Nanomaterials for Biosensing Applications in the Medical Field



Rakesh K. Sahoo, Saroj Kumar Singh, Rajaram S. Mane, and Shikha Varma

Abstract The combination of nanotechnology and biotechnology has emerged as an integrated technology for medical applications. Over the world, day by day, numerous researchers are developing novel materials using the suitable platform to detect pathogenic, mutagenic, or toxic compounds or any biological effect. This chapter addresses the classification of biosensors, especially for medical applications based on the two most important parameters: bio-recognition element and signal transduction. Furthermore, several grooming biosensing technologies are also addressed. Subsequently, more emphasis has been added to nanomaterial classification employed in the biosensors based on their chemical contents and structural dimensions. Additionally, more insight into the current challenges in the application of nanomaterials in biosensors, especially for medical applications, has been demonstrated.

Keywords Biosensors · Biomedical detection · Nanomaterials · 2D materials · Carbon materials

1 Introduction

The alarming rise in several pandemic and epidemic diseases like severe acute respiratory syndrome-coronavirus-2 (SARS-CoV-2), black fungus, cancer, etc., has forced the researcher to think up more advanced biological detection and monitoring systems

R. K. Sahoo · S. Varma (🖂)

S. K. Singh CSIR—Institute of Minerals and Materials Technology, Bhubaneswar 751013, India

R. S. Mane

Institute of Physics, Sachivalaya Marg, Bhubaneswar, Odisha 751005, India e-mail: shikha@iopb.res.in

School of Physical Sciences, Swami Ramanand Teerth Marathwada University, Nanded, Maharashtra 431606, India

[©] The Author(s), under exclusive license to Springer Nature Singapore Pte Ltd. 2023 313 R. S. Mane et al. (eds.), *Nanomaterials for Sustainable Development*, https://doi.org/10.1007/978-981-99-1635-1_12

to detect carcinogenic, mutagenic, and toxic elements [1–4]. Although modern technologies and industrialization have simplified our lives to a new level, others left behind several environmental issues leading to serious health issues [5]. Thus, it is highly desirable to design and explore the challenges in developing advanced detection and monitoring systems, especially bio-detection and bio-monitoring systems, to better human health. The potential of biosensors in various functional fields is schematically presented (Fig. 1). The technology that highly depends on genetically modified organisms can be treated as biosensor technology which is emerging in advancement. The research in biosensor had drawn attention when Gary Sayler's group reported the report of the genetically modified microbial biosensor in the early 1990s [6]. According to van der Meer and Belkin biosensor [7], a device detects the chemical and biological changes in the system and transforms them into a measurable signal when the biological materials interact with this engineered device. Based on the type of biological materials interacting with the bio-reporter, the sensor is nomenclatured by different names. When the biological material is an antibody or whole-cell or nucleic acid, it is termed as an immunosensor or microbial biosensor, or DNA aptamer [8], respectively. Basically, there are four major components in the biosensor namely; (i) bio-receptor, (ii) transducer, (iii) a signal processing unit, and (iv) a display or interface unit that showcases the output signal (schematically shown in Fig. 2).



Fig. 1 Potential of biosensor in various fields of application in schematic form



Fig. 2 Schematic of the biosensor

More research in experimental and theoretical aspects is timely required to use the biosensors as the first filter for pre-screening the samples. The potential of synergetic research in engineering with biology has an enormous potential in designing biosensors for advanced applications. Thus, it is highly desirable to understand the fundamental changes in the biological sensing behavior of living beings. Some of the natural examples are (i) vibration, tactile, and airflow sensors in spiders, (ii) fast response of the plants toward the change in luminous intensity, osmotic pressure, temperature, water availability, etc., (iii) the snapping system in venus flytrap, (iv) dogs possess a sense of smell far beyond the sensing behavior of the artificial sensor. More significantly, dogs can detection system is so sensitive that it can detect the concentration of parts per billion, and (v) the system for controlled bending of trees, etc. In the above, all cases functional outputs are highly correlated to materials behavior with biological needs.

The biosensing platform is expected to be mechanically robust, versatile, and high throughput that will simplify the life in developing individual medicine, in vivodrug development, genomic-proteomic research, and point-of-care medical diagnosis [9]. Integrated technologies where nanotechnologies coupled with micro-fabrication technologies are able to develop new biosensors for medical applications [10]. However, the above type of advanced biosensor fabrication is in the embryonic stage and needs more research and development to enhance sensitivity, specificity, and high throughput. In this contest, designing the building blocks of the biosensor, i.e., the sensing materials in different scales and dimensions, has received considerable attention. Especially, nanomaterials in various dimensions and squeezing the atomic scale dimension have demonstrated fascinating bio-molecule detection behaviors. The work of Nam et al. [11] using nanoparticles and Liber et al. [12] using nanowires to design ultrasensitive biosensor is the pioneer in this area of research.

Several up-to-date sensor platforms are tested and proposed [13-16], especially for bio-molecule detection; additionally, few integrated technologies are in the next research phase before the medical diagnosis [10, 17-20]. In this chapter, the classification of biosensors based on bio-recognition elements and signal transduction has been described. The electrochemical, optical, thermal, and piezoelectrical sensors

based on the signal transduction perspective are proposed. Additionally, the enzymatic, protein receptor, immunosensors, DNA aptamer, and whole-cell biosensors based on bio-recognition elements are expressed. In the next section of the chapter, the nanomaterials of different dimensions and compositions applied in biosensor design have thoroughly been elucidated. Very concisely, nanomaterials, especially the two-dimensional materials used in designing flexible energy harvest and sensing for biomedical applications, are presented. Further, the current challenge and future prospective design of nanomaterials for biosensing, particularly for biomedical applications, are outlined.

2 **Biosensors for Medical Applications**

Technically, the entire class of biosensors has been classified based on two critical perspectives out of several, i.e., signal transduction and biorecognition element. In the subsequent section, the above two prospectives are briefly elaborated as follows.

2.1 Signal Transduction Perspective

Based on the signal transduction perspective, biosensors are categorized as electrochemical, thermal, optical, and piezoelectric sensors [3]. The electrochemical sensors are the most advanced and vastly investigated sensors for vivo monitoring or on-site monitoring. Low detection limit, high sensitivity, and generalizability are the advantages of this sensor compared to other category sensors. This category of the sensor is miniaturized to a lab-on-chip. Based on the signal from the sensor measured, it is subcategorized as amperometric (measuring the current produced during oxidation and reduction of the electroactive species), voltammetry (measuring the change in voltage of the working electrode concerning the reference electrode), and conductometry (measuring the alteration in conductance due to biochemical reaction). During the ongoing COVID-19 pandemic, electrochemical biosensors have been considered a crucial tool for rapid, accurate, and large-scale diagnosis of severing acute respiratory syndrome-coronavirus-2 (SARS-CoV-2) [2, 4, 21–23]. During the biochemical reaction, the absorbed or emitted photons are measured through an optical transducer. The optical phenomena studied to observe the alteration in biological responses include fluorescence, surface plasma resonance, and absorption. In parallel, the advancement of fiber optics technology has boosted optical sensor research to an extent level.

A thermal sensor, the most basic version, is a thermometer that is used to measure body temperature. However, the temperature range and toxicity of mercury limit its uses. Modern thermal sensors or enzyme thermistors are designed with a principal component called a sensitive thermistor based on similar working mechanisms. The function of the thermistor is to accurately estimate the change in enthalpy of the system during the biochemical reactions [24]. The piezoelectric sensor reciprocates the relationship between the resonant frequency change with respect to the mass of the molecule absorbed or desorbed on the crystal surface. The direct, label-free interaction with analyte mode is an efficient way of piezoelectric sensing platform. It is observed that the antibody or antigen is the best bio-molecule to be compatible with the piezoelectric sensor surface [25, 26].

