wastewater (Zhang et al. 2020b). Their results showed that RE of 95.54% for TN and 96.10% for NH<sub>4</sub>-N<sup>+</sup> was obtained in real wastewater and biomass productivity reached up to 8.10 g/m<sup>2</sup>/day. Pine dust acted as a carrier for algal cells and performed the role of adsorbent as it removed 23.60% of COD, 37.30% of TN, 41.08% of NH<sub>4</sub><sup>+</sup>-N, and 17.07% of total phosphorus (TP) (Zhang et al. 2020b).

## 2.2.6 Synchronization of Microalgae with Other Species

Earlier in Sect. 2.1, the application of the microalgal-bacterial process has been discussed in detail as several researchers have focused on its application for wastewater treatment. Microalgae have also been used in synchronization with other species for wastewater treatment. Some of them have been explained in the upcoming sections.

### 2.2.6.1 Microalgae-Yeast Process

Yeast species are widely used in the baking, brewing, and pharmaceutical industries. But its application for wastewater treatment has not been thoroughly evaluated due to the assumption that it will not grow to its full potential in the non-sterile environment of wastewater (Walls et al. 2019). But the P and N content in the yeast cells are 3–5% and 10%, respectively, higher than the content in microalgal cells (0.87%: P; 6%: N) (Walker 1998; Dalrymple et al. 2013). Thus, yeast can remove the nutrients from the wastewater at a higher RE. Yeast also has good settling properties that can decrease the cost of the harvesting system (Walls et al. 2019). Therefore, the application of microalgae-yeast cells for wastewater emerged as a hot research topic during these years. The synergetic relationship between microalgae and yeast occurs in the same way as the microalgal-bacterial process (i.e., O<sub>2</sub> generated during the photosynthetic process of microalgae used by yeast for respiration in turn

generates CO<sub>2</sub>). Yeast cells can also trap the microalgal cells during harvesting, thus decreasing the cost of harvesting and dewatering. Walls et al. (2019) cultivated the *Scenedesmus* sp. and wild yeast in co-culture mode in a heterotrophic bioreactor, and they showed that this co-culture was efficient in 100% total ammonia nitrogen (TAN), 96% nitrate, and 93% orthophosphate. The biomass concentration of *Scenedesmus* sp. and yeast reached up to  $0.98 \pm 0.10$  g/L and  $4.2 \pm 0.1$  g/L, respectively (Walls et al. 2019). Yeast also offers the added advantage that it can be applied for aerobic fermentation for bioethanol production.

### 2.2.6.2 Microalgae-Macrophytes Process

*Lemna minor* belongs to the family of Lemnaceae, characterized as floating microphyte and smallest angiosperms having a rapid multiplication rate (Ekperusi et al. 2019). It is usually applied at the tertiary stage of the wastewater treatment process to treat effluent generated from the secondary treatment plant, mainly to remove toxic micropollutants and biomass production (Gatidou et al. 2017). It has also been applied for nitrogen removal, showing a high nitrogen uptake rate (Toyama et al. 2018). Recently, the co-culture of microalgae and macrophytes gained much importance for treating municipal wastewater by combining their synergistic effects. Kotoula et al. (2020) cultivated *Chlorella sorokiniana* UTEX 1230, *Lemna minor* in a SBR, and RE was 99% for COD and 88% for TKN, respectively 90% for NH<sub>4</sub><sup>+</sup>-N, and 91% for PO<sub>4</sub><sup>3-</sup>-P. *C. sorokiniana* was able to completely remove the COD while partially removing N and P. On the other hand, *Lemna minor* mainly contributed to the removal of nitrogen (Kotoula et al. 2020).

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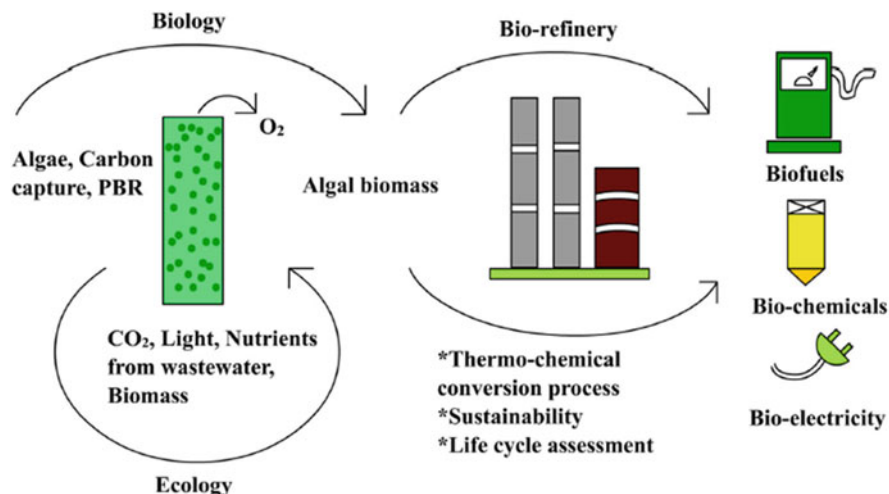
## 2.3 Microalgal Biorefinery Perception

As discussed earlier, high energy and cost are required during the microalgae-based wastewater treatment process, especially during the harvesting and dewatering process. The microalgae biorefinery approach (Fig. 2.2) has been proposed to compensate for the cost, where the microalgal biomass is transformed into various liquid and gaseous fuels, as explained below.

### 2.3.1 Liquid Biofuels

The demand for sustainable energy sources is increasing daily due to the increment of fuel load for the community, global warming effects, and decreasing petroleum reserves. In this context, liquid biofuels play a crucial role because they can put back fossil fuels and diminish carbon dioxide emissions (Williams and Laurens 2010). Some examples of liquid biofuels are bioethanol and biobutanol, which are fermentative biofuel that is derived from carbohydrates present in microalgal biomass.





**Fig. 2.2** Integration of microalgae-based wastewater treatment process with biorefinery concept (Arun et al. 2020b)

### 2.3.1.1 Bio-Oil

Bio-oil is obtained by pyrolysis and hydrothermal liquefaction (HTL) of biomass which refers to thermochemical conversion that leads to the polymerization of organic matter in an anaerobic environment (Sun et al. 2020). Initial steps of biomass degradation include degrading it into smaller compounds either individually or in combination with dehydrogenation, dehydration, decarboxylation, and deoxygenation. The obtained molecules are unstable and highly reactive, leading to cyclization, condensation, and polymerization, resulting in oily compounds and a great variety of molecular weight distribution (Arun et al. 2020b). Yang et al. (2007) noted that the quality of Bio-oil depends on the constituents of plant biomass like cellulose, hemicellulose, and lignin. It was found that cellulose, hemicellulose, and lignin degradation occurred at a temperature range of 220–315 °C, 314–400 °C, and 160–900 °C, respectively, and generated high solid residue (40%) (Yadav et al. 2020; Yang et al. 2007).

### 2.3.1.2 Biodiesel

In 1900, Rudolf Diesel initiated the production of methyl esters (commonly known as diesel) involving crops (Ramadhas et al. 2005). He considered it biodegradable, sustainable, and non-lethal (Demirbas and Fatih Demirbas 2011). Biodiesel consists of an extended chain of methyl ester and is renewable, non-hazardous, and eco-friendly fuel produced by oxidation and disintegration of biomass. Microalgae have been accepted as a good source of biodiesel production because of their high lipid content (50–70%) and multiplication rate (Satputaley et al. 2017). Biodiesel is highly viscous, due to which it accumulates on the fuel injector of the engines.

Processes like pyrolysis, dilution, and emulsification decrease viscosity (Marchetti et al. 2007).

Transesterification is a process through which triglycerides are converted into biodegradable, low atomic weight fatty acid methyl esters (FAMES) compounds suitable for engines. In the presence of methanol or ethanol, the rate of reaction is increased. Biodiesel production depends on the temperature, reaction time, catalyst load, and alcohol concentration (DuPont 2013). It was reported that transesterification, in combination with ultrasonication, reduces the reaction time that results in decreased working costs (DuPont 2013).

### 2.3.1.3 Bioethanol

It is the preferable liquid biofuel processed from the saccharification of carbohydrates and then alcohol fermentation (Ho et al. 2012). In alcohol fermentation, the components like starch, sugar, and cellulose present in biomass are converted into the fermentative fuel through the metabolic process of fungi, bacteria, or yeast in anaerobic conditions (Costa and de Morais 2011; Yadav et al., 2020). The United States Environmental Protection Agency reported that biofuels are receiving more attention all over the globe, in which bio-ethanol was the preferable biofuel in the last 10 years (Madakka et al. 2020). For the industrial fermentation process, *Saccharomyces cerevisiae* is the preferable strain (Suali and Sarbatly 2012). Through the glycolytic pathway, sugar converts into pyruvate followed by acetaldehyde synthesis, and carbon dioxide is liberated as a by-product. The produced acetaldehyde is then reduced to synthesize ethanol (Costa et al. 2015). In a study, it was mentioned that glucose resulted in ethanol (0.51 kg) and CO<sub>2</sub> (0.49 kg) per kg of substrate used (Hamed 2015). Another study reported that microalgae like *Chlorella vulgaris* yield around 65% ethanol converted from 37% starch content per dry cell weight (Brennan and Owende 2010). The anaerobic fermentation process for bioethanol production for algal biomass is a simple and easy process compared to other fermentative techniques.

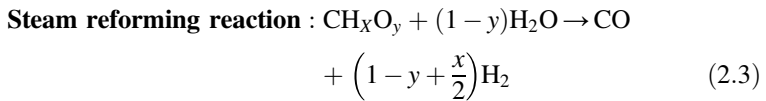
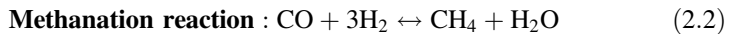
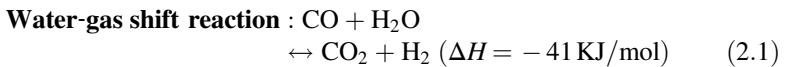
### 2.3.1.4 Biobutanol

In Liquid biofuels, biobutanol provides a high energy profile and may also bring back bioethanol in the future (Vivek et al. 2019). Yeast like *Clostridium acetobutylicum* can digest biomass feedstock (cellulose and starch) and produce biobutanol. Along with biobutanol, they also produce some valuable by-products like ethanol, acetone, and organic acids. Under favourable fermentation conditions, the maximum yield of biobutanol was 0.41 g/g of glucose; unexpectedly, it is less than bioethanol yield (0.5 g/g of glucose) (Chen et al. 2013). Biobutanol production is increased by adding butyrate into acetone-butanol-ethanol (ABE) fermentation because it enhances the metabolic route from acidogenesis to the solvent genesis acetoacetyl-CoA is transformed to butyl Co-A instead of acetoacetate (Kao et al. 2013).

## 2.3.2 Gaseous Biofuels

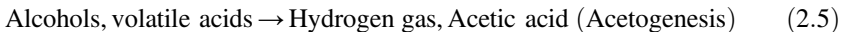
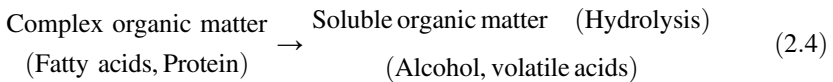
### 2.3.2.1 Biohydrogen

Biohydrogen production is achieved by conventional and anaerobic operations like reverse water gas shift reaction, gasification, water electrolysis, and steam methane reforming (Xue et al. 2013). In the ABE fermentation process, biohydrogen synthesis occurs synchronously with bioethanol and biobutanol. Photosynthetic microorganisms like *Rhodobacter sphaeroides* and *Rhodospseudomonas palustris* utilize organic matter present in microalgal biomass resulting in hydrogen and CO<sub>2</sub> generation (Lam and Lee 2011). In recent times hydrothermal gasification is the preferable technique for hydrogen production. Ma et al. (2017) reported that in the presence of a catalyst like alkaline biochar, gasification of biomass results in hydrogen yield of 89.13% (Ma et al. 2017). The gasification route was difficult to clear, but it was reported that it goes through several reactions like water gas shift, methanation, pyrolysis, steam reforming, and hydrolysis (Vo et al. 2020).



### 2.3.2.2 Biomethane

Biomethane is produced by the digestion of biomass anaerobically. In anaerobic digestion, organic matter is converted into biogas, CO<sub>2</sub>, methane (CH<sub>4</sub>), and trace gases. The three steps involved in anaerobic digestion activity are hydrolysis, fermentation, and methanogenesis (Pragya et al. 2013).



## 2.3.3 Bioelectricity

In recent years, microbial fuel cells (MFCs) from algal biomass have been a novel technology and attracting attention for bioelectricity generation (Chandrasekhar and

Venkata Mohan 2014). In MFCs, microorganisms are actively involved in bioelectricity generation; hence, they are referred to as a bioelectrochemical system (Deval et al. 2017). In microalgal MFCs, CO<sub>2</sub> is consumed by the photosynthesis process that results in organic biomass synthesis with simultaneous O<sub>2</sub> liberation. This liberated O<sub>2</sub> acts as an electron acceptor throughout the metabolism and ends up in the current synthesis. In MFCs, photosynthesis was also reported to be directly related to the light source intensity and cell density (Lee et al. 2015; Jadhav et al. 2019).

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## 2.4 Environmental Effect of Bio-Refinery Products

### 2.4.1 Carbon Footprinting

In the past century, the electrical energy and transportation zone restructured society by providing motorized movement to non-professional. It was reported that transportation (14%) and the electricity sector (25%) is responsible for GHG emission globally. Biofuels are eco-friendly as they have reduced the release of GHGs and CO<sub>2</sub> emissions. The car's lifespan determines the ecological impact of an automobile from manufacture to the level of its use. Well-to-Wheel (WTW) practice was developed to check the efficiency of vehicles. Basically, this WTW technique was separated into two steps, one is Well to Tank (WTT), and another is Tank to Wheel (TTW) (Strecker et al. 2014). The equal WTW technique calculates the carbon footprint estimation for electric vehicles. It was also reported that the lifetime of vehicles and carbon footprinting is affected by riding behaviour, use of gadgets (like air-conditioning, heating gadgets, defroster, etc.), and climate condition (Badin et al. 2013).

### 2.4.2 Negative Emission

The title “carbon negative” refers to the removal of carbon dioxide out of the common (natural) carbon cycle that includes carbon capture and segregation (CCS) through deposited biochar in soil and direct release of carbon dioxide in the wastewater for biomass farming. Here the released carbon dioxide will either be combined with the environment or treated as unfavourable depending on carbonaceous raw materials and the final target of carbon dioxide. Using 1 kg of microalgae biomass, approximately 2 kg (1.83 kg) of CO<sub>2</sub> gas can be isolated from the ecosystem (Rosenberg et al. 2011). This isolated carbon dioxide was transformed into gaseous and liquid fuels through thermochemical and biological processes. Recently, it was reported that through the gasification process, 33.5% of carbon dioxide is obtained from 15 g of *S. obliquus* biomass used (Arun et al. 2020a). Another study also reported that from 15 g of *A. fragilissima*, 34.1% of carbon dioxide and 29.5% of carbon dioxide were obtained by the HTL process and pyrolysis process, respectively. For microalgal biomass, the flow of carbon dioxide

was referred to as “carbon negative” because of its removal from the environment (Arun et al. 2020c).

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## 2.5 Conclusion

The current chapter concludes that microalgae present a promising approach for treating municipal wastewater, achieving high REs of up to 90%. Various advancements have been made in the microalgae-based wastewater treatment process, such as synchronizing microalgae with bacteria, yeast, and other species, PSBR, biofilm, and membrane technology. Out of all, the microalgae-bacterial process in the PSBR offers a cost-effective solution with high RE. Biofilm and membrane technology are also effective solutions, but the cost involved in these technologies is high, and, in the future, they may be a feasible solution after the decrease in cost. Integrating the biorefinery concept with the wastewater treatment process can decrease the cost of the process up to a suitable extent as the microalgal biomass can be transformed into various liquid and gaseous fuels and other by-products. This integration also decreases the net carbon emission in the atmosphere, decreasing the effect of global warming.

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# An Economic and Sustainable Method of Bio-Ethanol Production from Agro-Waste: A Waste to Energy Approach

# 3

Krishna Kant Pachauri

## Abstract

Rapidly increasing population and industrialization have led to a tremendous increase in energy consumption. This necessitates the exploration of sustainable and renewable methods of energy production to meet the increasing demand. Lignocellulosic agro-waste materials such as food processing and crop waste are attractive alternatives as raw materials for bioethanol production to meet the global market demand. Utilising these waste materials are also important from an economic and environmental perspective due to the low cost and the large availability of these cellulosic materials on the earth. Utilising agro-waste to produce bioethanol will also reduce the hazardous effects of phenolic compounds. The process broadly involves four significant steps: pretreatment, enzymatic hydrolysis, fermentation, and product recovery. However, there are several challenges and limitations at every step, such as agro-waste handling, transportation and removal of lignin & lignocellulose during the pretreatment, which increases the concentration of sugars used during enzymatic hydrolysis. Conversion of large chain polymers such as cellulose and hemicellulose into fermentable monomers is also a major challenge during enzymatic hydrolysis. Thus, developing an efficient strain for fermentation is essential to increase the production capacity. This chapter discusses the latest and cost-effective processes to produce bioethanol using agro-waste as raw materials.

## Keywords

Agro-waste · Bioethanol · Cellulosic materials · Fermentation · Waste management

K. K. Pachauri (✉)

National Institute of Technology Karnataka, Surathkal, Mangalore, Karnataka, India

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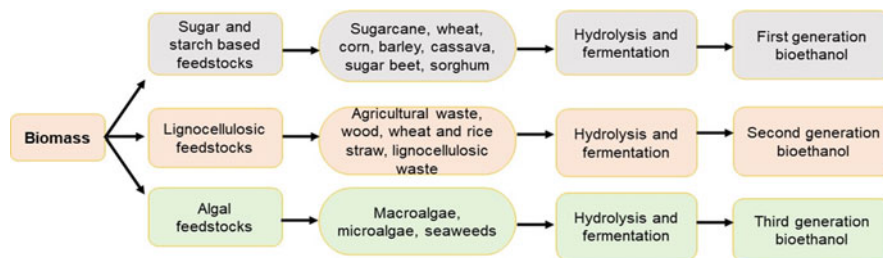
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### 3.1 Introduction

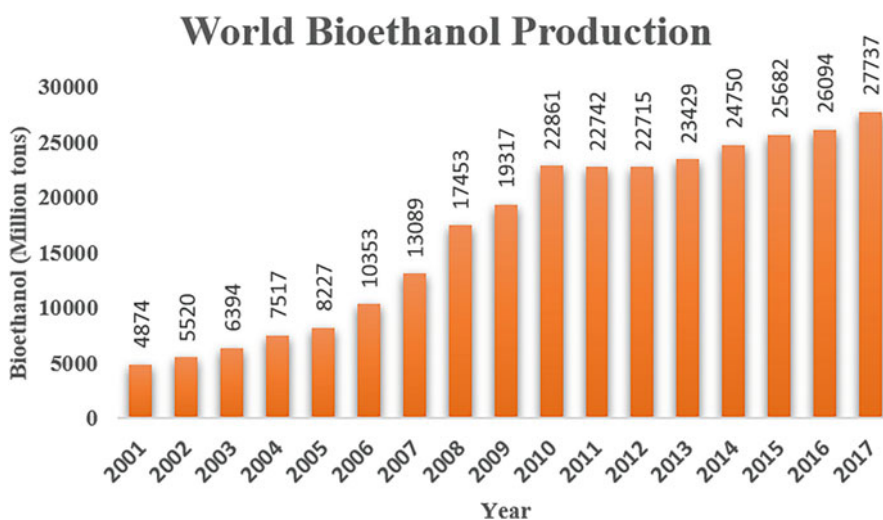
Rapidly increasing population, from 2.7 billion in 1955 to 7.9 billion in 2022 (projected to reach 8.0 billion by 2023) (<http://www.worldometers.info>), imposed a massive burden on energy resources and their utilization. Statistically, only 3% of the total global energy is supplied in the form of renewable energy, which is expected to increase up to 80% by 2050 (Mohapatra et al. 2019). In the last few decades, excessive use of fossil fuel rapidly increased air pollution, particularly in industrial and large urban areas. Furthermore, greenhouse gases are also generated by the combustion of fossil fuel which increases the global temperature and induces climate changes. The daily increasing energy demand, pollution level and limited and nonrenewable nature of fossil fuels have forced us to search for sustainable, efficient, and renewable energy resources.

The vast abundance, low cost, and enormous potential of plant biomass provide an excellent alternative source of biofuels (Haq et al. 2016). According to the studies, the world's annual lignocellulosic biomass production is around 181.5 billion tonnes, of which only 8.2 billion tonnes are utilized (Ashokkumar et al. 2022). China, the leader in the agricultural world, produces approximately 900 million tonnes of lignocellulosic waste annually. India also produces around 605 million tonnes of biomass waste (Zhao et al. 2022). Fuel production from renewable sources like plant materials could also help reduce the dependency on fossil fuels and global CO<sub>2</sub> production. Biofuels include bioethanol, biomethanol, bio-gas, biodiesel, biohydrogen, etc. Out of these, bioethanol from plant biomass is a promising way to tackle the global energy crisis and environmental issues (Naik et al. 2010). Bioethanol production from first-generation biomass such as corn and sugarcane is more common in the global bioethanol market. Almost 50 bn litres of first-generation bioethanol is produced annually. However, the major disadvantage of the first-generation bioethanol is the increasing price of food crops due to the increased production of these biofuels. Therefore, lignocellulosic biomass obtained from the non-food (Agricultural or residual forest materials) part of the plants can provide an excellent alternative to produce bioethanol, called second-generation biofuels. However, second-generation bioethanol production from lignocellulosic feedstocks is not cost-effective due to the critical barriers at the several steps of the production process. The third-generation bioethanol involves marine biomass (micro and macroalgae) as a feedstock. It has also gained worldwide popularity due to the unsustainability of first and second-generation bioethanol (Jambo et al. 2016). It also provides food security and less environmental impact (Fig. 3.1).

Globally, the United States (58%) and Brazil (27%) are leading bioethanol producers using corn and sugarcane as feedstock material, respectively (Kohler 2018). United States of America (USA) produced around 13.9 billion gallons of ethanol in 2020 (US Department of Energy, <https://afdc.energy.gov/data/>). Wheat, potato, and sugar beet are common feedstock materials for bioethanol production in European countries. Considering the food security issue of first-generation bioethanol, India's bioethanol production program depends on second-generation feedstock materials such as sugarcane molasses (Chandel and Sukumaran 2017).



**Fig. 3.1** Three different generations of bioethanol based on the feedstock used for the production



**Fig. 3.2** Status of world bioethanol production in million tonnes from 2001–2017. (Mohapatra et al. 2019)

India currently produces about 2% (4.26 bn liters from molasses-based and 2.58 bn litres from grain-based distilleries) of the total bioethanol production. It is expected to be increased up to 10 bn litres by 2025 targeted by Union Ministry of Petroleum & Natural Gas (MoPNG). The Indian government also launched its Ethanol-Blended Petrol Programme (EBPP) in 2003. According to the National Policy on Biofuels (2018), it is targeted to blend 20% ethanol under the Ethanol Blended Petrol (EBP) scheme by 2030 (Figs. 3.2).

## 3.2 Lignocellulosic Biomass

It is the most abundant raw material available for biofuel production (Madakka et al., 2020), majorly Bioethanol. It is a renewable organic material containing cellulose, hemicellulose, and lignin as three basic components. Lignocellulosic material has



enormous biotechnological value due to its chemical composition and properties (Pothiraj et al., 2006).

### 3.2.1 Cellulose

Cellulose is the most prevalent organic material on our planet earth, present in the cell wall of all plant materials. It is a non-toxic bio-degradable linear biopolymer containing several units of D-glucose linked with the  $\beta$ -1,4-glycosidic bond. Around 7000–15,000 subunits of glucose form a cellobiose chain after joining with the  $\beta$ -1,4-glycosidic bond, and these cellobiose chains are joined together by the hydrogen bonding and Vander-walls forces creating microfibrils. These microfibrils are joined together by hemicellulose, pectin, and other polymers and covered by the lignin forming a bundle of microfibrils called macrofibrils. This complicated cellulose structure makes it resistant to various biological and chemical reactions. The fermentable glucose unit is released from the complex cellulose molecule during the fermentation after the enzymatic hydrolysis of the  $\beta$ -1,4-glycosidic bond (Haq et al., 2016)

### 3.2.2 Hemicellulose

Hemicelluloses are the second principal component (20–35%) of lignocellulosic biomass. It is a heterogeneous polysaccharide containing pentoses (xylose, arabinose), hexoses (glucose, galactose, mannose), and sugar acids. The most common hemicellulose is xylan, found in nearly all agricultural residues. The dominant component of hemicellulose in hardwood and softwood is xylans and glucomannans, respectively. As compared to cellulose, hemicellulose is less complex, contains shorter chains of sugar units and is readily hydrolysed to fermentable sugar due to its breached and amorphous structure (hemicellulose bioconversion).

### 3.2.3 Lignin

Lignin is a heterogeneous polymer containing three aromatic units of *p*-coumaryl, coniferyl, and sinapyl alcohol linked together by different ether, ester, and carbon-carbon bonds (Hendriks and Zeeman 2009). The third major component of lignocellulosic biomass comprises around 15–25% of the total dry mass. The primary function of the lignin is to serve as a cementing material between the wood fibres and a stiffening material within the fibres. It also acts as a blockade to the enzymatic degradation of the cell wall. Lignin is also the most recalcitrant material because of the nonhydrolysable C-O-C and C-C bonds between its units. The rate of lignin degradation is much slower than the other non-cellulosic and cellulosic polysaccharides and proteins.

### 3.3 Raw Material for Bioethanol Production

Based on the raw material used, bioethanol is mainly divided into two categories. First-generation bioethanol is derived from raw materials containing starch and sugar such as rice, wheat, sugarcane, etc. Although the bioethanol produced using food materials is more economical, this can create a shortage of available food for the population. Exploring other alternative raw materials for bioethanol production that do not interfere with food security is necessary. Lignocellulose containing agro-waste materials such as crop residue, grasses, rice and wheat straws, sugarcane bagasse, etc., could be the best alternative raw material for bioethanol production. Bioethanol produced using these materials is referred to as second-generation bioethanol. The reduction of greenhouse gas emissions and renewability of the lignocellulose-rich waste material are the significant advantages of this second-generation bioethanol.

According to Sarkar et al. (2012), four primary agro-waste materials for bioethanol production are bagasse, rice straw, wheat straw, and corn straw, which are available throughout the year. Asia is the highest producer of rice and wheat straws, whereas America primarily produces bagasse and corn straws. Although the chemical composition of these materials varies, cellulose is commonly available as a major component. These agro-waste materials are also utilized for various other purposes, such as animal fodders, as a fuel to run boilers, as a domestic fuel, etc., in different quantities based on the requirement and the geographical regions (Sarkar et al. 2012).

Different types of feedstock materials are used to produce bioethanol. The overall process of fermentation also varies according to the raw material used for bioethanol production. Techniques like pretreatment, milling, and hydrolysis are not required in the case of the sugar-based feedstock materials, but these processes are necessary for the lignocellulosic feedstock materials. Liquefaction and saccharification processes are needed when the starch-based feedstock is used as a raw material. A detoxification unit is also considered in case of the toxic raw material used for the fermentation. Based on the chemical composition, raw materials are divided into four categories: sugar-based, starch-based, lignocellulosic-based, and algal-based materials.

#### 3.3.1 Sugar-Based Raw Material

Various raw materials like sugar beets, sugar cane, sweet sorghum, and sugar crops fall under the sugar-based feedstock category. High yield and low conversion cost are the two significant advantages of these sugar-based feedstocks. In contrast, seasonal availability is the major obstacle to the continuous supply of raw materials. Sugar cane byproducts like cane juice and molasses are the primary raw material for bioethanol production in Brazil (Zabed et al. 2014). In contrast, sugar beet is primarily used for bioethanol production in North America, Europe, and France (Ohlmaier-Delgado et al. 2021; Balat 2007). According to an estimate, around

25 gallons of bioethanol can be produced by one tonne of sugar beet. The byproducts (molasses) and other intermediates of sugar beet have high sugar content but require more energy and chemical processes than sugar cane. It is a more expensive raw material for bioethanol production than sugar cane. Sweet sorghum is also used as a raw material for China's bioethanol production. The plant's main stalk is the major sugar-containing portion, which is pressed using the rollers to recover the sugar material from the plant. The average output is 20 gallons of bioethanol from one tonne of sweet sorghum stalks (Sandesh Suresh et al. 2019). Since the stalk is only required for bioethanol production, the farmers use the sorghum grains as a food material.

### 3.3.2 Starch-Based Raw Material

This is the major feedstock material used for the bioethanol production obtained from the grains such as corn, wheat, and barley. These grains have high starch content, like 60–70% in the case of corn. This raw material is mainly used in North America and Europe for bioethanol production. Starch is found in the form of amylose and amylopectin in the grains. These polymeric structures are broken down to monomeric unit glucose by the hydrolytic action of enzymes, viz. glucoamylase,  $\beta$ -amylase, isomerase, etc.

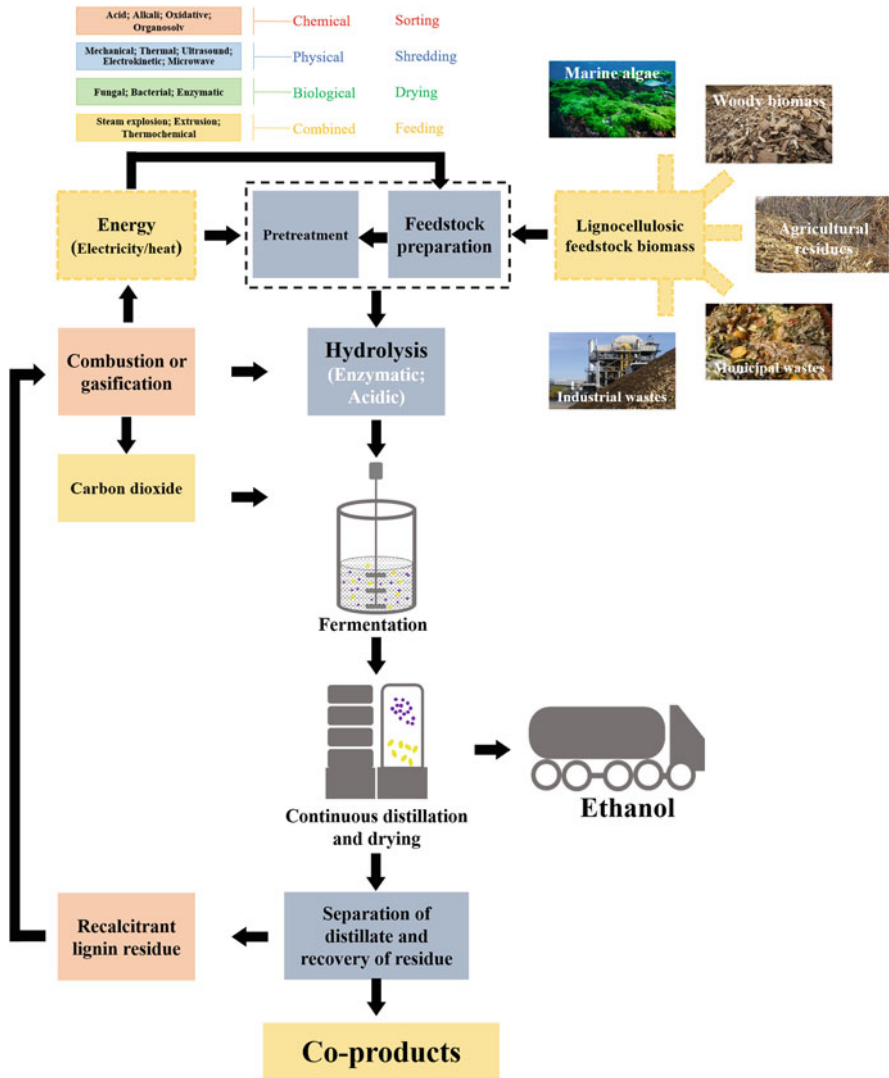
### 3.3.3 Lignocellulosic Raw Material

Bioethanol produced from the lignocellulosic waste material is called second-generation bioethanol. The feedstock used for second-generation bioethanol production generally contains agricultural waste materials (rice and wheat straw, corn residue, etc.), grasses, forestry and wood residues, etc. Treatment of these potentially valuable materials as waste raises many environmental concerns. Various prosperous efforts have been made to convert this waste material into valuable products like bioethanol. The world produces 731 million tonnes of rice waste, the highest waste generated annually. This large amount of generated rice straw could be used to produce 205 billion litres of bioethanol (Haq et al. 2016). At the same time, around 354 million tonnes of wheat straw are generated globally, which could be used to generate approximately 104 billion litres of bioethanol (Bhatia et al. 2012).

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## 3.4 Overview of Bioethanol Production from Lignocellulosic Agricultural Waste Materials

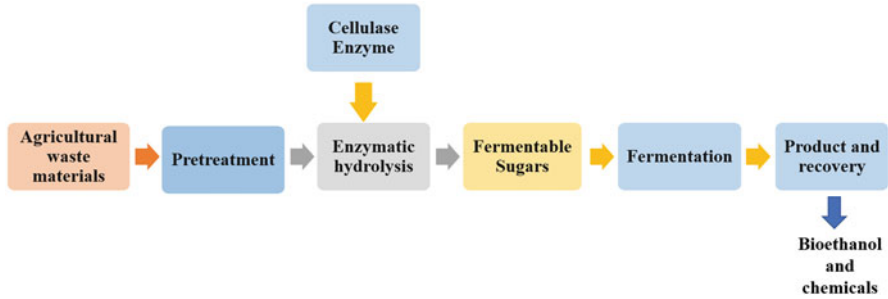
The conversion of lignocellulosic waste material into fermentable sugar is much harder than the sugar and starch-based feedstocks (Haq et al. 2016). The transformation of agricultural waste into ethanol is divided into four steps, which include



**Fig. 3.3** An overview of bioethanol production using lignocellulosic biomass

(1) pretreatment of the waste material, (2) enzymatic hydrolysis of the pretreated waste material, (2) fermentation, and (4) product recovery (Fig. 3.3).

Pretreatment is crucial step to increase the hydrolysis efficiency by increasing the pore size and reducing the crystallinity of the cellulose material. It also enhances the biodigestibility of the waste material and increases the product yield. Post pretreatment, the cellulosic microfibrils of the lignocellulosic biomass are exposed and become susceptible to the enzymatic and/or acid hydrolysis to produce fermentable sugar. The sugar is converted to ethanol by the action of microorganisms during

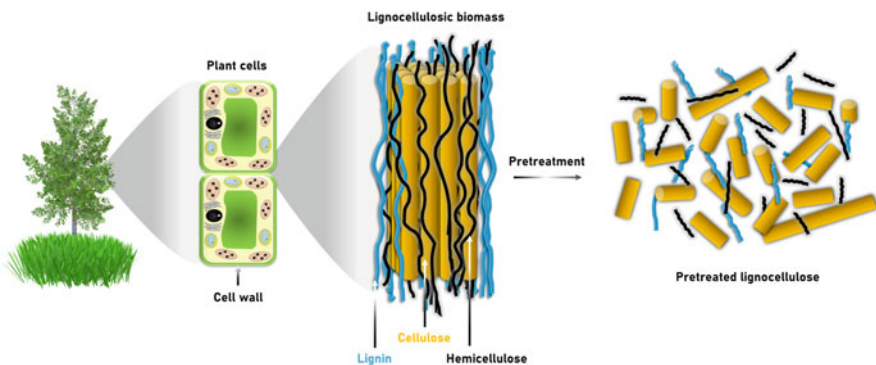


**Fig. 3.4** Schematic representation of major steps involved for producing ethanol using lignocellulosic biomass

the fermentation process. After fermentation, a dilute aqueous solution containing ethanol is obtained and concentrated into the anhydrous ethanol by various distillation methods. In some cases, pentose (xylose) detoxification is also done to remove the undigestible sugar. The complete process of lignocellulosic agricultural biomass to ethanol conversion is discussed in the following section (Fig. 3.4).

### 3.4.1 Pretreatment

It is the first and most crucial step in producing bioethanol from lignocellulosic agricultural waste materials. In this step, hemicellulose and lignin content are removed from the biomass and cellulose crystallinity is also reduced. Pretreatment also increases the material's porosity and the final yield of the fermentable sugars (Fig. 3.5). It inhibits carbohydrate degradation and reduces the production of toxic byproducts, hindering the hydrolysis and fermentation process. There are



**Fig. 3.5** Pretreatment of lignocellulosic biomass

specific goals to be fulfilled by an effective pretreatment process. An ideal pretreatment process should have the following qualities—

- It should be cost-effective and require minimum heat and power
- It Should have a high yield for multiple crops
- It Should provide highly digestible pretreated solid biomass
- It Should not be significant sugar degradation during the process
- There should be minimum production of toxic compounds
- There should be a high recovery of the valuable products derived from hemicellulose and lignin

Due to the diverse nature of lignocellulosic biomass, a universal pretreatment method is challenging to apply over different feedstock materials. Various pretreatment processes have been suggested during the last two decades and have been broadly classified into four categories (Table 3.1):

- Physical pretreatment (milling, grinding, irradiation, and pyrolysis)
- Chemical pretreatment (acid, alkali, ozonolysis, ionic liquids, and organic solvents)
- Physico-chemical pretreatment (steam explosion/hydrolysis, AFEX, and CO<sub>2</sub> Explosion)
- Biological (Fungi and bacteria)

#### **3.4.1.1 Physical Treatment**

Numerous mechanical (ball milling, grinding) and non-mechanical (irradiation) methods are considered as physical pretreatment of lignocellulosic wastes. Different electromagnetic rays, such as gamma rays, microwaves, and electron beams, are used during the irradiation method of hydrolysis of agronomic waste (Priyanka et al. 2018). Chipping, milling, and grinding are the most common ways of mechanical pretreatment.

##### **Milling**

This pretreatment method is frequently used to reduce the particle size of lignocellulosic biomass and increase the collective surface area for enzymatic action. Different types of milling, like ball milling, hammer milling, disk milling, etc. are used. Vibratory ball milling resulted in the most effective way of breaking down the biomass and reducing cellulose crystallinity compared to ordinary ball milling. The material size is reduced to 10–30 mm through chipping and 0.2–2 mm post-milling or grinding (Bhatia et al. 2012). Wet milling and dry milling are the two distinct methods of corn processing, and both these processes generate distinct co-products. The corn is passed through the milling hammer and divided into fine particles in the dry-milling process.

In contrast, corn is soaked in large steep tanks in a dilute sulfuric acid solution during the wet milling for 24 to 48 h. Dry milling is less labor-intensive and primarily used for ethanol production, while wet milling extracts high-value

**Table 3.1** Different pretreatment strategies along with their advantages and disadvantages

Type of pre-treatment	Pretreatment method	Advantages	Disadvantages	Reference
Physical	Milling	<ul style="list-style-type: none"> <li>• Size reduced up to 10–30 mm, no inhibitors production</li> </ul>	<ul style="list-style-type: none"> <li>• Higher power requirement</li> </ul>	Bhatia et al. (2012)
	Pyrolysis	<ul style="list-style-type: none"> <li>• Cellulose decomposes rapidly in the presence of oxygen</li> </ul>	<ul style="list-style-type: none"> <li>• High temperature requirement makes the process expensive</li> <li>• Production of residual char</li> </ul>	Sun and Cheng (2002)
	Irradiation	<ul style="list-style-type: none"> <li>• Uniformity, selectivity, less energy requirement, and short time duration</li> </ul>	<ul style="list-style-type: none"> <li>• Formation of the inhibitors</li> </ul>	Haq et al. (2016)
Chemical	Acid pretreatment	<ul style="list-style-type: none"> <li>• Cellulose more accessible to enzymes</li> </ul>	<ul style="list-style-type: none"> <li>• Formation of some inhibitory compounds (furfural)</li> <li>• Equipment corrosion and acid recovery problems (hydrolysis neutralization)</li> </ul>	Hoang et al. (2021)
	Alkali pretreatment	<ul style="list-style-type: none"> <li>• Non-corrosive and non-polluting chemicals</li> <li>• Lower temperature and pressure</li> </ul>	<ul style="list-style-type: none"> <li>• High cost of hydroxides</li> <li>• Chemical structure alteration of lignin in the biomass</li> </ul>	Mosier et al. (2005c)
	Organosolv pretreatment	<ul style="list-style-type: none"> <li>• Easy solvent recovery and high purity fractionation of the biomass</li> </ul>	<ul style="list-style-type: none"> <li>• Expensive nature of the process due to the use of organic solvents</li> <li>• High temperature and pressure</li> <li>• Formation of toxic inhibitors</li> <li>• Corrosion by organic acids</li> </ul>	Bensah and Mensah (2013)
	Ozonolysis pretreatment	<ul style="list-style-type: none"> <li>• High efficiency and mild operating conditions</li> <li>• Low production of inhibitory compounds</li> <li>• Selective lignin degradation</li> </ul>	<ul style="list-style-type: none"> <li>• Highly reactive, corrosive, flammable, and toxic nature of the ozone gas</li> </ul>	Travaini et al. (2016)
	Wet-oxidation pretreatment	<ul style="list-style-type: none"> <li>• No chemicals recovery required</li> <li>• Applies to the wide variety of woody biomass</li> </ul>	<ul style="list-style-type: none"> <li>• Large oxygen amount and catalyst cost are the major disadvantages</li> </ul>	Mcginnis et al. (1983)

	Ionic-liquid pretreatment	<ul style="list-style-type: none"> <li>• Green recyclable way of biomass pretreatment</li> <li>• No harmful and volatile organic solvents required</li> </ul>	<ul style="list-style-type: none"> <li>• Requirement of a massive quantity of expensive ionic-liquids</li> <li>• Energy intensive recycling process</li> <li>• viscous nature of the solution</li> </ul>	Zavrel et al. (2009)
Physico-chemical pretreatment	Steam explosion	<ul style="list-style-type: none"> <li>• Environment-friendly process</li> <li>• Low capital investment</li> <li>• Energy efficient process</li> </ul>	<ul style="list-style-type: none"> <li>• Less effective for softwoods</li> </ul>	Pielhop et al. (2016)
	Liquid hot water (LHW) pretreatment	<ul style="list-style-type: none"> <li>• No chemical input</li> <li>• Minimum waste and inhibitory product generation</li> <li>• Relatively lower total cost</li> </ul>	<ul style="list-style-type: none"> <li>• High temperature requirement</li> <li>• Alkaline addition to maintain a fixed pH range</li> </ul>	Weil et al. (1998)
	Ammonia fiber explosion (AFEX)	<ul style="list-style-type: none"> <li>• Minimum inhibitory product formation</li> </ul>	<ul style="list-style-type: none"> <li>• High ammonia cost</li> <li>• Environmental issues</li> </ul>	Bals et al. (2011)
Biological	Supercritical CO <sub>2</sub> explosion (SC-CO <sub>2</sub> ) explosion	<ul style="list-style-type: none"> <li>• Nonflammable, nontoxic, inexpensive, and environment-friendly method</li> <li>• Significantly less corrosive unlike other acid catalysed processes</li> </ul>	<ul style="list-style-type: none"> <li>• High pressure required</li> <li>• Large capital investment</li> </ul>	Sarkar et al. (2012)
	Bacteria and fungi	<ul style="list-style-type: none"> <li>• It is eco-friendly with no toxic compounds released in to the environment</li> <li>• Chemical recycling not required</li> </ul>	<ul style="list-style-type: none"> <li>• Lower rate of hydrolysis</li> <li>• Microorganism growth monitoring</li> <li>• Larger space required</li> </ul>	Sindhu et al. (2016)



co-products such as high fructose corn syrup (HFCS). A high energy requirement is the major disadvantage of this process (Mankar et al. 2021).

### **Pyrolysis**

This process is also called as thermal cracking due to the application of a high temperature. In this process, the material is heated at a high temperature of about 300 °C in non-oxidizing atmosphere. The rapid decomposition of biomass leads to the production of pyrolysis oils (bio-oils), pyro-gases (hydrogen and carbon monoxide gas), and solid residual char (Hosur et al., 2020; Yogalakshmi et al. 2022). The presence of oxygen can enhance the process. The decomposition can also occur at lower temperatures in the catalytic presence of sodium carbonate or zinc chloride to reduce the production of gases and other residues (Sun and Cheng 2002).

### **Irradiation**

This is an effective and easy-to-implement method of pretreatment. It enhances cellulase enzyme activity on lignocellulosic biomass by changing the ultrastructure of the cellulose. This treatment also degrades hemicellulose and lignin of the biomass. It is a short-duration process with uniformity, high selectivity, and requires less energy input (Haq et al. 2016; Cheng et al. 2011). Irradiation has also been performed along with other methods to improve the process of ethanol production. According to a recent study performed by Shangdiar et al. (2022), the hydrolysis time for sugar bagasse reduces up to 40–50% through microwave-assisted acid hydrolysis as compared to other conventional methods.

#### **3.4.1.2 Chemical Pretreatment**

It includes the application of dilute acids (HCl, H<sub>2</sub>SO<sub>4</sub>, organic acids), oxidizing agents (ozone and hydrogen peroxide), alkalis (Na<sub>2</sub>CO<sub>3</sub>, NaOH, Ca(OH)<sub>2</sub>, and NH<sub>3</sub>), SO<sub>2</sub> and CO<sub>2</sub> gases, organic solvents, and other chemicals for pretreating the biomass. These chemicals degrade the hemicellulose and remove the lignin from the lignocellulosic biomass materials (Nwosu-Obieogu 2016). These simple methods provide a good yield of fermentable sugars in a short duration (Sarkar et al. 2012).

#### **Acid Pretreatment**

It is recognized as one of the most crucial methods for solubilizing the hemicellulose portion of the biomass and increases the enzyme accessibility of cellulose. In this pretreatment, the waste undergoes treatment either with dilute or concentrated acids (usually 0.2%–2.5% w/w) at temperatures ranging from 130–210 °C. Sulfuric acid (H<sub>2</sub>SO<sub>4</sub>) is extensively used along with hydrochloric acid, phosphoric acid, and nitric acid (Cardona et al. 2010). Two approaches can be followed for acid pretreatment: at higher temperatures using dilute acids or at lower temperatures using concentrated acid. Both methods have their benefits and drawbacks (Taherzadeh and Karimi 2008). Relatively pure cellulose is obtained when acid pretreatment, which is responsible for hemicellulose removal, is followed by the alkali treatment to remove lignin (Wingren et al. 2003). Govumoni et al. (2013)

reported that the maximum glucose yield (65.2 g/L) was obtained by biomass pretreatment with 0.75% H<sub>2</sub>SO<sub>4</sub> at 100 °C temperature for 2 h, followed by the 1.5% NaOH treatment at 100 °C for 2 h. Sometimes the enzymatic hydrolysis step can be avoided because the acid pretreatment performs hydrolysis also, and converts biomass into fermentable sugars. In another strategy, Approximately 90% hemicellulose removal can be obtained using a mixture of sulfuric acid and acetic acid (de Moraes et al. 2011).

As per the literature reports, oxalic acid can replace sulfuric acid because it has high saccharification efficiency and is less lethal for the bioethanol-producing microorganism (Lee et al. 2009). As oxalic acid is costlier than sulfuric acid, from an economic point of view, using oxalic acid at a large scale for pretreatment is not possible. Conventional recovery methods such as ion exchange and adsorption can be applied to overcome this problem. Organic acids such as fumaric and maleic acid were also tested for their pretreatment efficiency. It was observed that dilute fumaric acid or maleic acid at 150 °C can be an excellent alternative to dilute sulfuric acid pretreatment (Kootstra et al. 2009). Generally, the use of concentrated acid is avoided due to the production of some inhibitory compounds (furfural) along with equipment corrosion and acid recovery problems (Alvira et al. 2010). The cost and the neutralization of these acids after treatment are the two significant challenges of this process.

### Alkali Pretreatment

Among all available methods, it is a frequently used technique because of some desirable features like using non-corrosive and non-polluting chemicals and requires lower temperature and pressure than other pretreatment methods (Mosier et al. 2005c). Alkali pretreatment causes swelling and decrystallization of cellulose, structural alteration of lignin by degrading ester and glycosidic side chains, and partial solvation of hemicellulose (Brodeur et al. 2011). Different reagents such as sodium hydroxide, calcium hydroxide, sodium carbonate, and ammonia have been explored to improve the enzymatic digestibility of biomass through alkaline pretreatment (Kim et al. 2016). Pretreatment with calcium hydroxide is most favoured because it is corrosion-free, economical, and possible recovery from the hydrolysate by reacting with the carbon-di-oxide (Mosier et al. 2005c). According to Sun et al. 1995, the optimum pretreatment results were obtained by treating the wheat straw using 1.5% sodium hydroxide for 144 h at a temperature of 20 °C. The treatment releases 60% lignin and 80% hemicellulose from the biomass. The use of alkaline hydrogen peroxide (AHP) is also an effective way to reduce the generation of biological growth inhibitors such as furfural and hydroxymethylfurfural (HMF) (Dutra et al. 2018). A comparative study of different chemical pretreatment strategies of corn stover between the alkaline, acid, and sulphite treatments found that cellulose saccharification yield was highest (65%) in the case of alkaline pretreatment (Yu et al. 2014). Various studies have been performed on different biomass materials to optimize the conditions of the alkali pretreatment for obtaining the maximum yield of fermentable sugar. A mixture of calcium hydroxide and biomass in the ratio of 1:10 at 50 °C for 24 h is the best-suited pretreatment condition

for switchgrass (Xu et al. 2010). Compared to the untreated biomass, the yield of glucose, xylose, and reducing sugars concentration increased by 3.15, 5.78, and 3.61 times under these conditions, respectively. Wang et al. 2016 suggested that high pressure-assisted alkali pretreatment (HPAP) of cotton stalks gives the highest yield of reducing sugar (271.70 mg/g) and ethanol (45.53%). During HPAP, dried powder of cotton stalk was mixed with 3% sodium hydroxide and kept for 40 min time at a high pressure of 130 kPa. Mainly four hydroxides (sodium, potassium, calcium, and ammonium) have been explored significantly for alkali pretreatment of different biomass materials. Calcium hydroxide (slake lime) is the most widely used pretreatment agent due to its availability and low cost (Kumar et al. 2009).

### Organosolv Pretreatment

In this pretreatment method, lignin and hemicellulose linkages are degraded after treating the lignocellulosic biomass through a solvent mixture with or without an acid catalyst (Haq et al. 2016). Different organic solvents with low (ethanol and methanol) and high boiling points (glycerol, ethylene glycol, and tetrahydrofurfuryl alcohol) have been tested for biomass pretreatment. Other compound classes such as ethers, phenols, ketones, and dimethylsulphoxide were also used for treating the biomass (Zhao et al. 2009). Generally, this pretreatment is operated at 160–220 °C, although, if the process is performed at higher temperatures (185–220 °C), fortification of external acid is not required because the acid generated from the biomass acts as a catalyst for the lignin-carbohydrate complex breakdown (Teramoto et al. 2008; Duff and Murray 1996). Studies have been performed to determine the optimum treatment conditions for obtaining the maximum glucose yield during the enzymatic hydrolysis step. According to Mesa et al. (2011), the best conditions for a dilute acid-pretreated sugarcane bagasse consist of 30% (v/v) ethanol for 60 min at 195 °C, which yield around 29.1 g glucose/100 g of sugarcane bagasse after hydrolysis of residue. A study performed by Araque et al. (2008) revealed the highest ethanol yield (99.5%) from the organosolv acetone-water pretreated wooden chips of the *Pinus radiata*. A 1:1 ratio of acetone and water was used for biomass pretreatment at pH 2.0 and a temperature of 195 °C for 5 min. The highest sugar concentration (31 g/L) was obtained by pretreating the rice straw using 75% (v/v) aqueous ethanol mixed with 1% (w/w) H<sub>2</sub>SO<sub>4</sub> at 150 °C for 60 min (Amiri et al. 2014). Generally, low molecular weight alcohols like ethanol and methanol are favoured for the treatment over high molecular weight alcohols due to their economic feasibility (Haq et al. 2016). Organosolv pretreatment has been recognized as an emerging method due to its inherent advantages like easy solvent recovery and high purity biomass fractionation into hemicellulose, cellulose, and lignin (Zhang et al. 2016). The expensive nature of the process due to the application of organic solvents at high temperature and pressure, formation of toxic inhibitors, and corrosion by organic acids are the significant challenges of this process (Bensah and Mensah 2013).

### Ozonolysis Pretreatment

Ozonolysis is becoming a widespread pretreatment method due to high efficiency and mild operating conditions. As the name implies, ozone is used to treat

lignocellulosic agricultural waste, the most potent oxidizing agent ( $E^\circ = 2.07 \text{ V}$ ,  $25^\circ \text{C}$ ). It is water soluble ( $110 \text{ mg/L}$ ,  $25^\circ \text{C}$ ) and can be quickly produced from oxygen through a strong endothermic reaction (Travaini et al. 2016). Recent studies suggested that ozone reacts more rapidly with insoluble lignin than carbohydrates, increasing biomass delignification, and enhancing the sugar release during enzymatic hydrolysis (Sankaran et al. 2020). Several studies have also been done on the pretreatment of different agricultural and forestry waste materials using ozonolysis. In a survey conducted by Travaini et al. (2013), sugarcane bagasse was pretreated by ozonolysis to enhance lignocellulosic digestibility. It was found that glucose and xylose contents were improved from 6.64% and 2.05% in raw bagasse to 41.79% and 52.44% in treated material, respectively. Ozonolysis has also been used for other feedstock materials such as maize stover (Li et al. 2015), corn straw (Shi et al. 2015), energy grasses (Panneerselvam et al. 2013), wheat and rye straw (García-Cubero et al. 2009), poplar sawdust (Vidal and Molinier 1988). The main advantages of ozonolysis are (1) Low production of inhibitory compounds, (2) Selective degradation of lignin, and (3) Ambient temperature and pressure conditions. Despite being so fruitful, ozone gas is highly reactive, flammable, corrosive, and toxic, which limits the process. (Travaini et al. 2016).

### Wet-Oxidation Pretreatment

In this process, agricultural waste is treated in the presence of oxygen and water at elevated temperatures and pressure (Schmidt and Thomsen 1998). Typically, the wet-oxidation process is operated at high temperatures ( $120\text{--}238^\circ \text{C}$ ) and oxygen pressure ( $120\text{--}480 \text{ psi}$ ) for 30 min. Acid formation occurs due to the dissolution of hemicellulose components, such as xylans, which are acidic in nature. The drop in pH due to the formation of acids makes the conditions favourable for hydrolytic reactions (Mcginnis et al. 1983). Pretreatment of several feedstock materials such as wheat straw (Schmidt and Thomsen 1998), softwood (Palonen et al. 2004), rice, sugarcane, peanuts and cassava (Carlos and Thomsen 2007), clover–ryegrass mixtures (Martín et al. 2008), and rape straw (Arvaniti et al. 2012) has been performed using this method. It is also applied to newspaper waste to increase its anaerobic digestibility (Fox and Noike 2004; Verma et al., 2018; Yadav et al., 2020). Moreover, the process offers certain advantages; (1) use of inexpensive materials (oxygen and water), (2) requires no chemicals recovery, (3) separation of biomass into the liquid (hemicellulose and lignin rich) and solid (cellulose rich) fraction, and (4) applies to the wide variety of woody biomass (Mcginnis et al. 1983).

### Ionic-Liquid Pretreatment

Ionic-liquids (ILs) are a “green” recyclable way of pretreating lignocellulosic materials. This method is an alternative to harmful and volatile organic solvents, which are exploited in various processes, including biomass pretreatment (Moniruzzaman and Goto 2019). ILs are organic salts with large organic cationic species and small inorganic anionic species with a  $< 100^\circ \text{C}$  melting point (Alayoubi et al. 2020). These liquids can disintegrate and solubilize the lignocellulosic biomass and enhance the availability of simple carbohydrates for fermentation. Various types

of ILs such as 1-ethyl-3-methylimidazolium acetate ([EMIM][OAc]), 1-allyl-3-methylimidazolium-chloride ([AMIM] [Cl]), 1-(2-Hydroxyethyl)-3-methylimidazolium-tetrafluoroborate ([HEMIM][BF<sub>4</sub>]), 1,3-Dimethylimidazolium-dimethylphosphate (ECOENG 1111P) (ECOENG) are explored by various researchers for the treatment of switchgrass, energy cane bagasse and other lignocellulosic material (Li et al. 2010b); Qiu et al. 2012); Zavrel et al. 2009). Aqueous ionic liquid containing the mixture of 1-ethyl-3-methylimidazolium acetate and water was also used for pretreating the straw. A higher sugar yield was obtained in the case of aqueous ionic liquid compared to the pure ionic liquid experimented under similar conditions (Fu and Mazza 2011). A study performed by Li et al. (2010b) showed that ionic liquid is more efficient for the treatment of switchgrass than dilute acid pretreatment. Although ionic liquids are a great alternative to the volatile and toxic organic solvent, a few challenges must be worked out before applying these liquids on an industrial scale. The major problems associated with this pretreatment method are the requirement of a massive quantity of expensive ILs, energy-intensive recycling process, and the viscous nature of the solution during the process (Zavrel et al. 2009).

### 3.4.1.3 Physico-Chemical Pretreatment

As the name says, this method allows the combined effects of physical and chemical ways to increase the digestibility of lignocellulosic materials. Steam explosion, liquid hot water (LHW), ammonia fiber explosion (AFEX), and CO<sub>2</sub> explosion are primary treatment methods in this category.

#### Steam Explosion

Steam hydrolysis (autohydrolysis) or explosion is the most widely used and environment-friendly physico-chemical process of lignocellulosic biomass treatment (Singh et al. 2015). This process includes heating the biomass by saturated steam under high pressure for an optimized period, following which the pressure is quickly released. This quick release of the pressure causes steam expansion inside the cellulosic matrix, breaking the cell walls and separating the individual fibers (Horn and Eijsink 2010). Pressure is a critical factor during the entire process because it is directly related to the temperature and impacts the hydrolysis kinetics of cellulose and other degrading products (Jacquet et al. 2015). The result of initial studies on steam explosion showed a cumulative effect of retention time and temperature on the pretreatment process called severity factor (S).

$$S = \log \left\{ \int_0^t \exp \left( \frac{T(t) - 100}{14.75} \right) dt \right\}$$

where  $S$  = severity factor,  $t$  = retention time (min.),  $T(t)$  = process temperature (°C), and 14.75 = activation energy of the process following Arrhenius law and first-order kinetics.

Although the normal range of temperature and retention time for the steam explosion is from 200–280 °C for 2–10 min, however, the optimum pretreatment results are obtained, either at high temperature for a brief time (270 °C for 1 min) or at a lower temperature for a longer time (190 °C for 10 min) (Singh et al. 2015). This pretreatment technique has been studied for bioethanol production using various feedstock materials like hardwood (Horn and Eijssink 2010), wheat straw (Ballesteros et al. 2006), corn stover (Yu et al. 2011), switch grass and sugarcane bagasse (Ewanick and Bura 2011), sunflower stalks (Vaithanomsat et al. 2009), pine (*Pinus patula*) (Chacha et al. 2011), eucalyptus wood (Martín-Sampedro et al. 2011), etc. The steam explosion is an effective pretreatment method for most feedstocks, but it is less promising in the case of softwoods (Pielhop et al. 2016) for two main reasons. First, the lower methoxy content of softwood lignin leads to its higher condensation and makes it chemically more resistant to deconstructing the lignin portion. Second, the partially acetylated glucomannans or galactoglucomannans group in the hemicellulose backbone, where xylose and arabinose amount is less (Nitsos et al. 2018; Singh et al. 2015).

### Liquid Hot Water (LHW) Pretreatment

LHW is an efficient and environment-friendly (chemical-free) pretreatment method utilized for enhancing enzymatic digestibility of lignocellulosic feedstock materials (Imman et al. 2018). In this technique, biomass is exposed to hot water at an elevated temperature (160–240 °C) and pressure (> 5 MPa) and kept for a limited time duration ( $\leq 1$  h). An optimized pH (4–7) is also necessary to maintain throughout the aqueous treatment, especially at high temperatures and pressures (Weil et al. 1998). During the process, hemicellulose decomposition occurs in three steps: generation of primary products, water dissolution of primary products, and further disintegration (Zhuang et al. 2016). In this pretreatment, around 20–30% of lignin is removed. A new pretreatment method was also developed with improved lignin removal by combining the LHW treatment with aqueous ammonia (Yu et al. 2013). Among the different available pretreatment methods, this method has several advantages, including no chemical input, minimum waste, and other inhibitory product generation, and relatively lower total capital investment due to no chemical requirement (Wells et al. 2020). This method has been investigated for the pretreatment of diverse feedstock materials such as corn fiber (Mosier et al. 2005a, b), yellow poplar wood sawdust (Weil et al. 1998), sugarcane bagasse (Yu et al. 2013), and wheat straw (Pérez et al. 2008).

### Ammonia Fiber Explosion (AFEX)

AFEX is a robust method of pretreatment for lignocellulosic material. It reduces the lignocellulosic recalcitrance and minimizes the production of inhibitory product formation during the pretreatment (Balan et al. 2009a). It is an ammonia-based physico-chemical pretreatment method utilizing the physical (high pressure and temperature) and chemical (ammonia) processes for efficient biomass pretreatment (Bals et al. 2011). In this process, the biomass is pretreated with the liquid anhydrous ammonia at high pressure and a temperature ranging from 60–100 °C for a variable

time (Alvira et al. 2010). The quick release of the pressure results in rapid ammonia gas expansion, causing swelling and physical disintegration of biomass fibers. During a typical AFEX pretreatment process, around 1–2 kg of ammonia/kg of dry-milled biomass is loaded in the AFEX reactor vessel for 30 min (Balan et al. 2009a). According to Bals et al. (2011), the four critical parameters in the AFEX treatment are ammonia to biomass ratio, moisture content, temperature of the reaction, and residence time, which can be variably used in treatment optimization. Various researchers have optimized the AFEX treatment parameters for different feedstock materials like switchgrass (Alizadeh et al. 2005), sweet sorghum (Li et al. 2010a), and hardwood of *Populus nigra* (Balan et al. 2009b). In a study by Bals et al. (2012), corn stover underwent AFEX pretreatment, revealing the flexibility in residence time and temperature during the treatment. According to this research, AFEX treatment of corn stover at the 40 °C temperature for 8 h long residence time produced an almost equal amount of sugar and ethanol as the conventional method of AFEX pretreatment using high temperature for a short duration. The pretreatment conditions and ammonia recovery processes significantly impact ethanol production's cost. Variations in ammonia loading and residence time contribute most towards the economic cost of the output. The study performed by Bals et al. (2011) can be utilized to evaluate the economic optimum of AFEX pretreatment conditions against the maximum yields.

### Supercritical CO<sub>2</sub> (SC-CO<sub>2</sub>) Explosion

In recent years, supercritical CO<sub>2</sub> (SC-CO<sub>2</sub>) is also being used for lignocellulosic biomass pretreatment due to a few advantages like nonflammable, nontoxic, inexpensive, and environment-friendly nature (Kim and Hong 2001). SC-CO<sub>2</sub> is readily accessible at a 31 °C critical temperature ( $T_c$ ) and 7.4 MPa pressure ( $P_c$ ) (Alinia et al. 2010). In this method, CO<sub>2</sub>, as a green solvent, is used to treat the biomass, which diffuses into the crystalline structure of the cellulosic biomass (Gu 2013). The subsequent release of CO<sub>2</sub> pressure causes cellulosic biomass disintegration and increases the accessibility of substrate surface area for enzymatic action during hydrolysis (Zheng et al. 1995). Although this method works similarly to steam and ammonia explosions, it is quite advantageous. It is more economical than the ammonia explosion and prevents inhibitor formation, which usually occurs in the case of a steam explosion (Sarkar et al. 2012). The moisture content present in the biomass during SC-CO<sub>2</sub> pretreatment also significantly changes the final sugar yield during enzymatic hydrolysis. According to Kim and Hong (2001), SC-CO<sub>2</sub> pretreatment at a pressure of 3100 psi and 165 °C temperature for 30 min with a moisture content of 73% showed significantly high net sugar yields of  $84.7 \pm 2.6\%$  in aspen (hardwood) and  $27.3 \pm 3.8\%$  in southern yellow pine (softwood). The untreated or without moisture SC-CO<sub>2</sub>-treated biomass gives almost same amount of sugar yields from aspen ( $14.5 \pm 2.3\%$ ) and southern yellow pine ( $12.8 \pm 2.7\%$ ). Another study on rice straw suggested that lignocellulosic biomass treated with CO<sub>2</sub>-added ammonia results in 97% ethanol yield (Cha et al. 2014). SC-CO<sub>2</sub> has been studied for pretreating various lignocellulosic biomass materials such as rice straw

(Gao et al. 2010), wheat straw (Alinia et al. 2010), sugarcane bagasse (Phan and Tan 2014), etc. to improve the final sugar yield during the enzymatic hydrolysis.

#### 3.4.1.4 Biological Pretreatment

Various chemicals, ionizing radiations, or combinations in different physical and chemical pretreatment methods affect enzymatic hydrolysis and fermentation by generating the process inhibitors. These processes also require special instruments and consume energy (Sindhu et al. 2016). Biological pretreatment is a suitable alternative that uses certain microorganisms (bacteria and fungi) to improve the digestibility of lignocellulosic biomass (Vasco-Correa et al. 2016). It is an ecofriendly and economically viable strategy devoid of chemical use, recyclable, and no toxic compound released into the environment.

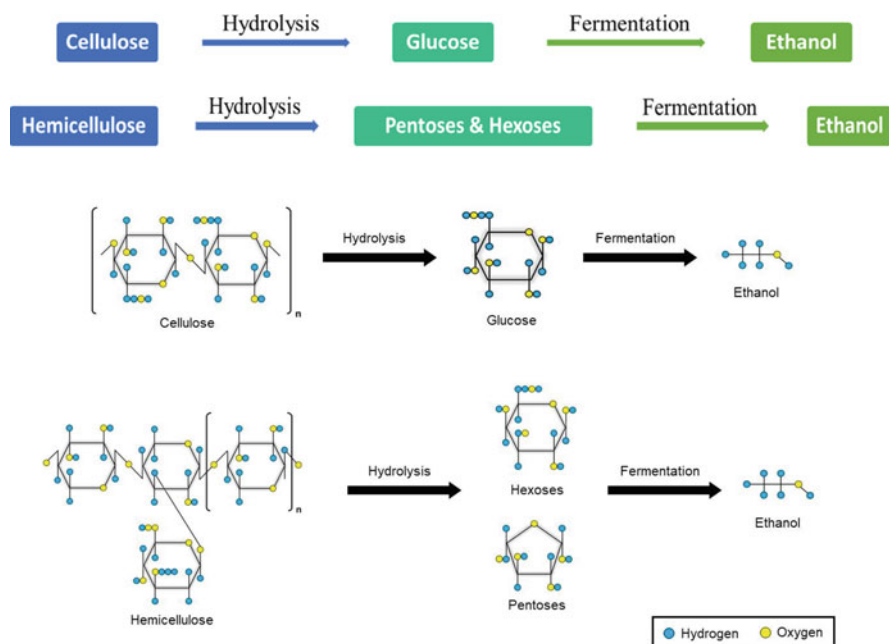
Different microorganisms like brown, white, or soft rot fungi and bacteria are used to disintegrate the biomass by releasing the hydrolytic (hydrolases) and ligninolytic enzymes (Sharma et al. 2019). Biological pretreatments are generally performed by growing microbes directly into the feedstock material or using the extracted enzyme. Efficient biodegradation can be achieved by the combined effect of the microbial community containing both fungi and bacteria (Vasco-Correa et al. 2016).

Few reports have manifested the process of biological pretreatment of different feedstock materials. In a survey by Suhara et al. (2012), 51 fungal strains belonging to white rot basidiomycete *punctularia sp.* were isolated from the decaying bamboo culm to check the selective lignin degradation by the microbes (Rai et al. 2020). A high lignin decomposition (>50%) and improved enzymatic hydrolysis of bamboo culm were observed after 12 weeks of pretreatment using *Punctularia sp.* TUF20056. In another study by Dhiman et al. (2015), rice straw and willow were exposed to simultaneous pretreatment and saccharification (SPS) using a mixture of oxidizing and hydrolytic enzymes obtained from a newly developed fungal consortium. This is the foremost study on environment-friendly and single-vessel SPS methodology, where 74.2% and 63.6% of saccharification were reported for rice straw and willow, respectively. Using a single vessel for pretreatment and hydrolysis makes this strategy more economical. Fungal pretreatment requires a long incubation time (from weeks to months), whereas it takes only a few hours or a day for bacterial and enzymatic pretreatment. Nevertheless, pretreatment with fungi (preferably white rot fungi) is predominantly used due to its high efficiency and increased yields (Zabed et al. 2019).

### 3.4.2 Hydrolysis of Lignocellulosic Biomass

After completing the pretreatment process, hydrolysis of lignocellulosic material is the next step of bioethanol production. In this aspect, feedstock material's cellulosic and hemicellulosic fraction is transformed into pentose and hexose sugars, thereby converting them to ethanol during fermentation. Two hydrolysis processes are





**Fig. 3.6** Schematic representations of hydrolysis products of cellulose and hemicellulose

utilized for ethanol production: acid-catalysed (dilute/concentrated acid) and enzyme-catalysed (Fig. 3.6).

The acid hydrolysis involves the exposure of lignocellulosic biomass to the acid for a certain time at a fixed temperature. Sulfuric acid ( $\text{H}_2\text{SO}_4$ ) and hydrochloric acids (HCl) are used for acid hydrolysis, whereas sulfuric acid is predominantly investigated (Tahezadeh and Karimi 2007). Cellulase enzymatic mixture is used during the enzymatic hydrolysis.

### 3.4.2.1 Concentrated-Acid Hydrolysis

It is an old process operated at low temperature ( $40\text{ }^\circ\text{C}$ ) and low pressure, which generally gives higher sugar yield and subsequently higher ethanol in comparison to dilute-acid hydrolysis. Concentrated  $\text{H}_2\text{SO}_4$  and HCl (30–70%) are used in this process to treat lignocellulosic biomass (Tahezadeh and Karimi 2007). However, the process is highly corrosive and requires expensive non-metallic construction of alloys or ceramics. The acid recovery is also essential in cutting down the commercial value of the final product. Various acid recovery techniques are explored, and it was observed that three methods, i.e. ion exclusion chromatography, solvent extraction, and electrodialysis, are the best performing and most widely used for acid recovery. These techniques are also applied on a large scale, and 90–99% of acid recoveries are reported with low sugar loss (Wolfaardt et al. 2021). Despite the shortcomings, concentrated acid hydrolysis is preferable because of the high sugar recovery efficiency of more than 90% in hemicellulose and cellulose sugars. The

only concern with the process is the environment and acid corrosion, which increase the cost.

### 3.4.2.2 Dilute-Acid Hydrolysis

It is undoubtedly the most commonly used chemical hydrolysis technique, which can be used either for pretreatment of the lignocellulosic biomass preceding the enzymatic hydrolysis or as a method of hydrolysis (Taherzadeh and Karimi 2007). In this process, 0.1–5% of mineral acids like HCl, H<sub>2</sub>SO<sub>4</sub>, and H<sub>3</sub>PO<sub>4</sub> are used at a high temperature of 210 °C. H<sub>2</sub>SO<sub>4</sub> is the most commonly used mineral acid for this process. The sugar yield depends on the process conditions like residence time, temperature, and acid concentration. The Scholler process was probably the first established dilute acid hydrolysis process where woody waste (sawdust and chips) was hydrolysed by using 0.5% H<sub>2</sub>SO<sub>4</sub> at 11–12 bar pressure for 45 min (Faith 1945; Zhou et al. 2021). Mainly, there are two ways of performing dilute acid hydrolysis: continuous flow process (5–10% of solid loading at high (>160 °C) temperature) and batch process (10–40% of solid loading at low (<160 °C) temperature). Generally, batch reactors are most widely used for hydrolysis at the pilot and lab scales. Dilute acid hydrolysis is also investigated in one and two stages to check the effect on hydrolysis. It was observed that glucose yield is better in two-stage hydrolysis, where the solid residuals obtained from the first stage hydrolysis are again subjected to the hydrolysis with the same or different process parameters. According to Karimi et al. (2006), 78.9% of xylan and 46.6% of glucan were digested to glucose and xylose after two-stage hydrolysis, whereas only 25.8% of glucose was yielded from glucan after a single stage of hydrolysis.

Although it is the most commonly used hydrolysis method, the process's main disadvantages are the generation of sugar by-products as fermentation inhibitors. The major by-product compounds generated during the process are furfural, 5-hydroxymethylfurfural, levulinic acid, and formic acid. These inhibitors cause a reduction in the sugar yield and inhibit microorganism growth during the subsequent fermentation process.

### 3.4.2.3 Enzymatic Hydrolysis

Enzymatic hydrolysis has evolved as an essential method because it requires much less energy, no acid recovery, mild environmental conditions (low temperature and neutral pH), and very few fermentation inhibitors are generated. The hemi (cellulolytic) enzymes can break the glycosidic bond of polymeric lignocellulosic biomass and convert it into monomeric forms like pentoses (arabinose, xylose) and hexoses (glucose, mannose, galactose). The optimal conditions for cellulases and hemicellulases are often similar and reported as 40–50 °C temperature and 4–5 pH (Maitan-Alfenas et al. 2015).

The enzymatic method of cellulose hydrolysis is the outcome of a synergistic action of three different enzymatic components of cellulase. These enzymatic components are (1) Endoglucanases—convert cellulosic polymers into the oligomeric form, (2) Exoglucanases—convert those oligomeric forms into the cellobiose, and (3)  $\beta$ -glucosidase—converts cellobiose to glucose. The required amount of

$\beta$ -glucosidase is necessary to provide during the reaction to avoid cellobiose inhibition. The hemicellulose fraction of the biomass is also hydrolysed by a group of enzymes referred to as hemicellulases. Xylan is the major polymer present in the hemicellulosic fraction of biomass. The enzymatic components of hemicellulases are (1) Endoxylanase—which converts xylan polymers into the oligomeric form by randomly acting upon the internal bond (Soni et al. 2020). (2)  $\beta$ -xylosidase—acts upon the non-reducing ends of the xylose chain to release xylose. (3)  $\alpha$ -arabinofuranosidase,  $\alpha$ -glucuronidase, and  $\alpha$ -galactosidase are other accessory enzymes responsible for the cleavage of various xylan constituents (Maitan-Alfenas et al. 2015). Various process parameters like pH, substrate concentration, temperature, enzyme loading, additives, etc., are essential in determining the hydrolysis efficiency. Enzyme dosage is a crucial factor, contributing up to 43.7% of the total ethanol production cost (Szczo drak and Fiedurek 1996).

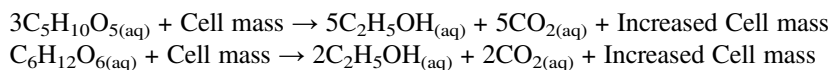
Several microorganisms produce these hemi (cellulolytic) enzymes, including native and genetically modified fungal species of *Aspergillus* sp. and *Trichoderma* sp. Various strategies have also been followed to enhance enzymatic hydrolysis, such as blending the enzymes of two different fungal species origin or using additives like non-ionic surfactants or non-catalytic proteins. According to Rocha-Martín et al. (2017), adding PEG4000 during hydrolysis accelerates the process and reduces the liquefaction time up to 25%.

Since the industrially produced cellulase (mainly produced from *Trichoderma* strains) lacks few necessary enzyme activities, isolation of cellulases from the plant pathogenic fungi (*Phoma exigua* ITCC 2049) is also an excellent alternative (Tiwari et al. 2013). These microorganisms produce the enzyme to digest the cell wall for invading the plant cell. Studies have also been performed on the recycling of enzymes to improve productivity. The most common approach to recycling the enzyme is recovering the enzyme associated with the insoluble biomass fraction after hydrolysis. According to Weiss et al. (2013), the enzyme loading could be decreased by up to 30% by recycling those insoluble biomass fractions to achieve the exact glucose yield under the optimized conditions.

### 3.4.3 Fermentation

It is a post-hydrolysis step in the processing of bioethanol from lignocellulosic biomass. The fermentation is performed using fungus, bacteria, or yeast under oxygen-free conditions. During the fermentation, pentose (arabinose, xylose) and hexose (glucose, mannose galactose) sugar obtained after hydrolysis are converted to ethanol and other products through microbial action. According to Lynd (1996), the maximum possible yield of ethanol production could be 0.51 (mass ethanol/mass carbohydrate) in the absence of cell mass production.

The major reactions that occur during the fermentation are:



After pretreatment and hydrolysis, fermentation is also an important step, where many advancements are needed. Ideally, the microorganism used during fermentation should have a few essential qualities like high ethanol production, ability to utilize multiple substrates, resistance against inhibitors produced during hydrolysis and fermentation, ability to retain functionality at high temperatures, high alcohol and sugar concentrations, and minimal by-products generations. Several microorganisms have reportedly been used for ethanol production from biomass. *Saccharomyces cerevisiae* is a widely used yeast for fermentation, producing a high ethanol yield up to 18% of the broth. This yeast can grow on both monosaccharides and disaccharides and is considered GRAS (generally recognized as safe) as a food additive for human consumption (Lin and Tanaka 2006).

### 3.4.3.1 Fermentation Using Yeast

Conversion of hexose sugars into ethanol can easily be accomplished by traditional fermentation cultures but not pentose sugars due to certain inhibitory substances. However, a few naturally occurring yeast strains (*Candida parapsilosis*, *Pichia stipitis*, and *Candida shehatae*) efficiently metabolize the xylose sugar using xylose reductase and xylitol dehydrogenase enzymes. No naturally occurring microorganism can effectively metabolize both pentose and hexose sugars into ethanol. Most microbes selectively use the substrate from a mixture of different carbon sources due to the carbon catabolite repression, ultimately reducing the process's efficacy. However, some researchers claim to resolve this selective substrate utilization barrier through metabolic engineering. New strains of popular microbial hosts like *E. coli* and *S. cerevisiae* have been developed through metabolic engineering to simultaneously utilize all sugar (pentoses and hexoses) components of lignocellulosic biomass (Zhang et al. 2011; Kim et al. 2010).

### 3.4.3.2 Fermentation Using Bacteria

Other than yeast, Gram-negative bacteria like *E. coli*, *Z. mobilis*, and *Klebsiella oxytoca* have also been engineered to search for industrially suitable microorganisms (Dien et al. 2003). *Z. mobilis* is the most commonly used high ethanol-yielding bacteria but only ferments hexose sugars. Work is also going on in this organism to introduce pentose sugars utilizing pathways through metabolic engineering. Thermophilic bacteria are also key of interest in producing ethanol due to their important advantages like higher operating temperature, broad substrate range, unique and thermostable hemi(cellulolytic) enzyme system, and low viscosity (Chang and Yao 2011). Various thermophilic bacteria including *Clostridium acetobutylicum*, *C. thermosulfurogenes*, *C. thermohydrosulfurium*, *C. thermosaccharolyticum*, *C. tetani*, *Kluveromyces marxianus*, *Thermoanaerobacterium saccharolyticum*, *Thermoanaerobacter ethanolicus*, *Geobacillus* sp., and *Pichia* sp. have been reported for their ethanologenic property (Arora et al. 2015). The external addition of hydrolytic enzymes for saccharification is not required in the case of thermophilic bacteria because they can produce hydrolytic enzymes and perform simultaneous saccharification and fermentation to make the process economical as well.

