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# Nanotechnology: Its Usages in Drug Delivery for the Treatment of Human Parasitic Diseases

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#### Abstract

The occurrence of infectious parasitic disease is the leading driver of mortality worldwide. Treatment of such parasitic disease is challenging due to the minimal target bioavailability of antiparasitic drugs, poor cellular uptake, nonspecific distribution at the target site and rapid elimination from the body. Further antiparasitic drug toxicity and prolonged therapeutic regimens also concerns us. Leading trends in nanotechnology can overpower these shortcomings in the form of an ideal nanocarrier system that can be designed and fabricated accordingly, where new formulations and the existing antiparasitic drugs in nano-sized delivery vehicle can be more promising in terms of minimized non-specific drug accumulation, desired antiparasitic drug availability at the site of action, reduced therapeutic dose and duration that is to be delivered etc. Through this chapter, we have highlighted the major challenges of conventional treatment approaches and presented nanotechnology as an imminent alternative treatment approach for the infectious parasitic disease. However, the unification of these two-research areas as "nano-antiparasitic medicines" can progress as a therapeutic strategic plan, minimising the burden of individuals suffering from this worldwide.

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#### Keywords

Nano carrier · Antiparasitic drugs · Nano-antiparasitic medicines · Therapeutic regimens

#### **Abbreviations**



# 9.1 Introduction

Infectious parasitic diseases are of major concern to public health worldwide. Their occurrence and pathological prevalence are closely related to geographic and socioeconomic factors (Cable et al. [2017](#page-12-0)). However, in India, their pervasiveness is determined by climate change, as such changes turn on favorable condition for the spread of vectorborne infectious parasitic diseases, it accounts for 17% of all infectious diseases. The most common vector-borne infectious parasitic diseases are visceral leishmaniasis, malaria, dengue, chikungunya, Japanese encephalitis, lymphatic filariasis etc. (Leal Filho et al. [2022\)](#page-13-0). Further, the causative agent (pathogen) for infectious parasitic diseases is usually known as parasites. These are broadly been classified as eukaryotic organisms ranging from single cellular protozoans, to large multicellular helminths responsible for severe disease onset in both animals and the human population. The various class of parasites that cause disease in the human population are protozoans, helminths, ectoparasites etc. Amongst protozoans there is subdivision based on their mode of movement this includes Sarcodina (Entamoeba); Mastigophora (Giardia and Leishmania); Ciliophora (Balantidium); Sporozoa (Plasmodium, Cryptosporidium). Similarly, sub division inhelminths is based on their shape this includes platyhelminths (Trematodes (flukes) and Cestodes (tapeworms)); acanthocephalins (worms reside in the gastrointestinal tract); nematodes (worms reside in the gastrointestinal tract, blood, lymphatic system or subcutaneous tissues). Under protozoan parasites severe health illness in human is



majorly caused by genus plasmodium, entamoeba, acanthamoeba, leishmania, trypanosoma and toxoplasma (David Sibley [2011\)](#page-13-0). While for helminths the human health complications are mainly related to genus of *ascaris*, *schistosoma* and tenia (Jiménez et al. [2016](#page-13-0)). For host-parasite interaction (Fig. 9.1).

The conventional treatment approach for controlling infectious parasitic diseases is dependent on the currently available antiparasitic drugs. Major issues with the conventional treatment approach are due to its insoluble nature, shorter half-life, and minimized bioavailability of antiparasitic drugs to the target site. However, for enhanced effective therapeutic response frequent long-term booster dosage is required based on parasitic life cycles. Such treatment repetitions might introduce deleterious consequences such as stress, drug resistance, etc. (Vercruysse et al. [2007\)](#page-14-0).