2.2 Bio-recognition Perspective

Based on the bio-recognition perspective, the biosensors are categorized as enzymatic, protein receptor, immunosensor, DNA aptamers, and whole-cell biosensors. Each of the categories is elaborated as below.

a. Enzymatic biosensors

This type of sensor enzyme is the primary component that recognizes and reacts with the analyte to produce the electrochemical outcome. The brief sketch consists of analytes, receptors, an electrochemical transducer, and a signal amplifier. Here, the enzyme acts as a catalyst. And the function of the electrochemical transducer is to convert the chemical signal from the bio-reaction into a measurable physical signal which is further amplified by an amplifier. Most enzyme-catalyzed reactions release oxygen, carbon dioxide, and residual ionic species, measured by a transducer [27]. Two types of analytical enzymes such as hydrolases and oxidoreductases are used in the enzyme biosensor.

b. Protein receptor-based biosensors

The role of protein is opposite to enzyme as discussed previously in enzyme sensor. In this case, the protein present in the cell membrane acts as a receptor and reacts in a non-catalytic way with the signal from the transducer and produces the detectable signal by the process of metabotropic receptors through enzyme secretion or ionotropic receptors. Optical transduction has a significant role in this type of sensing platform [28, 29].

c. Immuno-sensors

This is a solid-state device wherein the immunochemical reaction is coupled to a transducer which is a basic design to detect the direct binding between antibodies to an analyte. Due to direct detection, faster and more cost-effective detection is possible using this type of sensor. A most exciting feature of immune-sensor is their selective and sensitivity in detecting multiple analytes by designing new recombinant antibodies [30].

d. DNA aptamers biosensor

Aptamers are short, single-standard DNA or RNA, and less than a hundred nucleotides are arranged/assembled in a specific sequence. This aptamer can interact

selectively with superior specificity and affinity forms bonding with a particular type of analyte, virus, bacteria, proteins, small molecules, toxins, hormones, etc., by hydrogen or Van der Waal binding force for biosensing. The beauty of these aptamers is that they can rearrange to form a variety of shapes and dimensions [31, 32]. Compared to immune sensor, DNA aptamer sensor is more specific, stable, and has a simple detection ability and also the cost is relatively lower. Due to its high stability, low cost, and superior specificity the DNA aptamer sensor is considered an alternative to antibodies.

e. Whole-cell biosensor

This type of sensor consists of two working components, i.e., the sensing element and reporter. The reporter element is a gene or gene cassette that has catalytic as well as non-catalytic functions. Catalytically, it accelerates the biochemical reaction to a detectable signal, and in a non-catalytic way as coding for the genes for metabotropic or ionotropic signal generation. The sensing element observes the gene or sets of gene's transcription initiation point similar to a promoter. The microbial sensor is the widely used whole-cell biosensor [33, 34]. Functional information rather than analytical information can be obtained using the whole-cell biosensor. The functional information can be obtained from the living cells by understanding the stimulus on a living system which can be applied in pharmacology, toxicology, cell biology, and many more. For example, the bacteria whole-cell biosensors can be genetically modified to sense mercury, nitrogen oxide, and hydroxylated polychlorinated biphenyls in urine and serum.

2.3 Limitations of Bio-based Biosensor

The major limitations that lag behind the bio-based biosensor are

- 1. First, low sensing performance with the low sensitivity and high limit of detection value of the designed sensor.
- 2. Due to limited active catalytic sites and surface area, bio-based biosensors have less chemical and catalytic activity.
- 3. The mechanical and cyclic performance stability and work life span of this type of sensor are very low.
- 4. Relatively low diffusivity of the bio-based biosensors. Especially in the electrochemical sensing case where the rate change of Faradic current is proportional to the diffusivity of the analyte on the electrode probe surface.

The above short-coming of the bio-based biosensors are tactically overcome by the use of engineered nanomaterials in the biosensor. Nanomaterials based biosensors are the rapidly growing research, especially for biosensor applications. Nanomaterials are basically used as transducer materials in biosensor development.

3 Nanomaterials in Biosensors

3.1 Metal Oxide Nanostructures

The beauty of the metal oxide nanostructures lies in their inherent functional biocompatibility, abundant active surface area in absorbing the bio-molecules, and high catalytic property in immobilizing the biomolecules in a non-toxic way to enhance electron-transfer kinetics for effective sensing characteristics. Various metal oxides from metals like Fe, Zn, Ce, Mg, Zn, Cu, Ti, and Zr are extensively explored in the literature for biosensor applications (shown in Fig. 3) [35–37]. These oxides of different morphologies and dimensions are synthesized using various synthesis methods like hydrothermal, sol–gel, radio frequency sputtering, soft chemistry, etc. [3].



Fig. 3 Typical metal oxide nanostructures and their biosensing characteristics. The abbreviations in this picture can be read like this IEP as iso-electric point; ChO_x as cholesterol oxidase; GO_x as glucose oxidase; HRP as horseradish peroxidase; IgG as immunoglobulin G; Urs, as urease (adopted from Solanki et al. [35])

3.2 Chalcogenide Nanostructures

In biosensors, to enhance the optical, opto-electrical, electrical, and magnetic properties of the semiconducting oxides are deliberately used with metal oxide as a hybrid structure. Additionally, several semiconducting sub-atomic scale particles demonstrate fascinating biosensing characteristics. The use of semiconductor and semiconductor chalcogenide nanostructures is reported in optical transduction. As reported in the literature, semiconducting quantum materials are deliberately used in biosensing applications due to their superior photo-stability, size-dependent photoemission, and broad absorption. However, the structural defects in fine quantum dots enhance the radiative recombination leading to inaccurate emission estimation.

Several soft techniques are adopted to overcome such defects and make the transducer more sensitive toward analyte detection and bio-immobilization. Those are as follows.

- (i) In the case of CdS, a layer of ZnS is coated on the surface to form a core–shell structure which acts as a photo-quencher: *Encapsulation*
- (ii) Functionalization of the quantum dots to enhance biomolecule immobilization and minimize the chance of toxicity with a broad idea not to hamper the photophysical recombination: *Ligand exchange*
- (iii) An extension of the previous step where the quantum dots are coated with silica to enhance the stability: *Silanization*.

This type of non-radiative or Fluro-quenched nanostructures is used as Förster/fluorescence Resonance Energy Transfer (FRET), especially for detecting optical DNA and oligonucleotides. Recently, two-dimensional nanomaterials and their derived quantum structures have demonstrated high potential donors in FRET-based sensing applications. These materials are graphitic carbon nitride $(g-C_3N_4)$ [38], perovskite materials [39], selenium [40], 2D metal–organic/covalent organic frameworks [41], and their derived 2D quantum structures [42, 43]. The details of the above materials are tabulated in Table 1.