Thermophiles are also genetically engineered to overcome carbon catabolite repression. *Moorella thermoacetica* was genetically transformed by removing the two phosphotransacetylase genes, *pdul1* and *pdul2*, and incorporating a promoter-controlled native aldehyde dehydrogenase gene (*aldh*). The transformed thermophile shows a high tolerance and ferments glucose and xylose both for ethanol production (Rahayu et al. 2017). However, the industrial use of thermophilic microbes is still a great challenge because of the inherent low tolerance against ethanol and inhibitors produced during the pretreatment. Further trait improvement through metabolic engineering along with the production process optimization are also important aspects for achieving the reality of industrial production through thermophiles. (Chang and Yao 2011).

### 3.4.4 Strategies for Fermentation

Based on the events of hydrolysis and fermentation, various systems are developed for ethanol production using lignocellulosic biomass. These systems are classified as separate hydrolysis and fermentation (SHF), simultaneous saccharification and fermentation (SSF), simultaneous saccharification and co-fermentation (SSCF), simultaneous saccharification, filtration, and fermentation (SSFF), consolidated bioprocessing (CBP) and simultaneous pretreatment, saccharification, and fermentation.

#### 3.4.4.1 Separate Hydrolysis and Fermentation (SHF)

It involves the operation of hydrolysis and fermentation in different bioreactor vessels. The main advantage of this system is that both processes are carried out at their optimized parameters. Inhibition of enzymes by the cellobiose and glucose as end-products is the major drawback of this system. The reaction rate is reduced up to a large extent due to this enzymatic inhibition (Sasaki et al. 2014). The use of high enzyme concentration,  $\beta$ -glucosidase supplementation, sugar removal by ultrafiltration during hydrolysis, or implementation of SSF are a few strategies to reduce enzymatic inhibition (Sun and Cheng 2002).

#### 3.4.4.2 Simultaneous Saccharification and Fermentation (SSF)

During SSF, the production of reducing sugars through hydrolysis and its fermentation are performed simultaneously. This procedure was initialized by Takagi et al. (1977) to minimize the enzymatic inhibition by the hydrolysis end product. In this process, hydrolytic enzyme and fermenting microbe are added into the same reactor along with the biomass. Sugar produced after saccharification is consumed by the microorganism immediately. SSF process has a higher yield than SHF because of reduced enzyme inhibition. It also decreases the production cost by lowering the number of vessels and time of the process by using a single bioreactor for both hydrolysis and fermentation. The ethanol in the reactor vessel also reduces the chances of undesired microbial contamination. Sasaki et al. (2014) compare the production of acetone-butanol-ethanol (ABE) from acorns and wood chips through

SHF and SSF processes using *Clostridium acetobutylicum* NBRC13948 for fermentation. 15.45 g/L of ABE was obtained through SHF in 96 h of fermentation. However, 16.70 g/L of ABE was obtained through SSF in 120 h of fermentation without external addition of hydrolytic enzymes considering that *C. acetobutylicum* possesses amylolytic enzyme. Although SSF is a more advantageous process in terms of cost and time reduction, the major challenge in this process is optimizing process parameters for hydrolysis and fermentation. If the optimum conditions for both processes differ, it must be done at a suboptimum level. The ethanolic inhibition of microbes and enzyme is also a significant issue in this process (Sasaki et al. 2014; Sun and Cheng 2002).

#### **3.4.4.3 Simultaneous Saccharification and Co-Fermentation (SSCF)**

It is an improved SSF process and a feasible option ethanol production from xylose-rich biomass. It involves the co-fermentation of glucose and xylose sugars at a high concentration of water-insoluble solids (WIS) to achieve a high yield of ethanol with the help of genetically modified yeast strains (Olofsson et al. 2010). A recombinant yeast strain *Saccharomyces cerevisiae*, TMB3400 co-fermented xylose and glucose, yields a high final ethanol concentration (Öhgren et al. 2006). The presence of high glucose in the hydrolysate makes xylose utilization difficult due to the competitive inhibition of sugar transport. The prefermentation of initially present hexose into the slurry (Magnus et al. 2009) or the controlled addition of cellulase (Olofsson et al. 2010) could be possible ways to improve the xylose to ethanol conversion.

#### **3.4.4.4 Simultaneous Saccharification, Filtration, and Fermentation (SSFF)**

It is a novel technique of ethanol production developed by Ishola et al. (2013) to evade the disadvantages of SHF and SSF. The process of hydrolysis and fermentation are operated in separate bioreactors to provide the optimum conditions required for both. After hydrolysis, the filtrate which is rich in sugar is transferred to the fermentation vessel through a cross flow-filtration. At the same time, the fermented liquid is transferred to the hydrolysis vessel again. The fermenting organism is retained in the fermentation vessel by settling to reuse several times again. A comparative study between SSF and SSFF shows a negligible difference in the amount of ethanol (85.3% and 84.2%, respectively) obtained in both processes while using the same amount of slurry and enzymes, but the SSFF has an added advantage over SSF that new yeast supply is not needed for each batch in earlier case (Ishola et al. 2013). This process mainly provides two benefits over SSF and SHF: (1) Aforesaid, both hydrolysis and fermentation processes can be operated at their optimum conditions, and (2) The fermenting microorganism can be utilized for their full potential. However, there are a few weak spots in the process, such as the lifespan of the cross-filter membrane and the risk of cross-contamination in the yeast culture while using it several times.

#### 3.4.4.5 Consolidated Bioprocessing (CBP)

It is a recent, simplified, one-step process of converting the lignocellulose into the desired products without adding the enzyme using a single vessel (Singhania et al. 2022). This strategy applies to produce a wide range of products, mainly for ethanol production, which was commercially used by Olson et al. (2012). Generally, four important biological events, i.e. (1) production of cellulases and hemicellulases, (2) hydrolysis of pretreated biomass to sugars, (3) hexose sugars fermentation, and (4) pentose sugars fermentation, are performed separately or (some of them are combined in less highly integrated configurations). However, all these transformations are executed in a single reactor in one step, called CBP (Lynd et al. 2005).

Generally, the microorganism with combined properties of utilizing substrate and product formation are required for CPB. These microbes are not available naturally but could be developed using an organism development strategy. There are mainly two strategies followed to develop the microorganism with desired properties. (1) **Native cellulolytic strategy**—This approach involves engineering a naturally occurring microorganism that quickly degrades cellulose to improve its product-related properties, such as yield and titer. (2) **Recombinant cellulolytic strategy**—This strategy involves the use of non-cellulolytic microorganisms with desired product formation qualities like high yield and titer. The main objective of this strategy is the heterologous expression of a saccharolytic enzyme system in the organism. This approach has been applied to several host organisms such as *Bacillus subtilis*, *S. cerevisiae*, and *E. coli*. However, *S. cerevisiae* has been investigated most till date (Kashyap et al., 2019; Olson et al. 2012). CBP is a promising approach that reduces the overall cost of production by eliminating the need for external enzyme addition and circumventing the restrictions of the conventional workflow for bioethanol production. CBP also requires a lesser no. of reactor vessels, significantly reducing the maintenance and capital expenses of the process (Jouzani and Taherzadeh 2015). However, the low conversion efficiency is the major obstacle to commercializing the process (Singhania et al. 2022).

#### 3.4.4.6 Simultaneous Pretreatment, Saccharification, and Fermentation

It is based on a recent study conducted by Li et al. (2022) for bioethanol production through an integrated process using different microbes. In this method, pretreatment and saccharification of the lignocellulosic biomass have been performed by *Pecoramyces* sp. F1 (an anaerobic fungus), and the simultaneous fermentation was carried out by *Zymomonas mobilis* (a facultative anaerobic bacteria). Both of the microbes are co-cultured together to avoid the requirement for additional biomass pretreatment. According to Li et al. (2022), 0.32 g of ethanol yield was obtained from 1 g of glucose after continuously conducting the process for 4 days.

### 3.5 Ethanol Recovery

Ethanol can be easily recovered from the fermentation broth using recovery techniques, such as distillation, solvent extraction, gas stripping, steam stripping, membrane pervaporation, and adsorption. Out of these, distillation is the most commonly used separation technique for large-scale production. All ethanol, an almost equal amount of water, and a considerable quantity of other materials such as proteins, oil fibers, etc., are captured through the beer column during the first step of distillation. Furthermore, ethanol is purified with the aid of the stripper, rectifier, and molecular sieves by capturing the last bit of water and creating 99.6% pure ethanol (Kwiatkowski et al. 2006). However, the distillation process is unsuitable for small-scale production due to high energy demand. The ethanol recovery by distillation is not economically feasible if the ethanol concentration in broth is below 5% (Gírio et al. 2010).

Among all separation techniques, pervaporation is also the most promising recovery technique in terms of simplicity, less distillation, and energy consumption. This technique is ideal in case the fermentation broth is of low concentration and can be used before distillation process. Pervaporation works on the mechanism of solution diffusion mechanism under the influence of gradient force developed between the two sides of the membrane: the feed and permeate side. Moreover, there are two types of pervaporation processes: (i) vacuum pervaporation and (ii) sweep gas pervaporation (Huang et al. 2008). Choosing the right material for the membrane depends upon the particular component. Organic compounds will be found in the permeate in the case of the hydrophobic membrane. On the other hand, if the membrane is hydrophilic, the mixture feed gets dehydrated, and water will be recovered from the permeate (Zentou et al. 2019). Although pervaporation is the new membrane separation technique and has become economically competitive for some commercial processes, membrane fouling is the biggest challenge leading to the productivity loss. Repetitive cleaning is required to maintain the membrane permeability and to reduce microbial growth over the membrane.

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### 3.6 Conclusions

The continuous increase in worldwide energy demand and the reducing natural resources have created a challenging situation for researchers to think about alternate energy sources. Bioethanol production using lignocellulosic biomass is a significant alternative renewable resource for ethanol production. However, the process is not economical compared to the traditional first-generation bioethanol production using corn and sugarcane. The primary concern about first-generation bioethanol production is the food vs. fuel debate, and an increase in this type of bioethanol production may lead to a rise in food prices. Agricultural waste is renewable, cheap, abundantly available lignocellulosic biomass with no food value, and also, no extra land is needed to grow this material.



Bioethanol production using lignocellulosic waste material consists of four major aspects, i.e. feedstock material, pretreatment method, hydrolysis, and fermentation technology. Several pretreatment methods are available based on the feedstock material. A single pretreatment methodology cannot be applied to all the feedstock materials. The pretreated material is hydrolysed using cellulosic enzyme or acid hydrolysis technology. The enzymatic hydrolysis is a more robust method of saccharification. The major challenge during hydrolysis is achieving efficient cellulose and hemicellulose fibers depolymerization for further fermentation. The fermentation process also has hindrances, such as finding a suitable microorganism that utilizes both pentose and hexose sugars. However, this limitation is fulfilled by using a few transformed thermophiles, although they have less tolerance against continuously increasing ethanol concentration during fermentation. Concerning fermentation strategies like SHF, SSF, SSCF, SSFF and CBP have been explored to make the process more economical. These fermentation strategies also have certain limitations, like enzyme inhibition by the end product in the case of SHF. In contrast, optimizing similar process conditions for hydrolysis and fermentation is another challenging task in SSF. In conclusion, it is an excellent alternative for ethanol production using lignocellulosic biomass. Still, more research is needed to provide an efficient and economical strategies for feedstock collection, pretreatment, hydrolysis, and fermentation.

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# Sewage and Wastewater Management to Combat Different Mosquito Vector Species

# 4

Varun Tyagi, Santana Saikia, Dipanjan Dey, Anjana Singha Naorem, Vivek Tyagi, P. Chattopadhyay, and Vijay Veer

## Abstract

Vector-borne disease consists of 17% of the total infectious disease. As per the World Health Organization (WHO) report, more than 700,000 deaths occur annually due to vector-borne diseases caused by bacteria, parasites, or viruses. Mosquitoes are the deadliest vector that carries parasites to human and animal bodies. Mosquito breeding areas are diverse, and the magnitude of the mosquito population depends on the natural or artificial breeding areas hence the chances of mosquito vector-borne diseases increase. Most mosquitoes choose semi-arid regions where the wastewater irrigation system gives a constant water source for mosquitoes to breed. They prefer wastewater or sewage water for breeding or laying their eggs when suitable physical, chemical, and biological conditions are insufficient. In Urban areas, water pollution mainly occurs because of the continuous discharge of untreated wastewater into natural streams. Even water stabilisation ponds for urban wastewater treatment sometimes provide suitable breeding sites for mosquitoes. Ultimately these events contribute to the discrete of

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V. Tyagi (✉)

Defence Research Laboratory (DRDO), Tezpur, Assam, India

Eurofins Agrosience Services, Tiruppur, Tamil Nadu, India

S. Saikia · A. S. Naorem

Department of Zoology, Cotton University, Guwahati, Assam, India

D. Dey

Department of Zoology, Tihu College, Tihu, Assam, India

V. Tyagi

Ministry of Fisheries, Animal Husbandry and Dairying, New Delhi, India

P. Chattopadhyay · V. Veer

Defence Research Laboratory (DRDO), Tezpur, Assam, India

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natural waterways which associate with the increased population of mosquitoes and could cause a risk to human health. So, there is an urgent need to properly manage this sewage water/wastewater produced from households, industries, and other hospitals, private nursing homes, etc. Keeping all these in mind, in this chapter, we shall briefly discuss the relationship between wastewater and mosquito-borne illness and some management or control strategies of sewage water/wastewater to combat mosquito and mosquito-borne diseases.

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**Keywords**

Sewage water · Wastewater · Vector-borne disease · Mosquito

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## 4.1 Introduction

India is the second most populous country in the world, with a population of 1.21 billion per the 2011 census. With an increasing population, it is facing several challenges related to health problems. The disease epidemiology of India is very complex due to diverse ecological factors and various disease-causing vectors. Parasites cause vector-borne diseases, bacteria, and viruses, and these pathogens are transferred mainly by blood-sucking insects. According to a report published by WHO in 2020, more than 700,000 people die of the vector-borne disease every year, accounting for approximately 17% of the total deaths due to infectious diseases. Dengue is another fatal mosquito vector-borne disease. Globally, 3.9 million people across 129 countries are prone to Dengue, with an estimated 40,000 deaths occurring every year, wherein approximately 96 million cases are without any symptoms (WHO). Other viral diseases like Zika virus, Chikungunya fever, Yellow Fever, West Nile fever, and Japanese Encephalitis are also mosquito-borne. So, we can infer that mosquitoes are the deadliest animal globally, which spreads diseases and can cause millions of deaths in humans each year. The study report suggests *Culex* mosquitoes (especially *Culex quinquefasciatus* and *Culex tritaeniorhynchus*) prefer anaerobic water systems that receive untreated wastewater, where dissolved oxygen (DO) content is low. In typical laboratory conditions, mosquito size decreases due to population density. Still, the mosquitoes which are collected from sewage water emerge faster, sizes bigger, and the ratio of a female is more than male mosquitoes. These traits may help to regulate the population of mosquitoes.

According to the Down to Earth report, 78% of the generated sewage remains untreated in India. So, this sewage water is either directly released into the sea/river or remains logged in some areas, where it can play a significant role as a mosquito breeding bed. The anaerobic water bodies, where the levels of ammonium, biochemical oxygen demand (BOD), phosphorus, turbidity, etc. are high, show propagation of *Culex* species like *Culex pipiens*, *Culex quinquefasciatus*, and *Culex tritaeniorhynchus*. So, this has been a severe threat to the human population. The best approach to controlling the mosquito species takes the benefit of all the

mosquito life stages to attain control, using a combined tactic referred to as integrated pest management.

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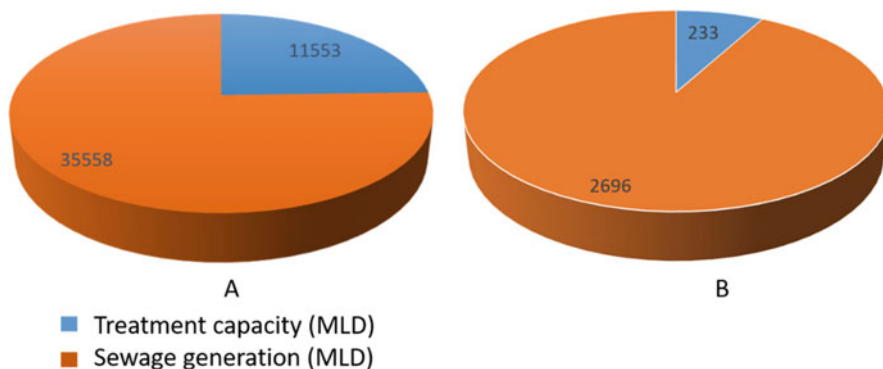
## 4.2 Indian Scenario of Wastewater and Sewage Problem

The water condition of India is very pathetic, and the quality of water has been degrading day by day. The municipal wastewater generated by urban areas is about 61,754 MLD (Kamble et al. 2019). But the available wastewater treatment in urban areas is only 22,963 MLD (CPCB 2016; Kamble et al. 2019). Hence, approx. 62% of untreated wastewater is directly released into the nearby water bodies. The water of rivers and lakes is getting polluted due to excessive anthropogenic activities. Despite several steps, including the National River and Lakes conservation plan, taken by the government and non-government organisations (NGOs), the water quality is still unsatisfactory (Goel 2006). The central fact behind this unsuccessful management is the poor coordination between the government and proper management plants, lack of human resources, and inadequate electricity facilities in India; without which industrial effluences can't be treated and directly released into larger water bodies.

Unmanaged water sources and man-made polluted water are pivotal sources for mosquito breeding. The adult mosquito population depends on the availability of both natural and artificial breeding sites for oviposition and thus increases the likelihood of vector-borne diseases (Banerjee et al. 2013). Household wastes also contribute to environmental pollution and affect natural constancy (Gomez-Dantes and Gutierrez 1992; Gupta et al. 1998; Hamer 2003; Nath 2003; Kumar et al. 2007; Sujauddin et al. 2008; Chakrabarti et al. 2009; Banerjee et al. 2013).

There are a total of 234 sewage wastewater treatment plants in India. Out of all these plants, most of them were formed under river action plans from 1978 to 1979 onwards (Kaur et al. 2012). These plants are situated in 5% of the town on the bank of some major rivers. As per the report of CPCB, total wastewater generation from Class I cities is 35,558 MLD, and Class II towns are 2696 MLD (Kaur et al. 2012 (Fig. 4.1).

Except for the domestic sewage, industries contributed 13,468 MLD wastewater, out of which only 60% is treated. Oxidation pond or activated sludge procedure is the most widely used technology in Class I cities, including 59.5% of total installed capacity (Fig. 4.1) (Kaur et al. 2012). Then the next technology is the Anaerobic Sludge Blanket Technology which covers 26% of the total installed capacity. In some cities, Waste stabilisation pond technology is also used. In India, there are no separate guidelines for wastewater management. However, some existing guidelines are based on environmental laws and legal provisions. Some of them include Constitutional Provisions on sanitation and water pollution; National Environment Policy, 2006; National Sanitation Policy, 2008; Hazardous Waste 6 (Management and Handling) Rules, 1989; Municipalities Act; District Municipalities Act etc.(Kaur et al. 2012), and so on. The state government is responsible for the management of sewage treatment and disposal.



**Fig. 4.1** Sewage generation and treatment capacity in 498 Class I cities and 410 Class II towns in India. (MLD million litres per day)

As per the report prepared by Water Act 1974, State Pollution Control Boards have all the authority to take necessary action against any defaulting agency. Water Act 1974 also focused on utilising treated sewage in irrigation. But all these efforts have been ignored by the State governments.

### 4.3 Relation of Water Pollution with Population and Rapid Industrialisation

Pollution cannot be eradicated from the earth as long as even a single man exists on earth because man is the main reason for pollution. Though the amount of pollution caused by a single man's daily activity is negligible, the combined effect of a larger population is still significant. The relationship between population density and water pollution is straight; therefore, the amount of water pollution in urban areas is more than that in a rural area with less population. In this context, urbanisation is crucial in establishing a relationship between pollution and population, particularly in a developing country. Over the last few decades, India has also witnessed a drastic rural-to-urban population migration, thereby creating higher population densities in cities. Urbanisation involves many construction activities, thereby creating energy demand and resource depletion which, in the long run, impact the quality of the environment and the health of flora and fauna. Population explosion in urban areas and unplanned human settlements create unhygienic living conditions. Moreover, rapid industrialisation in developing countries also accounts for the injudicious discharge of waste waters and industrial wastes, which sometimes find their way directly to the natural habitats. Hence, unplanned human settlements with improper sewage management, urbanisation, and industrialisation are some of the pollution factors, particularly water pollution. These conditions are suitable for the proliferation of a variety of mosquito species which, in the long run, are responsible for a wide range of diseases and health issues in humans.

## 4.4 Sewage and Waste Water Management

Sewage treatment sites can be a potential source of vector mosquitoes (Whelan 1981, 1984, 1988). Sewage effluents usually are nutrient-rich and can produce massive numbers of mosquitoes (Whelan 1988). As the sewage treatment sites are normally close to communities, these vectors have a significant role in public health problems. In developing countries like India, where rapid urbanisation occurs, proper sewage and solid waste disposal is a significant concern. Earlier research has also revealed that breeding mosquitoes, mainly Dengue vectors in solid wastes in North-Eastern India, pose a severe threat to the Dengue outbreak in this region (Rodrigues and Dantawate 1997; Barua and Mahanta 1996). The preliminary design, operation, and maintenance or defective mechanism of sewage effluent disposal are the primary cause of mosquito breeding in sewage treatment plants. These troubles can be rectified in the planning phase. Increased awareness among designers, operators, and regulators of sewage is highly needed to combat this problem.

## 4.5 Different Breeding Habitats

Mosquitoes act as a transmitting agent for varieties of vector-borne diseases. The breeding sites of mosquitoes are diverse, which creates trouble in regulating the vector mosquito population (Adeleke et al. 2008; Medronho et al. 2009; Irwin et al. 2008; Banerjee et al. 2010, 2013). In recent years, the distribution pattern of mosquito and mosquito-borne diseases has been changing due to the increasing rate of environmental sleaze, climate change, increased urbanisation, and the development of resistance in mosquitoes against pesticides and drugs (Gubler 1998; Patz et al. 1996; Jetten and Focks 1997; Simsek 2004). Evaluation of both basic and ecological data is obligatory before planning any mosquito control program (Simsek 2004). Particularly, for an effective program on vector control populations, information regarding the distribution and abundance of mosquito populations in different breeding habitats is crucial (Dutta et al. 2010).

*Aedes aegypti* is the primary causative vector of Dengue haemorrhagic fever in the Americas and Asia (Lenhart et al. 2005; Philbert and Ijumba 2013). This is also responsible for transmitting Yellow Fever in African urban and peri-urban areas. In some regions of India, it was reported to cause Chikungunya fever, Rift valley fever, and Nile encephalitis viruses in Eastern Africa (<http://www.24drtravel.com/travel-health-news> accessed on 24/6/2010; Philbert and Ijumba 2013). *Aedes aegypti* mosquitoes prefer to breed in domestic and semi-domestic polluted water (Philbert and Ijumba 2013). A study report suggested that all man-made containers filled with rainwater were also the leading breeding site of *Aedes aegypti*, in Dares Salaam, Tanzania (Surtees 1968; Philbert and Ijumba 2013). Trpis (1972) found that tyres, tins, wrecked motor cars, waterpots, snail shells, coconut shells, and tree holes as the most potent breeding habitats of *Ae. aegypti*, of which tyres were the most important and provided a constant source of *Ae. aegypti*.

Malaria is prevalent in the foothills areas of northeastern states of India viz. Assam, Arunachal Pradesh, Meghalaya, Manipur, and Tripura, except Sikkim (Mohapatro et al. 1998; Dutta et al. 2010). Two vector species of *Plasmodium falciparum*, viz. *Anopheles minimus* and *Anopheles dirus*, which is recently taxonomically revised as *Anopheles baimaii* (Sallum et al. 2005), have been playing a significant role in the transmission of Malaria in these areas (Prakash et al. 2006). As per the earlier reports, *Anopheles dirus* have the affinity to breed in temporary waterlogged areas (Dutta et al. 2010). According to a survey done by Dutta et al. (2010), it was observed that *Anopheles* mosquitoes choose forest fringe areas for breeding purposes. Some major preferable sites for vector breeding are shallow-water logging areas like ditches, animal hoof markings, and elephant footprints. Though most mosquitoes prefer to lay eggs in freshwater, species like *Anopheles gambiae* Giles (Roberts 1996) and *Anopheles stephensi* Liston (Roberts 1996) can breed in saline water; even in seawater, they can survive.

*Culex* mosquitoes are usually found in tanks with a wide variety of salinities (Roberts 1996). Studies reveal that *Culex* females prefer to lay eggs in 28% seawater; they avoid freshwater for oviposition. Ray and Choudhury (1988) found that maximum *Culex sitiens* survive in 55% seawater. *Culex quinquefasciatus* tend to oviposit in water rich in organic materials (Subra 1982; Roberts 1996) because of their attraction toward a high concentration of free ammonia and nitrates (Sinha 1976).

*Culex gelidus* was an exotic species of *Culex* and was first recorded in the Northern Territory in 1996 (Whelan et al. 2001). *Culex gelidus* is a primary vector of Japanese encephalitis. It is highly susceptible to Murray valley encephalitis virus, Kunjin virus, and Ross river virus, indicating that this species has a significant public health concern (Johnson et al. 2006). These mosquito species have been found breeding in wastewater ponds and high nutrients water bodies.

*Culex annulirostris* is found in shallow, vegetated freshwater swamps, streams, and lagoons. It prefers artificial breeding places like secondary sewage treatment and evaporation ponds and sewage pond effluent (Whelan 1984, 1988). The larvae of *Culex aanulirostris* are mostly found in still and sheltered areas where the larvae get protection from vegetation disruptive waves and aquatic predators. This is an important vector of Murray valley encephalitis, Kunjin virus, Ross river virus, and Barmah forest virus. Dengue is an urban disease directly related to the rapid development of urban areas. Therefore, with the growth of the urban regions, diseases like Dengue, Malaria, and other vector-borne diseases are also increasing.

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## 4.6 Common Vector-Borne Diseases in India

Malaria is a life-threatening disease transmitted by the bite of a female *Anopheles* mosquito. According to the World Health Organization (WHO 2021a), there were 229 million cases worldwide and approximately 409,000 deaths in 2019. Children below the age of 5 years are more vulnerable, as it was 67% of the total malaria deaths worldwide in 2019. Mainly five species of parasites can cause Malaria in

humans, and out of all these parasites, 2 of them- *P. falciparum* and *P. vivax* cause a significant threat to humans. In 2018, *P. falciparum* accounted for 99.7% of estimated Malaria cases in the WHO African Region, 50% of cases in the WHO South-East Asia Region, 71% of cases in the Eastern Mediterranean, and 65% in the Western Pacific. *P. vivax* is the predominant parasite in the WHO Region of the Americas, representing 75% of Malaria cases. According to the latest World Malaria report, released in December 2019, there were 228 million Malaria cases in 2018 compared to 231 million cases in 2017.

Chikungunya disease is transmitted to humans by the infected *Aedes* mosquito species. The main symptoms of the disease are fever and joint pain, and the other symptoms include headache, muscle pain, nausea, fatigue, and rash. There is no cure for this disease; only its symptoms can be relieved by treatment. Asia, Africa, and the Indian subcontinent are where Chikungunya mainly occurs. In 2015, a major outbreak of this disease affected many countries in the American region (WHO 2020b). The first and most significant outbreak of this disease occurred in the Islands of India in February 2005. In 2006 and 2007, there were several cases reported from India. Since 2005, Indonesia, India, Myanmar, Maldives, and Thailand have reported over 1,900,000 cases (WHO 2020a). Both *Ae. aegypti* and *Ae. albopictus* are the main causative organism for the Chikungunya outbreak. *Ae. aegypti* is restricted within the tropics and sub-tropics and *Ae. albopictus* occurs in temperate and even cold temperate regions. In recent decades, *Ae. albopictus* has spread from Asia and has become reputable in Africa, the Americas, and Europe. The last Chikungunya outbreak took place on 1st May 2019 in Congo. From 1st January to 14th April 2019, 6149 suspected cases of Chikungunya were reported in the country where approximately 54% of reported cases were female (WHO 2020b).

Dengue disease has been growing increasingly around the world over the last decades. Most of the cases are asymptomatic. Hence, many cases remain unreported. According to a report prepared by Bhatt et al. 2013, 390 million people are infected with Dengue every year. Another report suggests that 3.9 billion people are at risk of infection with the Dengue virus in 129 countries (Brady et al. 2012), out of which 70% of cases are from Asia (Bhatt et al. 2013). In 2000, the total number of reported Dengue cases to WHO was 505,430, approximately 2.4 billion, in 2010 and 4.2 million in 2019 (WHO 2021b). From this report, the severity of this disease can be easily observed. Dengue virus belongs to the family Flaviviridae, which consists of four types, viz. DENV-1, DENV-2, DENV-3, and DENV-4. Transmission of the Dengue virus to human occurs by the bite of female *Aedes aegypti* mosquitoes. *Aedes albopictus* is also a secondary vector for Dengue transmission in Asia.

Japanese encephalitis virus, another mosquito-borne flavivirus, is transmitted by *Culex* mosquitoes and is Asia's most important cause of viral encephalitis. According to a review report, globally, 68,000 cases are reported with approximately 13,600 to 20,400 deaths, and around three billion people are at risk of infection (WHO 2019). Though individuals of any age can be affected, this viral disease mainly targets children. The symptoms of Japanese encephalitis are mild headache and fever, but approximately 1 in 250 reported cases are clinical illnesses. In the case of children, gastrointestinal pain and vomiting may occur as initial symptoms. The

severity of this disease can be detected as high fever, headache, neck stiffness, disorientation, coma, seizures, paralysis, and ultimately, death.

Lymphatic filariasis or elephantiasis is caused by parasites that are transmitted to humans through the bite of infected mosquitoes. It is a very painful infection characterised by disfiguration of the infected region. Mosquitoes transmit the larvae to the skin of a human, from where they enter the body and migrate to the lymphatic vessels. Humans of all ages are affected by this disease. In endemic countries, this disease significantly impacts public health and the economy. Lymphatic filariasis affects over 120 million people in 72 countries throughout the tropics and subtropics of Asia, Africa, the Western Pacific, and parts of the Caribbean and South America (WHO 2021c).

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## 4.7 Mosquito Control Techniques

Mosquitoes can transmit life-threatening diseases worldwide, and it is on the rise in many tropical and subtropical countries. Therefore, mosquito control strategies are highly warranted. Many age-old practices for controlling the mosquito population have been in practice for a long time. Since the resistance of vectors against different pesticides is growing in many areas, controlling the vector population to combat different vector-borne diseases is challenging and equally effective (Poopathi and Tyagi 2006). The development of resistance against different synthetic drug vectors has made scientists think of alternate ways for vector control, which should be cost-effective, easily applicable, and environmentally friendly. As a result, interest in integrated vector control strategies has been developed (WHO 1982). A single method of controlling vectors was insufficient to give effective results, so the emphasis was laid on comprehensive mosquito control methods, including insecticides, biocontrol agents, and environmental management.

All the mosquito control measures can be divided into three categories:

1. Chemical-based control techniques.
2. Nonchemical-based control techniques.
3. Use of biocontrol agents.

### 4.7.1 Chemical-Based Control Techniques

Many advanced countries, including India, widely adopt Chemical-based mosquito control techniques. Various chemicals like insecticides, larvicides, and adulticides are normally sprayed on mosquito breeding sites. The other management techniques include using insecticide-impregnated paint, which is effective against *Culex quinquefasciatus* (Das et al. 1986; Poopathi and Tyagi 2006). Insecticide-impregnated mosquito nets are prepared with deltamethrinropes, which are helpful against *Anopheles* and *Culex* species (Sharma et al. 1989; Poopathi and Tyagi 2006).



- (a) *DEPA spray*: It is a multi-insect repellent spray developed by DRDO. The principal component of this spray is N, N Diethylphenylacetamide. This spray can give protection for up to 6–8 h from mosquito bites if it is sprayed on curtains, fabric, or skin. This product got approval from the Drugs Controller General of India (DCGI) and the Director General of the Armed Forces Medical Service (Das 2010).
- (b) *Anti-mosquito paint for rooms*: It is a quick-drying paint with an insecticidal property that lasts 2 years. It was developed by Defence Research Laboratory, DRDO, Tezpur. Besides decoration and preservation of wooden and metallic surfaces, it keeps away mosquitoes, cockroaches, and other insects by releasing insecticides from the paint.

#### 4.7.2 Non-Chemical-Based Control Techniques

The use of chemicals that are non-biodegradable in controlling insects often leads to the deposition of trash elements of the chemicals in the environment, and mosquitoes slowly develop resistance against those synthetic chemicals. Therefore, new techniques evolved gradually. This is achieved by spraying biodegradable non-ionic chemicals on the mosquito breeding sites, which results in the formation of monomolecular film (i.e. layer of the compound having a single molecule on the water surface and eventually leads to the death of the mosquito larvae). These chemicals are termed Surface Active Agents and are effective against *Culex quinquefasciatus*, *Anopheles stephensi*, and *Aedes aegypti* (Das et al. 1986; Poopathi and Tyagi 2006). Curtis and Minjas (1985) and Sharma et al. (1985) showed that expanded polystyrene beads (EPS) were very effective against *Culex quinquefasciatus* and *Anopheles stephensi*.

Some chemicals were observed to be chemically similar to natural juvenile hormones of insects, and these chemicals are called mimics or Juvenoids (Slama et al. 1974; Mulla 1995). Some other compounds inhibit the formation of cuticles in insect bodies. They are not chemically similar to juvenile hormones but can produce similar effects like JH (Mulder and Gijswijt 1973; Wellinga et al. 1973; Post et al. 1974; Grosscurt and Tipker 1980; Itoh 1981; Mulla 1991). All these chemicals are designated as Insect Growth Regulator (IGR). The publication of “Silent Spring” by Rachel Carson in 1960 raised awareness among the public about the bad impact of pesticides on the environment, characterised by high mammalian toxicity, poisoning risk to non-target organisms, and accumulation in the food chains (Mulla 1994). During the first quarter of the twentieth century, mosquitoes were controlled by source reduction through substances used to kill mosquito larvae, such as the use of petroleum oils and fishes that feed on mosquito larvae (Mulla 1994). Before the turn of the last century, interest has grown in the biological control of vectors (Lamborn 1890):

- (a) *Herbal mosquito repellents and herbal floating tablet mosquito larvicide*: DRDO, Tezpur identified nearly 20 different indigenous plants whose extracts

**Table 4.1** Names of essential oils obtained from different plant sources used as a larvicide against *Ae. albopictus* (Source of oils: Fragrance and Flavour Development Center, Kannur, U.P, India) (Tyagi et al. 2015)

Sl no	Name of essential oils	Botanical name	Extraction	Part used	Uses
1.	Amyris	<i>Amyris balsamifera</i> Linn.	Steam distillation	Wood	Antiseptic, antiaging, antistress, balsamic, sedative. It also acts as muscle relaxant, soothing agent
2.	Black pepper	<i>Piper nigrum</i> Linn	Steam distillation	Seed	Antiseptic, anticholerin, antiasthmatic, fever, cough
3.	Cinnamon	<i>Cinnamomum zeylanicum</i> Linn.	Steam distillation	Bark	Antioxidant, antiseptic, constipation, gastric, and irritation
4.	Dill	<i>Anethum graveolens</i> Linn.	Steam distillation	Seed	Antiseptic, stomachic, low blood pressure
5.	Jasmine	<i>Jasminum grandiflorum</i> Linn.	Hydro distillation	Flower	Dry skin, coughs, disorders of the chest
6.	Juniper	<i>Juniperus communis</i> Linn.	Steam distillation	Fruit	Antiseptic, obesity, urinary, antiseptic, digestive
7.	Thyme	<i>Thymus serpyllum</i>	Steam distillation	Leaves	Antiseptic, bronchitis, coughs and common cold, diarrhoea

and oils can be used as mosquito repellents. These were biodegradable as well as cost-effective. Herbal floating tablets are biodegradable and effective for 30 days. One single tablet is sufficient for one square meter of open water surface. These tablets are normally used in areas like tree wholes where water can be collected, and mosquitoes may lay eggs.

- (b) *Essential oils*: Different plant extracts for controlling mosquitoes have several beneficial impacts as they are a biodegradable, less hazardous, and rich source of stock (Tyagi et al. 2016). These plant products have efficient larvicidal (Vasudevan et al. 1989; Ansari et al. 2000; Anyanwu et al. 2001; Yadav et al. 2014, 2015; Tyagi et al. 2015) (Table 4.1), oviposition attractant, and ovicidal activity (Millar et al. 1992; Su and Mulla 1998, 1999; Ritchie 2001).

### 4.7.3 Biocontrol Method

Biocontrol agents for controlling mosquitoes can be categorised into eight types, out of which four are microbial agents (viruses, protozoans, fungi, and bacteria) and four are multicellular agents (nematodes, cyclopoid copepods, predaceous aquatic insects, and larvivorous fish) (Knight et al. 2003). Under specific conditions, particular species of parasites or predators can cause mortality in mosquito populations. However, in large-scale control, only some sets of these agents can be useful in constructed wetlands (Knight et al. 2003). These biological agents can be effective

only in the case of larvae of mosquitoes, but there are no effective biological control agents for adult mosquitoes. Mosquito-specific bacteria and larvivorous fishes are the most effective agents for controlling immature vector populations (Chapman 1985).

#### 4.7.3.1 Mosquito-Specific Bacteria

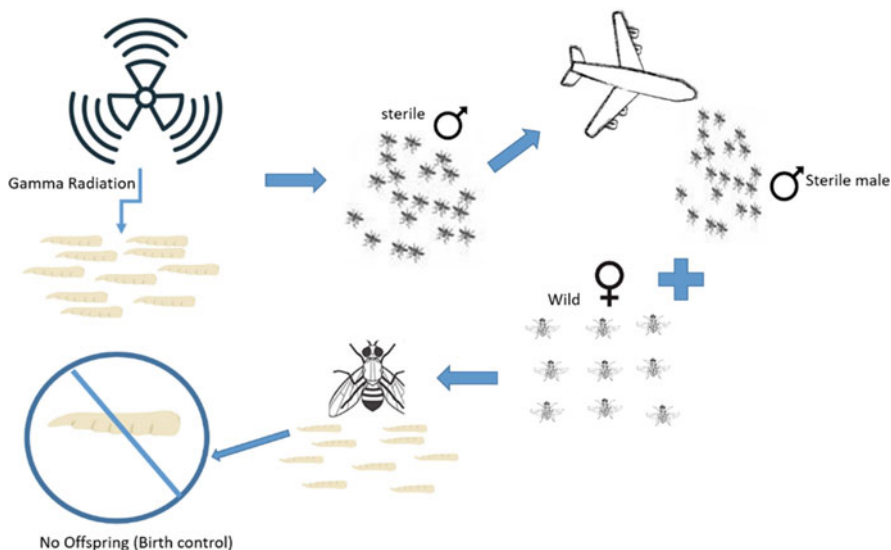
In the United States, two *Bacillus* species have been used against mosquitoes (Knight et al. 2003). *Bacillus thuringiensis* variety *israelensis* (Bti) was registered in 1981 for controlling mosquitoes, whereas *Bacillus sphaericus* (Bs) was approved for larval control in 1991 (Knight et al. 2003). In wastewater with suspended sediment or elevated organic matter, *Bacillus sphaericus* gives more effective results in controlling the mosquito population than *Bacillus thuringiensis israelensis*. *Bacillus* toxins have certain pathogenicities and are safe for humans and non-target organisms (Mulla 1990; Walton and Mulla 1992; Knight et al. 2003). The toxins released by *Bacillus* are short-lived and degraded quickly by UV light in aquatic environments (Kashyap et al. 2019). It is very effective in wetland treatment. The efficacy of the toxins against mosquito larvae depends on the water quality, larval density, solar radiation, vegetative cover, and flow regime (Walton and Mulla 1992).

#### 4.7.3.2 Larvivorous Fishes

Using *Gambusia affinis* fishes to control mosquitos' larvae is another technique that has been widely used for over 80 years. But recently, this control method has become controversial because *Gambusia* fishes affect local fish fauna's biodiversity and abundance (Gamradt and Kats 1996; Rupp 1996; Knight et al. 2003), particularly certain rare fishes in western drainages. These mosquito fishes can tolerate various environmental conditions (Knight et al. 2003). This method is effective in only a subset of habitats to which fish have been released (Rupp 1996).

Sterile insect technique (SIT) is an advanced technique that is potential in vector control programs. The technique has been developed and validated for methodical mass-rearing mosquitoes and eradicating and releasing *Aedines* and *Anophelines* (Lees et al. 2015). The Sterile insect technique was developed by E. F. Knippling and R. C. Bushland in 1930 (Vreysen et al. 2006). The first successful application of SIT was to control a devastating pest of cattle named Screwworm (*Cochliomyia hominivorax*) in Curacao in 1953. The whole process of SIT is mainly divided into three steps: mass rearing of the male insects, radiation-mediated sterilisation, and release of sterile insects (Reiter 2007). Successful mating of the sterile insect with the wild female will result in no offspring. The target pest population will decline if enough sterile males are released into the environment (Knippling 1955; Wilke et al. 2009). Eliminating the targeted vector population will reduce the transmission of vector-borne diseases; hence, this vector control method has been effective in many areas (Pates and Curtis 2005).

SIT is target specific (i.e. it acts only on the target insect species) and is also non-toxic since no toxic chemicals are used in the process. Hence it does not release any toxic agents into the environment, thereby reducing pesticide use and promoting integrated pest management (IPM). Some limitations of SIT lie in the fact that a good



**Fig. 4.2** Pictorial representation of the Sterile Insect Technique

laboratory set-up should be there, it should have a reliable supply of sterile insects, and released insects must be competitive with wild insects for mating. Laboratory rearing quality control issues, sterilisation quality control issues, high cost of laboratory set-up sometimes make it inconvenient to implement SIT (Fig. 4.2).

The sterile insect technique (SIT), applied as part of an area-wide IPM (AW-IPM) approach, offers considerable potential and has been used with great success against major pests of agricultural importance to establish pest-free areas (eradication), areas of low pest prevalence (suppression) or to maintain areas free of the pest through containment or prevention. Because of the increasing demand for environmental-friendly control tactics, it is anticipated that the SIT, as part of area-wide pest management approaches, will increasingly gain importance in the years/decades to come.

## 4.8 Conclusion

It would not be an exaggeration to refer to mosquitoes as the deadliest among the disease-causing vectors taking the lives of millions around the globe. The cosmopolitan distribution of mosquitoes renders them to come in direct contact with human beings, thereby making mosquito vector-borne disease a major global problem.

Moreover, high adaptability to changing environments and varied breeding grounds have increased the mosquito population worldwide. Research revealed that many species of mosquitoes, mainly *Culex* and *Anopheles*, could breed and proliferate in anaerobic water systems such as untreated wastewater containing very low DO.

Besides the problem of general water logging, which provides an ideal breeding ground for mosquitoes, another problem that emerged over the last few decades is water pollution in urban areas due to the continuous discharge of untreated wastewater and sewage into natural water bodies.

India, a populous and developing nation, is facing serious challenges regarding environmental management, conservation of natural resources, and achieving sustainable development goals. The water bodies are also not an exception in this context. The country's major rivers and lakes are facing severe pollution threats due to anthropogenic activities. In the name of urbanisation, sustainable development goals are being violated. So, there is an urgent need to properly manage wastewater, household, and industrial sewage. Sewage treatment sites should be located far away from human settlement areas since sewage is nutrient-rich, attracting mosquitoes. They should also be well-constructed and maintained. There should also be proper coordination between governments in the centre and state and local bodies to manage the adequate disposal of sewage.

Finally, it can be inferred that the pollution of water bodies is directly linked to the proliferation of mosquito vectors and outbreaks of mosquito vector-borne diseases. Hence, proper planning, management, and mass awareness regarding sewage and industrial waste disposal are essential to a healthy society besides achieving sustainable development goals.

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# Keratinase Role in Management of Poultry Waste

# 5

Manish Soni, Anjali Soni, Chinmay M. Joshi, Sunil Chhimpa, and Jayprakash Yadav

## Abstract

The poultry industry is one of the significant driving sectors in the food industry. On one hand, the enormous growth of this industry has boosted food safety. Still, on the other side, it also generates massive amounts of waste during various stages of food processing. Feathers, viscera, bones, and dead on arrival are some of the solid wastes which are generated. The poultry industry's most abundant wastes include feathers with approximately 90% protein content, mainly keratin protein. Enzyme technology has been one of the solutions for converting these wastes into valuable products, for example, amino acids, peptides, and other bioactive compounds having a physiological role. For this bioconversion, a keratinase enzyme is of utmost importance. Different microbes, bacteria, and fungi can degrade the feathers by secreting keratinase enzyme. This chapter gives an overview of poultry waste management through enzyme keratinase, its structure, different sources of the enzyme, production methods, and the role of the keratinase enzyme in bioconverting poultry waste into valuable products.

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M. Soni (✉)

Department of Biotechnology, School of Engineering and Technology, Jaipur National University, Jaipur, Rajasthan, India

A. Soni

B-447 Mahesh Nagar, Jaipur, India

C. M. Joshi

CUSILS, C. U. Shah University, Wadhwan City, Surendranagar, Gujarat, India

S. Chhimpa

Center for Converging Technologies, University of Rajasthan, Jaipur, Rajasthan, India

J. Yadav

Bioprocess Engineering Laboratory, Department of Biotechnology, Meerut Institute of Engineering and Technology, Meerut, Uttar Pradesh, India

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**Keywords**Chicken feathers · Keratinase · Keratin · Microorganism · Protease

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## 5.1 Introduction

The poultry industry is one of the significant and diverse elements of the food sector. The rise in products of the poultry industry is rapid due to population explosion, change in dietary habits, and lifestyle of people. The products of the poultry industry, such as meat, chicken, and eggs, are one of the primary protein sources in most people's diets. With the increasing amount of poultry products, a tremendous amount of waste is also generated. These wastes mainly include keratinous substances like horns, wool, feathers, pig bristles, etc. (Qiu et al. 2020). Li et al. 2020 have reported that more than 4.7 million tons of waste from chicken feathers alone were generated in 2019 from the poultry sector. These chicken feathers have lower risks for animals, the public, and the environment as they have been categorized under Class 3 animal by-products (Verma et al. 2017). The dry matter of feathers is 90% protein by mass (Ben Hamad Bouhamed and Kechaou 2017). Moreover, as a rich protein source, these waste products are potential sources for various valuable products, such as feed, fertilizers, antibacterial and antioxidant agents, and cosmetics (Callegaro et al. 2018; Gunasekaran et al. 2015; Lasekan et al. 2013). Cysteine, a sulphur amino acid, is the primary amino acid of these keratinous waste products.

Keratin protein being insoluble in water, it's difficult to dissolve and extract it from these waste products. So, cheap and eco-friendly methods and strategies must be designed to recover the keratin protein economically. Several physical, chemical, and biological extraction strategies today employ elevated temperatures resulting in the degradation of heat-labile amino acids and loss of nutritional value (Shavandi et al. 2017; Martinez et al. 2020). So, the alternative approach is the microbial keratinase enzyme catalyses keratin's biodegradation. Microbes are an eco-friendly approach for managing poultry waste products by converting these into valuable products for different use (Rai et al. 2020).

Keratinase are proteases belonging to the serine or metalloprotease family and can degrade keratin-rich proteins (Gupta and Ramnani 2006; Sahni et al. 2015). Feather proteins are processed into more digestible feather meal by traditional hydrothermal feather degradation. This process sustains the loss of essential amino acids and includes non-nutritive amino acids such as lanthionine and lysinoalanine. Therefore, biotechnological techniques and different microorganisms having keratinolytic enzyme activity are implemented to increase the nutrition content and value of poultry feathers to be used as feed supplements (Brandelli 2008; Gupta and Ramnani 2006; Onifade et al. 1998). Keratinase enzyme is capable of hydrolysing and transforming the keratin, applicable as animal feed (Onifade et al. 1998), and they can be used to produce nitrogen fertilizers as well (Ichida et al. 2001). Keratinases also find applications for producing biopeptides from keratin-rich

substrates and are used as antioxidants (Fakhfakh-Zouari et al. 2010). Another major biomedical application of the keratinase enzyme is its use in the degradation of prions (Yoshioka et al. 2007; Langeveld et al. 2003).

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## 5.2 By-Products of the Poultry Industry

Vast amounts of insoluble, recalcitrant, and non-degradable structural proteins like collagen, keratin, and elastin are the major animal waste products of the meat industry. Such by-products are also a rich source of protein that can be extracted and hydrolysed for use as feed or functional ingredients.

### 5.2.1 Feathers

A serious environmental problem is caused due to production of huge amounts of feather waste from the poultry processing industry. Keratin-rich feathers are used to produce feather meals, they can also be used for fertilizer and decorative purposes (Jayathilakan et al. 2012). The use of microbial enzymes in converting feather waste into usable form has been investigated for years (Gupta et al. 2013; Korniłłowicz-Kowalska and Bohacz 2011). Feather by-products are also a source of biohydrogen gas (Balint et al. 2005) and can be used to produce methane (Dudynski et al. 2012; Ichida et al. 2001).

### 5.2.2 Manure and Litter

Poultry processing plant waste is mainly found in the form of litter and manure which can be used as a surface of land or feed (Shih 1993; Simpson 1991). Also, such poultry manure is a promising source of methane production by certain anaerobic microorganisms (Salminen and Rintala 2002). Biogas is a major product of anaerobic digestion because it is a combustible fuel used to produce electricity or for heating or drying purposes (Verma et al. 2018).

### 5.2.3 Waste-Containing Collagen

Collagen is found in all animals and constitutes approximately 25% of the total protein content of skin and bones (Mayne and Brewton 1993; van der Rest and Garrone 1991). Collagen-rich by-products are denatured using heat treatment, and gelatin can be extracted from it. Waste hydrolysates of collagen have been studied for their antihypertensive properties. (Gomez-Guillen et al. 2011)., chicken bones are also considered a source of collagen to produce hydrolysates with unusual bioactivity (Huang and Liu 2010; Cheng et al. 2009). The bioactive peptides are

promising components for animal feed as they can employ physiological effects in-vivo.

#### 5.2.4 Miscellaneous By-Products

Blood, head, and gizzard are known to produce meal, while intestines, feet, and skin are considered a source of fat (Sams 2001). Keratinolytic microorganisms can degrade keratin-containing beaks or nails (Riffel and Brandelli 2006). It can be utilized with blood or viscera for animal feed production (Sams 2001). Generally, anticoagulant-treated blood is dried and concentrated to form a blood meal, a rich source of sulphur-containing amino acids like cysteine and methionine and basic amino acids like lysine and arginine (Marquez et al. 2005). Also, hatchery by-products contain unhatched eggs, discarded chicken, infertile eggs, and eggshells, which can be used as animal feed. These meals are a rich source of calcium, restricting their use in feed to up to 5% (Jayathilakan et al. 2012).

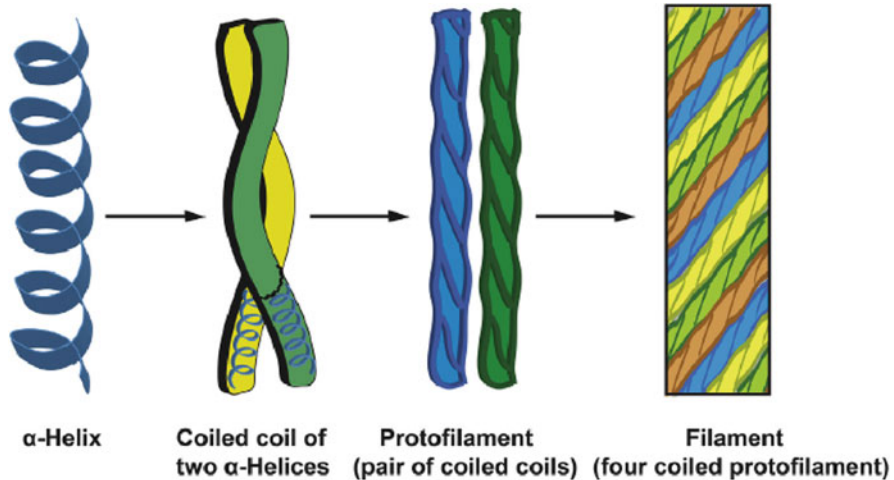
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### 5.3 Keratin

The principal protein constituent of feathers is keratin which makes up to 90% of its dry mass. It follows cellulose and chitin in abundance and is an insoluble structural protein (Lange et al. 2016). The primary source of keratin protein includes epithelial cells of vertebrates, hair, nail, and feathers of birds (Adelere and Lateef 2019). A compactly packed structure (alpha and beta sheets) makes the protein highly stable due to the non-accessibility of different proteolytic enzymes (Gupta and Ramnani 2006). Other weak interactions, disulfide bonds, hydrogen bonds, and hydrophobic interactions, provide a high degree of cross-linking to the keratin structure. Based on their secondary structure, keratins can be categorized into  $\alpha$ -keratin and  $\beta$ -keratin.  $\alpha$ -helical-coiled protein chains form the structure of  $\alpha$ -keratin, and  $\beta$ -sheets stack up to form the structure of  $\beta$ -keratin. (Qiu et al. 2020; McKittrick et al. 2012; Meyers et al. 2008).

#### 5.3.1 $\alpha$ -Keratin

The monomeric structure of  $\alpha$ -keratin consists of three regions - an N-terminal region (head region), a central rod region, and a C-terminal region (tail region) (Bragulla and Homberger 2009). The twisting of helices around themselves results in a quaternary structure of a coiled-coil dimer. The dimers combine into protofilaments and filaments (Fig. 5.1) (Hassan et al. 2020). Hair, nails, wool, horns, claws, and hooves are the significant source of  $\alpha$ -Keratin.



**Fig. 5.1** Structure of  $\alpha$ -Keratin (Hassan et al. 2020)

### 5.3.2 $\beta$ -Keratin

Chicken feathers are the major source of  $\beta$ -keratin. The enzyme keratinase is more effective in degrading  $\beta$ -Keratin than  $\alpha$ -keratin due to fewer disulfide bonds and a more porous structure (Gupta and Ramnani 2006).

Both forms of keratin-  $\alpha$ -keratin and  $\beta$ -keratin are present in different keratinous materials like- feathers, hair, bristles, wool, etc. There is a preferential expression of  $\alpha$ -keratin and  $\beta$ -keratin in feathers. (Ng et al. 2014).  $\alpha$ -keratin is the major component of feathers making up 41–67%, and the remaining 33–38% is  $\beta$ -keratin (Fraser and Parry 2008). Apart from feathers, other keratin-containing materials like hair, bristle, and wool preferentially contain  $\alpha$ -keratins which are 50–60%, and the rest is matrix proteins and  $\beta$ -keratins (Daroit and Brandelli 2014).

### 5.3.3 Hard Keratin and Soft Keratin

Based on the content of sulphur, keratin can be categorized into hard keratin and soft keratin. Hard keratin has more sulphur content compared to soft keratin. Feathers, horns, hair, and nails contain hard keratin, whereas skin and callus contain majorly soft keratin (Barone et al. 2005; Fraser and Parry 2008). Hard keratin is stiffer than soft keratin due to its high sulphur content, which can form more disulphide bonds. Moreover, soft keratin is more liable to hydrolysis by acid and alkali (Ng et al. 2014; Schrooyen et al. 2001).

## 5.4 Keratinase

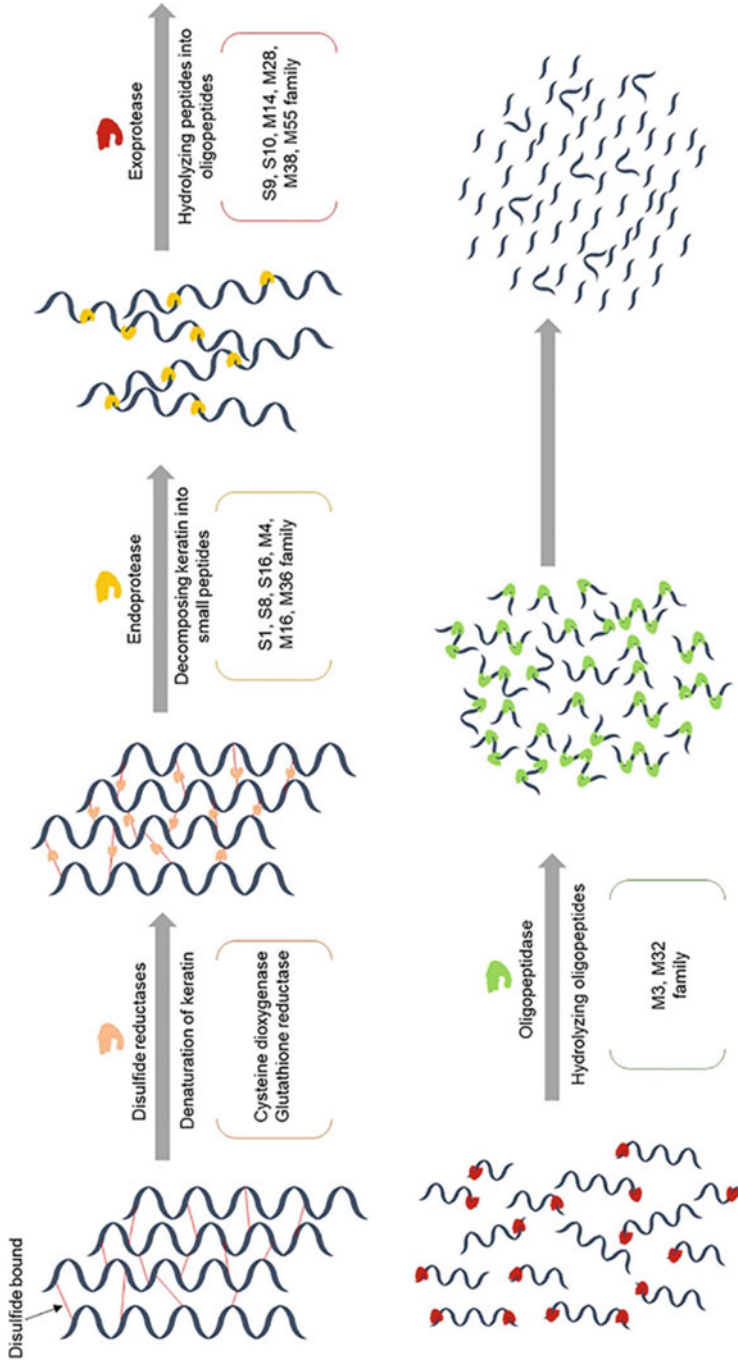
Enzymes are the catalysts for biological macromolecules produced by diverse species of different microorganisms, including bacteria, actinomycetes, fungi, and yeasts. Proteases are the enzymes that can destabilize the native protein structure. Keratinase (EC3.4.21/24/99.11) is a protease type with keratinolytic enzymatic activity (Lange et al. 2016). The enzyme keratinases identified till date belongs to serine family and metalloproteases family (Gupta and Ramnani 2006; Sahni et al. 2015). The mechanism of keratin degradation by enzyme keratinase is based on the reduction of disulphide bonds which occurs via a two-step reaction, namely sulfitolysis (SeS bond breakage) and proteolysis (Gupta and Ramnani 2006; Wang et al. 2015; Lange et al. 2016; Peng et al. 2019; Qiu et al. 2020). The steps involved in enzymatic keratin degradation are illustrated in Fig. 5.2.

Different keratin-rich substrates have been used to assay keratinolytic enzyme activity, including feather, pig bristles, wool cuticles, etc. Also, some coloured modified keratin derivatives are used, like azo-keratin and keratin azure (Gonzalo et al. 2020; Habbeche et al. 2014). Keratinase has been classified into different protease families based on their cleave pattern. They can be endo-protease, exo-protease, or oligo-peptidase. The detailed classification of keratinase into different protease families is illustrated in Table 5.1 (Qiu et al. 2020). Recently, novel keratinolytic enzymes have been identified belonging to the protease family M36 (Qiu et al. 2022). For this, they have used the bioinformatics tool Conserved Unique Peptide Patterns (CUPP).

## 5.5 Microbial Diversity of Keratinase

Several microorganisms can degrade keratin waste, including feathers, by secreting keratinases. Keratinases are a group of proteolytic enzymes (Tamreihao et al. 2019). A wide variety of microorganisms have been isolated and characterized from different keratin-rich environments and were found functional against the degradation of keratin-containing wastes (Chaturvedi et al. 2014). Bacteria, actinomycetes, and fungi are the main categories which include these keratin-degrading microorganisms (Calin et al. 2017; Bohacz and Kornilowicz-Kowalska 2019). Keratinases secreted by these microorganisms are involved in cleaving the disulphide bonds in keratin proteins. The keratinase enzyme's efficiency increases when combined with other proteases. These microorganisms secreting keratinase enzymes inhabit diverse environments- soil, water, and air rich in keratin sources (Qiu et al. 2020).

Among microorganism bacterial species such as *Bacillus licheniformis*, *B. subtilis*, and *Stenotrophomonas maltophilia* have the most potential to degrade keratin-rich waste material because of the suitable environmental conditions. *Bacillus sp.* is the most potent keratin-degrading sp. of all bacterial species. *Bacillus amyloliquefaciens* S13 produces two extracellular keratinolytic proteases having a molecular weight of 47 and 28 kDa, respectively (Hamiche et al. 2019; Kashyap



**Fig. 5.2** Steps involved in enzymatic keratin degradation (Qiu et al. 2020)



**Table 5.1** Protease family, characteristic features of Keratinase Enzyme (Qiu et al. 2020)

Family	Characteristics	Example	References
S1	Serine protease, Endoprotease, alkaline pH (9–12.5) Temp. 50–60° C	<i>Paenarthrobacter nicotinovorans</i>	Sone et al. (2015)
		<i>Nocardioopsis sp.</i> TOA-1;	Mitsuiki et al. (2004)
		<i>Streptomyces fradiae</i> var. k11	Li et al. (2007)
S8	Serine protease, Endoprotease, alkaline pH (7.5–11) Temp. 37–80° C	<i>Parengyodontium album</i>	Ebeling et al. (1974) and Jany et al. (1986)
		<i>Stenotrophomonas maltophilia</i>	Fang et al. (2014)
		<i>Fervidobacterium pennivorans</i>	Friedrich and Antranikian (1996) and Kim et al. (2004)
		<i>Thermoactinomyces sp.</i>	Wang et al. (2015)
		<i>Bacillus licheniformis</i>	Ramnani and Gupta (2004)
		<i>Bacillus subtilis</i>	Gupta and Singh (2013)
		<i>Bacillus amyloliquefaciens</i>	Yang et al. (2016)
		<i>Bacillus pumilus</i>	Fellahi et al. (2016)
		<i>Bacillus cereus</i>	Ghosh et al. (2009)
		<i>Bacillus halodurans</i>	Shrinivas and Naik (2011)
		<i>B. subtilis</i> dps3 (MW255302), <i>B. cereus</i> wps1 (MW255303) and <i>B. licheniformis</i> dcs1 (MW255304)	Liaqat et al. (2022)
		<i>Meiothermus taiwanensis</i>	Wu et al. (2017)
		<i>Stenotrophomonas maltophilia</i>	Jankiewicz et al. (2016)
		<i>Thermoactinomyces sp.</i> YT06	Wang et al. (2017)
		<i>Trichophyton benhamiae</i>	Jousson et al. (2004) and Solanki et al. 2019
		<i>Onygena corvina</i>	Huang et al. (2015)
		<i>Microsporium canis</i>	Descamps et al. (2002)
<i>Aspergillus niger</i>	Chen et al. (2015)		
		<i>Trichophyton mentagrophytes</i>	Yohko et al. (2014) and Abbas et al. 2022
S16	Serine protease, Endoprotease,	<i>Fervidobacterium islandicum</i>	Kang et al. (2020)
M4	Metallo protease, pH 9	<i>Pseudomonas aeruginosa</i>	Sharma and Gupta (2010)
		<i>Geobacillus stearothermophilus</i>	Gegeckas et al. (2015)
M16	Metallo protease, Endoprotease	<i>Fervidobacterium islandicum</i>	Kang et al. (2020)
M36	Metallo protease, Endoprotease	<i>Fusarium oxysporum</i>	Chaya et al. (2014)
		<i>Microsporium canis</i>	Brouta et al. (2002)
		<i>Onygena corvina</i>	Huang et al. (2015)

(continued)

**Table 5.1** (continued)

Family	Characteristics	Example	References
S9	Serine protease, exoprotease, pH 7–9	<i>Trichophyton rubrum</i>	Monod et al. (2005)
S10	Serine protease, exoprotease	<i>Trichophyton rubrum</i>	Zaugg et al. (2008)
M14	Metallo protease, exoprotease	<i>Trichophyton rubrum</i>	Zaugg et al. (2008)
M28	Metallo protease, exoprotease	<i>Trichophyton rubrum</i>	Monod et al. (2005)
		<i>Onygena corvine</i>	Huang et al. (2015)
		<i>Streptomyces fradiae</i>	Wu et al. (2010)
M38	Metallo protease, exoprotease	<i>Fervidobacterium islandicum</i>	Kang et al. (2020)
M3	Zinc dependent metallopeptidase	<i>Onygena corvine</i>	Huang et al. (2015)
M32	Zinc dependent metallopeptidase	<i>Fervidobacterium islandicum</i>	Lee et al. (2015)

et al. 2019b). *Bacillus cereus* strain isolated from the halophilic environment had keratinase enzyme activity (Arokiyaraj et al. 2019; Kashyap et al. 2019a). Strains of *Bacillus sp.* screened from the marine environment also have keratinase activity (Herzog et al. 2016).

Some other *sp.* of bacteria such as *Micrococcus*, *Pseudomonas*, *Paenibacillus*, *Serratia*, *Vibrio*, and *Stenotrophomonas* have been reported for their keratinase production when they are grown on keratin as a substrate and feather of birds (Laba et al. 2015; Chaturvedi et al. 2014; Paul et al. 2013; Khardenavis et al. 2009; Grazziotin et al. 2007; Fang et al. 2013).

Fungi and actinomycetes also have the ability to degrade keratin-rich waste products like feathers by secreting keratinase enzymes. The presence of hyphae facilitates Keratin degradation in fungi (Korniłłowicz-Kowalska and Bohacz 2011; Tridico et al. 2014). A heat-stable keratinase enzyme isolated from *Meiothermus taiwanensis* strain WR-220 can destabilize the highly recalcitrant sulphur bonded keratin protein (Wu et al. 2017). *Fusarium sp.* strain was also reported for its proficient keratin denaturation property (Calin et al. 2017). A list showing the microbial diversity of keratinase enzymes is illustrated in Table 5.2.

## 5.6 Role of Keratinase Enzyme in Waste Management and Production of Valuable Products

With the development in enzyme technology there is advancement in less energy-consuming techniques for the production of useful products using waste products of the poultry industry (Darah et al. 2013; Onifade et al. 1998). Proteases are one of the important enzymes for conversion of poultry waste into valuable products. Some of the applications of keratinase enzymes have been illustrated in Fig. 5.3.

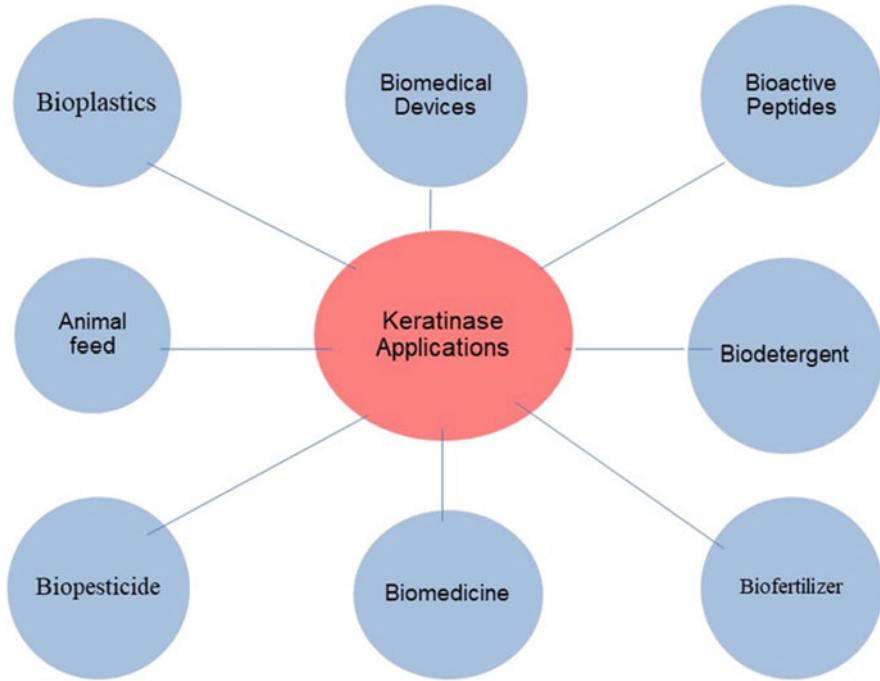
**Table 5.2** Microbial diversity of keratinase (Lange et al. 2016; Li 2019)

Species of microorganism	References
<i>Bacteria</i>	
<i>Bacillus amyloliquefaciens</i>	Hamiche et al. (2019)
<i>Bacillus cereus</i>	Arokiyaraj et al. (2019)
<i>Bacillus thuringiensis</i>	Sahni et al. (2015)
<i>Bacillus aerius NSMk2</i>	Bhari et al. (2018)
<i>Bacillus licheniformis</i>	Abdel-Fattah et al. (2018)
<i>Bacillus subtilis</i>	Liu et al. (2017)
<i>Bacillus pumilus</i>	Ramakrishna Reddy et al. (2017)
<i>Brevibacillus parabrevis</i>	Zhang et al. (2016)
<i>Chryseobacterium sediminis</i>	Kshetri et al. (2019)
<i>Stenotrophomonas sp.</i>	Herzog et al. (2016)
<i>Micrococcus sp.</i>	Laba et al. (2015)
<i>Serratia sp.</i>	Khardenavis et al. (2009)
<i>Fungi</i>	
<i>Trichoderma harzianum</i>	Bagewadi et al. (2018)
<i>Meiothermus taiwanensis</i>	Wu et al. (2017)
<i>Fusarium sp.</i>	Calin et al. (2017)
<i>Amycolatopsis</i>	Tamreihao et al. (2017)
<i>Streptomyces albidoflavus</i>	Bressollier et al. (1999)
<i>Trichophyton rubrum</i>	Sharma et al. (2012)
<i>Chrysosporium articulatum</i>	Bohacz (2016)
<i>Aphanoascus fulvescens</i>	Bohacz (2016)

### 5.6.1 Animal Feed

Keratin-rich feathers are a rich source of essential amino acids which form a part of animal feed. To preserve their nutritional value, these wastes need to be hydrolysed using keratinase enzymes. In order to manufacture hydrolysed feather keratin for feed formulation, keratinase from different sources have been investigated (Brandelli et al. 2010; Gupta and Ramnani 2006). *Bacillus spp.* is a significant source of keratinase enzyme. Commercial Versazyme<sup>®</sup> product, based on subtilisin-like keratinase obtained from *B. licheniformis*, a feed additive, was effectively tested (Odetallah et al. 2005).

Kim and Patterson 2000 have compared enzymatic and sodium hydroxide treatments for the processing of feathers and shown that alkaline treatment was a quick method for the separation of feathers from carcasses and feather-digesting enzymes improved the feather's nutritional quality.



**Fig. 5.3** Applications of Keratinase

### 5.6.2 Bio-Fertilizers

Bioconversion of feathers in the presence of keratinase enzyme has been used to prepare nitrogen fertilizers (Korniłowicz-Kowalska and Bohacz 2011; Vasileva-Tonkova et al. 2009). These form an inexpensive source of proteins which further improves and promotes the growth of roots and shoots in plants (Gurav and Jadhav 2013). Degradation of keratin by *Paecilomyces marquandii* fungal keratinase has resulted in products which are potentially useful for foliar fertilization. It has been also shown that enzymatic preparations result in higher amounts of amino acids as compared to microorganisms for keratin hydrolysis because the microbial cells consume some part of the solubilized products during development (Vesela and Friedrich 2009). Feather hydrolysates from *A. niger*, *B. cerus*, and *Amycolatopsis sp.* are used as biofertilizers due to presence of keratinolytic enzymatic activities (Adetunji et al. 2012; Tamreihao et al. 2017; Choinska-Pulit et al. 2019).

### 5.6.3 Bioactive Peptides

Hydrolysis of keratin-rich material is used to generate bioactive peptides. These bioactive peptides find major applications in the pharmaceutical and cosmetic

industry (Jin et al. 2018; Yeo et al. 2018). The hydrolysates from raw chicken feathers have shown inhibitory activities of antioxidants, angiotensin-converting enzyme (ACE), and dipeptidyl peptidase-IV (DPP-IV), indicating that these feather hydrolysates can be a source of bioactive peptides (Fontoura et al. 2014).

#### 5.6.4 Biomedical Devices

The products transformed from keratin obtained from human sources have been used in designing biomedical devices. For example, the keratin from human hairs as a scaffold is used in bone regeneration (Dias et al. 2010; de Guzman et al. 2013). Products like hydrogels have been obtained from keratin which has wound-healing properties (Wang et al. 2017).

#### 5.6.5 Biodetergents

The broad specificity of keratinase enzymes makes them an attractive candidate for biodetergent. Different microbial species have been exploited to formulate biodetergent. Alkaline keratinase from *Paenibacillus woosongensis* TKB2 has been used to formulate a biodetergent which was highly efficient in removing blood and egg yolk stains (Paul et al. 2014). Other potential microbial species used in biodetergent production are *Paecilomyces lilacinus* (Cavello et al. 2013), *Gibberella intermedia* (Zhang et al. 2016), and *B. pumilus* (Gong et al. 2015).

#### 5.6.6 Bioremediation and Biopesticide

Keratinolytic activity of enzyme is highly potent is reduction of toxicity from wastewater effluents from the leather industry (Qiu et al. 2020). It has been shown that nematodes and other entomopathogenic microorganisms can be suppressed by the action of keratinase enzymes (Brandelli et al. 2010; Gupta et al. 2013; Verma et al. 2017) and hence can be used as biological pesticide. A keratinase secreted by *Bacillus sp.* Has been reported to kill *Meloidogyne incognita* (a root-knot nematode) (Yue et al. 2011).

#### 5.6.7 Biomedicine

Keratinase enzyme also finds application in the biomedical field in treating corn, calluses, acne, scars, and other skin-related diseases (Gupta et al. 2013). For example, Pure100 keratinase is used in treating calluses, acne, and decontamination of prions (Gupta et al. 2013). Their use in the cosmetic industry is emerging at an accelerated rate as supplements for different purposes to beautify the skin (Anandharaj et al. 2016; Gupta et al. 2013).

Prions are infectious protein particles which cause contagious and fatal brain diseases (Saunders et al. 2008). In this disease, prion protein is misfolded (PrPSc) leading to change in secondary structure conformation from alpha helix to beta-sheets. Studies have shown that Keratinase enzyme can cleave  $\beta$ -plated protein found in patients suffering from prion disease. KerA from *B. licheniformis* PWD-1 was the first keratinase isolated and discovered to degrade and hydrolyse PrPSc (Langeveld et al. 2003). Other microbial species capable of degrading prion protein-PrPSc includes *Streptomyces sp.* (Tsiroulnikov et al. 2004), *Nocardiosis sp.* (Mitsuiki et al. 2004), *Thermoanaerobacter*, *Thermosipho*, and *Thermococcus sp.* (Suzuki et al. 2006).

### 5.6.8 Bioplastics

Bioplastics have been developed using chicken feather wastes as an alternative to conventional plastics which are based on petroleum products. Due to the presence of keratin, these bioplastics are resistant to high temperature, highly elastic, biodegradable, and biocompatible (Kota et al. 2014).

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## 5.7 Future Scope

Although substantial research is being carried out in the field of enzyme biology and keratinase enzyme in particular, a number of questions still need to be answered to fully understand the enzyme as a whole. For example, the relationship between the structure and the functional role of enzymes is still poorly understood. Substrate specificity, kinetics of enzyme, and biological diversity are some other areas to be explored in future. These studies will help in understanding the bioconversion mechanism and newer areas of applications of the enzyme keratinase.

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## 5.8 Conclusions

Industrially important enzymes have been under the scanner for mass production of the protein. Microbial keratinases having the capability to degrade recalcitrant keratin have been exploited for different applications in various industrial sectors like biomedicine, animal feed, biodetergent, biofertilizers. Microbial keratinase is more eco-friendly as compared to other chemical methods employed for enzyme production. Hence, they have been explored a lot in managing poultry waste and producing different valuable products from them.

**Conflict of Interest** The authors have no conflict of interest.

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# Biomedical Waste: Impact on Environment and Its Management in Health Care Facilities

# 6

Gyanendra Kumar Sonkar, Sangeeta Singh,  
and Satyendra Kumar Sonkar

## Abstract

Waste of any origin, if not properly disposed, possess a significant threat to the environment. Biomedical waste is a potential health hazard generated from institutions and laboratories providing health care facilities which includes all sorts of pathological, pharmacological, genotoxic, chemical, and radioactive wastes. About 20% of waste generated during patient care is hazardous and carries various health risks to hospital staff, patients, attendants, and the general population. Proper segregation and disposal of biomedical waste is the need of the hour as it will prevent contamination of groundwater sources that affect the health of humans and animals. Proper packaging and labelling of waste prevent the spread of infection through humans and animals. Biomedical waste is the source of water contamination and, if not rendered harmless before it is buried in land or disposed of in the water. Biomedical waste contaminates air if not segregated or incinerated properly, resulting in highly hazardous airborne particles of contagious diseases. The diagnostic laboratories using radioactive substances are potential pollutants of landfills and the atmosphere. The spread of air pollutants over huge areas of inhabited land has the potential to trigger several illnesses. Hence, there should be the management of biomedical waste at each level (i.e., places of its generation, collection, storage, transportation, treatment, and disposal). The stakeholders, including health care sector, state pollution control

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G. K. Sonkar (✉)

Department of Biochemistry, King George's Medical University, Lucknow, Uttar Pradesh, India

S. Singh

Department of Biosciences, Integral University, Lucknow, Uttar Pradesh, India

S. K. Sonkar

Department of Medicine, King George's Medical University, Lucknow, Uttar Pradesh, India

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board, and the municipal bodies, should work together to make the place safe for living with a neat and clean environment.

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**Keywords**

Biomedical wastes · Hazardous · Infection · Disposable · Incineration · Autoclave · Hydroclaving

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## **6.1 Introduction**

Human body is in communion with five basic elements of nature. With the passage of time since evolution, there has been an increase in human population, leading to a decline in natural resources. No one is really worried about the future generation as no one fears the laws these days. With global industrialization, the biggest problem that came on the way was pollution. Many flora and fauna are on the verge of extinction, and even humans are not spared, and what prevails is medically affected. In November 1992, under the umbrella of UNESCO, various scientific personnel came together to address pollution.

### **6.1.1 Definition of Biomedical Waste**

The medical documents related to waste management were first issued by World Health Organization (WHO) in 1996. As the medical sciences and its facilities have tremendously increased in the last few decades, it has led to a several-fold increase in biomedical waste (Sheth et al. 2006). The act of diagnosis, treatment, and immunization of human beings leads to generation of solid or liquid waste material. Wastes generated also add up to biomedical wastes. If these waste materials are not managed properly, they can be hazardous to health and the environment. All the hospital staffs are at risk to get various infections and injuries from these infectious materials. Diseases like hepatitis B, AIDS, etc., are on the increase, and these conditions have become critical public issues that need to be addressed. The technologies we are using for disinfecting these medical wastes are also adding toxic emissions polluting the environment. Hence to avoid these hazards, discriminate waste management system should be implemented in hospital infrastructure. Biomedical waste management (BWM) is a process that helps to ensure proper hospital hygiene and safety of health care workers and communities and the environment. BWM is concerned about planning and procurement, staff training and behaviour, proper use of tools, machines, and pharmaceuticals, proper methods applied for segregation, reduction in volume, treatment, and disposal of biomedical waste.