To overcome such limitations there is a prerequisite need for novel therapeutic approaches in the form of nanotechnology, integration of nanotechnology with parasitic disease management can design and fabricate nanomedicines with nanoparticles ranging from 1 to 1000 nm. It can have a substantial impact on parasitic diseases and its presence can aim for enhancing the efficacy of antiparasitic drugs at the target site. There are various types of nanomaterials such as organic nanocarriers that are made from desired synthetic or natural polymers, cholesterol, phospholipids, solid lipids etc. They can be designed in the form of nanospheres, nanoparticles, micelles etc. Other than this inorganic nanocarriers like metallic and non-metallic nanoparticles are also used (Sun et al. [2019\)](#page-14-0). The loading of antiparasitic drugs into the nanocarrier system is a physical or chemical reactive event which occurs through adsorption, encapsulation and conjugation process. Further, its release at the target site can be a sequential event that might occur through desorption, dissolution or degradation of antiparasitic drug from nanocarrier system. These nanocarriers can easily infiltrate into the biological system where it can shield the antiparasitic drug from enzymatic degradation causing sustainable, controlled release and accumulation of antiparasitic drugs at target site etc. (Negi

et al. [2013;](#page-14-0) Das and Chaudhury [2011;](#page-13-0) Chen et al. [2015\)](#page-13-0). However, effectiveness of therapeutic approach dose not completely depend on type of nanocarrier system and the properties of drug it also depends on the route of administration etc. (Chen et al. [2015,](#page-13-0) [2017\)](#page-13-0). At present, nanoparticles that have been explored so far exhibit the forthcoming potential for development of "nano-antiparasitic medicines" further it also highlights its other broad developmental aspects of antiparasitic drug delivery application.

# 9.2 Nanoparticles Physiochemical Characteristics and Its Effect on Activity of Antiparasitic Drugs

The physico-chemical characteristics of nanoparticles has an effect on the activity of antiparasitic drugs. Enhanced efficacy of antiparasitic drugs is also dependent on the size of nanoparticle as it plays crucial role in transportation of antiparasitic drugs and distribution in an in vivo system. Thus, can be assumed to have diverse inhibitory effect on parasites. Work led by Liu and colleagues have determined that variation in the size of radiolabelled liposomes will alter the distribution in an in vivo mouse model, where in the bloodpost 4 h treatment 60% of liposomes in the size range of 100–200 nm were identified and small size nanoparticles were easily eliminated through excretion. Antileishmanial effects of gold nanoparticles of smaller size was also reported in an in vitro system (Liu et al. [1992;](#page-13-0) Want et al. [2021\)](#page-14-0). Hence, selection of an appropriate size can be promising for longer retention and sustained targeted distribution of antiparasitic drugs. Therefore, optimization of the nanoparticles based on the shape and size can enhance the cellular entrance ability thus, can be promising for treating intracellular parasitic infections. Similarly, surface charges can also have an effect on the activity of antiparasitic drugs. Surface hydrophilicity or hydrophobicity can influence the kinetics in an in vivo system, its impact on protein binding extents will enable easier attaining of the estimated distribution and kinetics, modification with a desired polymer like polyethylene glycol (PEG) will increase nanoparticles surface hydrophilicity. It can significantly prolong the residence time, half-life and bioavailability of antiparasitic-loaded nanoparticles (Kumar et al. [2017](#page-13-0); Pensel et al. [2015;](#page-14-0) Fülöp et al. [2018\)](#page-13-0). Therefore, such modification of nanoparticles can be an important parameter for satisfactory sustained-release antiparasitic drugs (Fig. [9.2\)](#page-4-0).

## 9.3 Nano-Assisted Therapeutic Regime for Parasitic Disease

Current advancements and innovations in the nanotechnology field have been extensively studied in parasitic diseases, to overcome the several frailties associated with conventional diagnosis and therapeutics. As such, anti-parasitic drug delivery systems gained attention to ameliorate their bioavailability, controlled release and intracellular penetration activity. Therefore, drug-loaded nanoparticles hold significant promise to enhance efficacy and reduce the dose and side effects of drugs. This

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Fig. 9.2 Schematic representation of the effect of nanoparticles physiochemical properties on the activity of anti-parasitic drugs

section focuses on a variety of nanoparticles enlisted in Table [9.1](#page-5-0) used in drug delivery systems and therapeutics diagnostics include liposomes, polymeric nanoparticles, SLNs, nanosuspensions, metallic nanoparticles and others (Fig. [9.3\)](#page-9-0).