Bioluminescence resonance energy transfer (BRET) is another type of biosensing technique where semiconducting quantum nanostructures are used as the acceptor. It is a distance-dependent non-radiative energy transfer from a bioluminescent donor to a fluorescent acceptor through resonance energy transfer. Using this technique, the blood glucose level can be estimated from teardrops. Bioluminescence donors are natural enzymes collected from marine animals. Certain donors have specific functions based on their structural arrangement. Some of the BRET donor–acceptor pairs reported in the literature are listed below in Table 2. Recently, several quantum dots are used as the acceptor in BRET sensors due to their distinct advantages. Mattoussi et al. [60] tunable emission from Ag: ZnInSe QDs can be obtained by varying the In/Zn feeding ratio. This Ag: ZnInSe QDs demonstrates robust behavior in terms of tuning the emission to align the protein emission in the BRET sensor for several cycles. Some of the reported functionalized semiconducting quantum dots used in BRET sensors include polymer-coated CdSe/ZnS core–shell nanostructure [61],

Target analyst	Donor: acceptor pairs	Dynamic range	Detection limit	References
Bilirubin	MoS ₂ QDs: bilirubin	0.5–10.0 μm	2.1 nm	[44]
MicroRNA	MoS ₂ QDs: FAM-MBs	5–150 nm	0.38 nm	[45]
EP	MoS ₂ QDs: P _{EP-PEI} copolymers	0.2–40 μm	0.05 μm	[46]
AA	MoS ₂ QDs: P _{EP-PEI} copolymers	0.5–40 μm	0.2 μm	[46]
6-MP	MoS ₂ QDs: DAP	0.5–70 μm	0.29 μm	[47]
BSA	MoS ₂ QDs: RGO	5–50 nm	Not mentioned	[48]
Dopamine	MoS ₂ QDs-aptamer: MoS ₂ nanosheets	0.1–1000 nm	45 pm	[49]
BSA	MoS ₂ QDs: polyaniline	10–70 nm	9.86 nm	[50]
GSH	MoS ₂ QDs: R6G	5–50 nm	2.7 nm	[51]
Nitrite	MoS ₂ QDs: BSA-Au NCs	0.5–20 mg/l	0.67 nm	[52]
NFZ	WS ₂ QDs: NFZ	0.17–166 μm	0.055 μm	[53]
DNA	BP QDs: Dabcyl-L probe	4–4000 pm	5.9 pm	[54]
GSH	g-C ₃ N ₄ : MnO ₂	NM	0.2 μm	[38]
H_2O_2	g-C ₃ N ₄ : MnO ₂	0–130 μm	1.5 μm	[55]
Glucose	g-C ₃ N ₄ : MnO ₂	0–150 μm	1.5 μm	[55]
Ricin	g-C ₃ N ₄ : MnO ₂	0.25–50 µg/ml	190 ng/ml	[56]
Riboflavi	g-C ₃ N ₄ : riboflavi	0.4–10 μm	170 nm	[57]
Metronidazole	g-C ₃ N ₄ : metronidazole	0.01–0.10 µg/ml	0.008 µg/ml	[58]
Dopamine	BSA-Au NCs/g-C ₃ N ₄ : dopamine	0.05–8.0 µm	0.018 µm	[59]
Hg(II)	Perovskite: RBED	20–90 µm	2.36 µm	[39]

 Table 1
 Summary of the FRET sensing applications of 2D nanomaterials as donors

semiconductor polymer nanoparticles with poly[2-methoxy-5-((2-ethylhexyl)oxy)p-phenylenevinylene] (MEH-PPV) [62], carboxylated quantum dots (Qd-625) [63], annexin V·RLuc-QDs [64], and glutathione-coated CdSeTe/CdS QDs [65].

	I I I I I I I I I I I I I I I I I I I	8 11	
Bioluminescent proteins	Emission (nm)	Substrate	References
Vargula luciferase (Vluc) or Cypridina luciferase	460	Vagulin (Cypridina luciferin)	[66]
Bacterial luciferase (Lux)	490	FMNH ₂ long-chain aliphatic aldehydehdacf	[67]
Gaussia luciferase (Gluc)	480	Coelenterazine	[68]
Metridia luciferase	480	Coelenterazine	[69]
Renillaluciferase (Rluc)	480	Coelenterazine	[70]
Aequorin	469	Coelenterazine	[71]
Firefly luciferase (Fluc)	562	D-luciferin	[72]
Nanoluciferase (Nluc)	460	Furimazine	[73]

Table 2 Summary of bioluminescent proteins used in BRET sensing application

3.3 Magnetic Nanoparticles

Intrinsic magnetic nanoparticles [74] and functionalized or coated nanoparticles have been applied in various biological applications like DNA [75] or cell separation [76], biological missiles [77], radio-immunoassay [78, 79], and in several varieties of biomolecule immobilization [80-87], especially for biosensor applications. Core-shell nanostructures of Fe_3O_4 @polydopamine [88], Ferrocene-modified Fe₃O₄@SiO₂ nanoparticles [89], Au@Ni [90], Ag NPs@Fe₃O₄ [91], Fe₃O₄/Au $@\gamma$ -Fe₂O₃/Au [92], etc., are designed for biosensor applications. Grancharov et al. [93] reported that the functionalized magnetic nanoparticles are used as biomolecular labels in magnetic tunnel junction-based biosensor. Chuang et al. [94] interpreted the time scale of Brownian relaxation of magnetic nanoparticles suspended in liquid obtained from the susceptibility variation as a function of frequency as a bio-magnetic target molecule sensor. Simultaneous detection of the magnetic fieldassisted DNA hybridization is sensed using a spin valve sensor reported by Graham et al. [95]. Liu et al. [96] fabricated a phenol biosensor where carbon paste is used as the supporting substrate for chemically immobilized and functionalized core-shell magnetic nanoparticles.

Research on magnetic nanoparticle-based biosensors is limited to lab-scale devices and medical diagnosis instruments in miniature form for bedside medical diagnosis. Several NPs are used in medical diagnosis devices; their sensitivity, the minimum sample volume, and the analyte that can be detected using these instruments are listed in Table 3 which is adapted from Koh et al. [97]. One pioneer example is the μ -NMR designed by Weissleder et al. [10] using 39 nm functionalized iron oxide nanoparticles in the microfluidic network. Further, the improved version of the microcoils is embedded in PDMS to increase the filling factor and decrease the signal-to-noise ratio. Also, this instrument can detect a minimal amount of sample, i.e., 1 μ l of the device [19, 20, 98].

	Analyte	Magnetic particle/instrumentation	Sensitivity	Sample volume (µL)	References
MRSw type I	Nucleotide	CLIO, benchtop relaxometer	Low nM-pM	300	[99]
	Proteins	CLIO, benchtop relaxometer	Low nM	300	[100]
	Virus	CLIO, MRI	50 virus/100μL	100	[101]
	Bacteria	Core/shell, DMR	20 CFUb/100 μL (membranefitered)	5	[20]
	Cancer cell	Mn-MNP, DMR	2 cells/1 μL	5	[19]
MRSw type II	Antibody	MP, bench top relaxometer	< 1 pM	300	[102]
AC susceptometer	Antibody	Iron oxide nanoparticles	< 1 nM		[103, 104]
SQUID	Bacteria	Iron oxide nanoparticles	1.1 × 105 bacteria/ 20 μL		[103, 104]
	DNA	Magnetic bead	3–10 pM (signal amplification)		[105]
GMR	Protein	Cubic FeCo NP	2×106 proteins	2	[106]
	DNA	Antiferromagnetic NP	10 pM		[107]
	Protein	Iron oxide NP	2.4 pM		[108]

 Table 3
 Magnetic nanoparticle used in different medical diagnosis instruments with their sensitivity [97]

3.4 Carbon Nanostructures

The beauty of carbon-based nanomaterials from its bulk count part is

It is easy to electrochemically recognize a specific type of biomolecule (such as ascorbic acid and uric acid.) mixed with carbon nanomaterials and quantify it which is impossible with glassy carbon electrodes. In potentiodynamic analysis, carbon nanotubes act as an ion–to–electron transducer for biosensing analysis.

- a. The outstanding electrical transport properties of carbon nanomaterials like carbon nanotubes and graphene. Intrinsic single-wall carbon nanotubes and graphene possess ballistic transport properties with high electron mobility which is necessary for high-speed biosensors.
- b. Using carbon nanomaterial in particular single or bilayer defect-free graphene which has high conductivity with low thermal noise and due to fewer defects, the pink noise (*i*/*f* noise) is also very low and can be effectively utilized in designing ultrasensitive biosensors.
- c. For flexible biosensor design, carbon-based nanomaterials are considered the best selection based on cost, stability, and performance.

d. Carbon nanodots/quantum particles are the best fluorescent centers for effective optical biosensor applications.