In India, the implementation of disposal of hospital wastes without segregation is still a problem. At many places, it is very common to find huge dumps of biomedical waste. The municipal workers or the rag pickers who are working at these sites in order to separate syringes, bottles, disposables, etc. for the sake of reselling them,

incur the risk of getting infected by hepatitis B virus (HBV), hepatitis C virus (HCV), and human immunodeficiency virus (HIV) (Sulmer 1989).

According to Chartier et al. (2014), there are five principles that are widely used by several countries in their legislative and political systems.

1. The “polluter pays” principle is a policy implying to the basic rule that one should take responsibility of their own waste. It means that it’s the companies and individuals responsible, legally and financially, for the safe and efficient disposal of waste generated by them.
2. The “precautionary” principle states that even when there is no clear evidence of harm and or risk from any human activity, still significant and protective measures must be taken accordingly to minimize environment damage from biomedical wastes.
3. The “duty of care” principle creates a connection between the individual handling and managing wastes, thus creating an ethical responsibility for the person. The most efficient way to keep this principle operating is to include working environments with people with proper education and knowledge in this area of business.
4. The “proximity” principle recommends that waste be treated and disposed of in the nearest possible location to its source. This way risks are minimized for the health category, and logistic costs for waste managing are diminished.
5. The “prior informed consent principle” the principle is mentioned in various international treaties, and it is designed to protect the environment and public health from several kinds of hazardous wastes.

### 6.1.2 Generation of Biomedical Waste

The biomedical wastes (Fig. 6.1) are usually generated from hospitals, various health clinics, laboratories and research facility centres, veterinarian clinics, offices, banned drugs, as well as during a disease outbreak (Hegde et al. 2007). Rapidly increased medical waste brings big challenges to their treatment and disposal. For example, recent COVID-19 outbreak, which has been characterized as a pandemic, causes the increase in generation of medical waste during the care of COVID-19 patients and the situation may be much more serious as the outbreak spreads. If medical waste is not properly managed, it will pose a great threat to the environment and humans due to its toxicity and infectious nature (Cai and Du 2020).

### 6.1.3 Categories of Biomedical Waste

Out of the total biomedical wastes produced each day, approximately 15–20% is hazardously injuring humans, animals, and the environment (WHO 2018). Mixing the non-hazardous waste with hazardous waste makes the whole of the waste very infective. WHO has categorized biomedical wastes into eight types, whereas the





**Fig. 6.1** Biomedical wastes. (Source: Zafar 2020; <https://stock.adobe.com/search?k=“biomedical+waste”>)

Ministry of Environment and Forest in India (1998) has classified it into ten types (Kalpana et al. 2016) as follows:

Category 1	This includes human body parts, tissues, and other organs
Category 2	It includes several animal body parts, including tissues and bleeding parts of experimental animals used in research work or wastes generated from veterinary hospitals
Category 3	This includes wastes generated from Microbiology and Biotechnology laboratories, including research and industrial laboratories
Category 4	This constitutes waste that may cause punctures or cuts in body parts such as needles, syringes, scalpels, blades, glass, etc.
Category 5	All the medicines that have expired, contaminated, and discarded, including cytotoxic drugs, are included in this group
Category 6	This group comprises solid waste, i.e., those items contaminated with blood and body fluids
Category 7	This includes sharp, less solid wastes such as tabbing, catheters, and intravenous sets used for medical purpose
Category 8	This category includes liquid waste generated from laboratory and washing, cleaning, housekeeping, and disinfecting activities
Category 9	It includes ash generated from incineration of any bio-medical waste
Category 10	This group includes chemical and biological wastes.

## 6.2 Biomedical Waste Management Strategies

The management of biomedical waste is described as a multifaceted process that typically involves effective legislation, training, minimization, proper handling, segregation, storage, transportation, treatment, and safe disposal (Rao et al. 2004; WHO 2007).

### 6.2.1 Biomedical Waste Segregation and Storage

Segregation of biomedical waste is an important component of any waste management scheme (Fig. 6.2). It is an extensive challenge for the government and the health sector (Riyaz et al. 2010). It is still in its infancy all over the world (Arvind and Girish 2010). Proper management ensures that infectious waste is handled in accordance with established and acceptable procedures from the time of generation through treatment of the waste and its ultimate disposal (Sawalem et al. 2009). Proper container or color-coded bags must be used for each category of waste generated (Table 6.1) which will avoid environmental contamination and human health infection and help in segregating biomedical pollutants from non-pollutants. This practice reduces the total treatment cost, the impact of waste in the community, and the risk of infecting workers. Waste should be segregated into different



**Fig. 6.2** Biomedical waste segregation and storage. (Source: BMW Cell, KGMU, Lucknow)

**Table 6.1** Color-coding for biomedical waste segregation (Source: Biomedical Waste Management Rules, CPCB 2016)

S. No.	Category	Items	Container/ disposable bags color
1.	Non-Plastic infectious waste	Body parts of humans and animals and other items used in day to day procedures such as cotton dressings, plaster casts and other materials contaminated with blood	Yellow
2.	Plastic Infectious waste	Glucose bottles, hub removed syringe, catheters, intravenous sets, gloves, etc. which are disposable in nature	Red
3.	Sharp waste	Needles, scalpels, blade, etc.	Red (puncture proof)
4.	Glass Waste	Bottle, ampoules, slides, tubes, etc. made from glass	White
5.	Liquid waste	Wastes generated from washing, cleaning and disinfecting activities	Blue
6.	General waste	Papers, wrappers, Fruits and vegetables peel and leftover food and edibles, etc.	Black

categories at the site of generation (Park 1997; Rao et al. 2004). Segregation of biomedical wastes at source is a key, and it will help hospital authorities to save money on waste disposal (Vorapong 2009).

After segregation, biomedical wastes should have safe and secured storage. All the containers mentioned above should be spill-proof and strong enough to hold the designed volume and weight of wastes without getting damage and preferably having a cover lid that can be operated by a foot (Mastorakis et al. 2011). The biomedical wastes should not be stored beyond 48 h onsite, and hence they should be collected on a regular basis every day. It should be further seen that this storage area should not be accessible to unauthorized people such as patients or visitors (WHO 2005a, b). Large hospitals and institutions having different departments, laboratories and operating theatres (OTs), wards, etc., should have a centralized collection/storage room where the wastes can be collected before sending it to treatment or disposal site.

## 6.2.2 Biomedical Waste Handling and Transportation

Such wastes should be handled very carefully while it is being collected, stored, or during transportation. Time of collection should be well documented in duty charts and a copy of the same should be given to concerned waste collectors and supervisors. The waste bags should always be closed during transportation, with no leakage and no dragging of bags on the floor (Chandra 1999). The person collecting the waste should come in minimum contact to avoid infection. It should be done in the utmost safe manner while being transported outside the hospital



**Fig. 6.3** Proper collection and handling of biomedical wastes. (Source: BMWM Cell, KGMU, Lucknow)

premises to the site of disposal. The vehicle used for transportation within and outside the hospital premises should be covered and have proper door closure and avoid leakage. The reusable containers used during such transport should not have sharp edges or corners to easily be washed and disinfected (Fig. 6.3) (Pruss et al. 1999; Chandorkar and Nagoba 2004).

### 6.2.3 Treatment and Disposal of Biomedical Waste

In developing countries, the unsanitary disposal of waste has put millions of lives at risk because people often visit dumping sites scavenging for goods. Biomedical wastes are disposed of on the bare ground, discarded into water bodies, or thrown away casually, which raises health issues in the surrounding habitat. As in some countries like Pakistan, biomedical wastes are simply thrown out on the ground, mixed with ordinary waste, or buried without any appropriate measure (Mustafa and Anjum 2009). In India, the effective waste disposal system still lacks in many small hospitals and nursing homes except in a few large hospitals (Dwivedi et al. 2009). Even the Government and municipal hospitals are no better than the private nursing homes regarding their waste disposal. A large volume of infectious wastes is disposed of in burial pits located at hospital sites and in municipal landfills, both practices pose significant risks to humans, including direct contact and

contamination of surface water or groundwater (Rolando et al. 1997). Hence, before the actual disposal of biomedical waste, it should be disinfected, made environmentally non-toxic, and aesthetically acceptable. New processes and technologies are being introduced and marketed (Verma 2010; Diaz and Savage 2003; Mindrescu 2010). However, the final choice of treatment technology should be made carefully based on various factors, many of which depend on local conditions.

Broadly five methods viz. (a) chemical, (b) biological, (c) mechanical, (d) thermal, and (e) irradiation are being used in several places to treat biomedical wastes.

### 6.2.3.1 Chemical Processes

It is used for treating liquid wastes consisting of microorganisms, amount of contamination present, and biology of the microorganism (Patan and Mathur 2015). The wastes are first shredded, grinded and then mixed with chlorine dioxide, sodium hypochlorite, peracetic acid, lime solution, calcium oxide powder, and other inorganic chemicals. Anatomical wastes of humans and animals are treated with hot alkali in a stainless steel tank to disinfect them (Chartier et al. 2014).

### 6.2.3.2 Biological Processes

Using the naturally occurring aerobic and anaerobic processes, the organic substances are degraded, transformed, and stabilized into non-toxic end products (Verma et al. 2018). These fundamental processes are the basis for management strategies focusing on the biological treatment of organic waste materials. Biological degradation of waste materials is ambivalent and can lead to harmful effects if microbial activities occur under uncontrolled conditions in imbalanced systems (Bohm et al. 2011). Three changes occur during aerobic self-purification: **coagulation** of colloidal solids passing through the primary sedimentation stage; oxidation of carbon, nitrogen, and phosphorus; and nitrification. The basic requirements of any aerobic system for successful treatment of organic matter are a community of acclimatized **microorganisms**, adequate substrate (food), and a suitable environment (Scholz 2016). The organic wastes containing pathogens are destroyed using certain kinds of enzymes in the system. Digesting of such organic wastes with the help of worms (vermiculture) and composting. Deep burial is used successfully to decompose household kitchen wastes and hospital wastes such as placenta and other pathological wastes (Mathur et al. 2006). However, due care must be taken at such burial sites to restrict only authorized personnel and adequate precautions must be taken to prevent pollution and contamination of ground and surface water sources (Pruss et al. 1999) Furthermore, infectious and hazardous residues must be encapsulated with immobilizing agents prior to burial.

### 6.2.3.3 Mechanical Processes

This is done to reduce the bulk volume by more than 60%. It includes several processes such as granulating, pulverizing, shredding, grinding, mixing, agitating, and crushing of the biomedical wastes. This helps to facilitate further processes of treatment or disposal. Hence compaction and shredding are essentially the two

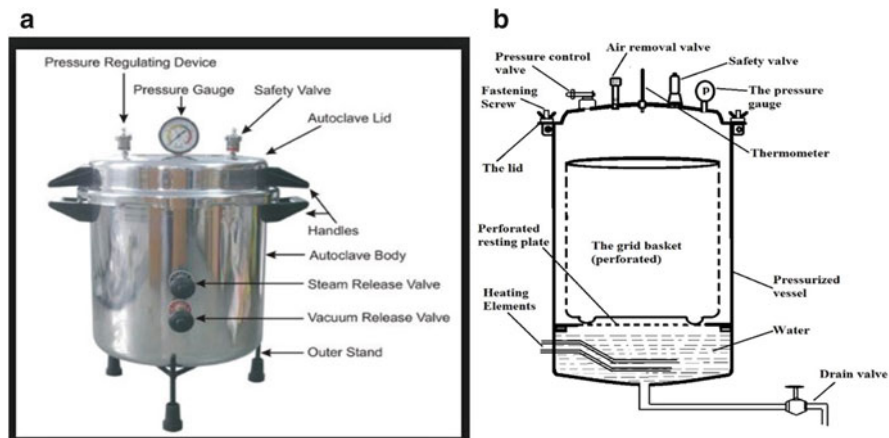
important mechanical methods. These two are not used for untreated wastes because it generates aerosol and spilling of microorganisms which can be health hazard such as tuberculosis (Acharya and Meeta 2000). Shredder's basic work is to shred sterilized/autoclaved biomedical wastes before they are disposed. It is mainly used in combination with an autoclave. This makes the wastes almost unrecognizable (Rasheed et al. 2005) and makes transportation easy. The problem with shredder is that its blade has to be regularly replaced due to its wear and tear process with preventive and breakdown maintenance every 6 months. Nowadays, electrically operated shredders are readily available. Mashing or shredding of solid biomedical waste can generate dust. If this dust becomes airborne, it can be a workplace hazard and a threat to the environment, hence, closed rooms and hood with ambient pressure are used for keeping mechanical equipment. The next equipment used is needle cutters and destroyers which can be either mechanical or electrical. Studies show that more than 20% of those, who administer injections, suffer "needle stick injuries". These are used at those locations where needles are used for blood collection or the immunization process and nursing stations and clinics. As per WHO report, 8–10 million Hepatitis-B, 2.3–4.7 million Hepatitis-C, and 80,000–160,000 HIV are estimated to occur from the reuse of syringe and needles without sterilization. The hospital staffs have plenty of chances of accidental needle stick injuries during administration of injections, drawing blood, and disposing used needles. Needles should be destroyed immediately after use since stick injury may occur at any stage after use (International Health Care Worker Safety Center 1998). These instruments help in avoiding the reuse of disposable syringes. There is an advantage of using electrical syringe cutters over mechanical ones as it can both cut and burn the needle and completely destroy it.

#### **6.2.3.4 Thermal Processes**

This method is regarded as the most revolutionary and universal method. This uses a high temperature, which leads to the destruction of microorganisms. Broadly two methods are known—(1) low heat systems (LHS) and (2) high heat systems (HHS). LHS operates at a temperature range of 93–177 °C and uses steam, hot water, or electromagnetic radiation to decontaminate the wastes. The two best known are Autoclave and Microwave. HHS usually requires very high temperatures to decontaminate the wastes. The best examples are incinerators, hydroclaving, and thermal plasma.

#### **Autoclaving**

It is simply also known as steam sterilization. It is used to sterilize or disinfect biomedical wastes before being disposed-off. There are two types of autoclaves in use (i.e., gravity type system or pre-vacuum-based system). The latter obtains the optimum result because it allows deeper sterilization of the contents, as it completely removes the air within, and allows high-temperature steam to penetrate and sterilize areas that would typically be occupied by ambient air, that is hard-to-reach (Baccini and Brunner 1991; Pruss et al. 1999). Gravity ones are used for non-porous items (i.e., those with hard surfaces). The third type of system is also in use, called the



**Fig. 6.4** (a) Vertical autoclave (source: [Pharmawiki](https://www.medicinesonline.com)). (b) Working of autoclave. (Source: <https://www.microsporemaster.com>)

retort type system, which operates at a much higher temperature and pressure than others (Sah 2007).

For carrying out the process of autoclaving, the wastes are kept inside a strong chamber and steam is introduced into it for a specified temperature, pressure, and time (Fig. 6.4a, b). This method applies to most biomedical wastes, especially microbiological ones; however, it is not suitable for pathological, cytotoxic, or other toxic chemical wastes (Shukla et al. 2013; Hegde et al. 2007). Steam sterilization should be carried out after separating infectious wastes from non-infectious hazards. Waste that contains antineoplastic drugs, toxic chemicals, or chemicals that would be volatilized by steam should not be steam-sterilized (Chandra 1999). These methods require simple maintenance procedures which are low cost and a popular technology in waste treatment. There is 30% reduction in waste volume if mechanical shredders are also used along with it. These can later be used compacted and used for land filling. Care should be taken not to treat anatomical or pathological wastes, or wastes containing low levels of radioactive substances or laboratory chemicals and organic solvents as operational malfunction may result in ineffective treatment.

### Microwave

It is used for disinfecting biomedical wastes using electromagnetic radiations (frequency between 300 and 300,000 MHz) in the presence of steam (Pruthivish et al. 1998). This is a relatively advanced and latest technology in the field of BMW. The wastes to be decontaminated are first shredded and mixed with steam in order to promote uniform heating and disinfecting. It is then subjected to microwave heating at 94 °C for a specified time. It is best suited for microbiology wastes, human blood, body fluids, and sharp wastes. However, it is not suitable for human and animal anatomical wastes and cytotoxins. After this, the wastes are allowed for disposal in other ways. The advantage of this method is that it can reduce the bulk volume of the



**Fig. 6.5** Incinerator. (Source: <https://www.Eco-business.com>)

waste tremendously at very minimum costs, with no emission of harmful gases, and no chemicals required (Sah 2007; Dumitrescu et al. 2007; Heberlein and Murphy 2008; Aravindan and Vsumathi 2015). It is fully computerized to handle. The use of this technology has started in the USA and European countries but is still not carried out in India.

### **Incineration**

This process uses a high-temperature dry oxidation process (Fig. 6.5). It helps in converting biomedical wastes into ash and gases. It consists of two chambers, outer and inner chambers with operating temperatures of 800–1000 °C and 850–1100 °C, respectively. There are two drawbacks of this system; the first one is that it can emit huge quantities of ash and several air pollutants such as particulate matter, metals, acid gases, oxides of nitrogen, carbon monoxide, etc. and secondly, it requires huge investment, operation, and maintenance costs together with costly emission control equipment (Nemathaga et al. 2008; Yang et al. 2009). However, such methods are being opposed by NGOs and common people in India and abroad. The setting of such facilities requires clearances as they involve risk of life due to occupational hazards and potential fire accidents. It is an old technology and was widely used in the past for all sorts of waste. However, biomedical waste, which is typically heterogeneous, is not acceptable for incineration if the combustible fraction is below 60%. Nowadays, incinerators are better equipped with pollution control



equipment that requires no pre-treatment of biomedical wastes. Since most of the biomedical waste can be incinerated, the waste does not always require sorting or separation prior to treatment. It can reduce the volume of the waste by 80% or more and solid mass by up to 85%, sterilize the waste, and reduce the need to pre-processing the waste before treatment (Goddu et al. 2007; Sorrels et al. 2017). The resulting incinerated waste can be disposed of in traditional methods, such as land filling. Modern incinerators can provide another benefit by creating heat to power boilers in the facility. It is recommended for human anatomical waste, animal waste, cytotoxic drugs, discarded medicines, and soiled waste (like dressings, plaster casts, cotton swabs, etc.) (Dumitrescu et al. 2007; McCormack et al. 1989).

### Hydroclaving

The instrument has a vessel, cylindrical in shape, double-walled, and mounted horizontally. It has a top-loading door and an unloading door at the bottom. There is a powerful motor with fragmenting/mixing arms inside it that slowly rotates the vessel. Steam is allowed to pass through the outer jacket with continuous tumbling. The optimum temperature required is 132 °C with a steam pressure of 36 psi for 20 min. During the whole process of treatment, the biomedical waste never comes in direct contact with steam. The entire process involving start-up to dehydration takes about 50 min. This helps hydroclave retain some steam back to the boiler (Sah 2007; Wallis 2010).

Moreover, it removes water from the waste and reduces the volume and weight significantly (85% and 60%, respectively) (Dumitrescu et al. 2007). However, one of the disadvantages of the hydroclave over the autoclave is that it takes more steam to heat up initially. It has to transfer the heat from the outer jacket into the vessel chamber through conduction. This initial high-energy requirement then diminishes for the continuing cycles (Fig. 6.6).

**Fig. 6.6** A Hydroclave.  
(Source: <https://www.healthmanagement.org>)

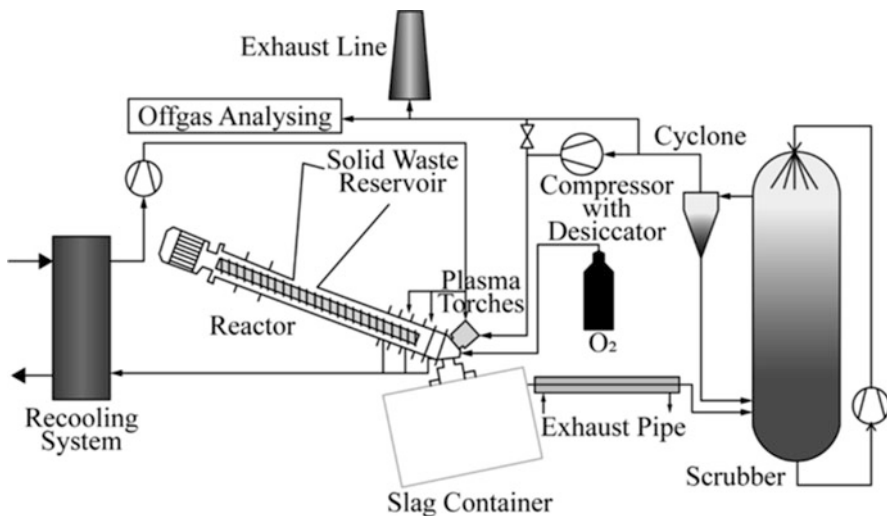


### Thermal Plasma

The technology has gained much importance these days because of generation of valuable co-products. It has attracted interest as a source of energy and spawned process developments (Heberlein and Murphy 2008). Traditional thermal technology for medical waste processing may cause indispensable secondary pollution such as dioxin, furan, heavy metals, and infectious materials that may remain in the solid residual. Thermal plasma technologies offer advantages of effectively treating medical waste due to its high temperature and energy density, lower pollutant emissions, rapid start-up and shut-down, and smaller size of the installation. These benefits play roles in treating medical waste on-site or off-site, especially when somewhere encounters an abnormally sharp increase in medical waste (Cai and Du 2020). This technique is already in commercial use for various industrial processes. Potential benefits are a more efficient use of energy, lower capital costs, and the substitution of exhaustible fossil fuels. This technology is also expected to have environmental benefits since the total gas flow rate is much smaller compared with conventional heating systems (Fig. 6.7) (Chang 2009).

#### 6.2.3.5 Irradiation Processes

one of the most advanced ways of degradation of pollutants from wastewater by the use of powerful gamma rays and beta rays (Lajayer et al. 2020) as well as less energetic Ultraviolet rays (UV) (Lee et al. 2015). The advantages of this technology are that it does not require chemical additives and no lethal by-products are produced (Chu et al. 2010).



**Fig. 6.7** Thermal plasma treatment of biomedical waste. (Source: <https://www.springer.com>)

### 6.3 Risks to Environment and Health

Disposal of waste has been known to be in civilization since 5000 BC. Since that time, the sewage system has been used to effectively dispose of waste in town planning. With urbanization and industrial development, the general public and social activists were not much aware of biomedical waste hazards and were not concerned about how the biomedical waste had to be disposed.

As per the WHO report, global life expectancy is increasing, but also there has been a steep increase in deaths due to increase in infectious disease. A study in the 1990s reported that infectious diseases such as tuberculosis, whooping cough, diarrhoea, pneumonia, etc. claimed more than 50,000 lives each day due to improper management of biomedical waste (Chitnis et al. 2002; Marinkovic et al. 2005).

With the introduction of hospitals across the cities, there was a problem in handling and disposal of waste generated during their care in hospitals. These were managed by untrained sweepers and some sanitary inspectors who did not have proper training in BMW (Park 1997). The improper management of biomedical wastes becomes a health hazard and spreads diseases in the population. They also add to environmental pollution and degradation. Hence, urgent protocols are needed to improve BMW, thereby eliminating occupational health hazards and protecting the environment.

Hospitals are involved in treating diseases, but it is also responsible for generating a large amount of biomedical waste. It has been known from several studies that patients acquire hospital-borne infections where the management of biomedical waste is poor. Though new drugs and technology for the management of diseases in the health care system are available, waste generation and their disposal have been neglected. Therefore, it is essential to take precautions in the design and organization of a hospital to minimize the risk of infection (Thomas and Timmreck. 2001).

A special attribute of biomedical waste is that even though it forms only a small part of the total solid waste, it can pollute and infect the whole solid waste if not taken care of properly. Once that happens, all the waste must be considered infected and treated as infectious waste. Improper handling, treatment, and disposal of biomedical wastes lead to pollution of air, water, and land (Sharma and Chauhan 2008). Indoors and outdoors environments can easily be affected by air pollution. The three types of air pollutions generated by biomedical waste are biological, chemical and radioactive. Indoor pollution can be due to pathogens in the form of spores that may remain suspended in the air for a long time.

On the other hand, open burning and incinerators add to the chemical pollution, which should be strictly avoided (Mandal and Dutta 2009). The dumping of biomedical waste can also pollute water bodies due to biological, chemicals, or radioactive substances in it. There is a serious threat to ground and surface water due to leakage of waterborne pathogens in the biomedical waste. Apart from harmful living organisms (pathogens), harmful chemicals, and heavy metals such as cadmium, lead, mercury, etc., present in the biomedical waste gets into the food chain after getting absorbed by plants. Salts of nitrates and phosphates that leach out into the landfills are also pollutants causing harm to crops, animals, and human beings (Mehta 1998).

Water pollution can alter the pH, BOD, DO, COD, etc. Toxins such as dioxins which are harmful to human and animal health have been present in water bodies near incinerator plants (Saini and Dadhwal 1995; Ravikant et al. 2002). Disposal of biomedical wastes inland gives rise to land pollution. Even liquid effluent after treatment is spread on land leading to land pollution. The dumping of biomedical waste in open land is the greatest cause of its pollution (Sharma and Mathur 1989).

In urban areas, improper practices such as dumping biomedical wastes in dustbins and open land and water bodies lead to diseases. Emission of harmful gases from incinerators and open burning can be carcinogenic and lead to respiratory problems (Manohar et al. 1998; Da Silva et al. 2005). Every day huge amounts of plastic wastes are thrown in the open, which choke animals upon eating them. Wastes containing sharp items can cause harm to humans and animals (Code and Christen 1999).

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## 6.4 Biomedical Waste Management Strategies

Management of biomedical wastes is a challenge for any city in a developing country due to a lack of funds and national regulations. In urban India, waste generation rates will reportedly reach 250 million tons annually by 2030, an increase of 130% from 2001 (Singh 2020). Hence each country has to frame their national legislation for the betterment of health care. It establishes legal controls and permits the national agency responsible for the disposal of biomedical waste, usually the Ministry of Health, to apply pressure for their implementation. The public, private, and informal sectors are the ones who are responsible for the waste management of any municipality. The central governments form the core of the public sector that consigns legal responsibilities of waste management to municipalities and local governments. Asian countries reportedly spend a significant \$25 billion each year on waste management, including BMW (Hoornweg and Thomas 1999), although this has not significantly improved waste management. These results in ineffective management practices including, lack of training, non-segregation, unsafe storage, lack of treatment, open dumping, and crude burning. So, the effective management of biomedical wastes requires sound legislation, training, safe handling, segregation, storage, transportation, treatment, and disposal practices (Mbongwe et al. 2008).

Apart from public sectors, private sectors have started to participate in the management of biomedical and general wastes in many developing countries (Post 1999). The advantage of having this sector is that it creates competition which ultimately brings down the management costs. This sector has less political interference and hence more effective in running the system smoothly (Zhu et al. 2008).

The third sector (i.e., the informal sector) is also a strong pillar in many developing countries. They contribute significantly to waste management and resource efficiency by collecting, sorting, trading, and sometimes even processing waste materials. Moreover, the informal sector activities are highly adaptable, flexible, and able to respond quickly to demand-driven forces. In India, the informal waste sector is socially stratified in a pyramid with scrap collectors (waste pickers and

itinerant waste buyers) at the bottom and re-processors at the top. Policies with the legal provision are necessary to assist in the effective management of biomedical waste (Phillips 1999).

Proper management of biomedical wastes starts at the source (i.e., segregation). This will help the medical/health care authorities to save money on the cost of disposal. Moreover, it will help to reduce the amount of infectious biomedical wastes from general wastes at the source. This saves more than 50% of costs, thereby minimizing health risks and costs of environmental hazards.

WHO (2005a, b) states that policies and plans should be implemented to ensure comprehensive waste management from production to disposal. It is required that hospitals and other areas that generate clinical waste comply with good practices and legislation regarding its disposal.

Still, there is no documentation of BMW policy, which leads to delay in final disposal.

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## 6.5 Handling of Biomedical Wastes During COVID-19 Pandemic

COVID-19 pandemic brought unprecedented challenges to all sectors including health care sector. This created panic in health sector and everyone was affected by it. Death toll started to increase with each passing day which roused a sense of fear even in health care professionals. This pandemic resulted in huge generation of BMW which presented a threat to the existing BMW infrastructure worldwide. Hence safe disposal of COVID-19 biomedical waste was a challenge (Dehal et al. 2022).

Increase in use of medical technologies in health care system to prevent spread of COVID-19 has generated tremendous amount of biomedical wastes raising fear among biomedical wastes handlers leading to occupational stress (Ma et al. 2020). Use of personal protective equipment (PPEs), boots, face shields gloves, goggles, along with sanitizers, masks, syringes, testing kits, etc. have added to the existing biomedical waste composition (Das et al. 2021; Praveena and Aris 2021). In spite of all these hazards, the knowledge about segregation and management helped reduce COVID-19 wastes.

Looking at the sensitivity of the situation and specific need of the local civic bodies, it was very urgent to evolve our own approach towards COVID-19 waste management. In India, the CPCB is responsible for the implementation of BMW (2016) rules. CPCB (2016) issued guidelines to treat BMW management as “essential services” and ensured the uninterrupted movement of vehicles and people involved in COVID-19 BMW management. There was adequate supply of yellow, red, white, and blue bags and containers to all the hospitals and as well as at the quarantine facilities so that proper segregation and collection of biomedical wastes can be done.

Our Medical University is one of the oldest and biggest in the country providing tertiary care to admitted patients. During the pandemic times, the majority of hospital

wards were converted into COVID facilities which few were still catering to non-COVID cases as well. This helped to segregate COVID and non-COVID wastes. Collection and transportation of COVID-19 wastes were carried out by dedicated staffs in PPE. These biomedical wastes were continuously handed over to the authorized agency for further processing.

Handling of solid and liquid COVID biomedical wastes should be done as per the guidelines recommended. That is, using color-coded bins for onsite segregation, carrier trolleys for handling of BMW generated at COVID-19 areas, regular cleaning of trolleys with 1–2% sodium hypochlorite solution, maintaining a separate record of COVID-19 related activities, liquid wastes should be treated chemically, personal protective equipment should be given to all persons involved in COVID-19 BMW handling, and should follow basic hygiene and infection-control measures with regular health screening (Arya and Mandavkar 2020; WHO 2020; Chand et al. 2021).

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## 6.6 Conclusion and Recommendations

The health care sector must understand the importance and seriousness of BMW and comply with the rules and regulations of their waste management policy. The responsibility lies at the first step of segregating biomedical wastes at the source of generation, collecting them in prescribed colour-coded bags, followed by safe transportation, applicable treatment, and proper disposal of biomedical wastes. Apart from this, training programs should be conducted in their set up for all and especially for those who are responsible for such management. These things have to be implemented effectively accountability should be fixed for each and every person involved in management of biomedical waste. This will help to protect not only our health but also our environment.

The following recommendations are to be noted:

- In coordination with the Ministry of Environment and other concerned ministries and local administration, any country's health ministry should specify the responsibilities towards managing biomedical waste within and outside the health care establishments.
- There is need for sustained cooperation among all key actors (government, hospitals, and waste managers) in implementing a safe and reliable medical waste management strategy, not only in legislation and policy formation but also particularly in its monitoring and enforcement. This can be achieved through the cooperation between the Ministry of Health, Environmental Quality Authority, Ministry of Local Government, and Non-Governmental Organizations working in related fields.
- It should be the responsibility of each health care facility (HCF) to ensure a safe and hygienic system of medical waste handling, segregation, collection, storage, transportation, treatment, and disposal, with minimal risk to handlers, public health, and the environment.

- All staff and waste handlers in each hospital should be well trained at the beginning of their work at hospitals and regularly updated with pre-employment and in-house specialized training, which provides them with a knowledge base about the process of waste management and associated health risks.
- Economically and environmentally sustainable technological options for waste treatment, which can be well operated and maintained, should be considered for medical waste management.
- There should be a hazardous waste landfill specially designed for the final disposal of treated hazardous healthcare waste. Its specifications are well known in the international literature, and we should benefit from that.
- There should be proper documentation on the quantity of medical waste generated per day/week/month/year to serve as a guide for effective and efficient planning.
- Waste should be segregated using management tools like colour-coding and proper labelling of waste containers. There should be appropriate and modernized methods of disposing of and treating medical waste.
- Infectious waste should be treated and disposed of separately from non-infectious waste.
- A waste management department headed by a waste management Officer should be in place to ensure effective supervision of the waste workers.
- There should be regular training programs for all categories of health workers concerning waste management.
- Waste management policy/legislation should be in place to regulate how waste would be managed.
- A waste management manual or guide document should be provided to guide waste handlers on how best to handle medical waste such as infectious and non-infectious.

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## **Part II**

# **Microbial Approach in Bioenergy Production**



# Microbial Intervention in Waste Remediation for Bio-Energy Production

# 7

Uma Chaurasiya, Akshay Joshi, Ashutosh Kumar, Wolfgang Merkle, Hans-Joachim Nägele, Deepak Kumar Maurya, Deepanshu Jayashwal, Nishtha Srivastava, and Vineet Kumar Maurya

## Abstract

The extensive exploitation of fossil fuels and the increasing global demand for energy entailed producing alternative fuels to swamp fossil fuels. Production of biofuels from biological, agricultural, municipal, and other waste products can be an alternative option to fossil fuels. Presently, biofuel production from waste products has marginally reduced the dependency on fossil fuels for energy. Eco-friendly renewable energy fuels such as biodiesel, bioethanol, biobutanol, biohydrogen, and biogas resulting from biomass conversion from agricultural waste, microalgae, or biological wastes have significantly contributed to the wellness of the economy as well as the environment. Biofuels are generated by biological processes such as fermentation via applications of suitable microorganisms from different genera with diverse biofuel production

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Akshay Joshi contributed as the first author.

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U. Chaurasiya

School of Studies in Botany, Jiwaji University, Gwalior, Madhya Pradesh, India

A. Joshi · W. Merkle · H.-J. Nägele

ZHAW School of Life Sciences and Facility Management, Wädenswil, Switzerland

A. Kumar (✉) · D. Jayashwal

ICAR - Indian Institute of Seed Science, Mau, Uttar Pradesh, India

D. K. Maurya (✉)

Agharkar Research Institute, Pune, Maharashtra, India

N. Srivastava

Department of Applied Science, Invertis University, Bareilly, Uttar Pradesh, India

V. K. Maurya

Department of Botany and Microbiology, H. N. B. Garhwal University, Srinagar Garhwal, Uttarakhand, India

mechanisms. The effect of wastes on the environment, potential waste products which could be used as raw material for biofuels production, types of biofuels produced from the waste products, and potential microorganisms used in biofuel production have been discussed in the present chapter. Emphasis has been given to putative biochemical pathways involved in bio-energy production, along with recent research and updates on utilising different sustainable resources for bio-energy production. Finally, the chapter has concluded with prominent challenges encountered during biofuel production from waste materials and potential mitigation strategies for them.

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**Keywords**

Biofuel · Bioethanol · Energy demand · Microbes · Sustainability

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## 7.1 Introduction

The world population is expected to reach 9.7 billion by 2050 (FAO 2009). The increased population requires food and energy security, along with the augmentation and up-gradation of current technologies used to dispose of agricultural, food, and other wastes in an eco-friendly manner. The rapidly depleting fossil fuel sources, increasing energy demand, and rising environmental pollution levels have pushed the world to look for alternative, sustainable, and environmentally safe energy sources. Waste, an inevitable by-product of day-to-day human activities, could be an alternative source of energy. Due to the widening industrialisation and rapidly growing demand for food supply, waste generation will be an unavoidable threat in the near future. The emission of greenhouse gases and the accumulation of solid wastes are the associated risk factors with the waste. Hence, converting waste into energy could be an effective method to mitigate the energy crisis and pollution. The conversion of biodegradable (agriculture and food wastes) wastes into biofuel is a good choice, which is being explored extensively for energy production.

The initial biofuel production approaches had severe drawbacks and needed inevitable improvement. For example, the production of first-generation biofuel (bioethanol from the substrates with high starch content, such as corn, wheat, etc.) uses to demand food materials for biofuel production. The negative aspect of this approach was that it required food crops. Hence it was snatching the food reserve as well as agricultural land. This increased the pressure on crop production from 2000 to 2015 (FAO and OECD 2019). FAO reviewed the first-generation biofuel production and warned about its dangers in 2009 (FAO 2009). If this approach was followed, it would have resulted in a serious risk to food security for humans and overuse of agricultural land. Hence, agricultural diversification and alternatives to food crops were searched for biofuel production. Currently, extensive research works on third and fourth generation biofuels.

Various technologies are being used for biofuel production from biodegradable and non-biodegradable wastes, which can be classified into biochemical and thermochemical processes (Jeguirim and Limousy 2018; Bharati et al. 2020). In biochemical processes, microorganisms play a crucial role in transforming organic biomass into biodiesel, bioethanol, and biogas. Whereas in thermochemical processes, bio-hydrogen and bio-oil are produced by combustion, gasification, and pyrolysis. The selection of the processes for biofuel production primarily depends on the feedstock's nature and available pre-treatment methods (Singh and Das 2019).

Recent technologies have shown the potential of microorganisms in the production of bioethanol and biogas. The innovation in bioethanol production from first- and second-generation biofuel using yeast and genetically engineered bacterial strains has been well known for the past few years. Recent studies also reveal the high yields of alcohol from syngas using acetogenic bacteria in indirect fermentation (Liou et al. 2005; Maurya et al. 2020). Similarly, processing algal lipids is a promising and carbon-neutral approach to converting sunlight and CO<sub>2</sub> into biodiesel. Hence, in this chapter, the classes of biofuels and the potential of microorganisms in converting deteriorating wastes into beneficial biofuels have been described in detail.

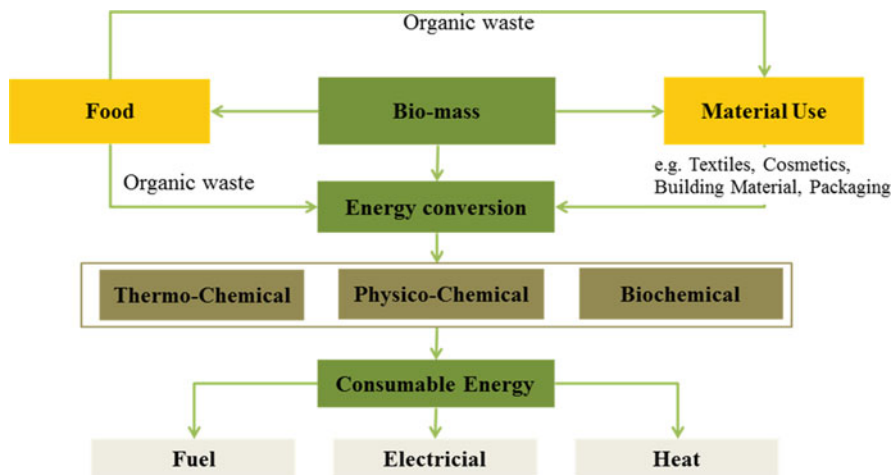
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## 7.2 Potential Biofuels Transformed from Wastes

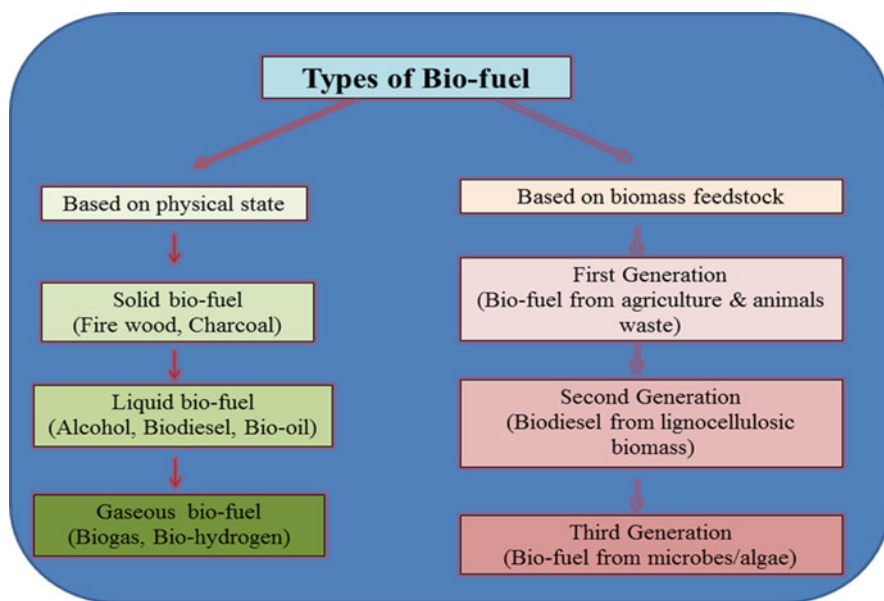
### 7.2.1 Types of Biofuels

Biomass is one of the most valuable sources as it supplies food, feed materials, and energy in a human-dominated ecosystem of the Earth. In the context of a renewed return to a so-called biobased economy, as it was practised for many centuries before industrialisation, a new focus will be laid on the production of food, feed, bio-based materials, and bioenergy from biomass. Therefore, new value chains will have to be developed that include the primary production of biobased resources, their conversion to higher-value goods, and their energetic use after their lifespan or from wastes produced alongside the value chains (Zörb et al. 2018) (Fig. 7.1).

Biomass can be converted into usable energy such as fuel, electricity, and heat via three different conversion pathways: thermo-chemical, physio-chemical, and biochemical pathways (Madakka et al. 2020). Various biomasses can be converted into energy carriers in solid, liquid, and gaseous forms using either of these three pathways (Fig. 7.1). Thermochemical conversion includes the processes of carbonisation, gasification, or pyrolysis and will result in solid, gaseous, and liquid forms of bioenergy. In Physico-chemical conversion, the biomass is given mechanical and chemical treatment, resulting in the extraction of plant oils. The plant oils are converted into biofuels after their transesterification. In biochemical conversion processes, alcoholic fermentation and anaerobic digestion transform the biomass into liquid, and gaseous energy carries.



**Fig. 7.1** Conversion paths from biomass to energy



**Fig. 7.2** Categorisation of biofuels based on their physical state and biomass Feedstock

Biofuels are renewable fuels derived from biomass through thermo-, physio-, or biochemical reactions. Depending on the feedstock used, three generations of biofuels are identified in the literature (Fig. 7.2). “First-generation” biofuels are based on food crops, such as wheat, barley, rapeseed, sugarcane, and corn, and thus have direct competition with food and feed. These raw materials have been the

subject of much debate worldwide as their use may lead to food shortages. For this reason, the use of “second-generation” or “advanced” biofuels, based on non-food crops and lignocellulosic material that will have reduced or no food competition, increased. To avoid any competition with food or feed a “third-generation” of biofuels based on algae or other microorganisms has been the focus of research as those resources will have only little land requirements (Loeffler et al. 2018; Zörb et al. 2018; Kumar et al. 2019a). Nowadays, research on “fourth-generation” which consists of combining genetically engineered feedstock with genomically synthesised microorganisms, is also being carried out to increase the efficiency of biofuel production from biomass (Mansoori et al. 2021).

Biofuels are classified into solid, liquid, and gaseous energy forms according to their physical properties (Fig. 7.2).

### 7.2.1.1 Solid Biomass

The use of solid biomass to derive energy is known as solid biofuels and has been classified into four well-known types of solid biofuels.

1. **Firewood:** Wood is the ancient biofuel source being used for thousands of years for the production of heat and light and other domestic purposes. Before its use as firewood, the wood needed to be dried with its moisture content reduced to about 10–25%. Compared to green firewood, dried wood burns more quickly and efficiently. But, the burning of firewood or fuelwood also produces hazardous greenhouse gases, which cause a negative impact on the environment.
2. **Woodchips:** wood chips are a processed form of firewood that is easier to handle and faster to burn. It is mostly used in areas where mechanical forestry equipment is available.
3. **Wood pellets:** In the wood pellets, the wood is converted into sawdust and processed at high temperatures. At high pressure, the temperature rises, and the lignin melts and glues the sawdust into pellets. Afterwards, the pellets are broken into pieces of 2–3 cm in length. Nowadays, wood pellets made from seed husk, formed after oil extraction, have a high demand for animal feed.
4. **Charcoal:** Charcoal has a much higher energy content compared to the other forms of wood biofuels. Charcoal is produced after the wood materials are heated below 400 °C temperature in the absence of air.

### 7.2.1.2 Liquid Biofuels

Liquid biofuels are transport fuels obtained from biomass. They are refined products of biomass feedstock. Bioalcohols (bioethanol and biomethanol) and biodiesel formed from bio-oil are examples of liquid biofuels.

1. **Bioethanol:** Bioethanol is produced by direct and indirect fermentation processes. In direct fermentation, ethanol is made from simple sugars obtained from either first-generation (wheat, beetroot, corn, and sugar cane) or second-generation biofuels (Stover, straw, stem, and stalks) sources (Elshahed 2010). In first-generation biofuel, extraction of sugar syrup is relatively simple. Hence,



microbial and enzymatic treatments are not required for pre-treatments. Sugar syrup is converted into ethanol using genetically engineered yeast and bacterial strains. Due to increasing debates on fuel Vs food during the past few years, various countries have moved from the first-generation biofuel to second-generation biofuels. In the second-generation biofuels, the lignocellulolytic microbial (bacteria and fungi) strains are used for the initial hydrolysis of complex sugars (polysaccharides) into simple sugars (oligo, di, or monosaccharides). These simple sugars are then subjected to microbial fermentation for bioethanol production (Lau and Dale 2009). Indirect fermentation is a promising approach for ethanol production. In this process, plant material is converted into syngas by pyrolysis. Syngas contains CO, CO<sub>2</sub>, and hydrogen (H<sub>2</sub>), which are then transformed into ethanol by anaerobic acetogenic bacteria (Tanner 2008).

2. **Biomethanol:** The preparation of biomethanol involves the gasification of carbohydrates from biomass and their partial oxidation. Compared to producing methanol from fossil fuels, the production of biomethanol from biomass is expensive. Hence only a tiny percentage of biomethanol is produced from biomass. Methanol is used as fuel, fuel additive, and an important base chemical for industries. Low flammability, high performance, and low emission of pollution are the advantages of using biomethanol (Pirola et al. 2018).
3. **Biodiesel:** Biodiesel consists of alkyl (C1-C4) esters of long-chain fatty acids. The production of biodiesel involves the transesterification of biological lipids (raw plant oil, animal fat, and waste oil) in the presence of methanol. A base is also used during the transesterification of lipids to form a liquid fuel. Biodiesel is used either as a substitute or as an additive for diesel. The lipids from photosynthetic algae are processed to produce biodiesel. This promising process is also popular as an eco-friendly and carbon-neutral process of biofuel production due to converting greenhouse gas CO<sub>2</sub> into biodiesel using sunlight. The process also has high carbon-fixation efficiency because the growth rate of microalgae is much faster than oil crops, and the extraction of oil exceeds about 80% of the dry biomass (Chisti 2007).
4. **Bio-oil:** Bio-oil is a pyrolysis product and comes along with other products such as biochar and syngas. Modification and optimising the conditions during pyrolysis can increase the amount of bio-oil. Bio-oil is a mixture of many compounds such as acids, alcohols, aldehydes, esters, ketones, sugars, alkenes, aromatic and nitrogen compounds, and many others. However, bio-oil is difficult to burn due to excess moisture. Moreover, it is also volatile, corrosive, and adhesive.

In recent studies, algae with high lipid profiles (e.g. arachidonic, eicosapentaenoic, and docosahexaenoic acids) have been used for the production of bio-oils. The major challenge in this process includes the development of low-cost extraction methods (Baskar et al. 2019).

### 7.2.1.3 Gaseous Biofuels

Gas and its products are extensively used for cooking, heating, transportation and electricity generation as they are very flexible in their use. Biogas, biohydrogen, and syngas are some types of gaseous biofuels.

1. **Biogas:** The anaerobic digestion of organic waste, sewage sludge, animal wastes, or energy crops using microorganisms leads to a mixture of gases known as biogas. This process works in four stages: hydrolysis, acidogenesis, acetogenesis, and methanogenesis. In the hydrolysis step, the microorganism ferment complex biomass into long-chain and short-chain volatile fatty acids. The product formed in acidogenesis is utilised by acetogenic bacteria to produce  $H_2$ ,  $CO_2$  and acetate, which is finally used up by methanogens to produce methane (Borja and Rincón 2017).

Biogas is composed of approximately 60–65% methane ( $CH_4$ ) and 30–35% carbon dioxide ( $CO_2$ ). However, the exact composition depends upon the feed material. Other gases  $H_2$ , hydrogen sulphide, and water vapours are also in lower amounts. Following the purification and concentration of biogas, it can be combined with heat and power units to generate heat and electricity. In addition, biogas can be injected into the gas grid or liquefied using pressure for fuel purposes.

2. **Biohydrogen:**  $H_2$  is an ecologically pure biofuel because it does not release any harmful gases upon combustion. Pyrolysis of biomass, such as waste, crop straw, municipal solid waste, crop grain residue, pulp waste, or manure slurry, results in the synthesis of biohydrogen.  $H_2$  is also formed as a final product in the fermentation process by the  $H_2$ ase enzymes in microorganisms (Vignais and Billoud 2007).

In photobiological  $H_2$  production, photosynthetic microbes such as *Cyanobacteria* and green algae are also well known to produce low-cost  $H_2$ . These photosynthetic microbes split the water molecules into electrons and oxygen. The hydrogenase enzyme can convert the produced electron into  $H_2$  (Prince and Ksheshgi 2005).

3. **Syngas:** Synthesis gas (syngas) is produced by pyrolysis or gasification of plant biomass or biobased gases. Carbon monoxide (CO) and  $H_2$  are the main components of syngas, accompanied by  $CO_2$ ,  $CH_4$ , hydrogen sulphide, water vapours, etc., depending on the biomass composition. Power to Gas technologies such as catalytic and biological methanation is becoming increasingly important (Martín 2016).

The syngas can be injected into the grid, liquified for fuel, and used to produce other fuels such as diesel. Moreover, syngas is the leading source for producing various chemicals such as ethanol, methanol and ethane. The  $H_2$  separated from syngas is used in fuel cells for electricity generation (Wu and Tu 2016).

### 7.3 Substrates for Biofuel Production

Due to the shortage of fossil fuels and environmental issues, renewable, environment-friendly fuels are becoming more important nowadays. Fuel crisis and treatment and proper usage of organic wastes are among the significant global challenges. Both challenges can be addressed by using organic wastes for biofuel production. Based on their origin, organic wastes can be classified into agricultural/forestry and non-agricultural/forestry wastes (Table 7.1). Agricultural wastes (by-products, co-products) are usually defined as non-food or feed plant or animal residues generated from either harvesting crops/trees or rearing animals. Compared to agricultural waste, the non-agricultural organic wastes (biowastes) include all organic wastes from the domestic, food, municipal, and industrial sectors.

All these wastes can generally be used for the production of biofuels. Depending on their composition (content of carbohydrates, proteins, lipids, cellulose, hemicellulose, lignin) and their dry matter they can be used to produce specific kinds of biofuels.

#### 7.3.1 Biofuels from Different Types of Biomass

Wastes with high content of dry matter like forestry residues and by-products from forest, straw, bagasse, solid animal waste, and other vegetal materials can be used to produce solid biofuels. These solid biofuels can substitute common wood-based biofuels. A homogenous fraction is a good choice for producing liquid biofuels from biowastes. Lipid-rich wastes from restaurants, catering, retail premises and food processing plants are suitable materials for producing liquid biodiesel. Waste biomass rich in starch, sugar, and lignocellulosic material is a good choice for the production of bioethanol and biomethanol (Yadav et al. 2020). However, this method is still in the infancy stage of development (Hirschnitz-Garbers and Gosens 2015). The production of bio-oils by pyrolysis of wastes is currently under optimisation at an industrial scale. Once optimised, this method can also use different biowastes to produce bio-oils (Karmee 2016). Gaseous biofuels (biohydrogen and syngas) are also released by pyrolysis or gasification of wastes.

**Table 7.1** Classification of organic wastes (modified—according to Pimiä et al. 2014)

Types	Organic wastes
Agricultural/forestry wastes	Forestry and agricultural residues, Manure
Non-agricultural/forestry wastes	food and kitchen waste
• Food waste	Household waste, Restaurant waste, Catering waste Retail premises waste, waste from food processing plants
• Industrial waste	Nature textiles, paper, processed wood
• Municipal waste	Garbage, Biodegradable garden and park waste, sewage sludge

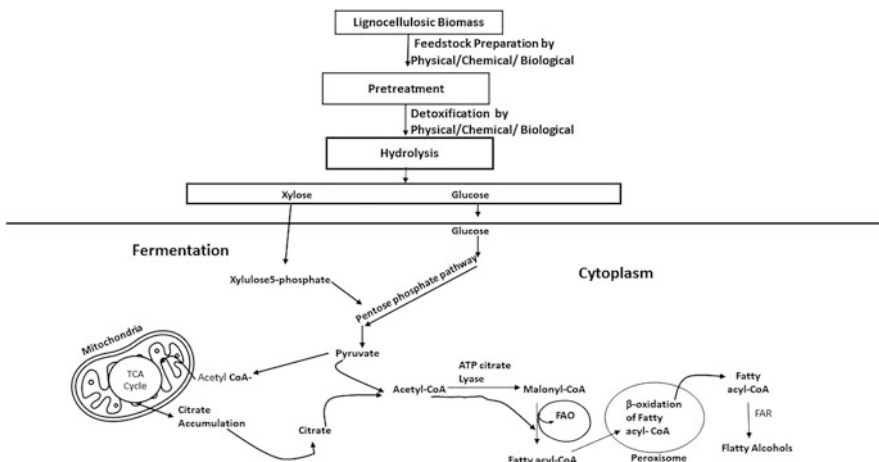
Unlike bio-oil production from waste, the production of biogas from waste is already an optimised method being practised worldwide. Biogas is another gaseous biofuel, produced utilising a variety of putrescible organic wastes, such as agricultural residues, manure, food wastes, industrial wastes, sewage, and the organic fraction of municipal solid waste (MSW). The high lignin and lignocellulosic contents lower the specific biogas yield (De Simio et al. 2008).

### 7.3.2 Pre-treatment of Waste Prior to Microbial Treatment

Biofuel usually starts with a preliminary feedstock preparation step involving cleaning and size reduction by milling, grinding, or chopping. All these steps consume a large amount of energy. Subsequently, the process follows four major steps: (1) pre-treatment, which involves degradation of the complex lignocellulosic network into smaller units, (2) Hydrolysis/saccharification to obtain fermentable sugars, (3) fermentation to convert sugars into ethanol, and (4) Purification (recovery and dehydration) to obtained good quality ethanol (Fig. 7.3).

#### 7.3.2.1 Pre-treatment

Naturally occurring forms (crystalline structure) of cellulose have high resistance to hydrolysis. The presence of lignin also limits enzymatic hydrolysis by adsorption of enzymes. Pre-treatment performs de-lignification, degradation of hemicelluloses and reduction in cellulose content. Pre-treatments can be physical (e.g. milling, grinding, and microwave), chemical (acid, alkali, ozonolysis, organosolv, and ionic liquids), physicochemical (steam explosion, ammonia fibre explosion, CO<sub>2</sub> explosion, liquid



**Fig. 7.3** Biochemical pathway of biofuel production from waste

hot water, and wet oxidation), or biological. During pre-treatment, lignocellulosic biomass several compounds such as (1) furfural and HMF (5-hydroxymethyl-2-furaldehyde), originating from the degradation of hexoses and pentoses, (2) acetic acid, originating from hemicelluloses, and (3) phenolic compounds originating from lignin are generated. These compounds are toxic to microorganisms, inhibit their growth, and extend the lag phase. So, several detoxification technologies are used to remove these toxic compounds.

### **7.3.2.2 Hydrolysis/Saccharification**

It is a crucial step in which sulphuric acid or hydrochloric acid or enzymes are used to convert cellulose and hemicelluloses into their monomers, i.e. fermentable sugars using the process of acid or enzymatic hydrolysis at low temperature, followed by microbial fermentation for the production of biofuel (Branco-Vieira et al. 2018).

### **7.3.2.3 Fermentation**

Different enzymes like xylanases, laccases, chitinases, cellulases, and proteases play a dedicated role in bioconversion. For example, xylan and cellulose as substrates are used for biofuel production. Bioconversion of the sugars to bioethanol occurs through fermentation, involving microorganisms (Adegboye et al. 2021; Soni et al. 2020).

### **7.3.2.4 Purification**

Lastly, the product obtained needs to undergo the process of purification and distillation, which involves separating the bioethanol, in pure form, from the fermentation broth. The quantity of bioethanol obtained from the fermentation process mainly depends on the amount of sugar produced during pre-treatment and hydrolysis/saccharification. The total yield of bioethanol can be measured in terms of the volume of ethanol produced per dry weight of raw material (Adegboye et al. 2021).

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## **7.4 Biological Agent in Biofuel Production from Waste**

### **7.4.1 Bacteria**

Microorganisms are considered alternative sources for the production of biofuels. Bacteria have significant advantages over higher plants and microalgae for synthesising intracellular as well as extracellular fatty acids to produce environment-friendly fuel oil (Kumar et al. 2020). Fast-growing bacteria can potentially use a wide range of feedstocks for biodiesel production. Bacteria effectively use agricultural by-products for their growth and utilise sugar and proteins pre-set in waste materials (Mihajlovski et al. 2020). Some of the well-known potential biofuel-producing strains of bacteria have been summarised in Table 7.2. Activated sludge contains a microbial population of heterotrophic bacteria responsible for wastewater treatment. These bacteria use the organic compounds in wastewater for their growth

**Table 7.2** Microorganisms in biofuel production

Organisms	Biofuel type	References
<i>Acinetobacter calcoaceticus</i>	Lipid	Choi et al. (2014), Moshtagh et al. (2021)
<i>Alkalibaculum bacchi</i>	Ethanol	Allen et al. (2010), He et al. (2022)
<i>Bacillus</i> sp. ( <i>B. mycoides</i> , <i>B. amyloliquefaciens</i> , <i>B. pumilus</i> )	Butanol	Kanno et al. (2013), Shabbir et al. (2022)
<i>Clostridium acetobutylicum</i>	Acetone, butanol, and ethanol	Ennis et al. (1986), He et al. (2022)
<i>Clostridium beijerinckii</i>	Isopropanol, butanol, and ethanol	Hettinga et al. (2009), Comwien et al. (2015), He et al. (2022)
<i>Clostridium carboxidivorans</i>	Ethanol, butanol	Fernández-Naveira et al. (2016), He et al. (2022)
<i>Clostridium phytofermentans</i>	Ethanol	He et al. (2022)
<i>Clostridium ragsdalei</i>	Ethanol	Devarapalli et al. (2017), He et al. (2022)
<i>Clostridium thermocellum</i>	Ethanol	Ng et al. (1981), He et al. (2022)
<i>Costridium saccharoperbutylacetonicum</i>	Butanol	Shukor et al. (2014), He et al. (2022)
<i>Cryptococcus curvatus</i>	Lipids	Yu et al. (2011), Kamal et al. (2022)
<i>E. coli</i>	Ethanol, 1-Propanol, 1-pentanol isobutanol, 1-butanol	Asghari et al. (1996), Zhang et al. (2008), Ku et al. (2022)
<i>Lactobacillus brevis</i>	Butanol	Russmayer et al. (2019), Esquivel-Hernández et al. (2022)
<i>Lipomycesstarkeyi</i>	Lipids	Yu et al. (2011), Zhang et al. (2022)
<i>Pseudomonas putida</i>	Butanol	Sahoo et al. (2019), Sarwar et al. (2022)
<i>Rhodococcus opacus</i>	Lipid	Le et al. (2017), Nair and Sivakumar (2022)
<i>Rhodospiridium Toruloides</i>	Lipids (Glucose and xylose)	Xie et al. (2012), Gao et al. (2022)
<i>S. cerevisiae</i>	Ethanol	Sharma et al. (2022)
<i>S. stipitis</i>	Ethanol	da Silva et al. (2022)
<i>Zymomonas mobilis</i>	Ethanol	Li et al. (2022)

and store the organic material in the form of lipid droplets. Oleaginous bacterial species belonging to the order Actinomycetales (*Mycobacterium*, *Streptomyces*, *Nocardia*, and *Rhodococcus*) can accumulate lipid up to 20% or more of their biomass (Cea et al. 2015). *Acidothermus*, *Bacillus*, *Clostridium*, *Pseudomonas*, and *Rhodothermus* degrade cellulose. A wide assortment of Gram-positive and

Gram-negative cellulose-degrading bacterial species includes *Clostridium thermocellum*, *Streptomyces* sp., *Ruminococcus* sp., *Pseudomonas* sp., *Cellulomonas* sp., *Bacillus* sp., *Serratia* sp., *Proteus* sp., *Staphylococcus* sp., and *Bacillus subtilis* (Kashyap et al. 2019; Khedr et al. 2019). *Geobacillus* is an obligate thermophilic bacteria which can generate and enhance the productivity of important bioenergy sources such as ethanol, isobutanol, 2,3-butanediol, biodiesel, and biogas at the temperature range of 35–75 °C (Novik et al. 2018).

Biogas is an effective source of renewable energy. Anaerobic microorganisms produce biogas by organic decomposition of domestic and agricultural waste as a substrate. CH<sub>4</sub> is the main combustible element of biogas, forming 50–75% volume of biogas. Remaining 25–50% volumes consists of non-combustible gaseous elements, such as CO<sub>2</sub>, N<sub>2</sub> (<1%), O<sub>2</sub> (0–1%), and nitrogen siloxanes (0–0.02%), halogenated hydrocarbons (<0.6%), CO <0.6%, hydrogen sulfide (0.005–2%), and water vapours (5–10%) (Wellinger and Lindberg 1999). *Thermovirga*, *Soehngenia* and *Actinomyces* are H group-containing bacteria that have more capacity to generate CH<sub>4</sub> than the black group. These microbial communities (black and H group) have been categorised with the help of Illumina sequencing. Archaeal species like *Methanosaeta*, *Methanolinea*, *Ethanospirillum*, and *Methanoculleus* are reported in both groups (Wang et al. 2017). Bioaugmentation strategies for enhancing biogas production plays a crucial role during the anaerobic degradation of cow manure. These bacterial strains include *Rikenellaceae*, *Clostridiaceae*, *Porphyromonadaceae*, *Bacteroidaceae*, and *Ruminococcaceae*. *Flavefaciens* and *Ruminococcus albus* showed CH<sub>4</sub> production at 41 °C (Ozbayram et al. 2018).

Biodiesel, consisting of mono-alkyl esters, is produced by the transesterification of edible and non-edible oil/fat from plant and animal origin. The use of biodiesel over conventional fossil fuel-based diesel offers several advantages, such as less emission of greenhouse gases, other gaseous pollutants and particulate matter (Behera et al. 2019). Oleaginous bacteria *Rhodococcus opacus* produce 80% biodiesel of its cellular dry weight using wastewater from corn stover (Le et al. 2017). Moreover, *Serratia* sp., a chemolithotroph, uses municipal secondary sludge as growth media for biodiesel production. These bacteria apply several strategies for their adaptation to produce lipids, bioplastics, exopolysaccharides and fatty acids (Kumar et al. 2020).

Bioethanol is an important alternative to fossil fuels and contributes to the economy by using domestic and environmental wastes. It is a safe, efficient and non-toxic biofuel produced without any by-products (Younesi et al. 2005; Eriksson and Kjellström 2010). The organic fraction of MSW comprises 50% lignocellulose-rich material. *Zymomonas mobilis* and *Rhodococcus opacus* have the potential of producing ethanol from MSW (Dornau et al. 2020). Brigham (2019) reported that Knallgas bacteria produce different types of high-energy-density transportation fuels by utilising CO<sub>2</sub>, H<sub>2</sub>, and O<sub>2</sub>. *Ralstonia eutropha* is a Knallgas bacterium, which has been genetically engineered to produce *n*-butanol, isobutanol, and terpene under chemolithoautotrophic conditions. Many extremophilic bacterial species, mainly thermophilic microorganisms, produce cellulase enzyme which increases the rates of cellulose hydrolysis. *Clostridium thermocellum*, *Thermoanaerobacter*

*thermohydrosulfuricum*, and *Clostridium stercorarium* subsp. *thermolacticum* not only efficiently degrades cellulose and hemicelluloses through hydrolysis but also readily ferments the pentose and hexose sugars (Di Donato et al. 2019). Ethyl alcohol is produced using syngas fermentation, in which anaerobic microorganisms (*Clostridium ljungdahli*, *C. tetanomorpum*, and *Clostridium* strain P11) utilise accessible carbon and energy source to produce ethanol biofuels (Williams et al. 2015; Kundiyana et al. 2010).

### 7.4.2 Yeast/Fungi

Fungi degrade the biomass of agricultural waste through biochemical and thermochemical processes to produce biofuels. Biochemical conversion leads to bioethanol and biodiesel production (Maurya et al. 2020). Endophytic fungi produce compounds such as alkanes, cyclohexanes, cyclopentane, alkyl alcohols/ketones, benzenes, and polyaromatic hydrocarbons found in biodiesel (Raven et al. 2019; Kumar et al. 2023). *Rhizopus Oryzae* fungi have been demonstrated to efficiently catalyse the methanolysis of vegetable oils for biodiesel production in solvent-free systems (Nagaraj et al. 2010). Some of the fungi used for biofuel production have been presented in Table 7.3.

Filamentous fungus *Aspergillus* sp. produces biodiesel with good fuel quality (acid number, 0.40 mg KOH/g of acid; iodine value, 11 g I<sub>2</sub>/100 g oil; density, 0.8342 g/cm<sup>3</sup>) using corncob waste liquor (CWL) as substrates (Subhash and Mohan 2011). Moreover, *Aspergillus niger* and *Trichoderma harzianum* have been reported to perform the alkali and enzymatic hydrolysis of rice husks (Solanki et al. 2019; Abbas et al. 2022). This hydrolysed husk can be used for bioethanol production via fermentation using *Saccharomyces cerevisiae* (Ahmad et al. 2017). Similarly, the co-culture of *Aspergillus niger* and *Saccharomyces cerevisiae* produce ethanol from the rice wastewater (Hatami et al. 2015; Gujjala et al. 2019). Furthermore, Subhash and Mohan (2015) reported that *Aspergillus awamori* uses CWL, paper mill effluent (lignocellulosic wastewaters) and cellulosic waste (de-oiled algae extract, DAE) as feedstock for single cell oil (SCO) production. DAE improvises biomass production by reducing production time; however, the high feedstock cost is a major limiting factor. Oleaginous fungi are cultured with lignocellulosic materials for lipid production, which produces biofuel at a comparatively lower cost due to the abundance of low-cost feedstock, such as glycerol, sewage water, whey and molasses. Oleaginous microorganisms have multiple advantages (Zheng et al. 2012), such as (1) capacity to accumulate 80% of lipid and increase the quality of fatty acids, (2) having good lipid profiles, suitable for making high-quality biodiesel, (3) capacity to utilise monosaccharides, glycerol, acetic acid, cereal, corncob, sweet sorghum, wheat straw, orange peel, apple pomace and oil for lipid production, (4) low capital cost and low energy expenditure is required for oil production, through solid-state fermentation, and (5) ease of oil harvesting from cell broth by using simple filtration after pellet formation, and reduction in the viscosity of the fermentation broth to



**Table 7.3** Role of important microbes in fuel production from different feed stocks

Organism	Biofuel	Feedstock	References
<i>Trichoderma asperellum</i>	Biohydrogen	Sweet sorghum	Shanmugam et al. (2018)
Consortium of <i>T. viride</i> and <i>A. niger</i>	Biohydrogen	Oat straw	Zhao et al. (2019)
<i>A. tubingensis</i> , <i>Trichosporono idesspathulata</i> , <i>Candida tropicalis</i> , <i>Rhodotorula mucilaginosa</i>	Biodiesel	Palm empty fruit bunch	Intasit et al. (2020)
<i>Mucor circinelloides</i>	Biodiesel	Sugarcane bagasse, corn milling	Carvalho et al. (2018)
<i>Penicillium citrinum</i>	Biodiesel	<i>Musa balbisiana</i> cola peels	Bardhan et al. (2019)
<i>Aspergillus awamori</i> , <i>Aspergillus oryzae</i>	Biohydrogen, Bioethanol	Food waste	Han et al. (2016)
<i>Gymnopus contrarius</i>	Biohydrogen	Rice straw	Sheng et al. (2018)
<i>Clostridium thermocellum</i>	Biohydrogen	Waste date palm	Swathy et al. (2020)
<i>Pleurotus ostreatus</i> , <i>Trametes versicolor</i>	Biogas	Chicken manure with sawdust and wheat straw	Basinas et al. (2022)
<i>Orpinomyces</i> sp., <i>Piromyces</i> sp., <i>Anaeromyces</i> sp., <i>Neocallimastix frontalis</i>	Biogas	Animal manure	Yıldırım et al. (2017), Bhujbal et al. (2022)
<i>Cladosporium</i> sp., <i>Verticillium</i> sp.	Biogas	Feathers, biological sludgeslime	Wrońska and cybulska (2018)

improve the mixing and mass transfer performance, compared to traditional high-cost centrifugation methods.

Oleaginous yeast such as *Rhodotorula glutinis* accumulates 25% lipid of its biomass for biodiesel production from monosodium glutamate wastewater (Zheng et al. 2012). *Saccharomyces cerevisiae* can use hexose monosaccharides (glucose, mannose, and galactose) and disaccharides (sucrose and maltose) to produce bioethanol via fermentation of lignocellulosic hydrolysates (Branco et al. 2019). Yeast strains such as *Kluyveromyces fragilis*, *Candida* sp., *Rhodospiridium* sp., *Rhodotorula* sp., and *Lipomyces* sp. accumulate 70% triacylglycerols of their biomass (Subhash and Mohan 2011). Hemicellulose and lignins of plant cell walls are acetylated, which yield acetic acid after hydrolysis as an unavoidable component. Acetic acid is toxic to the fermenting microorganisms, negatively influencing sugar fermentation and, subsequently, biofuel yield. Additionally, *Trichosporon fermentans* could be used for microbial lipid production from detoxified rice straw acid hydrolysate. But the obtained lipid content was lower than glucose as the sole carbon source (Huang et al. 2012). Yeast, *Saccharomyces cerevisiae*, is widely used for the production of ethanol from corn and sugarcane, but it cannot metabolise

xylose. But *Scheffersomyces stipitis* can convert xylose to xylulose by expression of nicotinamide adenine dinucleotide phosphate (NAD(P)H)-linked xylose reductase (XR) and nicotinamide adenine dinucleotide (NAD)-linked xylitol dehydrogenase (XDH) genes. This xylulose can be metabolised after its phosphorylation via the pentose-phosphate pathway (Wei et al. 2013). Moreover, endophytic fungal isolates *Colletotrichum* sp., *Alternaria* sp., and *Aspergillus* sp. have the ability of lipid accumulation, as whole-cell biocatalysts, under the nutrient optimum and nutrient-stressed conditions (Subhash and Mohan 2011).

Biogas production efficiency is influenced by the type and quality of the raw materials used. Waste products from the poultry industry, agricultural crop wastes, and animal residues fulfil the requirements of good raw materials due to having a significant proportion of fats and proteins (Wrońska and Cybulska 2018). Anaerobic fungi are known to produce plant carbohydrate hydrolysing enzymes for cell wall polysaccharide decomposition. Anaerobic fungi are promising candidates for mechanical and enzymatic degradation of plant polysaccharides to improve biogas production (Dollhofer et al. 2015). Anaerobic fungus *Piromyces rhizinflata* degrades volatile fatty acid and augments the lignocellulose biomass (corn silage and cattail) as feedstock for CH<sub>4</sub> and H<sub>2</sub> production (Nkemka et al. 2015). Similarly, the fungus *Auricularia auricula-judae* is used to decay sweet chestnut (*Castanea sativa*) leaves, hay and wood to decompose cellulose, hemicelluloses and lignin for the production of biogas (Mackulak et al. 2012).

### 7.4.3 Photosynthetic Microorganisms

Photosynthetic microorganisms, as a platform for biofuel production, have gained substantial recognition as an option that could significantly reduce environmental pollution by using CO<sub>2</sub> emitted from various sources (Machado and Atsumi 2012). These photosynthetic microorganisms directly fix CO<sub>2</sub> as their primary carbon source for biofuel production and replace the requirement of fermentable sugars. Algae and cyanobacteria are the pioneer and desired organisms for this strategy of biofuel production. Both these groups of organisms can grow much faster than plants, do not need arable land for their production and can be grown in submerged water (Dismukes et al. 2008). Research on algae has centred on enhancing their potential to produce large amounts of lipids pertinent to biodiesel production (Pate et al. 2011; Kumar et al. 2017). Cyanobacteria coupled with prokaryotic organisms such as *E. coli* is beneficial to both as a photosynthetic microorganism and naturally transformable host. Studies reveal that cyanobacteria have already been manipulated to produce a number of different biofuels (Dismukes et al. 2008; Machado and Atsumi 2012; Gao et al. 2016). For instance, *Synechococcus elongatus* sp. strain PCC 7942 was successfully manipulated for ethanol production via the external addition of enzymes such as pyruvate decarboxylase and alcohol dehydrogenase, redirecting the carbon from pyruvate (Deng and Coleman 1999). Continuous research works have significantly improved the production of ethanol using cyanobacteria (Gao et al. 2012, 2016). Further researches are being conducted

worldwide on other photosynthetic microorganisms to improve and strengthen the ability of biofuel production from waste.

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## 7.5 Waste Product Impact on Climate

Wastes are all the by-products released from industries, households, or other sources humans cannot use further. Waste management is a more significant challenge for both the small and big cities of developing countries. Urbanisation and increasing population are the major issues responsible for increasing the burden of waste. According to the Global Waste Management Outlook 2015 (GWMO), 2.0 billion tonnes/year of waste is produced by MSW and 7–10 billion tonnes from households, commerce, industries and construction site (Everett 2012; Al-Dhrub et al. 2017). These wastes may be in solid, liquid or gaseous forms whose disposal improperly leads to negative consequences on the health of humans, animals and the environment (Misra and Pandey 2005). Improper and uncontrolled disposal generates heavy metal pollution in the water, air, and soil. Open burning causes the release of CO<sub>2</sub>, SO and other air pollutants in the atmosphere. The release of waste in the water bodies also affects the aquatic ecosystems enhancing eutrophication (Ferronato and Torretta 2019). In the present climate change scenario, the melting of glaciers, increasing temperatures, seasonal variations, the emergence of various pathogens, and adverse consequences on agricultural production are the major threats to human society. Further, these wastes and their mismanagement will boost the future climate change rate. Nowadays, the conversion of different waste materials to generate energy and its use for societal welfare along with a significant positive impact on the environment is one of the top priorities (Tabasová et al. 2012; Kumar et al. 2019b). These strategies are required to control the rate of climate change and mitigate its adverse consequences.

Due to recent anthropogenic activities, the degree and amount of waste are increasing. The considerable increase in a waste generation began due to population explosion and industrialisation (Wilson 2007; Pikoń and Czop 2014). It has been reported that approximately 1.3 billion tonnes of MSW is generated per year, and it could rise to approximately 2.2 billion tonnes/year by the end of 2025 (Hoorweg and Bhada-Tata 2012). There are various waste management techniques through which the wastes can be transformed for the production of manures for agriculture purposes, eco-friendly energy sources, and pollution reduction (Widmer et al. 2005; Aljaradin and Persson 2012).

### 7.5.1 Impacts of Waste Disposal on the Environment

The waste material could be in solid, liquid or gaseous form and biodegradable or Non-biodegradable in nature. Food production through agriculture and its consumption is one of the main factors related to environmental impacts in the world. Food production involves using resources such as fuels, land, water and raw materials

linked to economic and environmental impacts. Most food packaging materials are made up of non-biodegradable plastics which are obstinate towards microbial disintegration and hence do not meet the requirements of compost forming (Pikoń and Czop 2014). Disposal of food wastes into water bodies affects the aquatic ecosystem, causing eutrophication and algal blooms due to increased nutrient concentration in water bodies (Scherhauser et al. 2018).

In developing nations, there is a major problem with management of solid waste (sewage and industrial sludge) due to several constraints; hence, landfilling with waste products in low-level areas is preferable. Sewage contains a large number of toxic substances which are harmful to human and animal health, as well as to the environment. MSWs majorly hold solid matter and are subject to landfilling for its management. The degradation of MSWs in landfills leads to the formation of different hazardous gases. The level of CO<sub>2</sub>, which usually remains high, regularly drops as the CH<sub>4</sub> concentration builds up if the degradation procedure is shifted from aerobic to anaerobic conditions. Other gases, including H<sub>2</sub>, nitrogen, etc., are produced in minor amounts during the degradation process. Burning solid waste at the landfilling site produces toxic gases that pollute the air, causing respiratory problems. These gases contribute to global warming and climate change. Solid waste undergoes a sequence of complex biochemical and physical processes, leading to the production of leachate and gaseous emissions. When leachates reach the water resources, they pollute surface water and groundwater (Aljaradin and Persson 2012).

### 7.5.2 Non-biodegradable Wastes

Hazardous and non-biodegradable solid wastes, which enter from the municipal waste directly disposed-off in the environment, play a significant role in environmental degradation. The majority of plastics are composed of polyaromatic hydrocarbon compounds and produce greenhouse gases, which cause a negative impact on the environment. Plastic restricts the water absorption in the soil due to seized soil capillaries and simultaneously affects the microbial diversity, water holding capacity, and loss of moisture content in the soil. More plastic waste in the soil environment triggers the process of soil infertility (Andreeßen and Steinbüchel 2019). Now a day's, the world is facing plastic waste pollution in the marine ecosystem also. Rivers are the indirect key carrier of plastic waste. Plastic waste harms many aquatic animals, and plastic pollution also decreases the aesthetic value of any water body.

The waste of glass industries is another unremarkable waste posing many challenges due to the high greenhouse gas emissions, rigorous energy use, and the intensive use of the Earth's natural resources. Discarding the glass waste in landfills is not offering environment-friendly management due to the non-biodegradable nature of glass waste and is triggering severe environmental soil pollution (Jani and Hogland 2014). Apart from municipal or industrial waste, E-waste comprises harmful materials that need proper management and recycling approaches to avoid environmental pollution (Gabra et al. 2019). E-waste is chemically and physically different from other forms of waste. The chemical composition of E-waste differs

depending on the age and quality of the discarded items. Most E-wastes contain a mixture of metals, particularly Cu, Al, and Fe, which are used in several kinds of plastics and ceramics. Discarded personal computers, laptops, washing machines, refrigerators and electrical wires are comprised of metal, plastics, electronic components and glass. Disposing of all this E-waste in the environment is polluting the water, soil, and air (Robinson 2009).

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## 7.6 Challenges in Biofuels Production from Waste

World socio-economic developments are mainly progressed by energy. Presently, the world's fuel demand of around 75% is compensated by non-renewable sources like petroleum and its derived fuel. As per the International Energy Report 2014, the global energy demand is expected to elevate by 37% by 2040 (Joshi et al. 2017). Therefore, research is being carried out in different parts of the world with a special focus on renewable sources to meet anticipatory growing energy demand. Hence, biofuels from waste biomasses could be a probable source to meet the global anticipatory energy demand.

There are several procedures and technologies by which renewable resources can generate biofuels (Joshi et al. 2017). The biofuels could be produced from enriched biochemicals produced by either microbiological agents such as bacteria, fungi, and microalgae or animals (Rodionova et al. 2017; Kumar and Banerjee 2019). For the last few decades, agriculture production has increased several folds. Simultaneously, food and agricultural waste also increased proportionally; hence, this waste production has been known to be the potential source of biofuels. However, algal biomass has recently been known to be a potential bioresource for producing different types of biofuels (Dragone et al. 2010; Rodionova et al. 2017).