#### 9.3.1 Liposomes

Liposomes are small artificial vesicle consisting of two or more lipid bilayer, which immobilized the drugs in bilayer or inner core when employed as a nano-carrier system, facilitates the targeted, controlled release of drug with decreased cytotoxicity. Currently, Calvo et al. constructed berberine (ber) loaded liposomes against Leishmania infantum infected Balb/c mice. This study demonstrated that Ber-loaded liposomes exhibited higher biocompatibility by increasing its selective index more than sevenfold in macrophages as confirmed by in vitro cytotoxicity assay and reduced parasitic burden in liver and spleen when compared to free drugs (Calvo et al. [2020;](#page-12-0) Frezza et al. [2013\)](#page-13-0). The evaluation of praziquantel loaded liposomes (Lip-PZQ) on Schistosomiasis mansoni (BH strain) showed the following dose for 45 days to mice model decreased the parasitic egg counts and worm in intestine and liver as compared to free drugs. Therefore, this formulation enhanced the bioavailability in host organisms via targeted delivered to liver site where it is absorbed by the tegument of S. mansoni (Frezza et al. [2013](#page-13-0)). Additionally, Voak et al. studied the biodistribution and pharmacodynamics of Amphotericin B encapsulated liposome nanoparticles (AmBisome) at different stages of Visceral leishmaniasis infected Balb/c mice. It has been reported that a higher dose (10 mg/kg) of AmBisome

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rasitic formulations studied in different narasitic diseases f,  $\frac{1}{2}$  and  $\frac{1}{2}$  of different nand manna Table 9.1 Tabular







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Fig. 9.3 Schematic representation of (a) conventional antiparasitic treatment approach, (b) nanoantiparasitic treatment approach [line in green color show effectiveness of anti-parasitic drugs while the line in green color with increased width shows enhance effectiveness due to the existence of nanocarriers as a drug transfer system for antiparasitic drugs distribution to target site]

kills parasite within the 48 h of 14 days of infection while drug potency was decreased when administered after 35 days of advanced infection. Drug accumulation distribution was found to be lower in spleen and liver on 35 days when compared to 14 days. This finding suggested that organ enlargement and other physiological factors are responsible for the distribution and potency of single dose of AmBisome (Voak et al. [2017](#page-14-0)). Moreover, Khodabandeh et al. demonstrated combinatorial therapy against resistant visceral leishmaniasis on 2-year-old boy. The patient was given treatment with liposomal amphotericin B, allopurinol and interferon gamma for 3 weeks. So, therefore together this treatment accelerated the complete cure. This study further suggested to identify resistant species of leishmania along with their response to different treatment (Khodabandeh et al. [2019\)](#page-13-0).

Apart from other, liposomes play a lead role in providing specific and controlled distribution, toxicity reduction, prolonged circulation and minimize side effects of anti-parasitic drugs. Their potency is further increased through modification of the surface by coating with suitable moieties.

#### 9.3.2 Solid Lipid Nanoparticles (SLNs)

Solid lipid nanoparticles (SLNs) are solid core lipid nonvehicle rapidly developed in current years. Mainly it is based on synthetic or natural lipids as a platform to endow drugs adsorption and encapsulation. The SLNs consolidate the advantages of conventional colloidal nano-carriers such as liposomes, oil-in-water emulsions and polymer nanoparticles, i.e., facile mass productions, highly compatible and degradable at physiological conditions. Currently, Khosravi et al. synthesized mannosylated functionalized solid-lipid nanoparticles loaded with paromomycin (PM-SLN-M) to combat acute toxoplasmosis. Although PM has high killing rate of intracellular Toxoplasma in low amount in comparison to PM-SLN-M but it possessed low cytotoxicity on Vero cells. This study suggested that PM-SLN-M exhibited remarkable activity against Toxoplasma without harming the cells of the host (Khosravi et al. [2020\)](#page-13-0). 5-Hydroxy-3methyl-5-phenyl-pyrazoline-1-(S-benzyl dithiocarbazate) (H2bdtc) loaded solid lipid nanoparticles (H2bdtcSLNs) to target Trypanosoma cruzi. Comparative study of benznidazole and H2bdtc-SLNs from in vitro and in vivo outcomes revealed that H2bdtc-SLNs mediate the parasitaemia reduction in mice compared to benznidazole. This study concluded that H2bdtc-SLNs formulation prevent inflammation and lesion in the liver and heart, which enhanced overall survivability in T. cruzi infected mice (Carneiro et al. [2014\)](#page-12-0). Moreover, Radwan et al. prepared formulation of praziquantel solid lipid nanoparticles (SLN-PZQ) against murine S. mansoni infection. The SLN-PZQ demonstrated superior anti-schistosomal activity along with significant bioavailability and sustained release of drug in advanced Schistosoma mansoni-infected groups were showed remarkable inhibition and reduction of worm population in both hepatic and intestinal tissue in comparison to free PZQ (Radwan et al. [2019\)](#page-14-0).