3.5 Hybrid Nanostructures

Hybrid nanomaterials are a promising platform for biosensor application, especially for the sensor in bio-medical diagnosis consisting of a unique conjugate of inorganic and organic components. The beauty of these hybrid nanomaterials lies in

- a. Fine inorganic nanoparticles (< 100 nm) have an enormous potential to be applied in electronics, catalysis, bio-medical, etc. However, for bio-medical applications, the inorganic particles must be bio-compatible and have colloidal stability in the aqueous environment without agglomeration and degradation. Thus, the organic material is widely hybridized with this inorganic particle to improve bio-compatibility, processability, and chemical stability.
- b. The organic/inorganic hybrids are mechanically robust and thermally more stable systems than individuals. Most importantly, the internal porosity of the hybrid can be tuned by anchoring the inorganic component which is highly desirable for ultrasensitive biosensor design and to increase the drug loading efficiency.
- c. The biological fluid when interacting with finer in-organic particles, the protein corona forms on the surface of the inorganic nanoparticles. The size and surface properties of the nanoparticles are highly dependent on the protein corona formation and cell-nanoparticle interaction. Additionally, the selection of organic components of the hybrid especially for biomedical application needs depth understanding of the protein corona formation and growth for effective biomedical application of the hybrid nanomaterial.

Based on recent literature, we are citing some of the recent works on the use of hybrid nanomaterials in biosensing applications. The list of carbon materials in hybrid form, reported in the literature in tabulated form (adapted from [3]) is cited in Table 4.

4 Challenges and Future Perspectives

Most importantly, the modern biosensor device can be miniaturized to a portable form for bedside clinical applications with effectively high throughput. Some of the new detection techniques that sound well from a scientific point of view and technological importance are grooming as next-generation electronic sensing chips such as field-effect electrolyte–insulator-semiconductor (FE–EIS) sensors and capacitive FE–EIS. Recently, the application of 2D materials like nanocarbon, metal dichalcogenides, hexagonal boron nitride, black phosphorous, and metal oxides has highly impacted the research in the FE–EIS-based sensors. However, there remain several challenges

Table 4 Represents the list of carbon	and carbon-based hybri	id nanomaterials employed in th	ne development of biosensors (A	dopted from Ref. [3])
Nanomaterial	Analyte	Transducer	Linear range	Detection limit	References
Ag@CQDs-rGO	Dopamine	Electrochemical	0.1–300 µm	0.59 nm	[109]
Ag NP-MWNT	Glucose	Electrochemical	0.025-1.0 mM	0.01 mM	[110]
Pd/Co-NCNT	Hydrazin	Electrochemical	0.05-406.045 µm	0.007 µm	[111]
Pd/CNF/[M3OA] ⁺ [NTF2] ⁻	H ₂		1.00–35.0 nM	0.33 nM	[112]
Cu NPs/Rutin/MWCNTs/IL/Chit/GCE	H ₂ O ₂	Cyclic voltammetry	0.35-2500 μM	0.11 µm	[113]
Cu/rGO-BP	Glucose	Electrochemical	0.1–2 mM	11 μm	[114]
Ni/Cu MOF	Glucose	FET	1 μM-20 mM	0.51 µM	[115]
NiO/PANINS	Glucose	Amperometric	1-3000 µM	0.06 µM	[116]
MnO-Mn ₃ O ₄ @rGO	H ₂ O ₂	Impedimetric	0.004–17 mM	0.1 μM	[117]
ZnO-rGO	Dopamine	Cyclic Voltammetric	0.1–1500 pM	$8.75\pm0.64~\text{pM}$	[118]
MoO ₃ @RGO	Breast cancer	Electrochemical	$0.001-500 \text{ ng mL}^{-1}$	0.001 ng mL^{-1}	[119]
Graphene QDs	Cu ²⁺	Electrochemical	0.015–8.775 μM	1.34 nM	[120]
Graphene QDs	Lung cancer ⁺	Fluorescence	0.1 pg mL^{-1} -1000 ng mL ⁻¹	0.09 pg mL^{-1}	[121]
CdTe/CdS//ZnS core/shell/shell QDs	l-ascorbic acid	Fluorescence	$8.0 \times 10^{-9} - 1.0 \times 10^{-7} \mathrm{M}$	$1.8 \times 10^{-9} \mathrm{M}$	[122]
NSET amptamer @Fe ₃ O ₄ @GOD and MoS ₂	Tumorcell(EpCAM)	Magnetic fluorescence	2–64 nM	1.19 nM	[123]
Au NPs@PDA@CuInZnS QDs	P53 gene	Electrochemiluminescenece	$0.1 - 15 \text{ nmol L}^{-1}$	0.03 nmol L^{-1}	[124]
CaM/SiNW-FETs	Protein	FET	10 ⁻⁸ -10 ⁻⁶ M	7 nM	[125]
G/Au NR/PT	HPV DNA	Electrochemical	1.0×10^{-13} -1.0 × 10 ⁻¹⁰ m L ⁻¹	$4.03 \times 10^{-14} \text{ m}$ L^{-1}	[126]
					(continued)

Nanomaterials for Biosensing Applications in the Medical Field

Table 4 (continued)					
Nanomaterial	Analyte	Transducer	Linear range	Detection limit	References
Graphene-Au NRs	NADHEthanol	Amperometric voltammetric	20–160 μM5–377 μM	6 μM1.5 μM	[127]
LAC-CNTs-SPCE	Para-cresol	Electrochemical	0.2–25 ppm	0.05 ppm	[127]
Co ₃ O ₄ -CNT/TiO ₂	Glucose	Photoelectrochemical	0-4 mM	$0.16 \mu M$	[128]
CNT thin-film transistor (TFT)	DNA	Thin film transistor (TFT)	1.6×10^{-4} -5 µmol L ⁻¹	$0.88 \mu g L^{-1}$	[129]
GQDs-MWCNTs	Dopamine	Electrochemical	$0.005 - 100.0 \ \mu M$	0.87 nM	[130]
CNT/Au NPs	Choline	Amperometric	0.05-0.8 mM	$15 \mu M$	[131]
PAMAM dendrimer	DENV 2E	Optical fiber	0.1 pM-1 μM	19.53 nm nM^{-1}	[132]
SAM/NH2rGO/PAMAM	DENV 2E	SPR	0.08 pM-0.5 pM	0.08 pM	[133]

326

in biosensor-based materials design, especially for medical applications which are as follows

- a. In enzyme-based biosensors, the presence of fouling agents and endogenous interfaces present in the sample has significantly hampered the sensor's sensitivity and specificity. Though this issue was partially addressed by making hybrid biomaterial, still the interface effect persists.
- b. Generally, doped semiconductor nanostructures have particular importance in biosensor design. However, the synthesis of doped semiconductor nanostructures is carried out in a harsh environment, and it isn't easy to achieve it on a large scale. Scale-up synthesis with high-quality control is highly desirable.
- c. Real-time in-vivo monitoring in complex media such as tissues and blood is still challenging. Moreover, it is highly desirable to establish a robust detection platform for in-vivo analysis, especially from a pharmacokinetic and pharmacodynamics point of view.
- d. Toxicity of the nanomaterials (carbon nanomaterials such as carbon whisker and carbon fiber.) in biosensors remains a significant challenge, especially for medical diagnosis.

