Chapter 6 Arbuscular Mycorrhizal Fungi: A Next-Generation Biofertilizer for Sustainable Agriculture

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Abstract Climate change has a significant impact on environmental conditions, which affects the growth and productivity of plants. As a result, sustainable crop production continues to be a major global challenge, attracting increasing attention from the scientific community in order to feed the world's growing population while reducing the use of conventional chemical fertilizers and pesticides. Arbuscular mycorrhizal fungi (AMF) are widely used to build symbiotic relationships with over 80% of the species of the land, including most of the cultivated plants. These fungi are of great interest because of their biofertilizer potential (microbial inoculants) in low-input and organic agriculture, which represents an adequate alternative tool for chemical fertilizers. Using AMF as biofertilizer enables plants to use mineral elements such as nitrogen and phosphorous effectively. In addition to an improvement in plant nutrition, AMF plays an important role in improving soil structure, fertility and heavy metal remediation. In conclusion, AMF can be used as a potential biofertilizer for control of environmental stress and may open new strategies to support agriculture and increase global food safety.

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6.1 Introduction

Food production needs to be doubled by 2050 in order to meet the demands of a growing population. The rising costs and adverse effects of chemical fertilizers on the environment and human health have pushed the agrarian community to look for substitutes for these chemical fertilizers (Srivastava et al. [2018](#page-15-0)). Biofertilizers are suitable alternatives to artificially synthesized fertilizers as they are less harmful to the environment, improve soil health and promote the quality and quantity of crop yield (Suhag [2016\)](#page-15-0). Biofertilizers are "microbial inoculants" that allow effective intake of mineral elements such as nitrogen and phosphorus and enhance drought tolerance, salt tolerance and improve plant health (Alori et al. [2017;](#page-11-0) Igiehon and Olubukola [2017](#page-13-0)). Most farmers in the world widely use living organisms like bacteria, fungi and cyanobacteria etc. as biofertilizers that are inoculated with seed or in soil to colonize the rhizosphere to increase the availability of nutrients (Sadhana [2014\)](#page-14-0). The use of symbiotic mycorrhizal fungi, particularly arbuscular mycorrhizal fungi (AMF), as biofertilizer has been adopted in agriculture systems because of their potential for improving soil quality, water stress tolerance, altering root architecture and pathogen resistance (Abbot and Robson [1991\)](#page-10-0). The mycorrhiza is an obligatory symbiotic association between fungi and the roots of higher plants (Sieverding [1995](#page-15-0)). The German Forest pathologist Frank invented the name mycorrhiza in 1885, which comes from two terms, the Greek word "mycos" meaning fungus, and the Latin word "rhiza" referring to fungal roots (Frank [1885](#page-12-0)). The AMF are ubiquitous endomycorrhiza that can inhabit a variety of ecosystems and form symbiotic association with roots of angiosperms and other plants (Gerdemann [1968](#page-12-0)) with more than 80% of land plant species including crops (Wang and Qiu [2006\)](#page-15-0). AMF also gives protection against abiotic stress (Auge [2001;](#page-11-0) Javaid [2007](#page-13-0)) and biotic stress to their host plants (Khaosaad et al. [2007\)](#page-13-0). Mycorrhiza also increase the fixation of nitrogen in nodule plants. Plants which receive good nutrition can withstand infections, and this is one strategy to combat diseases that are transmitted to the soil (Linderman and Davis [2004](#page-14-0)). The barrier created by Ectomycorrhizae when they cover the exterior surface of the root is the most obvious mechanism for protecting against illnesses (Castellano and Molina [1989\)](#page-12-0). The current chapter focuses on the significance of AMF as biofertilizers for sustainable agriculture, highlighting the importance of AMF and achievements in research related to their agricultural applications.

6.2 Development of Mycorrhizal Network

The AM fungi are classed as a separate phylum termed glomeromycota, which has roughly 150 species with considerable genetic and functional diversity (Smith and Read [2008](#page-15-0); Bucking et al. [2012\)](#page-11-0). A study by Hosny et al. ([1998\)](#page-13-0) indicates that asexually reproducing fungi have coenocytic hyphae and spores. The mycorrhizal fungi is not strictly a biofertilizer as it does not add mineral nutrition to soil like nitrogen-fixing bacteria but it improves the uptake of soil nutrients through arbuscules and improves plant development and soil health (Garg and Manchanda [2007;](#page-12-0) Solaiman [2014](#page-15-0)). An arbuscular is a tiny tree-shaped fungal structure that grows in the intercellular and intracellular regions of roots and is a key site for the exchange of nutrients between the two symbiotic partners (He and Nara [2007](#page-13-0)). A variety of genes and hormones initiate the symbiotic interaction between plant roots and the fungi. Strigolactones and lipochito-oligosaccharides generated by fungi are important in the development of the association (Mohanta and Bae [2015](#page-14-0); Sharma et al. [2021](#page-15-0)).

The symbiosis is established through a series of morphological and physiological interactions between the two hosts (Amalero et al. [2003\)](#page-11-0). The various developmental stages of the AM colony in the plant roots are as follows.

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AM fungi are obligatory biotrophs; they rely on their autotrophic host to complete their life cycle in a symbiotic association and generate the next generation of spores (Fig. [6.1\)](#page-3-0). Germination of fungal spores in a soil is the only plant-independent phase in the life cycle of mycorrhizal fungi (Bonfante and Bianciotto [1995](#page-11-0)). The spores germinate and grow into an extended mycelium for 2–3 weeks into extended mycelium, displaying apical dominance. The mycelium growth ceases after 2–4 weeks in the absence of an appropriate host. The presence of host root exudate stimulates intense hyphal growth and branching to increase the probability of contact with host roots (Paszkowski [2006\)](#page-14-0).

$6.2.2$ \mathcal{L} early Symbiotic Phase \mathcal{L}

Between the fungus and plant root epidermis AMF forms a cell-to-cell contact called appressorium (hypophodium). The formation of appressorium is the first morphological sign of symbiosis. The AM fungi penetrate into the roots of the host plant by penetrating the hyphae emerging from the appressorium. The hyphae successfully penetrates the cell wall using both mechanical and enzymatic catalysed mechanisms (Garcia-Garrido and Ocampo [2002](#page-12-0)).