In brief, the anti-parasitic activity has not been predominately studied. Therefore, SLNs offers promising substitute besides some other nanoparticles by facilitated the sustained drug release and specific targeting.

#### 9.3.3 Nanosuspensions

Nanosuspensions are defined by very fine submicron-dispersed colloid of solid drug nanoparticles stabilized by surfactants in aqueous medium. The potential advantages of nanosuspension offer efficient solubility, absorption, dissolution percentage and rate of drugs as well as mediates the prolonged release and reduces the systematic toxicity of drugs. It is an ideal operation to use nanosuspension apart from conventional dosage due to its affordable cost, high loading capacity, facile production and negligible adverse effects. In a study, Kayser prepared aphidicolin-loaded nanosuspensions (Aphi-loaded NSs) against leishmania-infected macrophages. This finding suggested that Aphi-loaded NSs easily phagocytosis by murine macrophage via passive target and enhanced the anti-leishmania activity approx. 140 times. This indicated that nanosuspension increased the payload of drugs and augment the sustained release of drugs to the infected site (Kayser [2000\)](#page-13-0). Recently, curcumin-nanoemulsion (CR-NE) was synthesized to treat acute and chronic toxoplasmosis in mice. This study revealed that CR-NE possessed the significant antitoxoplasmosis by inhibiting the growth of tachyzoites in Peritoneum were observed in both acute and chronic phase compared to curcumin (CR). In addition, decreased in cyst count were determined by the downregulation of BAG1 were maximum in CR-NE treated mice than CR (Azami et al. [2018](#page-12-0)). Zarenezbad et al. study the leishmanicidal activity using nanoemulsion-based nanogel Citrus limon essential oil against Leishmania tropica and Leishmania major. This finding suggested that the toxic effect of *Citrus limon* was found to be more significant compared to *Mentha*  piperita, Anethum graveolens. Moreover, an 80 μg/mL concentration of CLN gel responsible for the complete inhibition of both species of leishmania in in vivo mice model (Zarenezhad et al. [2021\)](#page-14-0).

#### 9.3.4 Polymeric Nanoparticles

Polymeric NPs are solid colloidal suspensions consisting of natural and synthetic polymers used for nanosized drug delivery system. NPs allows therapeutic agents could be encapsulated, entrapped, dissolved and conjugated to their surface. For the drug delivery, polymeric NPs can be employed in variety of forms such as in NPs, nanospheres, or nano-capsules based on preparation methods and their physiochemical properties. In the golden age of pharmaceutical nanocarriers, polymeric NPs have been considered for sustained drug release, and targeted delivery to specific organs and tissues and due to such versatile nature, it offers multiple cargo (proteins, peptides and genes) delivery.