5 Conclusions

This chapter comprehensively summarized the present scenario of nanomaterials in biosensors for medical applications. An attempt was made to summarize several chemical compositions and dimension nanomaterials applied in various biosensors in worldwide research. Additionally, the classification of biosensors based on the biorecognition and signal transduction mechanism was discussed. In recent decades, biosensors have demonstrated their potential to detect various quantitative and qualitative targets, especially for medical diagnosis. Due to the high stability and lower price, biosensors such as aptasensors and DNA-modified electrodes are being used as point-of-care devices for quick diagnosis of the SARS COVID-19 virus during ongoing pandemic emergencies across the globe. Modern biosensors have a vast perspective and high compatibility compared to conventional biosensors in medical applications due to their real-time diagnosis capability, high specificity, and sensitivity with minimal sample preparation.

References

- 1. https://www.who.int/news-room/fact-sheets/detail/cancer
- Yakoh A, Pimpitak U, Rengpipat S, Hirankarn N, Chailapakul O, Chaiyo S (2021) Paper-based electrochemical biosensor for diagnosing COVID-19: detection of SARS-CoV-2 antibodies and antigen. Biosens Bioelectron 176:112912
- Naresh V, Lee N (2021) A review on biosensors and recent development of nanostructured materials-enabled biosensors. Sensors 21(4):1109

- Raziq A, Kidakova A, Boroznjak R, Reut J, Öpik A, Syritski V (2021) Development of a portable MIP-based electrochemical sensor for detection of SARS-CoV-2 antigen. Biosens Bioelectron 178:113029
- 5. Manisalidis I, Stavropoulou E, Stavropoulos A, Bezirtzoglou E (2020) Environmental and health impacts of air pollution: a review. Front Public Health 8(14)
- Harms H, Wells MC, van der Meer JR (2006) Whole-cell living biosensors—are they ready for environmental application? Appl Microbiol Biotechnol 70(3):273–280
- van der Meer JR, Belkin S (2010) Where microbiology meets microengineering: design and applications of reporter bacteria. Nat Rev Microbiol 8(7):511–522
- Alhadrami HA (2018) Biosensors: classifications, medical applications, and future prospective. Biotechnol Appl Biochem 65(3):497–508
- Rocha-Santos TAP (2014) Sensors and biosensors based on magnetic nanoparticles. TrAC, Trends Anal Chem 62:28–36
- Lee H, Sun E, Ham D, Weissleder R (2008) Chip–NMR biosensor for detection and molecular analysis of cells. Nat Med 14(8):869–874
- Nam J-M, Thaxton CS, Mirkin CA (2003) Nanoparticle-based bio-bar codes for the ultrasensitive detection of proteins. Science 301(5641):1884–1886
- Patolsky F, Zheng G, Lieber CM (2006) Fabrication of silicon nanowire devices for ultrasensitive, label-free, real-time detection of biological and chemical species. Nat Protoc 1(4):1711–1724
- Hwang MT, Heiranian M, Kim Y, You S, Leem J, Taqieddin A et al (2020) Ultrasensitive detection of nucleic acids using deformed graphene channel field effect biosensors. Nat Commun 11(1):1543
- 14. Amoli V, Kim JS, Jee E, Chung YS, Kim SY, Koo J et al (2019) A bioinspired hydrogen bond-triggered ultrasensitive ionic mechanoreceptor skin. Nat Commun 10(1):4019
- Xue T, Liang W, Li Y, Sun Y, Xiang Y, Zhang Y et al (2019) Ultrasensitive detection of miRNA with an antimonene-based surface plasmon resonance sensor. Nat Commun 10(1):28
- Zhu Q-B, Li B, Yang D-D, Liu C, Feng S, Chen M-L et al (2021) A flexible ultrasensitive optoelectronic sensor array for neuromorphic vision systems. Nat Commun 12(1):1798
- Cai D, Ren L, Zhao H, Xu C, Zhang L, Yu Y et al (2010) A molecular-imprint nanosensor for ultrasensitive detection of proteins. Nat Nanotechnol 5(8):597–601
- McAlpine MC, Ahmad H, Wang D, Heath JR (2007) Highly ordered nanowire arrays on plastic substrates for ultrasensitive flexible chemical sensors. Nat Mater 6(5):379–384
- Lee H, Yoon T-J, Figueiredo J-L, Swirski FK, Weissleder R (2009) Rapid detection and profiling of cancer cells in fine-needle aspirates. Proc Natl Acad Sci 106(30):12459–12464
- Lee H, Yoon T-J, Weissleder R (2009) Ultrasensitive detection of bacteria using core-shell nanoparticles and an NMR-filter system. Angew Chem Int Ed 48(31):5657–5660
- Mahshid SS, Flynn SE, Mahshid S (2021) The potential application of electrochemical biosensors in the COVID-19 pandemic: a perspective on the rapid diagnostics of SARS-CoV-2. Biosens Bioelectron 176:112905
- Tran VV, Tran NHT, Hwang HS, Chang M (2021) Development strategies of conducting polymer-based electrochemical biosensors for virus biomarkers: potential for rapid COVID-19 detection. Biosens Bioelectron 182:113192
- Ranjan P, Singhal A, Yadav S, Kumar N, Murali S, Sanghi SK et al (2021) Rapid diagnosis of SARS-CoV-2 using potential point-of-care electrochemical immunosensor: toward the future prospects. Int Rev Immunol 40(1–2):126–142
- Ramanathan K, Danielsson B (2001) Principles and applications of thermal biosensors. Biosens Bioelectron 16(6):417–423
- Tombelli S (2012) 2—Piezoelectric biosensors for medical applications. In: Higson S (ed) Biosensors for medical applications. Woodhead Publishing, pp 41–64
- 26. Skládal P (2016) Piezoelectric biosensors. TrAC, Trends Anal Chem 79:127-133
- Rahimi P, Joseph Y (2019) Enzyme-based biosensors for choline analysis: a review. TrAC, Trends Anal Chem 110:367–374