Fig. 6.1 Life cycle of arbuscular mycorrhizal fungi

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The mycorrhiza colonizes roots by arbuscule, the tree-like fungal structures formed intracellularly subtended by intercellular hyphae in the cortical region. The structures are key sites for exchange of nutrients between two hosts (Dickson et al. [2007\)](#page-12-0). The periarbuscular membrane (PAM), a key interface for symbiotic interaction, keeps the fungus excluded from host cytoplasm. The exchange of nutrients between the two partners is mediated by membrane transport proteins such as P -type H^+ ATPase and phosphate transport (Bucher [2007](#page-11-0)).

The AM fungi can make a network of mycelia in plant roots and in the soil. Extraradical mycelium (ERM) grows in the soil (Fig. [6.2](#page-4-0)). They draw nutrients from the soil and deliver them to the plant's roots. The mycelium formed within the roots is called intraradical mycelium (IRM). The IRM releases nutrients at the interface and absorbs carbon from the plant roots in exchange. The absorbed carbon is utilized for expansion and spore formation by ERM. The spores can initiate colonization of nearby plants (Bucking et al. [2012\)](#page-11-0).

Fig. 6.2 Development of mycorrhizal network

6.3 Arbuscular Fungi as a Potential Biofertilizer

The AM fungi act as an important link between plants and the soil to achieve the goal of sustainable agriculture. They mediate nutrient transfer and therefore contribute to the maintenance of soil structure, soil nutrition and plant nutrition (Gentili and Jumpponen [2006](#page-12-0)). The high metabolic rate and efficient translocation of micronutrients and macronutrients from soil to plant mediated by fungi improve plant growth and yield in chick pea, custard apple and olive plantlets (Kumar et al. [2002;](#page-13-0) Briccoli et al. [2015\)](#page-11-0). Fungi can mobilize important nutrients like phosphorus (P), nitrogen (N) and act as a carbon sink in the soil (Bonfante and Genre [2010](#page-11-0)). As a result, the fungi have the potential to act as biofertilizer for sustainable agriculture (Giri et al. [2019\)](#page-12-0).

6.4 The Role of AMF in Improving Soil Health and Fertility

By enhancing soil nitrogen intake by the plant, fungal hyphae stabilize soil aggregates. Extracellular polysaccharide and glomalin exudates aid in the formation of network hyphae in the soil. The polysaccharide glomalin is the main contributor to soil formation because it promotes the development of organic matter and attachment of the hyphae to the soil (Adetunji et al. [2019](#page-10-0)). Soil aggregation improves soil health and quality by improving soil porosity, water-holding capacity, gaseous exchange, protecting organic carbon and promoting the growth of beneficial micro-fauna (Srivastava et al. [2018\)](#page-15-0). Several studies show that AMF reduces the harmful impact of heavy metal contamination in soil caused by anthropogenic activities and the usage of agrochemical products (Schützendübel and Polle [2002;](#page-14-0) Dong et al. [2008\)](#page-12-0). The fungi can absorb calcium, aluminium, cadmium, selenium and arsenic acid (Khan et al. [2000](#page-13-0); Al-Agel et al. [2005](#page-11-0)). It mitigates the effects of these heavy metals by immobilization, adsorption on the hyphal wall. The AMF also causes metal resistance in plants by altering metabolic processes such as phenylpropanoid pathway (Janeeshma and Puthur [2020](#page-13-0)).

6.4.1 **Role of AMF in Plant Nutrition** \mathcal{O}

Many studies have shown that AMF colonizes plant roots containing essential plant nutrients such as nitrogen (N), phosphorus (P), sulphur (S), potassium (K), calcium (Ca), copper (Cu), iron (Fe), and magnesium (Mg), among others. (Marschner and Dell [1994](#page-14-0)). AMF facilitates dissolution, transportation of immobile nutrients bound to rocks and mineralization of organic matter (Parihar et al. [2019](#page-14-0)).

6.4.1.1 Phosphorus (P) Absorption

Phosphorus is an essential nutrient for plants but difficult to absorb from soil due to low diffusion rate. Mycorrhizal fungi release an enzyme phosphatase that mobilizes organic P and increases its absorption by plants (Shen et al. [2011;](#page-15-0) Malla et al. [2004;](#page-14-0) Nath et al. [2018\)](#page-14-0). Phosphorus deficiency in plants inhibits photosynthesis, respiration and cell division, which subsequently reduces the yield (Baas and Kuiper [1989\)](#page-11-0). A study by Walder et al. ([2015\)](#page-15-0) indicates that the symbiotic interaction between the two hosts induces the expression of the Pi transporter in sorghum and flex plants. The fungal hyphae also reduce phosphorus leaching by different mechanisms involving extensive adsorption on the hyphae surface, storage of orthophosphate and polyphosphate in the hyphae and chelation of P with fungi exuded glycoprotein (Parihar et al. [2019](#page-14-0)).

6.4.1.2 Nitrogen (N) Absorption

The AMF can absorb N in both organic forms as amino acids (Whiteside et al. [2012](#page-15-0)) and inorganic form as nitrate and ammonium (Govindarajulu et al. [2005\)](#page-12-0). The extraradical mycelium of the fungi absorbs the inorganic forms. In the soil, the hyphae can take ammonium at a lower quantity than the roots (Johansen et al. [1994\)](#page-13-0). Ammonium transporters are found in arbuscules which provide nutrients to the host plant. The AM fungi-inducible nitrate and ammonium transporters have been identified in tomato and soyabean plants that facilitate the absorption of nutrients by the plants (Kobae et al. [2010\)](#page-13-0). Several amino acids such as glycine, cysteine, serine, arginine, aspartic acid, glutamine acids and cysteine etc. are absorbed by the fungi and then converted to ammonium for translocation at the symbiotic interface with the plant (Smith and Smith [2011\)](#page-15-0).