There are several prospects for the production of biofuels from wastes product that have been well recognised and exploited. Among them, biofuels by cyanobacteria or microalgae have been highly acknowledged (Demirbas et al. 2016; Heimann 2016; Rodionova et al. 2017; Chintagunta et al. 2020). Scott et al. (2008) have reported several benefits of using microalgae for biofuel production owing to high productivity compared to other bioresources. Besides the benefits of microalgae-based biofuels production, several challenges are still to be considered for commercial production of biofuels, such as ease and continuous accessibility of waste products, pre-treatment and processing of waste products that could be subjected to biofuel production. Appropriate selection of bioreactors for large-scale production of microalgal biomass, maintenance of contamination-free medium during the reaction, selection of superior microalgae strains and most important continuous supply of sterile medium as well as CO<sub>2</sub> for microalgae growth are the other aspects that need optimisations (Scott et al. 2008).

Food waste is the anon consumable source of lipids, carbohydrates, amino acids and phosphates. On average, food waste materials contain around 30% lipid and 50% carbohydrate (Pleissner et al. 2014, 2016). The waste food can be hydrolysed enzymatically, and the food wastes abundant in carbohydrates and lipids can be

subjected to bio-ethanol and biodiesel production, respectively. In the past few decades, focused research on the application of food wastes for producing biofuels has been going on globally. Sulaiman (2014) proposed a halal biorefinery to produce biofuels in Malaysia. Chinese Academy of Sciences reported using food waste to produce hydrolysates for bioethanol production (Yan et al. 2011; Karmee and Lin 2014). In Europe, potato peel has been utilised to produce bioethanol using environmentally benign biocatalytic methods with the involvement of liquefaction, saccharification and fermentation of peel (Arapoglou et al. 2010; Yan et al. 2011; Wang et al. 2017). The prime drawback of pre-treatment methods of waste products included the production of specific inhibitors for microbes that may interfere with the processing and production of biofuels. These inhibitors are formic acid, acetic acid, phenolic compounds, furan aldehydes, ionic lipids, and levulinic acid (Wang et al. 2018; Zhang et al. 2016).

Recent economics estimates that the costs of biofuel production from waste are 2–3 folds more expensive than petroleum fuels on an energy-equivalence basis (Lynch et al. 2016; Bušić et al. 2018). To lower the production cost of biofuel, several challenges are to be taken into consideration while converting waste biomass to biofuels, such as feedstock production, feedstock logistics, development of energy-efficient technologies (pre-treatment, enzyme hydrolysis, and microbial fermentation), separation of by-products (lignin and hemicelluloses), product development, the establishment of biofuel and biochemical standards, biofuel distribution and environmental impact minimisation. Some of the major drawbacks of pre-treatment procedures include the generation of by-products that works as inhibitors for microbial growth and fermentation. These compounds are formic acid, acetic acid, and levulinic acid (Wang et al. 2018; Zhang et al. 2016). The acetic acid in growing media potentially reduces the specific growth rate and biomass yield of *Saccharomyces cerevisiae* during ethanol production waste biomass (Pampulha and Loureiro-Dias 2000; Wang et al. 2018).

Similarly, phenolic compounds, furan aldehydes and ionic lipids also act as inhibitors to *S. cerevisiae* by decreasing specific cell growth rate and ethanol yield (Lin et al. 2015; Banerjee et al. 2019). All these constraints for biofuel production from wastes require high skill in agronomy, biomass logistics, biomass conversion, process engineering, chemistry, conversion technology, genetic engineering, microbial fermentation, economics, and environmental science (Rai et al. 2020; Kumaraswamy and Kashyap 2021). It is challenging to produce biofuel from waste and economically expensive over fossil fuel. However, developing recombinant strains through genetic engineering with high commercial potential, redefining effective pre-treatment processes, and increased access to waste bioresources could be a promising strategy for sustainable biofuel production.

## 7.7 Conclusion and Future Prospects

Presently, developed and developing nations are encountering several challenges pertinent to climate change, depletion of natural resources, environmental sustainability and energy security, and all of these directly or indirectly affect the environment. Hence, biofuels are supposed to be the most important to alleviate such energy crises sustainably. Furthermore, several biofuels of various classes could be produced from available indigenous resources and waste products generated from agriculture and food processing. Biomass generated as waste after processing agriculture and food is a potential feedstock for biofuel production. These biomasses are potentially converted into several biofuel products through the application of different microbes of the different genera (bacteria, fungi, and photosynthetic microbes). However, biofuel productions from waste products also have several constraints that must be overcome with an integrated application of technological advancement pertinent to strain improvement, adoption of improved protocol for pre and post-processing of biomasses, and control of microbial inhibitors to improve the yield and quality of biofuels. A combination of all these approaches and further researches in the area are expected to provide remedies for the existing energy crisis due to the depletion of non-renewable sources.

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# Role of Microorganisms in Biogas Production from Animal Waste and Slurries

# 8

Najib Lawan Yahaya, Mudassir Lawal, Abhishek Kumar Verma, Sudhir K. Upadhyay, and Ali Asger Bhojiya

## Abstract

Energy supply and waste management are two of the great challenges that humanity as a whole faces. The world's energy supply is mainly dependent on fossil fuels whose combustion leads to excessive carbon dioxide emission, which, when released into the atmosphere in greater concentration, causes global warming. Moreover, the amount of solid waste produced is increasing and is expected to grow rapidly in the next decades. Therefore, to meet these challenges in the future, it is necessary to use life-cycling technology as a robust tool capable of combatting environmental waste into energy. It is becoming apparent that the majority of organic waste from various agricultural and industrial sources can be converted by microorganisms into biofuels. These biofuels provide renewable energy sources that could significantly lower greenhouse gas emissions and ensure sustainable waste management. The concept of bioenergy production from waste has developed significantly over the last few decades. Biogas is among the gaseous biofuels produced by the anaerobic digestion of organic material, and recently, its production from animal waste such as cow dung is an economically viable way to reduce environmental pollution and provide an opportunity for effective waste management and production of valuable products. Biogas consists of mainly methane (CH<sub>4</sub>), carbon dioxide (CO<sub>2</sub>), and small amounts of hydrogen sulfide (H<sub>2</sub>S). This chapter focuses on the production of

N. L. Yahaya · M. Lawal · A. K. Verma

Department of Life Sciences, Faculty of Science and Technology, Mewar University, Chittorgarh, Rajasthan, India

S. K. Upadhyay

Department of Environmental Science, V.B.S. Purvanchal University, Jaunpur, Uttar Pradesh, India

A. A. Bhojiya (✉)

Department of Botany, U.S. Ostwal P.G. College, Chittorgarh, India

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biogas from animal wastes. The chapter will provide an overview of the concept of biogas production, microorganisms used in the production of biogas, the anaerobic digestion process, and the anaerobic digester. The chapter will also attempt to highlight the key stages involved in biogas production (hydrolysis, acidogenesis, acetogenesis, and methanogenesis), and the benefits of biogas. Details of factors influencing the production of biogas are also discussed.

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**Keywords**

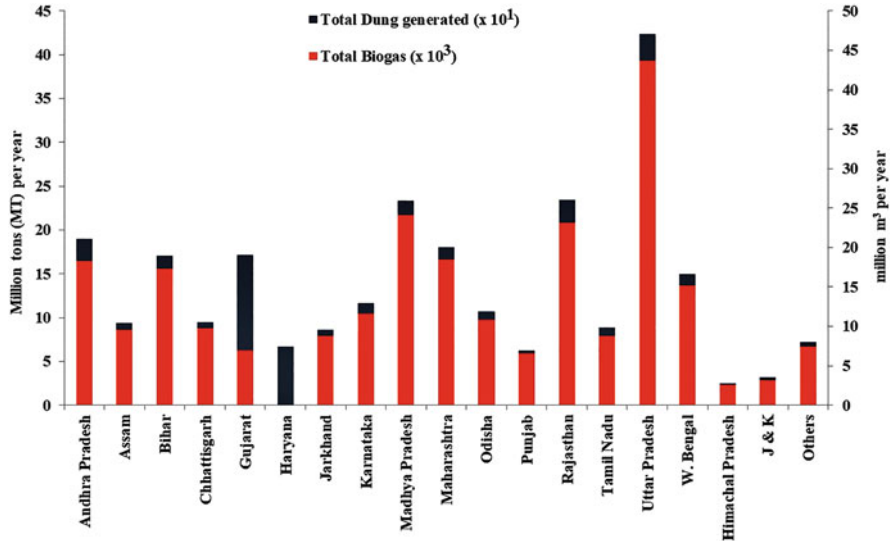
Global warming · Biofuels · Anaerobic digestion · Environmental pollution · Biogas

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## 8.1 Introduction

The present world energy supply is largely dependent on fossil-origin fuel such as petroleum, coal, and natural gas, etc. They are the ossified remains or impressions of dead plants and animals, which have been preserved in the Earth's crust for millions of years. Utilization of such resources converts carbon stored for millions of years into carbon dioxide (CO<sub>2</sub>), and its release into the atmosphere in greater concentrations causes global warming. For this reason, fossil fuels are non-renewable energy sources. One of the main threats to society today is the continuous increase in organic waste production. Therefore the task of waste management and inadequate energy supply are two of the enormous problems that are increasingly threatening the life of many people (Onwuliri et al. 2013). Sustainable management of waste as well as avoiding and reducing waste have become major priorities, representing a significant part of the public efforts to reduce pollution and greenhouse gas emissions and mitigate global climate changes.

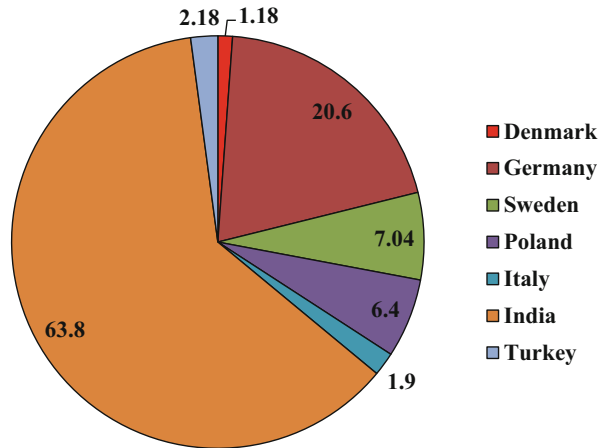
The decrease in the production of non-renewable energy sources along with the climate change problem has driven the search for renewable and more environmentally friendly energy sources as an alternative to fossil fuels which allow for sustainable development, as the system seems auspicious to achieve sustainable energy production without destructing our environment (Chojnacka et al. 2015). It is therefore important to implement a renewable energy system to replace fossil fuels. Research has shown that biogas is one such alternative energy source, particularly for the rural community (Raja and Wazir 2017). In contrast to fossil fuels, biogas is renewable energy as it is produced from biomass. Biogas will not only upgrade energy stability but also make a significant influence on the conservation of natural resources and environmental protection. It will increase the security of the energy supply, reduce dependency on fossil fuels and help to ensure sustainable development. Govarathanan et al. (2022) reviewed critically various research works and suggested that utilizing lignocellulosic (LC) biomass generates biogas at a high rate and also nanotechnology intervention was found to be very effective in biogas production (Yadav et al. 2020).



**Fig. 8.1** Quantifiable sources of livestock dung (MT per year) and potential for biogas generation (million m<sup>3</sup> per year) in India. (Adapted from Kaur et al. 2017)

Biogas is a promising renewable alternative to natural gas with similar applications. It is typically a mixture of different gases which primarily comprises methane (CH<sub>4</sub>), carbon dioxide (CO<sub>2</sub>), a small amount of hydrogen sulfide (H<sub>2</sub>S), moisture (H<sub>2</sub>O), and a few other gases formed in the absence of oxygen due to the breakdown of organic material (Liaquat et al. 2017). Nearly any organic waste materials can be biologically degraded and transformed into biogas and other energy-rich organic compounds by the process of anaerobic digestion, thereby enabling sustainable waste management (Goswami et al. 2016). Production of biogas through anaerobic digestion of animal waste converts these wastes into renewable energy. Biogas production from animal waste is an economically feasible way to reduce environmental pollution and produce valuable products, i.e., methane (Pampillón-gonzález et al. 2017). It is a very important renewable source of energy produced from organic materials like cattle dung, human waste, and different types of biomass. Therefore, biogas is a renewable energy source as it is wholly energy self-sustenance technology, independent of any fossil fuel, and reduces greenhouse gas emissions into the environment. State-wise generation of animal dung and the tentative theorized estimate of this untapped source for biogas production in India are shown in Fig. 8.1. The annual production of dung is estimated to be approximately 2600 million tons (MT), which is enormous in terms of volume, making it an important untapped energy source. The total dung generated which is mentioned in Fig. 8.1 comprises of large animal dung, small animal dung, pig dung, and poultry dung. Total potential biogas production from all dung sources was calculated in terms of annual yield measured in million m<sup>3</sup> per year.

**Fig. 8.2** Biogas potential of different countries (in billion m<sup>3</sup>). (Source: Karaca 2018)



When the biogas potentials of some other countries are examined, it is seen that India has good potential (Fig. 8.2).

## 8.2 Anaerobic Digestion and Biogas Production

Anaerobic digestion (AD) is a natural biological process whereby organic matter is decomposed and transformed by microorganisms into biogas in the absence of oxygen (Fedailaine et al. 2015). During the process, microorganisms digest plant and/or animal material in sealed containers, producing biogas. The process occurs in an anaerobic environment (oxygen-free environment) through the activities of a diverse group of microorganisms that break down the organic material and produce methane (CH<sub>4</sub>), carbon dioxide (CO<sub>2</sub>) in a gaseous form known as biogas, and other nutrient-rich compounds (Kythreotou et al. 2014). It is a complex process that involves two stages. At the initial phase of the process, degradation is executed by fast-growing, acid-forming microbes (acidogenic), where protein, carbohydrate, lipids, cellulose, and hemicellulose in the waste are hydrolyzed and metabolized into organic acids and volatile fatty acids (VFAs), along with carbon dioxide and hydrogen gases. At this stage, the decomposing products have noticeable, disagreeable, effusive odors from the organic acids, H<sub>2</sub>S, and other metabolic products (Liaquat et al. 2017). In the second phase of the process, most of the organic acids and other intermediary products of the earlier phases of the process are metabolized by methanogenic microorganisms, thereby producing biogas as the end-product, which comprises a mixture of different gases, as shown in Table 8.1.

Biogas production through anaerobic digestion (AD) is an environmentally friendly technology for bioenergy production utilizing the increasing amounts of organic waste produced worldwide. A wide range of waste streams, including industrial waste, domestic waste, human excreta, municipal wastewater, agricultural waste, animal waste as well as plant residues, can be treated with this technology. It

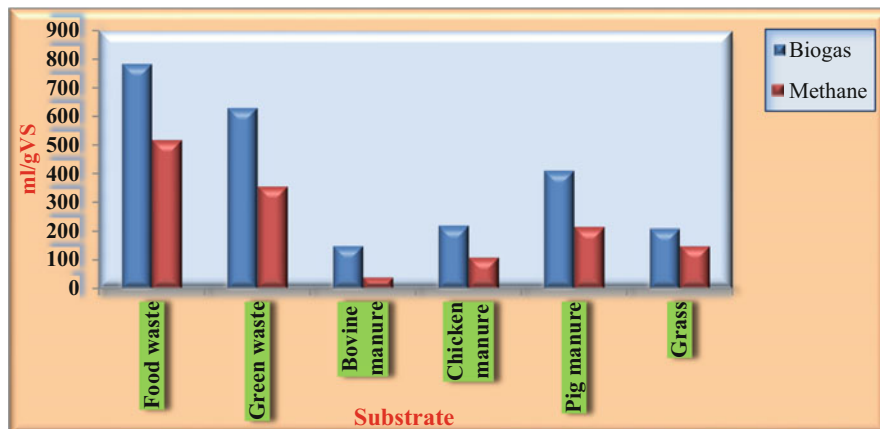
**Table 8.1** Typical percentage composition of biogas (Liaquat et al. 2017; Schnurer and Jarvis 2009)

S. no.	Biogases	Formula	Percentage %
1.	Methane	CH <sub>4</sub>	50–75
2.	Carbon dioxide	CO <sub>2</sub>	25–50
3.	Nitrogen	N <sub>2</sub>	0–10
4.	Oxygen	O <sub>2</sub>	0–2
5.	Hydrogen	H <sub>2</sub>	0–1
6.	Hydrogen sulphide	H <sub>2</sub> S	Traces
7.	Water vapor	H <sub>2</sub> O	Traces
8.	Ammonia	NH <sub>3</sub>	0–0.05

is an effective process to convert animal waste into profitable by-products as well as reduce the pollution of air, water, and soil caused by these wastes. The organic material in animal waste is easily decomposable, so a lot of microorganisms thrive in it. These microbes are mostly anaerobic and thus ideally suited to decompose the organic material in an anaerobic digester and produce biogas (Pampillón-González et al. 2017). The production of biogas through this process proffers significant benefits over other systems of bioenergy production and many other waste treatment processes. The major product of this process, i.e., the biogas, is a renewable energy source, while the by-product, i.e., the digester residue, can be used as a biofertilizer because of its high nutrient content available (Horváth et al. 2016). Biogas production is influenced by the amount of organic material and the number of anaerobic bacteria that degrade the organic material (Hidayati et al. 2018). Therefore, the quantity and quality of the biogas appear to be controlled by the type of biomass being digested and the microbial inoculum fed into the biogas plant. Biogas can be generated from nearly all types of biomass; nevertheless, animal waste and slurries represent one of the largest resources. Animal waste and slurries from cows, pigs, sheep, goats, and poultry have been estimated as among the major waste streams for biogas production, which, if left unprocessed or inadequately managed, may become a major environmental problem because of nutrient leaching (N, P), ammonia evaporation, and pathogen contamination. Among animal waste, it has been reported that pig manure produces a high yield of biogas and methane compared to other animal waste, as shown in Fig. 8.3 (Enzmann et al. 2018; Verma et al. 2018).

The purpose of using anaerobic digestion is usually related to waste management and energy production. The remaining digestate is an added benefit, which creates additional value. Hence, the practice of anaerobic digestion can assure appropriate waste management, production of biofertilizers, and improved environmental impact and sustainability (Luo et al. 2013). Anaerobic digestion (AD) technology is widely used in the treatment of organic wastes to achieve the reduction of the wastes with the simultaneous production of biogas, the technology allows the treatment of high organic loading wastes to reduce their volume and load while recovering biogas, which can be used to produce heat, electricity, and or upgraded to be biofuels for automotive vehicles (Awe et al. 2017; Madakka et al. 2020).

The anaerobic digestion technology has gained considerable momentum over a few years and it is considered a valuable technology for the production of renewable



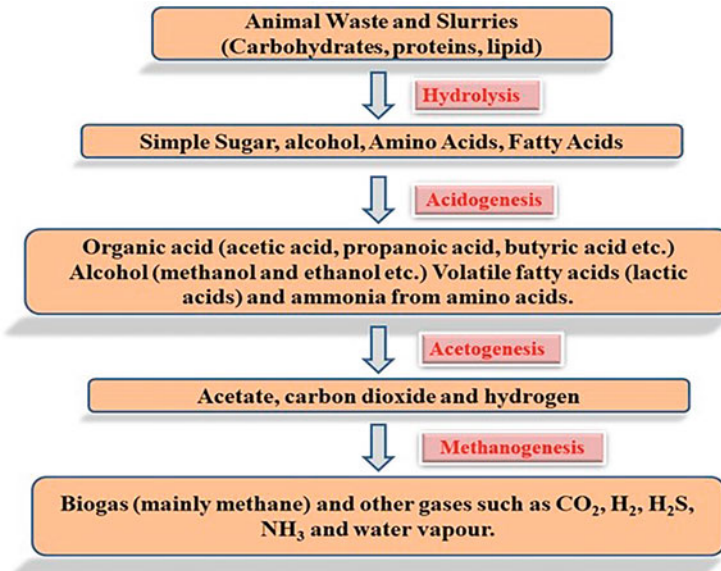
**Fig. 8.3** Biogas and methane contents of some organic waste in milliliter per gram volatile solids (mL/gVS) (Heo et al. 2003)

energy and offers a way to mitigate problems related to low access to energy (Anukam et al. 2019; Náthia-Neves et al. 2018).

The systems have undergone various modifications in the last decades to increase the efficiency of the process. An important milestone was the development of a new reactor design, i.e., the up-flow anaerobic sludge blanket (UASB) reactor, containing a well-settleable methanogenic sludge due to the formation of a dense sludge bed. Another technology making it possible to retain active biomass within the system was the application of membrane bioreactors (MBRs), which can also be utilized for the parting of inhibitory substances, which otherwise would negatively disturb the biological process (Mainardis et al. 2020). Additionally, advances in molecular biology techniques could provide scientists and students with a valuable tool to understand the complex microbiological processes involved in the anaerobic digestion of organic materials. By the application of these techniques, it would be possible to regulate and control the process and discover disturbances much earlier than using traditional process parameters for monitoring the process.

### 8.3 Stages of Biogas Production by the Anaerobic Digestion Process

Biogas production through anaerobic digestion (AD) of organic materials is the combinative activity of various microbial populations carried out by several different groups of bacteria and fungi such as hydrolyzing, acidifying, acetogenic, and methanogenic microbes, which in the final stage produce biogas mainly methane ( $\text{CH}_4$ ) and carbon dioxide ( $\text{CO}_2$ ) (Heeg et al. 2014). The production of biogas is usually carried out in four biological and chemical stages, i.e., hydrolysis, acidogenesis, acetogenesis, and methanogenesis. These four main stages account



**Fig. 8.4** Key steps of biogas production

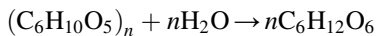
for the production of biogas from different organic matter as it takes place in an anaerobic reactor (Fig. 8.4). In the single-stage batch reactor, all wastes are loaded simultaneously, and all four processes are allowed to occur in the same reactor sequentially; the compost is then emptied at the end of a given retention period or cessation of biogas production (Kwietniewska and Tys 2014).

### 8.3.1 Hydrolysis

Hydrolysis is the first step in biogas production. In this step, the complex organic matter (polymers), that is, proteins, carbohydrates, and lipids (fats) are broken down and transformed into simple and smaller water-soluble compounds such as amino acids, fatty acids, and simple sugars, which in turn can be utilized by acidogenic bacteria (Chandra et al. 2012). During the hydrolysis process, hydrolytic bacteria present in the reactor secrete extracellular enzymes that convert complex organic substrates containing carbohydrates, lipids, and proteins into sugars, long-chain fatty acids, and amino acids, respectively (Li et al. 2011). However, certain substrates, such as lignin, cellulose, and hemicellulose, may find it difficult to degrade, and can be inaccessible to microbes due to their complex structures; enzymes are often added to enhance the hydrolysis of these carbohydrates (Lin et al. 2010).

From a chemical perspective, hydrolysis refers to the cleavage of chemical bonds by the addition of water. Cations and anions react with water molecules, altering pH

in the process to create a cleavage of H–O bonds. The reaction associated with this step is given below:

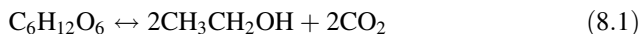


From the reaction, the hydrolysis of cellulose ( $C_6H_{10}O_5$ ) via the addition of water ( $H_2O$ ) to form glucose ( $C_6H_{12}O_6$ ) as the primary product and gives off  $H_2$ . The reaction is catalyzed by homogeneous or heterogeneous acids to produce glucose ( $C_6H_{12}O_6$ ) (Zupancic and Grilc 2007). Hydrolysis is the slowest step of biogas production, especially when solid waste substrates are used. The process rate depends on factors such as pH, particle size, enzyme production, diffusion, and enzyme adsorption on waste particles that are exposed to the degradation process. The magnification of the hydrolysis process increases the performance of digestion (Yu et al. 2016). Biological, chemical, and mechanical pre-treatments, or a combination of these can be used to accelerate hydrolysis, because they can cause lysis or disintegration of the substrate and allow the release of intracellular matter, allowing greater accessibility of anaerobic microorganisms, thus reducing the retention time in the digester (Ferrer et al. 2008).

### 8.3.2 Acidogenesis

This is the second stage of biogas production, where the products of the hydrolysis (water organic monomers of sugars and amino acids) are further broken down and converted mostly into several organic acids (acetic acid, propionic acid, butyric acid, succinic acid, pentanoic, etc.), VFAs (lactic acid), alcohols (methanol, ethanol), and ammonia (from amino acids) (Christy et al. 2014). Acidogenesis is usually the fastest step of biogas production and occurs due to the action of acidogenic fermentative microorganisms. With the rapidity of this stage, it is important to note that while the production of VFAs creates direct precursors for the final stage of methanogenesis, VFA acidification is widely reported to be a cause of digester failure (Akuzawa et al. 2011).

The exact compounds to be formed depend on the substrate and process conditions, as well as the microorganisms available. Studies have shown that volatile fatty acid concentrations can vary significantly for digesters operating at different pH, with different studies presenting seemingly contradictory results (Huang et al. 2015). The important acid in this stage is  $CH_3COOH$ , and it is the most significant organic acid used as a substrate by  $CH_4$ -forming microorganisms. Whereas the production of volatile fatty acids (VFAs) is increased when the process pH is greater than 5, the production of ethanol ( $C_2H_5OH$ ) is favored by a low pH value of less than 5 with the reaction process coming to a halt at a  $pH < 4$  (Bajpai 2017). Eqs. (8.1)–(8.3) present the reaction sequence that summarizes the acidogenic stage of biogas production (Barua and Dhar 2017).



Acetates,  $\text{CO}_2$ , and  $\text{H}_2$  pass through the basic pathway of transformation, while other products of acidogenesis play an insignificant role. As a consequence of these transformations, the new products may be directly used by methanogenic microbes as substrates and energy sources. This stage is very significant because it links the phase of fermentation with the phase of production of methane. Thus, more acid is produced to form elements of methanogens that generate methane gas (Ntaikou et al. 2010).

### 8.3.3 Acetogenesis

Acetogenesis is the third stage of biogas production. It is the process where acetogens produce acetate (a derivative of acetic acid) utilizing carbon and energy sources. In this phase, acetogenic microbes convert the compounds generated during the acidogenic phase, producing hydrogen, carbon dioxide, and acetate (Chandra et al. 2012). Acetogenic microbes digest the biomass to an extent from which, methanogens utilize it as a substrate to produce biogas (methane). This stage explains the efficiency of the production of biogas as, in the process of acetate reduction, more than 70% of  $\text{CH}_4$  is generated. Subsequently, acetate is the main intermediate product of the process of methane production (Gkamarazi 2015).

The stage involves coordination between the oxidizing microbes and the methanogenic microbes that are active in the next phase of the methane-producing process (Heeg et al. 2014). The reaction associated with this stage of AD is represented by Eqs. (8.4)–(8.6) (Anukam et al. 2019).



### 8.3.4 Methanogenesis

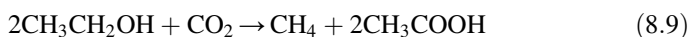
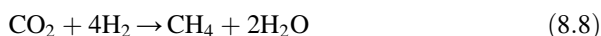
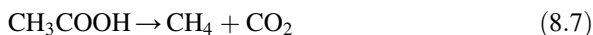
Methanogenesis is the final stage of the biogas production process. In this process, methanogens generate biogas from the end products of acetogenesis which consists mainly of methane ( $\text{CH}_4$ ) and carbon dioxide ( $\text{CO}_2$ ), but also comprises some other gaseous “impurities” such as hydrogen sulphide ( $\text{H}_2\text{S}$ ) (easily detectable by its smell



of rotten eggs), nitrogen, oxygen, and hydrogen (Chojnacka et al. 2015). The actual process of methanogenesis is very complex and needs explicit substrates and cofactors, the major substrates used are acetate, carbon dioxide, H<sub>2</sub>, formic acid, methanol, methylamine, and dimethyl sulfide. But two substrates, carbon dioxide and acetate, are the most commonly used (Costa and Leigh 2014). The pathway which precedes methane production exclusively depends on the methanogenic microbes and the availability of the substrate that favors the degradation process. Generally, there are six pathways of methanogenesis, each converting a different substrate into methane gas. The three major pathways are:

1. Hydrogenotrophic methanogenesis (production of methane by the reduction of H<sub>2</sub>/CO<sub>2</sub>)
2. Acetotrophic methanogenesis (production of methane by acetate decarboxylation)
3. Methylotrophic methanogenesis (production of methane by removal of the carboxyl group of methyl alcohols, methyl amines, etc.), (Slonczewski and Foster 2013)

The acetotrophic pathway is the main pathway of methane production in the anaerobic digestion process as 70% of the total methane generated during the process is through this pathway (Merlino et al. 2013), and the most commonly used pathway is hydrogenotrophic methanogenesis, which transforms carbon dioxide into methane by reduction of H<sub>2</sub>/CO<sub>2</sub> (Slonczewski and Foster 2013). The reaction equation representing the condition taking place in the methanogenesis step is represented by the following (Ostrem 2004):



The first Eq. (8.7) shows the conversion of CH<sub>3</sub>COOH into CH<sub>4</sub> and CO<sub>2</sub>. The CO<sub>2</sub> formed is reduced to CH<sub>4</sub> through H<sub>2</sub> gas in the second Eq. (8.8) and, lastly, Eq. (8.9), shows the production of CH<sub>4</sub> by decarboxylation of CH<sub>3</sub>CH<sub>2</sub>OH.

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## 8.4 Anaerobic Digesters

Anaerobic digesters are vessels in which a biochemical process is carried out and involve organisms or biologically active substances derived from such organisms.

Three basic types of digesters that have been executed in developing nations are floating-drum digester, fixed-dome digester, and tubular digester, all of which are wet digestion systems worked uninterruptedly under mesophilic conditions. These three types are easy to handle, low-cost, built with nearby available material, do not

have numerous moving parts and are thus less predisposed to failure. An additional digester type, the garage-type digester, which is worked as a dry digestion system in batch-mode, is considered another potential biogas technology suitable for low- and middle-income countries. Although this technology is being tested in some African countries like Ghana by converting a used shipping container, it is not yet ready for the commercial market as no viable low-cost design exists that has been successfully tested at full-scale (Vögeli et al. 2014).

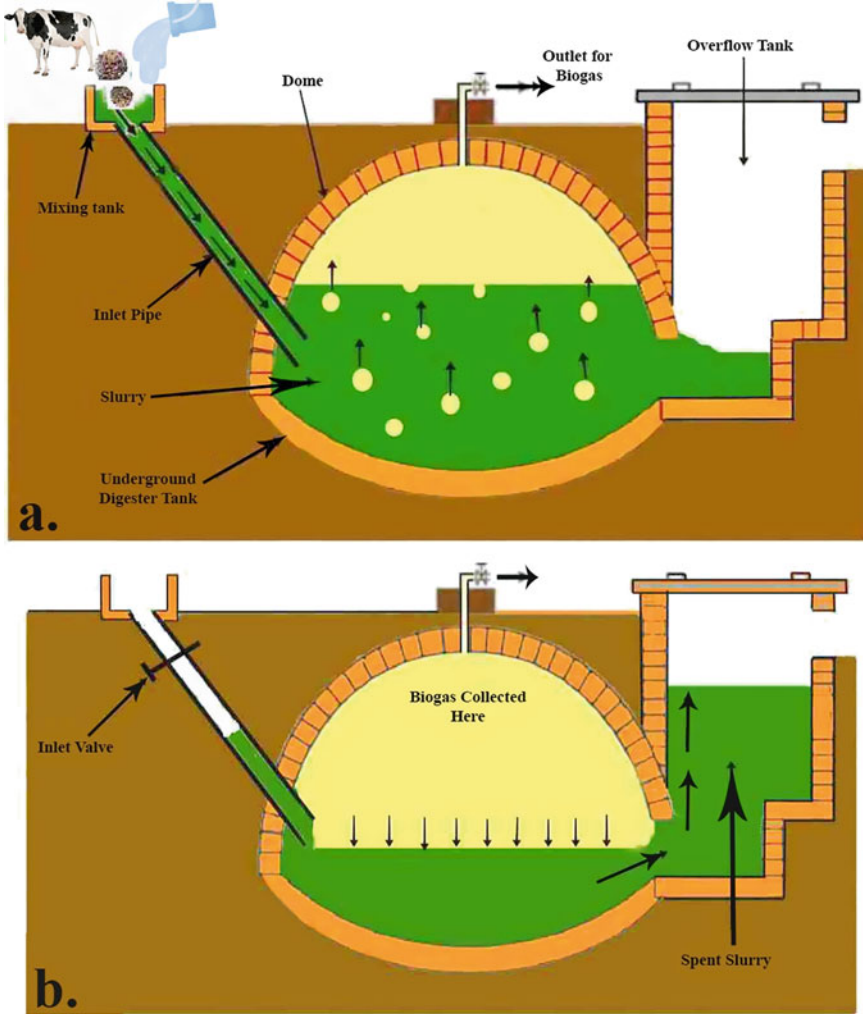
### 8.4.1 Fixed-Dome Digester

A fixed-dome plant is invented of a closed, dome-shaped digester with a fixed, feedstock inlet, a firm gas-holder, and the compensation tank or overflow tank. A schematic diagram is shown in Fig. 8.5. The digester stored the gas produced in the upper part of the reactor. The digestate was pushed into the compensation tank by the high pressure generated by gas produced in the digester with a closed outlet gas valve. The gas pressure falls and a relative amount of slurry flows back into the digester from the tank of compensation, once the gas valve is open for gas utilization. Given this design, gas pressure varies always, depending on gas production and use. Usually, such a plant is constructed underground, protecting the digester from low temperatures during cold seasons and at night. The internal pressure in the digester, which is normally 0.1–0.15, bar balances the surrounding soil up to the top of the gas-filled space (Werner et al. 1989).

Fixed-dome plants are only suggested for situations where experienced biogas technicians with specific technical skills in construction are available to ensure a gas-tight construction. In general, fixed-dome plants are characterized by modest initial cost and long operational life (about 15–20 years), since no moving or corroding parts are required. Though with time, the masonry building may become liable and spongy to cracking, resulting in gas leakages. Porosity may be counteracted with the use of special sealants; however, cracking often causes permanent leaks. The fluctuating gas pressure in this digester type might confound gas utilization (Nzila et al. 2012).

There are numerous designs of the fixed-dome digester such as the Chinese fixed-dome plant, the Indian *Deenbandhu*, or the CAMARTEC model developed in Tanzania. Fixed dome digester can be constructed in different sizes, typically ranging from 6 to 16 m<sup>3</sup>.

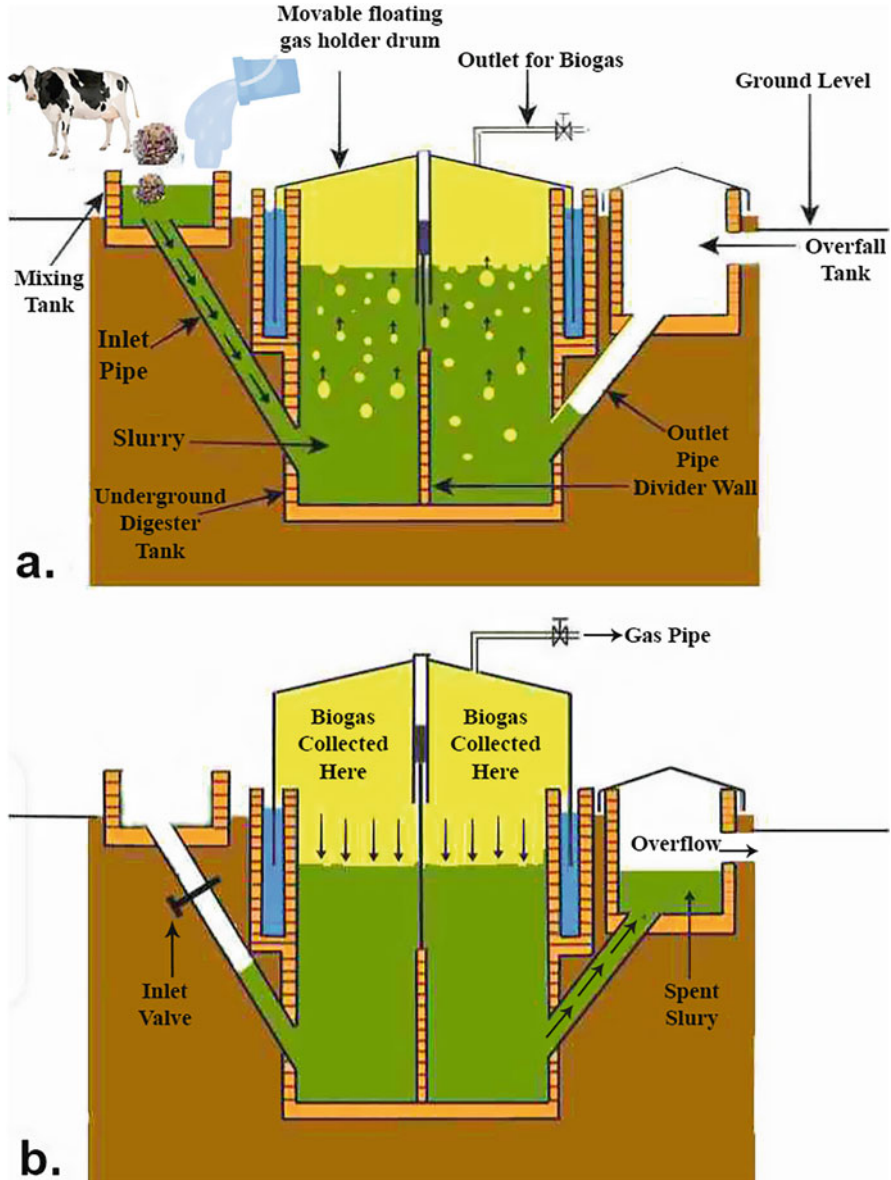
Nevertheless, the principle design elements of all fixed-dome digesters are the same. Generally, the fixed-dome digester type was classically used for cow dung-fed systems, but it is also appropriate for treating other waste types such as kitchen waste. Sometimes, toilets are also connected to the digester to treat the human waste product, which does not create significant problems.



**Fig. 8.5** Scheme of the fixed-dome digester; (a) production and collection of biogas, (b) digestate pushed into the overflow tank by the high pressure generated by gas

#### 8.4.2 Floating-Drum Digester

A floating-drum biogas plant contains a cylinder-shaped digester and a movable, floating gasholder (drum). The digester is mostly built underground (see Fig. 8.6), while the floating gasholder is above the ground. Smaller domestic-scale systems usually are above ground. The reactor part of the digester is typically made with bricks, concrete, or quarry-stone masonry and then plastered. The gas-holder is typically prepared from metal and is covered with synthetic paints, oil paints, or bitumen paints to protect it against corrosion. Conversely, it is important to ensure



**Fig. 8.6** Scheme of the floating-drum digester; (a) production and collection of biogas, (b) digestate pushed into the overflow tank by the high pressure generated by gas

sustained use by regular de-rusting, and the cover coating should be re-applied annually. A well-maintained metal gas-holder can be expected to last between eight to twelve (8–12) years in a dry climate or 3–5 years in humid areas. A proper

alternative to standard grades of steel is fiberglass-reinforced plastic or galvanized sheet metal (Nzila et al. 2012).

The generated gas collects in the gas drum, which falls or rises again, depending on the volume of gas produced and used. The drum level thus contains a valuable visual indicator of the quantity of gas available. The gas is provided at moderately constant pressure, which is contingent on the weight of the drum. Additional weights can be added on top of the gasholder, to increase gas pressure. Braces can be welded onto the inside of the drum which then helps to break up the scum layer when the drum is rotated (Vögeli et al. 2014).

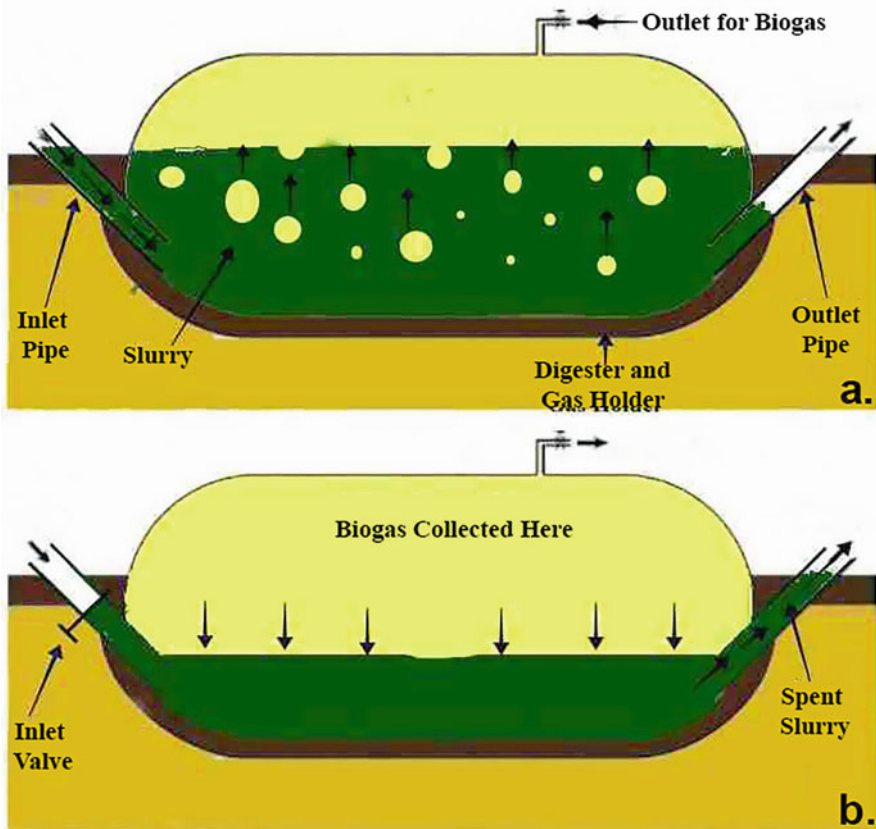
The gasholder is either a specifically constructed separate water jacket or floats directly on the fermenting slurry which reduces methane leakage, as shown in Fig. 8.6. A guiding frame constructed inside of the gas drum is an additional measure to prevent the tilting of the drum when it rises (see guide pole in Fig. 8.6). The design size of floating-drum plants is springy, with bioreactor sizes usually ranging between 1 and 50 m<sup>3</sup> (Vögeli et al. 2014).

### 8.4.3 Tubular Digester

A tubular biogas plant comprises a longitudinal-shaped heat-sealed, rubber bag (balloon) or weather-resistant plastic that serves as a digester and gas holder in one. The upper part of the balloon stores the gas produced. The outlet and inlet are attached straight to the skin of the balloon. No short-circuiting takes place as a result of the longitudinal shape, but since tubular digesters naturally have no stirring device, active mixing is incomplete and digestate flows through the reactor in a plug-flow manner. The pressure of the gas can be increased by placing weights on the balloon while taking care not to damage it. Figure 8.7 shows a schematic diagram of a typical tubular digester (Vögeli et al. 2014).

The advantage of these digesters is that they can be constructed at a low cost by standardized prefabrication. Furthermore, because of the shallow below-ground installation, they can be used in places with a high groundwater table. The plastic balloon though is quite liable to mechanical damage and has a comparatively short life span of 2–5 years (Nzila et al. 2012).

To prevent damage to and deterioration of the balloon, it is also very important to protect the bag from direct solar radiation with a roof. Moreover, a wire-mesh fence protects against damage by animals. This system can be modified for it to work at different altitudes and climates. For example, on the Bolivian Altiplano in west-central South America (more than 4000 m above sea level), biodigesters are surrounded in a polyethylene greenhouse, supported by two lateral adobe walls along the whole length of the shallow trench. A layer of 20 cm of insulating material (e.g., dry cereal straw and natural grass) can be used to decrease heat loss through the walls of the trench. The lateral walls accumulate the heat so that with freezing temperatures during wintertime nights, the digester remains operational of its high thermal inertia. Also, dark pipes are installed to pre-heat the water used for mixing the substrate before entering the balloon (Martí-Herrero 2008).



**Fig. 8.7** Scheme of tubular digester; (a) production and collection of biogas, (b) spent slurry pushed into the outlet pipe by the high pressure generated by gas

## 8.5 Microbes Involved in Biogas Production

Microbiology of anaerobic transformation of biological wastes is a method that involves numerous different kinds of microbes, such as hydrolytic, acid-forming, acetogenic, and methanogenic bacteria which produce  $\text{CO}_2$  and  $\text{CH}_4$  as the by-products of the digestion process. Each organic waste accounts for the degradation of a different type of compound.

### 8.5.1 Microbes Involved in Hydrolysis and Acidogenesis

The hydrolytic and acidogenic phases may be combined in the anaerobic acidogenic bacteria. The most commonly found acidogenic bacteria in digesters include species

of *Butyrivibrio*, *Propionibacterium*, *Selenomonas*, *Lactobacillus*, *Clostridium*, *Bacteroides*, *Bifidobacterium*, *Eubacterium*, *Ruminococcus*, *Acetivibrio*, *Peptostreptococcus*, *Peptococcus*, *Streptococcus*, and members of the *Enterobacteriaceae*. In mesophilic sewage sludges, there are usually between  $10^8$  and  $10^9$  hydrolytic bacteria per milliliter (Borja 2011; Kashyap et al. 2019).

### 8.5.2 Acetogenic Bacteria

Acetogenic species can be subdivided into those that do reduce protons to hydrogen obligately and those that are not obligately proton-reducing, that is, hydrogen-producing species during acetogenesis. The first group has a wide range, comprising the homoacetogens and species that may direct their metabolisms to proton reduction in the presence of a sufficient hydrogen-removing system. Homoacetogenic species are known in the genera *Acetobacterium*, *Acetogenium*, *Acetoanaerobium*, *Butyribacterium*, *Eubacterium*, *Clostridium*, and *Pelobacter* (Borja 2011).

In environments with sufficient  $H_2$  sinks, such as anaerobic digesters, many of the acidogenic bacteria direct their metabolism to acetogenesis. This facultative change in metabolism has been demonstrated in defined methanogenic co-cultures degrading alcohols, pyruvate, lactate, fructose, glucose, cellobiose, and cellulose. Obligately proton-reducing acetogenic microbes can only be grown in a sufficiently electron-removing environment, for example, in monoxenic culture with a hydrogen-removing or formate-removing species. The mixed culture concerning this type of “mutualistic” interaction is a culture containing the acetogenic bacteria and a hydrogen-removing bacterium such as a methanogen. *Desulfovibrio* spp. is obligatory proton-reducing acetogens when metabolizing ethanol or lactate in the absence of sulfate, and can be cultivated in mutualistic co-culture with methanogens. Some of the acetogens and their metabolizing substrate have been depicted in Table 8.2. The relative significance of formate and hydrogen in interspecies electron transfer essentials is to be established in different digesters and under different operating conditions (Borja 2011).

**Table 8.2** Acetogenic bacteria (Schiel-Bengelsdorf and Dürre 2012)

S. no.	Microbes	Metabolize/degrade [organic waste carbon chain acid ( $C_5$ ) to acetate ( $C_2$ )]
1.	<i>Methanobacterium suboxydans</i>	Pentanoic acid ( $C_5$ ) to propionic acid ( $C_3$ )
2.	<i>Methanobacterium propionicum</i>	Propionic acid ( $C_3$ ) to acetate ( $C_2$ )
3.	<i>Syntrophobacter wolinii</i>	Propionic acid ( $C_3$ ) to acetate ( $C_2$ )
4.	<i>Syntrophomonas wolfei</i>	Butyrate
5.	<i>Syntrophusbus wellii</i>	Benzoate
6.	<i>Desulfovibrio</i> spp.	Ethanol or lactate

### 8.5.3 Methanogens

Methanogenic microbes are present in sewage sludges at populations of up to  $10^8$  per milliliter and contribute up to 10% of the volatile solids. They are a morphologically diverse group of archaeobacteria unified by their capability to derive energy from methanogenesis. A limited range of substrates are utilized by the methanogens,  $H_2 + CO_2$  and acetate being the most important substrates in AD. Most methanogenic microbes utilize  $H_2$  and  $CO_2$ , but species of only two genera, *Methanothrix* and *Methanosarcina*, can produce methane from acetic acid. The species of methanogens that most commonly use  $H_2$  and  $CO_2$  as substrate and those that use acetate found in anaerobic digesters are described in Table 8.3.

Alternatively, hydrolysis is claimed to be rate-limiting when the biological waste contains much insoluble material (e.g., cellulosic compounds). Though, in the AD of soluble substrates, either methanogenesis from acetate or acetogenesis is considered to be rate-limiting. Under certain conditions, the rate of acetogenesis is controlled by the  $H_2$ -utilizing methanogens and so methanogenesis by either the acetate- or  $H_2$ -utilizing methanogens can be rate-limiting to the Anaerobic Digestion process (Schiel-Bengelsdorf and Dürre 2012).

**Table 8.3** Methanogens that most commonly use  $H_2$  and  $CO_2$  as well as acetate as a substrate are found in anaerobic digesters (Schiel-Bengelsdorf and Dürre 2012)

S. no.	Methanogens that use			
	$H_2$ and $CO_2$ as a substrate		Acetate as substrate	
	Genus	Species	Genus	Species
1.	<i>Methanobacterium</i>	<i>Bryantii, formicicum, wolfei, thermoautotrophicum, uliginosum, thermoalcaliphilum, thermoaggregans</i>	<i>Methanosarcina</i>	<i>Barkeri, mazei, acetivorans</i>
2.	<i>Methanobrevibacter</i>	<i>Arboriphilus, ruminantium, smithii</i>	<i>Methanothrix</i>	<i>Soehngenii, concilii</i>
3.	<i>Methanothermus</i>	<i>Fervidus</i>		
4.	<i>Methanococcus</i>	<i>Maripaludis, deltae, vanniellii, voltae, jannaschii, halophilus, thermolithotrophicus, frisius</i>		
5.	<i>Methanomicrobium</i>	<i>Mobile, paynteri</i>		
6.	<i>Methanogenium</i>	<i>Cariaci, marisnigri, olentangyi, tatii, aggregans, thermophilicum, bourgense</i>		
7.	<i>Methanospirillum</i>	<i>Hungatei</i>		
8.	<i>Methanoplanus</i>	<i>Limicola</i>		



### 8.5.3.1 Characteristics of the Methanogen Families, Substrates for Methanogenesis; Digester Input, and % of Biogas Produced

The two families of methanogenic microbes are Methanobacteriaceae and Methanothermaceae which are closely related. These methanogens have cell walls composed in part of pseudomurein (Kandler and König 1985). The Methanothermaceae also contain an additional surface layer composed of protein on their cell wall. The family of Methanothermaceae contains one genus, Methanothermus, and both species are extremely thermophilic bacilli with temperature optima of 83–88 °C. Like in many of the Methanobacteriaceae, the only substrate for methanogenesis is  $H_2 + CO_2$ . The family of Methanobacteriaceae contains two genera composed of mesophilic as well as thermophilic species. These genera, Methanobacterium and Methanobrevibacter, are bacilli that utilize either  $H_2 + CO_2$  alone or  $H_2 + CO_2$  and formate as substrates for methanogenesis (Miller and Wolin 1983).

Some of the most important and most distinctive features of all six families of methanogenic species, substrates for methanogenesis; digester input and % of biogas produced are summarized in Table 8.4 below:

**Table 8.4** Some characteristics of the methanogen families, substrates for methanogenesis; digester input, and % of biogas produced (Rosenberg et al. 2014)

S. no	Family	Characteristics	Substrates for methanogenesis
1	Methanobacteriaceae	Long or short rods, mostly Gram-positive; contain pseudomurein; nonmotile; GC content, 23–61 mol%	$H_2 + CO_2$ , formate, or alcohols But Cocci, utilize only $H_2 +$ methanol
2	Methanothermaceae	Rods; Gram-positive; contain pseudomurein; nonmotile; extreme thermophiles; GC content, 33–34 mol%	$H_2 + CO_2$
3	Methanococcaceae	Irregular cocci, Gram-negative; motile; GC content, 29–34 mol %	$H_2 + CO_2$ , and formate
4	Methanomicrobiaceae	Rods, spirals, plates, or irregular cocci; Gram-negative; motile or nonmotile; GC content, 39–61 mol%	$H_2 + CO_2$ , frequently formate and sometimes alcohols
5	Methanocorpusculaceae	Small, irregular cocci; motile or nonmotile; GC content, 48–52 mol%	$H_2 + CO_2$ , formate, and sometimes alcohol
6	Methanosarcinaceae	Pseudosarcina, irregular cocci, sheathed rods; substrates for methanogenesis are Gram-positive or negative; frequently nonmotile; GC content, 36–52 mol	Sometimes $H_2 + CO_2$ , acetate, and methyl compounds; formate is never used

### 8.5.3.2 Cooperation of Microorganisms in the Methane Fermentation Process

The four different groups of microorganisms that are responsible for the conversions of complex organic compounds to biogas mainly  $\text{CH}_4$  and  $\text{CO}_2$  are presented in Table 8.5. These groups of microbes may be counted among secondary fermentation bacteria (syntrophic and acetogenic bacteria), primary fermentation bacteria, and two types of methanogens belonging to the domain Archaea. These microbes occur in the ordinary environment and fulfill various roles during the process of anaerobic degradation of wastes (Conrad 1999). Syntrophy is a form of association of two metabolically different groups of bacteria, which permits the degradation of various substrates (Demirel and Scherer 2008).

Cooperation of the population of microbes permits the synthesis of certain products which are then used by a different group of bacteria. The bacteria which are involved in the production of methane belong to the domain Archaea and exhibit symbiosis relationships with other populations of microbes. They may develop only when hydrogen is used by hydrogenotrophs. Such cooperation between microbes producing hydrogen and using hydrogen was defined as the interspecific transfer of hydrogen (Conrad 1999). Syntrophy between microorganisms producing and using hydrogen allows for the growth and activity of these bacteria.

## 8.6 Factors Affecting Biogas Production

Biogas production through the anaerobic digestion process is influenced by a large number of factors that can influence digestion efficiency and the potential of biogas production (Mathew et al. 2015). Biogas production can be significantly improved with statistical optimization and pretreatment techniques (Gopal et al. 2021). Some of these factors are discussed below.

### 8.6.1 Temperature

Temperature is a critical and very important parameter to take into consideration during biogas production. It is the principal environmental factor affecting biogas

**Table 8.5** Microbial cooperation in organic matter degradation (Zieminski and Frac 2012)

Microorganisms	Electron donor	Electron acceptor	Product	Reaction type
Fermentative bacteria	Organic carbon	Organic carbon	$\text{CO}_2$	Fermentation
Syntrophic bacteria	Organic carbon	Organic carbon	$\text{H}_2$	Acidogenesis
Acetogenic bacteria	Organic carbon/ $\text{H}_2$	$\text{CO}_2$	$\text{CH}_3\text{COOH}$	Acetogenesis
Methogenic bacteria	Organic carbon/ $\text{H}_2$	$\text{CO}_2$	$\text{CH}_4$	Methanogenesis

digester performance (Mata-Alvarez et al. 2014). It affects the physical and physicochemical properties of the compounds present in the digester and the kinetics and thermodynamics of the biological process (Kougias et al. 2013). Temperature causes significant effects on the microbial community, interfering with the stability of the process, microbial growth, substrate utilization rate, and biogas yield (Khalid et al. 2011). The rate of biological reactions is designated by temperature. Temperature is a significant parameter that quite often has to be scrutinized, specifically, when there is a variation in the weather. There are three temperature ranges for biogas production, which are psychrophilic temperatures: 10–20 °C with an optimum at 25 °C; mesophilic temperatures: 20–45 °C with an optimum at 35 °C; and thermophilic temperatures: 50–65 °C with an optimum at 55 °C (Kothari et al. 2014).

There are mainly two temperature ranges that provide optimum digestion conditions for the production of methane—the mesophilic and thermophilic ranges. The types of active microbial consortia at the two temperature conditions are quite dissimilar. The choice of temperature condition will be determined by the type of expected outcome. However, the temperature should be appropriate to the type of microorganisms used. Thermophilic temperatures' condition is commonly used in large-scale biodigester (Kwietniewska and Tys 2014). This temperature condition requires higher energy costs and may favor the acidification of the reactor by inhibiting biogas production (Mao et al. 2015). Silwadi et al. (2022) investigated the effect of temperature on the enhancement of biogas production by anaerobic digestion of three different animal droppings, namely, cow, camel, and chicken. They found that digestion of cow, camel, and chicken manure at 37 °C increased the production by 2.2-, 2.1-, and 1.3-fold, respectively, compared to that obtained at 25 °C. Hossain et al. (2022) studied various factors which influence biogas production and found that biogas production rate and cumulative biogas production were found to increase with a rise in temperature.

## 8.6.2 pH

pH is one of the major operational factors that affect biogas production. During anaerobic digestion, different optimal pH values are required at different stages of biogas production. Each microbe grows much better at a certain pH value range, and the uttermost growth of the microbes occurs at an optimum pH value (Montañés et al. 2015). The optimum pH range to achieve high biogas yield in the anaerobic digestion process lies in the range of 6.5–7.5. During anaerobic digestion, the processes of hydrolysis and acidogenesis occur at acidic pH levels (pH 5.5–6.5), as compared to the methanogenic phase (pH 6.5–8.2) (Khalid et al. 2011). Methanogens are sensitive to acidic situations. The growth of microbes and the yield of methane could harmfully be affected by this acidic condition (Arsova 2010).

pH is a very important factor in the anaerobic digestion process. It provides an overview of the effectiveness of the process (Mathew et al. 2015). The lower pH is an indication of the failure of the system or low buffering capability that can inhibit digestion. High pH can also limit the methanogenesis process. The optimal pH value

is of great significance, and to keep a constant pH level, buffers such as lime and calcium carbonate need to be added to the system. To retain a steady pH value within the system, the interaction between the VFAs and bicarbonate concentration is crucial (Liu et al. 2008).

### 8.6.3 Nutrients Requirements

The nutrient requirement is a key concern for the steady execution of biogas production processes (Mathew et al. 2015). As for any biological processes, where microorganisms are involved, both the nutrient required in large and small quantities (macro and micronutrients) should be provided to the microorganisms in the right proportion to be able to achieve efficient biogas production. The nutrients should be found in abundance in the digester as the shortage of any of them may inhibit the process (Mara and Horan 2003). Insufficient availability of nutrient concentration may lead to low biogas yields and process uncertainty (Lebuhn et al. 2008). The macro and micronutrients are essential for the continual performance of the biogas production process (Bruni et al. 2010). Fundamental macronutrients such as carbon (C), nitrogen (N), phosphorus (P), and sulfur (S) are necessary for microbial growth and therefore must be provided to ensure efficient and stable biogas production. Among micronutrients iron (Fe), nickel (Ni), cobalt (Co), molybdenum (Mo), and tungsten (W) are the most important ones (Zandvoort et al. 2006). The growth of methanogenic microbes is reliant on many of these ions, so it is essential for all methanogenic pathways and thus their availability is necessary for biogas production. However, the exact quantity of the required ions should be determined individually in each case because it depends on the microbial consortia and the substrate used (Jagadabhi 2011).

### 8.6.4 C/N Ratio

The carbon/nitrogen ratio plays an important role in the anaerobic digestion process. It is the ratio between the amount of carbon and nitrogen contained in organic matter. The relation between the measure of carbon and nitrogen in organic matter is described by the C/N ratio. It is an important parameter in estimating nutrient deficiency and ammonia inhibition (Hartmann and Ahring 2006). Carbon present in organic matter is of great importance for biogas production. Nitrogen deficit can result in an inadequate consumption of the carbon source, which may result in the decline of microbial growth and lastly lead to a decrease in the biogas yield (Resch et al. 2011). Nitrogen is used as a nutrient by the microorganisms responsible for anaerobic digestion. Nitrogen compounds from organic waste are converted into ammonia in the anaerobic digestion process which contributes to maintaining the pH of the system stable during the process (Khalid et al. 2011). The optimal C/N ratio for anaerobic digestion of organic waste ranged from 20 to 35 (Mathew et al. 2015). A large carbon/nitrogen ratio is a sign of fast ingestion of nitrogen by methanogens,

which then leads to lower biogas yield, but if the ratio is low, an accumulation of ammonia occurs and pH values then may exceed 8.5, such condition can negatively influence the activity methanogenic bacteria (Kothari et al. 2014). Therefore, an optimal C/N ratio must be maintained to ensure efficient biogas production.

### 8.6.5 Agitation

The purpose of mixing the substrate in an anaerobic digester is to mix the new material with digestate containing the microbes (Mao et al. 2015). It is not essential, but always advantageous. Agitation is done to make sure that the contact between substrate and microbes is close and hence results in an enhanced digestion rate of the substrate (Hajji et al. 2016). Agitation enhanced biogas production by about 62% compared to gas production without agitation and thus increase biogas yield (Cavinato et al. 2013). The agitation has the advantage of bringing a homogeneous environment and maintaining a uniform slurry, thereby preventing scum formation. Scum can result in blockage of the gas pipe or potentially lead to foaming over the digester, avoids temperature gradients within the digester, and agitation also prevents grit deposition. Inappropriate mixing can interrupt the contact of microbes to the substrate and decrease biogas production, hence slow, occasional, and harmonious mixing of slurry which enhances biogas production is preferred (Prasad 2012).

### 8.6.6 Water Content

Water is an important nutrient for microorganisms' life and activity. It is an essential component of the organic matter breakdown process since it acts as a solvent and contributes to the mass transfer and diffusion of microorganisms, allowing interaction between the surface of the substrate with microbes involved in the anaerobic digestion process (Bollon et al. 2011). Biogas production from organic matter breakdown requires aqueous environments with water activity higher than 0.91 (Kwietniewska and Tys 2014). The highest methane production occurs at 60–80% moisture as high levels of moisture facilitate the digestion process (Khalid et al. 2011). The movement of bacteria and the activity of extracellular enzymes are highly determined by water content in the digester. The optimum water content of 60–95% has to be maintained in the digester. Although, the optimum moisture content varies with the different input materials, chemical characteristics, and degradation rates (Demetriades 2009).

### 8.6.7 Hydraulic Retention Time (HRT)

Hydraulic Retention Time describes the average time period for which the organic material remains inside the digester or the time required for a complete breakdown of

organic matter. Hydraulic Retention Time (HRT) can be expressed by the equation below:

$$\text{HRT} = V/Q$$

Where  $V$  is the reactor volume ( $\text{m}^3$ ) and  $Q$  is the flow rate of the fresh substrate ( $\text{m}^3/\text{day}$ ) (Kothari et al. 2014). Maximum biogas production occurs at the optimal value of HRT. Underloading and overloading reduce biogas production (Dobre et al. 2014). VFA will accumulate if the retention period is less than the optimal value, which will cause severe fouling and result in reduced biogas production. And if the retention period is above the optimal value, the biogas component will not be utilized effectively, hence biogas production will be reduced (Chen et al. 2016). Hydraulic retention time depends on the temperature of the system and the substrate to be digested. Usually, the HRT for mesophilic temperature conditions ranges from 10 to 40 days, while for the thermophilic condition, the time is shorter, 14 days (Kothari et al. 2014). In conditions where the influent streams have large solids concentrations, extensive retention times are vital to maximize biogas production (Khanal 2009). Hydrogen-producing bacteria prefer short retention times. In contrast to methane-producing bacteria, short retention times lead to a decrease in methane production.

### 8.6.8 Redox Potential

The redox potential of a digester is another important factor that affects biogas production. It is a measure of the oxidizability or reducibility of its content. Biogas production only proceeds in an environment free of oxygen (an anaerobic environment). The optimal value of the redox potential of a reactor must be less than  $-330$  mv for efficient biogas production (Weinrich et al. 2018).

### 8.6.9 Ammonia

Ammonia is frequently described as one of the impeding substances in the biogas production process. Free ammonia or ammonium ions are produced by the breakdown of nitrogenous matter in the digester, commonly present in the form of proteins (Chandra et al. 2012). Microorganisms need some ammonia to form cellular protoplasm for growth and reproduction (Lin et al. 2011). A healthy system will have an ammonia concentration of around 200 mg/L to support the anaerobic growth of the bacteria, while the increase in concentrations of ammonia greater than 1500 mg/L will cause inhibitory effects. This inhibition will cause inequity and accrual of intermediate digestion products such as VFAs which can result in acidification of the reactor, which in turn may result in a reduction in methane production. However, the effects of ammonia inhibition can be lessened by dilution with water in extreme

ammonia overloads, or altering feedstock to adjust C/N ratio in lesser overloads (Kayhanian 1999).

### 8.6.10 Organic Loading Rate (OLR)

Organic Loading Rate (OLR) is the amount of substrate (biomass) fed into or loaded to a unit of volume of the reactor under a unit of time. It signifies the quantity of substrate introduced into the digester in a given time. Organic Loading Rate is typically expressed in terms of kg volatile solid per m<sup>3</sup> per day [kg VS (m<sup>3</sup> day)<sup>-1</sup>], and can be defined by the equation:

$$OLR = Q.VS/V$$

Where OLR is the organic loading rate (kg VS substrate/m<sup>3</sup> digester/day), [kg VS (m<sup>3</sup> day)<sup>-1</sup>],  $Q$  is the fresh substrate added daily (kg/day),  $V$  is the volume of the bioreactor (m<sup>3</sup>) and VS stands for volatile solids [kg VS (kg)<sup>-1</sup>] (Kothari et al. 2014).

Biogas production is highly influenced by the organic loading rate. The organic loading rate depends on the types of biomass fed into the digester. Underloading and overloading reduce biogas production (Babae and Shayegan 2011). If OLR is increased, the metabolic activity of microbes will be high and hence improve biogas yield. Very high overloading of OLR leads to a significant rise in VFAs and causes its accumulation, which may result in acidification, a decrease in pH and the production of biogas, and may eventually result in system failure (Chen et al. 2016). This in turn influences the biological activity of microbes that generate methane as their growth is inhibited below a pH of 6.6, thus reducing the production of methane, which is the major product of biogas. To optimize digester efficiency and maximize methane production, it is therefore very crucial to assess the suitable OLR for a particular substrate.

### 8.6.11 Volatile Fatty Acids

Volatile Fatty Acids (VFAs) are also an important factor that affects biogas production. VFAs are needed in small quantities as part of the intermediary step for metabolic pathways of methane production by methanogens (Xu et al. 2018). It is estimated that to have a stable process of anaerobic digestion for the production of biogas, the volatile fatty acids, concentration, particularly acetic acid, should be below 2 g/L (Jain and Mattiasson 1998). VFAs can accumulate in a reactor when methanogens cannot keep up with the rate of degradation in the earlier digestion stages, causing a drop in the pH, which in turn will inhibit methanogens, and finally result in biogas digester failure (Yang et al. 2015).

### 8.6.12 Particle Size

The particle size of the substrate also affects biogas production. For microbes to digest, the large particle size of a substrate is problematic and may also result in reactor blockage. Small particles have a large area for adsorption of the substrate and thus allow for increased microbial activity, thus increasing the production of biogas (Sreekrishnan et al. 2004).

### 8.6.13 Inocula

Biogas production is not possible without a sufficient quantity of microbes that support biogas production. Inoculating the digester with microorganisms is necessary for the anaerobic digestion process. Diluted cow dung (optimally 1:1 ratio with water) is an ideal inoculate. At the start-up phase of biogas production, the bacteria population needs to be progressively acclimatized to the feedstock. This can be attained by gradually increasing the everyday feeding load which permits time to attain a stable microbial population. Some of the effluents are collected and inoculated back into the reactor. It is a way of inoculating fresh manure with active microbes. This inoculation of fresh manure can increase biogas production by up to 30% (Budiyono et al. 2014).

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## 8.7 Benefits of Biogas Technology

The production and use of biogas from anaerobic digestion provide socioeconomic and environmental benefits to society as a whole as well as the farmers involved. The use of the internal biogas production value chain boosts local economic potential, protects rural jobs, and strengthens regional financial strength (Saidmamatov et al. 2021). It contributes to the growth of the economy and society and increases living standards. Renewable energy sources are gaining popularity, and there is widespread interest in them. Biogas demand is gradually increasing as more people build biogas plants to supply biogas (Jørgensen 2009).

### 8.7.1 Reducing the Production of Greenhouse Gas

The use of fossil fuels such as crude oil, lignite, hard coal, and natural gas converts carbon deposited in the Earth's crust for hundreds of millions of years and releases it into the atmosphere as carbon dioxide (CO<sub>2</sub>). Carbon dioxide (CO<sub>2</sub>) is one of the constituents of greenhouse gas (GHG), thus global warming has resulted as a consequence of an increase in the current carbon dioxide (CO<sub>2</sub>) concentration in the atmosphere. On the other hand, the crucial difference, as compared to fossil fuels, is that the carbon in biogas was recently extracted from the environment by the plants' photosynthetic behavior (Tsaurai 2018). Thus, in a very short period



(between one and several years), the carbon cycle of biogas is closed. The generation of biogas by anaerobic digester also decreases methane and nitrous oxide emissions from the dumping and usage of untreated animal manure as fertilizer (Khayal 2019).

### **8.7.2 Source for Renewable Energy**

The present worldwide energy supply is dependent on fossil sources (crude oil, natural gas, lignite, hard coal). These are fossilized remains of dead animals and plants, which have been exposed to heat and pressure in the Earth's crust over millions of years. Fossil fuels are non-renewable resources; reserves are depleting much faster than new ones are being formed; as a result, the world's economies rely on crude oil today (Khan et al. 2017). There is some discrepancy among scientists on how long this fossil resource will last. Peak oil production is defined as "the point in time at which the extreme rate of the worldwide production of crude oil is reached, after which the rate of production enters its terminal decline." According to researchers, peak oil production has already happened or it is estimated to happen within the next period of time (Li 2008). The introduction and production of renewable energy systems such as biogas from anaerobic digesters would strengthen the reliability of the national energy supply and minimize reliance on imported fuels (Alhassan et al. 2019).

### **8.7.3 Low Input of Water**

As compared to other biofuels, biogas has several benefits. One of the benefits is that the method of anaerobic digestion requires the least amount of process water. This is an incredibly significant feature related to the assumed lack of water in many parts of the world (Khayal 2019).

### **8.7.4 Contribution to the EU Environmental and Energy Goals**

One of the key goals of European energy and environmental policy is to tackle global warming. The European targets for the development of renewable energy, the elimination of GHG emissions, and the effective management of waste are focused on the willingness of the Member States of Europe to take adequate steps to achieve them. The production and use of anaerobic digestion biogas have the potential to simultaneously comply with all three targets (Bartolini et al. 2017).

### **8.7.5 Reduction of Waste**

The ability to turn waste material into a valuable resource by using it as a substrate for anaerobic digestion is one of the key benefits of the biogas production process.

The overproduction of organic waste from manufacturing, agriculture, and households is a major problem affecting many developed countries. The production of biogas is an excellent way of coping with highly stringent national and European regulations in this region and of using organic waste for the production of energy, followed by the recycling of the digested substrate as fertilizer (Rai et al. 2020). Anaerobic digestion will also lead to a reduction in waste volume and waste disposal costs (Bong et al. 2017).

### 8.7.6 As an Excellent Fertilizer

A biogas plant is not solely an energy supplier but depends on the institutional structures and farmers' practices involved in making energy available. The digested substrate, commonly called the digestive, is beneficial nitrogen, phosphorus, potassium, and micronutrient-rich soil fertilizer that can be added to fields using the normal liquid manure application equipment. Due to higher homogeneity and nutrient abundance, better C/N ratio, and substantially decreased odor, digestive fertilizer performance has increased compared with raw animal manure (Kolar et al. 2011). Unpaprom et al. (2021) performed biogas production of crushed water hyacinth (WH) combined with swine dung (SD). The digestate from the biogas fermenter was confirmed to be an efficient alternative fertilizer with high nutrients (nitrogen, phosphorus, potassium) and environmentally friendly compared to chemical fertilizer.

### 8.7.7 Flexibility of Using Different Feedstock

For the production of biogas, different types of the feedstock may be used: animal manure and slurries, crop residues, organic waste from dairy production, food, and agro-industries, wastewater sludge, the organic component of municipal solid waste, household and catering organic waste, as well as energy crop waste. Biogas can also be obtained from landfill sites with unique infrastructure. The ability to use "wet biomass" types as feedstock, all characterized by a moisture content greater than 70% (e.g., waste sludge, animal slurries, flotation sludge from food manufacturing, etc.), is a significant benefit in biogas production. A variety of energy crops (grains, maize, rapeseed) have been primarily used as feedstock for the production of biogas in countries such as Austria and Germany. In addition to energy crops, biogas and fertilizer may be produced using all types of agricultural residues, degraded crops, unfit for food, or arising from unfavorable growing and weather conditions (Brémond et al. 2020).

### 8.7.8 Reduced Odor and Flies

Liquid manure, animal dung, and certain organic wastes are sources of constant, undesirable odor and attract flies at the time of their application and storage. Anaerobic Digestion eliminates these odors by as much as 75–85%. The digestive produced is nearly odor-free and the residual odors of ammonia fade soon after the application of fertilizer (Paolini et al. 2018).

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## 8.8 Future Prospects of Biogas Technology

The increasing energy demand compels the exploration of different types of waste and the development of new technologies for bioenergy production. Consumption of fossil fuels has contributed to detrimental effects on the environment and society (Korbag et al. 2020). Biogas is recognized as one of the leading bioenergy to address the existing environmental and energy challenges being faced by the world. It is an alternative energy source produced through solid waste management by the action of several microbes (Uche et al. 2020). The utilization of animal waste such as cow dung, pig dung, poultry dung, sheep dung, horse dung, etc. as the substrate for the production of biogas can effectively alleviate the shortage of energy and protect against environmental pollution (Gemechu 2020). Biogas is commonly used for cooking, lighting, heating, and power production and if purified further, it can be used as vehicle fuel (Roubik and Mazancová 2020).

The quantitative yield of biogas per unit weight of the substrate used differs from one type of substrate to another depending on the composition as well as the nature of the substrate. The methane content of biogas is the valuable portion of the gas and determines its calorific value (Nsair et al. 2020). Among the animal wastes that are used as substrate for biogas production, it has been reported that poultry waste has the highest methane content approximately 70% (Laiq Ur Rehman et al. 2019). Therefore, keen attention should be drawn to the utilization of several other types of animal waste that could have the potential to provide high methane content than poultry waste. Also, the degradation of organic waste material requires a co-ordinated action of several groups of microbial consortia with different metabolic capabilities (Palaniveloo et al. 2020). Conventional methods in molecular biology could help to classify only the most abundant microbial inhabitants found in the digester. Therefore, novel molecular biological techniques should be adopted that could provide a valuable tool for an improved understanding of this complex microbiological process, which in turn could help improve and control the process fruitfully in the future.

Biogas upgrading technologies are constantly being improved for better performance, enhanced upgrading efficiency, and low cost so that the technology gets a broader implementation globally. The current advancement of biogas upgrade techniques is illustrated by some innovative developments such as hydrate separation, cryogenic separation hybrid process, biological method, membrane enrichment, in situ upgrading, supersonic and industrial lung, multistage, and

high-pressure anaerobic digestion, though evaluated at laboratory and trial level (Olumide et al. 2017). However, commercial-scale optimization and testing are needed for these technologies to prove the full potential for biogas upgrading. Thus, there are still urgent needs for the development of novel anaerobic digestion technologies such as the development of a new reactor design to improve the efficiency of the process, increase biogas production rate and provide enormous potential concerning feasibility and technological simplicity with high efficiency. Also, research on the development of novel packing materials that can intensify mass transfer between gas and liquid and relatively low-pressure drop should be given utmost attention. There is also a need for the development of several computer models to model the biochemical anaerobic digestion process and regulate the process effectively. Better process management is essential for the future as well. Advanced monitoring and control systems will form part of the new epoch in the future of biogas plants and significantly contribute to process optimization (Theuerl et al. 2019). Operational process parameters like temperature, pressure, and flow rate of the gas should be optimized to decrease the large quantities of water needed, the cost for biogas compression, and water pumping.

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# Bioelectricity Generation from Organic Waste Using Microbial Fuel Cell

# 9

A. S. Zarena

## Abstract

Organic waste is a huge challenge and the scientific community is constantly striving to reduce organic waste emission. The motto of scientific community is “waste to watt” or “waste to energy.” This chapter emphasizes the application of microbial cells as electrochemical platforms for the conversion of organic waste for the production of fuels. Microorganisms play the most prominent role that is used to degrade the contaminants or substrates into harmless and valuable resources under mild operating conditions. In this technology, microorganisms act as biocatalysts to oxidize the substrate in the anode chamber from where the electrons are directed to the cathode as a result of electrical flow. Electricity generation by microbial fuel cell (MFC) is a pioneer in this issue. Like a battery, MFC uses chemical energy to generate electricity by using a natural process of cellular respiration of microorganism. MFCs have two electrodes each in the anode and cathode and they are held in separate chambers. The chambers can be with or without membrane. The anode chamber contains the anaerobic bacteria and the cathode chamber is aerobic. One of the best advantages of bacteria is that they can practically use nutrient that may be organic or inorganic. The oxidation process occurs within the bacteria living in the anode chamber. Electron bonds hold the food molecules together that bacteria eat. The bacteria break these bonds to release the electrons. The electrons released are captured to maintain a constant power density. Although the amount of fuel generated is low, nevertheless the technology is a hope for mitigating waste.

## Keywords

Microbial cell fuel · Bioelectricity · Electrode · Microbe · Organic waste

A. S. Zarena (✉)

Department of Biotechnology, Teresian College, Mysore, India

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## 9.1 Introduction

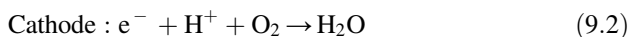
The global organic waste produced is alarming; most of the waste collected is dumped into landfills. This is not an effective way of disposing of the waste as this may further enhance the greenhouse effect by producing methane gas. The rapid consumption of non-renewable energy resources has led to the depletion of fossil fuels, an increase in CO<sub>2</sub> emission, and global warming, forcing the new world to look into alternative energy sources (Dhulipala et al. 2020). Organic waste and waste water are becoming a unique investment choice for developing biofuels because of the high organic contents, which could reduce the cost of biofuels production effectively (Owusu and Asumadu-Sarkodie 2016; Rai et al. 2020). Some specialized microorganisms have the ability to transfer electrons from the inner to the outer membrane of the cell via the electron transport chain. Researchers have used this phenomenon to explore new renewable energy generation methods based on microbial fuel cells (MFC) (Madakka et al. 2020). MFC is based on dual benefits for treating waste and producing energy from waste (Zhang et al. 2008). The entire concept of microbial fuel was an initiative by Michael Cresse Potter in 1911, wherein the first employed *Saccharomyces cerevisiae* and bacteria *Escherichia coli* for power generation in MFC (Potter 1911). For the last century, microbial fuel cell (MFC) has been used as an instrument in recovering resources from organic waste in generating biogas, dyes, electrical energy, biosurfactants, biofertilizers, bioplastics, pesticides, phenolic compounds, polyhydrocarbons, pharmaceutical products, textile, and removal of heavy metal (Sharma et al. 2020; Rai et al. 2020; Suresh et al. 2022). MFCs are also used in fertilizer production from human excreta and urine (Sabin et al. 2022).

Exo-electricigens are bacteria that can transfer electrons exogenously (outside the cell) to a terminal electron acceptor. A terminal electron acceptor's higher positive redox potential results in a higher energy gain. The process of generating electrons is known as electrogenesis, and the system or reactor is known as a microbial fuel cell (Logan 2009). The MFC architecture varies widely depending on the designer's need. A simple microbial fuel cell (MFC) consists of two compartments separated by a membrane or not, which allows the flow of electrons during the process (2004). The compartment consists of two electrodes, an anode and a cathode inoculated with microbes, and organic waste is added to the anode (Xu et al. 2017). Microorganisms use the organic matter contained in waste for their growth, nutrient, and reproduction. The metabolic processes by microorganisms produce several byproducts, such as protons and electrons that can be converted into energy (Clark and Pazdernik 2016). MFCs cannot operate at very low temperatures as the reactions inside the reactor take place at a very slow rate. Raw materials such as glucose, alcohol, butyrate, acetate, sodium acetate, sodium butyrate, and propionate are used as substrates for the organism to produce bioelectricity by chemical reduction (Logan 2008; Harnisch and Schröder 2010). Currently, the technology suffers in scaling-up with respect to designing and optimizing the physical and electrochemical parameters. Although microbes (biocatalysts) have a faster generation time, it is a poor conversion rate. Factors swaying MFC are type of microbial diversity,

electrodes, electron donor/acceptor, series and parallel connection, metal oxide, structure and concentration of organic pollutants, nature and resistance of electrolyte, circuit connection type (closed and open circuit), pH, temperature, and carbon source (Suresh et al. 2022).