Currently, Elmi et al. synthesized biogenic Chitosan nanoparticles from Penicillium fungi to target human protozoal parasites—Giardia lamblia, Plasmodium falciparum and Trichomonas vaginalis. This study demonstrated that nano-chitosan exhibited excellent anti-parasitic activity by inhibiting the growth rate of cultivated P. falciparum, T. vaginalis and G. lamblia by 59.5%, 99.4%, and 31.3%, respectively with negligible toxic effects. This finding concluded that green synthesized Chitosan nanoparticles combat parasitic infection based on dose-dependent (Elmi et al. [2021\)](#page-13-0). Although, polymer-based nanoparticles are extensively used to carry the drugs for intracellular parasites, for example, amphotericin B for Leishmania as reported in Asthana et al. [\(2015](#page-12-0)), and chloroquine and artemisinin for intracellular targeting of Plasmodium (Tripathy et al. [2013](#page-14-0)). In addition, layered assembly of gelatin nanoparticle encapsulated phthalocianato [bis(dimethylaminoethanoxy)] silicon (NzPC) and modified with polyelectrolytes (polystyrene sulfonate/ polyallylamine hydrochloride) [PGN-NzPc] for PDT (photodynamic therapy) implementation in combating promastigote form of Leishmania amazonensis. The PGN-NzPc facilitated the lower toxicity in the dark while in the presence of PDT trigged the 80% killing of Leishmania promastigotes by altering the morphology (de Souza et al. [2021\)](#page-13-0). Further Lima et al. prepared lignan  $(-)$ -6,6<sup> $\prime$ </sup>-dinitrohinokinin (DNHK) loaded into poly(lactic-co-glycolic acid) nanoparticles (DNHK-loaded PLGA) against Schistosoma mansoni. This study concluded that DNHK-loaded PLGA NPs augmented the sustained release of DNHK to the infection site and showed remarkable potency in killing 100% population of adult worms. This DNHK-PLGA NPs have enhanced anti-schistosomicidal activity (Lima et al. [2017\)](#page-13-0).

## <span id="page-12-0"></span>9.4 Future Perspective and Conclusion

Despite the quick expansion of parasitic diseases worldwide, exciting therapeutic approaches are essential to manage these diseases. But the existing conventional drugs for treating parasitic diseases are outdated, weak, ineffective with deadly toxicities, adverse effects and inducing elevated resistance to disease-causing pathogens. Various reports of re-purposing of conventional drugs via nanoformulations have shown significant responses by reducing their toxicity and enhancing the therapeutic efficacy coupled with low cost. Since, reports suggest that nanoformulations will increase efficacy by higher targeting efficiency, sitespecific delivery, and enhanced bio-availability of the drug at the disease site. Therefore, more emphasis is to be given to systematic investigation and research related to the selection designing and fabrication of nanocarriers. Here, the physiochemical characteristics of nanoparticles can be a conclusive factor playing a decisive role in determining the effectiveness of nanocarriers in delivering antiparasitic drugs at a desired locus with a hope for improved pharmacokinetic and pharmacodynamic properties. Moreover, the research method with powerful technologies together can advance the treatment approach with more precision in in vivo research and clinical studies. Currently, no nano-formulations of antiparasitic drug is available in the marketplace, lots of them are still in the process of better clinical trials, formulation designs, preclinical studies, and commercialization under the name of "antiparasitic nanomedicines". With constant efforts, efficient therapy will progress as an inevitable trend in pharmaceutical industries with antiparasitic nanomedicine shaving an infinite future against parasitic diseases controlling their spread.