6.4.1.3 Sulphur Absorption

Because of its redox characteristics and capacity to form disulphide bonds between cysteine amino acids, sulphur plays an important role in the biological function of many substances. Although plants absorb inorganic sulphate as their primary source of sulphur, 95% of soil sulphur is bonded in organic molecules. The form is not directly available to plants. The mycorrhizal fungi have sulphur transporters that make the element available to the plants (Giovannetti et al. [2014](#page-12-0)). The mycorrhizal plants can obtain sulphur from organic sources. Allen and Shachar-Hill [\(2009](#page-11-0)) observed 25% more sulphur content in plant roots with mycorrhizal association at moderate sulphur concentration as compared to nonmycorrhizal plant roots.

6.4.1.4 Potassium Absorption

Potassium is considered an important macronutrient for plants responsible for enzyme activation, regulation of stomatal opening and serving as an osmolyte in plant cells (Morgan and Connolly [2013;](#page-14-0) Kumar et al. [2020\)](#page-13-0). Although potassium is abundant in soil, it is not readily available to plants. The role of AMF in potassium uptake by host plants has received less attention. A study by Jianjian et al. [\(2019](#page-13-0)) observed the overexpression of potassium transporter protein in the roots of Lotus japonicas plants infected with AMF. Furthermore, AM symbiosis associated with potassium nutrition is correlated to alleviating abiotic stresses including salinity, drought, heavy metals and temperature stress (Berruti et al. [2016\)](#page-11-0).

$6.4.2$ Role of AMF in Plant Biotic Stress Tolerance

More than 90% of total mycorrhizal roots colonize the fungus in intercellular and intracellular tissues. It is proposed that plants can tolerate the intense mycorrhizal network by suppression of plant defence mechanisms against the AMF (Chen et al. [2018\)](#page-12-0). However, the general disease resistance of the plant is not attenuated. Indeed, plants show increased disease resistance against rhizospheric pathogens, pests and parasitic plants either by secreting repulsive exudates from mycorrhizal roots (Kwak et al. [2018](#page-13-0)). The AMF provides an effective way to control the biotic stress by improved nutrition and induction of plant defence process called as systemic acquired resistance (SAR). Additionally, plants exhibit fast and strong reactions against pathogens by a phenomenon called priming or induced systemic resistance (IRS) (Conrath et al. [2006](#page-12-0)). The AMF can directly interfere with plant pathogens either by release of antimicrobial substances or by competing with the pathogens for space and resources (Jacott et al. [2017](#page-13-0)). AMF-induced alleviated plant defence response against various biotic stress is as follows.

6.4.2.1 AMF and Parasite Tolerance

Plants colonized with AMF increase tolerance against parasitic Meloidogyne species of nematode. The AMF competes directly with the nematode for root space and reduces the process of reproduction (Dar and Reshi [2017](#page-12-0)). A study by López-Ráez et al. (2009) (2009) indicates that the AMF inhibit the growth of parasitic plants like *Striga* hermonthica in maize and Striga and Orobanche in sorghum. Thus, the presence of fungi in plant roots can act as biocontrol agents for sustainable agriculture.

6.4.2.2 AMF and Soil-Borne Pathogens

A number of reports have explained the positive effect of AMF-induced plant tolerance to biotic stress triggered by soil-borne pathogens. The symbiosis suppresses growth of fungi Fusarium, Macrophomina, Rhizoctonia, Verticillum, and oomycetes like Pythium and Phytophthora responsible for wilting and root rot disease (Hao et al. [2009;](#page-13-0) Harrier and Watson [2004;](#page-13-0) Whipps [2004\)](#page-15-0).

6.4.2.3 AMF and Insects

Rhizophagous insects are a common biotic stress for many plants. Hartley and Gange [\(2009](#page-13-0)) explained that the mycorrhiza can strongly influence the insect's growth by enhancing insect resistance of plant, but the effects may vary with the feeding mechanisms and lifestyle of the insects. Additionally, AMF-associated plant defence against insects is closely associated with levels of flavonoids and phenolic compounds in host plants (Wang et al. [2020\)](#page-15-0).

6.4.3 6.4.3 Role in Plant Abiotic Stress Tolerance

The plants confront abiotic stress like drought, salinity, extreme temperature, and heavy metals which show harmful effects on their growth and yield (Kumar et al. [2017;](#page-13-0) Nath et al. [2017](#page-14-0)). Abiotic stress can negatively affect plant survival and productivity. Therefore, it can act as a foremost threat to global food security (Kumar and Verma [2018](#page-13-0)). The AMF improves plants' tolerance to these abiotic stresses by various metabolic and physiological changes in plants (Malhi et al. [2021\)](#page-14-0). The role of AMF to combat various abiotic stresses is as discussed below:

6.4.3.1 AMF and Drought Stress Tolerance

Drought is a condition when water is unavailable to plants for its physiological functions. The environmental condition is also known as water stress (Subramanian and Charest [1998](#page-15-0)). The fluctuated transpiration rate generates reactive oxygen species (ROS), and consequently accelerates oxidative stress in plants (Auge [2001;](#page-11-0) Barzana et al. [2012\)](#page-11-0). Mycorrhiza can progress plant development and growth by enhancing root network and thickness, plant biomass and nutrient absorption and transport during drought conditions (Davies et al. [2002](#page-12-0)). The mycorrhizal inoculation facilitates synthesis of more dense hyphal networks and excretes glumalin which augment more water and nutrients absorption, which in turn improves soil quality (Gholamhoseini et al. [2013\)](#page-12-0).

The AMF symbiosis influences numerous biochemical and physiological processes such as (1) augmented osmotic regulation, (2) enhanced gas exchange, (3) absorption and transport of water and nutrients, and (4) better defence against oxidative stress (Marulanda et al. [2007](#page-14-0)). A study on Zea mays plants colonized with mycorrhiza Glomus intraradices reported expression of two aquaporin genes (Gint AQPF1 and Gint AQPF2) in root cortical cells holding arbuscules under drought stress (Moussa and Abdel-Aziz [2008;](#page-14-0) Li et al. [2013\)](#page-13-0). AMF-mediated enhancement in drought resistance has been demonstrated by (Li et al. [2019\)](#page-14-0) in C3 plant (Leymus chinensis) and C4 plant (Hemarthria altissima) observed because of alteration in expression of antioxidants enzyme.