## 9.2 MFC Working Principle and Electron Transfer

The significant components of MFCs are anode, cathode and membrane or separators. In MFCs, the anode chamber consists of organic matter and the exoelectrogenic bacteria that adhere to the anode surface and decompose the organic matter by oxidation of the substrates to produce CO<sub>2</sub>, protons, and electrons by the anaerobic process (Verma et al. 2018). The electrons generated from the metabolic activity of microorganisms are collected by cytochrome or redox protein and transferred to the cathode to react with a terminal electron acceptor (oxygen) through a copper electrical circuit and resistor. The flow of electrons through the external electric circuit is responsible for generating electric current (Logan and Regan 2006). Zhang et al. (2017) found that operating MFCs at a higher external resistance (1000 Ω) was feasible and then gradually switching to lower external resistances to facilitate higher current, increased energy output, and maximum power density. Concurrently, the H<sup>+</sup> ions flow through the semipermeable membrane combined with dissolved oxygen to form water molecules at the cathode. This process is driven by the electrochemical gradient resulting in a higher concentration of H<sup>+</sup> ions near the anode. The anode material acts as a catalyst for the transfer of the reaction while maintaining conductivity (Logan 2009).



The bacterial cells gain energy from pumping protons across the bacteria's inner membrane. This is responsible for forming a proton gradient, which produces ATP from ADP through ATPase and provides metabolic energy for the bacterium. The maximum current that an MFC can produce depends on the actual rate of substrate biodegradation and electron donor. The higher the positive redox potential of a terminal electron acceptor, the higher the energy gains for an organism (Harnisch and Schröder 2010).

### 9.2.1 Role of Microbial Fuel Cell (MFC)

- MFC system produces low amounts of sludge
- Recovers chemical energy from renewable sources like wastewater and organic matters

- Human waste is being reconnoitered as an efficient source to produce bioenergy or bioelectricity
- The concept used is “waste-to-energy”
- Generating electro carbon compounds from sequestration of CO<sub>2</sub> by employing anaerobic electrotrophic microbes as biocatalysts
- Onsite generation of biohydrogen and power in remote areas
- Potential application in groundwater to remove petroleum contamination
- It can be operated at ambient temperature and atmospheric pressure
- MFCs are used for the simultaneous removal of sulfide and nitrate from wastewater
- MFCs have desirable features of secondary storage batteries
- As biosensors in in situ monitoring and control for pollutant analysis, the advantages of using biosensors are miniaturization, easy operation, low cost, and safety

### 9.2.2 Limitation in Microbial Fuel Cell (MFC)

Besides the advantages of this technology, it still faces practical barriers such as low power, low efficiency, and current density that may be attributed to low-quality materials being used as anodes or material cost issues, especially the cathode and membranes if used (Yaqoob et al. 2020). The technology can have a brighter side as a new source of bioenergy, as researchers are extensively working on designs and configurations of electrodes and kinetics models for biofilm formation and planktonic performance (Solanki et al. 2020). Large-scale commercialization of MFC is the biggest obstacle due to its architecture (Logan and Regan 2006), membrane resistance during transportation of protons and problems in both chambers (Yaqoob et al. 2020). New materials, factors affecting the performance (electron transfer mechanisms, material and surface area of anode, cathode electrode, membrane, distances and flexibility), applications and cost-effectiveness for manufacturing MFCs have to be considered to extenuate electricity generation (He et al. 2017). Existing literature has pointed to greater power outputs between 2 and 5 W/m<sup>2</sup> and volumetric power over 100 W/m<sup>3</sup> if a smaller microbial fuel cell reactor is used for operation. Smaller the MFC, the better the operating condition (higher temperature and better conductivity). Reducing the distance between the anode and cathode can prevent fermentation and ohmic losses (Behera and Ghangrekar 2011; Yang et al. 2020a).

### 9.2.3 Mediators and Non-mediator MFCs

#### 9.2.3.1 Mediator-Less or Direct Electron Transfer Between the Cell Surface and the Electrode

It's the perfect alternative for producing electricity, mediator-less MFCs are operated with a dissimilatory metal-reducing microorganism primarily to the families of

Shewanella, Rhodoferrax, and Geobacter. Here the electron transport proteins present within the microbial cell transfer electrons from the cytoplasm to the outer membrane and finally to the anode. Electron transfer occurs through the outer membrane cytochrome or transmembrane and nanowires on the anode surface without any electron mediators. Bacterial nanowires are electrically conductive appendages composed of stacked cytochromes produced notably from the Geobacter and Shewanella genera and can form biofilms on the anode. The nanowires allow electricigens to use an electrode that is not in direct cell contact as the electron acceptor (Gorby et al. 2006). Chaudhuri and Lovley (2003) first reported a stable and long-term power generator by a mediator-less MFC using *Rhodoferrax ferrireducens*, that oxidized glucose to CO<sub>2</sub> and quantitatively transferred electrons to graphite electrodes. The H<sup>+</sup> diffusion also improved in the electrolyte fed with salt, thus enhancing current generation in membrane-less cathode chambers (Liu et al. 2015).

### 9.2.3.2 Mediator or Indirect Electron Transfer Mediator

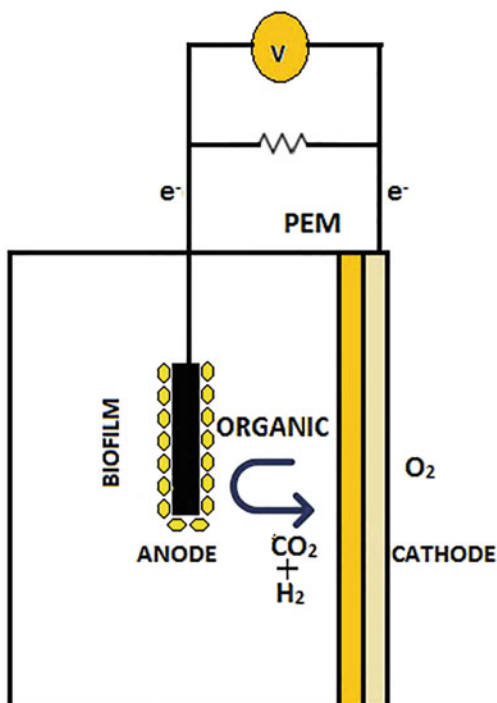
Here, a soluble mediator eliminates the direct interaction between the cells and the electron acceptor. In order to generate electricity, electro-active metabolites are used since microorganisms are electrochemically inactive for transferring electrons to the anode electrode. The electron mediators enter the bacteria cells, extract the electrons from the metabolic reactions of the electricigens, and supply these electrons to the anode of the MFC (He et al. 2017). *Lactococcus lactis* produces a natural mediator that produces quinones which are able to mediate electron transfer to extracellular electron acceptors such as Fe<sup>3+</sup>, Cu<sup>2+</sup> and hexacyanoferrate (Freguia et al. 2009). Depending on the microorganism species, involved mediators such as phenazine and pyocyanin may be natural. Neutral red, potassium ferricyanide and sulfate/sulfide anthracenedione, thionine, humic acid, meldola's blue (MelB) and 2-hydroxy-1,4-naphthoquinone (HNQ), riboflavin and methylene blue are used to increase the efficiency of microbial fuel cells and to reduce the activation energy (Li et al. 2014). However, as Cao et al. (2019) reported, the addition of mediators has attracted drawbacks to the working of MFCs as it could lead to relatively low current densities, expensive and toxic to the microorganisms. Also, separating these mediators from the solution is difficult as the mediators are water-soluble phenolic compounds. Some of the properties of mediators, as reported by Shukla et al. (2004), are (1) they should not interfere with the metabolites in the bacteria; (2) the mediators should be in an electrolyte solution and not adsorbed onto the microorganism; (3) the reduced mediator should easily diffuse out of the cell and move to the anode where they are oxidized; (4) the oxidized or reduced states of the mediator should be chemically stable and must be fast in the electrolyte solution.

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## 9.3 Materials and Architectures of Different Types of MFC

Depending on the availability of the substrate and microorganisms to metabolize the substrate, the power produced by MFCs may vary. The reactor is also affected by the rate of electron transfer from bacteria to the anode, cathode performance, the

**Fig. 9.1** Single chamber microbial fuel cell

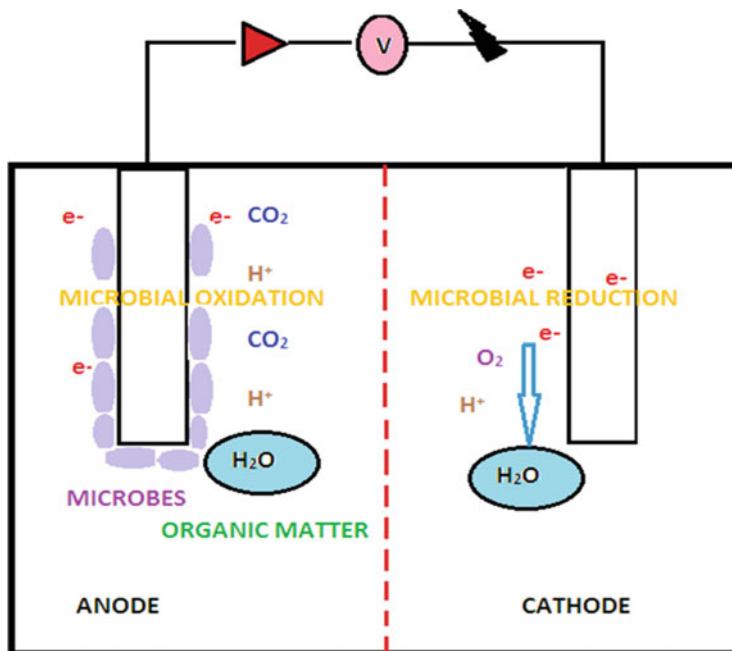


electrolyte, circuit resistance, proton mass transfer within the liquid, and the ion exchange (Liu et al. 2015). The anodic and cathodic chambers may or may not be separated by a proton exchange membrane (PEM), and different types of electrode material are being commercialized. Moreover, there are various influential factors for the performance of the MFC, such as temperature, pH, nutrients, and fuel cell configuration (Yaqoob et al. 2020). Figure 9.1 is a schematic illustration of single chamber microbial fuel cell.

### 9.3.1 Double-Chambered Fuel (DCF)

Double-chambered fuel is the most commonly used MFC. They are H shaped in structure and consist of a double-chamber with an anode and cathode chamber separated by a proton exchange membrane (PEM) or salt bridge. Oxidants such as ferricyanide and permanganate are used as a source of oxygen. In double-chambered MFC, the two chambers are connected by a circuit, and the sum of cations apart from H<sup>+</sup> transferred from the anode chamber to the cathode chamber is equal to the sum of e<sup>-</sup> transported through the circuit (Yap et al. 2020). The membrane (PEM) prevents oxygen diffusion into the anode and facilitates proton transfer from the anode to the cathode. The electrodes are in close proximity with the membranes resulting in higher oxygen diffusion from the cathode to the anode, thus increasing power





**Fig. 9.2** Double chamber microbial fuel cell

production and density (Choi et al. 2013). They are typically in batch mode and used in waste water treatment rather than electricity production (Du et al. 2007). Figure 9.2 schematic illustration of double chamber microbial fuel cell.

### 9.3.2 Single Chamber Fuel Cell (SCFC)

This fuel cell consists of a simple carbon electrode as an anode chamber and porous carbon exposed to air as a cathode. The cathodes are normally coated with graphite, in which electrolytes are poured into a steady state that prevents them from drying out. Single-chamber MFCs can achieve better performance than a two-chamber system due to the high mass transfer rate and oxygen concentration in the air compared to water (Fan et al. 2007a). A single chamber microbial fuel cell has an external cathode wall that is exposed to the atmosphere and eliminates oxygen (aeration) pumping to the cathodic chamber, thus reducing the cost (Cheng and Logan 2011). The advantage of using SCFC is less frequent oxidative media, aeration changing, and higher power generation (Logan et al. 2019). Eliminating the membrane in the chamber not only reduces the cost and complexity of MFCs but also increases the power density due to a decrease in internal resistance and is simpler to use than DCF.

### 9.3.3 Stacked MFC (SMFC)

In stacking, multiple cells are positioned in series or parallel connections. The voltage and current increase depending on the connection mode (parallel or series). The factor affecting the electricity production in stacked MFC is a microbial community, resistance, composition of the substrate, module configuration, anolyte and operation mode such as batch or continuous. It is possible to achieve improved voltage or current output by connecting several MFCs in series or parallel (Zhuang et al. 2012). Zhao et al. (2016) observed that when glycerol was used as a substrate, it is degraded faster in parallel connection than in series; they also noted that maximum power density increased with the increasing glycerol concentration in either of the connections. Generally, when MFC units are stacked in series, the voltage increases, whereas a parallel connection enhances the current (Aelterman et al. 2006).

Furthermore, switching from one connection mode to the other, the voltage output and microbial communities changed. For instance, when stacks were connected in series and then in parallel, microbial communities remained stable, but microbe abundance was affected when operated in parallel. The limitation of stacked MFC is a voltage drop due to voltage reversal, a cathode electrode and ionic conduction (Estrada-Arriaga et al. 2018).

### 9.3.4 Magnetic Fields Ceramic Microbial Fuel Cell (CMFC)

Ohmic losses are often a severe problem in the MFC reactor, sorted by ceramic-based stack MFC operating in super capacitive mode. This boosted power output and conductivity to a maximum of  $27.4 \text{ W/m}^3$  with an electrolyte solution of  $40.1 \text{ mS cm}^{-1}$ , thus reducing the overall system ohmic loss (Santoro et al. 2018). In another study, the efficiency of electricity generation was improved by replacing proton exchange membranes with ceramic membranes using microalgae *Spirulina platensis*. The results showed that the power generation could be boosted by 61% when implementing a 200 mT magnetic field (MF). The magnetic field affected the microorganism in both anode and cathode and improved the power density up to  $35.9 \text{ mW/m}^2$  and the current density of  $158.7 \text{ mA/m}^2$ . Ceramic microbial fuel cells (c-MFC) using diatoms have high energy conversion efficiency. The uniqueness of diatoms is they can fix 25% of atmospheric  $\text{CO}_2$ , hence releasing oxygen at longer hydraulic retention times (HRT). The hydraulic retention times (HRT) was  $32.2 \text{ W/m}^3$  at 24 h with constant power performance. These ceramic membranes are inexpensive when compared to other membranes (Walter et al. 2022). Though this technique is cost-efficient, it still suffers from calcium carbonate fouling (Chu et al. 2020).

### 9.3.5 Plant Microbial Fuel Cell (P-MFC)

Alternative approaches for power generation are being considered, such as plant microbial fuel cells (P-MFC). It is a novel technology that converts solar energy into electrical energy and is widely used in highly water-saturated ecosystems to produce sustainable energy. P-MFC is a reactor combining a microbial-based energy generation system and plants. Plants that can withstand waterlogged conditions, such as prickly pear, *Pachirama crocarpa*, *Populus alba*, *Opuntia* species (succulent plants), are widely utilized for sustainable electricity generation via plant-based biobattery technology. Despite the technology being initiated almost a decade ago, it is still considered in its infancy (Apollon et al. 2020; Lu et al. 2020). However, recent studies have revealed the beneficial roles of wetland plants in enhancing bioelectricity production within constructed wetland microbial fuel cells (CW-MFC). This enhancement can be attributed to the exudation of root oxygen, root exudates, and the removal of pollutants (Yang et al. 2021b). Performance of plant-MFC is governed by various parameters, such as selection of plant species, microbial flora in rhizosphere, design of MFC, electrode properties, inoculum characteristics, wastewater properties, factors like light intensity, and carbon dioxide concentration in air (Jadhav et al. 2021). Sharma et al. (2021) compared the cathode performance of wastewater containing plant *Canna indica* (PMFC) and the other having alga *Chlorella vulgaris* (AMFC). PMFC was deemed superior since its power output was six times higher ( $22.76 \text{ mW/m}^2$ ) than the AMFC ( $3.64 \text{ mW/m}^2$ ). Nguyen's studies have shown purple guinea grass cultivated in waterlogging could provide power densities of  $10.13 \text{ mW/m}^2$  two at the anode area. Soil water contents, ambient temperatures, photosynthesis, and photo-period were accredited to have a substantial role in controlling power and current outputs. At a lower temperature range of  $27\text{--}34 \text{ }^\circ\text{C}$ , a power density of  $0.6 \text{ mW/m}^2$  was obtained in waterlogging. The authors attributed the lower performance at low temperatures to the electroactive bacteria activities in the anode and the carbohydrate metabolism of plants (Nguyen and Nitisoravut 2019).

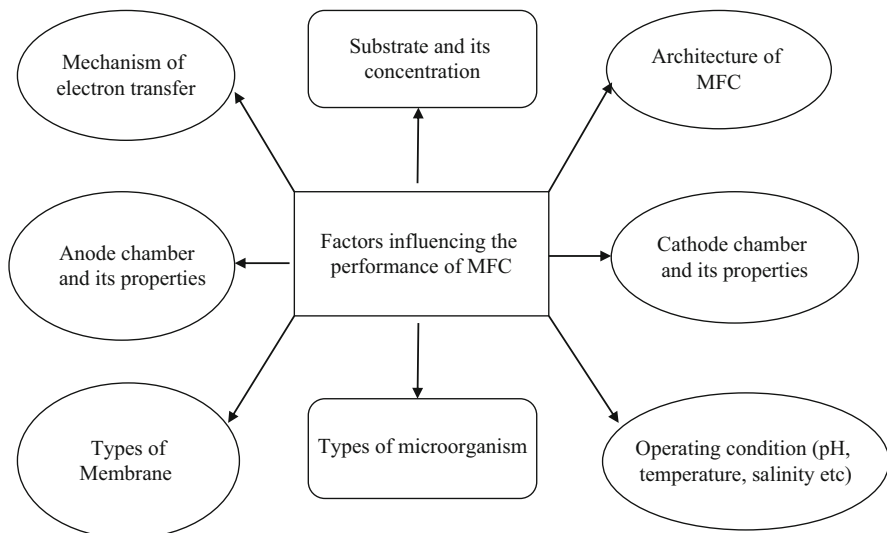
### 9.3.6 Photosynthetic Microbial Fuel Cell (Photo-MFC)

Photo-MFC can be considered the next-generation fuel cell for bioelectricity generation. Phototrophic prokaryotes (Anoxygenic phototrophic bacteria (APB)) are used to convert light energy into electricity through photosynthesis. As reviewed by Qi et al. (2018), at the anode, APB contains two pathways: APB can produce electrons by anoxygenic photosynthesis or endogenous respiration; hydrogen from APB photosynthesis is used as a medium for electron generation. The most frequently used APB were *Rhodospirillum*, *Rhodobacter*, *Rhodopseudomonas*, *Rhodovulum*, and *Chlorobium*. Photosynthetic MFCs provide treatment of biodegradable wastes by bacteria in the anode and remove carbon dioxide, phosphorus, and nitrogen in the cathode. The organic matter in the cathode could serve as nutrients for the algae, improving photo-MFC competence (Aiyer 2021). Sogani et al. (2021) investigated

the influence of a hybrid photo-assisted microbial fuel cell using *Rhodospseudomonas palustris* for the biodegradation of ethinylestradiol (EE2). An essential component of oral contraceptives that causes micropollutants in various wastewaters is highly recalcitrant. Degradation of EE2 to 89.82% with a maximum power density of  $0.633 \pm 0.04 \text{ mW/m}^2$  occurred at the bottom photo MFC along with top 63% bio-hydrogen production as a co-catabolite along with glycerol (Sogani et al. 2021).

## 9.4 Electrodes

The performance and cost of electrodes are the most critical aspects of designing an MFC. In recent years, a wide range of electrode materials and configurations have been tested and developed to enhance the performance of MFCs and lower material costs. The current trend in electrode modification with nanoparticles has become a new buzz to improve the performance of power outputs. According to Logan and Regan (2006), for an electrode to be ideal, the materials should possess certain features: (1) satisfactory conduction of electricity and little resistance; (2) corrosion resistance and chemical stability; (3) biocompatibility; (4) suitable toughness and mechanical strength; (5) high surface area. Figure 9.3 shows the factors affecting the microbial fuel cell.



**Fig. 9.3** Factors influencing microbial fuel cell performance

### 9.4.1 Cathode Electrode

The cathode electrode plays an important role in power generation. There are two potential methods of reducing cathode fuel oxygen levels. A direct 4-electron pathway can reduce oxygen to water or a 2-electron pathway to peroxide. The most desirable one is the 4-electron pathway (Panomsuwan et al. 2016). The drawback at the cathode is a low oxygen reduction reaction (ORR) kinetics which is improved by noble metals such as platinum (Pt), gold (Au), and palladium (Pd) (Khilari et al. 2015). Noble metals have outstanding electro-catalytic performance and four-electron transfer routes (Shabani et al. 2020). However, as reported, these noble metals come with a high cost, limited availability, poor stability, and surface poisoning. To overcome these problems, researchers have identified alternative solutions using tin oxide ( $\text{SnO}_2$ ), nickel-based composite, and sodium hexahydroxostannate ( $\text{Na}_2\text{Sn}(\text{OH})_6$ ) (Das and Jayaraman 2014). The cathode electrocatalyst developed using  $\text{Na}_2\text{Sn}(\text{OH})_6$  synthesized with a higher concentration of NaOH (2.0 M) showed higher ORR activity in terms of higher power density, the onset of potential and current density with a four-electron transfer process using pure and mixed inoculums. It proved to be a more cost-effective material for energy recovery in the MFC than  $\text{SnO}_2$  (Rout et al. 2020). The nickel-based composite showed promising high-effective oxygen reduction performance and outstanding power output with a power density of  $1421.4 \text{ mW/m}^2$  (Li et al. 2020a). Different approaches have been developed to enhance the activity of MFC by using earthenware and clayware as a membrane (Dhulipala et al. 2020; Suransh et al. 2020). Filtration types of membrane electrodes with Prussian blue (PB) doping and PVDF-PVC-PEG triblock copolymers prepared by the phase inversion process also exhibited superior ORR activity with the highest electrochemical activity and lowest charge transfer resistance (Yu et al. 2020). Current densities could be increased by utilizing modified polyaniline (PANI) polymers, such as fluorinated PANI (Yaqoob et al. 2021). Similarly, metal-free N/B-co-doped carbon-based catalyst (denoted as PANI/B-8) developed by pyrolysis of polyaniline and boric acid mixtures showed extraordinary enhanced kinetic activity toward ORR in alkaline electrolytes. This asymmetric neutral-alkaline microbial fuel cell (ANA-MFCs) markedly delivered an output power density twice as high than the symmetric MFCs (Hu et al. 2021). Among different types of co-catalysts, ten (weight %) hydrophobic Fe-N4/AC (activated carbon) air cathodes showed a simultaneous increase in the power density and Coulombic efficiency for electricity generation (Yang et al. 2020a, b). In open-air cathode MFCs, cation transfer through the membrane reduces the cathodic redox reactions by forming thick layers of carbonate salts on the surface of the electrode (Pham et al. 2003). Wetland-microbial fuel cells (CW-MFC) have shown to be extenuating to greenhouse gases. For instance, the roots of wetland plant *Acorus Calamus* L., when placed in anode, showed better microbial ecosystem for power generation. Correspondingly, carbon fiber felt (CFF) cathode showed lowest emission of methane  $0.77 \pm 0.04 \text{ mg/(m}^2/\text{h)}$  and nitrous oxide  $130.78 \pm 13.08 \text{ }\mu\text{g/(m}^2/\text{h)}$ . The maximum power density was  $2.99 \text{ W/m}^3$ . Thus proving to be eco-friendly in mitigation of greenhouse gases (Liu et al. 2022)

Air cathodes efficiently use oxygen from the air and avoid the need for aerating water or chemical catholyte (Fan et al. 2007a). Similarly, the addition of acetylene black (AB) into exfoliated porous graphitic carbon nitride (ep-GCN) cathode catalyst indicated excellent oxygen reduction reaction activity and was less cost-effective (Chakraborty et al. 2020). Copper (II) oxide (CuO) has shown extraordinary characteristics as the electrocatalyst for ORR in the cathodic chamber. A few advantages of using CuO are high specific surface area, high catalytic activity and easy synthesis, environmentally friendly, and good redox potential (Yadav et al. 2020b). On the other hand, they have a weak adsorption property that is overcome by heat treatment by immobilizing CuO particles on the electrode surface (Li et al. 2020b). Promising results were obtained with CuO as an electrocatalyst in removing caffeine waste and electricity generation. Results revealed that the CuO/C cathode achieved the highest caffeine removal (97.67%) and maximum power density ( $28.75 \text{ mW/m}^2$ ) under aerated conditions. The maximum power density and current density increased up to 51.79% and 36.84%, respectively, thus proving its economic performance (Yap et al. 2020). A consortium of microbial communities from various habitats is becoming a choice in replacing expensive platinum as a cathode catalyst in MFCs. Because of their low cost, environmental friendliness, and long-term sustainability, microbial biocathodes are gaining popularity. A comparative study for treating waste-activated sludge and power generation using MFC was elucidated in the anodic microbial consortium. The MFCs were supplied with two feed sludge matrices of freezing/thawing (F/T) liquid versus fermentation liquor for exploring cooperative interactions in anodic microbial consortia of MFCs. The F/T liquid cultivated main genera of *Azospira*, *Poivalibacter*, *Thauera*, *Terrimonas*, *Alicyclophilus*, *Dokdonella* and *Simplicispira*; the fermented liquor was enriched with *Phenylobacterium*, *Cellulomonas*, *Edaphobacter*, *Burkholderia*, *Clostridium*, *Sphingomonas*, *Leifsonia*, and *Microbacterium* in anodic biofilm. The study showed anodic fermentative bacteria in synergy with exoelectrogens microbial diversity, and larger functional genes played a collective role in more power generation through MFCs. The optimal power density of 0.152 and 0.182  $\text{mW/m}^2$  were produced from sludge F/T liquid and fermentation liquor (Xin and Qiu 2020).

### 9.4.2 Anode Electrode

The efficacy of electricity generation at the anode electrode depends on the material used as an electrode. Anode primarily serves as a current collector while providing a surface for biofilm development (Sarathi and Nahm 2013). Carbonaceous materials, stainless steel, copper, nickel, silver, gold, and titanium have been used as anode electrodes because they are highly stable. While the drawback of these metals is that they suffer from less electro-catalytic activity toward the redox reaction, and the metal ions could be poisonous to microbes, thus hindering the performance of MFC. This in turn reduces the degradation competence of the MFC (Suresh et al. 2022). The commonly used anode material is carbon in its various forms and configurations such as carbon-brush, felt, fiber, granule, mesh, nanotube, paper, plate, rod, graphite

embedded stainless steel frame, and titanium plates coated with mixed metal oxide. The implementation of anode surface modification by nanostructured materials has been extensively studied. The nanocatalyst has shown significant performance in the transfer of electrons to the electrode, enhancing the surface area to enrich bacteria adhesion and greater resistance against fouling (Li et al. 2019). For power generation, nanocatalysts such as iron oxide (FeO), iron (II) molybdate (FeMoO<sub>4</sub>), transition metal oxides or carbides such as ruthenium oxide (RuO<sub>2</sub>), manganese oxide (MnO<sub>2</sub>), and molybdenum have been used as electrodes (Yamashita and Yokoyama 2018). Scientist have also tried dual metal organic frameworks (D-MOFs), TiO<sub>2</sub>@ZIF-67/ZIF-8 composite (Zeolitic imidazolate frameworks). The maximal power density of TiO<sub>2</sub>@ZIF-67/ZIF-8 microbial fuel cell (MFC) was 341.506 mW/m<sup>2</sup> and continuous output voltage was 413.43 mV. The power density was 1.30 times higher ZIF-67/ZIF-8-MFC and 2.07 times of ZIF-67-MFC (164.836 mW/m<sup>2</sup>). The framework was able to maintain stable voltage output for 8.3 days (Yang et al. 2022). A novel anode electrocatalysts iron (II) molybdate coated on the graphite plate showed a fivefold reduction in resistance and a threefold increase in redox current. The power density (106.2 mW/m<sup>2</sup>) achieved was 1.4-folds higher than control electrodes. Considering the economy and high-performance FeMoO<sub>4</sub>, it can be successfully developed for enhancing bioelectricity generation in the MFC (Mohamed et al. 2020a). Graphene is used as both anode and cathode materials. As an anode, it improves the deficiency of electron transfer and bacterial attachment. When used as a cathode material, it supports the oxygen reduction reaction (Olabi et al. 2020). Chemically reduced graphene oxide (CGO) prepared using L-cysteine is considered the best choice as an anode electrode because of its high electrical conductivity, high surface area, great flexibility, and excellent mechanical properties (Pareek et al. 2019). Likewise, electrophoretic deposition of graphene oxide on the surface of carbon brush as anode significantly increased power density from 33 to 381 mW/m<sup>2</sup>, thus enhancing the performance and Coulombic efficiency of the MFC. Studies by Yaqoob et al. (2022) have shown anode electrodes consisting of graphene oxide (GO) and GO-polymer-metal oxide (GO-PANI-Ag) high productivity of 1.022 mW/m<sup>2</sup> and GO-PANI-Ag at 2.09 mW/m<sup>2</sup>. The biomass for this study was provided with oil palm trunk sap as organic substrate. The MFC was able to remove heavy metals such as Cd(II) (80.25%) and Pb(II) (78.10%). Polyaniline functionalized activated carbon (PANI-FAC) composite as a capacitive anode coated with stainless steel mesh improved the maximum power density to 322 mW/m<sup>2</sup> (Yellappa et al. 2020). The NiFe<sub>2</sub>O<sub>4</sub>-MXene@CF (Carbon felt) anode was considered preferable because of its low charge transfer resistance, high conductivity, and a large number of catalytically active sites. The power density was improved to 1385 mW/m<sup>2</sup> (Tahir et al. 2020). Similarly, polymerized nanofiber polyaniline (PANI) for carbon felt (CF) electrodes aimed at increasing the conductivity of the anodic electrode facilitated the adherence of exoelectrogenic yeast cells of *Cystobasidium slooffiae* JSUX1. This further improved bioelectricity generation in MFCs from using xylose as the substrate (Soni et al. 2020). An increased surface area of nanofiber PANI boosted the conductivity of the PANI/CF anode for a robust attachment of *C. slooffiae* JSUX1 to form a dense biofilm. The authors reported with

PANI/CF it was possible to achieve a derived power output about 2.2 times ( $119.35 \pm 3.27 \text{ mW/m}^2$ ) that of CF only ( $50.41 \times 6.9 \text{ mW/m}^2$ ). The maximum hydrogen yield was 25.83 mL (Moradian et al. 2022). Bioanode electrode synthesized using graphene oxide deposited on the surface of the carbon brush showed enhanced electron transfer rate and the bioactive surface area. The maximum power and current densities increased more than 10 and 6 times, and the columbic efficiency increased by 12 times when operated with waste water (Sayed et al. 2021).

### 9.4.3 Membranes

The use of membranes has its own merits and demerits. In addition to the high cost of membranes, MFC performance can be compromised by biofilm formation, fouling on the membrane surface, and increased oxygen permeability (Logan 2008; Choi et al. 2013). A variety of membranes are garnering renewed attention for use in MFC to facilitate the transport of protons from the anode to the cathode. Irrespective of the membrane material, they should have some key features such as (1) preventing direct electrical interaction between anodes and cathodes; (2) reducing the undesired crossover of oxygen and other substances; (3) maintaining effective transport of proton mass through the separator; (4) low internal resistance; (5) low mass transfer between oxygen-containing water of cathode and anaerobic anode; (6) high proton conductivity; (7) high energy recovery; (8) high ionic conductivity; (9) and long-term stability (Daud et al. 2015; Yang et al. 2019). Although the elimination of membrane has its advantage, the relatively broad electrode spacing leads to high internal resistance and restricts the electrode surface area and power density ratio. Therefore, further reduction in electrode spacing is required (Cheng et al. 2006). Membranes are classified based on their porous/nonporous nature. Nonporous membranes are subdivided into a cation exchange membrane (CEM), anion exchange membrane (AEM), and bipolar membrane (BPM). Porous membranes are categorized into UFM, MFM, and CMs (not within the scope of discussion).

*Ion exchange membranes* (IEMs) are a class of polymeric membranes containing highly swollen gels carrying fixed positive or negative charges. Ion-exchange membranes are permeable to ions of opposite charge (counter ions), but repel ions of the same charge (co-ions). The only exception is the protons (Luo et al. 2018). IEM has better selectivity, lower electrical resistance, and improved thermal, chemical, and mechanical properties. IEMs are categorized as cation exchange membranes (CEM) or PEM and anion exchange membranes (AEM) where the protons can permeate freely (Daud et al. 2015).

#### 9.4.3.1 Cation Exchange Membrane (CEM)

CEM is designed to allow the transfer of protons and cations through a membrane resulting in a net negative charge (Harnisch and Schröder 2010). Flat plate type MFC with Nafion PEM and anode assembly provides a larger surface area for the membrane and cathode (Kumar et al. 2017). Nafion, a perfluorosulfonic acid polymer, is an excellent choice as a proton exchange membrane because it has good



proton conductivity and chemical stability. The oxygen permeability through these membranes can reduce the Coulombic efficiency of the MFC (des Roches and Omiya 2014). Proton conducting membrane devices such as PEMFCs (Polymer electrolyte membrane fuel cells) and DMFCs (Direct methanol fuel cells) work better with the Nafion-based operation at low temperatures ( $<80\text{ }^{\circ}\text{C}$ ). Whereas at higher temperatures ( $120\text{--}200\text{ }^{\circ}\text{C}$ ), high-temperature hydrocarbon polymers poly (phenoxyphosphazene) (POP), sulfonated naphthalic polyimide, polybenzimidazole (PBI), alkyl sulfonated polybenzimidazole (PBI-AS), sulfonated poly (arylene ether ether ketone) (PEEK-SO<sub>3</sub>H), and sulfonated poly (arylene sulfone) (PSU-SO<sub>3</sub>H) are used (Shi 2014). Nafion-based PEM suffers from extreme biofouling; hence it is being replaced by non-fluorinate sulfonated membranes, which come at lower cost and higher energy recovery (Shabani et al. 2020). Fabricated ceramic separators like clayware ceramic pots used to treat rice-mill wastewater produced a power density of  $2.3\text{ W/m}^3$  (Behera and Ghangrekar 2011), and earthen CEM produced a maximum power output of  $16.8\text{ W/m}^3$  (Bhaduri and Behera 2023). Similarly, the terracotta flowerpot generated a maximum volumetric power density of  $14.59\text{ W/m}^3$ , which was 46% higher than Nafion as a PEM (Jana et al. 2012). Taşkan (2020) obtained a maximum power density  $26,680\text{ mW/m}^2$  and oxygen pressure of 10 psi with a sandwich-type microbial fuel cell having three chambers (2 anodes and 1 cathode) with a hollow fiber gas transfer membrane aerated cathode. Branched polyethyleneimine membrane (BPEI) has been shown to increase membrane permeability, improve mediator access to electron carriers and biofilm formation at the anode in the presence of *E. coli*, and neutral red as the mediator, the power densities generated were  $2.6\text{ mW/m}^2$  (Soh et al. 2020). Nanocomposite membranes are the other alternative for PEM. PEM used in fuel cells should possess the following properties: high proton conductivity, good mechanical strength, excellent chemical resistance, and good durability (Zakaria et al. 2016).

#### 9.4.3.2 Anion Exchange Membrane (AEM)

In an AEM, hydroxide ions are transferred from the cathode to the anode through the anion conducting polymer electrolyte, where it combines with hydrogen to form water during electrochemical oxygen reduction at the cathode to produce  $\text{OH}^-$ . In polymeric AEM, there is no liquid phase the positive charges, such as phosphate or carbonate, attached to the membranes facilitate the proton transfer by applying proton carriers (pH buffers) (Fan et al. 2007b). The ideal polymer for AEMs must have excellent  $\text{OH}^-$  conductivity, chemical and thermal stability, strength, flexibility, low gas permeability, low water drag, low cost, and good availability. The possible fuels used in AEM are hydrogen, methanol, ethanol, propanol, ethylene glycol, and sodium borohydride. AEMs suffer from poor solubility in low boiling solvents, chemical instability and low ionic conductivity. The synthesis of AEM is complex as it involves chloromethylation, which is a potent carcinogen (Hren et al. 2021).

#### 9.4.3.3 Bipolar Membranes (BPM)

A bipolar membrane is a double-layer structure comprising a cation exchange membrane and an anion exchange membrane, directly attached to one another. It

also has an interfacial layer where water dissociation occurs. The double layer enables the transport of protons and hydroxyl ion, and block co-ions. The innovation of these membranes is the separation of mono- and divalent ions, anti-deposition, anti-fouling, and water dissociation. However, in the use of such membranes, a pH gradient is the main concern (Kim et al. 2017). The bipolar membrane provides physical support for the embedded electrode. It minimizes electrode thickness, thereby reducing the distance between the structure which supplies protons and the electrode, thus minimizing ohmic losses (Mayerhöfer et al. 2020).

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## 9.5 Factors Responsible That Affect Performance of Microbial Fuel Cell

### 9.5.1 Effect of pH, Ionic Strength, and Temperature on Power Generation

By adding NaCl, Liu et al. (2015) noticed an increasing ionic strength of the solution from 100 to 400 mM, and the power output increased from 720 to 1330 mW/m<sup>2</sup>. This was perhaps because of the increased fluid access in the chamber with holes on both sides of the anode electrode and higher Pt content on the cathode (0.5 mg Pt/cm<sup>2</sup>). *Shewanella marisflavi* strain EP1 could generate a power density of 9.6 mW/m<sup>2</sup> when ionic strength was increased to 1146 mM (8% NaCl). Due to a reduction in internal resistance, increasing the ionic strength of the electrolyte significantly enhanced power output (Huang et al. 2010). Miyahara et al. (2015) observed the abundance of *Geobacter* bacteria increased when the NaCl concentration increased from 0 to 0.1 M but markedly reduced when the NaCl concentration was increased to 0.3 M due to intolerance. This indicated a strong correlation between the bacteria, ionic strength, and power output. Most reviewed studies reported that power density and temperature were exponential rather than linear. The influence of temperature had only a negligible effect in most of the studies suggesting the maximum power output drops at lower temperatures (10 °C) or higher temperatures of (55 °C) (Li et al. 2013). This is because MFCs cannot operate at extremely low temperatures because microbial reactions are sluggish at low temperatures or denatured at higher temperatures.

Nevertheless, a short-side-chain Hyflon<sup>®</sup> perfluorinated ionomer-based membrane produced a power density of 300 mW/cm<sup>2</sup> at 140 °C in the presence of 1 M methanol and air fed (Baglio et al. 2006). Wastewater-fed reactors were less susceptible to temperature than acetate-fed reactors (Heidrich et al. 2018). Although there is a contradiction with respect to the ideal operating pH in MFC, the most frequently mentioned is neutral pH (Borole et al. 2008). Low pH (<6) showed an adverse effect on the electrochemically active bacterial population resulting in a drastic fall in power output. Also, proton production is mainly related to the electrochemical oxidation of the organic fuels at the anode (Zhang et al. 2013).

## 9.5.2 Microbes as Biocatalyst Used in MFC

Inoculum selection, enrichment, operating conditions, and cell architecture impact the MFC reactor's start-up phase (Kumar et al. 2018). Bacteria generate electrical energy by the oxidation of organic matter and transfer the electrons to an electron acceptor outside of their cells; hence they are termed as "Exoelectrogens." These microbes can transfer the electrons directly from the cytoplasmic membrane to electron acceptors such as insoluble and soluble metals, flavins, or electrodes (Wu et al. 2013). The electrogenic bacteria only prefer non-fermentable substrate acetate and are capable of completely oxidizing acetate, whereas the fermentative bacteria convert carbohydrates into short-chain fatty acids and acetate (Yang et al. 2015). Proteobacteria ( $\alpha$ -proteobacteria,  $\beta$ -proteobacteria,  $\gamma$ -proteobacteria and  $\delta$ -proteobacteria) have the ability to directly transfer electrons to the electrode and represent the largest category of electricigens. Other bacteria used in MFC are archaea, cyanobacteria, firmicutes, yeast, and eukaryotic algae, which can oxidize organic compounds and transfer electrons to the anode (Cao et al. 2019). Primitive prokaryotes (Archaeobacteria) that can survive extreme conditions have been tested as possible sources for electricigens when complex compounds have to be degraded. Two halophilic archaea, *Haloferaxvolcanii* and *Natrialbamagadii*, used as a biocatalyst at the anode, were evaluated for electricity generation. Maximum power densities of 50.98 and 5.39  $\mu\text{W}/\text{cm}^2$  were obtained, which was higher when compared to mediator-less MFCs (Abrevaya et al. 2011). The exoelectrogenic bacteria preferably used in MFCs are dissimilatory metal-reducing bacteria such as *Geobacter* and *Shewanella* (Proteobacteria) referred to as metal-reducing microbes since they reduce the solid metal oxides (Cao et al. 2019). According to Bond and Lovley (2005), *Geobacter sulfurreducens* and *Rhodospirillum rubrum* could produce electricity by forming a monolayer directly on the anode electrode surface and use this as their end terminal electron acceptor in anaerobic respiration; hence, these microorganisms are also called anodophiles. Sulfur-reducers, especially *Desulfuromonas* and *Desulfovibrio*, could convert sulfate to sulfide, which is then oxidized to elemental sulfur and can be reduced again to sulfide. *Geobacter* species have several possible advantages over *Shewanella* species. *Shewanella* species incompletely oxidize a limited range of organic acids to acetate, inefficient since most of the electrons present in the initial fuel remain as acetate. *Shewanella* species appear to transfer electrons to anodes by releasing a soluble molecule that acts as an electron shuttle. On the other hand, *Geobacter* species can completely oxidize organic compounds to carbon dioxide with the recovery of >90% of the electrons available in the fuels as electricity (Bond et al. 2002; Kumar et al. 2019). *Geothrix fermentans* are iron-reducing acid bacteria capable of producing electron mediators that facilitate reduction reactions in graphite electrodes (Bond and Lovley 2005). A new model for nitrogen removal and power production was developed using MFCs with nitrite as an electron acceptor in the cathode (Jin et al. 2018). A novel denitrifying exoelectrogenic *Mycobacterium* sp. EB-1 revealed the strain was capable of producing electricity by direct electron transfer. Mutant strains of *S. oneidensis* and *S. putrefaciens* have shown improved performance and good

bacterial adhesion to the electrode, enhancing power generation. *S. oneidensis* MR-1 was constructed using the *yde H* gene from *E. coli* under the control of an IPTG-inducible promoter, and the strain *yde H* itself was under the control of a constitutive promoter. The recombinant *Shewanella* strains showed significant enhancement in biofilm formation and bioelectricity generation, which was about 2.8-fold of the original strain (Liu et al. 2015).

Mixed cultures were demonstrated to be beneficial compared to pure cultures due to the presence of different kinds of bacteria along with electricigens providing a high power density. Mixed culture minimizes the effects of oxygen diffusion into the anode chamber by scavenging any dissolved oxygen and maintaining anaerobic conditions in the anode chamber (Rabaey et al. 2004). Any oxygen diffusion into the system will result in substrate loss and reduced Coulombic efficiencies (Min et al. 2005). Activated sludge, anaerobic sludge, and domestic wastewater are excellent examples of mixed inoculum, including fermentative or methanogenic microorganisms carrying initial metabolism (Rout et al. 2020). A recent study also proved mixed, or co-culture of *Escherichia coli* and *Pseudomonas aeruginosa* generated a maximum power density of  $190.44 \text{ mW m}^{-2}$ , which was comparatively higher when the organism was used individually. The study further proved co-cultures when coupled with *Chlorella vulgaris* a synergistic effect was observed that improved mean power density from  $248 \text{ mW/m}^2$ , a 41.7% rise (Aiyer 2021). Khan et al. (2022) observed live diatoms (*Nitzschia palea*) in the anodic chamber could replace bacterial cell in generating electricity. Photosynthetic diatom microbial fuel cell (PDMFC) was supplied with f/2 media rich in nitrates, phosphates, metasilicates, trace metals, and vitamins as the anolyte. The maximum derived power output was  $12.62 \text{ mW/m}^2$  and coulombic efficiency of 22.95%. Besides the diatom cells showed about 64.28% increase in lipid production on 15th day compared to the 1st day. This was accompanied by formation of complex fatty acid methyl esters and carotenoids. Table 9.1 provides the list of biocatalysts, substrates, and electrodes involved in bioelectricity generation.

### 9.5.3 Organic Waste as Microbial Substrate

A great variety of substrates have experimented with high current production in MFC. Owing to the poor conversion of nutrients, the use of solid organic waste for electricity generation has drawbacks. Therefore, the nutrients need to be converted into monomers before being fed to the microbial cells. Enzymatic hydrolysis has been used to overcome the problem mentioned above (Ma et al. 2016). Increasing the substrate concentrations from 100 to 850 mg/L boosted the power output from 0.2 to  $1.2 \text{ W/m}^3$ ; however, concentrations higher than the above-mentioned were not beneficial (Jiang and Li 2009). Depending on the particular application for which an MFC is used, the metabolic substrate needed for electrogenic bacteria should be carefully selected, as not all electrogenic bacteria can completely oxidize multiple substrates. The substrate used includes carbohydrate (glucose, sucrose, maltose, galactose, fructose, sucrose, xylose, trehalose, rhamnose, cellulose, dextran), organic

**Table 9.1** List of microorganism substrates and electrodes used for bioelectricity generation in MFC

Organism	Type	Current density	Power density	Substrate	Cathode/anode electrode	References
<i>Acidiphilium cryptum</i>	Proteobacteria ( $\alpha$ )	–	12.7 mW/m <sup>2</sup>	Glucose	Anode: graphite (2.5 × 7.5 × 0.63 cm) felt connected to graphite rod. Cathode: platinum (5 × 5 cm) deposited carbon cloth	Borole et al. (2008)
<i>Acidiphilium</i> sp. strain 3.2 Sup 5	Proteobacteria ( $\alpha$ )	3 A/m <sup>2</sup>	–	Glucose	Anode and cathode: graphite felt	Malki et al. (2008)
<i>Actinobacillus succinogenes</i>	Proteobacteria ( $\gamma$ )	2.7 mA	348.6 mW	Glycerol	Anode and cathode: graphite	Zheng et al. (2020)
<i>Aeromonas hydrophila</i>	Proteobacteria ( $\gamma$ )	1.8 mA	–	Acetate	–	Pham et al. (2003)
<i>Aeromonas hydrophila</i>	Proteobacteria ( $\gamma$ )	8.77 mA/cm <sup>2</sup>	–	Chitin	Anode and cathode: carbon felt (3 cm × 3 cm × 3 mm)	Park et al. (2008)
<i>Aeromonas</i> sp. strain ISO2-3	–	–	800 mW/m <sup>2</sup>	Glucose	Graphite	Chung and Okabe (2009)
<i>Alcaligenes faecalis</i>	Proteobacteria	–	90 W/m <sup>3</sup>	Acetate and glucose	Anode and cathode: granular graphite matrix	Rabaey et al. (2004)
<i>Enterococcus gallinarum</i>	Mixed culture	–	644 mV	Dairy waste	Anode and cathode: copper electrode	Sanjay and Udayashankara (2020)
<i>Pseudomonas aeruginosa</i>						
Anaerobic and facultative microbes	Mixed culture	–	29.96 mW/m <sup>2</sup>	Bakery waste	Two-stage bioprocess method	Han et al. (2020)
Anaerobic sludge	Mixed culture	150 mA/m <sup>2</sup>	–	Solid potato waste	Anode and cathode: graphite electrode	Du et al. (2020)
Anaerobic microbes	Mixed culture	–	0.040–0.044 W/m <sup>2</sup>	Kitchen waste	Anode: stainless steel mesh Cathode: graphite plate	Dhulipala et al. (2020)

(continued)

Table 9.1 (continued)

Organism	Type	Current density	Power density	Substrate	Cathode/anode electrode	References
<i>Arcobacter butzleri</i> strain ED-1	Proteobacteria ( $\epsilon$ )	–	296 mW/L	Acetate	Anode and cathode: graphite felt with semidry cathodes	Fedorovich et al. (2009)
<i>Azoarcus</i> sp. and <i>Desulfuromonas</i> sp.	Proteobacteria ( $\delta$ )	–	488 mW/m <sup>2</sup>	Ethanol	Anode: plain porous carbon Cathode: carbon paper incorporating Pt catalyst	Kim et al. (2017)
<i>Bacillus subtilis</i>	Firmicutes	–	1.05 mW/cm <sup>2</sup>	Glucose	Anode: carbon cloth Cathode: platinum	Nimje et al. (2009), Kashyap et al. 2019
<i>Bacillus</i> , <i>Klebsiella</i> , and <i>Enterobacter</i> species	Firmicutes	–	0.0744 mW/m <sup>2</sup>	Yam ( <i>Dioscorea alata</i> ) waste	–	Fadzli et al. (2021)
<i>Bacteroidetes</i> , and <i>Proteobacteria</i>	Firmicutes	–	610 mW/m <sup>2</sup>	Citric acid waste	Dual-chamber MFC	Zhang et al. (2021)
<i>Citrobacter</i> sp. SX-1	Proteobacteria ( $\gamma$ )	58 mA/m <sup>2</sup>	88.1 mW/m <sup>2</sup>	Citrate, glycerol, sucrose	Anode and cathode: carbon cloth	Xu and Liu (2011)
<i>Clostridium beijerinckii</i>	Firmicutes	1.3 mA/cm <sup>2</sup>	–	Starch, glucose	Anode and cathode: woven graphite fuel cell cathode and anode	Niessen et al. (2004)
<i>Clostridium beijerinckii</i> SR1	Firmicutes	–	61.5 mW/m <sup>2</sup>	Sago hampas	Anode: carbon cloth (1.5 × 1.5 cm) cathode: 20% platinum on Vulcan carbon cloth	Jenol et al. (2020)
<i>Clostridium butyricum</i> EG3	Firmicutes	0.22 mA	–	Starch and glucose	Anode and cathode: graphite	Park et al. (2001)

<i>Comamonas denitrificans</i>	Proteobacteria (β)	–	35 mW/m <sup>2</sup>	Acetate	Anode: carbon paper or a graphite fiber brush	Xing et al. (2010)
					Cathode: Pt with PTFE diffusion layers on 30 wt% wet-proofed carbon cloth	
<i>Cupriavidus basilensis</i>	Proteobacteria (β)	902 mA/m <sup>2</sup>	44 mW/m <sup>2</sup>	Acetate	Anode: graphite rod	Friman et al. (2013)
<i>Desulfotulbus propionicus</i>	Proteobacteria (δ)	28.35 mA/m <sup>2</sup>	–	Fumarate, lactic, pyruvic, propionic acid	Cathode: carbon cloth	Holmes et al. (2004)
<i>Desulfotulvibrio desulfuricans</i>	Proteobacteria (δ)	233 mA/m <sup>2</sup>	–	–	Anode and cathode: graphite electrodes	Kang et al. (2014)
<i>Dysgonomonas oryzae</i>	Bacteroidetes	50 μA/cm <sup>2</sup>	–	Acetate, lactate	Cassette-electrode MFC	Kodama et al. (2012)
<i>Enterobacter cloacae</i>	Proteobacteria (γ)	493.8 mA/m <sup>2</sup>	4.9 mW/m <sup>2</sup>	Cellulose, sucrose, glycerol	Anode and cathode: carbon cloth	Rezaei et al. (2009)
<i>Enterobacter ludwigii</i>	Proteobacteria (γ)	440 mA/m <sup>2</sup>	–	Citrate, acetate and cellulose	Anode and cathode: carbon cloth	Feng et al. (2014)
<i>Escherichia coli</i>	Proteobacteria (γ)	–	600 mW/m <sup>2</sup>	Glucose	Anode: carbon/PTFE composite. Cathode: Nafion+Pt/C gas diffusion layer	Zhang et al. (2006)
<i>Escherichia coli</i>	Proteobacteria (γ)	–	120–140 mW/m <sup>2</sup>	Green bean sprouts	–	Mulyono (2020)
<i>Escherichia coli</i> K12	Proteobacteria (γ)	1.45 mA/cm <sup>2</sup>	6000 mW/m <sup>2</sup>	Glucose, sucrose	Anode: graphite cloth (30 × 25 mm) Cathode: graphite	Schröder et al. (2003)

(continued)

Table 9.1 (continued)

Organism	Type	Current density	Power density	Substrate	Cathode/anode electrode	References
<i>Escherichia coli</i>	Proteobacteria ( $\gamma$ )	1750 mA/m <sup>2</sup>	788 mW/m <sup>2</sup>	Glucose	Anode: Mn <sup>4+</sup> -graphite	Park and Zeikus (2003)
		325 mA/m <sup>2</sup>	91 mW/m <sup>2</sup>		Cathode: Fe <sup>3+</sup> -graphite	
<i>Geobacter metallireducens</i>	Proteobacteria $\delta$	–	40 mW/m <sup>2</sup>	Acetate	Anode: carbon paper Cathode: Pt catalyst	Min et al. (2005)
<i>Geobacter</i> spp.	Proteobacteria $\delta$	262 mA/m <sup>2</sup>	106 mW/m <sup>2</sup>	Sugar wastewater	Anode and cathode: graphite	Mohamed et al. (2020a)
<i>Geobacter sulfurreducens</i>	Proteobacteria $\delta$	456 mA/m <sup>2</sup>	188 mW/m <sup>2</sup>	Acetate	Anode and cathode: graphite	Nevin et al. (2008)
<i>Geobacter sulfurreducens</i>	Proteobacteria $\delta$	11,143 mA/m <sup>2</sup>	15 mW/m <sup>2</sup>	Acetate	Graphite	Bretschger et al. (2007)
<i>Geobacteria sulfurreducens</i> and <i>Shewanella oneidensis</i>	Proteobacteria $\delta$	3.74 mA/m <sup>2</sup>	45.50 $\mu$ W	Sucrose, acetate	Generation II fuel cell	Ieropoulos et al. (2005)
<i>Geobacteria sulfurreducens</i> PCA	Proteobacteria $\delta$	0.40 A	13 mW/m <sup>2</sup>	Acetate	Anode and cathode: graphite	Bond and Lovley (2003)
<i>Geopsychrobacter electrophilus</i>	Proteobacteria $\delta$	121.43 mA/m <sup>2</sup>	–	Fumarate	Anode and cathode: graphite	Holmes et al. (2004)
				Acetate, malic, fumaric and citric acid		
<i>Geothrix fermentans</i>	Acidobacteria	0.6 mA	–	Acetate	Anode and cathode: graphite electrode	Bond et al. (2002)
<i>Geothrix fermentans</i>	Acidobacteria	50 mA/m <sup>2</sup>	–	Acetate, propionate, malate, lactate, or succinate	Anode and cathode: graphite electrode	Bond and Lovley (2005)
<i>Gluconobacter oxydans</i>	Proteobacteria ( $\alpha$ )	–	7.23 mW	Glucose	Anode and cathode: cylinder graphite electrodes	Reshetilov et al. (2006)



<i>Haloflex volcanii</i>	Archaeobacteria	49.67 $\mu\text{A}/\text{cm}^2$	11.87 $\mu\text{W}/\text{cm}^2$	Acetate	Anode and cathode: plain carbon paper TGP-H-030 (Toray®, Tacoma, WA)	Abrevaya et al. (2011)
<i>Klebsiella pneumoniae</i> L17	Proteobacteria ( $\gamma$ )	0.08 mA	218.51 $\text{mW}/\text{m}^2$	Glucose, starch, lactic acid, lactate, fructose, sucrose, lactose, and maltose	Anode and cathode electrodes: carbon felt ( $4.5 \times 4.0 \text{ cm}$ )	Zhang et al. (2008)
<i>Lactobacillus plantarum</i>	Firmicutes	–	0.22 MW	Glucose	Anode: carbon cloth cathode: electrode platinum	Vega and Fernández (1987)
<i>Lactobacillus</i> , <i>Clostridium</i> <i>sensu stricto</i> and <i>Bacteroides</i>	Firmicutes	–	–	Watermelon rind	Membraneless biocathode microbial fuel cell (MB-MFC)	Yang et al. (2021a)
<i>Lactococcus lactis</i>	Firmicutes	3 $\text{A}/\text{m}^3$	–	Lactate, acetate	Anode: glass cylinder Cathode: Pt wire	Freguia et al. (2009)
<i>Lysinibacillus sphaericus</i>	Firmicutes	$\approx 270 \text{ mA}/\text{m}^2$	85 $\text{mW}/\text{m}^2$	Protein components	Anode and cathode: graphite felt	Nandy et al. (2013)
<i>Ochrobactrum anthropi</i> YZ-1	Proteobacteria ( $\alpha$ )	1027 $\text{mA}/\text{m}^2$	89 $\text{mW}/\text{m}^2$	Acetate	Anode: ammonia gas pre-treated plain carbon cloth Cathode: graphite fibers	Zuo et al. (2008)
Mixed microbial consortia	–	168.05 $\text{mA}/\text{m}^2$	2.01 $\text{W}/\text{m}^3$	Pharmaceutical waste	The paraboloid shape MFC Anode and cathode: graphite	Rashid et al. (2021)
Marine sediment sludge	–	–	2.08 mW	Orange peel waste	Multiple single solid phase MFC	Hariti et al. (2021)

(continued)

Table 9.1 (continued)

Organism	Type	Current density	Power density	Substrate	Cathode/anode electrode	References
Mixed culture Ochrobactrum (53%), Marinobacter (22%) and Rhodococcus (15%). Bacillus, Stenotrophomonas, Xanthobacter, Sphingomonas, Pseudomonas and Sedimentibacter	–	–	369 mW/m <sup>2</sup>	Aquaculture wastewater	Saline anode microbial fuel cell (SA-MFC) Anode and cathode: carbon felt separated by nafion	Pugazhendi et al. (2021)
<i>Propionibacterium freudenreichii</i> ET-3	Actinobacteria	–	–	Glucose	Anode and cathode: carbon Felt	Wang et al. (2008)
<i>Proteus mirabilis</i>	Proteobacteria ( $\gamma$ )	6 mA	–	Glucose	Anode: reticulated vitreous carbon (35 × 50 × 7 mm) Cathode: bright platinum foil (10 × 40 mm)	Thurston et al. (1985)
<i>Proteus vulgaris</i>	Proteobacteria ( $\gamma$ )	0.4 mA	–	Galactose	Anode: reticulated vitreous carbon Cathode: platinum (40 × 40 × 1 mm)	Kim et al. (2000)
<i>Rhodobacter sphaeroides</i>	Proteobacteria ( $\alpha$ )	–	790 mW/m <sup>2</sup>	Sistrom's minimal medium (nitrogen, succinate)	Anode and cathode: platinum-coated carbon paper	Cho et al. (2008)
<i>Rhodococcus pyridinivorans</i> HR-1	Proteobacteria ( $\alpha$ )	2.309 A/m <sup>2</sup>	0.336 W/m <sup>2</sup>	Acetate	Anode: unilaminar carbon cloth cathode: Pt/C	Cheng et al. (2020)
<i>Rhodoferrax ferrireducens</i>	Proteobacteria ( $\alpha$ )	74 mA/m <sup>2</sup>	33 mW/m <sup>2</sup>	Glucose, fructose, sucrose	Anode and cathode: graphite felt/rod/porous	Chaudhuri and Lovley (2003)

<i>Rhodospseudomonas palustris</i> DX-1	Proteobacteria (α)	0.99 mA/cm <sup>2</sup>	2780 mW/m <sup>2</sup>	Volatile acids, yeast extract, and thiosulfate	Anode: carbon paper		Xing et al. (2008)
					Cathodes Pt and carbon cloth		
<i>Rhodospirillum rubrum</i>	Proteobacteria (α)	–	1.25 W/m <sup>2</sup>	Light	Anode: carbon-based		Gomez et al. (2014)
					Cathode: chamber-stainless steel mesh		
<i>Shewanella algae</i> (MTCC-10608)	Proteobacteria (γ)	141 mA/m <sup>2</sup>	50 mW/m <sup>2</sup>	Dairy wastewater	Anode and cathode: acrylic (single chamber MFC)		Choudhury et al. (2021)
<i>Shewanella marisflavi</i> BBL25	Proteobacteria (γ)	6.850 mA/cm <sup>2</sup>	52.80 mW/cm <sup>2</sup>	Barley straw Miscanthus, Pine hydrolysate (Lignocellulose, glucose)	Anode: carbon felt		Gurav et al. (2020)
		6.661 mA/cm <sup>2</sup>	40.95 mW/cm <sup>2</sup>		Cathode platinum-coated carbon felt		
		6.294 mA/cm <sup>2</sup>	34.05 mW/cm <sup>2</sup>				
<i>Shewanella oneidensis</i> DSP10	Proteobacteria (γ)	100 mA/m <sup>2</sup>	24 mW/m <sup>2</sup>	Lactate	Anode and cathode: glassy carbon		El-Naggar et al. (2008)
<i>Shewanella oneidensis</i> MR-1	Proteobacteria (γ)	1100 mA/m <sup>2</sup>	167.6 mW/m <sup>2</sup>	Lactate	Anode and cathode: carbon cloth (2.5 cm × 2.5 cm)		Liu et al. (2015)
<i>Shewanella putrefaciens</i>	Proteobacteria (γ)	0.031 mA	0.19 mW/m <sup>2</sup>	Lactate	Anode: woven graphite		Kim et al. (1999)
<i>Shewanella putrefaciens</i>	Proteobacteria (γ)	312.5 mA/m <sup>2</sup>	10.2 mW/m <sup>2</sup>	Lactate, pyruvate, acetate, glucose	Anode and cathode: graphite		Park and Zeikus (2003)
					Anode and cathode: graphite rod		
<i>Spirulina platensis</i>	Proteobacteria (γ)	400 mA/m <sup>2</sup>	98 mW/m <sup>2</sup>	Cafeteria waste	Anode and cathode: graphite rod		Christwardana et al. (2020)
<i>Synechococcus</i> sp. and <i>Chlorococcum</i> sp.	Cyanobacteria	260 mA/m <sup>2</sup>	41.5 mW/m <sup>2</sup>	Kitchen waste	Anode and cathode: graphite electrode		Mohamed et al. (2020b)
		534 mA/m <sup>2</sup>	30.2 mW/m <sup>2</sup>				

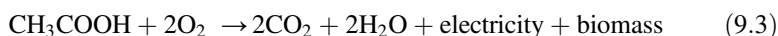
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Table 9.1 (continued)

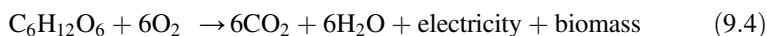
Organism	Type	Current density	Power density	Substrate	Cathode/anode electrode	References
Thauer, Nitrosomonas Desulfomicrobium Thiobacillus	Denitrifying bacteria	0.48 mA/cm <sup>2</sup>	1250 mW/m <sup>2</sup>	Acetate	Anode: carbon felts (3.0 cm × 3.0 cm). Cathode: manganese-based catalyzed carbon E4 air Cathode	Yang et al. (2019)
<i>Thermincola potens</i> strain JR	Firmicutes	50 mA/m <sup>2</sup>		Acetate	Anode and cathode: graphite blocks or graphite carbon fiber	Wrighton et al. (2011)
Thiobacillus, Afipia, Devosia Ignavi bacterium and Anaerolineaceae	Denitrifying bacteria and Proteobacteria	18–19 A/m <sup>3</sup>	0.518–0.594 W/m <sup>3</sup>	Acetate	Anode and cathode: graphite electrode	Zhao et al. (2016)
<i>Xanthomonas translucens</i> in synergistic with <i>Staphylococcus saprophyticus</i> ICBB 9554	Proteobacteria	–	0.33 mW/m <sup>2</sup>	Rice straw (cellulose)	Anode and cathode: carbon fiber	Khoirunnisa et al. (2020)

acids (acetate, butyrate, lactate, propionate, malate, succinate) (Bond and Lovley 2005), amino acids (serine, glycine, asparagine, aspartic acid, alanine, lysine, histidine, arginine), alcohols (methanol, glycerol, ethanol), inorganic compounds (sulfate, dye), and complex substrates (peptone, pectin, chitin, yeast extract, molasses) (Hu 2008; Lee et al. 2008; Chae et al. 2009) from waste waters, food waste, green waste, wood waste, brewery wastewater, industrial waste, sewage sludge, animal manure, slaughter houses, agriculture biomass, seafood biomass, food processing waste (Pant et al. 2010; Palanisamy et al. 2019; Hosur et al. 2020). The synergy between fermentative and electrogenic bacteria becomes a priority when a complex substrate is fed to MFCs. Using more complex substrates in combination resulted in a lower utilization rate and efficiency (Xiao and He 2014). The majority of the analyzed studies used acetate as a substrate to fuel the MFC, and the response was mixed. The power generation was higher in acetate-fed systems than in those produced with butyrate, propionate, and glucose, probably because of high degree of oxidation and energy efficiency in acetate (Yang et al. 2015). Bacteria in MFCs oxidize organic substrates, such as acetate, glucose, lignocellulose, and other sugars to produce electrons. The oxidation reaction is carried out by the anode, whereas the reduction process is carried out by the cathode (Eq. 9.3).

The overall biological reaction of acetate can be written as follows:



Another popular substrate for MFC is glucose, the overall biochemical reaction is written as in Eq. (9.4).



A comparative study of fermentable (glucose, glycerol) and non-fermentable (acetate, lactate) substrates showed glycerol performed more efficiently than acetate since fermentable substrate could augment the biodiversity and growth of biocathodic organism (Vicari et al. 2018). The electric current generation was significantly higher in Glucose-Fe(III) than with only glucose, suggesting the role of Fe(III) in electric current production (Gurav et al. 2020). Du et al. (2020) observed that there was a good relation between dissolved organic matter (DOM) coupled with electricity generation and total and viable bacteria. Their results demonstrated that mixing waste-activated sludge into solid potato enhanced the presence of the tyrosine-like aromatic amino acids and aromatic protein-like substances that promoted hydrolysis and humification of the solid potato. Studies have shown power output, and current density could be maximized by addition of antibiotics. Wen et al. (2011) have demonstrated that glucose–penicillin can be degraded to produce electricity in a single chamber MFC with an air-cathode. The maximum power density for glucose + penicillin ( $101.2 \text{ W/m}^3$ ) was sixfold higher than the sum of glucose ( $14.7 \text{ W/m}^3$ ) and penicillin ( $2.1 \text{ W/m}^3$ ) as the sole fuel. The maximum current density of penicillin ( $10.73 \text{ A/m}^2$ ) was 3.5-fold compared with that without penicillin ( $3.03 \text{ A/m}^2$ ). In the presence of the anode biocatalyst *Rhodococcus*

*pyridinivorans*, a remarkable increase in power production (1.64-fold) and current density (1.28-fold) was observed by applying livestock antibiotic salinomycin to sewage waste. Salinomycin, a cationic binding agent was able to transfer the cation to the cell membrane through protein transport, thus improving the power production (Cheng et al. 2020). Although lignocellulosic compounds derived from residues of agriculture are favorable for low-cost electricity generation, microorganisms in MFC cannot directly digest lignocellulosic biomass for energy production. It must be degraded into monosaccharides or other reduced matters (Yadav et al. 2020a; Yaqoob et al. 2021).

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## 9.6 Future Outlook and Conclusion

In reality, the success of an experiment is in scaling up from the lab to the field level. Strategy should be adapted to enhance the overall efficiency of oxygen reduction and increase in microbial fuel cell output. Deeper understanding of genetically engineered organisms and hybrid systems using recombinant technology can be used for strain improvement that can efficiently transfer electrons to anode. Using nanoparticles can increase the electron transfer mechanisms (Kumar et al. 2018). MFC technology in combination with other application should be focused such as bioremediation, proton generation, and biosensors for toxicity detection. Lately, microbial fuel cell(MFC)-based biosensors have been extensively developed as a novel alternative for water pollutant detection such as ammonia, styrene, nickel, and copper. The novel gene circuit engineered in *E. coli* Rosetta (sentinel Rosetta) was constructed by expressing *ribB* (riboflavin synthesis gene) and *OprF* (porin synthesis gene) with the promoters  $P_{cusC}$  and  $P_{T7}$ , enabled sensing  $Cu^{2+}$  and generating electricity (Zhou et al. 2021). Ammonium-based MFC biosensors have proven to indicate the presence of excess ammonium in waste water. Excess ammonium inhibits the activity of electrogenic bacteria in the anode chamber and subsequently affecting electricity production (Do et al. 2021). MFCs are successfully used to achieve efficient treatment of styrene-contaminated wastewater by using activated sludge as an inoculum with maximum power density of  $13.6 \text{ mW m}^{-2}$  and styrene removal was 100% (Oveisi et al. 2021).

Before commercialization of the technology, the reactor designs, operating conditions, data collection, interpretation, and kinetic models should be thoroughly investigated. Commercialization of the technology depends on cost-effectiveness, eco-friendliness, and safety. The surface area of the electrodes should be increased so that power generated within cells can be used to run other parts of a fuel cell (Rahimnejad et al. 2015). Long-term operation of the MFC must be carried out instead of short periods of time; this could be achieved by optimizing various parameters from laboratory scale to outdoor scale and can be made possible for power generation in outdoor scale (Pandit and Das 2015). One of the best examples of commercialization of MFC is in wastewater treatment in association with electricity production, reducing the biological oxygen demand (BOD) and chemical oxygen demand (COD) of effluents.

Although the MFC technology is convoluted, it is still gaining popularity as a promising future technology that can be used without polluting the environment for the simultaneous generation of energy and reduction of organic waste. The constant hunt for novel electrode materials for enhancing the power generation of MFCs has opened up new directions for fabricating novel electrodes. Biotechnology involving metabolic engineering can be applied to increase the rate of bacterial metabolism, which can lead to enhanced cell potential. The chapter focuses on physical and chemical parameters that influence better bioelectricity generation by careful monitoring of substrate, which can promote an electrochemically active microbial community to utilize waste. Careful reactor design, choice of compatible electrodes and membranes can have a dramatic influence on power and current density.

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# Bioremediation: Remedy for Emerging Environmental Pollutants

# 10

Arti Sharma and Sandeep Shukla

## Abstract

Bioremediation is one of the approaches to recycle wastes into another form that can be utilized by other microbes. At present, the environment is suffering from numerous environmental pollution problems. Microbes are the key players to overcome these challenges. Microorganisms persist everywhere on the planet because their metabolic activity is astonishing; then come into presence in all over range of ecological conditions. The microorganism's nutritional capability is completely varied and that's why it is used as bioremediation of environmental pollutants. Bioremediation is involved in eradication, degradation, immobilization, and decontamination of different chemical wastes and physically harmful materials from the environment via the all-inclusive and achievement of microorganisms. The principle is altering pollutants such as oil, heavy metal, hydrocarbons, pesticides, dyes, and so on. It is done by enzymatic way via breaking down, so it has great involvement to solve numerous environmental difficulties. There are two kinds of factors these are biotic and abiotic circumstances are determined rate of degradation. Presently, dissimilar methods and strategies are applied in the area in different part of the biosphere. For example, biostimulation, bioventing, bioaugmentation, biopiles, and bioattenuation are the common ones. All bioremediation methods have their own merits and demerits because they have their own specific uses.

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A. Sharma (✉)

Government Degree College Prithvipur, Niwari, Madhya Pradesh, India

S. Shukla

Department of Environmental Science, Gurugram University, Gurugram, Haryana, India

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**Keywords**Microbes · Pollutants · Bioremediation · Heavy metal · Pesticides

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**10.1 Introduction**

Due to rapid industrialization and modern agricultural practices, the environment in the past few decades has been polluted severely, which has resulted in pollution of air, water, soil, and even the food consumed by animals and humans. This problem is worldwide and may cause a threat to both the environment and human health (Manisalidis et al. 2020). The use of pesticides and herbicides helps to increase agricultural productivity; however, using these chemicals causes a huge loss of biodiversity and contaminates agricultural land. Based on the half-life, pollutants remain in the environment for a long period. Some of them fade away by microbial transformation into non-toxic by-products, while some pollutants such as polychlorinated dibenzodioxofurans (PCDDF), dichlorodiphenyltrichloroethane (DDT), hexachlorocyclohexane (HCH), dioxins, and chlordane may persist in the surroundings over different periods and enter the food web biomagnified (Guo et al. 2019). This uncontrolled release of lethal pollutants into environments is a serious concern. Conventional approaches such as pyrolysis, land-filling, and recycling for the removal of contaminants are not that efficient to end the production of toxic compounds (Ferronato and Torretta 2019; Rai et al. 2020). Thus, the use of microorganisms is more suitable than conventional methods for the remediation of toxic environmental pollutants. Bioremediation is an approach that causes restoration of the natural ecosystem by eliminating pollutants from the environment and also preventing further pollution. Bioremediation is more cost-effective than alternative methods of remediation, i.e., chemical as well as physical. Using bioremediation, the pollutants' toxicity can be reduced by applying the microorganisms that transform highly toxic pollutants into lesser non-toxic forms. Some of the xenobiotic compounds, e.g., nitrated aromatic compounds, highly halogenated, and a few pesticides are still not reported to be degraded by microorganisms (Arora 2020). Nevertheless, the efficiency of microbes depends on various factors, i.e., chemical nature of pollutants, concentration, availability and physiological features of the environment. Therefore, the components that affect the degradation potential of microbes are either concerned with nutritional necessities or ecological factors.

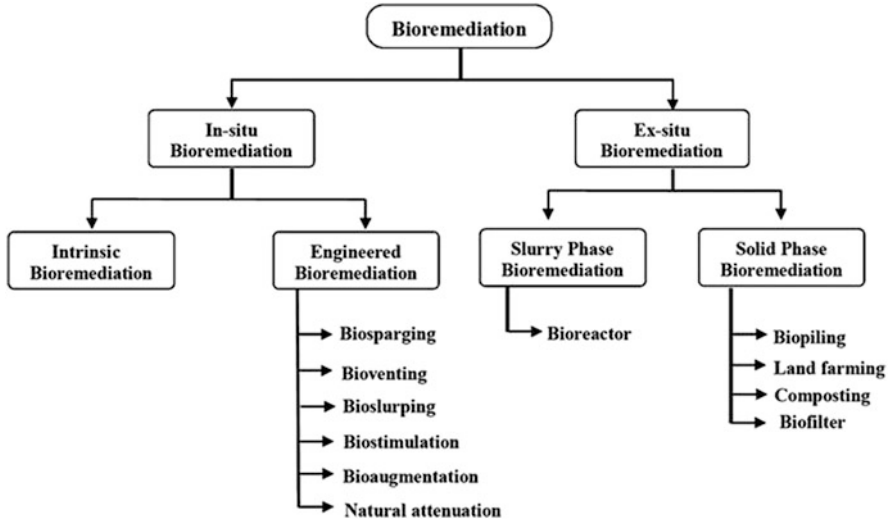
Further, based on the exclusion of toxic compounds and their transport methods, bioremediation is of the following two forms: *in situ* and *ex situ*. Moreover, recent methods incorporate the application of recombinant microorganisms for the effective degradation of pollutants. Under specific conditions for the remediation of different pollutants, recombinant microbes have been found to be successful as they have the genetic make-up to deal with pollutants. The elimination of numerous poisonous pollutants remains a problem for the environmental biotechnologists due to inefficient degradation by culturable microbes. The main hurdles for the use of recombinant microbes under field conditions are biological concerns and regulatory

restrictions (Ferronato and Torretta 2019). Despite the high efficiency of bioremediation, there are limited uses of recombinant microbes in the ecosystem due to the uncontrolled propagation and gene transfer. The present study's goals are to provide widespread details of combined approaches that have been accomplished for efficient evaluation of bioremediation processes (Srivastava 2021).

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## 10.2 Bioremediation

Bioremediation is an approach applied to remove ecological impurities from the ecosystem. It utilizes the living mechanisms inherent in microbes and plants to exclude hazardous pollutants and reconstruct the ecosystem to its original condition (Ancona et al. 2019). The basic concepts of bioremediation are to reduce the solubility of ecological impurities by redox reactions, changing pH, and adsorption of contaminants from the contaminated environment (Sharma 2020). Lot of work have been done on provoking pentachlorophenol biosorption by changing the pH levels in aqueous solutions. For the exclusion of pentachlorophenol from aqueous solution, the biosorption capabilities of *Aspergillus niger* (Gulzar et al. 2017) and *Mycobacterium chlorophenicum* (Das et al. 2015) were pH-dependent. It also evaluated the effect of pH on adsorption of pentachlorophenol by *M. chlorophenicum* and confirmed that pH values were a crucial parameter which affected pentachlorophenol adsorption. Several authors had performed many experiments, in which appropriate pH was used for best performance of microbes used in bioremediation. Bioremediation approaches are dependent on redox processes which focus on changing the microbiology and chemistry of water using selected reagents into contaminated water to enhance the degradation and eliminate numerous contaminants by in situ chemical oxidation reactions (Ojuederie and Babalola 2017). Redox reactions convert harmful contaminants into less toxic, mobile or inert stable compounds (Singh 2021). They play a crucial role in modifying toxic heavy metals such as As, Cr, Hg, and Se in soils and sediments into harmless forms (Ahemad 2019). A groundwater redox reaction is affected by the medium's physicochemical properties of the medium, but it can be improved using the addition of organic and inorganic alterations such as biochar and composts (Nejad et al. 2018). The use of compost in metal-mixed soils can cause modifications in the soil microbial population by altering pH, diminishing the solubility of heavy metals, and provoking microbial biomass and presented nutrients (Abedinzadeh et al. 2020). Biochar is a product of pyrolysis produced by manure crop residue as well as solid wastes. It can be utilized to enhance microbes for bioremediation to make the environment more suitable (Zahed et al. 2021). Several authors have explained that biochar is used as an actual agent in immobilization of organic pollutants and metals (Yaashikaa et al. 2020). Through biological pathway, Biochar has the capability to donate or accept electrons within their surroundings (Yaashikaa et al. 2020). Some scholars said that biochar may allow microbial electron shuttling processes (Pascual et al. 2020). The toxicity of heavy metals such as lead, arsenic, chromium, selenium, nickel, and copper rely on their oxidation



**Fig. 10.1** Bioremediation approaches for environmental clean-up (Sharma 2020)

states and is controlled by the redox reactions (He et al. 2019). Bioremediation depends on the prevailing environmental factors at the contaminated site and the nature of the organisms utilized as well as the degree of the pollutants in that environment (Ojuederie and Babalola 2017). Microbial bioremediation depends on the metabolic potential of the microbes to reduce ecological pollutants into modified innocuous forms via redox reactions (Ojuederie and Babalola 2017). Bioremediation can also be done through plants which remediate pollutants as well as contaminants from the environment. The bioremediation process carried out by plants is called phytoremediation. Heavy metals can be eliminated from the contaminated sites by plants (Nedjimi 2021). Bioremediation may be of two types, either *in situ* or *ex situ*. *In situ* bioremediation is the application of living treatment to clean up dangerous compounds present in the ecosystem and also to motivate microbes' capability to degrade contaminants or develop indigenous microbes to degrade contaminants present in environments using recombinant DNA technology (Goswami et al. 2018). Utilization of microorganisms for *in situ* bioremediation is affected using the non-availability of appropriate nutrient levels as well as environmental setting at the polluted site (Maulin 2014). *Ex situ* bioremediation is digging out the pollutant from its original location and transporting them to another site for treatment based on the pollutant type and depth of contamination, as well as geology of the contaminated site (Kumar et al. 2021). Figure 10.1 show the types of bioremediation and which have been explained one by one in the section below.