- Hock B, Seifert M, Kramer K (2002) Engineering receptors and antibodies for biosensors. Biosens Bioelectron 17(3):239–249
- Culver HR, Wechsler ME, Peppas NA (2018) Label-free detection of tear biomarkers using hydrogel-coated gold nanoshells in a localized surface plasmon resonance-based biosensor. ACS Nano 12(9):9342–9354
- Honda N, Inaba M, Katagiri T, Shoji S, Sato H, Homma T et al (2005) High efficiency electrochemical immuno sensors using 3D comb electrodes. Biosens Bioelectron 20(11):2306–2309
- Ellington AD, Szostak JW (1990) In vitro selection of RNA molecules that bind specific ligands. Nature 346(6287):818–822
- 32. Chinnappan R, Eissa S, Alotaibi A, Siddiqua A, Alsager OA, Zourob M (2020) In vitro selection of DNA aptamers and their integration in a competitive voltammetric biosensor for azlocillin determination in waste water. Anal Chim Acta 1101:149–156
- Woo S-G, Moon S-J, Kim SK, Kim TH, Lim HS, Yeon G-H et al (2020) A designed whole-cell biosensor for live diagnosis of gut inflammation through nitrate sensing. Biosens Bioelectron 168:112523
- 34. Guo M, Wang J, Du R, Liu Y, Chi J, He X et al (2020) A test strip platform based on a whole-cell microbial biosensor for simultaneous on-site detection of total inorganic mercury pollutants in cosmetics without the need for predigestion. Biosens Bioelectron 150:111899
- Solanki PR, Kaushik A, Agrawal VV, Malhotra BD (2011) Nanostructured metal oxide-based biosensors. NPG Asia Mater 3(1):17–24
- Sahoo RK, Das A, Samantaray K, Singh SK, Mane RS, Shin H-C et al (2019) Electrochemical glucose sensing characteristics of two-dimensional faceted and non-faceted CuO nanoribbons. CrystEngComm 21(10):1607–1616
- Rahman MM, Ahammad AJS, Jin J-H, Ahn SJ, Lee J-J (2010) A Comprehensive review of glucose biosensors based on nanostructured metal-oxides. Sensors 10(5):4855–4886
- Zhang X-L, Zheng C, Guo S-S, Li J, Yang H-H, Chen G (2014) Turn-on fluorescence sensor for intracellular imaging of glutathione using g-C₃N₄ Nanosheet–MnO₂ sandwich nanocomposite. Anal Chem 86(7):3426–3434
- 39. Huang Y, Yan F, Xu J, Bian Y, Zhang R, Wang J et al (2017) The FRET performance and aggregation-induced emission of two-dimensional organic-inorganic perovskite, and its application to the determination of Hg(II). Microchim Acta 184(9):3513–3519
- 40. Xing C, Xie Z, Liang Z, Liang W, Fan T, Ponraj JS et al (2017) 2D nonlayered selenium nanosheets: facile synthesis, photoluminescence, and ultrafast photonics. Adv Opt Mater 5(24):1700884
- Jiang X, Zhang L, Liu S, Zhang Y, He Z, Li W et al (2018) Ultrathin metal-organic framework: an emerging broadband nonlinear optical material for ultrafast photonics. Adv Opt Mater 6(16):1800561
- 42. Barua S, Dutta HS, Gogoi S, Devi R, Khan R (2018) Nanostructured MoS₂-based advanced biosensors: a review. ACS Appl Nano Mater 1(1):2–25
- Zhou J, Chen J, Ge Y, Shao Y (2020) Two-dimensional nanomaterials for Förster resonance energy transfer–based sensing applications. Nanophotonics 9(7):1855–1875
- 44. Shanmugaraj K, John SA (2019) Water-soluble MoS2 quantum dots as effective fluorescence probe for the determination of bilirubin in human fluids. Spectrochim Acta Part A Mol Biomol Spectrosc 215:290–296
- 45. Yu X, Hu L, Zhang F, Wang M, Xia Z, Wei W (2018) MoS2 quantum dots modified with a labeled molecular beacon as a ratiometric fluorescent gene probe for FRET based detection and imaging of microRNA. Microchim Acta 185(4):239
- Zhang F, Wang M, Zeng D, Zhang H, Li Y, Su X (2019) A molybdenum disulfide quantum dotsbased ratiometric fluorescence strategy for sensitive detection of epinephrine and ascorbic acid. Anal Chim Acta 1089:123–130
- Zhang F, Liu H, Liu Q, Su X (2018) An enzymatic ratiometric fluorescence assay for 6mercaptopurine by using MoS₂ quantum dots. Microchim Acta 185(12):540

- Swaminathan H, Ramar V, Balasubramanian K (2017) Excited-state electron and energy transfer dynamics between 2D MoS₂ and GO/RGO for turn ON BSA/HSA sensing. J Phys Chem C 121(23):12585–12592
- 49. Chen J, Li Y, Huang Y, Zhang H, Chen X, Qiu H (2019) Fluorometric dopamine assay based on an energy transfer system composed of aptamer-functionalized MoS₂ quantum dots and MoS₂ nanosheets. Microchim Acta 186(2):58
- Swaminathan H, Balasubramanian K (2018) Förster resonance energy transfer between MoS₂ quantum dots and polyaniline for turn-on bovine serum albumin sensing. Sens Actuators, B Chem 264:337–343
- Balasubramanian K, Swaminathan H (2018) Highly sensitive sensing of glutathione based on Förster resonance energy transfer between MoS₂ donors and Rhodamine 6G acceptors and its insight. Sens Actuators, B Chem 259:980–989
- 52. Li W, Shi Y, Hu X, Li Z, Huang X, Holmes M et al (2019) Visual detection of nitrite in sausage based on a ratiometric fluorescent system. Food Control 106:106704
- 53. Guo X, Wang Y, Wu F, Ni Y, Kokot S (2015) The use of tungsten disulfide dots as highly selective, fluorescent probes for analysis of nitrofurazone. Talanta 144:1036–1043
- Yew YT, Sofer Z, Mayorga-Martinez CC, Pumera M (2017) Black phosphorus nanoparticles as a novel fluorescent sensing platform for nucleic acid detection. Mater Chem Front 1(6):1130– 1136
- Zhou Y-J, Li L, Wan Y-H, Chen T-T, Chu X (2018) 2D g-C₃N₄–MnO₂ nanocomposite for sensitive and rapid turn-on fluorescence detection of H₂O₂ and glucose. Anal Methods 10(42):5084–5090
- Men C, Li CH, Wei XM, Liu JJ, Liu YX, Huang CZ et al (2018) A sensitive and low background fluorescent sensing strategy based on g-C₃N₄–MnO₂ sandwich nanocomposite and liposome amplification for ricin detection. Analyst 143(23):5764–5770
- Han J, Zou HY, Gao MX, Huang CZ (2016) A graphitic carbon nitride based fluorescence resonance energy transfer detection of riboflavin. Talanta 148:279–284
- Hatamie A, Marahel F, Sharifat A (2018) Green synthesis of graphitic carbon nitride nanosheet (g-C₃N₄) and using it as a label-free fluorosensor for detection of metronidazole via quenching of the fluorescence. Talanta 176:518–525
- Guo X, Wu F, Ni Y, Kokot S (2016) Synthesizing a nano-composite of BSA-capped Au nanoclusters/graphitic carbon nitride nanosheets as a new fluorescent probe for dopamine detection. Anal Chim Acta 942:112–120
- Mattoussi H, Mauro JM, Goldman ER, Anderson GP, Sundar VC, Mikulec FV et al (2000) Self-assembly of CdSe–ZnS quantum dot bioconjugates using an engineered recombinant protein. J Am Chem Soc 122(49):12142–12150
- 61. So M-K, Xu C, Loening AM, Gambhir SS, Rao J (2006) Self-illuminating quantum dot conjugates for in vivo imaging. Nat Biotechnol 24(3):339–343
- 62. Xiong L, Shuhendler AJ, Rao J (2012) Self-luminescing BRET-FRET near-infrared dots for in vivo lymph-node mapping and tumour imaging. Nat Commun 3(1):1193
- Kumar M, Zhang D, Broyles D, Deo SK (2011) A rapid, sensitive, and selective bioluminescence resonance energy transfer (BRET)-based nucleic acid sensing system. Biosens Bioelectron 30(1):133–139
- Tsuboi S, Jin T (2017) Bioluminescence resonance energy transfer (BRET)-coupled Annexin V-functionalized quantum dots for near-infrared optical detection of apoptotic cells. Chem-BioChem 18(22):2231–2235
- Jin T, Yoshioka Y, Fujii F, Komai Y, Seki J, Seiyama A (2008) Gd3+-functionalized nearinfrared quantum dots for in vivo dual modal (fluorescence/magnetic resonance) imaging. Chem Commun 44:5764–5766
- 66. Thompson EM, Nagata S, Tsuji FI (1989) Cloning and expression of cDNA for the luciferase from the marine ostracod *Vargula hilgendorfii*. Proc Natl Acad Sci 86(17):6567–6571
- 67. Waidmann MS, Bleichrodt FS, Laslo T, Riedel CU (2011) *Bacterial luciferase* reporters: the Swiss army knife of molecular biology. Bioeng Bugs 2(1):8–16