6.4.3.2 AMF and Salt Stress Tolerance

Salinity in soil is a prime problem for many plants growing in arid and semiarid regions (Giri et al. [2003;](#page-12-0) Al-Karaki [2006\)](#page-11-0). The high accumulation of salt in soil decreases aeration and porosity of soil and therefore affects water translocation, which results in drought-like stress (Mahajan and Tuteja [2005\)](#page-14-0). Plants under salt stress show decrease in rate of photosynthesis, reduction in activities of antioxidant enzyme, less stomatal conductance, decreased membrane stability, and low relative water content of the plants (Talaat and Shawky [2012](#page-15-0)). Salinity also causes oxidative stress in the plant by producing more reactive oxygen species (ROS) (Ahmad et al. [2010\)](#page-11-0). AMF inoculated plants develop strategies to enhance the antioxidant system which protects the plant cells from oxidative damage (Rai et al. [2011](#page-14-0)). The defence system develops superoxide dismutase (SOD) antioxidant enzyme that converts superoxide molecules to oxygen and hydrogen peroxide (H_2O_2) . Besides SOD, catalase (CAT) enzyme clears H_2O_2 by decomposing it to less reactive water and oxygen. These enzymes are continuously generated in the mitochondria, peroxisome and cytoplasm of the plant.

Several research investigations have reported on AMF's efficiency to promote growth, yield and development in plants subjected to salinity stress. AMF is effective for plants' response under different salt concentrations. Allium sativum plant inoculated with AMF showed expanded leaf area, more fresh and dry weights under high NaCl concentration (100 mM) as compared to the plant without mycorrhizal association (Borde et al. [2010](#page-11-0)). A study of Ghazi and Al-Karaki ([2001\)](#page-12-0) on tomato plant inoculated with fungi Glomus mosseae observed increase in biomass under moderate saline conditions. Under salt stress, AMF-inoculated rice plants preferably absorb more K^+ ion and avoid intake of Na^+ ions compared with control rice plants. The crop showed AMF induced more salt tolerance and crop yield (Mohsin et al. [2020](#page-14-0)). El-Nashar ([2017\)](#page-12-0) observed that the Antirrhinum majus plants improved their growth rate, their feed-water potential and their water efficiency. The favourable benefits of AMF association on biological parameters like photosynthetic rate, stomatal conductivity, and leaf water relationships under salt stress have been described by Ait-El-Mokhtar et al. [\(2019](#page-11-0)).

6.4.3.3 Heavy Metal Tolerance

Heavy metals like Cu, Co, Fe, Mn, and Zn are essential for plant growth. However, increased concentrations of these metals are hazardous to the plants due to the production of reactive oxygen species (ROS) by the plants (Palmer and Guerinot [2009;](#page-14-0) Puig and Penarrubia [2009](#page-14-0)). The AMF shows positive effects on plant growth under cadmium stress by lowering the levels of hydrogen peroxide and malonaldehyde (Hashem et al. [2016](#page-13-0)). A study by Yong et al. ([2014\)](#page-15-0) reported effective removal of heavy metals from polluted environments in clone of Schizosaccharomyces pombe. Metal dilution in plant tissues is also thought to be caused by increased growth or chelation in the rhizospheric soil (Kapoor et al. [2013;](#page-13-0) Audet [2014\)](#page-11-0). AMF would have reportedly bind Cd and Zn in the cortical cells and mental hyphae to restrict their intake by plant and increase growth, yield and nutrient status of plants (Andrade and Silveira [2008](#page-11-0); Garg and Chandel [2012](#page-12-0)).

6.4.3.4 Thermal Stress Tolerance

Thermal fluctuations lead to reduced germination, low rate of photosynthesis, retarded plant growth, yield and biomass production (Wahid et al. [2007;](#page-15-0) Hasanuzzaman et al. [2013\)](#page-13-0). A study by Maya and Matsubara ([2013\)](#page-14-0) reported the beneficial effects of AMF on plant growth and yield under thermal stress in Glomus fasciculatum. A number of reports suggest AMF improves growth rates in plants grown under low temperatures when compared to plants without mycorrhizal association (Zhu et al. [2010a](#page-15-0), [2010b](#page-15-0)). The AMF symbiosis supports the plants' survival under low temperature along with improved plant growth and development (Gamalero et al. [2009](#page-12-0); Birhane et al. [2012\)](#page-11-0). AMF strengthens the plant defence system, leading to more synthesis of various secondary metabolites and proteins (Abdel Latef and Chaoxing [2011b](#page-10-0)). It also facilitates the plant in efficient moisture retention, improved chlorophyll production and better osmotic adjustment capacity (Abdel Latef and Chaoxing [2011a;](#page-10-0) Zhu et al. [2010a,](#page-15-0) [2010b\)](#page-15-0). In addition, the AMF can maintain host plant moisture (Zhu et al. [2010a](#page-15-0)), boost secondary plant metabolites leading towards strengthening plant immune systems and enhance plant protein in support of cold stress conditions (Abdel Latef and Chaoxing 2011b).

6.5 Conclusion and Future Challenges

There are many factors, such as compatibility with the environment, competition with other soil organisms, and timing of the inoculation, that can affect the success of establishment of symbiosis. The use of AM fungi in agriculture requires the knowledge of its adaption in the target ecosystem and the establishment of a functional symbiosis in different types of soils.

The use of AM fungi as a biofertilizer is an economical, effective and eco-friendly approach toward the attainment of low-input farming. The symbiotic fungi develop an intensive network of mycelium that improves soil structure, fertility and plant health by efficient absorption of micro- and macronutrients from the soil. Fungi have a number of genes and molecular pathways that facilitate more effective nutrient uptake and transport to the plant roots as compared to plants without a mycorrhizal association. AMF association in plant roots is an effective tool to combat biotic and abiotic stresses responsible for loss of crop productivity and yield. Therefore, the AM fungi have potential biofertilizer to act as sustainable agriculture.