### 10.2.1 In Situ Bioremediation

There are two types of in situ bioremediation:

Intrinsic bioremediation and  
Engineered bioremediation

#### 10.2.1.1 Intrinsic Bioremediation

A type of bioremediation in which inert capability of naturally found microbes to degrade pollutants or contaminants without taking any engineered step to provoke the process. It degrades organic pollutants employing in situ microorganisms via a natural process known as natural attenuation. Potential intrinsic bioremediation of trichloroethylene (TCE) is being used, chlorobenzene as a primary substrate under aerobic and anaerobic conditions. Degradation of trichloroethylene is being dependent on degradation of primary substrate chlorobenzene. Microbial enumeration is accomplished to recognize the occurrence of intrinsic bioremediation. The existence of daughter compounds is an indicator of effective remediation.

#### 10.2.1.2 Engineered Bioremediation

A type of bioremediation that enhance the growth and degradative activity of microbes by using recombinant DNA technology that transports electron acceptors and supply nutrients or other growth enhancing materials. It is divided into six types. These are as follows: biosparging, bioventing, bioslurping, biostimulation, bioaugmentation, and natural attenuation. These are individually explained in the following sections.

##### Biosparging

Biosparging is the type of in situ bioremediation in which native microbes are used to degrade the organic constituents in the saturated zone. In Biosparging, nutrients are inserted into saturated zone to increase the biological activity to provoke the activity of native microbes. Biosparging can be used to reduce the concentration of petroleum ingredients that is dissolved in groundwater. It is the procedure in which pressurized air is pumped into a contaminated area to stimulate in situ aerobic biological activity. It targets chemical substances such as mineral oils, toluene, ethylbenzene, xylene, and naphthalene (BTEXN) that can be biodegraded in aerobic conditions (Soni et al. 2020; Verma et al. 2018; Yadav et al. 2020). It is used to treat soluble and residual contaminants in the saturated zone.

##### Bioventing

Bioventing was one of the first technologies that was applied in large scale in the 1990s. It is now mainly used in commercial applications. It is the type of bioremediation in which oxygen and nutrients are supplied into unsaturated zone. Oxygen is delivered into unsaturated zone via air movement through injection of air to enhance oxygen concentrations. This technique consumes the mandatory amount of oxygen

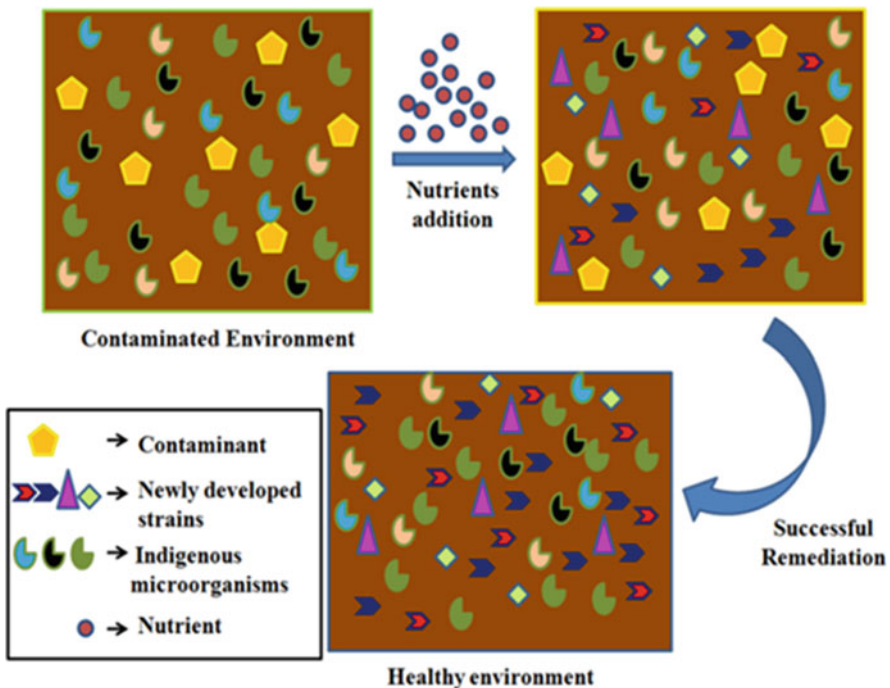
that is essential for degradation. It also reduces the volatilization and liberation of contaminants into the atmosphere.

### Bioslurping

Bioslurping combines bioventing and vacuum-enhanced free-product recovery. Bioventing boosts the aerobic bioremediation of hydrocarbon-impacted soils. Vacuum-enhanced free-product recovery eliminates light non-aqueous phase liquid from the capillary fringe and the water table. Bioslurping is less effective in low-permeability soils. The main limitation to air permeability is extreme soil moisture. Optimum soil moisture is very soil-specific and too much moisture can decrease air permeability of the soil apart from also decreasing its oxygen transfer capability. Microbial activity is inhibited when soil moisture is less.

### Biostimulation

Biostimulation refers to the addition of phosphorus, nitrogen, and oxygen into severely polluted sites to stimulate the native microbes to degrade the toxic contaminates. It modifies the environment to enhance the bioremediation. It is highly efficient, eco-friendly, and cost-effective for ecosystem. Figure 10.2 shows the outlines of biostimulation.



**Fig. 10.2** Depict the biostimulation bioremediation (Goswami et al. 2018)



### Bioaugmentation

Bioaugmentation is the technique of insertion of a precise combination of naturally occurring or genetically engineered microbial strains having higher capabilities in polluted sites for augmenting the natural degradation process. It is used for remediating soil as well as groundwater contaminated with tetrachloroethylene and trichloroethylene. Bacteria *Acinetobacter* and *Comamonas testosteroni* biodegrade 4-fluoroaniline and 3-chloroaniline in wastewater, respectively. Figure 10.3 shows the mechanism of bioaugmentation in which microorganisms convert contaminated environment into a contaminant-free environment.

### Natural Attenuation

Natural attenuation is the process that naturally transforms contaminants into less toxic forms. It attenuates pollution from soil and groundwater.

## 10.2.2 Ex Situ Bioremediation

It includes removal of waste materials and their collection from the polluted site or place to assist microbial degradation. There are two types of ex situ bioremediation:

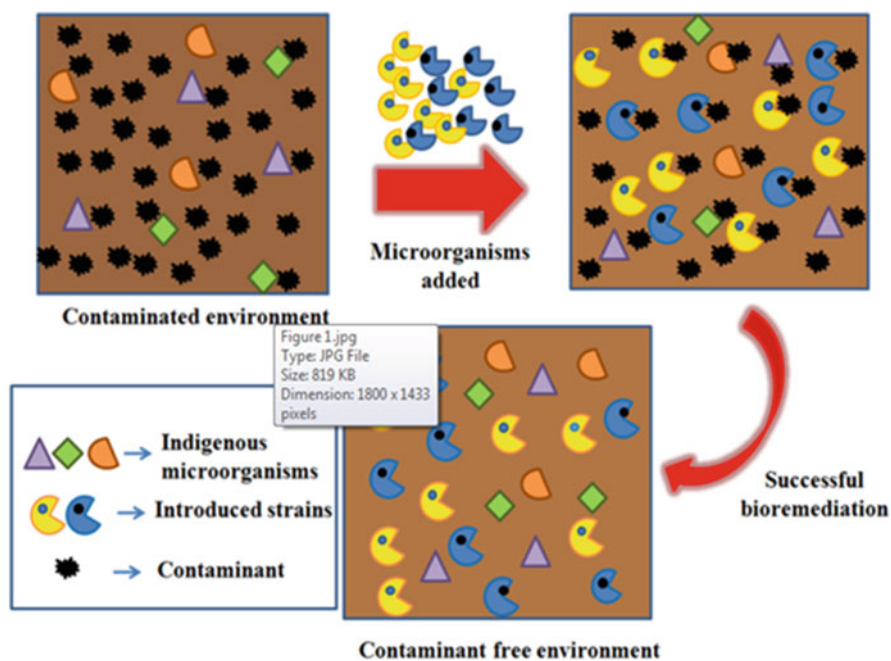


Fig. 10.3 Depict the bioaugmentation bioremediation (Goswami et al. 2018)

1. Slurry phase bioremediation
2. Solid-phase bioremediation

### **10.2.2.1 Slurry Phase Bioremediation**

It involves the treatment of a mixture of water and excavated soil in a bioreactor. The excavated soil is treated to separate stones and debris. An aqueous slurry is created by combining the contaminated soil with water and nutrients amount depends on altering the concentration of bio-degradation to occur. This is then placed into a bio-reactor. The slurry is mixed to retain solids suspended and microbes in contact with the soil impurities. Upon achievement of the process, the slurry is dewatered and the treated soil can be reinstated to its original position. Merely the polluted fines and collected wastewater require further treatment.

### **10.2.2.2 Solid Phase Bioremediation**

Solid phase ex situ bioremediation contains organic wastes (e.g., agriculture wastes, leaves, and manures, etc.) and problematic wastes (e.g., industrial and domestic wastes, etc.). It involves treatment of different solid wastes such as animal manures, municipal solid wastes, leaves, and agriculture wastes. Solid phase bioremediation is divided into four types such as biopiling, land farming, composting, and biofilter.

#### **Biopiling**

Biopiling is extensively used for remediating a wide range of petrochemical contaminates of soil. It involves the collecting of the soil into piles and provoking the biodegrading activity of microbial population by creating optimum growth conditions. It is used to treat non-halogenated volatile organic compounds and semi-volatile organic compounds. It is used recurrently to treat soils contaminated with petroleum hydrocarbons. Low weight petroleum products tend to vaporize from the pile owing to aeration, but the average and heavy petroleum hydrocarbons are degraded aerobically. Low levels of explosive residues, such as trinitrotoluene (TNT) and Royal Demolition Explosive (RDX) can also be treated, but less frequently. It is not used to treat inorganic contaminants and radionuclides.

#### **Land Farming**

Land farming is the treatment process that is accomplished in upper soil zone or in biotreatment cell. It has been proven most successful in treating petroleum hydrocarbons. Volatile hydrocarbons such as gasoline are treated very successfully. It has been used to treat surface soil contamination for hydrocarbons and pesticides. It enhances microbial degradation of hazardous compounds. As a rule of thumb, the higher the molecular weight, the slower the degradation rate. It means the more chlorinated or nitrated the compound, the harder it is to degrade.

#### **Compositing**

Compositing bioremediation remediates heavy metals, pesticides, and petroleum hydrocarbons from contaminated site. The benefits of compositing bioremediation are sequestering the precise contaminates, degrading the specific contaminates in

water and soil, and providing additional benefits associated with compost use such as provoking plant establishment and health, but it is not effective on some contaminates.

### **Biofilter**

Biofilter is the technology in which fuel hydrocarbon is passed through a soil bed where they sorb to the soil surfaces and are degraded by microbes in the soil. It is an important remediation method that can be useful in the removal of organic impurities from air and water. It also removes non-halogenated and is less effective for halogenated compounds. It is successfully used to control odors from compost piles. It is a highly effective air pollution control technology. It nearly changes all the contaminants to harmless products. Apart from that, it is a very low-cost technique.

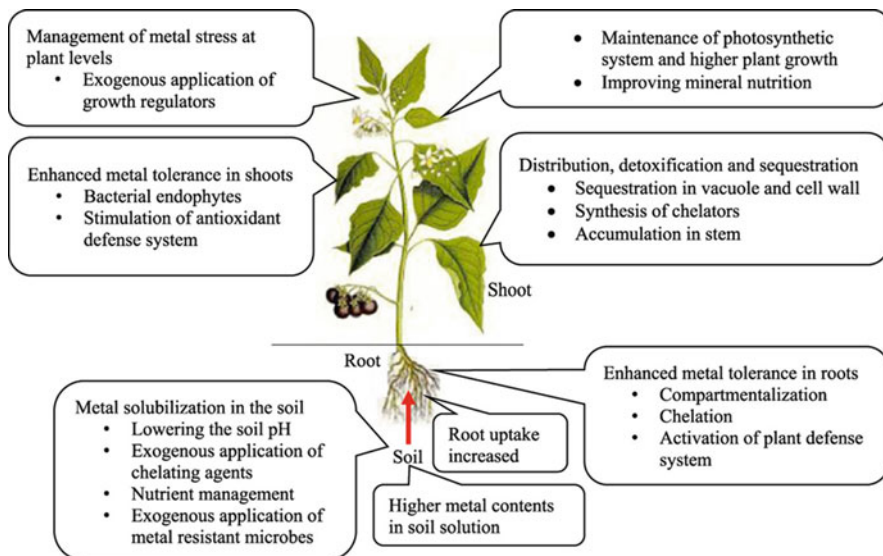
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## **10.3 Effects of Heavy Metals on the Environment**

Heavy metals with their non-biodegradability nature makes it stable to remove them from polluted biological tissues and it is a primary concern for worldwide health because of their fatal nature. Iron (Fe), manganese (Mn), cobalt (Co), copper (Cu), and molybdenum (Mo) heavy metals are required in minor quantities for the existence of living organisms, but at higher concentrations, they could be detrimental. The heavy metals Cd, Se, Ag, Hg, Cr, As, Zn, Au, and Ni are lethal heavy metals that pollute the environment and affects the soil quality and public health as well as crop production (Kaur et al. 2019). These metals are primary sources of life-threatening diseases in human being such as Alzheimer's disease, atherosclerosis, cancer, Parkinson's disease, etc (Uttara et al. 2009). Each metal toxicity is evaluated by absorbed dosage by the organisms and the duration of exposure. Heavy metal toxicity typically affects plants' physiological activities and are harshly hampered. For example, photosynthesis, respiration, electron transport chain, and cell division are affected by elevated levels of heavy metals as expected by laboratory experiments. Furthermore, high metal toxicity affects cytoplasmic enzymes in plants' cell and cell structures due to oxidative stress, which consequently affects metabolism and plant growth. Humans' exposure to Pb heavy metal could cause lethal health issues such as paralysis and lack of coordination. Severe exposure to Cd affects internal organs of the body such as the liver, kidney, and cardiac tissues. Arsenic is the most common cause of severe heavy metal poisoning in humans and causes respiratory organ failure such as lung cancer. Exposure of humans to Hg causes respiratory organ failure and speech impairment, hearing, and muscles dystrophy. It collects in the cells of microorganisms where it gets transformed to methyl mercury and becomes detrimental for aquatic lives. Consumption of these fish and other aquatic animals by humans can cause the transmission of toxic methyl mercury to humans. Due to the negative effects of these heavy metals, intensive efforts need to be made to efficiently eradicate them from the atmosphere and stabilize the ecosystem (Jaishankar et al. 2014).

### 10.3.1 Mechanism of Heavy Metal Remediation

Heavy metals remove important components in biological molecules and hamper the functions of the molecules. These alter enzyme activity, protein or membrane transporter structure or function, thus becoming toxic to plants (Thakur et al. 2016). The major treatment used for heavy metal deprivation include methods such as chemical precipitation, coagulation, electrodialysis, floatation, flocculation, ion exchange, evaporative recovery, nanofiltration, reverse osmosis, and ultrafiltration. Physicochemical methods such as extraction, soil washing stabilization, and immobilization are being also used for removal of heavy metals. These methods, even if effective, are usually expensive as a result of chemical reagent and high energy requirements, apart from production of secondary noxious end-products. To remove toxic metal contaminants from the atmosphere and stabilizing the ecosystem is to make use of native microbes to degrade such heavy metals. Engineered microorganisms can be used to treat polluted environments by altering toxic heavy metals into non-hazardous forms (Srivastava 2021). However, the bioremediation method will only be successful when microbes that have the capability to remediate and endure heavy toxicity are utilized. Microbes are crucial to remediate heavy-metal-contaminated surroundings as they have a variety of ways to endure metal toxicity. Microbes that can change the oxidation state of several heavy metals have been broadly studied. Heavy metals bioremediation will be fruitful if a group of bacterial strains is employed rather than using a single strain culture. The synergistic effect of a group of bacteria on the mixture of Cd, Pb, and Cu heavy metal bioremediation from contaminated soils using the following strains of *Viridibacillus arenosi*, *Sporosarcina soli*, *Enterobacter cloacae* and *E. cloacae* were studied (Kashyap et al. 2019; Li et al. 2019). Bacterial mixtures had larger resistance for the remediation of heavy metals than using a single strain. Heavy metals are the key environmental pollutants and the assembly of these metals in soils are dangerous for agricultural manufacture owing to the toxic effects on crop development and food quality. **Phytoremediation** is an important and low-cost tool which is used for the remediation of metal-contaminated soils. *Solanum nigrum* is the best example which is widely used for the remediation of heavy metal-contaminated soils owing to its capability for metal uptake and endurance. *S. nigrum* can tolerate huge amounts of heavy metals by enhancing the activities of antioxidant enzymes and metal deposition in non-active parts of the plant. A summary of heavy metal uptake and tolerance in *S. nigrum* is given in Fig. 10.4. Both endophytic and **soil microbes** can play a role in augmenting metal tolerance in *S. nigrum*. Additionally, optimization of soil management practices and exogenous application of amendments can also be used to enhance metal uptake and tolerance in this plant (Muhammad et al. 2017).



**Fig. 10.4** Mechanism of heavy metal remediation by *Solanum nigrum* (ur Rehman et al. 2017)

## 10.4 Potential Hazards of Textile Wastewater

Textile wastewater containing hazardous dyes has adverse impacts on the human lifecycle and water resources. The textile dyes substantially affect the quality of water bodies, impair photosynthesis, inhibit plant growth, and provide recalcitrance and bioaccumulation. It increases Biological Oxygen demand (BOD) and Chemical Oxygen demand (COD) and may boost mutagenicity and carcinogenicity (Al-Tohamy et al. 2022). The presence of dyes in water has hostile environmental influences due to their carcinogenic nature. Dyes inhibit the dissemination of sunlight into the water. It changes the color of water and, apart from that, affects the photosynthetic reaction that damages aquatic life. The presence of chlorine and metals in textile wastewater could be injurious for certain forms of marine life. These dyes and pigments can damage water quality by eutrophication and disturb the ecological conditions of the aquatic flora and fauna. Dyes cause severe human health problems, and they can also cause a series of long-term harmful effects if they reach human organs via the food chain (Khan and Malik 2014).

### 10.4.1 Treatment of Dyes

Physicochemical and biological are two major techniques for the remediation of dyes. The physicochemical approach used for treating the textile effluents. These are oxidation, flocculation, coagulation, precipitation, bleaching, membrane filtration,

ion-exchange, and adsorption. The physicochemical techniques that are employed for dye remediation also have demerits such as high cost, high-energy requirement as well as generation of secondary waste. Besides these conventional methods, bioremediations have recently received considerable attention as a relatively low-priced and reasonably good treatment choice for textile effluents.

#### **10.4.1.1 Physicochemical Methods**

Numerous physicochemical techniques have been used for the removal of dyes from wastewater. These contain adsorption, membrane separation, coagulation, flocculation, ion-exchange, photo degradation, and oxidation (Rajasulochana and Preethy 2016). However, these methods have economic and technical obstacles, such as high cost and generation of huge amounts of sludge and detrimental by-products as well as low viability on a commercial scale. Flocculation and coagulation approaches are effective for the decolorization of dye-containing wastewater. Coagulation approach employs ferrous sulfate and ferric chloride for the uptake of dyes from textile wastewater (Yaseen and Scholz 2019). Nevertheless, studies have also described the fruitful applications of other coagulants such as poly-aluminum chloride, magnesium chloride, and aluminum chloride (Gautam and Saini 2020) for the remediation of textile wastewater. However, coagulation has certain demerits such as high cost, low decolorization efficiency, and the generation of substantial amounts of sludge.

#### **10.4.1.2 Biological Methods**

Besides the physicochemical methods, biological methods are an alternate choice because they have low operating cost. They also convert harmful and toxic materials into harmless as well as non-toxic products. Numerous bioremediation techniques for the elimination of textile dyes are discussed in the following sections. Bioremediation is an approach in which either organic wastes are degraded naturally into harmless compounds or their concentration is minimized to a standard range (Kumar et al. 2020; Uday et al. 2016). Microbes used in the bioremediation approach consume the environmental contaminants as food and break them down. Nutrients supply and other constituents are vital for the degradation of harmful substances. Enzymes are responsible to enhance the metabolic reactions. Different enzymes are responsible to degrade numerous dyes. The environmental conditions play a crucial role in the bioremediation approach because it affects the microbial growth that is crucial for the bioremediation. For an effective bioremediation process, the environmental circumstances can be improved to promote microbial growth, thereby enhancing the degradation productivity of the microbes (Kanissery and Sims 2011).

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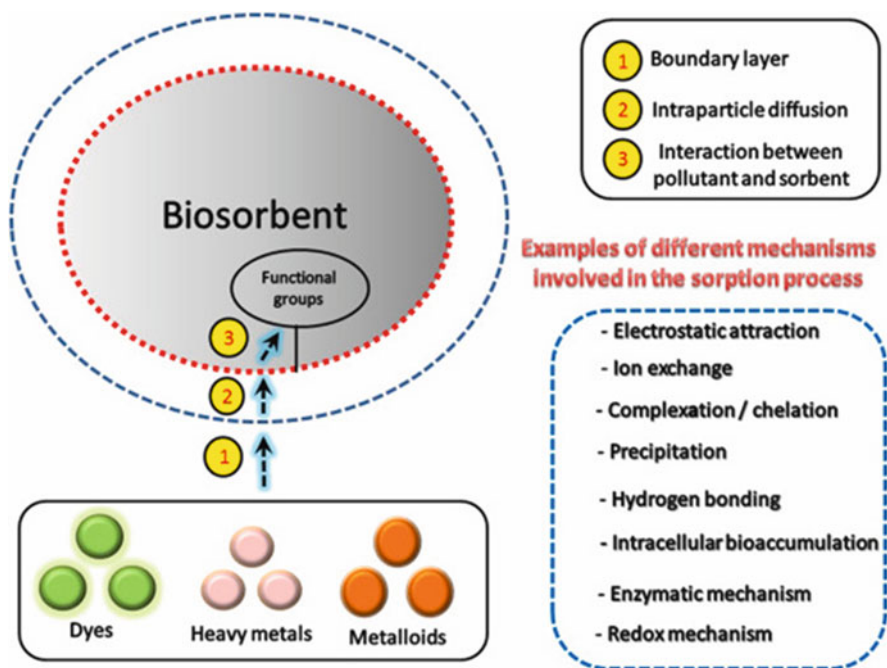
## **10.5 Degradation of Dyes by Bacterial Strains**

Large amounts of sludge are produced due to using these high-cost physicochemical methods, using which result in a secondary level of air and water pollution. Due to that, there is an urgent need for cheap and eco-friendly removal techniques for

polluting dyes. Biological processes is the potential alternative to conventional physiochemical method because they contain several microbes such as bacteria, fungi, yeast, and algae which are used to make the environment eco-friendly in nature. Bacteria can attain a higher degree of dye-degradation and process the complete mineralization of textile dyes under optimum conditions. Recently, the biological processing of textile effluent has been described as more cost-effective and eco-friendly than physiochemical techniques (Roy et al. 2020).

## 10.6 Mechanisms of Bacterial Dye Degradation

By using biosorption, desulfonation, deamination, and reduction of azo bond techniques, bacteria perform the decolorization process of dye. Electrons are produced during acetate, and sulfide oxidation results in azo bonds in the dye are fragmented. Azo reduction usually occurs by the degradation of aromatic amines (Ramalho et al. 2004). Biosorption is the technique to remove the dye or minimize the concentration of dye, heavy metals, and metalloids in a large amount of wastewater (Fig. 10.5).



**Fig. 10.5** Mechanism involved in the biosorption process (Elgarahy et al. 2021)

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## 10.7 Mechanisms of Fungal Dye Degradation

Enzymatic degradation as a dominant mechanism is used in fungal dye remediation. Enzymatic bioremediation is an ecological, economical, as well as innovative technique. This process explores the typical characteristics of microorganisms or genetically modified organisms of producing specific enzymes to metabolize the pollutant, transforming the toxic form into a nontoxic form, and sometimes into new products. The enzymes involved in bioremediation processes are laccases, dehalogenases, and hydrolases. Laccases are enzymes capable of catalyzing the oxidation of phenolic compounds, aromatic amines, and their compounds. Dehalogenases degrade a wide range of halogenated compounds by cleaving C–X bonds (X = halogen atom). Hydrolases break chemical bonds using water and convert larger molecules into smaller molecules, decreasing their toxicity. These enzymes facilitate the cleavage of C–C, C–N, S–N, S–P and C–P bonds. Other mechanisms are also involved; these are desulfonation, deamination, and hydroxylation as well as demethylation. Biosorption was the primary mechanism for the removal of Reactive Blue 19 (RB19), RB, AR57, and RBB by several fungal strains. The elimination of RB5, Acid Red 97 (AR97), Reactive Blue 49 (RB49), and Acid Violet 43 (AV43) by fungal strains using reduction of azo bond (Ihsanullah et al. 2020; Sabuda et al. 2020).

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## 10.8 Mechanisms of Algal Dye Degradation

The crucial mechanism for algal remediation of textile dyes is biosorption. The adsorption of reactive dyes onto dried *Chlorella vulgaris* was principally a physical adsorption method, and it is exothermic in nature (Aksu and Tezer 2005). The degradation of Rhodamine B (RB) dye into CO<sub>2</sub> and H<sub>2</sub>O by *Coelastrella* spp. (Baldev et al. 2013). The removal of CR textile dye by *Haematococcus* spp. involves azo dye reduction and adsorption mechanism (Mahalakshmi et al. 2015).

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## 10.9 Mechanisms of Dye Degradation by Yeast

Adsorption, asymmetric cleavage of the azo bond, and hydroxylation are the crucial mechanisms for the removal of dye by yeast. Azo-dye Acid Red B (ARB) dye is decolorized via yeast under aerobic conditions. The ARB dye was transformed into *ortho*-hydroxyl compounds upon further oxidation (Jamee and Siddique 2019).

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## 10.10 Bioremediation Applications

Bioremediation must be considered as appropriate methods that can be applied to all states of matter in the environment such as, solids, liquids, gases, and saturated and vadose zones. The main methods of bioremediation are natural bioremediation and



biostimulation. The biological community misused for bioremediation contain native soil microflora. Apart from that, higher plants can be manipulated to enhance toxicant removal called phytoremediation, especially for remediation of metal contaminates.

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### 10.11 The Advantage of Bioremediation

There are many advantages of bioremediation (Tyagi and Kumar 2021), and these are as follows:

1. Bioremediation is a natural process and takes a little time to effect adequate waste-treatment process for contaminated material such as soil.
2. Microbes able to degrade the contaminant, the biodegradative populations, become reduced. The treatment products are commonly harmless, including cell biomass, water, and carbon dioxide. It needs very less effort and can be commonly carried out on-site regularly without disturbing normal microbial activities.
3. This also eradicates the transporting of amounts of waste off-site and the possible threats to human health and the environment. It is functional as a cost-effective process as compared to other conventional methods that are used for clean-up of toxic hazardous waste regularly for the treatment of oil-contaminated sites.
4. It supports complete degradation of the pollutants; many of the toxic hazardous compounds can be transformed to less harmful products and the disposal of contaminated material.

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### 10.12 The Disadvantage of Bioremediation

It is restricted for biodegradable compounds since not all compounds are disposed by whole degradation process. There are new products of biodegradation that can be more toxic than the original compounds and persist in the atmosphere. Biological processes are ecofriendly and inexpensive. It includes the occurrence of metabolically active microbial populations, appropriate environmental growth circumstances, obtainability of nutrients and contaminants. It is demanding to encourage the process from preliminary study to largescale field operations. Pollutants might be existing in solids, liquids, and gases in all three states. It frequently takes larger than other treatment such as excavation and incineration. Study is required to develop and engineer bioremediation skills that are suitable for sites with complex mixtures of pollutants that are not uniformly dispersed in the atmosphere.

## 10.13 Conclusions

Biodegradation is ecofriendly and an attractive route to remediating, cleaning, and managing as well as improving method for resolving unhygienic atmosphere via microbial activity. The speed of undesirable waste substances degradation is determined in competition within microorganisms like bacterial, fungi, and algae's inadequate supply with nutrient, rough external abiotic circumstances, and low bioavailability. Bioremediation depends on several factors which hold, but are not restricted to, budget and concentration of pollutants. It may be used to treat a wider range of pollutants. In contrast, in situ techniques have no supplementary cost for excavation; but, on-site installation charge of equipment, committed with meritoriously and control the subsurface of polluted site can decrease some unproductive in situ bioremediation approaches. Geological features of contaminated sites, including soil and pollutant type as well as depth, human habitation, and performance of every bioremediation approach, should be incorporated in determining the most suitable and operative bioremediation technique for the successful treatment of polluted sites.

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# Rhizoremediation: A Plant–Microbe-Based Probiotic Science

# 11

Neha Sharma and Sandeep Sharma

## Abstract

Global health is at the tipping point with the emitters of a myriad of anthropogenic environmental pollutant chemicals by industries. From these sites, an array of xenobiotic compounds, i.e., polycyclic aromatic hydrocarbons (PAHs), polychlorinated biphenyls (PCBs), pesticides, heavy metals, etc. get released, entering into our food chain, thereby threatening our lives too. The establishment cost for the remediation of these recalcitrant compounds with the traditional techniques (landfilling, incineration) is quite high. So, an alternative, safe, economical, ecofriendly, biological-based method is required. Microbes assisted remediation, rhizoremediation, appears to be particularly effective for the degradation of specific xenobiotic compounds in the rhizosphere due to the higher microbial communities than the non rhizospheric or bulk soil. Root exudates (such as organic acids, carbohydrates, phenolic compounds, etc.) in the rhizospheric region act as inducers for the catabolic genes during rhizosphere colonization to degrade the various xenobiotic compounds. The key step involved in degradation mechanism is the activation or reduction of pollutant molecule by bacteria such as *Pseudomonas*, *Alcaligenes*, *Sphingomonas*, *Rhodococcus*, and *Mycobacterium*, creating reactive sites for the next reactions, converting substrates into acetyl-CoA, which is catabolized in Kreb's cycle. Fungi generally co-metabolize organic pollutants, but they do grow on some aliphatic and

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N. Sharma (✉)

Department of Microbiology, Punjab Agricultural University, College of Basic Science and Humanities, Ludhiana, Punjab, India

e-mail: [neha-mb@pau.edu](mailto:neha-mb@pau.edu)

S. Sharma

Department of Soil Science, Punjab Agricultural University, College of Agriculture, Ludhiana, Punjab, India

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aromatic compounds by extracellular oxidation or intracellular catabolism. Although the process involved in rhizoremediation occurs through natural process, it can be optimized with the suitable plant-microbial interaction using individual strain or consortium to increase the microbial population density. However, studies on potential microbial communities, their selection from the niche area, characterization with their degradation capacity, proliferation in the applied root system can be a novel and useful tool to improve the plant.

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**Keywords**

Rhizoremediation · Xenobiotics · Degradation · Rhizosphere

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## 11.1 Introduction

### 11.1.1 Concept and Definition

Amidst the rise of global demand for food supply, one of the major prospects of shrinking agricultural land with industrial development is the release of a myriad of undesirable xenobiotic compounds. The majority of the compounds are released by agrochemicals, refineries, pharmaceutical, and petrochemical industries. The existence and survival of these toxic substances not only lead to their accumulation in the environment but also enter our food chain too. Most of these compounds being recalcitrant by nature, persist for long periods and cause lethal impacts. So, the cleaning up of these contaminated sites has become a major health issue and has received global attention by the environmentalists. To circumvent the toxic substances, various physical, chemical strategies such as dig-and-dump approach (landfill), incineration have been developed. However, these methods did not achieve success due to their high establishment cost with respect to the greater proportion of contaminated sites. To overcome them, an opportunity prevails to switch to a natural, inexpensive, eco-accommodating, microbial bioprocess to utilize waste with the minimal efforts (Rai et al. 2020; Sharma et al. 2020a). So, “rhizoremediation” describes such a low-cost bioremediation of xenobiotics through plant-microbe-based probiotic science, in which rhizospheric flora catalyzed the degradation and mineralization of xenobiotics.

“Rhizoremediation” is defined as the process involving the degradation of specific contaminants in the rhizosphere with the catalytic activities of microorganisms, particularly recalcitrant compounds. The term is derived from two words, “rhizo” means the root (i.e., rhizospheric region around the root; 1 mm) and “remediation” refers to the process involved to degrade recalcitrant compounds. In simple words, the process involved the release of root exudates structurally similar with the contaminants that leads to the colonization of those strains which are able to metabolize the compounds. The present article represents the features, mechanism involved to understand the microbial dynamics for the facilitation of a safer technology.

### 11.1.2 History

The roots secrete an enormous range of compounds into the surrounding soil. The particular region around the plant root was coined as “rhizosphere” by German agronomist and plant physiologist Lorenz Hiltner in 1904 (>100 years ago). Later on in 1920, first phenomenon of root exudation was explained by Knudson with the indication regarding microbial abundance in the rhizospheric region. Newman, in 1985, reported that plant roots can release 10–40% of their total photosynthetically fixed carbon; however, the composition and amount of the released compounds vary with the type of plant species, climatic conditions, nutrient deficiency or toxicity, physicochemical and biological properties of the surrounding soil. The plant rhizosphere–microbe relationships create a desirable niche for the proliferation of microbial communities. The first investigation towards the degradation of toxic compounds in the rhizospheric region reported the action of microbial species with the main emphasis on herbicides and pesticides degradation (Hoagland 1994). Later on, various studies on the suitable plant species in combination with microbes were done for remediating from recalcitrant compounds (Qiu et al. 1994; Kuiper et al. 2001). Various researchers have reported the degradation of Polycyclic Aromatic Hydrocarbons and Biphenyls; however, the persistent microbial population involved has not been studied in detail till now. The establishment of a plant species for rhizoremediation directly depends on the below-ground root system involved as they primarily harbor the degradative bacteria and metabolites (1° and 2°) produced (Kuiper et al. 2004; Salt et al. 1998). Similarly, Siciliano et al. (2003) reported that the plant species having extensive branched root system increases the degradative microbes involved.

## 11.2 Role of Microorganisms for the Remediation of Pollutants

Beneficial microbes in the rhizosphere aid in nutrient acquisition by producing the metabolite as well as degrade a variety of xenobiotics and PAHs (Olanrewaju et al. 2017; Di Benedetto et al. 2017). Some of these mechanisms include bioremediation, biofertilization, and biocontrol. Colonizing microorganisms can be detected attached to the root as free organisms in the rhizosphere (e.g., attracted to the root environment by nutrients present in exudates), or as endophytes (Solanki et al. 2023). Some of the examples are mainly species of *Arthrobacter*, *Aspergillus*, *Bacillus*, *Geobacillus*, *Pseudomonas*, *Rastonia*, *Rhodococcus*, *Rhodopseudomonas*, *Xanthomonas* (Ali et al. 2015; Kashyap et al. 2019; Saraf et al. 2014).

The majority of the rhizospheric bacteria and fungi produce Volatile Organic Compounds (VOCs) as metabolites having PGPR properties and help in signal talk between the plants and their associated rhizospheric microbes (Ali et al. 2015). *Bacillus cepacia*, *B. subtilis*, *Pseudomonas fluorescens*, *P. trivialis*, *S. maltophilia*, and *S. plymuthica* are some of the microbial species involved in producing VOCs (Ali et al. 2015; Saraf et al. 2014).



**Table 11.1** Role of involved microbial enzymes for the biotransformation processes

S. no	Property	Purpose	Examples	
1.	Siderophore production	Availability of metals	<i>Azotobacter vinelandii</i> , <i>Bacillus megaterium</i>	Ferreira et al. (2019)
2.	Biosurfactant production	Solubilize hydrophobic pollutants	<i>Alcaligenes faecalis</i> , <i>P. extremaustralis</i>	Rani et al. (2020)
3.	Laccase, dioxygenase, peroxidase	Degradation of various aromatic compounds	<i>Trametes versicolor</i> , <i>Pseudomonas</i> sp., <i>P. chryso sporium</i>	Kumar and Chandra (2020)
4.	Nitrilase	Aliphatic, aromatic nitriles	<i>Aspergillus niger</i> , <i>Pseudomonas</i> sp.	Badoei-Dalfard et al. (2016)
5.	Nitroreductase	Explosives	<i>P. putida</i> , <i>Comamonas</i> sp.	Ojuederie and Babalola (2017)
6.	Cytochrome P450 monooxygenase	Aliphatic and aromatic hydrocarbons	<i>P. chryso sporium</i>	Hou and Majumder (2021)
7.	Dehalogenase	Halogenated aliphatic and aromatic hydrocarbons	<i>Xanthobacter autotrophicus</i> , <i>Sphingobium chlorophenolicum</i>	Ang et al. (2018)

For the biotransformation of the recalcitrant compounds, the degraders initiate the degradation by the action of intracellular enzymes, namely, dehalogenases, dehydrogenase, dioxygenases, oxygenases, phosphatases, nitroreductases, nitrilases, and lignolytic enzymes (Yadav et al. 2020), or possessing the siderophore or biosurfactant properties (Table 11.1). The mechanism involved for rhizoremediation routes through jasmonic acid and ethylene-based pathway, which are triggered by the immune system of the host plant (Berendsen et al. 2012; Nambara 2013). During the process, the rhizospheric microbes interact with the host leads to the activation of jasmonic acid, which resulted in Induced Systemic Resistance (ISR) development. The induced ISR affects the jasmonic acid–ethylene pathway by increasing the expression during pathogen localization (Zamioudis and Pieterse 2012).

Many microbial species have been reported as PAH degraders, but their activity has mainly been measured under controlled conditions like pure culture and batch experiments. Few reports have considered their activity in soil and in the rhizosphere (Joner et al. 2001; Nichols et al. 1997). The mechanistic interactions between plants and microbial degradation processes are poorly known. Bacteria and fungi are able to use PAH as a source of carbon via specific metabolic pathways that include ring fission. It is assumed that bacteria can transform or degrade 2–4 rings PAH (Kanaly and Harayama 2000) whereas fungi, especially ligninolytic species, can also degrade higher molecular weight compounds (Novotny et al. 1999; Schützendübel et al. 1999).

## 11.3 Essential Factors for Rhizoremediation

The control and optimization of a rhizoremediation process is a complex system of many factors, namely, prevalent microbial population in the niche, availability of contaminants, environmental factors (type of soil, temperature, pH, the presence of oxygen or other electron acceptors, and nutrients).

### 11.3.1 Prevalent Niche Microflora

The microorganisms have pushed their boundaries of life everywhere in each possible direction. They can be easily adapted in almost any environmental conditions, such as at subzero temperatures, in extreme heat, desert conditions, in water, with an excess of oxygen, and in anaerobic conditions, with the presence of hazardous compounds or on any waste stream (Verma et al. 2018). The main requirements for their broad spectrum are an energy and a carbon source that make them an ideal for the purpose of remediation (Merino et al. 2019). Based on the oxygen requirements, rhizospheric microorganisms can be aerobic or of anaerobic type. Aerobic bacteria, recognized for their degradative abilities, are *Alcaligenes*, *Mycobacterium*, *Pseudomonas*, *Rhodococcus*, and *Sphingomonas*, which have been reported for pesticides and hydrocarbons degradation, have oxygen-dependent metabolism (Bala et al. 2022). The initial enzyme in the pathway for aerobic degradation, methane monooxygenase, is active against a wide range of compounds, including the chlorinated aliphatics trichloroethylene and 1,2-dichloroethane. Many of these bacteria utilize the amendments as the sole source of carbon and energy leading to the increase in the microbial biomass with respect to the unamended ones (Sharma et al. 2015). Nowadays, there is an increasing interest in anaerobic bacteria used for rhizoremediation of polychlorinated biphenyls (PCBs) in river sediments, dechlorination of the solvent trichloroethylene (TCE), and chloroform (Tegene and Tenkegna 2020). In addition, ligninolytic fungi such as the white rot fungus *Phanaerochaete chrysosporium* have the ability to degrade an extremely diverse range of persistent or toxic environmental pollutants (Table 11.2). Common substrates used include straw, sawdust, or corn cobs. Moreover, a new class of microbes, methylotrophs, utilizing methane for carbon and energy are found to have a broad substrate range.

### 11.3.2 Availability of Contaminants

The concentration and biochemical quality of biomass present in the soil determine the decomposition rate, thereby affecting the predominance of specific microbial communities (Sharma et al. 2020b). It is generally observed that biodegradation rate of xenobiotic compounds is low even if the nature of compound is biodegradable. The possible reason for the slow degradation might be due to the bioavailability of compounds to microorganisms. Therefore, availability of the contaminants is

**Table 11.2** List of plant–microbe combinations used for the rhizoremediation of various pollutants

Plant	Pollutant	Microbes	References
<i>Ocimum basilicum</i>	Polychlorinated biphenyls	<i>Pseudomonas</i> , <i>rhizobium</i> , <i>bacillus</i>	Sanchez-Perez et al. (2020)
<i>Euphorbia mili</i> , <i>Syngonium podophyllum</i>	Benzene	<i>Pseudomonas</i> , <i>Enterobacter</i>	Sriprapat and Thiravetyan (2016)
<i>Aloe vera</i>	Formaldehyde	<i>Rhizosphere microorganisms</i>	Yang et al. (2020)
<i>Populus alba</i>	1,4-dioxane	<i>Actinomycetes</i>	Simmer et al. (2020)
<i>Arabidopsis thaliana</i>	Chloromethane	<i>Hypomicrobium</i> sp.	Nadalig et al. (2011)
<i>Zea mays</i>	Lindane	<i>Streptomyces</i> sp.	Simon Sola et al. (2019)
<i>Brassica napus</i>	Phenol and Cr	<i>Pantoea</i> sp.	Ontanon et al. (2014)

considered to be the most important for the degradation of recalcitrant components. Bioavailability is defined as the extent of a contaminant that actually interacts with the biological membranes of an organism. However, in case of hydrophobic nature of pollutants, biodegradation occurs only in their aqueous phase. Bouchez et al. (1995) studied that the phenanthrene biodegradation took only their dissolved state which was utilized by the microflora. The studies reported that the factor hindering the process is the mass transfer rate, which can be overcome with the dissolution of solid to liquid phase. So, the above-mentioned factors, alone or in combination, directly affect the process of rhizoremediation.

### 11.3.3 Environmental Factors

#### 11.3.3.1 Nutrients

Carbon is the most basic element of living forms and is needed in greater quantities than other elements. In addition to hydrogen, oxygen, and nitrogen, it constitutes about 95% of the weight. Phosphorous and sulfur contribute with 70% of the remainders (Bala et al. 2022). The nutritional requirement of carbon to nitrogen ratio is 10:1, and carbon to phosphorous is 30:1. Nutrients C:N:P = 120:10:1 molar ratio N and P for microbial growth. The specificity to degrade the xenobiotic compounds are associated with the available nutrient ratio which is necessary to induce the chemotactic response. As a result, the degrading microorganisms induce the pathway by catabolizing the compounds while using sole carbon and energy source.

#### 11.3.3.2 pH

pH is among the main factors that directly affect the rate of biodegradation of the pollutants present in the soil. As pH affects the activation of biochemical reactions involved, thereby directly affecting the extent of microbial colonization. Thus the specific enzymes required for the rhizoremediation are pH-dependent, thereby

making the microbial process dependent on optimum pH too. Singh et al. (2006) reported that degradation of pesticides was less in acidic soils as compared to the neutral and alkaline soils.

### 11.3.3.3 Type of Soil

The effectiveness of rhizoremediation is affected by the physicochemical properties of the soil, namely, amount and nature of clay, moisture, nutrients, organic matter, temperature, pH, redox conditions, which not only affects the colonization but also the transport of chemicals into soil. Moreover, a linear correlation was found with the moisture of the soil for the mineralization of relative pesticides (Schroll et al. 2006).

Soil organic matter, nutrient rich, supports the growth of biodegradation flora and controls the adsorption of contaminants, thereby affecting the degradation process. The formation of soil organic matter is a continuum of progressively decomposing processes. The other dominant factor is physical interparticle interaction, i.e., porosity of soil (Lou et al. 2022). In fine-grained soils, hydrocarbons are capable of inducing changes in particle texture, to considerably reduce the number of micropores and the overall surface, while the macropore features remain approximately the same. In the case of coarse-grained soils, contamination can create hydrocarbon-coated particles and fill both macropores and micropores (Rajabi and Sharifipour 2019).

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## 11.4 Mechanism: Plant–Microbe Interactions

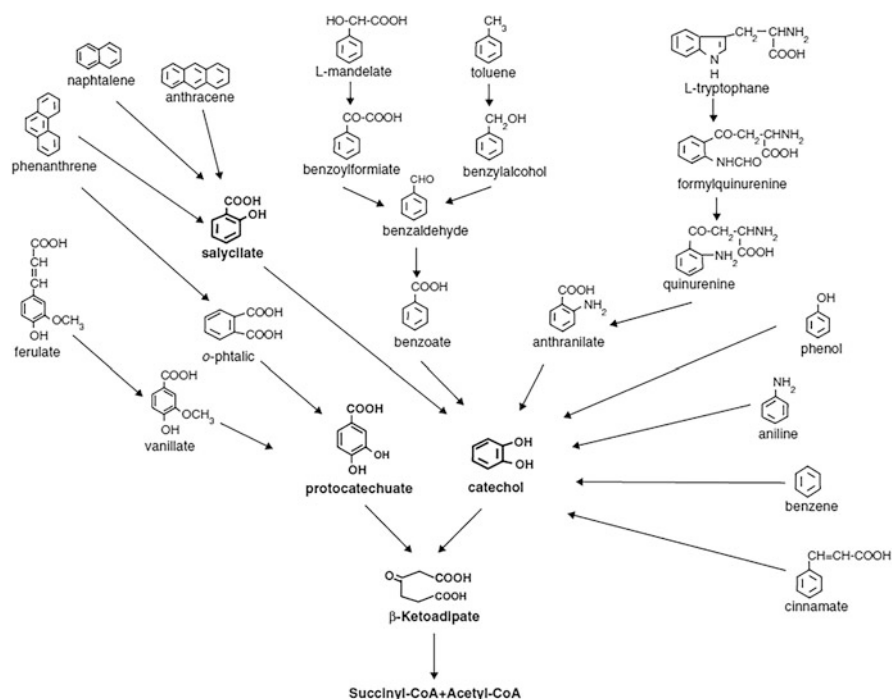
For a successful rhizoremediation strategy, the microorganisms involved must be able to proliferate in the presence of pollutant and have the specific operative catabolic pathways for the remediation. The mechanism for rhizoremediation involves:

1. Root exudation and colonization
2. Regulation of catabolic genes

Interacting with the pollutants: Rhizobiome in action.

### 11.4.1 Root Exudation and Colonization

In the rhizosphere, root secretions secrete attractant or repellent as signal to evoke the signaling pathway between roots and rhizospheric microbes (Lakshmanan et al. 2014). These signaling pathways regulate the interactions among the plants, microbes, and plant–microbe forms (Fig. 11.1) to induce responses (Moe 2013; Mommer et al. 2016). Not every root exudation is directly involved in the plant growth and nutrition. Some of them act as signaling molecules which mediate interactions in rhizobiome (Kumar et al. 2023). The exudates include sugars



**Fig. 11.1** Convergence of the bacterial degradation pathways of different aromatic compounds into a central metabolic pathway (Segura and Ramos 2013)

(arabinose, glucose, fructose, maltose, mannose, oligo-forms), amino acids (asparagine, aspartate, arginine, cysteine, glutamine), organic acids (acetic-, ascorbic-, benzoic-, ferulic, malic acids), phenolic compounds (coumarin), and high molecular weight compounds (enzymes, vitamins, flavonoids, fatty acids, auxin, alkaloids, gibberellin, nucleotides, steroids, tannins, terpenoids, polyacetylenes) (Gunina and Kuzyakov 2015; Hayat et al. 2017).

Plant roots not only provide nutrients to the microbes but also provide a large surface area for the colonization of microflora. During root exudation, the roots release root exudates structurally similar with the contaminants such as phenylpropanoid act as inducer of *Pseudomonas putida*, also *p*-cymene, limonene, and isopropene induce PCB degradation in *Arthrobacter* (López-Farfán et al. 2019). As a result, the biodegradative microorganisms using their attachments (such as surface proteins, capsular polysaccharides, or flagella) get attached to the plant roots by the process of chemotaxis. Diversity in root exudation leads to the generation of different microbial communities specific to each plant species. Using the In Vivo Expression Technology (IVET), transcriptomics and mutants defective studies in motility, mechanism involved during root colonization and recognition of catabolic

gene cascade which get activated during colonization are now being discovered (Bala et al. 2022; Xu et al. 2022).

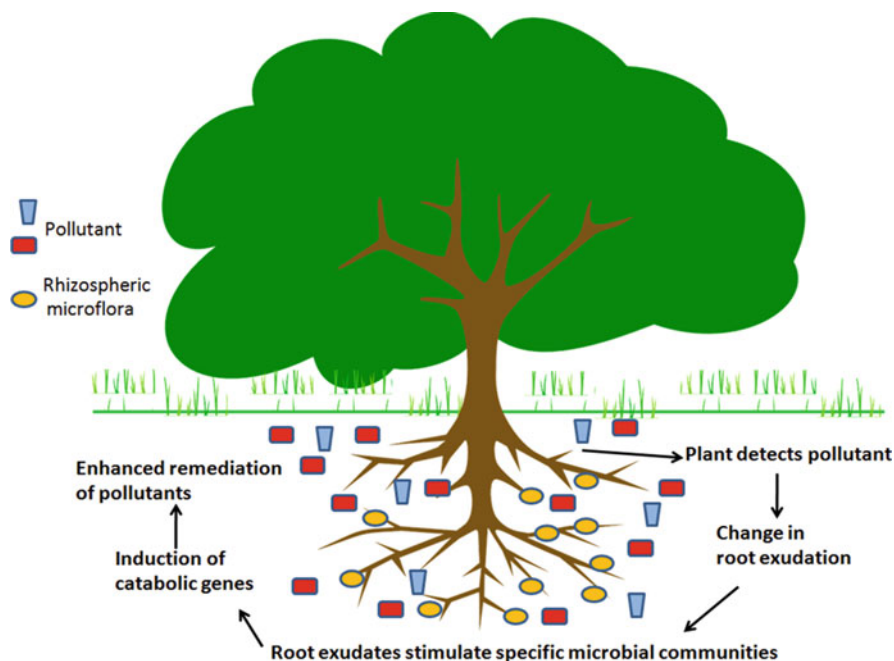
### 11.4.2 Regulation of Catabolic Gene Cascade

The selection of an appropriate plant species for pollutant degradation is considered as a main aspect for rhizoremediation; however, plant–microbe combination is also of major concern. Root exudates release a variety of organic acids, inorganic compounds, fatty acids, nucleotides, sugars, and secondary metabolites that lead to the colonization of the specific microflora. After colonization, expression of specific catabolic genes gets induced in the presence of root exudates (Rani et al. 2020). For the specific gene expression, about 200 homologues of promoter regions have been identified in different strains of *Pseudomonas*, such as 20 genes have been identified in *P. fluorescens*, 28 genes in *P. putida* KT2440. The functions of these specific genes help the microorganism in chemotaxis, motility, transport, secretions, stress mechanisms, energy metabolism, detoxification, and protein synthesis (Table 11.2). The specificity in the type of secretions helps directly or indirectly in the determination and the regulatory control of the specific species of microorganisms (Supreeth 2022).

Small quorum sensing (QS) signals are found to play a role in establishing the density of specific microbial population (Venturi and Keel 2016). In the rhizosphere, communication signaling contains a cascade complex of regulatory responses that reacts to a specific compound by activating the transcription of particular loci. Many of the inhabiting species of Ascomycetes secrete signaling molecules, mainly alcohols that actively participate in specific developmental processes in plants (Benocci et al. 2017). AHLs (QS signaling molecule) are also found to regulate the activation of plant genes, induction of systemic resistance with respect to stress in plants and effectors for plant growth (Venturi and Keel 2016). The firstly studied AHL-QS system contains Lux I synthase family that forms the AHL molecules on interaction with the regulation of LuxR family, thereby leading to increase in the gene expression and alters the community of the rhizobiome (Lareen et al. 2016). One of the significant signaling networks can be observed in legumes, as they possess nitrogen-fixing symbionts to establish a stable communication network for plant growth.

### 11.4.3 Interacting with the Pollutants: Rhizobiome in Action

The mechanisms involved in plant–microbe interactions are complex. This process involves various levels of communications between organisms, activation and inactivation of genes, induction and repression of responses to various signals, and various pathways elicited in responses. Recently, pattern recognition receptors (PRRs) have emerged to study the plant immune responses. These PRRs act as molecular signatures that are species specific of each class of microbe to interact with



**Fig. 11.2** Positive feedback loop mechanism involved for the rhizoremediation of pollutants

the plants. Various model systems related to legume–rhizobia have been studied to uncover associated molecular determinants in symbiosis (McCormick 2018; Wood and Stinchcombe 2017). With the release of flavonoids from roots of the legumes, host-specific transcriptional activation of nod factor (NF) (i.e., lipochitooligosaccharides) takes place. These nod factors account for rhizobia–host specificity (Behm et al. 2014).

With the activation of specific promoters, transcriptional induction of catabolic genes leads to the establishment of microflora to metabolize the pollutants (Bala et al. 2022). There is a proposed pathway to sufficiently reduce the toxicant present in the soil (Fig. 11.2). When plant detects the pollutant in the soil, it alters the rate of exudation with the accordance of concentration of toxicant. This change in the root exudation evokes an increase in the relative abundance of those microbial communities which are best able to metabolize. Wu et al. (2006) reported about the greater prevalence of alkane monooxygenase (catabolic gene) in the rhizospheric region than the bulk soil for the decontamination of hydrocarbons. The substituents of aliphatic- or aromatic-hydrocarbons of pollutants are metabolized by the plants due to their structural similarity with the plant metabolites, resulting in their complete degradation or mineralization. Sometimes pollutants cannot be directly assimilated by the microbes that oxidize them, but may instead be further transformed by other populations (Supreeth 2022). These relationships significantly enhance mineralization of recalcitrant pollutants and prevent the accumulation of

toxic intermediates. This mechanism operates as a positive feedback loop until the concentration of the toxicant gets significantly reduced.

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## 11.5 Advantages and Disadvantages of Rhizoremediation

Although adopting various physicochemical remediation options are cost-effective, but side-by-side also pose a threat to the humans and the environment. As these treatments are prohibitive in the larger areas having lesser levels of contamination (Cunningham and Ow 1996). Therefore, considering a safe, economic, biological treatment is considered to be safer (Cunningham et al. 1996; Doty 2008).

In recent years, the approach for using these biological treatments achieved different success rates, namely, landfarming, composting, and though they appear to be promising, sometimes provoke the mobilization of the contaminant. Despite the number of researches reporting the screening of microbes having the ability for remediation, however, most of the attempts against pollutants remained unsuccessful (Cerniglia 1993; Parales and Haddock 2004). The reasons behind the delay in success might be due to factors affecting, like soil (type, moisture, temperature), toxicity of the contaminant, the inability of allochthonous microorganisms to compete with the existing autochthonous microflora for pollutant removal that directly influence the process (Goldstein et al. 1985; Head 1998).

Autochthonous (indigenous) microorganisms present in polluted environments hold the key to solving most of the challenges associated with biodegradation and rhizoremediation of recalcitrant compounds (Verma and Jaiswal 2016), provided that environmental conditions are suitable for their growth and metabolism. Another major advantage of rhizoremediation technique is that the process does not require extensive preliminary assessment of polluted site prior to remediation; this makes the preliminary stage short, less laborious, and less expensive. Additionally, the usage of plants in the polluted site confers additional advantage of accumulating some metals which can be recovered after remediation (called phytomining). A study by Wu et al. (2015) reported the potential applications (food, feedstuff, biofortification of agricultural products) of Selenium-enriched material recovered from remediation sites. Therefore, for rhizoremediation selection of suitable plant–microbe combination is a better approach to treat the diverse range of pollutants. Although rhizoremediation is a promising option, it also has drawbacks as the process takes much time due to the slow growth of plants and is limited by climate change and soil characteristics. Moreover, root exudates hinder the process by increasing the dissolution rate of pollutants that can be introduced in the soil environment. Pollutants, beyond a level, prove toxic to plants and their associated microorganisms (van Dillewijn et al. 2008), as microorganisms can convert the contaminant into their more toxic form or can mobilize the contaminant from where it can be entered into the human food systems.



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## 11.6 Cost-Effectiveness

In spite of the billions in funding and development of newer technologies and programs aimed at restoring heavy metal-polluted soils, the severity of heavy metal problems is increasing alarmingly every year around the world. This is partly due to the lack of awareness, but largely due to economic constraints mostly in developing countries (Wu et al. 2015). However, the rhizoremediation is inexpensive when employed as it curbs the cost of transport, recycling, and monitoring. Further, since the rhizoremediation approach involves the use of cheap renewable resources like PGPR having multiple properties, this technology could be more profitable than any other remedial technology. The biocontrol activities like antagonism and competition for nutrients and niches (Lugtenberg and Kamilova 2009) add further strength to the economic friendliness of rhizoremediation approach by cutting off the costs for pesticides and thereby circumventing phytopathogens naturally. Thus, rhizoremediation approach is made environmentally as well as economically more pragmatic. Rhizoremediation approach is aesthetically pleasing and low cost, uses solar energy, requires minimal maintenance, presents no need for further recycling, and preserves the soil fertility and ecology. As a result, this strategy is gaining wider acceptance (Olanrewaju et al. 2017). However, how this technology could be useful in the rehabilitation of metal contaminated but non-agricultural soils with poor nutrients or nutrient-deficient soils is indeed a challenge before scientists.

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## 11.7 New Insights

The studies on connecting the regulation of catabolic genes involved in the rhizosphere with the selection of the suitable plant–microbe types will have a greater impact in this approach. The studies on the biodegradability of pollutants are still lacking (Supreeth 2022). More insight into the transportation and assimilation of recalcitrant compounds by the plants need to be explored. The fate of contaminants should be extensively studied during rhizoremediation to avoid undesirable effects during field testing. Exploration in molecular signaling, genes involved between plant types and microbes, and exploiting these for the elimination of contaminants are to be considered. These studies can provide insight to study the underlying mechanism involved during microbe–plant interactions for the activation of regulatory catabolic cascade involved in polluted soils.

The monitoring of capable gene by “-omics” technique will allow the selection of catabolic genes for rhizoremediation (Kiely et al. 2006). The improvement in the analysis of metagenomics will possibly reveal new degradative capacities (genes) that will be worth introducing into strains with other interesting traits (i.e., good root colonization abilities). The signals that plant and microbes exchange when they recognize each other will have to be interpreted and the molecular basis of the specific interactions between certain plant genotypes with specific bacteria will need to be dissected (Lou et al. 2022). Information that can be derived from these studies

may provide further insights on how to design a successful rhizoremediation strategy.

Finally, more studies about the impact of using recombinant microorganisms over indigenous microbial communities are needed to meet with safety requirements, especially with the increasing need for recombinant microbes to deal with highly toxic chemicals, such as dioxins and PCBs (Hou and Majumder 2021). Molecular techniques such as “omics” (genomics, metabolomics, proteomics, and transcriptomics) have contributed toward better understanding of microbial identification, functions, metabolic, and catabolic pathways, in this way overcoming the limitations associated with microbial culture-dependent methods. Nutrient limitation, low population or absence of microbes with degradative capabilities, and pollutant bioavailability are among the major pitfalls which may hinder the success.

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## 11.8 Conclusion

Due to greater advantages, rhizoremediation is gaining wider acceptance. Besides remediation and earning, it ensures food security for humans and safeguard them from a lot of ailments. However, large-scale field trials and their assessments are required to guarantee the practicability of rhizoremediation. Although some studies about the selection of suitable plant–microbe combinations have been done. However, further inoculation studies have been required related to the assessment of potential probiotic rhizobacteria with the ability of rhizoremediation in rhizospheric region to yield a useful novel system. This can be an interesting tool to further improve and develop remediation techniques into a widely accepted technique.

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**Part III**

**Biotechnological Approach**



# Microbial Fermentation System for the Production of Biopolymers and Bioenergy from Various Organic Wastes and By-Products

# 12

Jayprakash Yadav, Sambit Ray, Manish Soni,  
and Brijendra Kumar Kashyap

## Abstract

The microbial fermentation process or MFP is a technique used in several sectors to produce natural, novel, eco-friendly, and pragmatical products for human beings. The MFP technique has been extensively studied and applied in pharmaceutical, dairy, fruit juice, and agricultural sectors and industries. Consequently, by-products in the form of solid and liquid wastes are generated in various sectors and business establishments, making waste management difficult. Hence, for the management of the waste generated by these industries, the by-products were used as a substrate for producing biopolymers and bioenergy by the action of microbes. Moreover, microbes utilise these by-products generated by various industries in their metabolic pathway to produce biopolymers and bioenergy as end products during fermentation processes. The aerobic fermentation process has been mainly used for biopolymer production, and the anaerobic fermentation process is used for bioenergy, such as biogas and bio-hydrogen. Several microbes have been reported, such as *Bacillus* spp., *Nocardia* spp., *methylophilus*,

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J. Yadav (✉)

Department of Biotechnology, Meerut Institute of Engineering and Technology, Meerut, Uttar Pradesh, India

S. Ray

Department of Ceramic Engineering, National Institute of Technology, Rourkela, Odisha, India

M. Soni

School of Engineering and Technology, Department of Biotechnology, Jaipur National University, Jaipur, Rajasthan, India

B. K. Kashyap

Department of Biotechnology Engineering, Institute of Engineering and Technology, Bundelkhand University, Jhansi, Uttar Pradesh, India

e-mail: [brijendrakashyap@bujhansi.ac.in](mailto:brijendrakashyap@bujhansi.ac.in)

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*Alcaligenes* spp., *Rhizobium* spp., *Azotobactor* spp., *Pseudomonas* spp., and recombinant *Escherichia coli*, by researchers in their research work. This chapter summarises the conversion of the complex substrate (waste) to the transparent substrate (waste), microbial strains, and fermentation techniques to produce biopolymers and bioenergy. This information is beneficial for selecting a suitable substrate source for a particular product generation with a known fermentation process and/or modifying the existing fermentation process.

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**Keywords**

MFP · Biopolymer · Bioenergy · Sustainable waste · Microbes

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## 12.1 Introduction

For the last few decades, researchers have been looking for suitable technologies which can be helpful in reducing the excretion of pollutants produced from the conventional management technologies of organic waste disposal, to transform organic waste into eco-friendly products (bioenergy and biomaterials). In the upcoming future, recently developed strategies for waste management, in addition to the existence of a favourable environment, can wisely swap conventional products (fossil fuels) with organic waste or power crops as a substrate for the production of energy and materials (plastics) and also play an important role to reduce the emission of greenhouse gas (GHG) into the environment (Bauen et al. 2009).

Biodegradable materials have the potential to replace conventional materials and bioactive compounds synthesised from renewable sources. Renewable resources such as organic waste can be utilised to produce several valuable products, like bioethanol and the bioactive compound obtained by sugar metabolism (Mezule et al. 2015; Liguori et al. 2016). During the microbial fermentation process, some microorganisms can produce biopolymers as an extracellular substance known as exopolysaccharides (EPS). These EPS, after proper downstream and purification processes, can be used as an absorbent, lubricants, adhesives, and cosmetics in the chemical, packaging, food, and cosmetics industries (Pepe et al. 2013). Current technological traits have discovered the value of bioactive compounds as key to biopolymers (succinic acid) (Ventorino et al. 2016, 2017) and 2,3-butanediol (Saratale et al. 2016) resultant from lignocellulosic biomass. Moreover, many biopolymers such as polyhydroxyalkanoates (PHAs), polylactides, aliphatic polyesters, and polysaccharides (Kumar et al. 2020; Lee 2000) have also been successfully investigated as bioplastics since their physical and chemical qualities perform similarly to typical artificial plastics (Steinbüchel and Fuchtenbusch 1998). Over them, PHAs have drawn a lot of attention because of their ability to biodegrade in a variety of conditions within a year (Cavalheiro et al. 2009). Some bacterial species such as *Alcaligenes* spp., *Azotobactor* spp., *Pseudomonas* spp., *Bacillus* spp., methylotrophs, and recombinant *E. coli* have been reported for the PHA production using different sustainable waste as low-cost substrates (Kashyap et al. 2019). Although, organic waste and by-products have been used as a useful substrate

for PHA production to replace conventional plastics. The production cost of PHA has been influenced by the used substrate, fermentation processes, and downstream processes. For the cost-effective production of PHA, a significant determination has been dedicated to strain improvement, more efficient fermentation, and the PHA recovery process to commercialise PHAs (Salehizadeh and Van Loosdrecht 2004).

For the industrial application of PHAs production, future scenarios are mainly engrossed in promoting cost-effective substrate, upgraded microbes culturing strategies, and recovery process technology, which is required for reducing production costs (Huang et al. 2005). As a result, a variety of low-cost substrates were investigated for the production of biopolymers, including cellulosic and hemicellulosic materials, sugar, oils, starch-based materials, whey, glycerol, fatty acids, molasses, and sucrose, as well as biological matter obtained from wastes and wastewater, with promising results (Castilho et al. 2009).

Moreover, similar substrates are highlighted with the ability to synthesise biopolymers as a source of bioenergy (biomethane and biohydrogen) which is obtained by the anaerobic digestion process. Hence, such substrates can be simultaneously utilised to synthesise bioenergy and biopolymers and also get the most valorisation once they are used as biological waste.

The organic compound metabolises into methane, CO<sub>2</sub>, water, and ammonia by a series of biochemical reactions in the metabolic process of present bacterial consortia called anaerobic digestion (Verma et al. 2018). During the primary process, organic compounds' complex biomolecules are broken down and hydrolysed into biodegradable products and soluble matters using extracellular enzymes (Panico et al. 2014). In the following process, complex biomolecules are hydrolysed into unstable fatty acids (VFAs) using acidogenic microorganisms, also known as the acidogenic phase (Sans et al. 1995). Then, the products of acidogenic phase undergo acetogenic phase. In the acetogenic phase, the end products are acetic acid, hydrogen, and CO<sub>2</sub>. In the final phase (methanogenic phase), methane-producing archaea utilise the end products of acetogenic phase to produce methane (Chynoweth et al. 2001). Similar substrates responsible for methanogenic metabolism are commonly utilised as precursors of the production PHAs (Patel et al. 2011). Hence, this appraisal provides information about the current technology for driving PHAs and biogas, focusing on utilising organic substances and by-products as raw goods to significantly lower production costs. Furthermore, this appraisal explores the efficiency of all biological processes while developing an advanced exclusive integrated strategy that can simultaneously synthesise biopolymers and bioenergy.

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## 12.2 Biodegradable Polymers (PHAs Production and Classification)

Polyhydroxyalkanoates (PHAs) are a group of bio-based and biodegradable polymers which resemble conventional plastics (Morais 2013; Koutinas et al. 2007). Numerous bacteria and extremophilic archaea acquire PHAs in their cytoplasm as water-insoluble granules helping in microbial survival during starvation

**Table 12.1** Bacterial species used for the production of PHAs and their derivatives

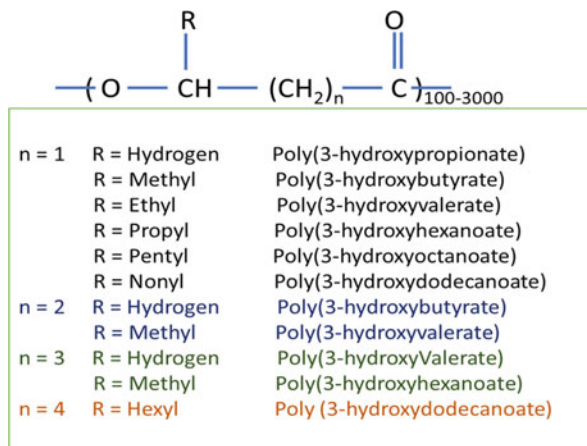
Bacteria name	Polymers	References
<i>Pseudomonas putida</i>	PHA	Cai et al. (2009)
<i>Azotobacter chroococcum</i> and <i>Azotobacter vinelandii</i>	PHA	Borrero-de Acuña et al. (2014)
<i>Bacillus subtilis</i>	PHB	Mohapatra et al. (2017)
<i>Bacillus cereus</i>	PHB	Panda et al. (2018), Hassan et al. (2019)
<i>Bacillus megaterium</i>	PHB	Sharma and Bajaj (2015), Yustinah et al. (2019), López et al. (2012)
Recombinant <i>E. coli</i>	PHA	Akdoğan and Çelik (2018), Pradhan et al. (2018), Wang et al. (2013)
<i>Burkholderia sacchari</i>	PHB	Alarfaj et al. (2015), Fei et al. (2016), Orita et al. (2014)
<i>Alcaligenes latus</i>	PHB	Soto et al. (2019), Khanafari et al. (2006)
<i>Cupriavidus nector</i>	PHB	García et al. (2014), Lee (2000)
<i>Methylobacterium extorquens</i>	PHB	Mahishi et al. (2003)
<i>Lactobacillus casei</i>	PHB	Inan et al. (2016), Park et al. (2019)
<i>Vibrio proteolyticus</i>	PHA	Iyapparaj et al. (2013)
<i>Ralstonia eutropha</i>	PHA	Tohyama et al. (2002), Kucera et al. (2018)
<i>Halomonas halophila</i>	PHB	Saratale et al. (2019), Kucera et al. (2018)
<i>Bacillus</i> spp.	P(HB-co-HV)	Hong et al. (2019)

and under adverse environmental conditions. PHAs and their derivatives have been accumulated by several microbes listed in Table 12.1 as energy-preserving components/granules (Reddy et al. 2003). The carbon source present in an excess amount in media has been utilised by a bacterial cell for cell growth, and other nutrient components such as nitrogen, oxygen, phosphorus, and sulfur have been used in low quantity for limited growth (Anderson and Dawes 1990). Bacterially synthesised PHAs are the unit of decomposable thermoplastic elastomers which are presently in use and are applied to be used in different sectors such as medical science, pharmaceutical industries, and the agricultural sector (Suriyamongkol et al. 2007).

PHAs can be categorised into two groups on the basis of C-atoms present on the side of a polymer as follows: a polymer having 3–5 C-atoms is known as short-chain length (SCL) PHAs, and having 6–14 C-atoms is known as medium-chain length (MCL) PHAs (Anderson and Dawes 1990). The physical properties of these polymers depend on the functional group present on their side chain. The SCL-PHAs have good properties such as brittle, crystalline, and stiff polymers, containing a high melting point and a low glass transition temperature. On the other hand, MCL-PHAs have less crystallinity and tensile strength and lower melting points.

PHAs have been represented by the common structural formula shown in Fig. 12.1, where  $n$  is equal to the number (1, 2, 3, and 4), and an alkyl group representing R. P(3HB) is the most commonly known monomer of the PHAs family.

**Fig. 12.1** Basic structural formula of PHAs. P(3HB) is a commonly used homopolymer for several studies of PHA, for which  $n = 1$  and R = methyl group



Copolymers of polyhydroxybutyrate (PHB) have been produced during fermentation using co-feeding strategies of the different substrates. Copolymers such as 3-hydroxybutyrate (3HB) and 4-hydroxybutyrate (4HB). The 3HV can be combined and form PHB molecule and poly(3-hydroxybutyrate-co-3-hydroxyvalerate) [P(3HB-3HV)], leading to an extra fragile compound than P(3HB) (Reddy et al. 2003).

Hence, an environmental load of chemically derived polymers can be reduced by utilising bio-based polymers (i.e. PHAs), which have biodegradable and biocompatible properties. Biocompatibility is the property of any compound that cannot produce toxins during its decomposition so that it can replace petrochemical-based plastics in environmental science and medical research. Conventional plastics take several years to degrade and also produce toxins during degradation. But, naturally produced polymers can be degraded in the presence of some indigenous microorganisms (bacteria and fungi) within a year. Specifically, some isolated indigenous aerobic and anaerobic PHA-degrading bacteria, for example, *Comamonas* sp. (Jendrossek et al. 1993), *Pseudomonas lemoignei* (Delafield et al. 1965) from the soil, *Alcaligenes faecalis* (Tanio et al. 1982) and *Pseudomonas fluorescens* from activated sludge (Mergaert et al. 1994), and *Pseudomonas stutzeri* from lake water (Mukai et al. 1994), and fungi (*Aspergillus fumigatus*) (Mergaert et al. 1994), have been isolated from several environments' sources. These microbes have the specific mechanism for secretion of extracellular PHA-depolymerase enzyme to degrade PHAs in water-soluble monomers and oligomers, which can be utilised as a carbon source (of methane beneath anaerobic environments) (Xu et al. 2010).

Hence, the biodegradability of PHAs and their derivatives have reduced the plastic waste accumulation in the environment that can be generated by a human being (Atlić et al. 2011). Moreover, biopolymers (PHAs) are better than petrochemically synthesised polymers, such as polyethylene and polypropylene in terms of sustainability and environmental safety (Atlić et al. 2011), but the recognition and more general usage of these eco-friendly PHAs are associated with the cost

of the end product (Carpine et al. 2020; Chanprateep 2010; Valentino et al. 2017). The recent PHA cost has been reported from €2.2 to 5/kg<sup>-1</sup> which is reliant on the monomer composition and specifically higher for the copolymers (Castilho et al. 2009; Carpine et al. 2020; Chanprateep 2010), which is lower than the reported initially past ten-decade range from €10 to 12/kg<sup>-1</sup> (Carpine et al. 2020). The production costs of currently used PHAs are very high and cannot compete with commercially generated polymers, with production costs of less than €1.0/kg<sup>-1</sup> (Carpine et al. 2020; Valentino et al. 2017). Even though the production costs of PHA products are expensive, these products possess valuable demand in countries like the UK, Italy, Japan, Brazil, the USA, and the People's Republic of China for their biocompatible and biodegradable properties (Tian et al. 2009; Lee et al. 1999).

### 12.2.1 PHA Production Using Suitable Substrate and Bacterial Strains

Several microorganisms can synthesise PHAs under optimised culture conditions and when grown with a suitable substrate, known as precursors. Microbes utilise these compounds in excess amounts in media as the sole carbon (energy) source for their growth (Rai et al. 2020). Furthermore, PHAs are also accumulated when cell growth is weakened or limited due to the lack of other nutrients such as nitrogen, oxygen, phosphorus, and sulfur (Wong and Lee 1998). Hence, PHAs might be produced by varying several growth parameters such as temperature, pH, aerobic, and anaerobic conditions.

Appropriate substrates have been reported for the production of PHAs as follows: agroindustry waste (e.g. sugarcane), CO<sub>2</sub> (Tsuge et al. 2002), renewable resources (e.g. starch) (Koutinas et al. 2007; Xu et al. 2010; Halami 2008), cellulose (Vandamme and Coenye 2004), and sucrose (Jiang et al. 2008; Page et al. 1992; Reddy et al. 2003), chemicals (e.g. propionic acid (Suriyamongkol et al. 2007), waste materials (e.g. molasses) (Page et al. 1992; Yilmaz et al. 2005), whey (Ahn et al. 2001; Koller et al. 2008; Nikel et al. 2006), and fossil resources, such as low-rank coal (Füchtenbusch and Steinbüchel 1999). Among them, waste materials and renewable resources are rationally considered appropriate and hopeful substrates and avoid the utilisation of fossil resources (environmental issues and high cost).

In the coming sections, the research idea is classified based on the various substrates utilised by the microbes for the mass production of PHAs, as represented in Table 12.2. The obtained result is denoted as follows: PHAs content (PHA, %) and PHAs amount (PHAS, g/L), estimated by the formula represented in Eqs. (12.1) and (12.2), respectively. The equations are:

$$\%PHAs = \frac{mPHAs}{mcells} \times 100 \quad (12.1)$$

**Table 12.2** Summary of PHA production using starch-based substrate by several microbes

Strain	Type of PHA	Operation mode	Incubation period (h)	PHA concentration (g/L)	PHA content (%)	References
<i>Bacillus</i> sp. CFR-67	PHA	Batch	72	5.9	59	Sharma and Bajaj (2015)
<i>B. thuringiensis</i> IAM 1207	PHA	Batch	48	2.6	72.8	Mendonça et al. (2014)
<i>Bacillus cereus</i> CFR06	PHA	Batch	72		48	Nascimento et al. (2016)
<i>Cupriavidus</i> sp. KKU38	PHA	Batch	96	2.43	61.6	Poomipuk et al. (2014)
<i>Bacillus tequilensis</i> MSU 112	PHA	Sequencing batch reactor (SBR)	24	3.346	79.2	Chaleomrum et al. (2014)
Mixed culture	PHA	SBR fed-batch	24		Up to 65	De Grazia et al. (2017)
<i>Bacillus cereus</i> 64-INS	PHA	Batch fermentor	21	2.78	60.53	Ali and Jamil (2014)
<i>Haloferax mediterranei</i>	PHA	Fed-batch	75	20	50.8	Bhattacharyya et al. (2012)
Engineered <i>Escherichia coli</i>	PHB	Batch	72	1.24	57.4	Bhatia et al. (2015)
<i>Corynebacterium glutamicum</i>	PHB	Batch	72	0.39	6.4	Song et al. (2013)

$$[\text{PHAs}] = \frac{\% \text{PHAs}}{100} \times \text{DCW} \quad (12.2)$$

Where mPHAs are denoted by the quantity of PHAs in mg, mcells represent the quantity of freeze-dried biomass of cells in mg, and DCW represents the dry cell weight in g/L.

### 12.2.2 Starch-Based Substrate

Starch is a widely available raw material as a renewable carbon source. Liquefaction and saccharification are the processes that convert starch into glucose by hydrolysis process. During PHA production, starch has to be converted into glucose as PHA-synthesising bacteria cannot degrade starch due to a lack of amylase enzyme. For starch degradation, commercially available enzymes are frequently used, but they contribute to a rise in the cost of manufacturing and processing of glucose. The reported PHA production using a different form of the starch-based substrate during the fermentation process is listed in Table 12.2. Hence, starch-based substrates are suitable materials for synthesising PHAs for P(3HB) production. Still, PHA production is firmly determined using bacterial species, which work under several biotechnological processes in the presence of carbon sources in traditional environments. One of the valuable results has been obtained from *C. nector* NCIMB 1159 culture using wheat and hydrolysed waste potatoes under nutrient ( $\text{N}_2$  and phosphorus)-limiting strategies during batch and fed-batch fermentation, respectively (Xu et al. 2010).

### 12.2.3 PHAs Production Using Molasses and Sucrose as a Carbon Source

Molasses is a popular effluent of the sugar manufacturing and processing industry, which is a cheaper carbon source than glucose. Molasses are considered potential feedstock because of their richness, low cost, and high sugar content. However, sucrose in molasses is required to transform into its monomers, fructose and glucose, by pretreatments for microorganism consumption during fermentation. Molasses was reported as a cost-effective substrate for PHAs production through the fermentation processes. The reported PHA production using molasses and sucrose as a carbon source during the fermentation process is listed in Table 12.3. The different carbon source was studied for the maximum PHB production in the batch fermentation process of *Bacillus megaterium*. The maximum biomass was reported with 3% molasses, while the best PHA and PHB yield was found to be 46.5% and 46.3% per mg dry cell weight with 2% molasses after a 48-h incubation period (Gouda et al. 2001).