- Tannous BA, Kim D-E, Fernandez JL, Weissleder R, Breakefield XO (2005) Codon-optimized Gaussia Luciferase cDNA for mammalian gene expression in culture and in vivo. Mol Ther 11(3):435–443
- 69. Stepanyuk GA, Xu H, Wu C-K, Markova SV, Lee J, Vysotski ES et al (2008) Expression, purification and characterization of the secreted luciferase of the copepod Metridia longa from Sf9 insect cells. Protein Expr Purif 61(2):142–148
- Paulmurugan R, Gambhir SS (2003) Monitoring protein-protein interactions using split synthetic renilla luciferase protein-fragment-assisted complementation. Anal Chem 75(7):1584–1589
- Mithöfer A, Mazars C (2002) Aequorin-based measurements of intracellular Ca²⁺-signatures in plant cells. Biol Proced Online. 4:105–118
- 72. Coleman SM, McGregor A (2015) A bright future for bioluminescent imaging in viral research. Futur Virol 10(2):169–183
- 73. Song G, Wu Q-P, Xu T, Liu Y-L, Xu Z-G, Zhang S-F et al (2015) Quick preparation of nanoluciferase-based tracers for novel bioluminescent receptor-binding assays of protein hormones: Using erythropoietin as a model. J Photochem Photobiol, B 153:311–316
- Haun JB, Yoon T-J, Lee H, Weissleder R (2010) Magnetic nanoparticle biosensors. WIREs Nanomed Nanobiotechnol 2(3):291–304
- 75. Gómez Pérez A, González-Martínez E, Díaz Águila CR, González-Martínez DA, González Ruiz G, García Artalejo A et al (2020) Chitosan-coated magnetic iron oxide nanoparticles for DNA and rhEGF separation. Colloids Surf A 591:124500
- Cui Y-R, Hong C, Zhou Y-L, Li Y, Gao X-M, Zhang X-X (2011) Synthesis of orientedly bioconjugated core/shell Fe₃O₄@Au magnetic nanoparticles for cell separation. Talanta 85(3):1246–1252
- 77. Ghanbari Adivi F, Hashemi P (2021) Removal of histamine from biological samples by functionalized Fe₃O₄@Agarose@Silica nanoparticles and its fast determination by ion mobility spectrometry. Coll Surf B 203:111717
- Zhao J, Zhu Z-Z, Huang X, Hu X, Chen H (2020) Magnetic gold nanocomposite and aptamer assisted triple recognition electrochemical immunoassay for determination of brain natriuretic peptide. Microchim Acta 187(4):231
- 79. Cao W, Liu B, Xia F, Duan M, Hong Y, Niu J et al (2020) MnO₂@Ce6-loaded mesenchymal stem cells as an "oxygen-laden guided-missile" for the enhanced photodynamic therapy on lung cancer. Nanoscale 12(5):3090–3102
- Bayramoglu G, Kayili HM, Oztekin M, Salih B, Arica MY (2020) Hydrophilic spacerarm containing magnetic nanoparticles for immobilization of proteinase K: employment for speciation of proteins for mass spectrometry-based analysis. Talanta 206:120218
- Suo H, Xu L, Xue Y, Qiu X, Huang H, Hu Y (2020) Ionic liquids-modified cellulose coated magnetic nanoparticles for enzyme immobilization: Improvement of catalytic performance. Carbohyd Polym 234:115914
- Vasić K, Knez Ž, Konstantinova EA, Kokorin AI, Gyergyek S, Leitgeb M (2020) Structural and magnetic characteristics of carboxymethyl dextran coated magnetic nanoparticles: from characterization to immobilization application. React Funct Polym 148:104481
- Darwesh OM, Ali SS, Matter IA, Elsamahy T, Mahmoud YA (2020) Chapter Twenty enzymes immobilization onto magnetic nanoparticles to improve industrial and environmental applications. In: Kumar CV (ed) Methods in enzymology, vol 630. Academic Press, pp 481– 502
- Fauser J, Savitskiy S, Fottner M, Trauschke V, Gulen B (2020) Sortase-mediated quantifiable enzyme immobilization on magnetic nanoparticles. Bioconjug Chem 31(8):1883–1892
- Esmi F, Nematian T, Salehi Z, Khodadadi AA, Dalai AK (2021) Amine and aldehyde functionalized mesoporous silica on magnetic nanoparticles for enhanced lipase immobilization, biodiesel production, and facile separation. Fuel 291:120126
- 86. Kharazmi S, Taheri-Kafrani A, Soozanipour A (2020) Efficient immobilization of pectinase on trichlorotriazine-functionalized polyethylene glycol-grafted magnetic nanoparticles: a stable and robust nanobiocatalyst for fruit juice clarification. Food Chem 325:126890

- Sahin S, Ozmen I (2020) Covalent immobilization of trypsin on polyvinyl alcohol-coated magnetic nanoparticles activated with glutaraldehyde. J Pharm Biomed Anal 184:113195
- Martín M, Salazar P, Villalonga R, Campuzano S, Pingarrón JM, González-Mora JL (2014) Preparation of core–shell F_e3O₄@poly(dopamine) magnetic nanoparticles for biosensor construction. J Mater Chem B 2(6):739–746
- Qiu J, Peng H, Liang R (2007) Ferrocene-modified Fe₃O₄@SiO₂ magnetic nanoparticles as building blocks for construction of reagentless enzyme-based biosensors. Electrochem Commun 9(11):2734–2738
- Gao X, Du X, Liu D, Gao H, Wang P, Yang J (2020) Core-shell gold-nickel nanostructures as highly selective and stable nonenzymatic glucose sensor for fermentation process. Sci Rep 10(1):1365
- Mazhani M, Alula MT, Murape D (2020) Development of a cysteine sensor based on the peroxidase-like activity of AgNPs@Fe₃O₄ core-shell nanostructures. Anal Chim Acta 1107:193–202
- 92. Mahmoudi-Badiki T, Alipour E, Hamishehkar H, Golabi SM (2017) A performance evaluation of Fe₃O₄/Au and γ-Fe₂O₃/Au core/shell magnetic nanoparticles in an electrochemical DNA bioassay. J Electroanal Chem 788:210–216
- 93. Grancharov SG, Zeng H, Sun S, Wang SX, O'Brien S, Murray CB et al (2005) Biofunctionalization of monodisperse magnetic nanoparticles and their use as biomolecular labels in a magnetic tunnel junction based sensor. J Phys Chem B 109(26):13030–13035
- Chunga SH, Hoffmann A, Bader SD, Liu CBK, Makowski L, Chen L (2004) Biological sensors based on Brownian relaxation of magnetic nanoparticles. Appl Phys Lett 85(4):2971
- Graham DL, Ferreira HA, Feliciano N, Freitas PP, Clarke LA, Amaral MD (2005) Magnetic field-assisted DNA hybridisation and simultaneous detection using micron-sized spin-valve sensors and magnetic nanoparticles. Sens Actuators, B Chem 107(2):936–944
- Liu Z, Liu Y, Yang H, Yang Y, Shen G, Yu R (2005) A phenol biosensor based on immobilizing tyrosinase to modified core-shell magnetic nanoparticles supported at a carbon paste electrode. Anal Chim Acta 533(1):3–9
- 97. Koh I, Josephson L (2009) Magnetic nanoparticle sensors. Sensors 9(10):8130-8145
- Grimm J, Perez JM, Josephson L, Weissleder R (2004) Novel nanosensors for rapid analysis of telomerase activity. Can Res 64(2):639–643
- 99. Perez JM, Josephson L, O'Loughlin T, Högemann D, Weissleder R (2002) Magnetic relaxation switches capable of sensing molecular interactions. Nat Biotechnol 20(8):816–820
- Kim GY, Josephson L, Langer R, Cima MJ (2007) Magnetic relaxation switch detection of human chorionic gonadotrophin. Bioconjug Chem 18(6):2024–2028
- 101. Perez JM, Simeone FJ, Saeki Y, Josephson L, Weissleder R (2003) Viral-induced selfassembly of magnetic nanoparticles allows the detection of viral particles in biological media. J Am Chem Soc 125(34):10192–10193
- Koh I, Hong R, Weissleder R, Josephson L (2008) Sensitive NMR sensors detect antibodies to influenza. Angew Chem Int Ed 47(22):4119–4121
- 103. Fornara A, Johansson P, Petersson K, Gustafsson S, Qin J, Olsson E et al (2008) Tailored magnetic nanoparticles for direct and sensitive detection of biomolecules in biological samples. Nano Lett 8(10):3423–3428
- 104. Grossman HL, Myers WR, Vreeland VJ, Bruehl R, Alper MD, Bertozzi CR et al (2004) Detection of bacteria in suspension by using a superconducting quantum interference device. Proc Natl Acad Sci 101(1):129–134
- 105. Strömberg M, Göransson J, Gunnarsson K, Nilsson M, Svedlindh P, Strømme M (2008) Sensitive molecular diagnostics using volume-amplified magnetic nanobeads. Nano Lett 8(3):816–821
- 106. Srinivasan B, Li Y, Jing Y, Xu Y, Yao X, Xing C et al (2009) A detection system based on giant magnetoresistive sensors and high-moment magnetic nanoparticles demonstrates zeptomole sensitivity: potential for personalized medicine. Angew Chem Int Ed 48(15):2764–2767
- 107. Fu A, Hu W, Xu L, Wilson RJ, Yu H, Osterfeld SJ et al (2009) Protein-functionalized synthetic antiferromagnetic nanoparticles for biomolecule detection and magnetic manipulation. Angew Chem Int Ed 48(9):1620–1624