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References

- Abbot LK, Robson AD (1991) Factors influencing the occurrence of arbuscular mycorrhizae. Agric Ecosyst Environ 35:121–150
- Abdel Latef AA, Chaoxing H (2011a) Effect of arbuscular mycorrhizal fungi on growth, mineral nutrition, antioxidant enzymes activity and fruit yield of tomato grown under salinity stress. Sci Hortic 127:228–233
- Abdel Latef AA, Chaoxing H (2011b) Arbuscular mycorrhizal influence on growth, photosynthetic pigments, osmotic adjustment and oxidative stress in tomato plants subjected to low temperature stress. Acta Physiol Plant 33:1217–1225
- Adetunji CO, Kumar D, Raina M, Arogundade O, Sarin NB (2019) Endophytic microorganisms as biological control agents for plant pathogens: a panacea for sustainable agriculture. In: Varma A, Tripathi S, Prasad R (eds) Plant biotic interaction-state of the art. Springer Nature, Switzerland. isbn:978-3-030-26656-1
- Ahmad P, Jaleel CA, Sharma S (2010) Antioxidative defense system, lipid peroxidation, proline metabolizing enzymes and biochemical activity in two *Morus alba* genotypes subjected to NaCl stress. Russian J Plant Physiol 57:509–517
- Ait-El-Mokhtar M, Laouane RB, Anli M, Boutasknit A, Wahbi S, Meddich A (2019) Use of mycorrhizal fungi in improving tolerance of the date palm (*Phoenix dactylifera* L.) seedlings to salt stress. Sci Hortic 253:429–438
- Al-Agel A, Sylvia DM, Ma LQ (2005) Mycorrhizae increase arsenic uptake by the hyperaccumulator Chinese brake fern (Pteris vittata L.). J Environ Qual 34:2181–2186
- Al-Karaki GN (2006) Nursery inoculation of tomato with arbuscular mycorrhizal fungi and subsequent performance under irrigation with saline water. Sci Hortic 109:1–7
- Allen JW, Shachar-Hill Y (2009) Sulfur transfer through an arbuscular mycorrhiza. Plant Physiol 149:549–560
- Alori ET, Dare MO, Babalola OO (2017) Microbial inoculants for soil quality and plant health. In: Sustainable agriculture reviews, pp 281–307
- Amalero EG, Guido LI, Graziella BE, Philippe LE (2003) Methods for studying root colonization by introduced beneficial bacteria. Agronomie 23:407–418
- Andrade SAL, Silveira APD (2008) Mycorrhiza influence on maize development under Cd stress and P supply. Braz J Plant Physiol 20(1):39–50
- Audet P (2014) Arbuscular mycorrhizal fungi and metal phytoremediation: ecophysiological complementarity in relation to environmental stress. In: Ahmad P, Rasool S (eds) Emerging technologies and management of crop stress tolerance. Academic, San Diego, pp 133–160
- Auge RM (2001) Water relations, drought and vesicular–arbuscular mycorrhizal symbiosis. Mycorrhiza 11:3–42
- Baas R, Kuiper D (1989) Effects of vesicular-arbuscular mycorrhizal infection and phosphate on Plantago major ssp. pleiosperma in relation to internal cytokinin concentrations. Physiol Plant 76:211–215
- Barzana G, Aroca R, Paz JA (2012) Arbuscular mycorrhizal symbiosis increases relative apoplastic water flow in roots of the host plant under both well-watered and drought stress conditions. Ann Bot 109:1009–1017
- Berruti A, Lumini E, Balestrini R, Bianciotto V (2016) Arbuscular mycorrhizal fungi as natural biofertilizers: let's benefit from past successes. Front Microbiol 6:1–13. [https://doi.org/10.3389/](https://doi.org/10.3389/fmicb.2015.01559) [fmicb.2015.01559](https://doi.org/10.3389/fmicb.2015.01559)
- Birhane E, Sterck F, Fetene M, Bongers F, Kuyper T (2012) Arbuscular mycorrhizal fungi enhance photosynthesis, water use efficiency, and growth of frankincense seedlings under pulsed water availability conditions. Oecologia 169:895–904
- Bonfante P, Bianciotto V (1995) Presymbiotic versus symbiotic phase in arbuscular endomycorrhizal fungi: morphology and cytology. In: Vanna A, Hock B (eds) Mycorrhiza: structure, function, molecular biology, and biotechnology, pp 229–247
- Bonfante P, Genre A (2010) Mechanisms underlying beneficial plant-fungus interactions in mycorrhizal symbiosis. Nat Commun 1:48
- Borde MY, Dudhane MP, Jite PK (2010) AM fungi influences the photosynthetic activity, growth and antioxidant enzymes in Allium sativum L. under salinity condition. Notulae Sci Biol 2(4): 64–71
- Briccoli BC, Santilli E, Lombardo L (2015) Effect of arbuscular mycorrhizal fungi on growth and on micronutrient and macronutrient uptake and allocation in olive plantlets growing under high total Mn levels. Mycorrhiza 25:97–108
- Bucher M (2007) Functional biology of plant phosphate uptake at root and mycorrhiza interfaces. New Phytol 173(1):11–26
- Bucking H, Liepold E, Ambilwade P (2012) The role of the mycorrhizal symbiosis in nutrient uptake of plants and the regulatory mechanisms underlying these transport processes. In: Dhal NK, Sahu SC (eds) Plant science. IntechOpen. <https://doi.org/10.5772/52570>
- Castellano MA, Molina R (1989) Mycorrhizae. In: Landis TD, Tinus RW, McDonald SE, Barnett JP (eds) The container tree nursery manual, Agriculture handbook. 674, vol 5. U.S. Department of Agriculture, Forest Service, Washington, DC, pp 101–167