**Table 12.3** Summary of reported research on PHA production using molasses as a substrate

Bacterial strain	Monomer of PHA	Operation mode	PHA production time (h)	PHA concentration (g/L)	PHA content (%)	References
<i>Cupriavidus necator</i>	PHB	Batch	60	0.78	27.30	Sen et al. (2019)
<i>Bacillus megaterium</i> ATCC 6748	PHB	Batch	45	1.23	35.00	Chaijammrus and Udpuay (2008)
Recombinant	PHA	Batch (molasses)	72	3.06	75.5	Saranya and Shenbagarathai (2011)
<i>Escherichia coli</i>	PHA	Batch (sucrose)	72	2.50	65.1	Gouda et al. (2001)
<i>Bacillus megaterium</i>	PHA	Batch	48	–	46.5	Page (1992)
	PHB			–	46.3	
<i>Azotobacter vinelandii</i> UWD	PHB	Batch (beet molasses)	24	7.4	65	Page (1992)
		Batch (cane molasses)		6.5	69	
<i>Cupriavidus necator</i>	PHB	Fed-batch	45–50	80–100	65–70	Page (1992)
<i>Azotobacter vinelandii</i> UWD	PHB	Fed-batch	35	23	66	Nonato et al. (2001)
<i>Bacillus megaterium</i> BA-019	PHB	Fed-batch	24	72.7	42	Kulpreecha et al. (2009)
<i>Pseudomonas aeruginosa</i>	PHA	Batch	54	5.60	76.5	Kulpreecha et al. (2009)
<i>Halomonas halophila</i>	PHA	Batch	72	64.06	2.57	Kucera et al. (2018)



### 12.2.4 Lignocellulosic Waste Material Used as a Substrate for PHAs

Lignocellulose is one of the abundant biopolymers that are found in waste biomass generated from plants. In general, the chemical composition of lignocellulose is 5–25% of lignin (complex polyphenolic structure), 40–80% of cellulose (b-D,1–4 glucan), and 10–40% of hemicellulose (D-arabinose, D-xylose, D-mannose, D-glucose, D-galactose, and sugar alcohols) (Obruca et al. 2015; Werpy and Petersen 2004; Yadav et al. 2020). Enzymes such as cellulase, hemicellulase, and ligninase are lignocellulose-degrading enzymes converting lignocellulose into sugar and biofuels (Yadav et al. 2020). Lignocellulosic feedstocks for industrial biorefinery crops are primarily wooden residues, agricultural waste, grasses, and stable municipal waste (Delmas 2008). These industrial feedstock can be used for the production of bioenergy and biopolymers, as cellulose and hemicellulose during fermentation process produces biofuels (bioethanol), biochemicals (lactic acid, succinic acid, and xylitol), and biopolymers such as polyhydroxyalkanoates (PHAs) (Madakka et al. 2020; Werpy and Petersen 2004). The reported PHA production using lignocellulosic materials as a carbon source during the fermentation process is listed in Table 12.4.

### 12.2.5 Whey-Based Culture Media Used as a Substrate for PHAs

Whey, an affordable renewable industrial waste (by-product), constitutes an excellent applicant for PHA synthesis (Choi and Lee 1997). Whey is the most important by-product of cheese production, representing 80–90% of the quantity of milk remodelled. Fifty per cent of the whey produced is utilised for making valuable products that include food ingredients and human and animal feed. However, the remaining is considered waste (pollutant) as a result of high biological oxygen demand (BOD) (Wong and Lee 1998). Whey has been used to make PHB in flask cultures and laboratory-scale fermenters using recombinant *E. coli* strains to carry the PHA biosynthesis genes of various species (Lee 2000; Choi and Lee 1997; Wong and Lee 1998; Ahn et al. 2000; Nikel et al. 2006). Using the wild-type strains of *Hydrogenophaga pseudoflava* DSM 1034 and *Sinorhizium meliloti* 41, the possibility for direct conversion of whey lactose to PHA was also examined by Povo and Casella (2003). Studies have shown that halophilic archaeon *Haloferax mediterranei* and the eubacteria *Pseudomonas hydrogenovora* and *H. pseudoflava* can utilise whey lactose as raw material for PHA production (Povo and Casella 2003; Koller et al. 2008). Moreover, it has been reported in decreasing the manufacturing cost for PHA production by developing higher bacterial strains and environment-friendly methods for fermentation and recovery (Solaiman et al. 2006; Lee 1996).

Conversely, whey might be an attractive potential raw material for PHA manufacturing. Still, the lack of, more importantly, PHA microorganisms to utilise the most of lactose has restricted its use as an attainable carbon supply (Pantazaki et al. 2009). *Thermus* species have been known to make the most of disaccharides corresponding to lactose. *Thermus thermophilus* HB8 and associated species mature

**Table 12.4** Summary of reported research on PHA production using lignocellulosic waste as a substrate

Bacterial strain	Monomer of PHA	Fermentation type	Fermentation time (h)	PHA concentration (g/L)	PHA content (%)	References
<i>C. necator</i>	PHB	Batch (sugarcane bagasse)	–	6.27	69.60	Dietrich et al. (2019)
<i>B. cepacian</i>	PHB	Fed-batch (sugar maple wood chips)	–	8.72	–	
<i>Burkholderia</i> sp. F24	PHB	Fed-batch (sugarcane bagasse)	–	12.25	50	
<i>Halomonas halophila</i>	PHA	Batch	72	61.95	2.17	Kucera et al. (2018)
<i>Burkholderia sacchari</i>	PHB	Fed-batch		72	105	Obruca et al. (2015)
<i>Bacillus megaterium</i>	PHB	Batch		51.6	12.5	
<i>Ralstonia eutropha</i>	PHB	Batch	36	70	10.10	Saratate et al. (2019)
<i>Burkholderia sacchari</i> IPT 101	PHB	Batch	25	62	2.73	Silva et al. (2004)
<i>Burkholderia cepacia</i> IPT 048	PHB	Batch	25	53	2.33	Silva et al. (2004)
<i>R. eutropha</i> ATCC 17699 ( <i>C. necator</i> )	PHB	Batch	48	75.5	11.4	Saratate and Oh (2015)

at temperatures from 50 to 85 °C, with an optimum at 70 °C (Wong and Lee 1998). *Thermus* sp. IB-21 has three thermostable lactose-hydrolases, two  $\beta$ -glycosidases (bglA and bglB), and one  $\beta$ -galactosidase (bgaA). The evaluation of *T. thermophilus* HB27 genome revealed that the diversity of lactose-hydrolases is frequent in *Thermus* sp. (Ahn et al. 2000). Because *T. thermophilus* ATCC 27634 (HB8) demonstrated a limp beta-galactosidase activity compared to different *Thermus* species, *Thermus* strains are separated into three groups based on  $\beta$ -galactosidase activity (Kim et al. 2000). The reported PHA production using lignocellulosic materials as a carbon source during the fermentation process is listed in Table 12.5.

### 12.3 Integrated Systems to Simultaneously Produce PHAs (Intracellular Products) and Biosurfactants (an Extracellular By-Product)

Bacterial strains which show maximum PHA production are used for industrial-scale production of PHAs to minimise the cost of biopolymers. These bacterial strains use the generated waste materials from the environment and convert them into important extracellular and intracellular by-products such as PHAs and exopolysaccharides (EPS). PHAs are an intracellular form of carbon and energy reserve, whereas EPS and biosurfactants are secreted as extracellular materials to prevent the cell from dehydration and predation. These materials have industrial attention, such as laundry powder and textile softener (Khosravi-Darani et al. 2013), and are additionally used in several other industries such as cosmetics, food, chemical, and packaging as a lubricant, adhesives, absorbents, and cosmetics (Khosravi-Darani et al. 2013; Kahar et al. 2004). Different bacterial genera such as *Bacillus*, *Enterobacter*, *Rhodococcus*, *Pseudomonas*, *Acinetobacter*, and *Arthrobacter* produce biosurfactants, organised as amphipathic molecules with polar and non-polar heads (Jiang et al. 2008). Biosurfactant formation is primarily influenced by carbon sources such as alkanes, lipids, sugars, and waste materials; hence, these compounds are available in a broad spectrum of chemicals. The primary function of biosurfactants is to minimise surface and interfacial tension, which forms microemulsions (Ibrahim and Steinbüchel 2009). Rhamnolipids are commonly studied biosurfactants. *P. aeruginosa* IFO3924 can synthesise PHAs and rhamnolipids simultaneously (Zhu et al. 2010). In this experiment, batch culture was performed at 30 °C in a 3-L fermentor equipped with an agitator, and decanoate (7 g/L) was used as a sole carbon source. PHA content of 23% of DCW and rhamnolipid amount 298 mg/L were obtained after 72 h of cultivation.

EPS (a mixture of high molecular polymers) is another type of extracellular polymeric substance which supplies carbon units when the substrate is limited. Bacterium *R. eutropha* was reported for the simultaneous production of EPS (extracellular products) and PHB (intracellular products). This study produced EPS as a growth-associated product, while PHB production was observed under nitrogen-limiting and cell-growth conditions. The polymers' production was reported using

**Table 12.5** Summary of reported research on PHA production using a whey-based culture media as a substrate

Bacterial strain	Monomer of PHA	Fermentation type	Fermentation time (h)	PHA concentration (g/L)	PHA content (%)	References
<i>Thermus thermophilus</i> HB8	PHA	Batch	24	–	35	Pantazaki et al. (2009)
Recombinant <i>E. coli</i> (R. <i>eutropha</i> genes) GCSC 6576	PHB	Fed-batch	49	69	85.18	Kucera et al. (2018)
Recombinant <i>E. coli</i> ( <i>A. latus</i> genes) CGSC 4401	PHB	Fed-batch	37.5	96.2	80.50	Suk Ahn et al. (2001)
Recombinant <i>E. coli</i> ( <i>A. latus</i> genes) CGSC 4401	PHB	Fed-batch	36.5	168	87	Suk Ahn et al. (2001)
<i>B. megaterium</i> Tt3	PHA	Shake flask	48	2.20	–	Israni et al. (2020)
<i>Hydrogenophaga pseudoflava</i>	PHBHV	Batch	48	4.2	–	Ahn et al. (2000)
Recombinant <i>E. coli</i> ( <i>C. necator</i> genes) GCSC 6576	PHB	Fed-batch with oxygen limitation	52	25	80	Kucera et al. (2018)
		Fed-batch without oxygen limitation	35	32	57	

**Table 12.6** Summary of reported research on PHA production coupled to metabolites used in industry

Bacterial strain	Monomer of PHA	PHA concentration (g/L)	PHA content (%)	Produced metabolites (g/L)	References
<i>Pseudomonas aeruginosa</i> IFO3924	PHA	0.5	23	Rhamnolipids 0.3	Pantazaki et al. (2009)
<i>Ralstonia eutropha</i> ATCC 17699	PHB	12.7	62	EPS 0.18	Wong and Lee (1998)
<i>Azotobacter beijerinckii</i> WDN-01	PHB	2.73	80.50	EPS 1.2	Ahn et al. (2000)
<i>Azotobacter chroococcum</i> 6B	PHB	0.74	28	EPS 2.1	Ahn et al. (2000)
<i>Pseudomonas mendocina</i> NK-01	PHA	0.316	25.3	Alginate oligosaccharides 0.57	Israni et al. (2020)

glucose and nitrogen at concentrations of 40 and 3 g/L, respectively, as listed in Table 12.6.

## 12.4 Bioenergy Manufacture Using Industrial and Agricultural Waste

### 12.4.1 Biogas Production (Anaerobic Digestion)

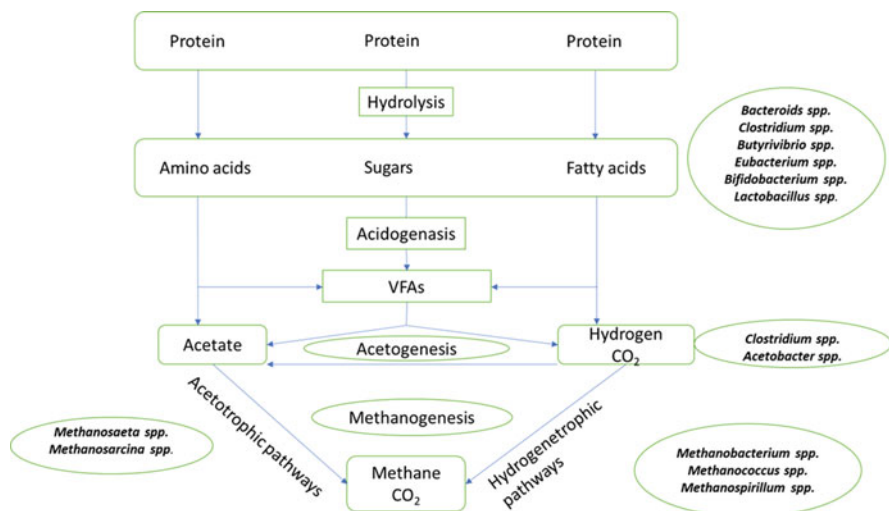
Anaerobic digestion is a fused biological process that minimises the organic content of effluent obtained from municipal wastewater treatment plants, stabilising the sludge (Appels et al. 2008). In the past few decades, anaerobic digestion has been applied for energy generation from solid wastes (biological waste and vitality crops) in fashion and also reducing the solid waste landfill problem (de Mes et al. 2003; Lettinga 2001).

Dung et al. analysed official data on food waste produced from 21 countries and assessed bioenergy production potential based on anaerobic digestion for biomethane, estimating a methane potential of up to 379.769 kWh/year (Dung et al. 2014).

Remediation processes performed using anaerobic digestion are flexible because they can have multiple combinations depending on the number of stages (one or two stages) as follows: (1) can function at different temperatures, mainly at 35 °C (mesophilic microbes) and at 55 °C (thermophilic microbes); (2) can be performed in batch, semi-batch, and continuous operations; (3) can be conducted in thoroughly agitated or plug flow reactors; and (4) can be carried out with less than 10% solid content in mass (wet system) or more than 20% solid content (steam system),

preceded by several revolutionary pretreatments to extend waste solubilisation (Mancuso et al. 2017).

Treating biological waste via anaerobic digestion have shown an edge in financial and environmental advantage (Lettinga 2001; Mancuso et al. 2017; Barton et al. 2008); and at the end of the anaerobic process, the waste matters are decomposed and are highly stable and less toxic to the environment. Hence the natural gas produced by anaerobic digestion has been successfully used as biogas which can be utilised for feeding into household gas networks (Fahimnia et al. 2015) as a substitute to petroleum gases, and the leftover matter can be utilised as fertiliser (Tambone et al. 2009; Rehl and Müller 2011). Due to the process, the environment's CO<sub>2</sub> balance does not alter and does not involve in global warming (Abbasi et al. 2012). Anaerobic digestion efficiency and results are dependent on the processing environment (Mata-Alvarez et al. 2000; Atasoy et al. 2018; Ariunbaatar et al. 2015), such as pH, nutrients component, temperature, availability of inhibitors (Ariunbaatar et al. 2015), utilisation of substrate and particle size, presence of micronutrient and the microbial strain used as inoculum. Anaerobic digestion is performed in the presence of microbial combinations (both bacteria and Archaea). Each trophic assembly of microbial commodity contains several microorganisms that play an essential role in the metabolic reactions (Kundu et al. 2017). A massive syntrophic association happens between different microbial consortia since biochemical reactions have occurred in series (Fig. 12.2). During anaerobic digestion, bacteria play an essential role in hydrolysis and acetogenesis.



**Fig. 12.2** Schematic representation of methane by a biological process with intermittent products such as VFAs, acetate, hydrogen, and CO<sub>2</sub>

Anaerobic bacterial species have been reported, such as *Streptococcaceae* and *Enterobacteriaceae*, which belong to genera of *Bifidobacterium*, *Butyrivibrio*, *Bacteroides*, *Clostridium*, *Eubacterium*, and *Lactobacillus* (Gerardi 2003). These bacteria are preferably subjected to the anaerobic digestion process. Thus, the bacterial species *Clostridia* fermented the protein hydrolysates to VFAs and also released CO<sub>2</sub> and hydrogen (H<sub>2</sub>) during anaerobic digestion.

In anaerobic digestion, Archaea plays a vital role in the methanogenic phase. Methanogenic Archaea (anaerobe) can convert fermentation products to methane (Gonzalez-Martinez et al. 2016). Among them, a few bacteria such as *Methanosaeta*, *Methanothrix*, and *Methanosarcina* genera produce methane using acetic acid as substrate, and these methanogens are known as acetoclastic or acetotrophic methanogens. Moreover, some other consortia of methanogens produce methane using water and CO<sub>2</sub> and methyl compounds such as *Methanobacterium*, *Methanococcus*, *Methanospirillum*, or *Methanomassiliicoccus* (Raposo et al. 2012).

For biogas production, the bacterial strain mentioned in Fig. 12.2 are capable of utilising all types of wastes matters that include animal manure, agriculture waste (organic), effluent from wastewater treatment plants, dairy wastes, waste from food processing industries, organic fraction of municipal solid waste (OFMSW), fruit and vegetable waste, and power crops are suitable substrate that can be used in an anaerobic digesters (Raposo et al. 2012).

Organic waste found from agriculture waste, meals waste, and OFMSW is mainly made up of metabolised carbohydrates. Feeding these wastes is not the proper way: VFAs obtained by the acidification process of anaerobic digestion tend to synthesise, triggering an instant drop of pH value, which prevents the action of methanogenic Archaea (Siegert and Banks 2005) and primes to a deficit of the process.

Protein-rich wastes are commonly obtained from the meat and fish processing industries, slaughterhouses, and animal farmhouses (slurry and manure) and are considered to have a low C/N ratio that can inhibit microbial growth and proliferation activities (Callaghan et al. 2002; Cuetos et al. 2008; Edström et al. 2003; Chen et al. 2008b). Besides, proteins inside the anaerobic digestion process are converted to ammonia as an end product, which is relatively noxious to microbes (Nielsen and Angelidaki 2008) and must be measured when searching for economical processes for ammonia removal (Limoli et al. 2016).

Estimating the amount of methane produced from a specific substrate can be commonly obtained through a specific biomethane potential test (BMP). This test generates the experimental value for the specific biomethane production that can be correlated with the anaerobic biodegradation potential of the system. However, the BMP results can fluctuate for the same substrate, as the anaerobic degradation can be affected by parameters such as temperature, mixing intensities of the matters, pH of the medium, substrate/inoculum (S/I) ratios, substrate particle size and distribution, liquid/volume ratios, nutrient content of the medium, inoculum, and pretreatment process (such as mechanical, thermal, and chemical treatments) and co-mixing of the substrates (Esposito 2012). Table 12.7 represents the methane yields from different substrates (Raposo et al. 2012).

**Table 12.7** Methane yields obtained from the reported solid waste. Adapted from Raposo et al. (2012)

Substrates	Yields (mL CH <sub>4</sub> g <sup>-1</sup> VS <sub>added</sub> )	References
Glucose	335	Dussadee et al. (2017)
Food wastes	245–510	Pagliano et al. (2017)
Fruit and vegetable wastes	470	Scaglione et al. (2008)
Apple fresh wastes	317	Buffiere et al. (2006)
Banana peelings	289	Buffiere et al. (2006)
Cooksfoot	325	Mähner et al. (2005)
Cellulose	354–375	Owens and Chynoweth (1993)
Cabbage leaves 2 mm size	309	Pagliano et al. (2017)
Carrot peelings	388	Buffiere et al. (2006)
Kitchen waste	432	Neves et al. (2006)
Leather fleshing	490	Shanmugam and Horan (2009)
Cauliflower leaves	341–352	Zubr (1986)
OFMSW	353	El-Mashad and Zhang (2010)
Lettuce residues	294	Buffiere et al. (2006)
Orange peelings	279	Buffiere et al. (2006)
Maize residues	317	Dinuccio et al. (2010)
Mandarin peels 2 mm size	486	Gunaseelan (2004)
Pineapple peel	400	Kapdan and Kargi (2006)
Rape oilseed	800–900	Hansen et al. (2004)
Paper and cardboard	109–128	Pommier et al. (2010)
Potato waste	320	Parawira et al. (2004)
Rice straw	347–367	Sharma et al. (1988)
Algal biomass	640	Zhen et al. (2016)
Sugar beet	340	Lehtomäki et al. (2008)
Starch	348	Lehtomäki et al. (2008)

### 12.4.2 Biohydrogen Production

Hydrogen is considered an excellent supply of vitality because it represents a clear flammable and can be simply convertible to electrical energy (Kapdan and Kargi 2006). Organic hydrogen manufacturing is said to be biogas manufacturing for two primary reasons: to i) similar to industrial processing method, and b) appropriate alike substrates for biogas production. Biohydrogen and biogas production have similar biological process that produce biohydrogen when the hydrogen gas using microorganisms such as *homoacetogens* and *methanogens* are inactivated. The inhibition is achieved via heat treatment of the inoculum to inactivate all the microorganisms, leaving behind only spore-forming fermenting bacteria (Angenent et al. 2004). *Clostridium* and *Thermoanaerobacterium* are the most common bacteria employed during dark fermentation for the production of biohydrogen. Furthermore, multiple investigations have shown that mixed cultures in batch or in bioreactors can produce biohydrogen (Shin et al. 2004; O-Thong et al. 2009; Ismail



et al. 2010; Prasertsan et al. 2009; Ghimire et al. 2015). The benefits of utilising combined cultures for biohydrogen production includes no sterilisation, high adaptive ability of microbial diversity, the ability to make use of a mixture of substrates, and the potential of acquiring a steady and continuous course of biohydrogen production (Ismail et al. 2010).

Furthermore, the identical biological substrates, similar to dense surplus (solid waste), might be subjected to delivering biogas-biohydrogen, hence shifting residues accurately into a source of bioenergies (Angenent et al. 2004). Several hydrogen-producing processes via microbial fermentation has been widely reported and studied in depth. However, hydrogen production by using photosynthetic bacteria, algae, and fermentative microorganisms in bioreactors is the most sustainable and optimum method for hydrogen production.

The natural process of biohydrogen production is known as autotrophic conversion. In this process, photosynthetic microorganisms, i.e. microalgae, convert photovoltaic energy to hydrogen (Ghimire et al. 2015). In autotrophic process, autotrophic microorganisms such as purple non-sulfur bacteria utilise the by-products of dark fermentation for the production of hydrogen gas via photofermentation and simultaneously break down the VFAs (Lindblad 2004; Das and Veziroğlu 2001; Miyake et al. 1999; Lo et al. 2008). But, this photofermentation process has a few limitations that include drop in hydrogen production with time, a lack of genetically modified photosynthetic microorganisms, and reduction in hydrogen conversion efficiency of photobioreactors (Chen et al. 2008a). Chen et al. (2008a) developed a novel photobioreactor (PBR) that boosts phototrophic hydrogen synthesis by utilising acetate as the sole carbon source during the fermentation process by using *Rhodospseudomonas palustris* WP3–5. The photobioreactor was brightened by combinatorial light sources and enhanced hydrogen production by up to 62.3% compared to the conventional photobioreactor.

In heterotrophic circumstances, the fermentation occurs in two different ways: photosynthetic bacteria perform photofermentation, while anaerobic microbes undertake gloomy fermentation (in dark condition), in which biohydrogen is produced by carbohydrates metabolism in anaerobic surroundings (Pradhan et al. 2015; Ghimire et al. 2015). Diverse rumen bacteria, likely *Clostridia*, methanogenic archaea, methylotrophs, or aerobic microbes (*Alcaligenes* spp., *Bacillus* spp.) and facultative anaerobic microbes (*E. coli*, *Enterobacter* spp., *Citrobacter* spp.) have been reported for the darkish fermentation process of biohydrogen production. *Clostridium butyricum* and *Clostridium articum*, in particular, produce butyric acid and propionate as essential products, both of which are important for hydrogen synthesis during anaerobic digestion. Photofermentation occurs beneath anaerobic environments concerning purple bacteria (non-sulfur photosynthetic bacteria) using brightness (light) as an energy source for hydrogen production (Eroglu and Melis 2011). The purple bacteria have the property to metabolise the organic acids and give biohydrogen as an end product with simultaneous production of PHB beneath anaerobic conditions.

As for methane production through anaerobic digestion, biohydrogen may be produced by different bacterial strains utilising several biological matters

(substrates). For example, Cappelletti et al. (2012) focused their research on hydrogen production from cheese whey and molasses with the goal of repurposing food craft wastes by using mesophilic, thermophilic, and hyperthermophilic bacteria as inoculum for hydrogen production. Amongst them, *Thermotoga* strains confirmed the furthestmost promising results specially, *T. neapolitana* was the most studied and essential strain. Experiments on *T. neapolitana* using various biological substrates, such as rice straw, beet pulp pellet, corn starch, and rice flour, established and supported the conclusion ((Nguyen et al. 2010a, b). Such substrates are particularly appropriate for producing H<sub>2</sub> due to their straight forward biodegradability and are also handy due to their current in numerous carbohydrate-rich waste waters and agricultural residues (Davila-Vazquez et al. 2008). Diverse substrates, primarily cast offs for biohydrogen manufacture, are protein- and fat-rich wastes. *C. butyricum* strain was subjected to the production of H<sub>2</sub> using the sucrose-based medium as a substrate during the fermentation process (Chen et al. 2005). In actuality, *C. butyricum* CGS5 can proficiently synthesise hydrogen (2.78 mol H<sub>2</sub> mol<sup>-1</sup> sucrose) on a substrate (iron-containing medium) (Lo et al. 2008). A similar bacterial strain (*C. butyricum* CGS5) was reported for hydrogen production, which is isolated from the environmental source (soil samples) with nine cellulolytic bacterial strains classified in the class of *Cellulomonas* sp. and *Cellulosimicrobium cellulans* by Lo et al. (2008). From the isolated strains, only *C. butyricum* CGS5 showed effective H<sub>2</sub> synthesis of 17.24 mmol H<sub>2</sub> g cellulose<sup>-1</sup> using rice husk hydrolysates as the sole substrate.

All biotechnological hydrogen synthesis processes have specific limits since a substantial part of the used substrate is transformed into numerous soluble metabolic goods as opposed to hydrogen. Hence, the key lateral product of darkish fermentation is VFAs and diverse components, like alcohols (Kumar et al. 2016). As a result, the effluent derived from the fatty acid-rich fermentation process can be used as a suitable substrate for biologically synthesising polyesters, such as polyhydroxyalkanoate, which has an immense market potential (Park et al. 2017; Morgan-Sagastume et al. 2010; Chen 2009).

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## 12.5 Integrated Process Systems for Bioenergy Synthesis from Industrial and Agricultural Sustainable Substances

### 12.5.1 Coupled Synthesis of PHAs and Bioenergy from Carbon-Based Wastes

Organic waste has been degraded into methane (CH<sub>4</sub>) and carbon dioxide (CO<sub>2</sub>) by several steps' course with the ability to synthesise hydrogen and bioplastics (from VFAs) as intermediates (Patel et al. 2011). During the anaerobic digestion process, the biomass is metabolised in the primary zone and hydrolysis–acidification occurs. During this process, the produced acids are metabolised by aerobic fermentation and produced biopolymers and biogas as secondary metabolites.

A PHA manufacturing arrangement, in its record inclusive outline, is constructed of four predominant bioprocess stages (Fig. 12.3) as follows: (1) feedstock production, (2) biomass selection, (3) PHA production, and (4) PHB extraction.

The basic arrangement could be performed using artificial substrate by eliminating stage 1 from the cycle, utilising pure culture by eliminating stage 2 from the cycle, or using both artificial substrate and pure culture to eliminate stages 1 and 2 from the cycle. The goals of every stage are as follows: (1) producing organic acids from complex organic substances (e.g. carbohydrates fructified wastes); (2) choosing the microbial strain from the mixed culture population that can produce maximum PHA production beneath defined dynamic feeding conditions (Serafim et al. 2008); (3) synthesising PHAs subjecting choose culture; and (4) extracting PHAs from microbes.

In the first stage, the dark fermentation could be resourcefully conducted. This method cultivates according to the anaerobic fermentation process in which the series of biochemical reactions have occurred and exclusion of the last stage that is clogged following several approaches. These approaches are setting HRT (hydraulic retention time), maintaining the pH level at 5.5, introducing toxins (chemical compounds) to methanogens, and accomplishing thermal shocks.

The dark fermentation cycle could be improved for producing VFAs and H<sub>2</sub>. The H<sub>2</sub> is a by-product that is generated from the metabolic reaction during the biological process and VFAs, altered by the following actions: (1) the operative settings (i.e. temperature, HRT, pH, OLR [organic loading rate], and SRT [solids retention time]); (2) orientation of the dark fermentation and substrate feeding strategies; and (3) the reactor feeds by using different types of organic waste (Fig. 12.4).

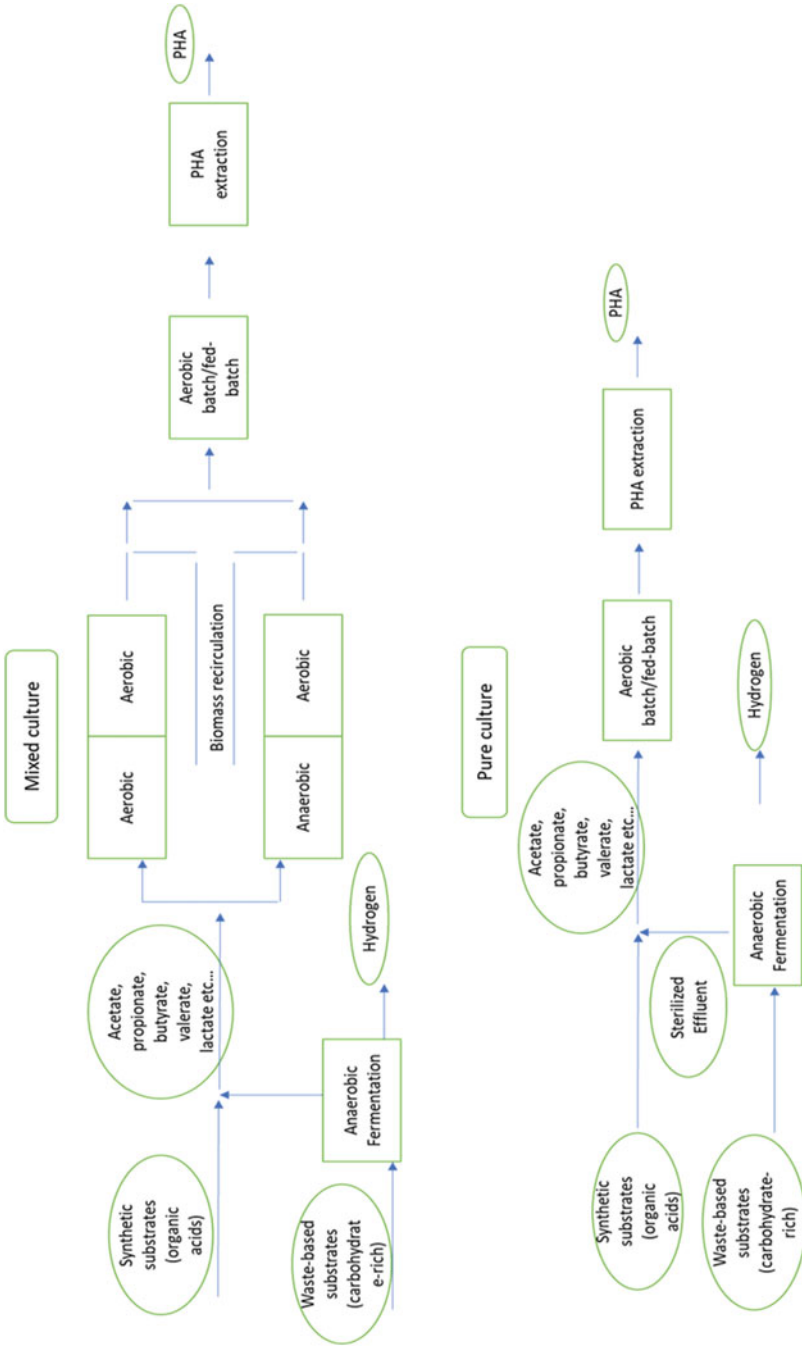
Many microbes have been reported for the PHAs production, such as *A. eutrophus*, *B. megaterium*, *Rhizobium*, *A. beijerincki*, *Nocardia*, and *P. oleoverans* commonly using formic acid, acetic acid, and propionic acid as substrates (Suriyamongkol et al. 2007). *A. eutrophus* and *A. beijerincki* have been reported as suitable microbes for PHAs synthesis and showed maximum PHAs content of up to 70% of DCW, beneath the nitrogen and phosphorus limiting strategies whereas *Rizobium* sp. and *Pseudomonas* sp. showed PHAS accumulation up to 60% of DCW (Suriyamongkol et al. 2007).

PHAs production has been seen in some other bacterial strains beneath adverse environments with different PHAs yields. Amongst them, H<sub>2</sub> and PHAs production have been obtained from several purple non-sulfur bacteria, similar to *R. sphaeroides*, *Rhodospirillum rubrum*, *Rhodopseudomonas palustris*, *Rhodopseudomonas palustris*, and *Bacillus* sp., beneath nutrient-limiting strategies (Singh Saharan et al. 2014).

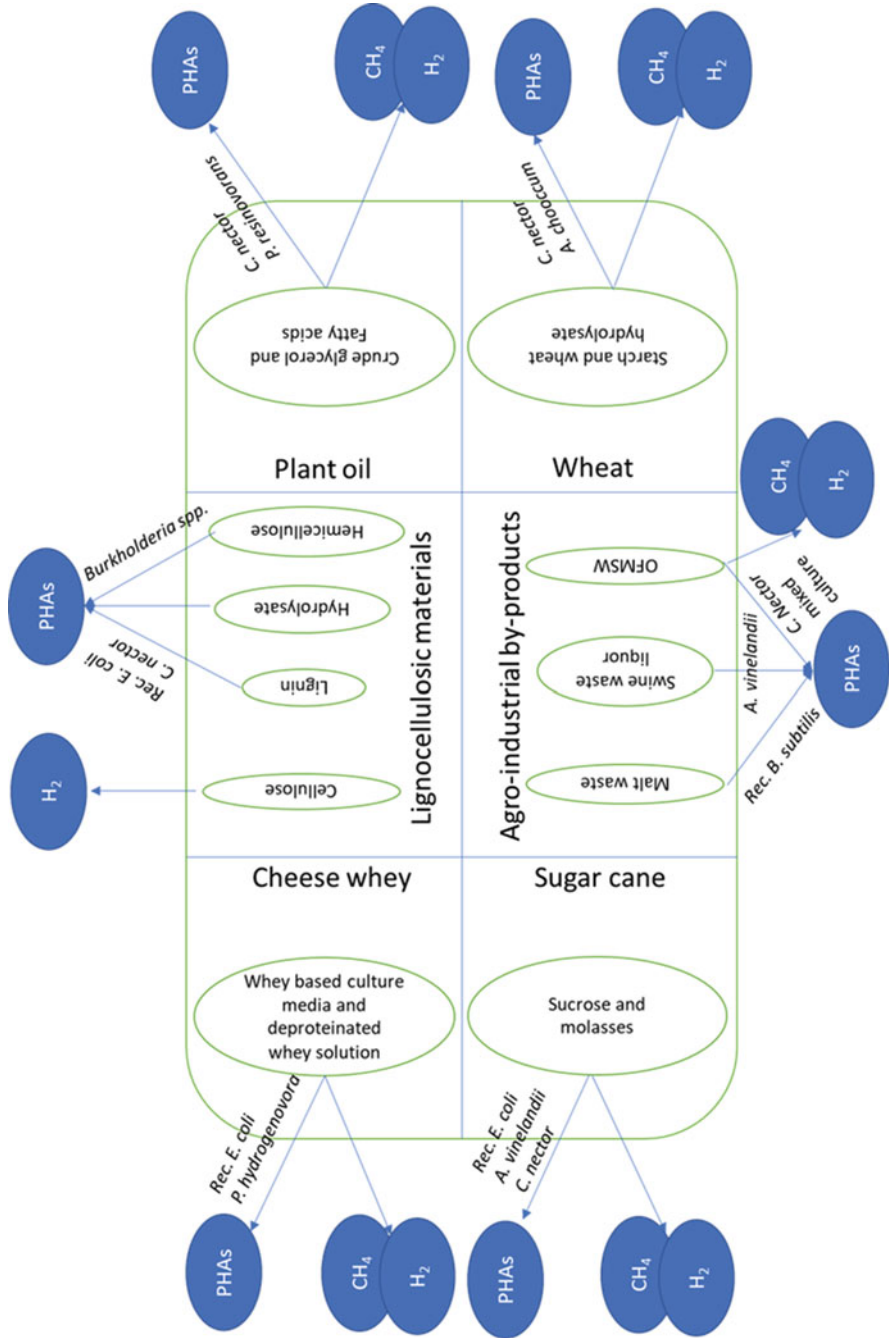
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## 12.6 Conclusions

With advanced and eco-sustainable skills, organic waste can be utilised to produce bioenergy and biochemicals by successful action of biological processes, individually or simultaneously. Bioprocesses can provide bioenergy or commercially



**Fig. 12.3** Cycle of polyhydroxyalkanoates (PHAs) production system. (Adapted from Serafim et al. 2008)



**Fig. 12.4** The organic wastes used for the production of PHAs and bioenergy, and also summarises the different bacterial species for by-products transformation

scalable chemicals while reducing pollution based on practical viability, economics, societal needs, and ease of use. Biologically formed plastics (bioplastic) can replace petrochemically created plastics, using competent bacteria fed with organic wastes and by-products as a substrate if the product is cost-effective.

In this context, different organic substrates and by-products can be used to produce bioenergy (hydrogen and methane) and biopolymers (PHAs). Otherwise, the review highlights the possibility of integrating the two production processes to design a unique energy and biopolymer production system. The integrated system seems to be a flexible process that aims (1) to produce organic acids from complex organic solid wastes rich in carbohydrates; (2) to use selected microbial strains or mixed cultures that show the highest capacity for PHA accumulation under specific dynamic feeding conditions; and (3) to produce bioenergy or accumulate PHAs by microorganisms from acidogenic effluents.

This integrated system represents new perspectives on the use of valorising organic substrates, organic waste, and their by-products for the production of both bioenergy and PHAs.

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**Conflict of Interest** There is no conflict of interest to declare.

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# Nanotechnology: Opportunity and Challenges in Waste Management

# 13

Arun Sharma, Brijendra Kumar Kashyap, Om P. S. Patel,  
and Arun Pareek

## Abstract

As a developing country, India has practiced fast-paced urbanization over the past three decades: from 1990 to 2021, its urban population increased by 260 million. This chapter provides a combination of literature and experimental data to trace the potential of waste materials for sustainable energy production in India. In the context of the Swachh Bharat Mission undertaken by the Ministry of Urban Development, is scientific treatment of Municipal Solid Waste (MSW) produced in India. Most of the successful technologies in the waste-to-energy sector were designed mainly in developed countries, including gasification, pyrolyzation, anaerobic digestion, and landfill gas recovery, which were suitable to handle segregated waste, which may be biodegradable, non-biodegradable, and hazardous. Solid waste management (SWM) is a significant problem for many urban local bodies (ULBs) in India, where it is a major challenge in metro cities like Mumbai, Chennai, Delhi, Kolkata, Hyderabad, Bangalore, Pune, and Ahmedabad, with high population density. Nanotechnology has emerged as a multipurpose proposal that could provide efficient, lucrative, and eco-friendly solutions to produce energy from waste compounds. Recent advances are explored to develop the opportunities of utilizing nanotechnology to address

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A. Sharma (✉)

Department of Chemistry, MITS Group of Institutions, Pali, India

B. K. Kashyap

Department of Biotechnology Engineering, Institute of Engineering and Technology, Bundelkhand University, Jhansi, Uttar Pradesh, India

O. P. S. Patel

Department of Technical Education, Government Polytechnic Naraini, Banda, Uttar Pradesh, India

A. Pareek

Department of Chemistry, Samrat Prithviraj Chauhan Government College, Ajmer, India

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the following points: (1) Nanomaterials for waste treatment; (2) green energy production; (3) management of waste materials; (4) manufacturing advancement and chemistry; and (5) reuse and waste utilization. In addition to the practical challenges mentioned above, we also describe community perspectives and provide an outlook on the role of nanotechnology in the application of energy production from waste materials for sustainable development.

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**Keywords**

Nanotechnology · India · Waste management · Chemistry · Waste to energy

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### 13.1 Introduction

India is a diverse country with many different religious cultures, groups, and traditions. Current systems in India face many environmental challenges mainly associated with the generation of a large volume of waste material and its inappropriate collection, transportation, improper treatment, and inadequate disposal. Sustainable solid waste management (SWM) is challenging in India because of rapid population growth (Kaur and Deswal 2019). Current SWM systems are inappropriate, with a negative impact of waste on the environment, public health, and economy. Here, there is an urgent need to move to more development in social, economic, and environmental areas that require new technologies for waste management. Ministry of Environment and Forests (MoEF) has already introduced waste Management and Handling Rules (Sambyal 2020). This chapter reviews the challenges, barriers, and opportunities associated with improving waste management and energy production in India with the help of nanotechnology.

To produce energy, waste-to-energy technologies may also recover useful materials and free land from dumping issues. A significant increase in the use of waste-to-energy technologies has been proposed. This depends on various factors such as climate, location, demographics, and other socioeconomic factors. The thermal treatment of residual waste such as combustion, pyrolysis, etc., can provide heat and power. Due to various operational and design problems, thermal treatment has not worked effectively. For example, in 1987, the first large-scale MSW incinerator built at Timarpur, New Delhi, could process 300 tonnes/day and cost Rs. 250 million (US\$ 5.7 million). The plant failed because of seasonal variations in waste composition and properties, poor waste segregation, inappropriate technology selection and maintenance, and operational issues (Gidarakos et al. 2006) Despite this experience, nanotechnology can play a key role in future waste management in India for waste-to-energy production. In addition, energy generation from waste would have significant social, environmental, and economic benefits for India.

Nano technological products and Nanomaterials are expected to contribute significantly to a clean environment from waste and protect the climate by reducing greenhouse gases and hazardous wastes and producing pollution-free energy (Guerra et al. 2018). Nanomaterials reveal exceptional chemical and physical properties, which make them attractive for improving novel and environmentally friendly

products (Rahimi and Doostmohammadi 2019). In future, nanotechnology may contribute significantly to climate protection and solving our energy-related problems. Some specific examples of nanotechnology applications that are widely used to benefit the environment include highly efficient, ecofriendly, and reusable batteries, the use of titanate nanofibers for the removal of radioactive ions from water which also act as a good adsorbing material, magnetic nanocomposites for clean-up of oil spills, nanofilters (graphene nanoflakes) for water purification, artificial photosynthesis in generation of hydrogen-powered technologies with the help of nanostructured polymeric materials and many more (Verma et al. 2021; Wiek et al. 2012).

In the twenty-first century, the world is facing a drastic waste disposal problem. India is a growing industrialized country that needs to be more focused in this context. Pyrolysis, sanitary landfills, and incineration, etc. are the commonly used methods which are especially non-ecofriendly, expensive, and time-consuming (Kawai and Osako 2013). Waste treatment is more effective by using nanotechnology modification concepts based on efficient nano-filters and Ag, Cu, Zinc oxide, TiO<sub>2</sub> nanoparticles (NPs), Carbon Nanotubes (CNT), etc (Nath 2018; Zhang et al. 2016). As compared to other methods, nanotechnology provides the best possible solutions for solving the issues of waste disposal. According to central pollution control board, India is the tenth most industrialized country in the world with about 88% of industrial clusters scattered all over the country (Sunil et al. 2017). Pulp and paper industries, thermal power plants, textiles, and steel and iron industries are mainly responsible for river water pollution and the effects can be seen in the case of Plachimada, Kerala and in the Tungabhadra sub basin, Karnataka (Panigrahi and Pattnaik 2019; Yadav et al. 2020). Due to this river water pollution, health, environment, and economy around these rivers are adversely affected. Over the past two decades, approximately more than ₹1500 Crore has been spent on the River Yamuna's water treatment by the Government of India, but is still found to be very toxic (Parween et al. 2017). In Yamuna river water, several unidentified by-products have been found during its recent examination. The main source of wastewater are household waste, street sweepings, commercial waste, clinics and dispensaries, hotel and restaurants, construction and sludge, as observed by the national solid waste association of India. At present, blazing in air, disposing in ocean, sanitary landfills, incineration, manure formation, ploughing in farms, crush, mixing, and disposing of waste into sewers, etc. are commonly used methods of waste disposal (Ahuja 2017). Approximately 48% of waste produced is organic in nature, which can be easily converted into reusable, high quality compost, and the remaining waste can be recycled to obtain useful materials (Pappu et al. 2007; Rai et al. 2020). Generally, public think about waste management of solid materials that will be the ultimate solution for its disposal and typically the land filling is the first click in their minds. To get rid of waste, the most important aspect is the proper channel treatment for getting the ultimate state of waste management. In the natural environment, active microbes decompose the wet matter to produce manure in the optimum composition of water, oxygen, nitrogen, and carbon (Gupta et al. 2018). Active microbes successfully do the composting and break down wet organic matter

into composite. Since, many strategies have been followed from time to time, these days, nanotechnology is the new hope for efficient management in the advancement of waste disposal treatment (Dermatas et al. 2018). Nanotechnology can provide solutions for challenging social issues in terms of reducing waste production, cleaning up industrial contamination, recycling and reusing wastewater that is safe for drinking and good for living of aquatic biota. Nanotechnology is the most effective tool for the treatment of waste disposal as it makes the filters, sensors, metal removal screeners in combination with solar energy that makes it more effective and efficient when used for its process. By using nano-level filtration process, many industrial pollutants such as Bisphenol-A, Alkyl Phenol, Phthalates, etc. could be separated from polluted water (Singha and Kumar Mishrab 2020). Nano-filtration process is combined with other technologies in many industrial waste-treatment plants to produce effluents with less concentration of industrial waste as well as produce energy.

### 13.2 Waste Generation in India

Rapid population growth in India has played a major role in increasing MSW. Compared with 1028 million population of India in 2001, it has been enhanced in 2013 and was 1252 million (Census of India 2011), and currently is 1591 million (Worldometer 2021). Based on Worldometer elaboration of the latest United Nations data, the current population of India is 1591 million (Worldometer 2021).

As shown in Table 13.1, rapid population magnification in every decade is found. Ahmedabad (6.3 million), Hyderabad (7.7 million), Bangalore (8.4 million), Chennai (8.6 million), Kolkata (14.1 million), Delhi (16.3 million) and Greater Mumbai (18.4 million) are the main listed metro cities in India (Bhattacharyya

**Table 13.1** Population growth between 1911 and 2011 in India. Source: Provisional Population Totals-India, 2011 (Census of India 2011)

Census year	Population $\times 10^6$	Decadal growth $\times 10^6$	Progressive growth rate (compared with 1911)
1911	252	13.7	5.75
1921	251.3	-0.8	5.42
1931	278.9	27.6	17.02
1941	318.6	39.7	33.67
1951	361.1	42.4	51.47
1961	439.2	78.1	84.25
1971	548.1	108.9	129.94
1981	683.3	135.1	186.64
1991	846.4	163.1	255.05
2001	1028.7	182.3	331.52
2011	1210.2	181.4	407.64

**Table 13.2** Census of India (2011), CPCB Report 2011. Major cities in India and per capita waste generation data (2010–2011) as given (Census of India 2011)

Metro city	Population (2011) $\times 10^6$	Total waste generated (tonnes/day)	Waste generation (g/capita/day)
Ahmedabad	6.3	2300	0.36
Hyderabad	7.7	4200	0.54
Bangalore	8.4	3700	0.44
Chennai	8.6	4500	0.52
Kolkata	14.1	3670	0.26
Delhi	16.3	5800	0.41
Mumbai	18.4	6500	0.35

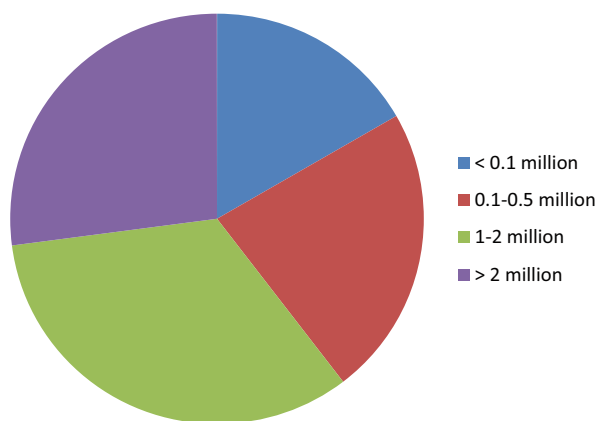
2017). As shown in Table 13.2, these metro cities have high waste generation per capita (Rimaitytė et al. 2012).

Successful waste management planning involves forecasting of future waste generation that is fundamental to estimating the quantity of waste and characteristics of MSW in India (Kumar et al. 2017). Various factors such as living standards, the extent and type of commercial activity, eating habits, and season play key roles to determine the quantity of MSW generated (Kirubakaran et al. 2005). Approximately 133,760 tonnes MSW is generated in India per day, in which approximately 91,152 tonnes MSW is collected and approx. 25,884 tonnes MSW is treated (Joshi and Ahmed 2016). As shown in Table 13.3, in small towns and in cities, MSW generation per capita in India is found to be approximately 0.17 kg/person/day and approximately 0.62 kg/person/day, respectively (Kumar et al. 2009).

Various factors such as population density, economic status, level of commercial activity, culture, and city/region may alter waste generation rate. High MSW generated in states of Maharashtra (115,364–19,204 tonnes/day), Uttar Pradesh, Tamil Nadu, West Bengal (11,523–15,363 tonnes/day), Andhra Pradesh, Kerala (7683–11,522 tonnes/day) and Madhya Pradesh, Rajasthan, Gujarat, Karnataka and Mizoram (3842–7662 tonnes/day). Lower waste generation is observed in Jammu and Kashmir, Bihar, Jharkhand, Chhattisgarh, Orissa, Goa, Assam, Arunachal Pradesh, Meghalaya, Tripura, Nagaland, and Manipur (less than 3841 tonnes/day) (Rajamanikam et al. 2014).

### 13.2.1 Waste-to-Energy in India

Material recovery techniques are one of the most promising approaches which can be used to solve the problems associated with improper waste disposal. In India, inert and high moisture content fractions are separated from the source, increasing the potential for thermal recovery. In thermal recovery, residual waste is processed that will leave over after all commercially feasible and recyclable materials are extracted (Jouhara et al. 2018). Energy is produced, valuable materials are recovered, and useless land could be used for waste dumping using Waste-to-energy technologies.

**Table 13.3** Per capita waste generation in Indian cities. Source: (Mondal et al. 2018; Kumar et al. 2009)**Per capita waste generation in Indian cities**

Cities with a population range	Number of cities including	Waste generation rate (g/capita/day)
<0.1 million	8	0.17–0.54
0.1–0.5 million	11	0.22–0.59
1–2 million	16	0.19–0.53
>2 million	13	0.22–0.62

Determination of the exact composition of residual waste and MSW is vital for energy recovery, is very much changing in India and usually increases with the increasing amount of high calorific waste (Al-Khateeb et al. 2017). Many things such as location, climate, demographics, and other socioeconomic factors are significantly responsible for increased waste-to-energy technologies (Malinauskaite et al. 2017; Rana et al. 2017; Gómez et al. 2009).

The most widely used waste-to-energy technology is combustion to provide combined heat and power from the residual waste (Oko and Nwachukwu 2018). To reduce dumping, an integrated waste management system would play a significant role in India. Recycling techniques are also convenient to adopt for achieving waste-to-energy. In India, industries are keen to use unsegregated low-calorific value waste that can be processed by the available waste-to-energy technologies such as combustion, incineration, gasification, pyrolysis, production of refuse-derived fuel, and gas-plasma technology, etc. are currently being developed (Chand Malav et al. 2020).

In India, waste-to-energy advancement is based on a build, control, and transfer model. The increased use of waste-to-energy techniques would effectively generate clean and reliable energy from a renewable fuel resource and minimize land disposal, reducing dependence on fossil fuels and, moreover, reducing greenhouse gas emissions (Chaudhary and Pathak 2020; Madakka et al. 2020). Furthermore, India

would have significant social and economic benefits via the generation of energy from waste compounds. However, some difficulties are found in India, which would appear in the pathway of waste-to-energy. Due to various operational and design problems, many amenities have not worked effectively. Despite these experiences for future waste management in India, the waste-to-energy technique will play a key role in its development.

### 13.2.2 Manufacturing Advancement and Chemistry

This chapter describes waste management in developing countries, especially in the Indian subcontinent, which comprises industrial hazardous waste, medical waste, solid waste, wastewater, agricultural waste, construction and household waste. The current scenario aims to reevaluate the already developed strategies that are particularly associated with toxic materials as waste management. India, as one of the most progressive developing countries, is deficient in a systematic practical approach to control waste management programs formulated by government and local bodies; shows a lack of ability to efficiently gather and administer wastes, and minimize the unconstructive impact of such activities. Inadequate hazardous waste treatment and its final disposal are not being properly addressed with existing regulations and regulatory frameworks due to fragmented responsibilities among local authorities and government sanitation departments. To improve the current situation, this chapter provides the best practical solutions for hazardous waste management (Mmereki et al. 2016). It is necessary to develop innovative ideas in developing countries and medical engineering background for the designing of low-cost simple-to-use devices, equipment, infrastructure requirements, and sophisticated controls to treat and detoxify these wastes with less human physical involvement to reduce the danger for environmental and public health impacts. The most suitable approach to detoxifying the surface and surrounding area is using hazardous waste disposal with cost-effective technologies, easily driven in the face of limited infrastructure, technical expertise and knowledge for developing countries (Nandi 2014). Phytoremediation is one of the most popular long-lasting, and highly efficient innovative processes which involve tree plantation, preserving environmental resources such as soil, water conservation, etc. and detoxifying hazardous wastes, generally used in China, India, Sri Lanka, Nepal, Bhutan, Pakistan, etc. (Sharma et al. 2018). Hazardous waste management is becoming a major issue in developing countries owing to the variety of waste streams consisting of toxic materials, which adversely affect the environment and health of living beings. Hence, numerous realistic methodologies are suggested, which include the following point, e.g.:

- Develop awareness in public places about recycling wastes
- Reduce waste production at its origin of source
- Build capacity and develop manpower for waste recycling
- Controlled waste management systems, which continuously monitored, reported on time and evaluated performances

- Appropriate infrastructure development and timely implement technical guidance as given by experts
- Reformation of regulatory frameworks and local bodies' reinforcement
- Recognizing and developing appropriate technologies for waste treatment and disposal
- Specified most promising and state-of-the-art technologies and selected proposals for financial support
- The local waste management system is rejuvenated and acquires maintenance

The most effective approach to increase revenue in industrial manufacturing is waste removal. There are many ideas to classify waste minimization, which mainly includes all practices such as prevention of waste, recycling and reuse that may reduce the large amounts of waste entering the environment and polluting it at various levels (Rosenfeld and Feng 2011). More specifically, practices generally adopted by industries for waste minimization include:

- Modified designs of products
- Customized inventory management
- Altering operational procedures and maintenance
- Material research and change with the best alternative
- Equipment substitution or cost-effective modifications
- Reuse and recycle waste materials

Adopting various approaches for waste management in manufacturing industries could minimize waste production and maximize output in energy generation. In this manner, metallurgical, pharmaceutical, food processing, plastic, and polymer-based manufacturing factories can control their processing cost when using waste management techniques and help eliminate waste (Woodard and Curran Inc. 2006). The more waste is produced in manufacturing industries, the more negatively does it affect the production cost and requires a larger area. To overcome these issues, industries are more focused on minimizing waste production and eliminating unwanted materials, drastically increasing productivity and lowering the manufacturing cost per unit article based on total waste management programs. Technocrats utilize various waste management strategies that have solved traditional problems to reduce the quantity of waste produced during manufacturing, which gives a large amount of a manufacturer's profits in the business. Many researchers found solutions for traditional issues to innovate better tools to reduce waste generation. DuPont has developed a methodology that systematically identifies prospects to decrease the amount of waste produced by industries (All Answers Ltd. 2018). Studies performed by DuPont examined waste assembly practices in reverse, beginning with the waste streams and tracing back to their source, addressing significant challenges at each step to minimize or eliminate the overall quantity of waste produced. The process operates as a screening tool for potential waste minimization protocols, vital which techniques are paramount and that can be applied easily. The government is seriously concerned and calling for various



proposals from young technocrats helpful in waste reduction and with environment protection as their center of attention that benefits society and the manufacturer's profit. Subsequently, a list for the waste reduction or minimization process is mentioned as follows:

- *Resource utilization.* Proper utilization of raw materials can reduce or minimize waste production at the individual level as well as benefit various industries.
- *Scrap Material Recycling.* In this process, manufacturing plants can reuse or recycle unwanted materials into a purposeful design, reducing pollution and creating more technological development opportunities.
- *Process monitoring and Quality control improvement.* This is the most widely adopted strategy by various industries to provide the best quality for their consumers. Frequently monitoring the equipment improves efficiency and gives maximum output.
- *Waste Exchange.* one of the most feasible approaches for reducing waste is to find a type of waste produced in a manufacturing unit that could be used up as starting materials for another route of processing units.
- *Supply chain.* In practice of many commercially available platforms those having trend of using variety of undesirable packaging materials which will be responsible for generating one more cause of waste. It can be reduced at the time of packaging and delivered as it is ordered. These supply chain modification strategies significantly reduce waste materials production and maintain an ecofriendly atmosphere.

Developing more environmentally benign chemical products and processes encompassing the design, manufacture, and use of efficient, effective, and safe materials are applied with Sustainable Chemistry (Parmar et al. 2018). Concerning the Sustainable Development Goals, the idea of Sustainable Chemistry might provide an essential tool to reach these objectives, incorporating a large number of targets for proving solutions for harmful chemicals and waste management (Friege 2017).

Sustainable chemistry contributes to the most promising approaches for providing maximum energy output via reducing waste materials and resolve environmental challenges: reuse/recycle of waste and recovered at the source of its generation in order to get useful fractions which can be exploited in the formation of another product (Barra and González 2018). Subsequently, the fundamentals of Sustainable Chemistry are as follows:

- Advanced resource organization and growing exercise of waste-derived renewable assets exclusive of endangering foodstuff production
- “Benign by design” means using less toxic and biodegradable compounds under natural conditions, creating a pollution-free environment
- “Design for recycling” is useful for designing products which are possible to recycle and convert into other useful materials

Novel technological developments in the twenty-first century give chemical solutions for reducing plastic waste compounds in degradable components and detoxifying hazardous liquid wastes (Thiounn and Smith 2020). Due to inappropriate liquid waste management, the extensive use of toxic chemicals by many industries, especially in manufacturing plants, causes major global challenges. Liquid wastes pollute water resources and fertile agricultural soil and seriously affect the environment causing irreversible damage, if not properly rectified (Redouane and Mourad 2016). Recently, researchers also developed a method to purify toxic chemicals using Bismuth oxychloride (BiOCl) nanoparticles, which showed excellent photo catalytic power to decontaminate toxic chemicals in the presence of sunlight under atmospheric conditions (Liu and Peng 2020). This technology can be used to destroy organic dyes in sunlight by their photocatalytic degradation. Reactive oxygen species contain oxygen free radicals, peroxides, and superoxides produced at the surface of nanoparticles in the presence of sunlight and are capable of destroying harmful organic dyes. In the presence of sunlight, Bismuth oxychloride nanoparticles have the potential for complete degradation of methylene blue chosen as a reference material which resembles in molecular characteristics to a large number of chemical wastages as produced by many industries (Huang et al. 2013). Studies reveal that the Bismuth oxychloride nanoparticles-based photo degradation process takes up to 4 h under optimized reaction medium, temperature, humidity, etc. One of the most effective properties of these nanoparticles is that they can be reused for 4–5 cycles, associated with 80% retention efficiency. However, when compared with solid waste, liquid waste is mainly associated with disposal problems due to the formation of several toxic by-products since their management requires special attention for researchers (Rasalingam et al. 2014). Nanomaterials-based newly developed techniques are most promising for waste-to-energy sustainable growth with 30–40% cost reduction in the treatment of liquid waste that saves expenses and gives industries more turnovers. Waste-to-energy can be produced by the process known as incineration, which is generally used for solid waste to burn finally after numerous steps of recycling.

### **13.2.3 Barriers and Changes Required to Improve Waste Management in India**

The present situation of SWM in India is unfortunate since the finest and most suitable processes, from waste collection to disposal, are not properly used. The main limitation associated is the availability of qualified waste management professionals and deficiency of training protocols in SWM in technical subjects, as well as less conscientiousness in existing SWM organizations all over India (Khajuria et al. 2010). Municipal authorities are responsible for supervising municipal solid waste (MSW) in India, but are facing financial issues regarding appropriate recycling of collected waste, its storage, disinfection, final treatment, and disposal. To achieve the goal of effective SWM, proper planning for MSW is very much required starting from waste collection to its final disposal under the supervision of

the government regulatory framework for its strategic execution in India (Narayana 2009).

Inadequate environmental awareness, joined with short stimulus, has repressed improvement in innovations and the acceptance of recent technologies that could make over waste management in India. Community thoughts is one of the most paramount factor in India for strengthening SWM. The Centre of visualization for waste management in India is the reuse of wastes as assets with increased extraction value, recycling, and recovery to generate revenue and obtain energy efficiency (Sridevi et al. 2012). Local urban authorities have to be responsible for waste management associated with the respective urban local body. The respective authority that may be a Chairman, Director and commissioner must have to honestly straightforward for guidance, performance, and evaluation of waste management systems. Waste management requires a sufficient budget from cumulative Indian society as part of their service to achieve the goal of sustainable development (Rawat et al. 2013).

To develop SWM in India, a well-established and autonomous organization is essential to control waste management. In the absence of comprehensible guidelines and a set of laws, it is complicated to achieve modernization in waste minimization. The waste management division wants to incorporate smart and beneficial production with comprehensible performance desires enforced by the local body framework, with monetary penalties applied in case waste management services are not effectively working. Budget for waste management authorities; provide necessary resources, and funding for infrastructure must be increased from waste producers output and converted to get other valuable materials having some application. A standard charge of ₹1 /person/day would collect approximately ₹50,000 crore per annum. This intensity of financial support would be enough to afford successful waste management throughout India (Yadav et al. 2010).

In future, the quantity of waste production and its characterization data is crucial as this establishes the suitability of diverse waste management and recovery options. Necessary equipment and vehicles are required and must be procured by State-level regulatory authorities to monitor primary and secondary waste collection, proper transport, treatment, and ultimate disposal. Waste in the streets and littering is the foremost problem in India that severely impacts communal health. Nagpur has practiced a method for sweeping roads in which every worker sweeps a fixed road length and collects the waste materials. Centre for Development Communication's scheme, the Swatchata Doot Aplya Dari (sanitary worker at the doorstep), was selected as a model of good tradition by UN-HABITAT in 2007 (Nolan 2015).

Waste segregation at the source must be involved in waste management practices to allocate much more proficient quality extraction and recycling. Inorganic separating of dry and biodegradable wet waste would have considerable advantages and should be the waste producer's liability. In continuing waste management preparation requires creative schemes developed by urban local sanitation authorities involved in the private sector and NGOs. The roles and responsibilities to bring sustainable structures must be formulated, with regular monitoring and evaluation, to achieve systematic progress. Knowledge should be mutually transformed by

**Table 13.4** Scientific, engineering, and organizational challenges for waste management in India (De Snyder et al. 2011)

Scientific challenges	Engineering challenges	Organizational challenges
Cost of reusable and recyclable materials	Waste collection, segregation, transportation and systematic disposal	Structuring of the waste management system and organizational setup
Disposal of produced waste and remnants	Practicable cost-effective technological solutions in terms of recycle, recovery, reuse, and reduce	Training and awareness for waste management
Solution for eco-friendly revitalization		

**Fig. 13.1** Scientific, engineering, and organizational challenges for waste management in India



connecting diverse areas of India and different societal groups. Many research institutions, R&D organizations, private sector companies, and NGOs are working independently or in collaboration on solid waste management using a holistic approach, and upcoming waste management in India must engage widespread contribution of the informal segment all over the system.

Establishing training and capacity building at every level, from school education to research and development, is necessary. The awareness should be spread to each person in the waste management system about the importance of waste management and the negative effects of poor waste management on the atmosphere and public health (Agarwal Siddharth et al. 2007). This strategy will effectively develop responsible citizens who consider waste as a resource opportunity, as shown in Table 13.4 and Fig. 13.1.

### 13.3 Nanomaterials for Waste Treatment

Depleting water resources reduces drinking water as limiting resources because of a speedily growing population and climate change that results in the occurrence of prolonged droughts and floods in many areas of the world. Therefore, reprocessing or recycling water available in any form will help to take the edge off this challenge. Conversely, huge quantities of liquid wastes and effluents, produced by commercial, manufacturing, domestic, municipal, and natural activities, can produce alarming situation to the ecosystem and individual health (Harrington 1978). Many previously developed chemical, biological, and physical technologies have been used to purify water and waste treatment from water bodies to produce pollutant-free water. Moreover, these traditional methods are especially concerned on primary treatment of impure water that is mainly based on the manual removal of insoluble solid particulate matter and high amounts of toxic elements such as phosphorus, heavy metals, nitrogenous compounds and other ionic impurities, which are dangerous for the environment, if released into it (Rajasulochana and Preethy 2016). Thus, the most recent technology, i.e., nanotechnology, is effective in the advanced treatment of waste-water via nanomaterials and nanosorbents. Various forms of nanomaterials have been developed to enhance productivity and boost the elimination of selective components from wastewater, such as filtration membranes, separating highly efficient molecular sieves, catalytic and absorption materials, etc. This chapter addresses the latest progress in wastewater management by means of nanotechnology and the implications associated with significance in water purification in developing countries, with special emphasis on using nanomaterials in wastewater treatment to develop strategies companionable to technologies related to wastewater management. Additionally, latent planning for using nanomaterials in wastewater treatment applications, their limitations, and official frameworks was also considered (Nnaji et al. 2018).

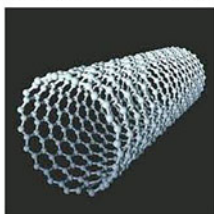
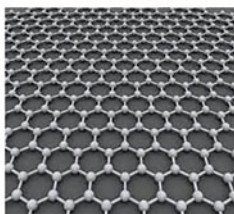
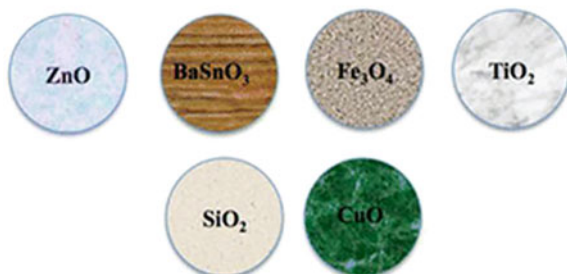
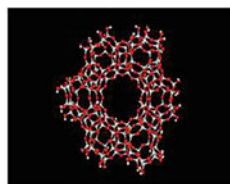
In the course of fast expansion of compounds of a fast expansion of compounds based on nanomaterials and sustainable nanotechnology to resolve environmental challenges has given consideration for growing trepidation in the last two decades. There are enormous applications of nanotechnology for wastewater treatment to improve better performances and higher efficiencies of water purification techniques as well as eliminate contaminants from water streams to achieve sustainable approaches for safe and sound water supply (Khan et al. 2012). This chapter describes modern technological advancements using nanostructure compounds to treat wastewater. This involves the synthesis of tremendously efficient and highly effective nanomaterials with unique chemical and physical properties, mainly carbon-based nanomaterials, i.e., carbon nanospheres, CNTs, carbon quantum dots, and metallic nanomaterials. The metallic nanomaterials (metal and metal oxides nanoparticles and noble metallic nanoparticles) were especially targeted due to their characteristically strong performance and suitable mechanisms for eliminating and absorbing various contaminants which are organic-based, inorganic-type and inert in nature. Many issues have been realized concerning the large-scale application of nanomaterials in wastewater treatment; these are mainly

aggregation or agglomeration of particles, complicated segregation, leaching when coming into contact with water, which are associated with adverse toxic effects forced on the environment and the health of living beings. Nanocomposite compounds are one of the most promising and evolutionary materials exhibiting compatibility with functional ligands to improve absorption of many impurities of different phases and properties. This chapter especially focuses on environmentally friendly nanocomposites, including organic and inorganic materials, nanofilms, ultrafine membranes, and magnetic nanoparticles. The perspective and applications of these nanocomposites and nanosized compounds are briefly discussed (Zhang et al. 2016).

Suitable nanomaterials and their selective approach for targeted toxins and contaminants under optimized conditions may alter their efficiency in waste treatment. Since waste management mainly depends on the selection of appropriate nanomaterials, among the large variety nanomaterials synthesized in the laboratory, some are found suitable in this context and have been synthesized in required quantities at the industrial level. Currently, metal nanomaterials are most commonly used, especially iron-based nanoparticles and carbon nanomaterials. On the other hand, zero-valent iron (nZVI) is the most widely used nanomaterial in environmental protection and is synthesized in optimum quantities (Li et al. 2017). Another widespread metal-based nanomaterial is titanium dioxide ( $\text{TiO}_2$ ) nano-sized particles. These nanoparticles are mostly used as excellent photocatalysts due to their outstanding photosensitivity and heavy metals adsorbing agents (Kinsinger et al. 2015). These nanomaterials generate hydroxyl radicals when exposed to ultraviolet (UV) light or direct sunlight, which are highly reactive and can quickly oxidize contaminants. In advanced oxidation processes, these hydroxyl radicals are applied for water treatment. For optimum photocatalytic activation in the ultraviolet (UV) radiation by Titanium dioxide ( $\text{TiO}_2$ ) nanoparticles, having large band gap energy can provide maximum output. These nanoparticles act as excellent catalysts and remain the same during degradation, since they are chemically stable in water and can easily undergo different reduction processes. These ( $\text{TiO}_2$ ) nanoparticles exhibit strong antimicrobial activity due to the formation of hydroxyl radicals possessing low toxicity, and could be synthesized in low-cost scheme (Azizi-Lalabadi et al. 2019).

Carbon nanotubes (CNTs) are the most commonly used carbon-based nanomaterials. CNTs are cylindrical shaped nanostructures, and allotropic forms of carbon. CNTs can be classified as single-walled CNTs and multi-wall CNTs, based on their synthesis. CNTs have a large specific surface area which acts as highly assessable adsorption site for toxic substances. Their modifiable surface chemistry (CNTs tubular structure) and their compatibility for surface functionalization boost their sorption capabilities. Consequently, CNTs are excellent as adsorbing materials for heavy metals, contaminants in polar and non-polar organic forms, and oils (Poudel and Li 2018). Compared to other nanomaterials, CNTs can be regenerated and reused without change in properties.

Many other metallic nanomaterials, consisting of silver, zinc, bimetallic nanoparticles, polymeric nano adsorbents with various shapes, and magnetic

**1. Carbon nanotubes (CNTs)****2. Graphene****3. Fullerene****4. Metal oxide nanoparticles****5. Zeolite**

**Fig. 13.2** Carbon Nanotubes (CNTs), Graphene, Fullerene, and Metallic nanomaterials consisting of oxides of silicon, zinc, barium, titanium, copper, iron, and Zeolite (Yang et al. 2013)

nanoparticles (as shown in Fig. 13.2) are also efficient for the management of liquid waste. Apart from the aforementioned nanomaterials, these specified nanoparticles are also utilized in technologies for developing waste management strategies (Yang et al. 2013). Principles used for developing these techniques are based on nanomembrane ultra filtration, using adsorption and separation of undesired materials with nanosized photo catalysts. Nanoparticles exhibit size-dependent properties and are capable of absorbing pollutants on their surface due to their high surface-to-volume ratio. In recent years, membrane filtration and separation process attracted researchers for removing a large number of waste materials (Singha and Kumar Mishrab 2020). Separating membranes in different pore sizes may act like a barrier wall for noxious waste molecules. The electrochemical deposition technique is applied to embed nanomaterials in the template of ultrafine nanocomposite membranes and can modify their surface. CNTs and other nanomaterials can be applied to improve the physical strength and mechanical properties of polymeric nanocomposed membranes, including its efficiency toward fouling in corrosive medium and retaining in liquid contaminants with excellent permeability for getting desirable quality of the purified water. The advanced oxidation photocatalytic process can oxidize many contaminants and microorganisms present in the waste and decompose them into non-toxic and environmentally safe compounds. Titanium dioxide ( $\text{TiO}_2$ ) nanoparticles are used in heterogeneous photocatalysis and degrade many organic contaminants. However, separation and

catalytic processes are combined to develop new strategies for waste materials. This combination will direct the foundation of membrane-based photocatalytic reactors, which will be proficient in managing liquid waste channels by real-time preservation of the catalytic nanoparticles (Dermatas et al. 2018). Traditionally, the most economical, technical, and feasible option is the use of an adsorption-based technique. Study in treating dumped waste using the adsorption technique has resulted in the creation of definite compounds for the expulsion of metallic substances in the solution. These materials are mainly natural products like Peat Kaolin, Zeolite, activated carbon, clay, aluminosilicates, and polysaccharides.

Carbon-based nanomaterials, especially CNTs, are being used as exceptional adsorbents having excellent efficiency due to their large specific surface area. Compared to generally used carbon-based powder and activated granular carbon, the Multiwalled carbon Nanotubes (MWCNTs) possess excellent metal-ion sorption capacity. In the application of filtration, nanomaterials with controlled shapes, densities, and dimensions can be used to build structures. Cylindrical nanoporous membranes easily filter out small microorganisms (sizes ranging from 28 to 65 nm) (Singh et al. 2020). CNTs are widely used for water filtration in various forms. In the thrust area of science and technology, many neutral (zero-valent) metal nanoparticles, such as Al, Zn, Fe, and Ni, are used to treat water pollution. At the nano-range highly reducing metals, such as Fe, Al, Ni, and Zn in the presence of water, zero valent aluminum (Al) is thermodynamically unstable. It is responsible for the deposition of oxides/hydroxides on the peripheral surface. It impedes 100% transfer of electrons from the metallic peripheral surface to the contaminant compounds (Jiang et al. 2018).

### 13.3.1 Nanotechnology for Green Energy Production

Nanostructure materials such as CNTs, graphenes, fullerenes, and quantum dots are currently being used to make lightweight, cost-effective, and more efficient solar cells. The improved surface-area-to-volume ratio of these nanoparticles increased the collection of solar radiation at the panel by revealing more conducting surfaces to solar light. Moreover, lead selenide (PbSe) nanoparticles result in more conductivity since more electrons are ejected (and therefore gain more electricity) when struck by quanta of light. Additionally, nanotechnology modifies the structural properties and design of photovoltaic cells and enhances windmills' effectiveness. CNTs' surface modification by epoxy groups provide good-strength long blades and reduce windmill weight. Thus the amount of electricity generated by such windmills is greater than conventional windmills. Nano paints increase the life period and durability of turbines and blades of windmills (Echiegu 2016). Furthermore, the world faces many great challenges, e.g., food, water, energy, shelter, healthcare, employment, electronic devices, cars, and aeroplanes, etc., in reducing time and minimizing efforts of manpower on the Earth's global environment and climate. Nanotechnology is one of the most promising approaches as multipurpose utilities could provide solutions for many challenging issues toward achieving highly efficient, cost-effective, and



environmentally friendly global sustainability challenges facing society. This chapter is committed to utilizing nanotechnology to achieve sustainable development in producing energy from waste. The main highlights in topical advancements and advancements and giving up the opportunities associated with utilizing nanotechnology to deal with worldwide challenges in (1) water purification and treatment; (2) clean and green energy technologies; (3) management of greenhouse gases; (4) utilization and supply of materials; and (5) hygienic manufacturing and green chemistry. Aforementioned, many technical challenges are greatly demanded to outlook societal perspectives and the responsibility of nanotechnology in the convergence of knowledge, skills, and civilization for achieving sustainable improvement (Diallo et al. 2013). These are some selected nanomaterials and nanoparticles that are presently being utilized as building blocks to expand the upcoming invention of sustainable commodities and technologies with respect to water purification and treatment, energy production, its renovation, and storage, management of greenhouse gases, utilization and supply of materials, and cost-effective manufacturing using green chemistry.

Industrialized manufacturing is indispensable to a sustainable social economy. In both developed and developing countries, this is the key mechanism that drives innovations and develops higher job values (Malinga et al. 2013). Industrialized manufacturing has an intense environmental footprint. Firstly, it needs many resources such as materials, energy, and water. Secondly, it produces many types of wastes, e.g., gaseous, liquid, and solid, and toxic compounds that require proper disposal of such materials and convert them into risk-free products. Nanotechnology is rising as an empowering tool for eco-friendly manufacturing and green chemistry in various areas such as chemical, semiconductor, materials design and processing, petrochemical, pharmaceutical, and many other industries (Shannon et al. 2008; Park et al. 2012). Opportunities and challenges impart on the state-of-the-art nanomanufacturing and sustainability various strategies, adopted for achieving waste to energy (Brinker and Ginger 2011). In order to achieve the target of “long-term sustainability,” one needs to reduce the cost of manufacturing articles and industrial tools. This is one of the most popular and feasible assembly-based processes. It is being used because of its rapidity, simplicity, easy to work at room temperature, and the ambient pressure conditions. It could even be extensively diminished by minimizing the utilization of materials, energy, water, and waste generation. Nanotechnology and biotechnology, in combination, open new horizons to develop detoxified and “green chemistry” routes for synthesizing functional nanomaterials using bacteria, fungi, and plants that can provide environmentally acceptable solutions (Brundtland 1987).

### 13.3.2 Nanotechnology for Management of Waste Materials

Nanotechnology is one of the most promising approaches in this context for reducing the amount of waste generated in various forms. Its efficiency toward energy production is significantly based on the choice of appropriate nanomaterials for the

targeted impurities. It is feasible due to the availability of large numbers of already synthesized nanomaterials at the existing conditions. Carbon-based and iron-based nanomaterials are selectively manufactured at the industrial level among various nanomaterials produced in laboratory-scale conditions so far. Zero-valent iron (nZVI) is the most extensively used nanomaterial specifically designed to protect environmental issues. Several research groups have reached the desirable results for the effectiveness of nZVI material to get a reductive breakdown of organic impurities, especially for chlorinated hydrocarbons (Lacalle et al. 2020). Moreover, nZVI has a very high surface-to-volume ratio to absorb heavy metals, e.g., hexavalent chromium, effectively on their surface and convert them into non-toxic compounds via a mechanism followed by redox reactions at the surface of nZVI particles (Dong et al. 2019). Another approach of nZVI is to eliminate the inorganic components (e.g., nitrates) to provide highly efficient treatment for wastewater streams. Considerable utilization of nZVI is well acknowledged to its perspective for getting modified surfaces to strengthen its selectivity in the direction of definite contaminants and toxic materials and its stabilization. Furthermore, nZVI, combined with noble characteristics holding metals such as palladium or platinum, called “bimetallic nZVI,” increases their surface area effectively and frequently catalyzes the redox reactions, thereby enhancing the rate of reaction when reacting with targeted contaminants (Ma and Zhang 2008). The main disadvantage associated with nZVI nanoparticles is their possible aggregation or agglomeration with challenging storage problems which may reduce their high reactivity with respect to time elapsed. In particular, the reactivity of nZVI nanoparticles toward various naturally occurring water constituents is also found significantly, affecting their potential reactivity and surface inactivation when reacting with the target contaminants.