- Osterfeld SJ, Yu H, Gaster RS, Caramuta S, Xu L, Han S-J et al (2008) Multiplex protein assays based on real-time magnetic nanotag sensing. Proc Natl Acad Sci 105(52):20637–20640
- 109. Han G, Cai J, Liu C, Ren J, Wang X, Yang J et al (2021) Highly sensitive electrochemical sensor based on xylan-based Ag@CQDs-rGO nanocomposite for dopamine detection. Appl Surf Sci 541:148566
- 110. Chen L, Xie H, Li J (2012) Electrochemical glucose biosensor based on silver nanoparticles/multiwalled carbon nanotubes modified electrode. J Solid State Electrochem 16(10):3323–3329
- 111. Zhang Y, Huang B, Ye J, Ye J (2017) A sensitive and selective amperometric hydrazine sensor based on palladium nanoparticles loaded on cobalt-wrapped nitrogen-doped carbon nanotubes. J Electroanal Chem 801:215–223
- 112. Afzali M, Mostafavi A, Nekooie R, Jahromi Z (2019) A novel voltammetric sensor based on palladium nanoparticles/carbon nanofibers/ionic liquid modified carbon paste electrode for sensitive determination of anti-cancer drug pemetrexed. J Mol Liq 282:456–465
- Roushani M, Dizajdizi BZ (2015) Development of nonenzymatic hydrogen peroxide sensor based on catalytic properties of copper nanoparticles/Rutin/MWCNTs/IL/Chit. Catal Commun 69:133–137
- 114. Zhu T, Wang X, Chang W, Zhang Y, Maruyama T, Luo L et al (2021) Green fabrication of Cu/rGO decorated SWCNT buckypaper as a flexible electrode for glucose detection. Mater Sci Eng, C 120:111757
- 115. Wang B, Luo Y, Gao L, Liu B, Duan G (2021) High-performance field-effect transistor glucose biosensors based on bimetallic Ni/Cu metal-organic frameworks. Biosens Bioelectron 171:112736
- 116. Kailasa S, Rani BG, Bhargava Reddy MS, Jayarambabu N, Munindra P, Sharma S et al (2020) NiO nanoparticles—decorated conductive polyaniline nanosheets for amperometric glucose biosensor. Mater Chem Phys 242:122524
- 117. Li Y, Tang L, Deng D, He H, Yan X, Wang J et al (2021) Hetero-structured MnO-Mn₃O₄@rGO composites: synthesis and nonenzymatic detection of H₂O₂. Mater Sci Eng, C 118:111443
- 118. Verma S, Arya P, Singh A, Kaswan J, Shukla A, Kushwaha HR et al (2020) ZnO-rGO nanocomposite based bioelectrode for sensitive and ultrafast detection of dopamine in human serum. Biosens Bioelectron 165:112347
- Augustine S, Kumar P, Malhotra BD (2019) Amine-functionalized MoO3@RGO nanohybridbased biosensor for breast cancer detection. ACS Appl Bio Mater 2(12):5366–5378
- 120. Wang Y, Zhao S, Li M, Li W, Zhao Y, Qi J et al (2017) Graphene quantum dots decorated graphene as an enhanced sensing platform for sensitive and selective detection of copper(II). J Electroanal Chem 797:113–120
- 121. Kalkal A, Pradhan R, Kadian S, Manik G, Packirisamy G (2020) Biofunctionalized graphene quantum dots based fluorescent biosensor toward efficient detection of small cell lung cancer. ACS Appl Bio Mater 3(8):4922–4932
- 122. Huang S, Zhu F, Xiao Q, Su W, Sheng J, Huang C et al (2014) A CdTe/CdS/ZnS core/shell/shell QDs-based "OFF–ON" fluorescent biosensor for sensitive and specific determination of 1ascorbic acid. RSC Adv 4(87):46751–46761
- 123. Cui F, Ji J, Sun J, Wang J, Wang H, Zhang Y et al (2019) A novel magnetic fluorescent biosensor based on graphene quantum dots for rapid, efficient, and sensitive separation and detection of circulating tumor cells. Anal Bioanal Chem 411(5):985–995
- 124. Liu Y, Chen X, Ma Q (2018) A novel amplified electrochemiluminescence biosensor based on Au NPs@PDA@CuInZnS QDs nanocomposites for ultrasensitive detection of p53 gene. Biosens Bioelectron 117:240–245
- 125. Lin T-W, Hsieh P-J, Lin C-L, Fang Y-Y, Yang J-X, Tsai C-C et al (2010) Label-free detection of protein-protein interactions using a calmodulin-modified nanowire transistor. Proc Natl Acad Sci 107(3):1047–1052
- 126. Huang H, Bai W, Dong C, Guo R, Liu Z (2015) An ultrasensitive electrochemical DNA biosensor based on graphene/Au nanorod/polythionine for human papillomavirus DNA detection. Biosens Bioelectron 68:442–446

- 127. Li L, Lu H, Deng L (2013) A sensitive NADH and ethanol biosensor based on graphene–Au nanorods nanocomposites. Talanta 113:1–6
- 128. Zhao K, Veksha A, Ge L, Lisak G (2021) Near real-time analysis of para-cresol in wastewater with a laccase-carbon nanotube-based biosensor. Chemosphere 269:128699
- 129. Li W, Gao Y, Zhang J, Wang X, Yin F, Li Z et al (2020) Universal DNA detection realized by peptide based carbon nanotube biosensors. Nanoscale Adv 2(2):717–723
- Huang Q, Lin X, Tong L, Tong Q-X (2020) Graphene quantum dots/multiwalled carbon nanotubes composite-based electrochemical sensor for detecting dopamine release from living cells. ACS Sustain Chem Eng 8(3):1644–1650
- 131. Wu B, Ou Z, Ju X, Hou S (2011) Carbon nanotubes/gold nanoparticles composite film for the construction of a novel amperometric choline biosensor. J Nanomater 2011:464919
- 132. Mustapha Kamil Y, Al-Rekabi SH, Yaacob MH, Syahir A, Chee HY, Mahdi MA et al (2019) Detection of dengue using PAMAM dendrimer integrated tapered optical fiber sensor. Sci Rep 9(1):13483
- 133. Omar NAS, Fen YW, Abdullah J, Mustapha Kamil Y, Daniyal WMEMM, Sadrolhosseini AR et al (2020) Sensitive detection of dengue virus type 2 E-proteins signals using selfassembled monolayers/reduced graphene oxide-PAMAM dendrimer thin film-SPR optical sensor. Sci Rep 10(1):2374