- Chen M, Arato M, Lorenzo B, Eva N, Didier R (2018) Beneficial services of arbuscular mycorrhizal fungi—from ecology to application. Front Plant Sci 9:1–14
- Conrath U, Beckers GJM, Flors V, Garcia-Agustin P, Jakab G, Mauch F (2006) Priming: getting ready for battle. Mol Plant Microbe Interact 19:1062–1071
- Dar MH, Reshi ZA (2017) Vesicular arbuscular mycorrhizal (VAM) fungi—as a major biocontrol agent in modern sustainable agriculture system. Russ Agric Sci 43(2):138–143
- Davies FT Jr, Olalde-Portugal V, Aguilera-Gomez L, Alvarado MJ, Ferrera-Cerrato RC, Boutton TW (2002) Alleviation of drought stress of Chile ancho pepper (Capsicum annuum L. cv. San Luis) with arbuscular mycorrhiza indigenous to Mexico. Sci Hortic 92(3–4):347–359
- Dickson S, Smith FA, Smith SE (2007) Structural differences in arbuscular mycorrhizal symbioses: more than 100 years after Gallaud, where next? Mycorrhiza 17:375–393
- Dong Y, Zhu YG, Smith FA et al (2008) Arbuscular mycorrhiza enhanced arsenic resistance of both white clover *(Trifolium repens Linn.)* and ryegrass *(Lolium perenne L.)* plants in an arseniccontaminated soil. Environ Pollut 155(1):174–181
- El-Nashar YI (2017) Response of snapdragon Antirrhinum majus L. to blended water irrigation and arbuscular mycorrhizal fungi inoculation: uptake of minerals and leaf water relations. Photosynthetica 55(2):201–209
- Frank B (1885) Über die auf Wurzelsymbiosenberuhende Ernährunggewisser Bäumedurchunterirdische Pilze. Berichte der DeutschenBotanischen Gesellschaft 3:128–145
- Gamalero E, Lingua G, Berta G, Glick BR (2009) Beneficial role of plant growth promoting bacteria and arbuscular mycorrhizal fungi on plant responses to heavy metal stress. Can J Microbiol 55:501–514
- Garcia-Garrido JM, Ocampo JA (2002) Regulation of the plant defence response in arbuscular mycorrhizal symbiosis. J Exp Bot 53(373):1377–1386
- Garg N, Chandel S (2012) Role of arbuscular mycorrhizal (AM) fungi on growth, cadmium uptake, osmolyte, and phytochelatin synthesis in Cajanus cajan (L.) Millsp. under NaCl and Cd stresses. J Plant Growth Regul 31(3):292–308
- Garg N, Manchanda G (2007) Symbiotic nitrogen fixation in legume nodules: process and signaling: a review. Agron Sustain Dev 27:59–68
- Gentili F, Jumpponen A (2006) Potential and possible uses of bacterial and fungal biofertilizers. In: Rai MK (ed) Handbook of microbial biofertilizers. Food Products Press, New York, pp 1–28
- Gerdemann JW (1968) Vesicular-arbuscular mycorrhizae and plant growth. Annu Rev Phytopathol 6:397–418
- Ghazi J, Al-Karaki GN (2001) Mycorrhizal influence on fruit yield and mineral content of tomato grown under salt stress. J Plant Nutr 24:1311–1323
- Gholamhoseini M, Ghalavand A, Dolatabadian A (2013) Effects of arbuscular mycorrhizal inoculation on growth, yield, nutrient uptake and irrigation water productivity of sunflowers grown under drought stress. Agric Water Manag 117:106–114
- Giovannetti M, Tolosano M, Volpe V, Kopriva S, Bonfante P (2014) Identification and functional characterization of a sulfate transporter induced by both sulfur starvation and mycorrhiza formation in Lotus japonicus. New Phytol 204:609–619
- Giri B, Kapoor R, Mukerji KG (2003) Influence of arbuscular mycorrhizal fungi and salinity on growth, biomass and mineral nutrition of Acacia auriculiformis. Biol Fertil Soils 38:170–175
- Giri B, Prasad R, Wu Q-S, Varma A (2019) Biofertilizers for sustainable agriculture and environment. Springer International Publishing (ISBN: 978-3-030-18932-7). [https://www.springer.](https://www.springer.com/gp/book/9783030189327) [com/gp/book/9783030189327](https://www.springer.com/gp/book/9783030189327)
- Govindarajulu M, Pfeffer P, Jin H (2005) Nitrogen transfer in the arbuscular mycorrhizal symbiosis. Nature 435:819–823
- Hao Z, Fayolle L, Van TD, Gianinazzi-Pearson V, Gianinazzi S (2009) Mycorrhiza reduce development of nematode vector of grapevine fanleaf virus in soils and root systems. In: Boudon-Padfieu E (ed) Extended abstract 16th meeting of ICVG, Dijon, France, pp 100–1001
- Harrier LA, Watson CA (2004) The potential role of arbuscular mycorrhizal (AM) fungi in the bioprotection of plants against soil-borne pathogens in organic and/or other sustainable farming systems. Pest Manag Sci 60:149–157
- Hartley SE, Gange AC (2009) Impacts of plant symbiotic fungi on insect herbivores: mutualism in a multitrophic context. Annu Rev Entomol 54:323–342
- Hasanuzzaman M, Gill SS, Fujita M (2013) Physiological role of nitric oxide in plants grown under adverse environmental conditions. In: Tuteja N, Gill SS (eds) Plant acclimation to environmental stress. Springer, New York, pp 269–322
- Hashem A et al (2016) Alleviation of cadmium stress in Solanum lycopersicum L. by arbuscular mycorrhizal fungi via induction of acquired systemic tolerance. Saudi J Biol Sci 23:272–281
- He XH, Nara K (2007) Element biofortification: can mycorrhizas potentially offer a more effective and sustainable pathway to curb human malnutrition? Trends Plant Sci 12:331–333