Additionally, Titanium dioxide ( $\text{TiO}_2$ ) nanoparticles exhibit excellent activity as photocatalysts due to their high photosensitivity and behave as good adsorbing material for heavy metals. In presence of ultraviolet (UV) radiation, titanium oxide ( $\text{TiO}_2$ ) nanoparticles generate hydroxyl radicals which can oxidize contaminants and show high reactivity for waste materials. The water treatment method is generally termed advanced oxidation process, where hydroxyl radicals are significantly used. Wide bandgap energy is associated with  $\text{TiO}_2$  nanoparticles so, for maximum photocatalytic activation, UV radiation is highly required. These  $\text{TiO}_2$  nanoparticles are chemically stable, insoluble in water, exhibit strong antimicrobial activity, act like a catalyst, remain unaffected during the degradation process, and easily go through different reduction processes. Many applications are reported for hydroxyl radicals produced by  $\text{TiO}_2$  nanoparticles with considerable advantages such as low toxicity and cost-effectiveness. Nanotechnology can be employed with waste management technologies either in situ or ex situ. A large number of advantages are associated with in situ technologies where the nanotechnology is utilized in the form of an absorbent reactive barrier, e.g., nanoporous membranes. The reactive region in nanoporous membranes is perpendicularly in the stream pathway of the targeted subsurface cloud of pollutants in general, caused by negligent waste management practices (Dadrasnia et al. 2013). Nanoparticles of nZVI exhibit tremendous physical and chemical properties for significant removal of various pollutants by their

adsorption on nanoparticles' surface using latest technologies such as photocatalytic absorption, ultrafine nanomembrane filtration, and specified segregation of contaminants.

In recent years, membrane filtration and separation process has also gained significant interest in developed countries. Nanoporous membrane size is comparable with the size of contaminant molecules, which act as barriers to removing pollutants. Nanomaterials such as CNTs and polymeric nanocomposites can be applied for developing ultrafine membranes formed by available traditional methods, which include surface deposition and embedment. This significantly improves the mechanical strength and enlarges water permeability as opposed to the fouling membrane surface.  $\text{TiO}_2$  nanoparticles can reduce a variety of organic contaminants by heterogeneous photocatalysis mechanisms. Owing to their unique properties and extraordinary functional compatibility, nanomaterials are widely used for waste management compared to the already developed conventional management technologies. It is generally expected that nanomaterials' function will decrease the overall waste management cost, fulfill energy demands, and increase the process efficiency followed via a simple mechanism. However, their elevated production charge restricted their industrial manufacturing for a time period. More importantly, there are still significant awareness gaps pertaining to their fate, and transportation associated with severe environmental damage and affecting living beings must be well measured and identified prior to the extensive use of nanotechnology in the waste management sector (Bora and Dutta 2014).

### 13.3.3 Nanotechnology for Reuse and Waste Utilization

At the present time, people are well aware regarding recycling, which includes sanitization of waste materials such as washed bottles, spoiled cans, and unused cardboard into big sizes, decorating recycling materials and placed at appropriate places. Subsequently, these recyclable materials are transported to the compilation plant, where they are segregated, cleaned, and converted into fresh materials designed to provide modified components useful in manufacturing. The most promising advantage of recycling is that it reuses raw materials that would not be used anywhere, protects valuable natural resources, and reduces energy consumption and the amount of waste sent to landfills and incinerators (Denison 1996). This reduces greenhouse gas emissions and lowers the overall production costs, saving the environment by protecting it from the leaching of toxic chemicals from dumped sanitary landfills. Technological development requires the recycling of nanomaterials to make use in many household products or creates useful desirable materials due to their unique chemical and physical properties. Many household products are available in the market that incorporate various nanomaterials. For example, television and computer displays incorporate catalytic gold nanoparticles and inorganic quantum dots. An inevitable consequence of this surge in use is that nanomaterials are increasingly prevalent in waste streams—the more we buy something, the more it gets thrown out! Only a few recycling and reuse strategies have

been developed for nanoparticles. To date, nanomaterials designing, recycling, and reuse strategies are very much challenging. Researchers have made efforts to obtain their practical utility, which is relatively simple, low cost, fast, and energy efficient. Some research groups have used powerful magnets to separate iron-based nanomaterials from complex mixtures, wastewater treatment, and powdered solid waste (Testa-Anta et al. 2018). New methods such as extraction techniques, separation columns, etc. have been developed to reuse high-priced gold nanoparticles from different liquids and mixtures. A newly developed concept utilizes waste materials to synthesize various nanomaterials using a top-down approach. For example, unwanted plastic material and polythene bags have been utilized to produce small-size carbon nanoparticles (less than 10 nm in size) known as carbon quantum dots with interesting photo-optical properties and emerging applications as imaging agents (Meng et al. 2010; Abdelbasir et al. 2020). These carbon quantum dots were prepared by cleanly cutting the plastic bags into tiny particles and heating these small pieces in the optimized concentration of hydrogen peroxide ( $H_2O_2$ ) solution, easily available in a first aid box as an antiseptic liquid. Furthermore, the waste repurposing method requires a furnace and sand to form silicon carbide nanoparticles using heat discarded compact discs (CDs). Silicon carbide nanoparticles exhibit outstanding thermal, physical, chemical, and mechanical properties. The use of CDs for nanomaterials generation is remarkable due to the increasing use of electronic devices. The rapid urbanization is responsible to create environmental non-degradable wastes. In addition to electronic waste, glass is another commonly used household product, and its various forms for different purposes are available in the market. Mainly, silicon element in the form of silicates is found as a basic constituent in glass manufacturing. A research group has recently developed a method to convert this waste glass (silica) into silicon nanomaterials with different shapes that can be the best alternative for energy storage in the form of long-lasting and rechargeable batteries (Abhuri et al. 2020). Since the method described is highly efficient and can be used as one of the best tools for providing electrical energy from waste broken glass articles. In this straightforward method, firstly, glass bottles are crushed into small pieces, then washed with isopropanol solution, a very commonly used ingredient in many household materials (such as antiseptics, disinfectants, and liquid detergents), then uniformly mixed with salt. This mixture is then put in the kiln for heating in the presence of magnesium to produce silicon nanoparticles. In the furnace, magnesium enables the silica to be converted into silicon; salt plays an important role in adsorbing the produced heat during the chemical reaction and prevents the agglomeration of silicon nanoparticles due to their high surface-to-volume ratio and provide stability. The developed synthetic scheme provides highly active silicon nanoparticles in the era of energy production to make a very effective, lightweight, rechargeable energy storage lithium-ion fuel cell battery. This is one of the best alternatives to conventional, large-sized, lead-acid-free batteries found to be used in most vehicles. The intrinsic significance of nanoparticles which require recuperating and recycling such valuable materials, has developed the impetus among researchers for recycling nanomaterials and generating them from traditional waste components. Using green chemistry

approaches, such methodologies can be implemented on a large scale, which helps to reduce energy consumption and minimize waste accumulation. These reasons attract the scientific community to recycle nanomaterials in various forms of environmental protection with sustainable growth. It will be useful for developing countries to achieve the goal of waste-to-energy production in the coming years (Bankar 2018). Production and processing are the backbones of global development, using ways to reduce the amount of waste production and life-lasting, user-friendly products are constantly rising. As estimated in studies, till year 2025, solid wastes generated will reach about 2 billion tonnes/year; it is highly required to develop new technological process for effective recycling and reuse of waste materials. The amount of waste generated is minimized in three ways, and the impact of nanotechnology can be seen. First is the recycling of nanomaterials, second is nanoprocessing for recycling solid wastes, and the last is upgrading existing processes. Nanoprocess for waste management is very effective in providing technological intelligence, innovative products, easy procedures, and a wide application range for many industries such as nanovarnishing, ultrafine nanocoatings, nano healthcare equipment, nanomedicine in pharmaceuticals, and nanomanufacturing. To generate renewable energy sources, significant contribution of nanotechnology is found in current industrial development. It is essential for reducing environmental pollution and continuous sustainable growth, protection/preservation of the ecosystem, and waste management (Vega-Baudrit 2017).

Various sectors, including plastic technology, printer ink, textiles factories, cosmetic materials, sunscreen lotions, surface cleaning materials, vehicle modifications, and games commodities, require engineered nanomaterials (ENMs) such as titanium dioxide (nano-TiO<sub>2</sub>), zinc oxide (ZnO), silver nanoparticles, gold nanoparticles, C<sub>60</sub> fullerene nanostructures, carbon Nanotubes (CNTs), graphene sheets, and silica (SiO<sub>2</sub>) nanoparticles that have been integrated (Jeevanandam et al. 2018; Santos et al. 2015). It depends on available resources; regulatory authorities may alter extensively from nation to nation and affect the quantity of recycled waste and recovery of materials. In many regions, “zero waste” or “circular economy” strategies are adopted for getting better efficiencies in recycling and recovery of waste materials which depends on the collection system, type of composed waste, and existing technological development. Moreover, ENM-containing wastes are hazardous to the environment since they require advanced chemical, biological, thermal, or physical treatment for their reuse and recycling (Part et al. 2018).

Traditional methods available for wastewater treatment are costly; in several cases, not efficient due to the scarcity of treatment process. Consequently, novel approaches are regularly being required, which may be used as a way of enhancement in conventional wastewater management methods. The chapter provides an outline of improvement in the area of nanotechnology for waste management, observes the potential of nanomaterials on human strength and the ecosystem, and encourages novel techniques for minimizing waste and producing energy by means of nanotechnology. The greatest emphasis has been placed on synthesizing advanced nanomaterials for removing toxic compounds using nanoporous membrane, photo degradation of pollutants using catalytic nanoparticles, disinfectants based on

non-toxic nanomaterials for wastewater treatment and adsorption of pollutants on their surface (Maksimović and Omanović-Miklićanin 2017). Various nanomaterials are in diverse applications of research and development owing to their unique functionality, chemical reactivity, and physical properties. Some nanoparticles detoxify contaminants (catalytic oxidation process), while others can filter these contaminants (nanomembrane-based separation and isolation process) (Yunus et al. 2012). Dioxins are efficiently absorbed by carbon nanotubes (CNTs), which are more effective than activated carbon. Most nanomaterials and nanotechnological processes still need to be developed in the new stage of scientific research.

Furthermore, analytical procedures and laboratory tests are essential to better perceive these processes and characterize their broad application potential. Recently developed technologies are concerned about their prospective impacts on the global market, regarding maintenance and easy-to-recycle nanomaterials and protect environmental risks due to their accumulation in contaminants. Much effort is required to curtail the key limitations of nanotechnology and acquire complete environmental benefits via solving waste minimization in terms of environmental protection. Two essential features that make nanoparticles very good adsorbing substances are their large surface area and multifunctionality, making them competent to react chemically or physically with various targeted molecules. Such properties give long-term stability to nanoparticles for various contaminants and make them suitable for photocatalytic degradation of toxic species in wastewater streams. Carbon Nanotubes (CNTs) have many applications compared to activated carbon in terms of absorption efficiency, surface area, 3D-arrangement of carbon atoms, unique mechanical, electrical, physical, chemical, and functionalization compatibility to bind with a variety of contaminant molecules, e.g., heavy metals, pathogens, and organic pollutants. This is why they are called “material of the twenty-first century.” Besides carbon nanotubes, several metal-based nanoparticles such as iron oxides ( $\text{Fe}_x\text{O}_y$ ), silicon (Si), titanium (Ti), and tungsten (W) also exhibit adsorption characteristics for many substances and unstable radioactive elements. Adsorption strength depends on pH, temperature, type of absorbent, and environmental conditions to reduce and recycle waste materials (Sadegh et al. 2017).

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## 13.4 Conclusion

Conventional technologies, i.e., pyrolysis and incineration, dealing with waste disposal treatment, are costly, time-consuming, with difficult operating protocols. It is not much effective for generating sufficient energy as compared to the more efficient, recently developed nanotechnology. This mainly includes ultrafine nanofiltration, carbon nanotubes (CNTs), nanoparticles, nanoadsorbents in treatment of dumped waste owing to their high surface-to-volume ratio. Moreover, nanotechnology is the most promising and advanced technology to provide an efficient approach to waste disposal treatment. In India, SWM is mainly due to increasing population growth and dense slum areas in metro cities. Inadequate waste infrastructure and dumping in open areas create major issues in the present scenario. It is time

to spread community awareness toward waste materials and their potentially harmful effects on the environment as well as on human health. Proper waste management is essential for sustainable development with special emphasis on minimization of waste production, recycling, treatment, and disposal to develop technology for achieving the goal of waste-to-energy. India faces challenges concerning the appropriate implementation of waste management policy, technological development, availability of resources, and well-trained personnel. In the absence of these primary prerequisite conditions, India continuously suffers severe impacts of untreated waste disposal on public health and faces many environmental issues.

The current situation of hazardous waste management in India is not satisfactory. This chapter incorporates the management of hazardous waste, such as biomedical waste, factory hazardous waste, household hazardous waste, etc. in developing countries. There are several key shortcomings with respect to hazardous waste management in developing countries associated due to lack of information on the amount of hazardous waste produced, lack of competence and awareness; very less inducement or penalty; lack of roles and responsibilities for stakeholders; limited resources, and lack of infrastructure; insufficient organizational framework, lack of technically expert personnel, economic assistance, how to treat, testing instrumentations, and facilities; lacking integrated skeleton regarding the supervision and management of hazardous wastes. Furthermore, insufficient waste collection, improper treatment and disposal systems, and apathetic management by the government make it complex for the local regulatory sectors to recognize the purpose of achieving appropriate and strategic management of solid waste. There is an urgent need to adopt best practices of hazardous waste management from developed countries, and its successful implementation in developing countries comes together with the context. In this context, nanotechnologies have been confirmed to be most promising and very efficient under normal laboratory conditions concerning wastewater treatment.

Moreover, nanotechnological developed tools are commercially applicable, cost-effective, practically usable, minimize waste generation, and eliminate awful environmental and public health impacts. This chapter exclusively focuses on the possibilities of nanotechnology in India with regard to energy produced by the waste treatment. Nanoparticles acquire high surface area, making them suitable candidates for absorbing contaminant molecules and for developing various sensors to detect pathogens, viruses, hazardous chemicals, etc. present in wastewater, soil, and the environment. Nanotechnology exhibits excellent compatibility with the available waste treatment methods. In developed countries, absolute sewerage setup with wastewater treatment plants incorporating advanced technology is already working. Specific nanomaterials can improve the efficiency of the existing system with the least amount of variations to the existing infrastructure. Advancement in the practicability of nanotechnology for water treatment strongly needs appropriate infrastructure in developing countries, especially for undeveloped provinces worldwide. In the framework of hi-tech growth and potential outlook for recovering competence and minimizing cost, essentially three varieties of nanomaterials preserve the most promising in the field: mainly nano-sized

adsorbents, ultrathin nanomembranes, and nanocatalysts. The aforementioned challenges concerning commercialization of nanomaterials, broad characterization spectrum, cost of production, technical characteristics, environmental impacts, etc. can be merely regarded as temporary interference. A warm association among every concerned establishment is demanded to overcome these issues. It can be anticipated that the practical approach of highly developed nanotechnology, coupled with cautious supervision designed to circumvent unwanted consequences, can build an enormous contribution to this area of research and establish itself as a superior waste treatment solution.

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# 'Omics' Approaches for Structural and Functional Insights of 'Waste to Energy' Microbiome

# 14

Ashutosh Kumar, Neeraj, Uma Chaurasiya, Deepak Kumar Maurya, Surochita Basu, Aniruddha Kumar, Sapan Patel, and Vineet Kumar Maurya

## Abstract

'Microbiome' represents all the microorganisms present in a given environment which can have large boundaries like forest or ocean ecosystems, small boundaries like pond, tree, pits, etc., or even smaller niches like the human gut. A microbiome is not merely a legion of microbes but interacts actively with its environment. Waste deposits, like waste landfills, sanitary landfills, waste

Ashutosh Kumar and Neeraj contributed equally with all other contributors.

A. Kumar

ICAR - Indian Institute of Seed Science, Maunath Bhanjan, Uttar Pradesh, India

Neeraj

Department of Computer Science and Engineering, H. N. B. Garhwal University (A Central University), Srinagar Garhwal, Uttarakhand, India

U. Chaurasiya

School of Studies in Botany, Jiwaji University, Gwalior, Madhya Pradesh, India

D. K. Maurya

Agharkar Research Institute, Pune, India

S. Basu

Department of Botany, Tripura University, Suryamaninagar, Tripura, India

A. Kumar

Department of Environmental Microbiology, Babasaheb Bhimrao Ambedkar University, Lucknow, Uttar Pradesh, India

S. Patel

School of Studies in Botany, Jiwaji University, Gwalior, Madhya Pradesh, India

V. K. Maurya (✉)

Department of Botany and Microbiology, H. N. B. Garhwal University (A Central University), Srinagar Garhwal, India

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disposal plants, sewage treatment plants, and wastewater-based bioreactors, are also unique types of ecosystem which have a distinct microbiome associated with it called 'microbiome of wastes' (MoW). Waste is not merely a garbage collection but has certain types of microbiome related to it, interacting actively with the waste and degrading it. 'Waste' and 'Energy' are the two major global concerns, and the concept of 'Energy from Waste' (EFW) can simultaneously deal with both of these problems. The study of MoW for EFW requires the isolation and identification of microorganisms present in it and their functional characterisation. Due to inherent limitations, the traditional culture-based techniques are inadequate for studying the complete diversity of a microbiome. Hence 'Omics'-based approaches are utilised for MoW research. Omics approaches involve meta-genomics, meta-transcriptomics, meta-proteomics, and metabolomics, which are used for studying various aspects of a microbiome. Metagenomics approaches are based upon the DNA isolation and amplification of different 16SrRNA/18SrRNA regions, followed by their phylogenetic analyses. Metagenomics provides accurate information about the complete microbial diversity of a microbiome, but does not provide any information about physiological processes of a microbiome. Hence, metatranscriptomics and metaproteomics approaches are used to analyse the genes, proteins, and enzymes being expressed by a microbiome, which reflects the physiological conditions of that environment. Primary and secondary metabolites of microbiome also affect the physiochemical condition of an environment, which are studied using metabolomics approaches. While the metagenomics and metatranscriptomics approaches are dependent upon the sequencing and their alignments, the meta-proteomics and metabolomics approaches depend upon mass spectrometry and database searching. An integrated Omics approach of metagenomics, metatranscriptomics, metaproteomics, and metabolomics is required for a comprehensive analysis of MoW. The Omics approaches, their brief methodology, advantages, and limitations are described in this chapter. Besides, computational technologies, which are the core of all the Omics approaches, have also been highlighted, and the development of dedicated computational algorithms are the need of the day.

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**Keywords**

Microbiome · 16SrRNA sequencing · DNA extraction · Peptides · MALDI-TOF-TOF · Nano-LC-MS-MS · Algorithms · Waste to energy

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## 14.1 Introduction

Surging demand for energy, along with other necessities like food, fibre, and shelter, is expected to increase with the global population (from ~7.7 billion in 2020 to ~9.7 billion, expected by 2050). Rapidly exhausting conventional energy sources

such as fossil fuel reserves, forest woods, etc. have led us to shift to renewable energy resources like solar energy (Kabir et al. 2018; Sharif et al. 2021), tidal energy (Chowdhury et al. 2021), hydropower (Yuksel 2010; Ope Olabiwonnun et al. 2021), wind energy (Neto et al. 2020), nuclear energy (Saidi and Omri 2020; Azam et al. 2021), etc.; moreover, carbon emissions during energy consumption also needs to be minimised. Another concern is waste production; the increased human population exacerbates the global waste production problem. The concept of 'energy from waste' (EFW) can be of dual benefit: on the one hand, it would reduce the global burden of waste, and on the other, it would fulfil the increased global demand for energy (Sharma et al. 2020; Srivastava et al. 2020; Munir et al. 2021). The burning of waste is the most common method for energy recovery, but it produces harmful chemicals. Production of methane in waste landfill areas and biogas plants is another form of EFW, which are more environmentally friendly than burning wastes (Lee et al. 2017; Glushkov et al. 2019). Common forms of EFW, like the production of methane, biogas, syngas and ethanol, are the product of physiochemical activities of 'microbiome of waste' (MoW). Hence, proper research on the microorganisms, their activities, enzymes, and metabolites present in an MoW would enable more efficient production of EFW. Information about the chemical nature of waste and the microorganism capable of degrading it can be utilised to identify the microbes capable of degrading the waste having known chemical composition. This could be used to construct more efficient EFW plants, where specific types of wastes can be converted into energy using selected microorganisms. Biotechnology, molecular biology, and microbiology-based approaches provide limited information about any 'biome', such as the presence of a finite number of genes, enzymes, or life forms (Kumar et al. 2015b; Bharati et al. 2020; Maurya et al. 2020). 'Omics' technologies – metagenomics, metatranscriptomics, metaproteomics, and metabolomics – being used in various facets of science are capable of furnishing the detailed information about MoW, and can be utilised for improvement of EFW technologies to make them more efficient, feasible, and globally acceptable (Jiang et al. 2019; Lee et al. 2019; Kumar et al. 2021a, b; Xu et al. 2022).

Based on the role of microorganisms, the technologies for EFW can be categorised into two groups: one that involves purely physical and chemical reactions without any involvement of microorganisms, and the second one that involves microorganisms for converting waste into energy (Karagoz et al. 2020). This chapter is focused on microorganisms in relation to EFW, i.e. 'microbiome of waste (MoW)', the use of 'Omics' techniques for the analyses of MoW, and their role in EFW.

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## 14.2 Microbiomes

Microbiome represents all the microorganisms present in and interacting within a given environment; this environment can have large boundaries like ecosystems, small boundaries like pond, tree, pits, etc., or an even smaller niche like the human gut. Mohr gave the term 'Microbiome' in 1952 (Mohr 1952). Microorganisms are



ubiquitous and play an important role by performing a variety of activities, often counted as 'indirect benefits' (Berg et al. 2020). Due to a microorganism's ubiquitous nature and ability to survive any extreme environment, the wastes are also occupied by various types of microorganisms which constitute the 'microbiome of wastes'. These microorganisms degrade the biodegradable waste into simple molecules and work as natural scavengers. Microbes could completely degrade almost all the waste of biological origin within due course of time. Still, the rate of waste generation overcomes the natural rate of waste degradation by microbes, resulting in piles of waste (of even biodegradable wastes in nature) everywhere. Apart from the slower speed of natural biodegradation by microbes, the inability of microbes to degrade non-biodegradable materials like plastic, rubber, etc. is another limitation of the natural biodegradation process (Adebayo and Obiekezie 2018; Moharir and Kumar 2019; Srivastava 2019; Rastogi et al. 2020). Although some research on plastic degrading bacteria have been published in recent years, most of them are at the laboratory level only (Urbanek et al. 2018; Yuan et al. 2018; Gambarini et al. 2021). Biotechnological approaches focusing on selected microbes from a 'microbiome', or 'Omics' approaches focusing on all microbes and their functional aspects of a 'microbiome' are preferably used for the study of a 'microbiome' (Kim et al. 2017; Rai et al. 2020; Zhang et al. 2019b; Fenske et al. 2020; Maurya et al. 2020).

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### 14.3 Waste and Energy

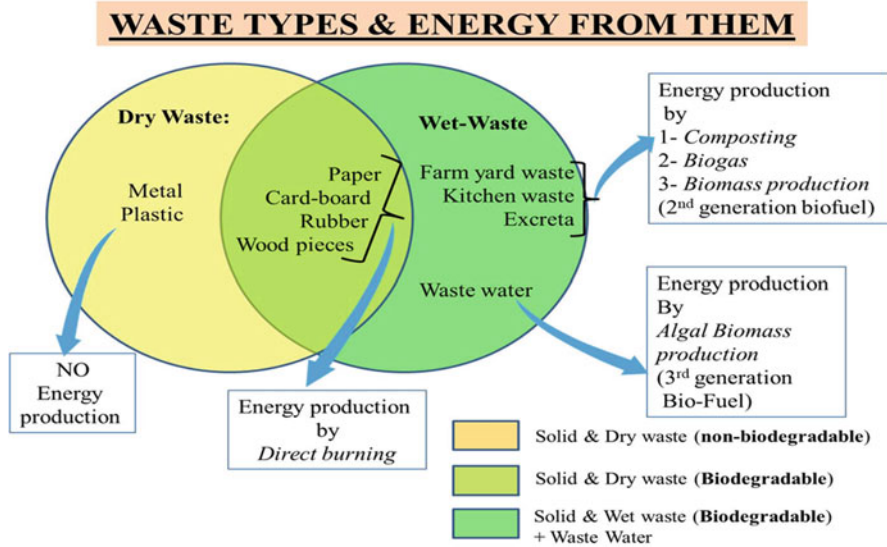
The problem of waste is not new to mankind, and has existed since the inception of civilisation when humans lived as nomadic groups. The only thing that has changed from nomadic to urban lifestyle, with respect to waste generation, is the 'waste quality'. Waste produced by nomads or rural inhabitants was mostly biodegradable, and much of this waste could be recycled. In contrast, most of the waste produced in urban areas is non-biodegradable in nature, leading to more waste accumulation and pollution than recycling. All human activities lead to the generation of some sort of wastes, and with rapidly increasing population and industrialisation, the quantity of waste is growing, and its quality is worsening. Waste generation is a never-ending process, and the waste generation rate is much faster than waste treatment, resulting in the worldwide accumulation of waste materials. Presently, ~7.6 billion humans produce 2.01 billion tonnes of waste globally per year. This amount is expected to increase by 70% by the year 2050 if proper waste management methods are not adopted (Kaza et al. 2018; David et al. 2020). Dumping waste, a common practice in rural and underdeveloped areas, occupies a significant area of land. In India, approximately 1400 km<sup>2</sup> of the area would be required for solid waste dumping by 2047, which could be otherwise used for other useful purposes. Besides wastage of land, the other negative aspects of waste are foul smell, unpleasant look, and pollution of land, water, and air. Air and water pollution, consequently, cause many human diseases like asthma, allergy, cancer, etc (Kumar and Agrawal 2020).

Waste can be categorised as biodegradable/non-biodegradable (based on nature of waste), solid waste/liquid wastes (based on physical nature of garbage), and agricultural waste/household waste/municipal waste/industrial waste (based on origin). Whatever the nature of waste, its disposal is always a problem, as any single waste disposal technology can't be applied to all sorts of garbage. Waste management deployed for handling the waste includes waste sorting, treatment, and recycling in an environment-friendly manner. Waste treatment and management strategies depend entirely on the waste's composition because biodegradable waste can be simply converted into valuable products by dumping it into pits (covered or open). In contrast, non-biodegradable wastes need different methodologies for their disposal altogether. Global analysis of wastes composition shows that non-biodegradable materials like plastic (12%), glass (5%), and metals (4%) altogether form 21% of the waste, bio-degradable materials like food and green wastes (44%), paper and cardboard (17%), rubber and leathers (2%), and wood (2%) altogether account for 65% of the waste, while the remaining 14% waste is composed of other uncategorised materials (Kaza et al. 2018; Yadav et al. 2020).

Besides the composition of waste, the economy and infrastructure of a nation also affect the mechanisms adopted for waste disposal. Landfilling, dumping, incineration, recycling, and composting are common waste management methods, but each has inherent limitations. Landfilling and dumping occupy a vast land area, which could be otherwise used for other more important purposes; incineration releases CO<sub>2</sub> and other harmful gases to the environment, causing air pollution. Recycling and composting are better options than landfilling, dumping, and incineration in terms of pollution and land requirements. Globally, landfilling is used to dispose of 36.6% of waste, followed by dumping garbage in an open area, which accounts for the disposal of 33% of waste. Although food and green wastes account for 44% of global waste, only 5.5% of waste is disposed of by composting. Recycling accounts for 13.5% of waste, while 11.1% of waste is incinerated. Amongst these waste disposal methods, only the sanitary landfill method, which accounts for 7.7% of waste disposal, is used to generate EFW in the form of gas. Another form of EFW is the production of biogas from biowaste materials using biogas plants (Horgan and Kenny 2011; Kumar et al. 2017; Singh et al. 2018; Lakshmikanthan 2019).

Waste management at the site of its production could be the best solution; however, it can be applied where most of the waste is bio-degradable and can be converted into compost, but this cannot be applied to other sorts of wastes, and availability of land for waste management is also limited. Waste management strategies usually convert waste into products other than energy; energy generation from these products, however, needs to be focused upon (Das et al. 2019).

Energy is one of the primary requirements of the time, and whose demand is continuously increasing. Continuous depletion of conventional energy sources (wood, coal, and petroleum) may lead to the problem of a global energy crisis in the future, and the world needs alternative and sustainable energy resources to deal with it. India is the third topmost consumer (~9%) of the total energy of the world, after China (23%) and the USA (17%). Energy availability, security, negative effects of fossil fuels on the environment, and improved standard of living have forced the



**Fig. 14.1** Types of waste and energy from waste

world to look for green, sustainable, and renewable energy alternatives. Hydropower and nuclear energy have been used for decades to meet the increased energy demands. Solar energy, wind, and tidal energy are also gaining attention as alternative energy resources. Energy generation from the tonnes of waste accumulated worldwide could be an alternative approach with immense potential for sustainable energy and is beneficial for waste management and energy generation (Zong et al. 2018). Energy from waste (EFW) is the process of conversion of wastes into energy in either electricity, heat, biogas (methane), biofuel, or synthetic fuel. EFW can be categorised into three categories: (1) energy from direct combustions with or without recovery of heat; (2) energy from the combustion of methane gas produced in sanitary landfills; (3) energy from the combustion of methane produced from anaerobic digestion of organic wastes; and (4) energy stored in the microbial cells in the form of lipids, which can be further converted into biofuels. The first three categories utilise solid waste, while the fourth utilises liquid waste (water).

All sorts of waste can't be used for EFW, and technology used for EFW depends upon the physical, chemical, and biological nature of waste. As shown in Fig. 14.1, only solid biodegradable wastes are utilised for direct energy generation by incineration. Sanitary landfills also generate combustible gases that can also be converted to energy. Liquid biodegradable waste, especially wastewater from households, can be used for indirect energy generation by utilising it for third generation of biofuel production (Chauhan and Maurya 2018; Gajraj et al. 2018). Microorganism-based EFW technologies involves 'anaerobic digestion' (biogas and alcohol production), 'fermentation' (alcohol production), 'landfill or Sanitary landfills with gas capture' (for methanol production), 'microbial fuel cells' (electricity, hydrogen generation),

'biochemical conversion' (biogas or biomass production), and 'carbon assimilation' (algae-based biofuel production). Integration of information related to microorganisms present in MoW, enzymatic reactions, and biochemical pathways helpful in converting waste material into energy-yielding forms is required to establish EFW technologies.

#### 14.4 'Omics' Approaches for Waste to Energy Microbiome

Among the vast microbial diversity, only few types of microbes are useful for EFW. The microbes that contain enzymes essential for converting complex waste biomass into gaseous or liquid biofuels are the only valuable microbes for EFW. The intervention of various 'Omics' technologies enable detailed analysis of MoW and different 'Omics' technologies are used to study different aspects of EFW using microorganisms, as shown in Fig. 14.2.

'Omics' refers to the collective technologies used to study diversity, functional role, and interaction of the pools of biological molecules and other entities that enable the characterisation of organisms' structure, function, and dynamics in an ecosystem. This ecosystem can be as small as a single cell or vast as a pond, forest, wasteland, etc. 'Omics' provides a holistic view about microbial diversity (metagenomics), functional diversity of mRNA (metatranscriptomics), enzymes and proteins (metaproteomics), and organic molecules (metabolomics), playing an active role in an ecosystem. 'Omics' technologies are known as the 'system biology approach' because all the components of a process or ecosystem can be analysed by combining these technologies. 'Omics' technologies deal with all the analytes

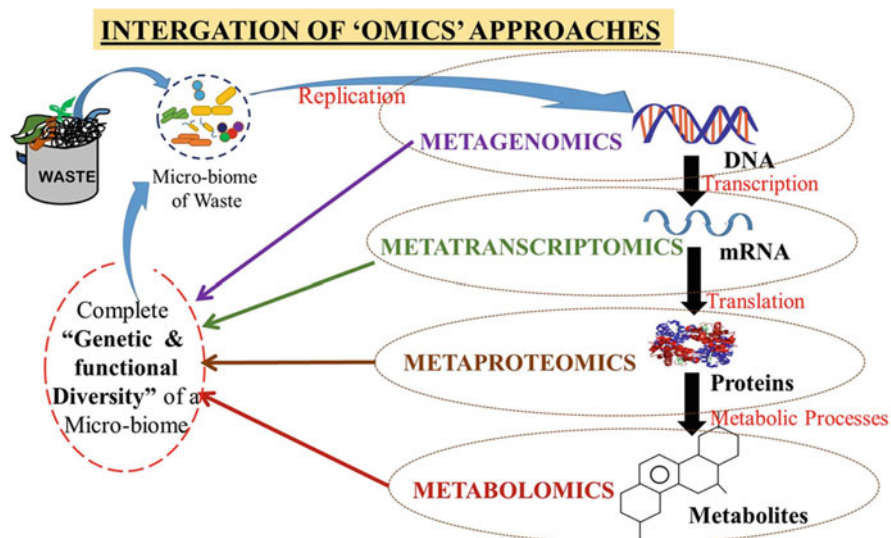


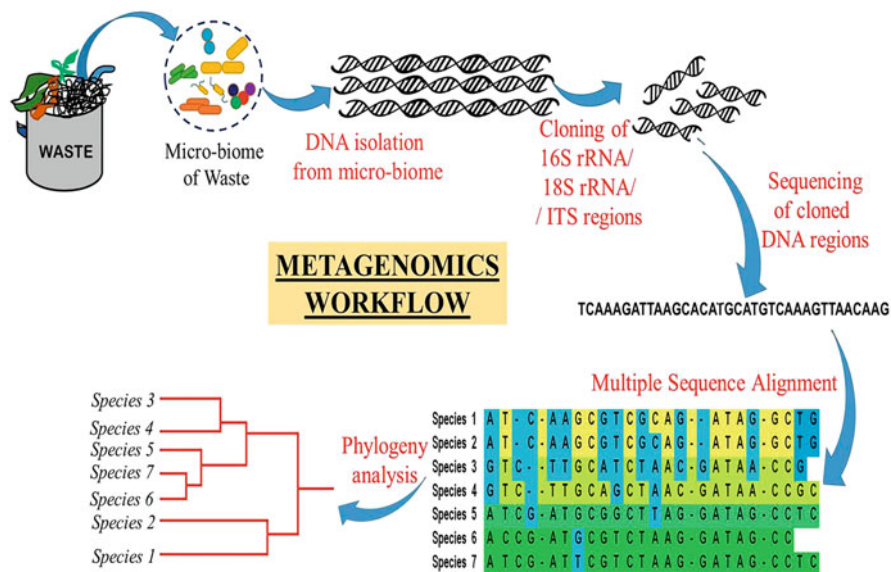
Fig. 14.2 Integration of 'Omics' approaches for different aspects of EFW

(DNA, mRNA, proteins, metabolites) in a non-biased and non-targeted manner. ‘Omics’ technologies are ‘high-dimensional’ technologies because of the nature of data generated by them. Data analysis requires strong computational infrastructure, novel algorithms and dedicated software. ‘Omics’ approaches are ‘top-down’ approaches where different components are studied together, and the metabolic networks are reconstructed (Horgan and Kenny 2011; Singh et al. 2018). In addition, Omics approaches can also be utilised for ‘bottom-up’ approaches, such as bioprospecting, in which the information obtained from omics experiments is used for improving the desired character of a life form (Kumar et al. 2019b). The following sections have discussed the details of various ‘Omics’ technologies and their specific application for the study of MoW.

#### 14.4.1 Metagenomics Technologies for EFW Microbiome

Different microorganisms performing various biological activities occupy a waste deposition site, which plays a vital role in waste degradation/disposal. Both types and number of microorganisms present in waste constitute the ‘microbiome of waste’. The classical microbiological technologies are useful for the analysis of only culturable microbial diversity, while the diversity of non-culturable microbes is out of their limits. Data on diversity analysis say that culturable diversity constitutes approximately only 1% of the total microbial diversity, while unculturable microbes constitute 99% (Amann et al. 1995). Metagenomics is used to analyse microbes’ remaining 99% non-culturable diversity (Cardenas and Tiedje 2008). The idea of ‘Metagenomics’ was first proposed by Pace et al. in 1986, and the term ‘Metagenome’ was coined by Handelsman et al. in 1998. Metagenomics can be divided into two categories: structural metagenomics and functional metagenomics. Structural metagenomics aims to analyse the structure (population compositions and population dynamics) of non-culturable microbial communities, which is determined by inter and intraspecific competitions, physiochemical and climatic parameters. Microbial community structure analysis allows a deeper understanding of the relationships between the individual components that build a community and is essential for deciphering ecological or biological functions amongst its members (Tringe et al. 2005; Vieites et al. 2008). The functional metagenomic approach aims to identify genes responsible for coding enzymes, proteins, and metabolites involved in a metabolic pathway. It differs from structural metagenomics in activity-based screenings rather than 16SrRNA-based analysis (Alves et al. 2018). The metagenomics analysis of an MoW enables the identification of microbes capable of degrading waste into metabolites, suitable for EFW.

Metagenomics-based microbial diversity analysis of the operational and non-operational municipal landfill sites demonstrated the dominance of Proteobacteria (55.7%) in both types of landfills. Bacteroidetes, Acidobacteria in active landfills, and Firmicutes, Actinobacteria at closed landfill waste sites were other dominant species after Proteobacteria (Zainun and Simarani 2018). Microbial community data collected from the three landfill sites showed that these landfill sites



**Fig. 14.3** Different steps of a metagenomics analysis

were occupied by different microbial communities dominated by the members of Proteobacteria and Chloroflexi. These sites were present with enzymes responsible for the degradation of dioxin, styrene, furfural, steroid, hydrocarbon, and cellulose. Enzymes involved in the biosynthesis of streptomycin, carbapenem, and monobactam were also reported. Different enzymes and microbes indicate that landfill sites can be exploited to develop an effective bioremediation process (Thakur et al. 2020). Zhang et al. (2017) used pyrosequencing-based metagenomics to demonstrate that the abundance of the predominant phyla Firmicutes, Elusimicrobia and Proteobacteria were selectively enhanced by 1.7–2.9 times after supplementing the medium with activated carbon. Using metagenomic analysis, Suksong et al. (2016) demonstrated that the process of anaerobic digestion is accelerated by the inoculation of *Ruminococcus* sp. (bacteria), *Clostridium* sp. (bacteria), and *Methanoculleus* sp. (Archaea) (Verma et al. 2018). There are many studies on community dynamics analysis, analysis of methanogenic pathways, microbial community structure relationship between diversity and function at the genome level, and the effects of environment and microbial communities on anaerobic digestion process for biogas generation from waste (Kumaraswamy and Kashyap 2021; Zhang et al. 2019a).

The workflow of metagenomics is shown in Fig. 14.3. The first step of a metagenomic experiment is sampling and extracting DNA from these samples (Verma et al. 2018). Collected samples must represent all the cells present at the sampling site. Underground and running water samples yield meagre amounts of DNA, which is insufficient for further steps of DNA amplification. Multiple displacement amplification (MDA) using random hexamers and phage phi29 polymerase increases

DNA yields from such samples. This method can amplify as minimum as femtograms of DNA to make micrograms of PCR product (Thomas et al. 2012). After DNA extraction, the selected segment of DNA is amplified using primers specific for that segment. The extracted DNA is amplified using primers specific for variable regions (V1–V9) of 16S rRNA (for bacteria and Archaea), 18S rRNA (for eukaryotic microbes), and ‘Internal Transcribed Spacer’ (ITS, ITS1 and ITS2), in particular ITS2 between the 5.8S and 28S rRNA genes (for fungi) (Ghosh et al. 2019). The amplified DNA sequences from either 16S rRNA, 18S rRNA and ITS segments are sequenced using an automated DNA sequencing technology. Although the slowest and costliest sequencing technology, Sanger sequencing technology is the gold standard for DNA sequencing. The technologies based on Sanger’s method have a minor error rate of 0.3%. The second-generation technologies, which are faster than the Sanger sequencing method, include (1) 454 Pyrosequencing technology by Roche; (2) ion-torrent Semi-conductor sequencing; (3) reversible terminator-Illumina/Solexa sequencing; and (4) Supported Oligonucleotide Ligation and Detection (SOLiD) technology by Applied Biosystems. The error rate in second-generation sequencing technologies is 0.1–1%. Third-generation technologies, which are even cheaper and faster than second-generation sequencing technologies, include (1) Single Molecules Real Time (SMRT) sequencing approach by Pacific Biosciences, and (2) Oxford Nanopore Sequencing (ONT). Third-generation sequencing technologies have the highest error rate of 12–15%. DNA sequencing data is a huge volume of data ranging from gigabytes to terabytes. These sequence data consist of smaller segments called ‘contigs’, of 200–400 nucleotide length. These contigs are assembled into larger fragments using reference-based assembly methods or de novo assembly methods. After assembly, the DNA contigs are grouped together into similar types of groups representing individual genome or genomes, and this process is called Binning. Binning can be either composition-based or similarity/homology-based binning. After binning, the functional, positional, and species information are assigned to each DNA sequence through the ‘Annotation’ process. The phylogenetic map of DNA sequences is prepared based on the annotation results. The process of assembly, binning, annotation, and phylogeny analysis are performed using dedicated statistical programmes, and algorithms contained in specialised software. A list of various software used for these purposes can be accessed in the work of Ghosh et al. (2019) and Zhang et al. (2019a). A review on plastic waste degrading microbial communities and other aspects of plastic waste has been presented by Akan et al. (2021), while Li et al. studied the effect of temperature change on the different microbial communities in food waste-based bioreactors using 16s rRNA sequencing (Li et al. 2022). They found that the temperature change significantly affected the bioreactor’s bacterial and archaeal community structure, and *acs*, *metF*, *coaA*, *mer*, *mch*, and *ftr* genes were upregulated in thermophilic reactors compared to the mesophilic bioreactors (Li et al. 2022).

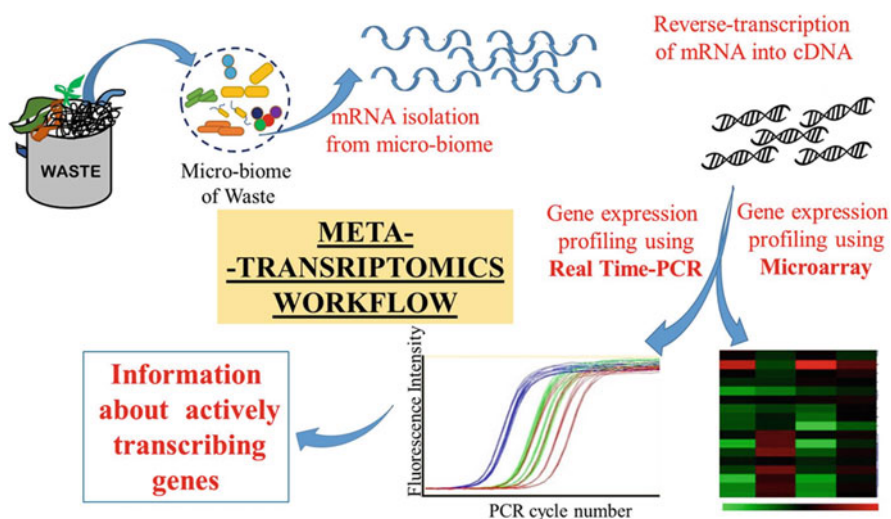
Metagenomics has a distinct advantage in identifying the non-culturable diversity of any microbiome, which is impossible for traditional microbial culture-based methods. It is a robust technology and can be used for the study of complete microbial diversity, comparison of microbial diversity of two/several different microbiomes and to study the dynamics of microbial communities over time. The

limitation of metagenomics is that it cannot provide the information about enzymes, proteins, and metabolites present in a microbiome, which determine the physiochemical nature of a microbiome. Other omics approaches could fulfil this limitation of metagenomics.

### 14.4.2 Metatranscriptomics Technology for EFW Microbiome

Metagenomics approaches reveal only the presence of organisms or genes and do not reveal anything/much about their activity. The information about actively transcribing genes by a microbiome is provided by 'metatranscriptome' analysis, which was introduced in the late nineteenth century. It gives insights into how a microbial community responds over time to their changing environmental conditions. Metatranscriptomics is culture-independent profiling of actively transcribed DNA from a given microbial community at a particular time under defined conditions. Metatranscriptomics elucidates the three important aspects of a given microbial community; gene expression abundance, gene activity diversity, and comparative gene expression analysis. Mass spectroscopy-coupled proteomics or meta-proteomics approaches are also used to provide gene expression data. Still, they require a reference genome or a reference meta-genome for peptides matching-based identification. Metatranscriptomics doesn't require a reference genome or meta-genome and can detect relatively low amounts of non-coding RNAs too, which are not detected by proteomics approaches (Warnecke and Hess 2009; Shakya et al. 2019).

The workflow of metatranscriptomics is shown in Fig. 14.4. The first step of a metatranscriptomics experiment is the extraction of mRNA from a given sample.



**Fig. 14.4** Different steps of a metatranscriptomics analysis



Techniques are available to directly extract mRNA from bacteria, archaea, fungi, and other eukaryotes, making the technology more accessible. As the mRNA degrades faster than DNA, the extracted mRNA is reverse transcribed into cDNA using reverse transcriptase. If it is impossible to prepare cDNA, then the mRNA sample must be stored in a deep freezer in an RNase-free buffer. Preparation of cDNA from mRNA involves three essential steps: (1) extraction of total RNA from the given sample; (2) mRNA enrichment by removing rRNA from the sample; (3) synthesis of cDNA synthesis from mRNA. Enrichment of mRNA is essential because it constitutes only 1–5% of the total RNA present in the cell. Suppose the initial concentration of cDNA is very low for DNA sequencing methods. In that case, it can be amplified using either of the following: RNA polymerase, multiple strand displacement amplification (MSDA), and emulsion PCR. Emulsion PCR is unbiased and promising for amplifying a very small amount of cDNA. As mentioned in the metagenomics section, the cDNAs are sequenced using automated DNA sequencers. Further steps of metatranscriptomics involve assembly, binning, annotation and analysis of the data, which are similar to the metagenomics data processing and analysis, as mentioned in Sect. 14.4.1 above (Warnecke and Hess 2009; Jouzani and Sharafi 2018; Shakya et al. 2019).

Metatranscriptomics provides deep insight into actively transcribing genes. Unlike PCR, it doesn't require gene-specific primers. Hence, all actively expressing genes are identified in a metatranscriptome study. The limitation of metatranscriptomics is that it relies upon mRNA, which is very unstable. Therefore proper care needs to be taken to avoid mRNA degradation. In addition, the deficient concentration of mRNA is also an issue with some samples. The limitation of metatranscriptomics is that all the mRNA of a cell is never translated into protein, hence it doesn't provide an accurate picture of protein translation status. Moreover, metatranscriptomics also remains silent about post-translational changes and protein isoforms, which are studied using metaproteomics.

Metatranscriptomics analysis of activated sludge from a municipal wastewater treatment plant in Hong Kong showed the expression level of the enzymes: ammonia mono-oxygenase (amoA, amoB, amoC) and hydroxylamine, related to nitrification was higher in activated sludge. In addition, genes responsible for denitrification were also actively expressed in the sludge. At the same site, metagenomics analysis confirmed the dominance of oxidising ammonia bacteria, such as *Nitrosomonas*, *Nitrospira*, and other non-ammonia oxidisers archaea (Yu and Zhang 2012). Metatranscriptomics analysis of wastewater effluent-based bioreactor revealed much about enzymes and microbial communities involved in nitrogen metabolism of wastewater. The analysis showed that the enzymes related to nitrification pathways (Ammonia mono-oxygenase, Hydroxylamine oxidase, and Hydroxylamine reductase) and denitrification pathways (nitric-oxide reductase, nitrous-oxide reductase, nitrate reductase and NO-forming nitrite reductase) were abundant in the waste water.

Moreover, high gene expression levels for enzymes related to energy production and growth were also active in the wastewater. The abundance and expression level of different enzymes involved in nitrification, denitrification, ammonification, and

nitrogen fixation was found to be affected by the environmental conditions of the bioreactor (Crovadore et al. 2017). Recent metatranscriptomics studies involving microbial communities involved in xenobiotic degradation, methane production, and bio-composting potential of microbial communities of waste have been listed in Table 14.1 under the meta-transcriptomics section (Ding et al. 2020; Braga et al. 2021; Kakuk et al. 2021; Russell et al. 2021).

### 14.4.3 Metaproteomics Technology for EFW Microbiome

The biological activities of any microorganism or in any biome are the results of enzymes present either inside the microbial cell or secreted outside. Identification of enzymes playing an active role in waste decomposition and converting complex biomolecules into simpler ones could be useful for EFW. Enzymes are proteins in nature and can be analysed using proteomics technology. Metagenomics approaches provide information about the composition, diversity, and dynamics of a microbial community, but remain silent about the presence of enzymes and overall metabolic activities of the microbial communities. For these two purposes, metatranscriptomics and metaproteomics technologies are used. Although metatranscriptomics provides important information about actively transcribing genes in a microbial community, it is also true that total mRNA is never translated into proteins. Metatranscriptomics also remain silent about post-translation modifications (PTMs) of the proteins. These bottlenecks of metagenomics and metatranscriptomics are overcome by metaproteomics technology, which gives accurate information about the presence of enzymes and other proteins, and PTMs-induced modification in proteins. Metaproteomics is defined as the identification of all the proteins being expressed within an ecosystem at a given time (Wilmes and Bond 2004). Proteomics technologies have evolved from dimensional Poly Acrylamide Gel Electrophoresis (PAGE) to two-dimensional PAGE, followed by the development of gel-free technologies such as shotgun proteomics analysis and Nano-LC-MS/MS. Metaproteomics approaches are used to study any biome's functional diversity. Metaproteomics studies give information about the substrate and enzymes at any given waste decomposition site. The study about enzymes and proteins present in any biome, like the waste biome, provides insight into waste degradation pathways.

A metaproteomics workflow is shown in Fig. 14.5, which includes the following basic steps: sample preparation (protein extraction and purification), protein separation, protein digestion, mass spectrometry of digested peptides, and bioinformatics-based identification of proteins. Sample preparation is the first and most critical step of metaproteomics studies. For comparative proteomics of two or more samples, the experimental designing and sampling must be performed in such a way that the effect of other factors is minimised. After sampling, total proteins are extracted from the samples. There are many protein extraction methods, and the selection depends upon the type of sample and biological samples in it. A review by Siggins et al. (2012) and Heyer et al. (2015) mentions the protein extraction method from diverse samples. A fundamental consideration in a protein extraction protocol is the location

**Table 14.1** Example of different OMICS studies in relation to waste microbiome

<i>Metagenomics studies</i>					
S. No.	Sample and sampling site	Microbes identified	Role of microbes	Technology used	Reference
1	Activated sludge of a municipal waste water treatment plant	Proteobacteria, Actinobacteria, Bacteroidetes, Firmicutes Ammonia oxidising bacteria such as <i>Nitrosomonas</i> sp. and <i>Nitrospira</i> sp. and other non-ammonia oxidisers archaea	Bacteria were dominating compared to eukaryotes. Lowest abundance of Archaea and viruses.	Illumina sequencing	Yu and Zhang (2012)
2	Improvement of biogas (methane) production from solid-state anaerobic digestion (SSAD) of oil palm biomass. Total solids (TS) contents, feedstock to inoculum (F:I) ratios and carbon to nitrogen (C:N) ratios.	<i>Ruminococcus</i> sp., <i>Thiomargarita</i> sp., <i>Clostridium</i> sp. and <i>Anaerobacter</i> sp., <i>Sporobacterium</i> sp., <i>Thiomargarita</i> sp., <i>Saccharofermentans</i> sp., <i>Oscillibacter</i> sp., and <i>Sporobacter</i> sp., <i>Anaerobacter</i> sp. Archeal community; <i>Methanoculleus</i> sp., <i>Desulfurococcus</i> sp., <i>Methanospirillum</i> sp., and <i>Methanosarcina</i> sp. and <i>Methanomassiliicoccus</i> sp.	F:I ratio of 2:1, C:N ratio of 30:1 and 16% TS were found better for enhanced biogas production	Polymerase chain reaction-denaturing gradient gel Electrophoresis (PCR-DGGE)	Suksong et al. (2016)
3	Wastewater effluent-based bioreactor	Burkholderia, three other bacterial genera, Proteus, Vibrio, and Curvibacter	Bacteria involved in Ammonia assimilation, nitrate/nitrite ammonification, and denitrification	Illumina sequencing.	Crovadore et al. (2017)

4	Industrial biogas reactor with food waste as feedstock, operated at 60 °C	<p>Nitrosomonas, Methylococcus, Brucellaor Lactobacillus</p> <p>Proteolytic bacterium <i>Coprothermobacter proteolyticus</i>, <i>Thermoanaerobacteriales</i>, <i>Thermacetogenium</i>, <i>Anaerobaculum</i>, <i>Synergistales</i> and multiple associated bacteria</p> <p><i>Dictyoglomi</i> and <i>Planctomycetes</i></p> <p><i>Atribacteria</i></p> <p>Cellulytic bacteria</p> <p><i>Synergistales</i> and <i>Synergistales</i></p> <p><i>Anaerobaculum spp.</i></p>	<p>Syntrophic oxidising</p> <p>Bacteria play important roles in anaerobic oxidation and methane production</p>	16s RNA Illumina sequencing	Hagen et al. (2017)
5	Biogas (Methane) production from anaerobic co-digestion of food waste (FW) and waste activated sludge (WAS) were investigated and effect of biological pretreatment on Microbiome composition was studied	<p>Bacteroidetes, Chloroflexi, Proteobacteria, Firmicutes</p> <p><i>Syntrophomonas</i> sp., <i>Proteinihilum</i> sp., <i>Bacteroides</i> sp., <i>Petrimonas</i> sp., <i>Methanosarcina</i> sp., <i>Methanobacterium</i> sp. and <i>Methanosarcina</i> sp.</p>	<p>Methane production increased 24.6% after biological pretreatment. Anaerobic co-pretreatment reduced the abundance of filamentous bacteria of genus <i>Levilinea</i></p>	Pyrosequencing	Zhang et al. (2017)
6	Active (operational) and closed (non-operational) municipal landfills of Malaysia	<p><math>9.16 \times 10^7</math> bacteria in closed landfill and <math>1.52 \times 10^7</math> bacteria in active landfills were observed.</p>		Illumina sequencing of 16S rRNA amplicon	Zainun Simarani (2018)

(continued)

Table 14.1 (continued)

		Proteobacteria Bacteroidetes, Acidobacteria, Firmicutes, Actinobacteria, Gemmatimonadales, Chloroflexi, Parvarchaeota, Verrucomicrobia, Tenericutes ( <i>in order of dominance</i> )			
7	Three landfill sites of India, situated near New Delhi, Chandigarh, and Himachal Pradesh	Proteobacteria, Chloroflexi, Firmicutes, Chloroflexi, Bacteroidetes		Illumina sequencing of 16S rRNA amplicon and V3–V4 regions	Thakur et al. (2020)
8	Anaerobic digestion of waste-activated sludge with help of hydrochar which increased the methane production rate	<i>Syntrophomonas</i> sp. FDU0164, <i>Methanosarcina</i> sp. FDU0106, <i>Firmicutes</i> sp. FDU0048, <i>Proteiniphilum</i> sp. FDU0082, and <i>Aminobacterium mobile</i> FDU0089	Degradation of organics, which could be related to the enhanced methane yield	Metagenomics, Nucleotides Sequencing by Illumina,	Shi et al. (2021)
9	Food waste-based bioreactors	<i>acs</i> , <i>metF</i> , <i>coaA</i> , <i>mer</i> , <i>mch</i> , and <i>ftr</i> genes were upregulated under high temperature conditions. And significant change in bacterial and archaeal communities was observed under different temperature conditions.	Effect of temperature on bacterial communities was studied	16s RNA Illumina sequencing	Li et al. (2022)
<i>Metatranscriptomics studies</i>					
<b>S. No.</b>	<b>Sample and sampling site</b>	<b>Gene/mRNA identified</b>	<b>Role of gene/mRNA</b>	<b>Technology used</b>	<b>Reference</b>
1					

	Activated sludge sample, collected from the aeration tank of a wastewater treatment plant at Stanley, Hong Kong	Verrucomicrobia, <i>Nitrospirae</i> sp., <i>Euryarchaeota</i> sp., Ammonia mono-oxygenase (amoA, amoB, amoC) and hydroxylamine	Enzymes involved in oxidation of organic material and nitrogenous waste	cDNA preparation and Illumina sequencing. Meta Genome Rapid Annotation using Subsystem Technology, v3.1 (MG-RAST)	Yu and Zhang (2012)
2	Wastewater effluent-based bioreactor	Enzymes related to nitrification pathways (Ammonia mono-oxygenase, Hydroxylamine oxidase, and Hydroxylamine reductase) and denitrification pathways (Nitric-oxide reductase, Nitrous-oxide reductase, Nitrate reductase and NO-forming Nitrite reductase) were high. Enzymes involved in energy production and growth were also upregulated	Enzymes involved in nitrification, denitrification, ammonification and nitrogen fixation were affected by the changes in physiochemical conditions of the bioreactor	cDNA preparation and Illumina sequencing. MG-RAST	Crovadore et al. (2017)
3	To assess composting potential of bacteria under bioreactor and traditional mode	<i>Psychrobacter</i> , <i>Galbibacter</i> , <i>Pseudomonas</i> , <i>Staphylococcus</i> and <i>Flavobacterium</i>	Composting process through microbial diversity	Illumina MiSeq sequencing of cDNA of metatranscriptomics	Ding et al. (2020)
4	Methane production	<i>Methanosarcina</i> , <i>Methanoculleus</i>	Microbial community in the CH <sub>4</sub> formation and CO <sub>2</sub> mitigation Power-to-Gas process	Metagenomic-assembled genomes (MAGs)	Kakuk et al. (2021)
5	Xenobiotic compounds such as pesticide degradation in soil	<i>Afpia</i> , <i>Sphingopyxis</i> , <i>Pseudomonas</i>	Enzymatic degradation via peroxidases, oxygenases, and hydroxylases	Metagenomic and metatranscriptomic sequencing	Russell et al. (2021)
6					

(continued)

Table 14.1 (continued)

	Assessment of variation in abundance of bacteria involved in the composting through hydrogenases and N <sub>2</sub> O reductase enzymes	<i>Rhodothermus marinus</i> , <i>Thermobispora bispora</i>	Enzymatic degradation via hydrogenases and N <sub>2</sub> O reductase	Metagenome-assembled genomes	Braga et al. (2021)
<i>Metaproteomics studies</i>					
<b>S. No.</b>	<b>Sample and sampling site</b>	<b>Microbes/proteins/enzymes identified</b>	<b>Role of proteins/enzymes</b>	<b>Technology used</b>	<b>Reference</b>
1	Degradation polychlorinated dioxins	<i>Spingomonas wittichii</i> RW1, catechol 1,2-dioxygenase, adenosylhomocysteinase	Dibenzofuran degradation pathway	Difference gel electrophoresis (DIGE) and matrix-assisted laser desorption/ionisation mass spectrometry (MALDI-MS)	Colquhoun et al. (2012)
2	Industrial biogas reactor with food waste as feedstock, operated at 60 °C	Enzymes involved in the Wood-Ljungdahl pathway and β-oxidation of fatty acids. Proteins of <i>Methanosaeita</i> bacteria, adapted for growing under high ammonia concentration	Syntrophic oxidising Bacteria play important roles in anaerobic oxidation and methane production	ID-SDS-PAGE based separation of intracellular and secreted proteins of bacteria, followed by Nano-LC-MS/MS based identification	Hagen et al. (2017)
3	Toxicity of silver nanoparticles	<i>Paracoccus denitrificans</i>	Transcriptional analysis indicated that Ag NPs restrained the expression of key genes related to denitrification	Analysing the transcriptional and proteomic responses of bacteria	Zheng et al. (2018)
4	Three landfill sites of India, situated near New Delhi, Chandigarh, and Himachal Pradesh	Enzymes responsible for dioxin degradation, styrene degradation, steroid degradation, streptomycin biosynthesis, carbapenem	Enzymes involved in degradation of xenobiotics	PICRUSt bioinformatics tool followed by KEGG annotations	Thakur et al. (2020)

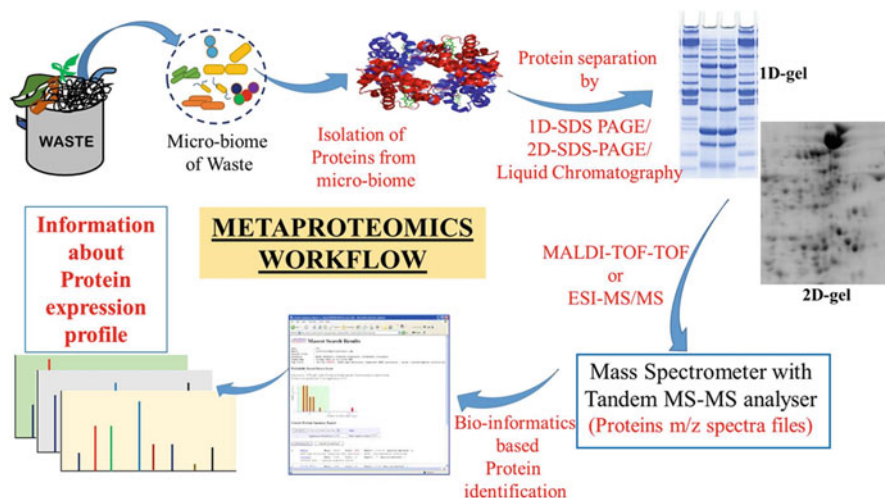
		biosynthesis, monobactam biosynthesis, furfural degradation pathways and plant cell wall degrading enzymes		
5	Antimony, Sb(III) resistance and transformation	<i>Acinetobacter johnsonii</i> JH7	Expression and activities of anti-stress enzymes were enhanced under Sb(III) stress	Reduction of phosphate-specific transporter could decrease Sb(V) uptake Gu et al. (2020)
6	Effect of ammonia toxicity on active methanogens	<i>Desulfo vibrio</i> , performs Biotransformations of propionate and butyrate to acetate and finally to the methane	Ammonia restrained the enzyme synthesis process by inhibiting the RNA polymerase (subunits A' and D) during transcription	Metagenomic and metaproteomic Liu et al. (2021)
7	(1) Environmental, (2) multidrug resistant (MDR) clinical and (3) susceptible clinical strains	<i>Pseudomonas aeruginosa</i> , Serine protein kinase and arginine/ornithine transport ATP-binding proteins	Upregulation of chitin binding and BON domain proteins	2-D DIGE and liquid chromatography tandem mass spectrometry quadrupole time-of-flight (LC-MS QTOF) Liew et al. (2021)
<i>Metabolomics studies</i>				
<b>S. No.</b>	<b>Sample and sampling site</b>	<b>Metabolites identified</b>	<b>Role of metabolites</b>	<b>Technology used</b>
1	Diverse role of <i>Pseudomonas</i> in environment	<i>Pseudomonas</i> . Poaeamide analogue and a molecular subfamily of cyclic lipopeptides, bananamides	Scavenge nutrients, sense population density and enhance or inhibit growth of competing microorganisms	Mass spectrometry-based molecular networking Nguyen et al. (2016)
2	Replacement of chemical fertilisation in banana cultivation in Colombia	<i>Bacillus amyloliquefaciens</i> and <i>Pseudomonas fluorescens</i>	Secondary metabolites and growth parameter of plants	SEM, FISH-CLSM Gamez et al. (2019)
3	Glyphosate-based herbicides sludge of municipal wastewater treatment plant	<i>Pseudomonas</i> sp., <i>Actinobacteria</i> and <i>Serratia</i> sp.	Esters, either those of phospholipids or	Fourier transform infrared (FTIR) spectroscopy analyses Grube et al. (2019)

(continued)



Table 14.1 (continued)

	(Daugavgriva, Riga, Latvia), agricultural soil and plant tissue	<i>Pseudomonas</i> sp. LAM1902	poly- $\beta$ -hydroxybutyrate indicates degradation		Weimer et al. (2020)
4	Mechanism of Nicosulfuron Biodegradation	<i>Pseudomonas aeruginosa</i>	Nicosulfuron was degraded by LAM1902 mainly via breaking the sulfonylurea bridge		Wang et al. (2020)
5	Analyses of genomic and metabolomic pathways of strains with antifungal properties	<i>Pseudomonas aeruginosa</i>	Phenazine-1,6-dicarboxylic acid (PDC) and pyocyanin (PYO)	Whole-genome resequencing of the two strains	Xu et al. (2021)
6	Atmospheric-pressure cold plasma can significantly change bacterial metabolites	<i>Pseudomonas aeruginosa</i>	Plasma-activated water treatment, the carbohydrate metabolism of the bacteria was inhibited and the metabolic processes of protein and amino acid decomposition were enhanced	Gas chromatography time-of-flight mass spectrometry (GC-TOF-MS) and Kyoto Encyclopedia of Genes and Genomes (KEGG) analysis	Karadzic et al. (2021)
7	Bioaccumulation and biomagnifications in environments	<i>Pseudomonas aeruginosa</i>	<i>P. aeruginosa</i> as biomarkers of presence of heavy metals and organic pollution	Comparative proteomics	



**Fig. 14.5** Different steps of a metaproteomics analysis

of proteins (extracellular or intracellular) and the selection of cell lysis methods in the case of intracellular proteins. The lysis method must be robust but not harsh to the proteins, and in the case of extracellular proteins, the cell lysis step is not required. Physical methods (ultra-sonication, homogenisation, grinding with liquid nitrogen, French press, freeze-thaw) and chemical methods (use of detergents) are used for cell lysis. The next step after cell lysis is the precipitation of proteins from the samples. Trichloro acetic acid (TCA), TCA-Acetone, and Ammonium sulphate are the common reagents used for protein precipitation. Precipitated proteins are separated from soluble impurities by centrifugation, followed by washing the protein pellet with acetone to remove the remaining impurities in the purified pellet of proteins. Either of Lowry's method estimates purified proteins, Modified Lowry's method, Bradford's method, and the Bi-cinchoninic acid method. The next step is the separation of proteins for mass-spectrometry-based identification. Due to complex nature of samples, a high number of proteins in the sample and the inability of a mass-spectrometer to identify more than 4–5 intense peaks in a sample, the proteins are separated before their processing for mass spectrometry analysis. PAGE-free liquid chromatography methods, PAGE-based methods: 'One-dimensional SDS PAGE' (1-D) (Kilambi et al. 2016) and 'Two-dimensional gel electrophoresis' (2-DE) (Maurya et al. 2014) can be used for protein separation. 2-DE has an inherent limitation in dealing with the proteins having a very low abundance, very high molecular weight and extreme (too basic or too acidic) pI (iso-electric point); hence 1-D methods are preferred. Liquid chromatography (LC) methods are also a method of choice in which proteins are digested inside the solution without separating them on 1-D or 2-DE gels, followed by their separation on a chromatographic column (Kilambi et al. 2016). The only problem with LC is the column clogging by impurities in the digested peptides samples. 1-D gel-based separation (MudPIT)

analysis separates the impurities by retaining them in the gel, thus preventing the chromatographic columns from clogging. Proteins are reduced with 'Dithiothreitol' and alkylated with 'Iodoacetamide' before digestion into peptides. Proteins are digested into peptides using trypsin (a serine protease). Although other enzymes are also used for the digestion of proteins, trypsin is most widely used and is the 'gold-standard'. Trypsin digestion of proteins can be performed on proteins separated in the form of sliced bands/spots of a PAGE gel (in-gel digestion) or protein solubilised in buffer (in-sol digestion), inside micro-centrifuge tubes. The digested peptides are de-salted using 'C-18 reverse phase resin', and the salt-free purified peptides are injected into a mass spectrometer for their identification. 'Matrix Assisted Laser Desorption Ionization' (MALDI) and 'Electro Spray Ionization' (ESI) are the two popular ionisation methods used in an MS for ionisation of the peptides for their identification. The ionised peptides are sorted based on their mass and charge ( $m/z$ ) ratio. Nowadays, in addition to molecular weight analysis of peptides, their de novo sequencing is also possible using tandem MS-MS, in which the first MS separation allows the selection of abundant peptides peaks and the second MS further fragments the selected peaks and provides the sequence of peptide in peak. The  $m/z$  data obtained from a tandem MS-MS is analysed using bioinformatics platforms. Mascot server from <https://www.matrixscience.com/> and 'ProteinProspector' (version 6.2.2) of <https://prospector.ucsf.edu/> is the platform which analyses  $m/z$  data files from an MS-MS system and identifies the proteins present in the sample based on genome database searching (Heyer et al. 2015, 2019; Yang et al. 2020).

Although metaproteomics is a robust and widely accepted technology for global-proteomic profiling of a sample, it has certain inherent limitations too. Due to pH limitations of gel-based methods, the proteins with extreme pI can't be separated on gels and hence become out of scope of gel-based methods. Although the gel-free method (Nano-LC-MS-MS) has overcome these limitations, the separation of peptides and clogging of LC columns remains challenging. In addition, special protocols are required for analysis of PTMs in the samples. Dependency on the gene-sequence database is the most significant limitation of proteomics technology, because if no gene sequence for a corresponding new protein is available, it will remain unidentified.

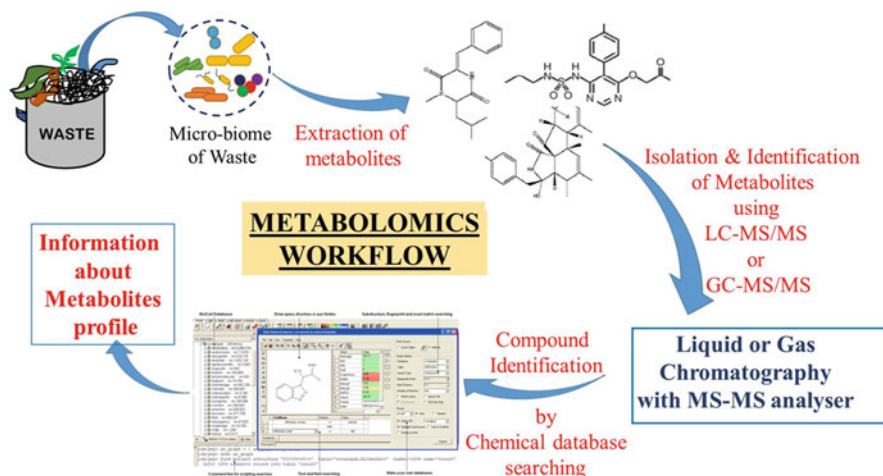
Metaproteomics analyses from different types of samples (human biology, soil, marine, and freshwater ecosystems, natural and bioengineered systems) have been reviewed by Siggins et al. (2012). Methods of protein sampling to MS-based protein identification from the different samples and their outcomes have been mentioned in the review work. In relation to EFW, the meta-proteomics analysis of an industrial food waste-based biogas reactor operating at thermophilic condition (Hagen et al. 2017) revealed multiple unculturable bacteria to syntrophically oxidise acetate and longer chain fatty acids to hydrogen and carbon dioxide that are subsequently converted to methane. However, the metaproteomic analysis of biogas plants remains constrained by sample complexity, impurities, and lack of protein identification technologies (Heyer et al. 2015). These studies highlight the importance of metaproteomics for efficient energy production by elucidating the critical steps in

converting waste to energy. Enzymes in the degradation of different organic compounds (furfurals, dioxin degradation, styrene, steroid degradation, and plant cell wall) were upregulated in three landfill sites in India (Thakur et al. 2020). Other studies involving metaproteomics analysis of methane-producing bacteria (Liu et al. 2021), antimony-resistant bacteria (Gu et al. 2020), and multidrug-resistant bacteria (Liew et al. 2021) are listed in Table 14.1.

#### 14.4.4 Metabolomics Technology for EFW Microbiome

In an ecosystem like EFW biome, the life forms, proteins, enzymes, and chemical molecules play a very crucial role in maintaining that ecosystem. The methods for analyses of all life forms, mRNA, and proteins have been discussed in the preceding sections. The chemical molecules (metabolites) produced and metabolised in any ecosystem are identified by metabolomics technology. Metabolomics is the technology for providing holistic information about the metabolites in any cell, tissues, biological fluid, or ecosystem. These metabolites tell about the chemical reactions, starting material, intermediates, and the end products of those reactions in an ecosystem. Metabolite profiling first appeared in the literature in the 1950s. Prof. Jeremy Nicholson proposed the concept of metabolomics, which offered the idea of analysing all metabolites of an ecosystem or sample. Metabolomics studies are generally focused on small molecules with a relative molecular weight of <1000 Da. Metabolomics gives information about the relationship between metabolites and the physiochemical conditions of an ecosystem; it also reflects the effect of changing an ecosystem's physical, chemical, or biological conditions (Yang et al. 2019). In addition, metabolites produced by plants (Meena et al. 2020; Negi and Maurya 2020), fungi (Gangwar et al. 2020), and microorganisms are responsible for their medicinal properties, too (Kumar et al. 2015a, 2019a; Bharati et al. 2020; Maurya et al. 2021; Yadav et al. 2021).

The workflow of metabolomics is shown in Fig. 14.6. Metabolomics encompasses two fundamental techniques of chemistry: (1) techniques used for the separation of chemical molecules: Liquid chromatography (LC) and Gas chromatography (GC); and (2) techniques used for the identification of chemical molecules: Mass spectroscopy (MS) and Nuclear magnetic resonance (NMR). Like other 'omics' technologies, sample preparation is a very critical step of metabolomics and requires a due care to avoid the degradation of chemical molecules in the sample. After sampling, the chemical molecules from a sample are separated using their inherent physiochemical properties. This step decreases the chemical complexity of the sample and makes the identification of chemicals in it more accessible. Liquid chromatographic methods use different types of stationary and mobile phase to separate metabolites by exploiting their physical and chemical properties. At the same time, gas chromatography separates only volatile molecules. After the chromatography-based separation of metabolites, mass spectrometry (MS) techniques are unquestionably used to identify various chemical molecules at a high-throughput scale. MS techniques can identify different chemical molecules



**Fig. 14.6** Different steps of a metabolomics analysis

present in a sample without separating them in pure form. NMR is used to determine the three-dimensional structure of any chemical molecule isolated from the sample, but essentially needs that molecule to be in pure form. The data obtained from MS or NMR is analysed using bioinformatics or chemo-informatics software to identify metabolites using various algorithms. Further, the metabolic pathways can be constructed using specialised bioinformatics software (Rochfort 2005; Yang et al. 2019).

Metabolomics has its unique advantage of dealing with chemical molecules only. Dependency on 'chemical molecules databases' for identifying metabolites separated by LC-MS/MS or GC-MS/MS is the main limitation of metabolomics analysis. The more the chemical database is updated, the better the metabolomics results will be.

There are many reports on the application of metabolomics for the analysis of waste ecosystems. Zhen et al. (2018) analysed water effluent plants' effect on drinking water quality by cell-based metabolomics. They found that the impact of the water effluent treatment plant on the studied site was not significant. However, analyses of hydrophilic and lipophilic metabolomes indicated a gradient of response intensities with a distance of the sampling sites from the wastewater treatment plant. Metabolomics was also used to discriminate the toxicity generated by pyrazinamide and its metabolic products (pyrazinoic acid and 5-hydroxy pyrazinoic acid) (Rawat et al. 2018). Guan et al. (2018) derived the relationship between pectinase activity and two metabolic pathways (fatty acid synthesis pathways and TCA) using two strains DY1 than DY2 of *Bacillus licheniformis*, for elucidating the metabolic mechanism of the fermentation process (Kashyap et al. 2019).

### 14.4.5 Need of Computational Algorithms for 'Omics' Analysis

All the 'Omics' approaches generate high-volume data, and in the case of metaproteomics and metabolomics, the data can be multidimensional too. Such high volume and multidimensional data need advanced computer algorithms for deriving complete gene sequence from contigs, sequence alignments, sequence annotation, and phylogenetic analysis for metagenomics and metatranscriptomics. Development in computer science has enabled a user with minimum computer hardware information but without knowledge of programming language and computer, algorithms to use the software packages used for 'Omics' data analysis comfortably (Chamrad et al. 2004; Veltri 2008). Surface Vector Machine (SVM), Decision tree (DT), Random forest (RF), Artificial neural networking (ANN), *k*-nearest neighbors (KNN), Correlation-based feature selection (CBFS), Logistic Regression (LR), Principal component analysis (PCA), Principal component extraction (PCE), minimum Redundancy Maximum Relevance (mRMR), Genetic Algorithms (GA), etc., are the few algorithms used for the processing of genome sequences via applying clustering, classification and feature selections techniques. Metaproteomics and metabolomics analysis require specialised algorithms for dealing with mass spectrometry data (Neeraj et al. 2020). Original data from a mass spectrometer needs pre-processing algorithms for binning, alignments, base line subtraction for original data to improve MS data analysis.

Further steps of mass spectrometry data analysis require algorithms for peak selection, fragmentation-based peak analysis, and identification of fragmented analyte in peak (peptide or metabolite) by database searching. In addition, a database for complex genomics, transcriptomics, proteomics, metabolomics, and pathways data analysis also requires dedicated algorithms. All the algorithms work together in a software package, which provides a graphic user interface for controlling different steps of experimentation and analysing the results of those analyses. However, significant advances in high-throughput measurement techniques, sophisticated in silico data processing/analysis tools, and the development of dedicated algorithms need to be addressed in future research (Kumaraswamy and Kashyap 2021).

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## 14.5 Non-omics Technologies for EFW Microbiome

Besides the 'omics' technologies discussed above, there are other classical technologies that can be used for study of EFW microbiome. Microscopy and culture-based technologies are used for identification of culturable microorganisms and their characteristics. DGGE (Denaturing Gradient Gel Electrophoresis) is used to study of DNA-based diversity of non-culturable microbes. Enzyme activity assay like ELISA (Enzyme Linked Immunosorbent Assay) or qualitative/quantitative tests for biochemicals, can be used for identification of enzymes and metabolites present at an EFW microbiome. Test for the urease enzyme proposed by Tabatabai (Tabatabai 1994) for assaying the enzyme in soil is one such example (Kumar et al. 2015a).

## 14.6 Conclusion and Future Outlook

Renewable energy sources provide an alternative option for fossil fuel energy with dual benefits of energy security and environmental safety. Energy from waste adds one more dimension to these benefits, in which the problem of waste is mitigated along with energy production. Continuous development, urbanisation, and consumerist lifestyle generate tonnes of waste worldwide. A significant portion of waste is biodegradable and combustible in nature, which is used for energy production. Production of biofuels and biogas from biodegradable waste is a sustainable method of energy production from waste, but is in initial phase of development. Conversion of waste into chemical forms feasible for EFW is a complex interplay of microorganism, proteins, and chemical entities present in waste, which is a natural and slow process. Use of 'Omics' technologies has enabled the identification of microorganisms, actively transcribing genes, proteins, enzymes, and metabolites involved in the process of energy production from waste. Although worldwide researches are going on EFW and have provided significant information, but still this knowledge is insufficient for adoption of EFW at commercial scale. Despite various 'Omics' researches, scientists are still unable to find the microorganisms or groups of microorganisms that can efficiently degrade all types of waste for EFW. These limitation of EFW technologies are due to two reasons, firstly due to the diversity of EFW, researches are limited and most of them focus on similar types of aspects. Secondly, the 'omics' technologies have their inherent limitations due to which a consolidated information and feasible methods for efficient EFW technologies are still lacking. Integration of 'omics' approaches (metagenomics, metatranscriptomics, metaproteomics, and metabolomics) is required for comprehensive analysis of waste microbiome. Also refining these tools and integrating them into intense research designs for deciphering MoW systems' biology remains to be accomplished yet. In future, more intense researches on EFW would produce more information, filling the knowledge gap in EFW, enabling the sustainable and useful treatment of wastes to generate energy.

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# Corrections to: Current Research Trends and Applications in Waste Management

Brijendra Kumar Kashyap and Manoj Kumar Solanki

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## Correction to:

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The original version of this book was inadvertently published with errors. The following errors have been corrected with this correction.

1. The Editor Brijendra Kumar Kashyap's affiliation has been updated to read as Department of Biotechnology Engineering, Institute of Engineering and Technology, Bundelkhand University, Jhansi, Uttar Pradesh, India
2. In Chapter 10, the author Sandeep Shukla's affiliation has been corrected to read as Department of Environmental Science, Gurugram University, Gurugram, Haryana, India

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The updated version of the book can be found at  
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