#### References

- Amer EI, Abou-El-Naga IF, Boulos LM, Ramadan HS, Younis SS (2022) Praziquantelencapsulated niosomes against Schistosoma mansoni with reduced sensitivity to praziquantel. Biomedica 42(1):67–84
- Asthana S, Jaiswal AK, Gupta PK, Dube A, Chourasia MK (2015) Th-1 biased immunomodulation and synergistic antileishmanial activity of stable cationic lipid–polymer hybrid nanoparticle: biodistribution and toxicity assessment of encapsulated amphotericin B. Eur J Pharm Biopharm 89:62–73
- Azami SJ, Teimouri A, Keshavarz H, Amani A, Esmaeili F, Hasanpour H et al (2018) Curcumin nanoemulsion as a novel chemical for the treatment of acute and chronic toxoplasmosis in mice. Int J Nanomed 13:7363
- Cable J, Barber I, Boag B, Ellison AR, Morgan ER, Murray K et al (2017) Global change, parasite transmission and disease control: lessons from ecology. Philos Trans R Soc Lond B Biol Sci 372(1719):20160088
- Calvo A, Moreno E, Larrea E, Sanmartín C, Irache JM, Espuelas SJP (2020) Berberine-loaded liposomes for the treatment of Leishmania infantum-infected BALB/c mice. Pharmaceutics 12(9):858
- Carneiro ZA, da S Maia PI, Sesti-Costa R, Lopes CD, Pereira TA, Milanezi CM et al (2014) In vitro and in vivo trypanocidal activity of H2bdtc-loaded solid lipid nanoparticles. PLoS Negl Trop Dis 8(5):e2847
- <span id="page-13-0"></span>Chen A, Shi Y, Yan Z, Hao H, Zhang Y, Zhong J et al (2015) Dosage form developments of nanosuspension drug delivery system for oral administration route. Curr Pharm Des 21(29): 4355–4365
- Chen X, Li J, Huang Y, Wei J, Sun D, Zheng N (2017) The biodistribution, excretion and potential toxicity of different-sized Pd nanosheets in mice following oral and intraperitoneal administration. Biomater Sci 5(12):2448–2455
- Darvishi MM, Moazeni M, Alizadeh M, Abedi M, Tamaddon A-M (2020) Evaluation of the efficacy of albendazole sulfoxide (ABZ-SO)–loaded chitosan-PLGA nanoparticles in the treatment of cystic echinococcosis in laboratory mice. Parasitol Res 119(12):4233–4241
- Das S, Chaudhury A (2011) Recent advances in lipid nanoparticle formulations with solid matrix for oral drug delivery. AAPS PharmSciTech 12(1):62–76
- David Sibley L (2011) Invasion and intracellular survival by protozoan parasites. Immunol Rev 240(1):72–91
- de Souza C, Carvalho JA, Abreu AS, de Paiva LP, Ambrósio JA, Junior MB et al (2021) Polyelectrolytic gelatin nanoparticles as a drug delivery system for the promastigote form of Leishmania amazonensis treatment. J Biomater Sci Polym Ed 32(1):1–21
- Elmi T, Rahimi Esboei B, Sadeghi F, Zamani Z, Didehdar M, Fakhar M et al (2021) In vitro antiprotozoal effects of nano-chitosan on Plasmodium falciparum, Giardia lamblia and Trichomonas vaginalis. Acta Parasitol 66(1):39–52
- Frezza TF, Gremião MPD, Zanotti-Magalhães EM, Magalhães LA, de Souza ALR, Allegretti SM (2013) Liposomal-praziquantel: efficacy against Schistosoma mansoni in a preclinical assay. Acta Trop 128(1):70–75
- Fülöp V, Jakab G, Bozó T, Tóth B, Endrésik D, Balogh E et al (2018) Study on the dissolution improvement of albendazole using reconstitutable dry nanosuspension formulation. Eur J Pharm Sci 123:70–78
- Jiménez B, Maya C, Velásquez G, Torner F, Arambula F, Barrios J et al (2016) Identification and quantification of pathogenic helminth eggs using a digital image system. Exp Parasitol 166:164– 172
- Kayser O (2000) Nanosuspensions for the formulation of aphidicolin to improve drug targeting effects against Leishmania infected macrophages. Int J Pharm 196(2):253–256
- Khodabandeh M, Rostami A, Borhani K, Gamble HR, Mohammadi M (2019) Treatment of resistant visceral leishmaniasis with interferon gamma in combination with liposomal amphotericin B and allopurinol. Parasitol Int 72:101934
- Khosravi M, Mohammad Rahimi H, Doroud D, Mirsamadi ES, Mirjalali H, Zali MR et al (2020) In vitro evaluation of mannosylated paromomycin-loaded solid lipid nanoparticles on acute toxoplasmosis. Front Cell Infect Microbiol 10:33
- Kumar R, Pandey K, Sahoo GC, Das S, Das V, Topno R et al (2017) Development of high efficacy peptide coated iron oxide nanoparticles encapsulated amphotericin B drug delivery system against visceral leishmaniasis. Mater Sci Eng C Mater Biol Appl 75:1465–1471