- Hosny M, Gianinazzi-Pearson V, Dulieu H (1998) Nuclear DNA contents of 11 fungal species in Glomales. Genome 41:422–429
- Igiehon NO, Olubukola OB (2017) Biofertilizers and sustainable agriculture: exploring arbuscular mycorrhizal fungi. Appl Microbiol Biotechnol 101(12):4871–4881
- Jacott CN, Murray JD, Ridout CJ (2017) Trade-offs in arbuscular mycorrhizal symbiosis: disease resistance, growth responses and perspectives for crop breeding. Agronomy 7:75
- Janeeshma E, Puthur JT (2020) Direct and indirect influence of arbuscular mycorrhizae on enhancing metal tolerance of plants. Arch Microbiol 202:1–16
- Javaid A (2007) Allelopathic interactions in mycorrhizal associations. Allelopath J 20:29–42
- Jianjian L, Junli L, Jinhui L, Miaomiao C, Yujuan H, Yuan T, Aiqun C, Guohua X (2019) The potassium transporter SlHAK10 is involved in mycorrhizal potassium uptake. Plant Physiol 80(1):465–479
- Johansen A, Jakobsen I, Jensen ES (1994) Hyphal N transport by a vesicular-arbuscular mycorrhizal fungus associated with cucumber grown at three nitrogen levels. Plant Soil 160:1–9
- Kapoor R, Evelin H, Mathur P, Giri B (2013) Arbuscular mycorrhiza: approaches for abiotic stress tolerance in crop plants for sustainable agriculture. In: Tuteja N, Singh Gill S (eds) Plant acclimation to environmental stress. Springer, pp 359–401
- Khan AG, Kuek C, Chaudhry TM, Khoo CS, Hayes WJ (2000) Plants, mycorrhizae and phytochelators in heavy metal contaminated land remediation. Chemosphere 41:197–207
- Khaosaad T, Garcia-Garrido JM, Steinkellner S, Vierheilig H (2007) Take all diseases is systemically reduced in roots of mycorrhizal barley plants. Soil Biol Biochem 39:727–734
- Kobae Y, Tamura Y, Takai S, Banba M, Hata S (2010) Localized expression of arbuscular mycorrhiza-inducible ammonium transporters in soybean. Plant Cell Physiol 51:1411–1415
- Kumar A, Verma JP (2018) Does plant-microbe interaction confer stress tolerance in plants: a review? Microbiol Res 207:41–52
- Kumar R, Jalali BL, Chand H (2002) Influence of vesicular arbuscular mycorrhizal fungi on growth and nutrient uptake in chickpea. J Mycol Plant Pathol 32(1):11–15
- Kumar M, Prasad R, Kumar V, Tuteja N, Varma A (2017) Mycorrhizal fungi under biotic and abiotic stress. In: Varma A, Prasad R, Tuteja N (eds) Mycorrhiza. Springer International Publishing AG, pp 57–70
- Kumar P, Tapan Kumar T, Singh S, Tuteja N, Prasad R, Singh J (2020) Potassium: a key modulator for cell homeostasis. J Biotechnol 324:198–210. <https://doi.org/10.1016/j.jbiotec.2020.10.018>
- Kwak MJ, Kong HG, Choi K, Kwon SK, Song JY, Lee J, Lee PA, Choi SY, Seo M, Lee HJ, Jung EJ (2018) Rhizosphere microbiome structure alters to enable wilt resistance in tomato. Nat Biotechnol 36(11):1100–1109
- Li T, Hu Y, Hao Z, Li H, Wang Y, Chen B (2013) First cloning and characterization of two functional aquaporin genes from an arbuscular mycorrhizal fungus Glomus intraradices. New Phytol 197:617–630
- Li J, Meng B, Chai H, Yang X, Song W, Li S, Sun W (2019) Arbuscular mycorrhizal fungi alleviate drought stress in C3 (Leymus chinensis) and C4 (Hemarthria altissima) grasses via altering antioxidant enzyme activities and photosynthesis. Front Plant Sci 10:499
- Linderman RG, Davis EA (2004) Evaluation of commercial inorganic and organic fertilizer effects on arbuscular mycorrhizae formed by Glomus intraradices. Hortic Technol 14(2):196-202
- López-Ráez JA, Matusova R, Cardoso C, Jamil M, Charnikhova T, Kohlen W, Verstappen F, Ruyter-Spira C, Bouwmeester HJ (2009) Strigolactones: ecological significance and use as a target for parasitic plant control. Pest Manag Sci 64:471–477
- Mahajan S, Tuteja N (2005) Cold, salinity and drought stresses: an overview. Arch Biochem Biophys 444:139–158
- Malhi GS, Kaur M, Kaushik P, Alyemeni MN, Alsahli AA, Ahmad P (2021) Arbuscular mycorrhiza in combating abiotic stresses in vegetables: an eco-friendly approach. Saudi J Biol Sci 28(2):1465–1476
- Malla R, Prasad R, Kumari R, Giang PH, Pokharel U, Oelmueller R, Varma A (2004) Phosphorus solubilizing symbiotic fungus Piriformospora indica. Endocytobiosis Cell Res 15(2):579-600 Marschner H, Dell B (1994) Nutrient uptake in mycorrhizal symbiosis. Plant Soil 159:89–102
- Marulanda A, Porcel R, Barea JM, Azcón R (2007) Drought tolerance and antioxidant activities in
- lavender plants colonized by native drought-tolerant or drought sensitive Glomus species. Microb Ecol 54:543–552
- Maya MA, Matsubara Y (2013) Influence of arbuscular mycorrhiza on the growth and antioxidative activity in Cyclamen under heat stress. Mycorrhiza 23(5):381–390
- Mohanta TK, Bae H (2015) Functional genomics and signaling events in mycorrhizal symbiosis. J Plant Interact 10(1):21–40
- Mohsin SM, Hasanuzzaman M, Nahar K, Hossain M, Bhuyan MH, Parvin K, Fujita M (2020) Tebuconazole and trifloxystrobin regulate the physiology, antioxidant defense and methylglyoxal detoxification systems in conferring salt stress tolerance in Triticum aestivum L. Physiol Mol Biol Plants 26(6):1139–1154
- Morgan JB, Connolly EL (2013) Plant-soil interactions: nutrient uptake. Nat Educ Knowl 4:2
- Moussa HR, Abdel-Aziz SM (2008) Comparative response of drought tolerant and drought sensitive maize genotypes to water stress. Aust J Crop Sci 1:31-36
- Nath M, Bhatt D, Prasad R, Tuteja N (2017) Reactive oxygen species (ROS) metabolism and signaling in plant-mycorrhizal association under biotic and abiotic stress conditions. In: Varma A, Prasad R, Tuteja N (eds) Mycorrhiza. Springer International Publishing AG, pp 223–232
- Nath M, Bhatt D, Bhatt MD, Prasad R, Tuteja N (2018) Microbe-mediated enhancement of nitrogen and phosphorus content for crop improvement. In: Prasad R, Gill SS, Tuteja N (eds) Crop improvement through microbial biotechnology. Elsevier, pp 291–301
- Palmer C, Guerinot ML (2009) A question of balance: facing the challenges of Cu, Fe and Zn homeostasis. Nat Chem Biol 5:333–340
- Parihar M, Meena VS, Mishra PK, Rakshit A, Choudhary M, Yadav RP, Rana K, Bisht JK (2019) Arbuscular mycorrhiza: a viable strategy for soil nutrient loss reduction. Arch Microbiol 201(6): 723–735
- Paszkowski U (2006) A journey through signalling in arbuscular mycorrhizal symbioses. New Phytol 172(1):35–46
- Puig S, Penarrubia L (2009) Placing metal micronutrients in context: transport and distribution in plants. Curr Opin Plant Biol 12:299–306
- Rai MK, Kalia RK, Singh R, Gangola MP, Dhawan AK (2011) Developing stress tolerant plants through in vitro selection—an overview of the recent progress. Environ Exp Bot 71:89–98
- Sadhana B (2014) Review article arbuscular mycorrhizal fungi (AMF) as a biofertilizer—a review. Int J Curr Microbiol App Sci 3(4):384–400
- Schützendübel A, Polle A (2002) Plant responses to abiotic stresses: heavy metal-induced oxidative stress and protection by mycorrhization. J Exp Bot 53(372):1351–1365
- Sharma A, Singh A, Raina M, Kumar D (2021) Plant-fungal association: an ideal contrivance for combating plant stress tolerance. In: Prasad R, Nayak SC, Kharwar RN, Dubey NK (eds) Mycoremediation and environmental sustainability. Springer Nature, Singapore
- Shen J, Yuan L, Zhang J, Li H, Bai Z, Chen X, Zhang F (2011) Phosphorus dynamics: from soil to plant. Plant Physiol 156(3):997–1005
- Sieverding E (1995) Vesicular-arbuscular mycorrhiza management in tropical agrosystems. AGRIS, GTZ No. 224. Eschborn 371
- Smith SE, Read DJ (2008) Mycorrhizal symbiosis, 3rd edn. Academic, Amsterdam
- Smith SE, Smith FA (2011) Roles of arbuscular mycorrhizas in plant nutrition and growth: new paradigms from cellular to ecosystem scales. Annu Rev Plant Biol 62:227–250
- Solaiman ZM (2014) Contribution of arbuscular mycorrhizal fungi to soil carbon sequestration. In: Mycorrhizal fungi: use in sustainable agriculture and land restoration. Springer, Berlin, pp 287–296
- Srivastava P, Saxena B, Giri B (2018) Arbuscular mycorrhizal fungi: green approach/technology for sustainable agriculture and environment. In: Mycorrhiza—nutrient uptake, biocontrol, eco restoration, 4th edn. https://doi.org/10.1007/978-3-319-68867-1_20
- Subramanian KS, Charest C (1998) Arbuscular mycorrhizae and nitrogen assimilation in maize after drought and recovery. Physiol Plant 102:285–296
- Suhag M (2016) Potential of biofertilizers to replace chemical fertilizers. Int J Adv Res Sci Eng Technol 3(5):163–167
- Talaat NB, Shawky BT (2012) 24-Epibrassinolide ameliorates the saline stress and improves the productivity of wheat (Triticum aestivum L.). Environ Exp Bot 82:80–88
- Wagg C, Pautler M, Massicotte HB, Peterson RL (2008) The co-occurrence of ectomycorrhizal, arbuscular mycorrhizal, and dark septate fungi in seedlings of four members of the Pinaceae. Mycorrhiza 18(2):103–110
- Wahid A, Gelani S, Ashraf M, Foolad MR (2007) Heat tolerance in plants: an overview. Environ Exp Bot 61:199–223
- Walder F, Brule D, Koegel S, Wiemken A, Boller T, Courty PE (2015) Plant phosphorus acquisition in a common mycorrhizal network: regulation of phosphate transporter genes of the Pht1 family in sorghum and flax. New Phytol 205:1632–1645
- Wang B, Qiu YL (2006) Phylogenetic distribution and evolution of mycorrhizas in land plants. Mycorrhiza 16:299–363
- Wang C, Tian B, Yu Z, Ding J (2020) Effect of different combinations of phosphorus and nitrogen fertilization on arbuscular mycorrhizal fungi and aphids in wheat. Insects 11(6):365
- Whipps JM (2004) Prospects and limitations for mycorrhizas in biocontrol of root pathogens. Can J Bot 82:1198–1227
- Whiteside MD, Gercia MO, Treseder KK (2012) Amino acid uptake in arbuscular mycorrhizal plants. PLoS One 7(10):e47643
- Yong X et al (2014) Enhanced cadmium resistance and accumulation in Pseudomonas putida KT2440 expressing the phytochelatin synthase gene of Schizosaccharomyces pombe. Lett Appl Microbiol 58:255–226
- Zhu XC, Song FB, Xu HW (2010a) Arbuscular mycorrhizae improve low temperature stress in maize via alterations in host water status and photosynthesis. Plant Soil 331:129–137
- Zhu XC, Song FB, Xu HW (2010b) Effects of arbuscular mycorrhizal fungi on photosynthetic characteristics of maize under low temperature stress. Acta Ecol Sin 21:470–475