- Leal Filho W, Ternova L, Parasnis SA, Kovaleva M, Nagy GJ (2022) Climate change and zoonoses: a review of concepts, definitions, and bibliometrics. Int J Environ Res Public Health 19(2):893
- Lima TC, Lucarini R, Luz PP, de Faria EH, Marçal L, Magalhães LG et al (2017) In vitro schistosomicidal activity of the lignan  $(-)$ -6, 6'-dinitrohinokinin (DNHK) loaded into poly (lactic-co-glycolic acid) nanoparticles against Schistosoma mansoni. Pharm Biol 55(1): 2270–2276
- Liu D, Mori A, Huang L (1992) Role of liposome size and RES blockade in controlling biodistribution and tumor uptake of GM1-containing liposomes. Biochim Biophys Acta 1104(1):95–101
- Muga JO, Gathirwa JW, Tukulula M, Jura WGO (2018) In vitro evaluation of chloroquine-loaded and heparin surface-functionalized solid lipid nanoparticles. Malar J 17(1):133
- Nahar M, Dubey V, Mishra D, Mishra PK, Dube A, Jain NK (2010) In vitro evaluation of surface functionalized gelatin nanoparticles for macrophage targeting in the therapy of visceral leishmaniasis. J Drug Target 18(2):93–105
- <span id="page-14-0"></span>Negi JS, Chattopadhyay P, Sharma AK, Ram V (2013) Development of solid lipid nanoparticles (SLNs) of lopinavir using hot self nano-emulsification (SNE) technique. Eur J Pharm Sci 48(1–2):231–239
- Pensel PE, Gamboa GU, Fabbri J, Ceballos L, Bruni SS, Alvarez LI et al (2015) Cystic echinococcosis therapy: albendazole-loaded lipid nanocapsules enhance the oral bioavailability and efficacy in experimentally infected mice. Acta Trop 152:185–194
- Radwan A, El-Lakkany NM, William S, El-Feky GS, Al-Shorbagy MY, Saleh S et al (2019) A novel praziquantel solid lipid nanoparticle formulation shows enhanced bioavailability and antischistosomal efficacy against murine S. mansoni infection. Parasit Vectors 12(1):304
- Safarpour H, Majdi H, Masjedi A, Pagheh AS, de Lourdes Pereira ML, Rodrigues Oliveira SM et al (2021) Development of optical biosensor using protein A-conjugated chitosan–gold nanoparticles for diagnosis of cystic echinococcosis. Biosensors (Basel) 11(5):134
- Saqib M, Ali Bhatti AS, Ahmad NM, Ahmed N, Shahnaz G, Lebaz N et al (2020) Amphotericin B loaded polymeric nanoparticles for treatment of leishmania infections. Nanomaterials (Basel) 10(6):1152
- Sinha J, Mukhopadhyay S, Das N, Basu MK (2000) Targeting of liposomal andrographolide to L. donovani-infected macrophages in vivo. Drug Deliv 7(4):209–213
- Sun Y, Chen D, Pan Y, Qu W, Hao H, Wang X et al (2019) Nanoparticles for antiparasitic drug delivery. Drug Deliv 26(1):1206–1221
- Tripathy S, Mahapatra SK, Chattopadhyay S, Das S, Dash SK, Majumder S et al (2013) A novel chitosan based antimalarial drug delivery against Plasmodium berghei infection. Acta Trop 128(3):494–503
- Varela-Aramburu S, Ghosh C, Goerdeler F, Priegue P, Moscovitz O, Seeberger PH et al (2020) Targeting and inhibiting Plasmodium falciparum using ultra-small gold nanoparticles. ACS Appl Mater Interfaces 12(39):43380–43387
- Vercruysse J, Schetters T, Knox D, Willadsen P, Claerebout E (2007) Control of parasitic disease using vaccines: an answer to drug resistance? Rev Sci Tech 26(1):105
- Voak AA, Harris A, Qaiser Z, Croft SL, Seifert K (2017) Pharmacodynamics and biodistribution of single-dose liposomal amphotericin B at different stages of experimental visceral leishmaniasis. Antimicrob Agents Chemother 61(9):e00497–e00417
- Want MY, Yadav P, Khan R, Chouhan G, Islamuddin M, Aloyouni SY et al (2021) Critical antileishmanial in vitro effects of highly examined gold nanoparticles. Int J Nanomed 16:7285
- Zafar A, Ahmad I, Ahmad A, Ahmad M (2016) Copper(II) oxide nanoparticles augment antifilarial activity of Albendazole: in vitro synergistic apoptotic impact against filarial parasite Setaria cervi. Int J Pharm 501(1–2):49–64
- Zarenezhad E, Agholi M, Ghanbariasad A, Ranjbar A, Osanloo M (2021) A nanoemulsion-based nanogel of Citrus limon essential oil with leishmanicidal activity against Leishmania tropica and Leishmania major. J Parasit Dis 45(2):441–448